Supporting information

Stable and Reusable Ni-based Nanoparticles for General and Selective Hydrogenation of Nitriles to Amines

Zhuang Ma,^a Vishwas G. Chandrashekhar,^a Bei Zhou,^a Asma M. Alenad,^b Nils Rockstroh,^a Stephan Bartling,^a Matthias Beller^{*},^a and Rajenahally V. Jagadeesh^{*a}

^a Leibniz-Institut für Katalyse e.V., Albert-Einstein-Str. 29a, Rostock, D-18059, Germany

^b Chemistry Department, College of Science, Jouf University, P.O. Box: 2014, Sakaka, Saudi Arabia

*E-Mails: matthias.beller@catalysis.de; jagadeesh.rajenahally@catalysis.de

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S1. Materials and methods

All nitriles were obtained commercially from various chemical companies. Unless otherwise stated all reagents were used directly without purification. Nickle (II) nitrate hexahydrate (cat no.139267-100G), cobalt (II) nitrate hexahydrate (cat no. 239267-100G), copper (II) nitrate trihydrate (cat no. 61194-100G), iron (III) nitrate nonahydrate (cat no. 216828-100G) and manganese (II) nitrate tetrahydrate (cat no. 63547-100G) were purchased from Sigma Aldrich. Silica suspension (Silica LUDOX® AS-40 colloidal silica, cat no. 420840-1L) was purchased from Sigma Aldrich.

Preparation of catalytic materials was performed in a tube furnace (Lenton Thermal Design Ltd). All catalytic reactions were carried out in 300 mL autoclave (PARR Instrument Company). To avoid unspecific reactions, all catalytic reactions were carried out either in glass vials, which were placed inside the autoclave, or glass/Teflon vessel fitted autoclaves.

GC and GC-MS were recorded on Agilent 6890N instrument. GC conversions and yields were determined by GC-FID, HP6890 chromatograph with FID detector, column HP530 m x 250 mm x $0.25 \ \mu m$.

NMR spectra were recorded on Bruker 300 Fourier, Bruker AV 300 and Bruker AV 400 spectrometers. Chemical shifts are reported in ppm relative to the deuterated solvent. Coupling constants are expressed in Hertz (Hz). The following abbreviations are used: s = singlet, d = doublet, t = triplet and m = multiple. The residual solvent signals were used as references for ¹H and ¹³C NMR spectra (CDCl₃: $\delta H = 7.26$ ppm, $\delta C = 77.12$ ppm; DMSO-d₆: $\delta H = 2.50$ ppm, $\delta C = 39.52$ ppm).

High resolution mass spectra (HRMS) were obtained either from a MAT 95 XP from Thermo (EI) or from an HPLC system 1200 and downstream ESI-TOF-MS 6210 from Agilent (ESI).

XRD powder patterns were recorded on a Panalytical X'Pert diffractometer equipped with a Xcelerator detector or on a Panalytical Empyrean diffractometer equipped with a PIXcel 3D detector system, both used with automatic divergence slits and Cu K α radiation (40 kV, 40 mA). Cu β -radiation was excluded by using nickel filter foil. Peak positions and profile were fitted with Pseudo-Voigt function using the Panalytical HighScore Plus software package Phase identification was done by using the PDF-2 database of the International Center of Diffraction Data (ICDD)

Scanning transmission electron microscopy (STEM) measurements were performed with an aberration-corrected JEM-ARM200F (JEOL, Corrector: CEOS) at 200 kV. The microscope was equipped with an Enfinium ER (GATAN) electron energy-loss spectrometer for chemical analysis. High-Angle Annular Dark Field (HAADF) and Annular Bright Field (ABF) detectors were used for general imaging, the Annular Dark Field (ADF) detector was used for position control for EELS acquisition. EELS was done at a camera length of 4 cm, an illumination semi angle of 27.8 mrad and a filter entrance aperture semi angle of 41.3 mrad. The solid samples were deposited without any

pretreatment on a holey carbon supported Cu-grid (mesh 300) and transferred to the microscope.

The X-ray Photoelectron Spectroscopy (XPS) measurements were performed on an ESCALAB 220iXL (Thermo Fisher Scientific) with monochromated Al K α radiation (E = 1486.6 eV). Samples are prepared on a stainless-steel holder with conductive double-sided adhesive carbon tape. The electron binding energies were obtained with charge compensation using a flood electron source and referenced to the C 1s core level of adventitious carbon at 284.8 eV (C-C and C-H bonds). For quantitative analysis the peaks were deconvoluted with Gaussian-Lorentzian curves using the software Unifit 2021. The peak areas were normalized by the transmission function of the spectrometer and the element specific sensitivity factor of Scofield.

S2. Procedure for the preparation of catalysts

In a 250 mL dried round bottom flask, 280 mg Ni(NO₃)₂·6H₂O was dissolved in 50 mL distilled water and then 1.25 g silica suspension (Silica LUDOX® AS-40 colloidal silica) was added. The round bottom flask containing reaction mixture was stirred at 65 °C for 5 h. Then, the water was removed by rotary evaporator and the obtained solid was dried overnight in an oven at 110 °C. The dried solid material was grinded to a fine powder and calcined in a tube furnace at 600 °C for 6 h under air atmosphere. Then, the calcined material was reduced under 10% H₂/N₂ flow at different temperature (300 °C, 400 °C, 500 °C, 600 °C and 700 °C) for 5 h. The resulting materials are represented as M-NPs@SiO₂-T, where M and T denotes metal and reduction temperature.

Elemental analysis of the optimal catalyst, Ni-NPs@SiO₂-500 (wt%: Si=33.4%, C=43.5%, H=9.7%, Ni=8.9% by ICP-OES).

Same procedure has been applied for the preparation of other catalytic materials such as, Fe-NPs@SiO₂-500, Mn-NPs@SiO₂-500, Co-NPs@SiO₂-500 and Cu-NPs@SiO₂-500 using $Fe(NO_3)_3 \cdot 9H_2O$, $Mn(NO_3)_2 \cdot 4H_2O$, $Co(NO_3)_2 \cdot 6H_2O$, and $Cu(NO_3)_2 \cdot 3H_2O$, respectively.

S3. Characterization of catalysts

<u>XRD</u>



Figure S1. XRD patterns of NiO@SiO₂ calcined at 600 °C without reduction catalyst.



Figure S2. XRD patterns of Ni-NPs@SiO₂-300 catalyst.



Figure S3. XRD patterns of Ni-NPs@SiO₂-400 catalyst.



Figure S4. XRD patterns of Ni-NPs@SiO₂-500 catalyst.



Figure S5. XRD patterns of Ni-NPs@SiO₂-600 catalyst.



Figure S6. XRD patterns of Ni-NPs@SiO₂-700 catalyst.







Figure S8. Selected STEM-HAADF images of Ni@SiO₂-500 showing the (partial) presence of an oxidic layer/shell around a metallic Ni core.

<u>STEM</u>



Figure S9. Selected STEM-HAADF images of Ni-NPs@SiO₂-500R (after one run) showing the presence of Ni (oxide) at the interfaces of the support particles.



Figure S10. STEM-ADF image of Ni-NPs@SiO₂-500 (left) and corresponding Ni L edge electronenergy loss (EEL) spectra of the highlighted areas (right). In area 1, the L3/L2 ratio implies the presence of metallic Ni, while the edge shape of spectrum 2 points towards the occurrence of NiO^{S1}. The EEL spectra are background subtracted and deconvolved.

<u>S4. General procedure for the hydrogenation of nitriles to primary amines</u>

The magnetic stirring bar and 0.5 mmol of corresponding nitrile were transferred to 8 mL glass vial and then 2 mL MeOH were added. Next, 15-30 mg of catalyst (Ni-NPs@SiO₂-500; 4.5-9.0 mol% Ni) was added and the vial was fitted with septum, cap, and needle. The reaction vials (8 vials with different substrates at a time) were placed into a 300 mL autoclave. The autoclave was flushed with hydrogen twice at 20 bar pressure and then it was pressurized with 5-7 bar ammonia gas and 35 bar hydrogen. The autoclave was placed into an aluminium block preheated at 60 and 80 °C and the reactions were stirred for required time. After the completion of the reactions, the autoclave was cooled to room temperature. The remaining ammonia and hydrogen were discharged and the vials containing reaction products were removed from the autoclave. The solid catalyst was filtered off and washed thoroughly with ethyl acetate. The reaction products were analyzed by GC-MS. The corresponding primary amines were purified by column chromatography. Then, the resulted free amines were converted to their respective hydrochloride salt and characterized by NMR and HR-MS analysis. For converting into hydrochloride salt of amine, 1-2 mL methanolic HCl (1.5M HCl in methanol) was added to the ether solution of respective amine and stirred at room temperature for 4-5 h. Then, solvent was removed by rotary evaporation, and the resulted hydrochloride salt of amine is dried under high vacuum. The yields were determined by GC for the selected amines: After completion of the reaction, n-hexadecane (50 µL) as standard was added to the reaction vials and the reaction products were diluted with ethyl acetate followed by filtration using plug of silica and then analyzed by GC.



Figure S11. a: Reaction pathway for the catalytic hydrogenation of nitriles. b: control experiments.

Table S1. Hydrogenation of 5-chloropicolinonitrile to (5-chloropyridin-2-yl)methanamine: Testing of different solvent^a.

	Ni-NPs@SiO ₂ -500			
CI 1	35 bar H ₂ , 5-7 bar NH ₃ 80 °C, 24 h			u
Entry	Solvent	Conv (%)	Yield (%) of	
			2	3
1	MeOH	>99	88	11
2	EtOH	90	79	10
3	t-BuOH	76	43	33
4	i-PrOH	91	80	11
5	Toluene	40	21	19
6	THF	54	29	25
7	MeCN	-	-	-
8	H ₂ O	-	-	-
9ь	MeOH	>99	85	13

Reaction conditions^a: 0.5 mmol 5-chloropicolinonitrile, 30 mg Ni-NPs@SiO₂-500 (9.0 mol% Ni), 35 bar H₂, 5-7 bar NH₃, 2 mL solvent, 80 °C, 24 h. ^b: same as "a" using NiCl₂ as nickel precursor. Conversions and yields were determined by GC using n-hexadecane standard.

Table S2. Hydrogenation of 5-chloropicolinonitrile to (5-chloropyridin-2-yl)methanamine: Testing of different noble metal catalyst.

 $\underbrace{\operatorname{Cl} \underbrace{\underset{1}{\overset{N}{\operatorname{Noble metal catalyst}}}_{1} \underbrace{\operatorname{Noble metal catalyst}}_{20 \text{ bar H}_2, 5-7 \text{ bar NH}_3,}}_{1} \underbrace{\operatorname{Cl} \underbrace{\underset{2}{\overset{N}{\operatorname{N}}}}_{2} + \underbrace{\operatorname{Cl} \underbrace{\underset{N}{\overset{N}{\operatorname{N}}}}_{3} \underbrace{\underset{N}{\overset{N}{\operatorname{N}}}}_{3} \underbrace{\operatorname{Cl}}_{4} + \underbrace{\operatorname{Cl} \underbrace{\underset{N}{\overset{N}{\operatorname{N}}}}_{7} \underbrace{\operatorname{NH}}_{4} + \underbrace{\operatorname{NH}}_{7} \underbrace{\operatorname{Cl}}_{4} + \underbrace{\operatorname{Cl} \underbrace{\underset{N}{\overset{N}{\operatorname{N}}}}_{7} \underbrace{\operatorname{Cl}}_{4} + \underbrace{\operatorname{Cl} \underbrace{\underset{N}{\operatorname{N}}}}_{7} \underbrace{\operatorname{Cl}}_{4} + \underbrace{\operatorname{Cl} \underbrace{\underset{N}{\operatorname{N}}}_{7} \underbrace{\operatorname{Cl}}_{4} \underbrace{\operatorname{Cl}}_{7} \underbrace{\operatorname{Cl}}_{4} + \underbrace{\operatorname{Cl}}_{7} \underbrace{\operatorname{Cl}}_$

Catalyst	Conv (%)	Yield of 2 (%)	Yield 013 (%)	Yield 014 (%)	Yield 01 / (%)			
5% Ru/C	42	11	<5	<5	17			
5% Rh/C	35	9	<5	<5	11			
10% Pd/C	50	19	<5	<5	21			
5% Pt/C	20	8	-	-	10			
Reaction conditions: 0.5 mmol nitrile, 4 mol% catalyst, 20 bar H ₂ , 5-7 bar NH ₃ , 2 mL MeOH,								
40 °C, 12 h.								



Table S3. Hydrogenation of (5-chloropyridin-2-yl)methanamine to product 3 and 4.

Figure S11. Hydrogenation of 5-chloropicolinonitrile to (5-chloropyridin-2 yl)methanamine: Testing different reaction time, temperature, catalyst amount and H₂ pressure. Reaction conditions: 0.5 mmol 5-chloropicolinonitrile, $10-30 \text{ mg Ni-NPs} @SiO_2-500 (3.0-9.0 \text{ mol}\% \text{ Ni})$, $15-35 \text{ bar H}_2$, $5-7 \text{ bar NH}_3$, 2 mL MeOH, $60-80 \degree$ C, 6-24 h. Conversions and yields are determined by GC using n-hexadecane

standard.



Figure S12. Hydrogenation of 5-chloropicolinonitrile to (5-chloropyridin-2yl)methanamine: Kinetic profiles for recycled experiments.

S5. Catalyst recycling

The magnetic stirring bar, 10 mmol benzonitrile, 320 mg Ni-NPs@SiO₂-500 (4.8 mol% Ni) were transferred to 300 mL glass fitted autoclave and 40 mL of dry MeOH was added. The autoclave was flushed with hydrogen twice at 20 bar pressure and then it was pressurized with 5-7 bar ammonia gas and 35 bar hydrogen. The autoclave was placed into an aluminum block heated at 60 °C and stirred for 20 h. After the completion of the reaction, the autoclave was cooled to room temperature. The remaining ammonia and hydrogen were discharged, and reaction products were removed from the autoclave. To the reaction products, 500 μ L n-hexadecane as standard was added. The catalyst was separated by centrifugation and the centrifugate containing reaction products were subjected to GC analysis. The separated catalyst was washed with water, methanol and ethyl acetate and then dried under vacuum. The dried catalyst was used for the nextrun without further purification or reactivation.

S6. Procedure of reaction upscaling

To a glass fitted 300 mL autoclave, the magnetic stirring bar and corresponding nitrile were transferred and then 30-70 mL of dry MeOH were added. Next, the required amount of catalyst (Ni-NPs@SiO₂-500, 4.5-9.0 mol%; 30 mg for each 0.5 mmol substrate **27**; 15 mg for each 0.5 mmol substrate **35**, **39** and **72**; 20 mg for each 0.5 mmol substrate **83** and **103**) was added. The autoclave was closed and flushed with hydrogen twice at 40 bar pressure and then it was pressurized with 5-7 bar ammonia gas and 35 bar hydrogen. The autoclave was placed into an aluminium block preheated at 60 and 80 °C and the reactions were stirred for required time. After completion of the reaction, the

autoclave was cooled to room temperature. The remaining ammonia and hydrogen were discharged, and the reaction products were removed from the autoclave. The solid catalyst was filtered off and washed thoroughly with ethyl acetate. The reaction products were analyzed by GC-MS and the corresponding primary amines were purified by column chromatography (silica; n-hexane-ethyl acetate mixture).

S7. References

S1. Jeangros, Q.; Hansen, T. W.; Wagner, J. B.; Dunin-Borkowski, R. E.; Hébert, C.; Hessler-Wyser,
A. Oxidation mechanism of nickel particles studied in an environmental transmission electron microscope. Acta. Mater 2014, 67, 362-372.

S8. NMR data

¹**H NMR (300 MHz, DMSO-***d*₆) δ 8.35 (dd, *J* = 2.5, 0.8 Hz, 1H), 7.85 – 7.78 (m, 1H), 7.45 (d, *J* = 8.2 Hz, 1H), 3.73 (s, 2H), 2.87 (s, 2H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 149.19, 148.54, 139.42, 139.18, 124.17, 42.70. HRMS (ESI-TOF): calcd for C₆H₇ClN₂ [M+H] 143.0376; found 143.0375.

NH3⁺CI⁻

¹H NMR (300 MHz, DMSO-*d*₆) δ 8.83 (s, 5H), 7.60 (d, *J*=4.7 Hz, 2H), 4.07 (s, 2H). ¹³C NMR (75 MHz, DMSO-*d*₆) δ 150.16, 143.41, 124.53, 41.41. HRMS (ESI-TOF): calcd for C₆H₈N₂ [M+H] 108.0687; found 109.0765.



¹H NMR (300 MHz, DMSO- d_6) δ 8.52 (d, J = 2.2 Hz, 1H), 8.41 (dd, J = 4.8, 1.7 Hz, 1H), 7.74 (dddt, J = 7.8, 2.4, 1.6, 0.7 Hz, 1H), 7.32 (ddd, J = 7.8, 4.8, 0.9 Hz, 1H), 3.73 (s, 2H). ¹³C NMR (75 MHz, DMSO- d_6) δ 149.17, 147.94, 139.83, 135.23, 123.73, 43.59.

Br N

¹H NMR (300 MHz, DMSO- d_6) δ 8.32 (dq, J = 2.4, 0.8 Hz, 1H), 7.75 – 7.69 (m, 1H), 7.57 (d, J = 2.4, 0.8 Hz, 1H), 7.75 – 7.69 (m, 1H), 7.57 (d, J = 2.4, 0.8 Hz, 1H), 7.75 – 7.69 (m, 1H), 7.57 (d, J = 2.4, 0.8 Hz, 1H), 7.75 – 7.69 (m, 1H), 7.57 (d, J = 2.4, 0.8 Hz, 1H), 7.75 – 7.69 (m, 1H), 7.57 (d, J = 2.4, 0.8 Hz, 1H), 7.75 – 7.69 (m, 1H), 7.57 (d, J = 2.4, 0.8 Hz, 1H), 7.75 – 7.69 (m, 1H), 7.57 (d, J = 2.4, 0.8 Hz, 1H), 7.75 – 7.69 (m, 1H), 7.57 (d, J = 2.4, 0.8 Hz, 1H), 7.57 (d, J = 2.4, 0.8 Hz, 1H), 7.75 – 7.69 (m, 1H), 7.57 (d, J = 2.4, 0.8 Hz, 1H), 7.57 (d, J = 2.4,

8.2 Hz, 1H), 3.70 (d, J=0.7 Hz, 2H), 1.95 (s, 2H).
¹³C NMR (75 MHz, DMSO-d₆) δ 149.81, 139.91, 139.41, 139.02, 127.91, 42.77.
HRMS (ESI-TOF): calcd for C₆H₇N₂Br [M+H] 186.9871; found 186.9870.

¹**H NMR (300 MHz, DMSO-***d*₆) δ 8.70 (s, 3H), 8.39 (s, 2H), 8.15 – 8.06 (m, 2H), 7.11 – 7.05 (m, 1H), 3.94 (d, *J* = 5.2 Hz, 2H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 154.21, 145.59, 136.75, 118.66, 113.61, 38.85. HRMS (ESI-TOF): calcd for C₆H₉N₃ [M+H] 124.0797; found 124.0874.

¹H NMR (300 MHz, DMSO- d_6) δ 8.71 (d, J = 2.2 Hz, 1H), 8.06 – 7.96 (m, 1H), 7.80 (pent, J = 3.6 Hz, 1H), 3.86 (s, 2H), 2.28 (d, J = 5.2 Hz, 2H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 149.67, 145.28, 145.24, 144.82, 143.88, 136.94, 136.91, 127.74, 124.12, 120.56, 120.50, 116.88, 43.17.

HRMS (ESI-TOF): calcd for C₇H₇F₃N₂ [M+H] 177.0639; found 177.0641.

¹**H NMR (300 MHz, DMSO-***d*₆) δ 9.05 – 8.96 (m, 3H), 8.69 (dd, *J* = 8.3, 2.0 Hz, 1H), 7.98 (d, *J* = 8.3 Hz, 1H), 4.24 (q, *J* = 5.7 Hz, 2H), 2.78 (s, 3H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 153.67, 146.91, 141.82, 131.63, 127.76, 38.99, 19.37.

HRMS (ESI-TOF): calcd for C₇H₉ClN₂ [M+H] 156.0640; found 157.0643.

¹**H NMR (300 MHz, DMSO-***d*₆) δ 8.72 – 8.69 (m, 1H), 8.54 (dd, *J* = 2.7, 1.5 Hz, 1H), 8.48 (d, *J* = 2.6 Hz, 1H), 3.86 (d, *J* = 0.7 Hz, 2H), 2.01 (s, 2H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 158.82, 144.03, 143.86, 143.07, 45.87.

HRMS (ESI-TOF): calcd for C₅H₇N₃ [M+H] 109.0640; found 110.0718.



¹**H NMR (300 MHz, DMSO-***d*₆) δ 9.10 (d, *J* = 47.0 Hz, 2H), 8.72 (s, 3H), 8.36 (s, 1H), 4.05 – 3.95 (m, 2H), 2.51–2.44 (m, 3H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 163.38, 161.30, 144.54, 108.62, 35.38, 21.42.

HRMS (ESI-TOF): calcd for C₆H₉N₃ [M+H] 138.0905; found 138.0983.



¹H NMR (300 MHz, DMSO-*d*₆) δ 8.05 (ddd, *J* = 7.9, 1.4, 0.7 Hz, 1H), 7.90 (ddd, *J* = 8.0, 1.3, 0.6 Hz, 1H), 7.46 (ddd, *J* = 8.1, 7.2, 1.4 Hz, 1H), 7.38 (ddd, *J* = 7.7, 7.2, 1.3 Hz, 1H), 4.14 (s, 2H).
¹³C NMR (75 MHz, DMSO-*d*₆) δ 178.94, 153.86, 134.99, 126.22, 124.83, 122.57, 122.53, 44.38.
HRMS (ESI-TOF): calcd for C₈H₈N₂S [M+H] 164.2213; found 164.2215.



¹**H NMR (300 MHz, DMSO-***d*₆) δ 8.55 (d, *J* = 1.5 Hz, 1H), 8.42 (d, *J* = 1.6 Hz, 1H), 3.81 (s, 2H), 2.45 (s, 3H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 155.46, 151.49, 143.37, 142.47, 45.50, 21.13. HRMS (ESI-TOF): calcd for C₆H₉N₃ [M+H] 124.0875; found 124.0877.

NH₃⁺CI[−]

¹H NMR (300 MHz, DMSO-*d*₆) δ 8.28 (s, 3H), 7.73 (d, *J* = 31.7 Hz, 2H), 6.73 (s, 1H), 3.90 (s, 2H).
 ¹³C NMR (75 MHz, DMSO-*d*₆) δ 144.16, 142.38, 119.35, 111.60, 33.83.



¹**H NMR (300 MHz, DMSO-***d*₆) δ 8.70 (s, 3H), 7.72 (dd, *J* = 1.9, 0.9 Hz, 1H), 6.65 – 6.39 (m, 2H), 4.04 – 4.02 (m, 2H).

¹³C NMR (**75** MHz, DMSO-*d*₆) δ 148.15, 143.97, 111.37, 110.72, 35.35.



¹H NMR (300 MHz, DMSO-*d*₆) δ 7.62 – 7.48 (m, 2H), 7.30 – 7.18 (m, 2H), 6.77 (q, *J* = 0.9 Hz, 1H),

3.89 (d, *J* = 0.9 Hz, 2H).

¹³C NMR (**75** MHz, DMSO-*d*₆) δ 157.85, 154.67, 128.71, 124.15, 123.13, 121.20, 111.32, 104.06, 45.48.

HRMS (ESI-TOF): calcd for C₉H₉NO [M+H] 147.1771; found 147.1776.



¹**H NMR (300 MHz, DMSO-***d*₆) δ 8.55 (s, 3H), 7.53 (d, *J* = 4.8 Hz, 1H), 7.30 (s, 1H), 7.22 (s, 2H), 7.10 (s, 1H), 4.20 (s, 2H).

¹³C NMR (**75** MHz, DMSO-*d*₆) δ 141.10, 134.82, 134.26, 130.77, 128.90, 126.31, 124.81, 124.22, 39.29.

HRMS (ESI-TOF): calcd for C₉H₁₀NS₂ [M+H] 195.0176; found 195.0180.



¹**H** NMR (400 MHz, DMSO-*d*₆) δ 9.28 (dd, *J* = 5.1, 1.5 Hz, 1H), 9.09 – 9.04 (m, 1H), 8.94 (s, 3H), 8.48 – 8.41 (m, 2H), 8.28 (dd, *J* = 8.8, 1.9 Hz, 1H), 8.04 (dd, *J* = 8.4, 5.1 Hz, 1H), 4.31 (q, *J* = 5.8 Hz, 2H).

¹³C NMR (101 MHz, DMSO-*d*₆) δ 146.86, 144.90, 139.50, 135.83, 134.80, 129.50, 128.48, 123.11, 122.86, 42.17.



¹**H NMR (300 MHz, DMSO-***d*₆) δ 11.50 (s, 1H), 8.66 (s, 3H), 7.51 – 7.35 (m, 2H), 7.27 – 7.02 (m, 2H), 6.68 (ddd, *J* = 3.1, 1.9, 1.0 Hz, 1H), 4.24 (q, *J* = 5.8 Hz, 2H).

¹³C NMR (**75** MHz, DMSO-*d*₆) δ 136.29, 127.05, 126.17, 125.43, 121.24, 119.38, 112.29, 99.75, 40.44.



¹**H NMR (300 MHz, DMSO-***d*₆) δ 8.55 (s, 3H), 7.61 (q, *J* = 8.5 Hz, 4H), 7.41 (d, *J* = 2.1 Hz, 2H), 6.28 (t, *J* = 2.0 Hz, 2H), 4.02 (s, 2H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 140.28, 131.31, 130.94, 119.49, 119.40, 111.12, 42.02. HRMS (ESI-TOF): calcd for C₁₁H₁₃ClN₂ [M+H] 208.0767; found 208.0771.

¹H NMR (300 MHz, DMSO-*d*₆) δ 8.65 (s, 3H), 7.18 (d, *J* = 2.2 Hz, 1H), 7.07 – 6.82 (m, 2H), 6.03 (q, *J* = 2.1 Hz, 2H), 3.90 (q, *J* = 5.7 Hz, 2H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 147.68, 128.16, 123.39, 110.04, 108.63, 101.65, 42.38.

¹**H NMR (300 MHz, DMSO-***d*₆) δ 8.58 (s, 3H), 7.08 (d, *J* = 2.0 Hz, 1H), 6.96 (dd, *J* = 8.3, 2.1 Hz, 1H), 6.85 (d, *J* = 8.3 Hz, 1H), 4.23 (s, 4H), 3.86 (q, *J* = 5.7 Hz, 2H).

¹³C NMR (**75 MHz, DMSO-***d*₆) δ 143.93, 143.59, 127.45, 122.56, 118.43, 117.45, 64.58, 64.54, 42.03.



¹**H NMR (300 MHz, DMSO-***d*₆) δ 8.80 (s, 3H), 8.05 (dd, *J* = 2.2, 0.9 Hz, 1H), 7.70 (ddd, *J* = 8.4, 2.3, 0.8 Hz, 1H), 7.47 – 7.33 (m, 1H), 4.16 (q, *J* = 5.8 Hz, 2H), 3.91 – 3.69 (m, 4H), 2.99 – 2.78 (m, 4H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 155.04, 130.47, 127.11, 127.06, 127.01, 126.96, 126.76, 126.71, 126.66, 126.61, 126.54, 124.84, 124.42, 123.99, 122.94, 121.38, 66.76, 52.83, 48.96, 37.77.
HRMS (ESI-TOF): calcd for C₁₂H₁₆ClF₃N₂O [M+H] 296.7180; found 296.7183.



¹**H NMR (300 MHz, DMSO-***d*₆) δ 8.65 (s, 3H), 7.54 – 7.38 (m, 2H), 7.09 – 6.94 (m, 2H), 4.49 (t, J = 5.0 Hz, 2H), 4.00 – 3.86 (m, 6H), 3.60 – 3.43 (m, 4H), 3.23 (d, J = 11.2 Hz, 2H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 157.99, 131.13, 127.39, 115.16, 63.56, 62.94, 55.12, 52.08, 41.98. HRMS (ESI-TOF): calcd for C₁₃H₂₀N₂O₂ [M+H] 236.3151; found 236.3155.

¹**H NMR (300 MHz, DMSO-***d*₆) δ 8.67 (s, 3H), 7.54 (d, *J* = 1.6 Hz, 1H), 7.45 – 7.34 (m, 3H), 7.23 – 7.14 (m, 2H), 7.08 (dt, *J* = 7.8, 0.8 Hz, 1H), 6.19 (t, *J* = 2.3 Hz, 1H), 5.51 – 4.96 (m, 2H), 4.33 – 3.88 (m, 2H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 163.79, 160.56, 142.56, 139.51, 139.29, 139.24, 134.30, 129.06, 128.98, 128.87, 122.62, 122.47, 115.88, 115.60, 84.48, 72.85, 42.38.

HRMS (ESI-TOF): calcd for C₁₅H₁₅ClFNO [M+H] 279.7390; found 279.7411.



CI⁺H₃N

¹**H NMR (300 MHz, DMSO-***d*₆) δ 10.73 (s, 1H), 8.67 (bs, 3H), 7.64 – 7.52 (m, 3H), 7.50 – 7.42 (m, 2H), 7.20 – 7.06 (m, 2H), 5.14 (q, *J* = 13.0 Hz, 2H), 3.97 (q, *J* = 5.8 Hz, 2H), 3.00 (q, *J* = 7.1 Hz, 2H), 2.63 – 2.57 (m, 6H), 2.24 (t, *J* = 8.1 Hz, 2H), 1.49 (ddt, *J* = 21.8, 14.0, 7.1 Hz, 2H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 161.82 (d, *J* = 341.1 Hz), 143.99, 141.11, 138.79, 133.91, 128.73, 126.87 (d, *J* = 7.7 Hz), 122.09 (d, *J* = 8.0 Hz), 115.04 (d, *J* = 20.7 Hz), 90.04, 71.38, 56.34, 41.93, 41.83, 37.37, 19.10. White solid.

HRMS (ESI-TOF): calcd for C₂₀H₂₅FN₂O [M+H] 329.2001; found 329.2004.



¹**H NMR (300 MHz, DMSO-***d*₆) δ 8.74 (bs, 3H), 7.82 – 7.77 (m, 1H), 7.50 – 7.37 (m, 6H), 7.28 – 7.23 (m, 1H), 5.01 (s, 2H), 3.90 (q, *J* = 5.8 Hz, 2H), 2.95 – 2.88 (m, 2H), 2.23 – 1.78 (m, 8H), 1.66 – 1.54 (m, 2H), 1.39 – 1.21 (m, 2H), 0.82 (t, *J* = 7.3 Hz, 3H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 179.10, 172.81, 140.81, 139.32, 134.16, 131.43, 130.04, 129.73, 128.74, 128.35, 127.97, 127.24, 71.52, 43.60, 36.78, 27.00, 26.45, 25.23, 21.48, 13.44. White solid. HRMS (ESI-TOF): calcd for C₂₅H₃₁N₃O [M+H] 390.2561; found 390.2565.



¹H NMR (300 MHz, MeOD) δ 8.36 (s, 1H), 8.03 (s, 1H), 7.38 – 7.08 (m, 8H), 6.97 – 6.90 (m, 1H), 3.85 – 3.67 (m, 4H).

¹³C NMR (75 MHz, MeOD) δ 152.51, 141.15, 129.96, 129.50, 129.40, 128.91, 68.15, 53.19. Off white solid.

HRMS (ESI-TOF): calcd for $C_{17}H_{19}N_5$ [M+H] 294.1655; found 294.1601.



¹**H NMR (300 MHz, DMSO-***d*₆) δ 8.62 (s, 3H), 7.62 – 7.33 (m, 2H), 7.20 (d, *J* = 7.7 Hz, 2H), 3.94 (s, 2H), 2.31 (s, 3H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 138.12, 131.56, 129.49, 129.45, 42.31, 21.23.

NH₃⁺CI⁻

¹H NMR (300 MHz, DMSO-*d*₆) δ 8.78 (s, 3H), 7.74 – 7.57 (m, 6H), 7.53 – 7.31 (m, 3H), 4.06 (s, 2H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 140.60, 140.03, 133.79, 130.14, 129.46, 128.13, 127.20, 127.16,

NH₃⁺CI⁻



¹**H NMR (300 MHz, DMSO-***d*₆) δ 9.97 – 8.20 (m, 3H), 8.18 – 8.14 (m, 1H), 8.02 – 7.96 (m, 2H), 7.70 (dd, *J* = 7.1, 1.2 Hz, 1H), 7.63 – 7.55 (m, 3H), 4.50 (s, 2H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 133.66, 131.14, 130.96, 129.31, 129.09, 127.57, 127.15, 126.64, 125.83, 123.97, 39.75.



¹**H NMR (300 MHz, DMSO-***d*₆**)** δ = 9.01 – 8.51 (s, 3H), 7.84 – 7.51 (m, 2H), 7.40 – 7.07 (m, 2H), 4.14 – 3.84 (s, 2H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 162.50 (d, *J* = 244.5 Hz), 131.90 (d, *J* = 8.5 Hz), 130.88 (d, *J* = 3.0 Hz), 115.74 (d, *J* = 21.4 Hz), 41.79.



¹**H NMR (300 MHz, DMSO-***d*₆) δ 8.72 (s, 3H), 7.67 – 7.52 (m, 2H), 7.48 (dt, *J* = 8.6, 2.3 Hz, 2H), 4.01 (s, 2H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 133.62, 131.51, 131.49, 128.93, 41.83.



¹**H NMR (300 MHz, DMSO-***d*₆) δ 8.60 (s, 3H), 7.82 – 7.74 (m, 2H), 7.37 – 7.29 (m, 2H), 3.97 (s, 2H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 137.73, 134.32, 131.78, 129.43, 129.01, 95.31, 42.04.



¹**H NMR (300 MHz, DMSO-***d*₆) δ 8.67 (s, 3H), 7.66 (ddd, *J*=6.2, 4.0, 2.2, 1H), 7.59 – 7.48 (m, 1H), 7.48 – 7.26 (m, 2H), 4.12 (s, 2H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 132.41, 130.76, 129.32, 129.25, 128.35, 127.55, 43.65.

Br NH₃⁺Cl⁻

¹H NMR (300 MHz, DMSO- d_6) δ 8.73 (s, 3H), 7.64 – 7.55 (m, 2H), 7.55 – 7.46 (m, 2H), 3.99 (q, J = 5.6 Hz, 2H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 134.00, 131.83, 122.13, 41.88.

Br NH₃⁺Cl⁻

¹H NMR (300 MHz, DMSO-*d*₆) δ 8.71 (s, 3H), 7.94 – 7.20 (m, 4H), 4.02 (s, 2H). ¹³C NMR (75 MHz, DMSO-*d*₆) δ 137.26, 132.27, 131.63, 131.11, 128.65, 122.07, 41.84.



¹H NMR (300 MHz, DMSO-*d*₆) δ 8.81 (s, 3H), 7.77 (s, 4H), 4.12 (q, *J* = 5.8 Hz, 2H).
¹³C NMR (75 MHz, DMSO-*d*₆) δ 139.33, 130.28, 129.92, 129.49, 129.07, 128.65, 126.40, 125.84, 125.78, 125.73, 125.68, 122.80, 42.00.

¹**H NMR (300 MHz, CDCl₃)** δ 7.58 (dq, *J* = 2.0, 1.0 Hz, 1H), 7.51 – 7.38 (m, 3H), 3.91 (d, *J* = 1.9 Hz, 2H), 1.61 (s. 2H).

¹³C NMR (**75** MHz, Chloroform-*d*) δ 144.02, 130.98, 130.55, 130.49, 130.47, 128.89, 126.03, 123.82, 123.77, 123.61, 123.56, 122.42, 45.93.

¹**H NMR (400 MHz, DMSO-***d*₆) δ 8.72 (s, 3H), 7.67 (d, *J* = 7.7 Hz, 2H), 7.39 (d, *J* = 7.5 Hz, 2H), 4.04 (s, 2H).

¹³C NMR (101 MHz, DMSO-*d*₆) δ 148.69, 134.12, 131.70, 121.54, 120.49 (q, *J* = 256.4 Hz), 41.78.

HO, NH₃+CI-

¹**H NMR (300 MHz, DMSO-***d*₆) δ 8.57 (s, 3H), 7.48 – 7.26 (m, 4H), 4.48 (s, 2H), 3.95 (t, *J* = 5.7 Hz, 2H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 143.31, 132.73, 129.22, 126.93, 62.91, 42.39.



¹**H NMR (300 MHz, DMSO-***d*₆) δ 8.72 (s, 3H), 8.00 – 7.87 (m, 2H), 7.81 – 7.62 (m, 2H), 4.12 (s, 2H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 140.15, 132.86, 130.31, 119.06, 111.54, 42.11.



¹H NMR (300 MHz, DMSO-*d*₆) δ 8.63 (s, 3H), 7.87 – 7.82 (m, 2H), 7.69 (d, *J* = 8.0 Hz, 2H), 7.44 (s, 2H), 4.09 (d, *J* = 5.7 Hz, 2H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 144.54, 138.32, 129.90, 126.20, 42.07.

HRMS (ESI-TOF): calcd for C₇H₁₀N₂O₂S [M+H] 187.0541; found 187.0543.



¹H NMR (300 MHz, DMSO-*d*₆) δ 8.68 (s, 3H), 8.12 – 8.04 (m, 1H), 7.93 – 7.89 (m, 2H), 7.62 – 7.56 (m, 2H), 7.43 (s, 1H), 4.07 (d, *J* = 5.5 Hz, 2H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 167.87, 137.66, 134.58, 129.18, 128.11, 42.20.

¹H NMR (300 MHz, DMSO- d_6) δ 8.82 (s, 3H), 8.00 – 7.94 (m, 2H), 7.74 – 7.64 (m, 2H), 4.10 (q, J = 5.6 Hz, 2H), 3.85 (s, 3H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 166.37, 139.93, 129.87, 129.72, 129.66, 52.71, 42.13.



¹H NMR (300 MHz, DMSO-*d*₆) δ 8.83 – 8.50 (s, 3H), 7.67 – 7.60 (d, *J* = 7.5 Hz, 2H), 7.51 – 7.45 (d, *J* = 7.6 Hz, 2H), 4.01 – 3.93 (s, 2H), 1.30 – 1.20 (s, 12H). ¹³C NMR (75 MHz, DMSO-*d*₆) δ 137.78, 134.92, 129.44, 128.79, 84.18, 42.49, 25.12.

NH3⁺CI⁻

¹**H NMR (300 MHz, DMSO-***d*₆) δ 8.65 (s, 3H), 7.67 – 7.26 (m, 2H), 7.10 – 6.71 (m, 2H), 4.11 – 3.81 (m, 2H), 3.74 (s, 3H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 159.74, 131.08, 126.47, 114.31, 55.67, 42.05.



¹**H NMR (300 MHz, CDCl₃)** δ 7.23 (td, *J* = 7.9, 1.3 Hz, 1H), 6.95 – 6.83 (m, 2H), 6.77 (dq, *J* = 8.3, 1.6 Hz, 1H), 3.82 – 3.79 (m, 2H), 3.78 (dd, *J* = 2.1, 1.0 Hz, 3H).

¹³C NMR (75 MHz, Chloroform-*d*) δ 159.85, 144.93, 129.56, 119.35, 112.63, 112.24, 55.17, 46.42.



¹H NMR (400 MHz, DMSO-*d*₆) δ 8.48 (s, 3H), 7.58 – 6.87 (m, 4H), 3.93 (s, 2H), 3.83 (s, 3H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 157.61, 130.71, 130.58, 122.29, 120.71, 111.34, 55.99, 37.86.



¹H NMR (300 MHz, DMSO- d_6) δ 8.61 (s, 3H), 7.93 (s, 1H), 7.84 (dd, J = 13.4, 8.6 Hz, 2H), 7.62 (d, J = 8.0 Hz, 1H), 7.35 (d, J = 2.5 Hz, 1H), 7.24 – 7.17 (m, 1H), 4.13 (d, J = 5.7 Hz, 2H), 3.88 (d, J = 1.2 Hz, 3H).

¹³C NMR (**75** MHz, DMSO-*d*₆) δ 158.12, 134.49, 129.75, 129.64, 128.45, 128.39, 127.54, 119.61, 106.35, 55.71, 42.76, 31.17.



¹**H NMR (300 MHz, DMSO-***d*₆) δ 8.59 (s, 3H), 7.48 – 7.30 (m, 7H), 7.06 – 7.00 (m, 2H), 5.13 (s, 2H), 3.99 – 3.84 (m, 2H).

¹³C NMR (**75** MHz, DMSO-*d*₆) δ 158.77, 137.44, 131.07, 128.92, 128.30, 128.10, 126.73, 115.25, 69.62, 42.04.



¹**H NMR (400 MHz, DMSO-***d*₆) δ 8.40 (s, 3H), 7.18 (dd, *J* = 8.2, 6.7 Hz, 1H), 7.09 (d, *J* = 7.5 Hz, 2H), 4.00 (q, *J* = 5.7 Hz, 2H), 2.41 (s, 6H).

¹³C NMR (101 MHz, DMSO-*d*₆) δ 138.53, 131.28, 129.25, 128.78, 36.72, 19.94.



¹H NMR (300 MHz, DMSO-*d*₆) δ 8.72 (s, 3H), 7.61 (dd, *J* = 6.3, 2.5 Hz, 1H), 7.52 – 7.36 (m, 2H), 4.17 – 3.93 (m, 2H).

¹³C NMR (**75** MHz, DMSO-*d*₆) δ 161.30, 158.03, 129.38, 129.34, 126.32, 126.27, 126.18, 126.06, 123.01, 122.79, 117.41, 117.10, 35.90, 35.85.

¹**H NMR (400 MHz, DMSO)** δ 8.75 (s, 3H), 7.88 (d, *J* = 2.0 Hz, 1H), 7.68 (d, *J* = 8.3 Hz, 1H), 7.54 (dd, *J* = 8.3, 2.1 Hz, 1H), 4.03 (q, *J* = 5.6 Hz, 2H).

¹³C NMR (101 MHz, DMSO-*d*₆) δ 135.70, 131.72, 131.49, 131.41, 131.07, 130.02, 41.33.



¹**H NMR (300 MHz, DMSO-***d*₆) δ 8.82 (s, 3H), 7.51 (ddq, *J* = 7.5, 1.9, 0.6 Hz, 1H), 7.41 – 7.28 (m, 2H), 4.11 (s, 2H), 2.36 (d, *J* = 0.7 Hz, 3H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 136.69, 133.41, 132.31, 131.65, 128.45, 127.26, 40.34, 20.59.

Br NH₃+CI-

¹H NMR (300 MHz, DMSO-*d*₆) δ 8.67 (s, 3H), 7.79 (d, *J* = 2.2 Hz, 1H), 7.59 – 7.03 (m, 2H), 3.98 (s, 2H), 2.34 (d, *J* = 4.2 Hz, 3H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 137.80, 134.40, 133.03, 131.51, 128.93, 124.44, 41.53, 22.58.



¹**H NMR (400 MHz, DMSO-***d*₆) δ 8.76 (s, 3H), 7.98 (d, *J* = 2.1 Hz, 1H), 7.84 (d, *J* = 8.5 Hz, 1H), 7.71 (ddd, *J* = 8.5, 2.1, 0.9 Hz, 1H), 4.19 (s, 2H).

¹³C NMR (101 MHz, DMSO-*d*₆) δ 138.16, 135.05, 131.03, 129.45, 128.71, 128.66, 126.52, 126.22, 38.73.



¹H NMR (300 MHz, DMSO-*d*₆) δ 8.91 (s, 3H), 8.36 (s, 2H), 8.11 (s, 1H), 4.26 (s, 2H).
¹³C NMR (75 MHz, DMSO-*d*₆) δ 137.97, 131.29, 130.86, 130.41, 129.98, 125.52, 122.42, 121.90, 41.51.



¹**H NMR (300 MHz, DMSO-***d*₆) δ 8.42 (s, 3H), 7.20 (d, *J* = 8.6 Hz, 1H), 6.84 (d, *J* = 8.6 Hz, 1H), 3.98 – 3.65 (m, 11H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 154.42, 151.84, 141.81, 125.21, 119.87, 108.14, 61.51, 60.87, 56.44, 37.25.



¹H NMR (300 MHz, DMSO-*d*₆) δ 8.52 (s, 3H), 7.43 (s, 2H), 6.36 (s, 2H), 4.06 – 3.73 (m, 2H).
 ¹³C NMR (75 MHz, DMSO-*d*₆) δ 141.63, 130.10, 129.59, 123.92, 122.94, 117.99, 41.29.



¹**H NMR (300 MHz, DMSO-***d*₆) δ 8.80 (s, 3H), 7.95 – 7.01 (m, 2H), 4.05 (s, 2H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 152.06, 152.01, 151.93, 151.88, 148.78, 148.73, 148.65, 148.60, 140.87, 140.66, 140.46, 137.56, 137.35, 137.15, 132.07, 132.00, 131.96, 131.89, 131.84, 131.78, 114.72, 114.64, 114.53, 114.44, 41.36.



¹H NMR (300 MHz, DMSO-*d*₆) δ 8.91 (s, 3H), 7.69 – 7.58 (m, 1H), 4.03 (t, *J* = 1.5 Hz, 2H).
¹³C NMR (75 MHz, DMSO-*d*₆) δ 157.91, 157.87, 157.75, 157.62, 154.63, 154.51, 154.45, 154.40, 154.33, 152.73, 152.59, 152.52, 152.30, 151.78, 151.64, 151.56, 149.42, 149.34, 149.20, 149.13, 149.07, 148.99, 148.45, 148.38, 148.31, 148.24, 148.17, 148.09, 138.58, 138.45, 138.38, 138.24, 135.33, 135.19, 135.12, 134.98, 134.91, 108.96, 108.90, 108.73, 108.67, 108.61, 108.44, 108.39, 102.65, 102.60, 102.36, 102.31, 102.27, 102.22, 101.98, 101.93, 30.09.
HRMS (ESI-TOF): calcd for C₇H₅NF₄ [M+H] 180.0436; found 180.0441.

MH₃⁺CI

¹H NMR (400 MHz, DMSO-d₆) δ 8.37 – 7.57 (m, 3H), 2.73 (d, J=8.1 Hz, 2H), 1.53 (pent, J=6.2 Hz, 2H), 1.27 (t, J=9.4 Hz, 6H), 0.90 – 0.83 (m, 3H).
¹³C NMR (101 MHz, DMSO-d₆) δ 39.19, 31.20, 27.36, 25.99, 22.37, 14.33.

NH3⁺CI⁻

¹**H NMR (300 MHz, DMSO-***d*₆) δ 5.66 (s, 3H), 2.62 (t, *J* = 7.3 Hz, 2H), 1.45 (q, *J* = 7.1 Hz, 2H), 1.25 (s, 14H), 0.89 – 0.82 (m, 3H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 40.44, 31.80, 30.25, 29.48, 29.28, 29.22, 26.67, 22.58, 14.36.

NH3⁺CI⁻

¹H NMR (300 MHz, DMSO- d_6) δ 8.11 (s, 3H), 2.87 – 2.60 (m, 2H), 1.55 (pent, J = 7.2 Hz, 2H), 1.24 (s, 18H), 0.90 – 0.82 (m, 3H).

¹³C NMR (**75** MHz, DMSO-*d*₆) δ 39.14, 31.78, 29.53, 29.50, 29.43, 29.34, 29.20, 29.04, 27.38, 26.36, 22.58, 14.42.

HRMS (ESI-TOF): calcd for C₁₂H₂₇N [M+H] 186.2222; found 186.2226.

NH₃⁺CI⁻

¹**H NMR (300 MHz, DMSO-***d*₆) δ 8.05 (s, 3H), 2.69 (s, 2H), 1.51 (s, 2H), 1.21 (s, 22H), 0.86 – 0.80 (m, 3H).

¹³C NMR (**75** MHz, DMSO-*d*₆)δ 39.15, 31.74, 29.49, 29.40, 29.30, 29.16, 29.01, 27.37, 26.33, 22.54, 14.41.

HRMS (ESI-TOF): calcd for C₁₄H₃₁N [M+H] 214.2534; found 214.2537.



¹**H NMR (300 MHz, CDCl₃)** δ 2.70 (t, *J* = 7.0 Hz, 2H), 1.50 – 1.42 (m, 2H), 1.26 (d, *J* = 7.0 Hz, 30H), 0.87 (t, *J* = 6.5 Hz, 3H).

¹³C NMR (75 MHz, CDCl₃) δ 42.06, 31.94, 29.71, 29.68, 29.64, 29.48, 29.38, 26.88, 22.71, 14.14. HRMS (ESI-TOF): calcd for C₁₇H₃₇N [M+H] 256.3004; found 256.3004.



¹H NMR (300 MHz, CDCl₃) δ 2.61 (t, J = 6.9 Hz, 2H), 1.36 (pent, J = 6.6 Hz, 4H), 1.19 (s, 30H), 0.84 – 0.78 (m, 3H).

¹³C NMR (75 MHz, CDCl₃) δ 42.19, 33.79, 31.94, 29.71, 29.67, 29.53, 29.38, 26.92, 22.71, 14.13. HRMS (ESI-TOF): calcd for C₁₈H₃₉N [M+H] 270.3161; found 270.3162.

¹**H NMR (300 MHz, DMSO-***d*₆**)** δ 8.07 (s, 6H), 2.73 (q, *J* = 6.3 Hz, 4H), 1.55 (q, *J* = 7.3 Hz, 4H), 1.30 (d, *J* = 13.3 Hz, 8H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 39.12, 28.71, 27.31, 26.16.

-CI⁺H₃N

¹**H NMR (300 MHz, DMSO-***d*₆) δ 8.04 (s, 6H), 2.73 (q, *J* = 6.7 Hz, 4H), 1.54 (q, *J* = 7.4 Hz, 4H), 1.27 (s, 12H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 39.44, 29.13, 28.95, 27.37, 26.29.

¹**H NMR (400 MHz, DMSO-***d*₆) δ 8.01 (s, 3H), 3.04 – 2.54 (m, 4H), 1.24 (s, 20H), 0.86 (d, *J* = 6.3 Hz, 3H).

¹³C NMR (101 MHz, DMSO-*d*₆) δ 39.16, 31.76, 29.48, 29.40, 29.31, 29.18, 29.01, 27.39, 26.49, 26.32, 22.56, 14.43.



¹H NMR (400 MHz, DMSO- d_6) δ 8.06 (s, 3H), 2.69 (s, 2H), 1.51 (q, J = 7.6 Hz, 2H), 1.33 (pent, J = 8.1 Hz, 2H), 1.16 (s, 12H), 0.66 (t, J = 7.5 Hz, 2H).

¹³C NMR (101 MHz, DMSO-*d*₆) δ 83.15, 38.99, 29.88, 25.43, 25.13, 21.15.



¹H NMR (300 MHz, DMSO-*d*₆) δ 8.20 (s, 3H), 2.57 (d, *J* = 6.5 Hz, 2H), 1.79 – 1.48 (m, 6H), 1.15 (t, *J* = 10.9 Hz, 3H), 0.90 (t, *J* = 11.7 Hz, 2H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 44.80, 35.76, 30.30, 26.13, 25.54.



¹H NMR (300 MHz, DMSO-*d*₆) δ 5.33 (s, 3H), 2.31 (s, 2H), 1.93 (pent, *J* = 3.1 Hz, 3H), 1.69 – 1.56 (m, 6H), 1.47 (d, *J* = 3.0 Hz, 6H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 52.58, 39.75, 36.95, 33.05, 28.12.

¹**H NMR (300 MHz, DMSO-***d*₆) δ 8.26 (s, 3H), 7.06 (t, *J*=7.2, 1H), 6.90 (d, *J*=8.7, 3H), 2.93 – 2.71 (m, 4H), 2.13 (s, 3H).

¹³C NMR (**75** MHz, DMSO-*d*₆) δ 137.99, 137.76, 129.62, 128.87, 127.65, 126.02, 40.44, 33.22, 21.39.



¹**H NMR (300 MHz, DMSO-***d*₆) δ 8.24 (s, 3H), 7.51 – 7.04 (m, 5H), 3.04 (dd, *J* = 12.0, 7.0 Hz, 1H), 2.99 – 2.79 (m, 2H), 1.95 – 1.66 (m, 1H), 1.52 (ddq, *J* = 14.2, 9.0, 7.2 Hz, 1H), 0.67 (t, *J* = 7.3 Hz, 3H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 141.50, 129.07, 128.37, 127.38, 45.26, 44.17, 26.28, 11.87.

NH₃⁺Cl⁻

¹**H NMR (300 MHz, DMSO-***d*₆) δ 8.49 (s, 3H), 8.20 (d, *J* = 8.0 Hz, 1H), 7.95 – 7.87 (m, 1H), 7.80 (dd, *J* = 6.1, 3.3 Hz, 1H), 7.57 – 7.48 (m, 2H), 7.41 (q, *J* = 3.0 Hz, 2H), 3.44 (dd, *J* = 10.1, 5.9 Hz, 2H), 3.06 (dt, *J* = 11.0, 5.6 Hz, 2H).

¹³C NMR (**75** MHz, DMSO-*d*₆) δ 134.06, 133.93, 131.75, 129.16, 127.85, 127.35, 126.84, 126.29, 126.14, 124.03, 30.60.

NH₃⁺CI[−]

¹**H NMR (300 MHz, DMSO-***d*₆) δ 8.29 (s, 3H), 7.22 (dd, *J* = 8.2, 5.1 Hz, 2H), 7.05 (t, *J* = 8.4 Hz, 2H), 3.00 – 2.75 (m, 4H).

¹³C NMR (**75** MHz, DMSO-*d*₆) δ 163.09, 159.88, 134.07, 134.04, 131.04, 130.93, 115.84, 115.56, 40.39, 32.42.



¹**H NMR (300 MHz, DMSO-***d*₆) δ 8.26 (s, 3H), 7.09 (d, *J* = 8.1 Hz, 2H), 6.79 (d, *J* = 8.1 Hz, 2H), 3.63 (s, 3H), 2.92 – 2.74 (m, 4H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 158.42, 130.07, 129.71, 114.41, 55.52, 40.63, 32.46.



¹H NMR (300 MHz, DMSO-d₆) δ 9.68 – 9.16 (s, 1H), 8.48 – 7.83 (s, 3H), 7.07 – 6.99 (m, 2H), 6.77 – 6.70 (m, 2H), 2.98 – 2.86 (ddt, J=14.2, 9.1, 5.0 Hz, 2H), 2.83 – 2.74 (dd, J=9.3, 5.8 Hz, 2H).
¹³C NMR (75 MHz, DMSO-d₆) δ 156.66, 129.98, 127.76, 115.85, 40.71, 32.61.



¹**H NMR (300 MHz, DMSO-***d*₆) δ 7.89 (s, 3H), 7.38 – 7.13 (m, 5H), 2.96 (s, 2H), 0.97 – 0.90 (m, 2H), 0.80 (s, 2H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 141.52, 129.00, 128.97, 127.26, 47.59, 23.94, 13.55.



¹**H NMR (400 MHz, DMSO-***d*₆) δ 8.05 (s, 3H), 7.33 (d, *J* = 7.1 Hz, 2H), 7.22 (d, *J* = 7.2 Hz, 3H), 3.39 (s, 2H), 2.31 (dq, *J* = 47.0, 9.5 Hz, 4H), 1.89 (dq, *J* = 114.7, 9.0 Hz, 2H).

¹³C NMR (101 MHz, DMSO-*d*₆) δ 146.12, 128.92, 126.84, 126.53, 48.34, 45.07, 30.99, 15.57.

¹**H NMR (300 MHz, DMSO-***d*₆) δ 8.19 (s, 3H), 7.40 – 7.21 (m, 10H), 4.46 (t, *J* = 7.8 Hz, 1H), 3.52 (d, *J* = 7.9 Hz, 2H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 141.70, 129.21, 128.34, 127.41, 49.03, 42.94.

¹**H NMR (300 MHz, DMSO-***d*₆) δ 8.10 (s, 3H), 7.50 – 6.85 (m, 5H), 3.05 (dd, *J* = 12.8, 9.5 Hz, 1H), 1.86 – 1.47 (m, 5H), 1.45 – 1.30 (m, 1H), 1.30 – 0.53 (m, 5H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 139.97, 129.22, 128.77, 127.31, 49.25, 41.35, 40.31, 31.26, 29.72, 26.04.

¹**H NMR (300 MHz, DMSO-***d*₆) δ 8.20 (s, 3H), 6.80 – 6.73 (m, 2H), 6.65 (d, *J* = 7.9 Hz, 1H), 3.62 (d, *J* = 10.3 Hz, 6H), 2.88 (s, 2H), 2.76 (t, *J* = 7.4 Hz, 2H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 149.13, 147.95, 130.20, 120.93, 112.92, 112.39, 56.01, 55.93, 40.57, 32.89.

¹H NMR (300 MHz, DMSO-*d*₆) δ 9.10 – 8.66 (d, *J*=22.4 Hz, 2H), 8.47 – 7.80 (s, 3H), 6.74 – 6.61 (m, 2H), 6.50 – 6.44 (m, 1H), 2.97 – 2.83 (t, *J*=7.9 Hz, 2H), 2.78 – 2.64 (dd, *J*=9.6, 6.2 Hz, 2H). ¹³C NMR (75 MHz, DMSO-*d*₆) δ 32.80, 40.78, 116.28, 116.54, 119.67, 128.45, 144.52, 145.76.

¹**H NMR (400 MHz, DMSO-***d*₆) δ 8.30 (s, 3H), 8.04 (d, *J* = 1.7 Hz, 2H), 7.96 (dt, *J* = 1.9, 1.0 Hz, 1H), 3.15 (s, 4H).

¹³C NMR (101 MHz, DMSO-*d*₆) δ 141.49, 131.22, 130.90, 130.57, 130.49, 130.45, 130.25, 127.93, 125.22, 122.51, 121.02, 120.98, 120.94, 119.80, 39.60, 32.65.

¹H NMR (400 MHz, DMSO-*d*₆) δ 8.47 (ddd, *J*=4.9, 1.9, 1.0 Hz, 1H), 7.68 (td, *J*=7.6, 1.9 Hz, 1H), 7.29 – 7.07 (m, 2H), 2.92 – 2.85 (m, 2H), 2.79 (td, *J*=6.8, 1.2 Hz, 2H), 1.45 (s, 2H).
¹³C NMR (101 MHz, DMSO-*d*₆) δ 160.74, 149.42, 136.73, 123.62, 121.63, 42.39, 42.37.

¹H NMR (300 MHz, DMSO-*d*₆) δ 8.27 (s, 3H), 7.32 – 7.24 (m, 2H), 7.23 – 7.16 (m, 3H), 2.79 – 2.69 (m, 2H), 2.68 – 2.60 (m, 2H), 1.94 – 1.83 (m, 2H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 141.38, 128.85, 128.74, 126.45, 38.72, 32.36, 29.13.

¹H NMR (300 MHz, DMSO-d₆) δ 8.23 (s, 3H), 7.17 (t, J = 7.6 Hz, 2H), 6.88 (d, J = 7.9 Hz, 2H), 6.77 (t, J = 7.3 Hz, 1H), 3.15 (t, J = 7.0 Hz, 2H), 2.86 (s, 2H), 1.90 (pent, J = 7.2 Hz, 2H).
¹³C NMR (75 MHz, DMSO-d₆) δ 145.21, 129.62, 120.19, 115.97, 42.77, 37.10, 25.99.

¹H NMR (300 MHz, **DMSO-***d*₆) δ 7.66 (s, 3H), 7.13 (t, *J* = 7.7 Hz, 2H), 6.82 – 6.44 (m, 3H), 3.32 (t, *J* = 7.2 Hz, 4H), 2.80 (d, *J* = 7.5 Hz, 2H), 1.83 (pent, *J* = 7.3 Hz, 2H), 1.04 (t, *J* = 6.9 Hz, 3H). ¹³C NMR (75 MHz, **DMSO-***d*₆) δ 147.89, 129.57, 115.69, 112.28, 47.17, 44.56, 37.30, 26.00, 12.49.

¹**H NMR (300 MHz, DMSO-***d*₆) δ 10.89 (s, 1H), 8.21 (s, 3H), 7.53 – 7.47 (m, 2H), 7.40 (ddt, *J* = 7.9, 6.3, 1.1 Hz, 2H), 7.35 – 7.30 (m, 1H), 7.30 – 7.23 (m, 2H), 7.20 (d, *J* = 2.4 Hz, 1H), 6.82 (dd, *J* = 8.7, 2.4 Hz, 1H), 5.12 (s, 2H), 3.06 (s, 4H).

¹³C NMR (75 MHz, DMSO-*d*₆) δ 152.58, 138.26, 132.09, 128.81, 128.20, 128.09, 127.66, 124.53, 112.64, 112.32, 109.81, 102.27, 70.36, 39.16, 23.59.

HRMS (ESI-TOF): calcd for C₁₇H₁₈N₂O [M+H] 267.1497; found 267.1503.

¹**H NMR (300 MHz, CDCl₃)** δ 8.06 (s, 1H), 7.91 (s, 1H), 7.27 – 7.26 (m, 1H), 7.05 – 7.01 (m, 2H), 5.30 (s, 2H), 2.73 (s, 4H), 1.24 (s, 16H).

¹³C NMR (**75 MHz, CDCl**₃) δ 152.02, 148.14, 143.23, 134.49, 124.37, 123.66, 54.39, 54.02, 39.83, 26.41. Colorless oil.

HRMS (ESI-TOF): calcd for C₁₇H₂₇N₅ [M+H] 302.2271; found 302.2279.

S9. NMR spectra

270 260 250 240 230 220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 f1 (ppm)

200928.f330.10.fid Zhuang Ma ZM1-89 PROTON DMSO {C:\Bruker\TopSpin3.6.0} 2009 30

- 2.52 DMSO - 2.51 DMSO - 2.51 DMSO - 2.50 DMSO - 2.49 DMSO — 3.53 Н2О _ N___ H₂N, . NH₃⁺ CĪ











200914.339.10.fid Zhuang ma, Zm592 Au1H DMSO {C:\Bruker\TopSpin3.6.0} 2009 39

















200316.f337.10.fid Ma/ ZM 1-524 PROTON DMSO {C:\Bruker\TopSpin3.6.0} 2003 37

— 3.47 Н2О NH₃⁺ Cī













200303.f332.10.fid Ma/ ZM 1-509 PROTON DMSO {C:\Bruker\TopSpin3.6.0} 2003 32





210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 f1 (ppm)













200305.f351.10.fid Zhuang Ma ZM 1-460 PROTON DMSO {C:\Bruker\TopSpin3.6.0} 2003 51



[\]NH₃⁺Cī



270 260 250 240 230 220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 f1 (ppm)























270 260 250 240 230 220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 f1 (ppm)











270 260 250 240 230 220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 f1 (ppm)





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270	260	250	240	230	220	210	200	190	180	170	160	150	140	130	120	110	100	90	80	70	60	50	40	30	20	10	0	-10
f1 (ppm)																												







200511.f321.10.fid Zhuang Ma ZM-537 PROTON DMSO {C:\Bruker\TopSpin3.6.0} 2005 21




200421.f321.10.fid Zhuang Ma ZM-546 PROTON DMSO {C:\Bruker\TopSpin3.6.0} 2004 21

























270 260 250 240 230 220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 f1 (ppm)







270 260 250 240 230 220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 f1 (ppm)









210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 f1 (ppm)











270 260 250 240 230 220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 f1 (ppm)

















270 260 250 240 230 220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 fi (ppm)







210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 f1 (ppm)











270 260 250 240 230 220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 f1 (ppm)








270 260 250 240 230 220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 f1 (ppm)





270 260 250 240 230 220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 f1 (ppm)







210824.317.10.fid Zhuang Ma ZM 153 Au1H DMSO {C:\Bruker\TopSpin3.6.2} 2108 17

- 3.41 H2O _ NH₃⁺CĪ HQ, HO

DMSO DMSO DMSO DMSO

5125



114



270 260 250 240 230 220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 fl(ppm)



270 260 250 240 230 220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 f1 (ppm)





н | ∕ NH₃⁺ CĪ



-3 f1 (ppm) -1 -2

200306.f335.10.fid Zhuang Ma ZM 1-136 PROTON DMSO {C:\Bruker\TopSpin3.6.0} 2003 35

— 3.28 Н2О н₃с∕



119



