



Synthesis and anti-rheumatoid arthritis activities of 3-(4-aminophenyl)-coumarin derivatives

Yuhang Miao , Jie Yang , Yinling Yun , Jie Sun & Xiaojing Wang

To cite this article: Yuhang Miao , Jie Yang , Yinling Yun , Jie Sun & Xiaojing Wang (2021) Synthesis and anti-rheumatoid arthritis activities of 3-(4-aminophenyl)-coumarin derivatives, Journal of Enzyme Inhibition and Medicinal Chemistry, 36:1, 450-461, DOI: [10.1080/14756366.2021.1873978](https://doi.org/10.1080/14756366.2021.1873978)

To link to this article: <https://doi.org/10.1080/14756366.2021.1873978>



© 2021 The Author(s). Published by Informa UK Limited, trading as Taylor & Francis Group.



Published online: 08 Feb 2021.



Submit your article to this journal [↗](#)



Article views: 30



View related articles [↗](#)



View Crossmark data [↗](#)

RESEARCH PAPER



Synthesis and anti-rheumatoid arthritis activities of 3-(4-aminophenyl)-coumarin derivatives

Yuhang Miao, Jie Yang, Yinling Yun, Jie Sun  and Xiaojing Wang

Institute of Materia Medica, Shandong First Medical University & Shandong Academy of Medical Sciences, Jinan, China

ABSTRACT

Rheumatoid arthritis is a chronic systemic disease characterised by an unknown aetiology of inflammatory synovitis. A large number of studies have shown that synoviocytes show tumour-like dysplasia in the pathological process of RA, and the changes in the expression of related cytokines are closely related to the pathogenesis of RA. In this thesis, a series of novel 3-(4-aminophenyl) coumarins containing different substituents were synthesised to find new coumarin anti-inflammatory drugs for the treatment of rheumatoid arthritis. The results of preliminary activity screening showed that compound **5e** had the strongest inhibitory activity on the proliferation of fibroid synovial cells, and it also had inhibitory effect on RA-related cytokines IL-1, IL-6, and TNF- α . The preliminary mechanism study showed that compound **5e** could inhibit the activation of NF- κ B and MAPKs signal pathway. The anti-inflammatory activity of compound **5e** *in vivo* was further determined in the rat joint inflammation model.

ARTICLE HISTORY

Received 12 November 2020
Revised 9 December 2020
Accepted 5 January 2021

KEYWORDS

3-(4-Aminophenyl)-coumarin; rheumatoid arthritis; RA-FLSs proliferation inhibition; anti-inflammation

Introduction

Rheumatoid arthritis (RA) is the most common chronic inflammatory joint disease and a progressive autoimmune disease¹. The main pathological feature of RA is the recurrent chronic inflammation in the synovial tissue of the joint, which gradually infiltrates the cartilage and subchondral bone in the joint, and significantly affects the joints, organs, and systems, and eventually leads to irreversible joint deformities and dysfunction². At present, the research on the pathogenesis of RA is still a top priority³. During the pathological process of RA, synovial cells exhibit tumour-like abnormal proliferation, and invade into cartilage and bone, producing a variety of cytokines, leading to chronic inflammation of cartilage and bone⁴. There are two types of synovial cells: macrophage-like type A cells, with filamentous pseudopodia, invagination of serosa, vesicles, mitochondria, lysosomes, cytoplasmic fibres, and Golgi bodies, which have phagocytic function⁵; type B fibroblast-like synoviocytes (FLSs), which have a high concentration of endoplasmic reticulum structure. RA-FLS is an important effector cell in the course of RA. The over proliferation of RA-FLSs will lead to damage of joint and chronic persistent inflammation^{6–9}. Therefore, inhibiting the proliferation of RA-FLSs is an important therapeutic strategy in the clinical treatment of RA. There are four main types of drugs for clinical treatment of RA: non-steroidal anti-inflammatory drugs such as aspirin¹⁰, glucocorticoid drugs such as Prednisone¹¹, disease-altering drugs such as methotrexate (MTX)¹², and biological agents such as TNF- α inhibitors^{13,14}. Most of these RA treatment drugs have many side effects, and the biological agents are relatively expensive, leading to a huge burden on patients and delay of disease treatment. Therefore, it is imperative to develop new anti-RA drugs with high efficiency, low cost, and few side effects.

Coumarin (called Benzopyran one or o-hydroxycinnamic acid-8-lactone), which is composed of fused benzene and α -pyranone ring, is a kind of rich phenolic derivatives^{15,16}. Coumarins have a variety of biological activities including anti-tumour¹⁷, antibacterial¹⁸, antioxidant¹⁹, anti-Alzheimer's disease²⁰, and anti-inflammation²¹. There have been many reports on the anti-inflammatory activity of coumarin derivatives, and the relationship between structure and activity of (SAR) has also been explored, it was found that the introduction of the electron-rich hydrophobic group into the 3-position of coumarin nucleus could enhance the anti-inflammatory activity^{22,23}. The therapeutic effect of coumarin on RA has been reported before²⁴. In recent years, many coumarin derivatives obtained from natural products related to the treatment of RA have been found, such as Daphning, a coumarin derivative extracted from natural product *Daphne odora* Var. It has a good therapeutic effect on autoimmune arthritis²⁵. Synthetic coumarin derivatives are also widely used as in anti-RA. It has been found that scavenging reactive oxygen species, inhibiting the activation of NF- κ B and the synthesis of TNF- α can effectively treat RA^{26,27}; Multi-target drugs (such as NSAIDs, CORMs) and carbonic anhydrase inhibitors (CAI) can be used as hybrid compounds to treat RA through synergy, which is an important research direction for anti-RA drugs in the future^{28,29}. Carbonic anhydrase (CAs) isoforms are overexpressed in the synovium of RA patients and there is the interaction between various CAs subtypes and arthritis-like diseases, so carbonic anhydrase inhibitors have been paid more and more attention in the development of anti-RA drugs³⁰. Coumarin derivative is a kind of suicided CAIs, which is hydrolysed by esterase CA activity and binds to the active site of carbonic anhydrase, so coumarin has a unique and new carbonic anhydrase inhibition mechanism^{31,32}. 3-(4-aminophenyl)-coumarin derivatives are derived from 3-aryl coumarin, and

CONTACT Jie Sun  sunjie310@126.com; Xiaojing Wang  xiaojing6@gmail.com  Institute of Materia Medica, Shandong First Medical University & Shandong Academy of Medical Sciences, No. 6699, Qingdao Road, Huaiyin District, Jinan 250000, China

© 2021 The Author(s). Published by Informa UK Limited, trading as Taylor & Francis Group.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

the positional isomer structure of 3-aryl coumarin is similar to that of flavonoids and isoflavones. From the structure-activity relationship analysis, the electron-rich hydrophobic group introduced at position 3 can enhance the anti-inflammatory activity, and the hydroxyl group at position 7 or 8 can interact with the receptor through an intermolecular hydrogen bond. Joint inflammation can lead to elevated ROS, which can erode bones and joints and cause pain. The persistent pain behaviour in RA is related to the increase of inflammatory mediators in ROS. Flavonoids have the effect of anti-oxidation and relieving pain³³. Therefore, we speculate that 3-(4-aminophenyl)-coumarin derivatives may also have antioxidant imbalance activity. We tend to explore the anti-RA activity of 3-(4-aminophenyl)-coumarin derivatives through structural extension, which provides a new type of coumarin structure for the treatment of RA, but also a new direction for structural modification of coumarin compounds (Scheme 1).

Results and discussion

Synthesis of novel 3-(4-aminophenyl)-coumarin compounds containing various substituents

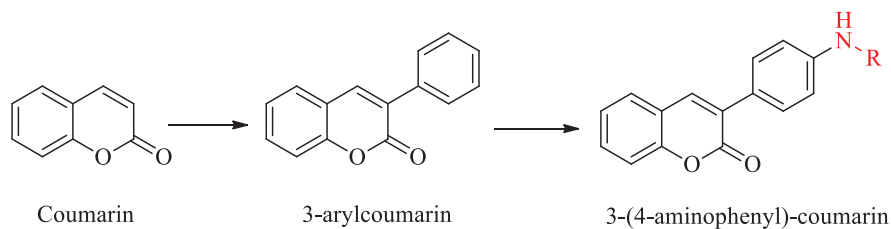
The 3-(4-aminophenyl)-coumarin derivatives were synthesised using the synthesis method previously reported by our group, and the synthesis route is shown in Scheme 2. The intermediate 2 was

synthesised from substituted p-amino-phenylacetic acid and o-hydroxybenzaldehyde by Perkin reaction and acidified with hydrochloric acid to give 3-(4-aminophenyl)-coumarin intermediate 3. In the next step, a series of substituted benzoyl chlorides were synthesised from benzoic acid. The target compound was obtained by amide condensation of 3 with substituted benzoyl chloride in the mixed solution of pyridine and acetone (Table 1, 4a-4q). The synthetic route of 3-(4-aminophenyl)-coumarin derivatives with heterocycles is shown in Scheme 3. Hydroxy benzotriazole (HOBt) and dicyclohexylcarbodiimide (DCC) were added to the anhydrous toluene solution containing compound A-E. Then the intermediate 3 was put into the reaction solution, and the reaction mixture was stirred at room temperature for 8-16 h to obtain 5a-5h³⁴.

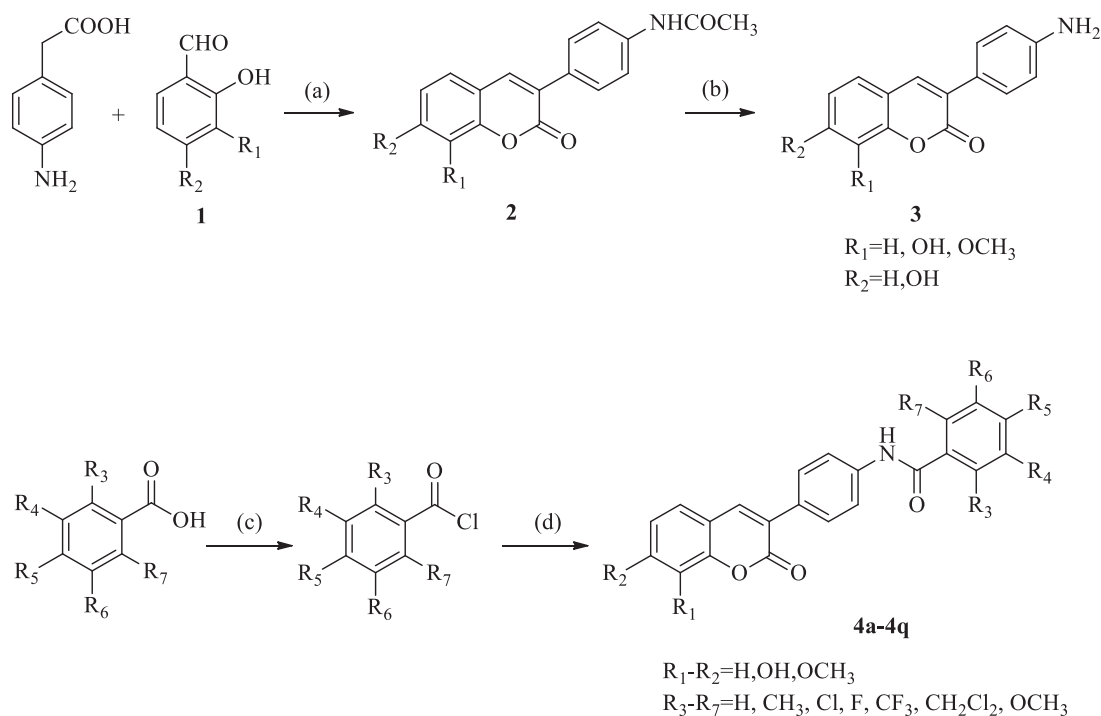
Biology

Effect of compounds on proliferation of RA-FLS

Rheumatoid arthritis fibroblast-like synovial cells (RA-FLSs) are the main effector cells of RA, which exhibit abnormal activation and proliferation in synovial lesions, and mediate joint destruction and synovitis during RA pathological process³⁵. Recent studies have found that RA-FLS cells activated in the chronic inflammatory environment have unique morphology and show characteristics similar to tumour cells, so the inhibition of RA-FLS cell proliferation is very important for the treatment of RA³⁶. We first tested



Scheme 1. Structural derivation of 3-(4-aminophenyl)-coumarins.



Scheme 2. The synthetic route of 3-(4-aminophenyl)-coumarins 3 and target product 4. Reagents and conditions: (a) acetic anhydride, Et₃N, 115 °C. (b) HCl ethanol. (c) COCl₂, DCM, reflux. (d) 3-(4-aminophenyl)-coumarin 3, acetone, pyridine, and RT.

RA-FLS proliferation inhibitory activities of 25 3-(4-aminophenyl) coumarin derivatives using MTT assay with MTX as a reference compound. As shown in Table 2, some of the coumarin

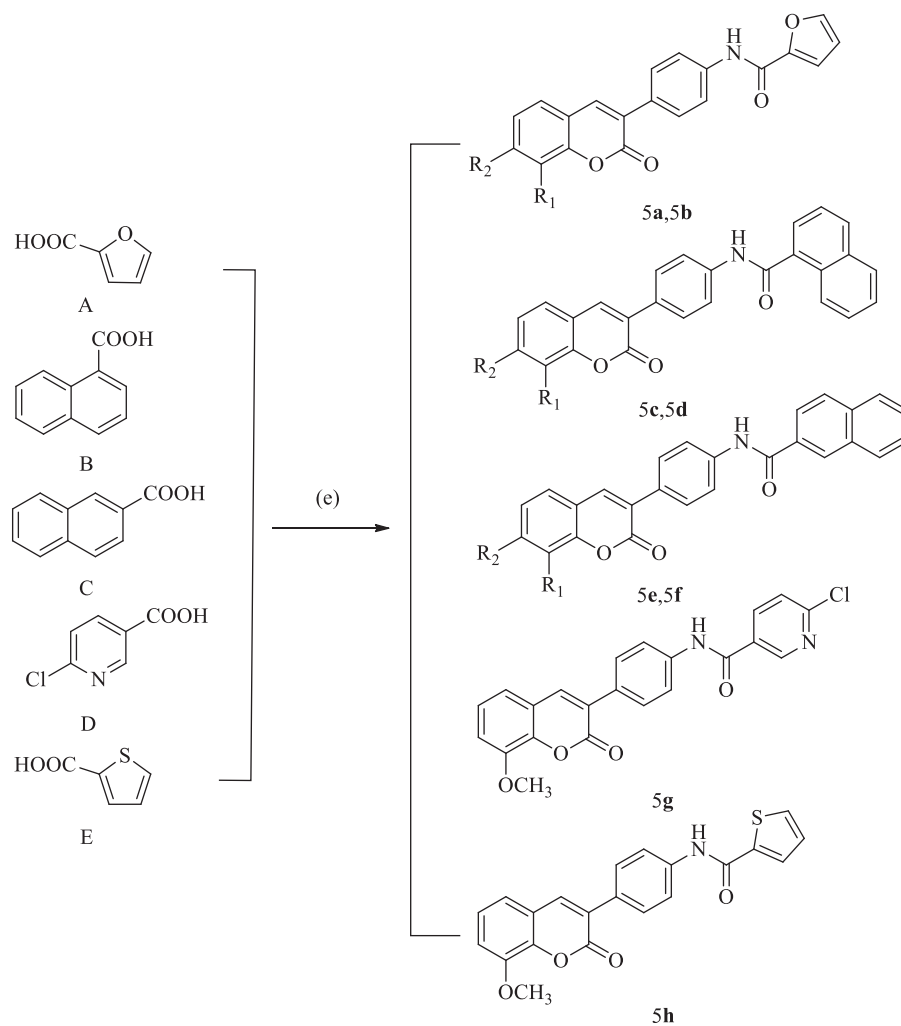
Table 1. Structures of compounds 4a–4q, 5a–5h.

Product	R ₁	R ₂	R ₃	R ₄	R ₅	R ₆	R ₇	Yield (%)
4a	H	OH	H	H	F	H	H	70
4b	H	OH	H	H	CH ₃	H	H	58
4c	H	OH	H	H	CH ₂ Cl	H	H	77
4d	H	OH	H	CH ₃	H	H	H	85
4e	H	OH	H	Cl	H	H	H	81
4f	H	OH	F	H	H	H	H	72
4g	H	OH	H	H	Cl	H	H	80
4h	H	OH	H	CH ₃	H	CH ₃	H	72
4i	OCH ₃	H	H	H	H	F	H	75
4j	OCH ₃	H	H	H	F	H	H	78
4k	OCH ₃	H	H	H	CF ₃	H	H	68
4l	OCH ₃	H	H	H	CH ₂ Cl	H	H	70
4m	OCH ₃	H	H	CH ₃	H	H	H	87
4n	OCH ₃	H	F	H	H	H	H	82
4o	OCH ₃	H	OCH ₃	H	H	H	H	77
4p	OCH ₃	H	Cl	Cl	H	H	H	62
4q	OH	OH	H	H	CH ₂ Cl	H	H	58
5a	H	OH						70
5b	OCH ₃	H						75
5c	H	OH						78
5d	OCH ₃	H						82
5e	H	OH						65
5f	OCH ₃	H						72
5g	OCH ₃	H						65
5h	OCH ₃	H						72

derivatives exhibited an inhibitory effect on RA-FLS proliferation. Among these compounds, heterocyclic substituted compound **5e**, with substitution of side-chain phenyl by naphthalene has the strongest activities ($IC_{50} = 1.78 \pm 0.02 \mu M$), which was even stronger than MTX ($IC_{50} = 5.00 \pm 0.04 \mu M$). When R₁ is hydrogen and R₂ is hydroxyl substitution, the activities of compound **4e** ($IC_{50} = 5.60 \pm 0.29 \mu M$) and **4g** ($IC_{50} = 9.13 \pm 0.02 \mu M$) are comparable to that of MTX. We, therefore, infer that R₄ or R₅ chlorination is significantly beneficial to the activity of RA-FLS proliferation inhibition. The activity of compound **4p** ($IC_{50} = 7.61 \pm 0.13 \mu M$) is better than **4l** ($IC_{50} = 15.71 \pm 0.11 \mu M$) and **4o** ($IC_{50} = 16.57 \pm 0.04 \mu M$), suggesting that the RA-FLS inhibitory activity of R₂ hydroxyl substituted compounds is better than that of R₁ methoxy substituted compounds (Figure 1).

Effect of compounds 4e, 4p, and 5e expression levels of RA-related cytokines

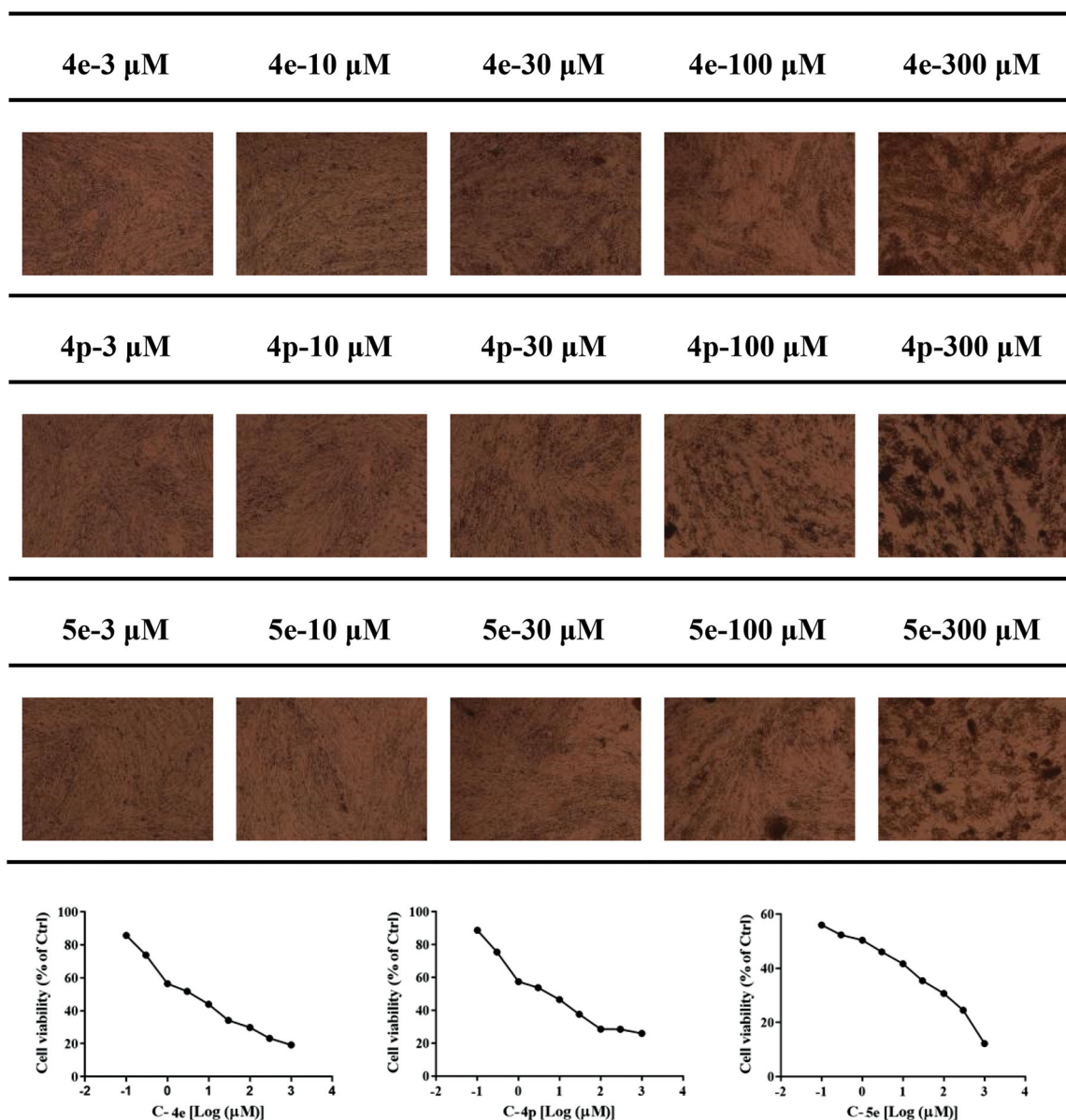
Cytokines are critical local protein mediators involved in almost all important biological processes, including cell growth, activation, and differentiation, inflammation, immunity³⁷. It is well known that the imbalance between the activities of pro-inflammatory and anti-inflammatory cytokines in rheumatoid joints is conducive to the induction of autoimmunity and chronic inflammation, resulting in a joint injury. Therefore, the expression of related cytokines is closely related to the pathogenesis of RA³⁸. Cytokines



Scheme 3. General synthetic route to compounds 5a–5h. Reagents and conditions: (e) 3-(4-aminophenyl)-coumarin 3, DCC, HOBT, toluene, and RT.

Table 2. Effect of compounds on proliferation of RA-FLSs. (mean \pm SD, $n = 3$)

Compound	IC ₅₀ value (μ M) RA-FLSs cell proliferation inhibitory activity	Compound	IC ₅₀ value (μ M) RA-FLSs cell proliferation inhibitory activity
4a	10.55 \pm 0.28	4n	>50
4b	13.93 \pm 0.02	4o	16.57 \pm 0.04
4c	>50	4p	7.61 \pm 0.13
4d	>50	4q	17.39 \pm 0.33
4e	5.60 \pm 0.29	5a	>50
4f	18.59 \pm 0.04	5b	>50
4g	9.13 \pm 0.02	5c	13.07 \pm 0.02
4h	>50	5d	19.83 \pm 0.03
4i	8.79 \pm 0.19	5e	1.78 \pm 0.02
4j	>50	5f	10.40 \pm 0.32
4k	>50	5g	24.72 \pm 0.03
4l	15.71 \pm 0.11	5h	11.74 \pm 0.29
4m	>50	Methotrexate	5.00 \pm 0.04

**Figure 1.** The cell proliferation inhibitory activity of compounds 4e, 4p, and 5e.

respond comprehensively to a variety of stimuli in the process of immunity and inflammation, involving both pro-inflammatory and anti-inflammatory activities. Cytokines such as IL-1 and tumour necrosis factor (TNF) have pro-inflammatory activity, while IL-10

mainly plays an anti-inflammatory role³⁹. IL-1 is a prototype proinflammatory cytokine and has two forms, IL-1 α and IL-1 β , with indistinguishable biological activities in most studies⁴⁰. IL-1 affects almost all cell types and usually acts synergistically with another

pro-inflammatory cytokine, TNF. Although IL-1 is well known to up-regulate host defence and act as an immune adjuvant, it is also a highly inflammatory cytokine⁴¹. Inhibition of IL-1 was first applied for the treatment of RA. IL-1 α is characterised by its immediate effect after release without further maturation. By binding to IL-1R1, IL-1 α can trigger inflammation through induction of IL-1 β activation and production of other cytokines, such as TNF or IL-6. IL-1 β is the most functional member of the IL-1 family and high levels of IL-1 β are found in synovium and fluid of RA patients⁴². As another important intercellular messenger, TNF- α has drawn more and more interest in the development of therapeutic methods for RA in recent years. It is generally believed that IL-1 plays a key role in joint destruction, while TNF- α is also involved in the systemic inflammatory process⁴³. Numerous studies on synovial tissue of RA have demonstrated that TNF- α is a potential therapeutic target for RA, so the discovery of novel TNF- α inhibitors is of great significance for treatment of RA^{44,45}.

Compounds **4e** ($IC_{50} = 5.60 \pm 0.29 \mu\text{M}$), **4p** ($IC_{50} = 7.61 \pm 0.13 \mu\text{M}$), and **5e** ($IC_{50} = 1.78 \pm 0.02 \mu\text{M}$) showed an excellent inhibitory effect on RA-FLS proliferation. We next used these optimised compounds to further explore their anti-RA activities and related mechanisms. We first detected the effect of these compounds on the contents of cytokine IL-1 α , IL-1 β , and TNF- α released by RA-FLSs with stimulation LPS. The results showed that the three compounds could concentration-dependently reduce

the secretion of IL-1 α , IL-1 β , and TNF- α . Compound **5e** exhibited stronger activity than Compounds **4e** and **4p** (Figure 2).

RA-FLS cells were cultured at 37°C for 24 h, then the cells were divided into four groups: blank control group, model group (LPS), LPS + compound group, and LPS + MTX group. Different concentrations of compounds and 1.0 $\mu\text{g}/\text{mL}$ MTX, were added to incubate for 48 h, and then LPS was added to stimulate the secretion of proinflammatory cytokines at the final concentration of 5 $\mu\text{g}/\text{mL}$. After 24 h of incubation, pro-inflammatory cytokines including IL-1 α , IL-1 β , and TNF- α were measured by ELISA method.

Multiple cytokine IL-10 is an effective inhibitor in immune cells including macrophages, T cells and NK cells, and it can block the synthesis and activation of cytokines in these cells. Different from pro-inflammatory factors IL-1 and TNF- α , IL-10 has effective anti-inflammatory properties^{46,47}. We examined the effects of **4e**, **4p**, and **5e** on cytokine IL-10. The results showed that all the derivatives could increase the amount of IL-10, which preliminarily indicated that the derivatives were beneficial to the fight against RA disease (Figure 3).

We also detected the effect of these compounds on the content of IL-10 released by RA-FLSs stimulated by LPS, and the results showed that **4e**, **4p**, and **5e** could significantly increase the content of IL-10 released by RA-FLSs. These results indicate that compounds **4e**, **4p**, and **5e** could inhibit the production of pro-inflammatory cytokines and enhance the production of anti-inflammatory cytokines, therefore exert anti-inflammatory functions.

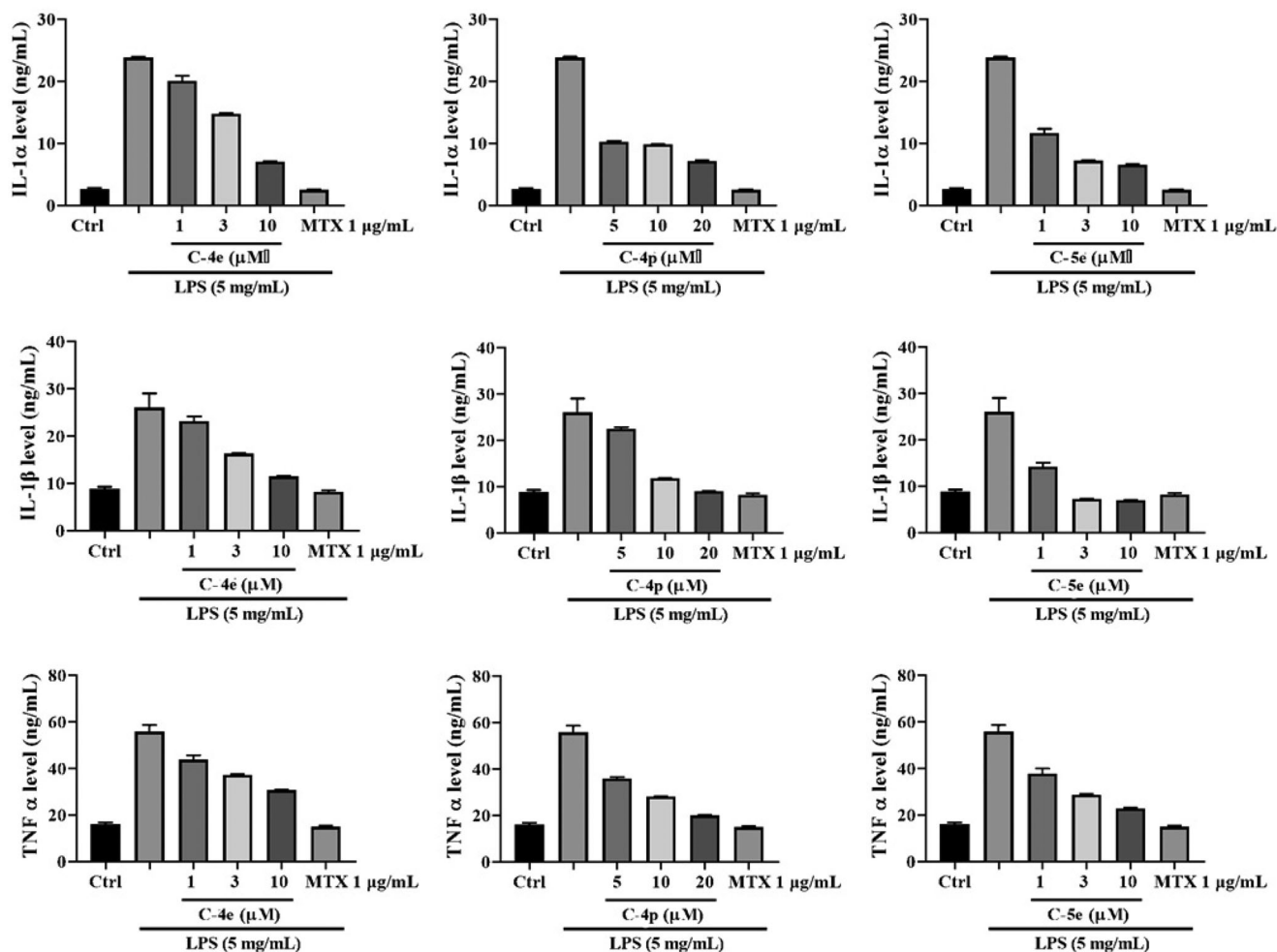


Figure 2. Effect of compounds **4e**, **4p**, and **5e** on secretion of IL-1 α , IL-1 β , and TNF- α .

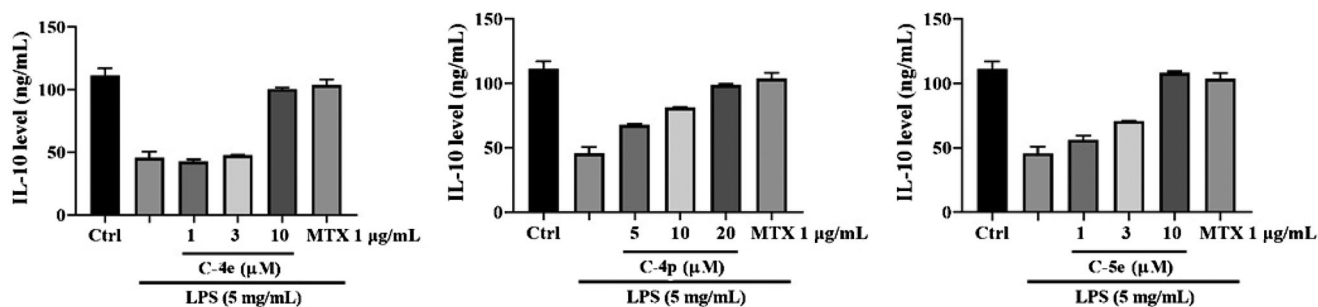


Figure 3. Effect of compounds 4e, 4p, and 5e on secretion of cytokine IL-10.

Compound **5e** has the strongest activity among them, which might be a potential anti-RA drug candidate⁴⁸.

Effect of compound 5e on luciferase activity of NF-κB

The activation of a series of signal pathways is a typical feature of chronic synovitis in RA. These signal pathways induce phosphorylation of cytoplasmic proteins in synovial cells, and phosphorylation of transcription factors and nuclear proteins such as NF-κB⁴⁹, thus promoting cell proliferation and activation. On the other hand, inflammatory factors such as TNF-α, IL-1α, and IL-1β in turn regulate cell membrane receptors and activate the mitogen-activated protein kinase (MAPK) signal pathway. Therefore, investigation of the above signalling events is of great significance for the in-depth understanding of the pathogenesis of RA and the development of new drugs for treatment. NF-κB-Luc luciferase reporter gene plasmid is a reporter gene used to detect the transcriptional activity of NF-κB. The enhanced activity of NF-κB stimulated by LPS leads to a significant increase in the expression of inflammatory proteins. We next treated RA-FLSs with compound **5e** and tested the luciferase activity of NF-κB. Similar to MTX, compound **5e** can significantly reduce the activity of NF-κB, which is of great significance in inhibiting inflammatory factors and reducing inflammation (Figure 4).

Effect of compound 5e on MAPK signal pathway

MAPK mediates important pathways in the eukaryotic signal transduction network, which plays a key role in regulating gene expression and promoting disease development^{50,51}. In mammals, five different MAPK-mediated signal pathways regulate cell growth and differentiation. JNK and p38 MAPK signalling pathways play an important role in stress responses such as inflammation and apoptosis⁵². We next treated RA-FLSs with compound **5e** and tested the changes in MAPK-mediated signal pathways. The results showed that LPS stimulation significantly increased the levels of phospho-extracellular signal-regulated kinases (ERK)1/2 and phospho-p38, and compound **5e** could significantly reduce the levels of phospho-ERK1/2 and phospho-p38. These results indicate that compound **5e** can effectively suppress the phosphorylation and activation of ERK1/2 and P38 in the MAPK signal pathway (Figure 5), which is well known to cause suppression of NF-κB activity. In contrast, MTX only suppressed the phosphorylation of ERK1/2 but not the p38.

Anti-RA activity of compound 5e in vivo

To establish an arthritis model, we used Freund's complete adjuvant to induce arthritis in rats via injection of CFA into the left hind paw of rats^{53,54}. After injection of CFA into SD rats, the

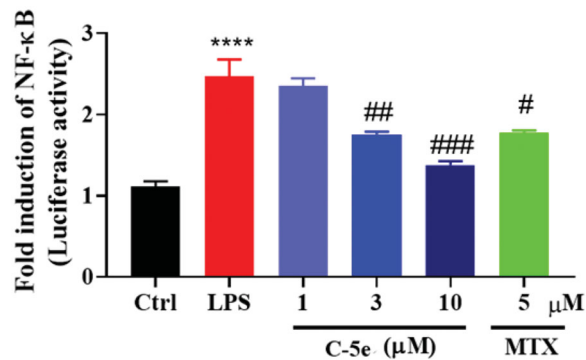


Figure 4. Inhibitory effect of compound **5e** on luciferase activity of NF-κB. RA-FLS cells were cultured at 37°C for 24 h, then NF-κB luciferase plasmid and control sea kidney plasmid were transfected for 48 h. Different concentrations of compound **5e** and LPS (final concentration: 5 μg/mL) were added to stimulate the secretion of proinflammatory cytokines. After incubation for 48 h, the cells were collected and the luciferase activity of NF-κB pathway was detected by dual luciferase system. # $p < 0.05$, ## $p < 0.01$, ### $p < 0.001$, compared with model group; **** $p < 0.0001$ compared with ctrl group.

results showed that 80–90% of the rats showed paw swelling within 14–28 days (Figure 6, model group). The model rats were treated with compound **5e** of different concentrations once a day from day 14 to day 28. MTX was used as a positive control. The experimental results are shown in Figure 6. After successfully establishing the model, we tested the pain threshold and motor function of RA rats. The reflex latency (paw withdraw thermal latency, PWTL) of rat foot contraction induced by thermal stimulation was used as an index of hindlimb pain threshold by the hot plate method. As shown in Figure 7, the PWTL of the model group was shorter than that of the control group ($p < 0.001$). After the intervention of compound **5e** in the low-dose group and high-dose group, the results showed that the PWTL of RA rats was prolonged, and the prolongation of PWTL was positively correlated with the drug concentration⁵⁵. It is proved that compound **5e** reduces thermal hyperalgesia in RA rats and has an analgesic effect on RA rats. In the motor function test, the left gait width and hind limb gait width of RA rats before and after treatment were measured by gait analysis. Compared with the control group, the left stride length of the model group was shorter, and the hindlimb stance width was wider, and the motor function of rats was blocked ($p < 0.001$). After treatment with compound **5e** in low-dose group and high-dose group, it was found that compound **5e** could improve the exercise ability of rats (Figure 8).

We then treated RA rats with compound **5e**. In normal rats, no inflammation pathological features were observed, the

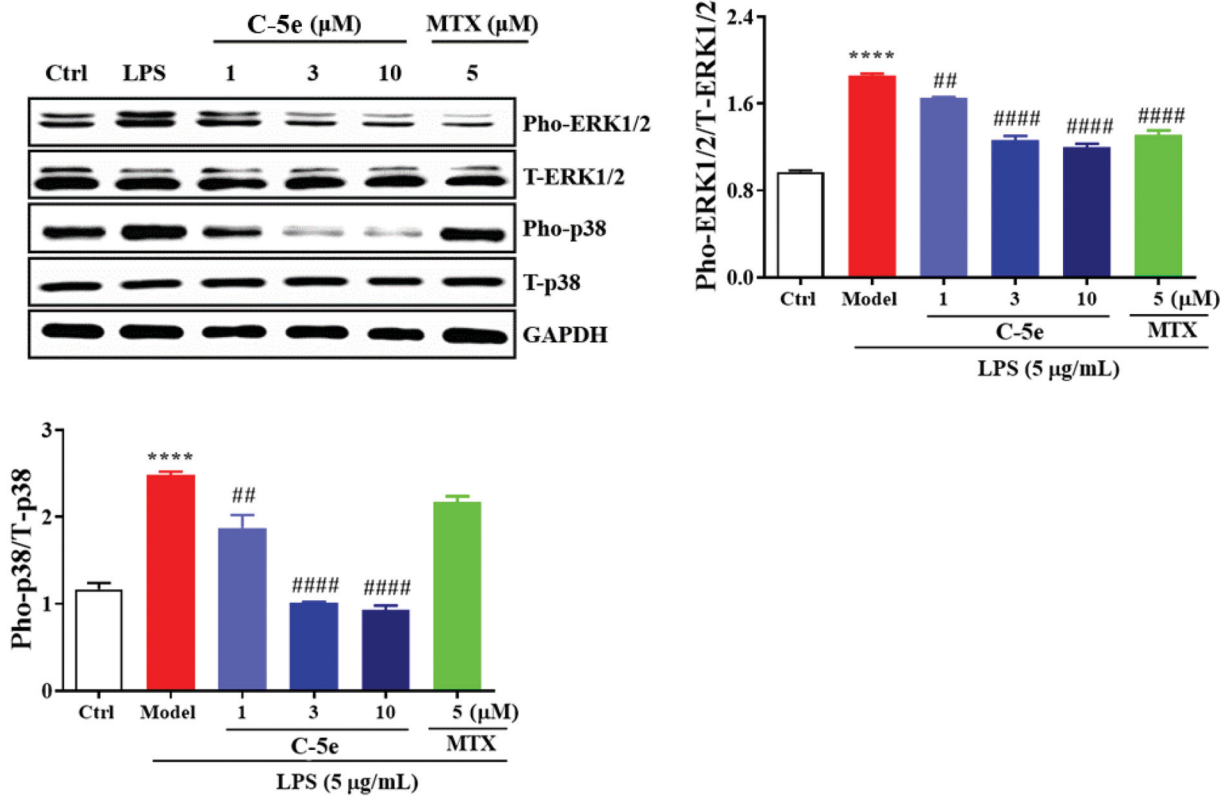


Figure 5. Inhibitory effect of compound 5e on MAPK signal pathway. RA-FLS cells were cultured at 37°C for 24 h. The cells were preincubated with compound 5e of different concentrations for 1 h, and then treated with LPS (final concentration: 5 μg/mL) for 72 h. Collected cells were used for Western blotting analysis of anti-ERK1/2, anti-pho-ERK1/2, anti-p38 and anti-pho-p38 antibodies. ## $p < 0.01$, #### $p < 0.0001$ compared with model group; **** $p < 0.0001$ compared with ctrl group.

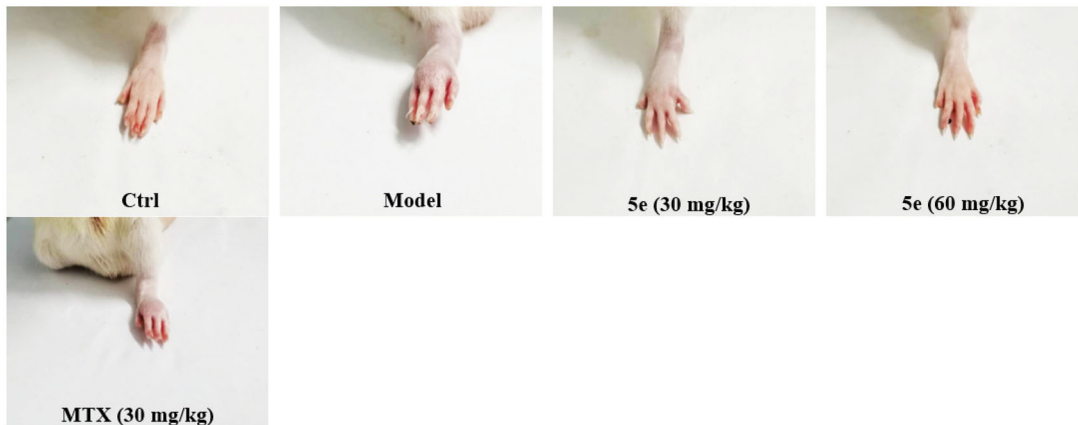


Figure 6. Establishment and treatment of CFA rat model of RA. Normal control group (Ctrl, saline), model group (Model, saline), high dose compound 5e (60 mg/kg) group, low dose compound 5e (30 mg/kg) group, positive control MTX (30 mg/kg) group. ($n = 5$ per group).

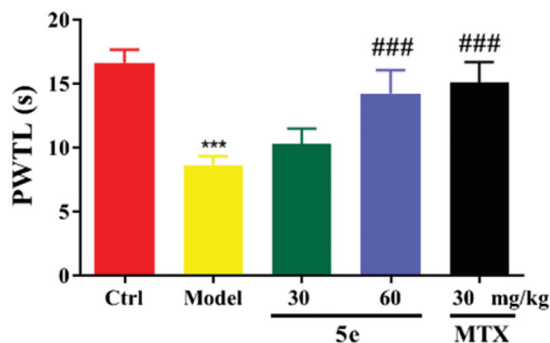


Figure 7. The analgesic effect of compound 5e on rats was detected by the method of thermal pain threshold of rat claws. ### $p < 0.001$, *** $p < 0.001$. ($n = 5$ per group).

surface of cartilage was smooth and free of cracks, and the cells were evenly distributed (Figure 9(A)). In the arthritis model group, obvious inflammation pathological features were observed, including disorder of chondrocytes and vacuolisation of some cells, which indicate decreased activity or died cells, and destroyed cartilage surface (Figure 9(B-1,B-2)). After treatment with 30 mg/kg of compound 5e, the inflammation was significantly improved, the vacuole of chondrocytes decreased, and the destruction of the cartilage surface was recovered (Figure 9(C)). After treatment with 60 mg/kg of compound 5e, almost no inflammatory cell infiltration was observed, the surface of cartilage tended to be smooth, indicating a better therapeutic effect than low dose compound 5e (Figure 9(D)).

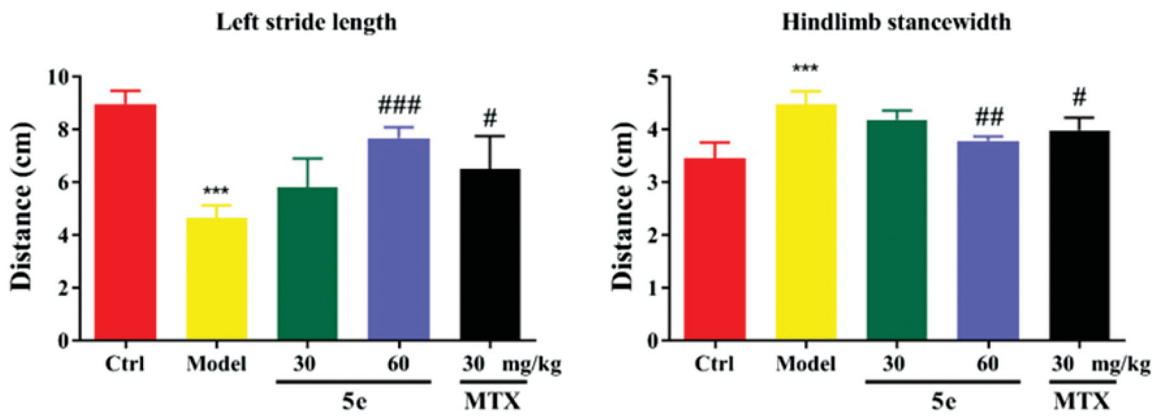


Figure 8. Motor function test of SD rats. Place the rat in the plexiglass chamber of the treadmill (25 cm long and 5 cm wide), set the treadmill speed to 18 cm/s, the shutter speed to take pictures: 40 ms, and record 5 s for each animal. The motor function of RA rats was explained by the motor parameters of left stride length and hindlimb stance width. # $p < 0.05$, ## $p < 0.01$, ### $p < 0.001$, compared with model group, *** $p < 0.001$ compared with ctrl group. ($n = 5$ per group).

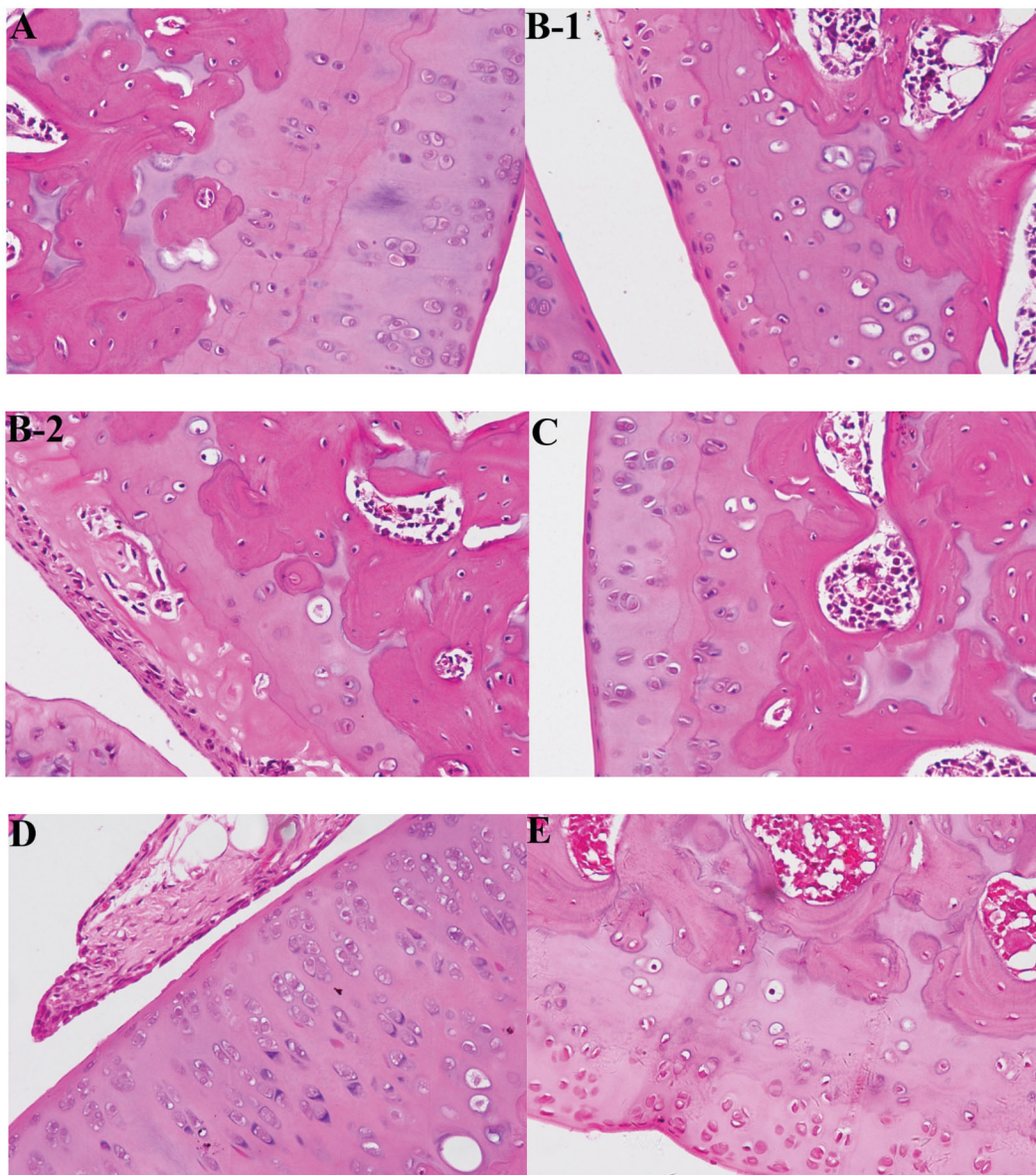


Figure 9. Compound 5e recovers RA histopathological features of rats. (A) Normal control group (Ctrl, normal saline), (B-1, B-2) model group (model, normal saline), (C) low-dose compound 5e group (30 mg/kg), (D) high-dose compound 5e group (60 mg/kg), and (E) positive control MTX (30 mg/kg). ($n = 5$ per group).

Conclusion

In this study, we screened the anti-RA activity of a series of 3-(4-aminophenyl)-coumarins synthesised by our team. Compound **5e** had the strongest inhibitory effect on the proliferation of RA-FLS₅. We further discussed the anti-inflammatory activity of compound **5e** and its related mechanism. We found that compound **5e** could inhibit the expression of inflammation-related cytokines (IL-1 α , IL-1 β , and TNF- α) induced by lipopolysaccharide. The preliminary mechanism study found that compound **5e** has an effect on inflammation-related signal transduction, which can inhibit the activity of nuclear factor-mitogen-activated protein B (NF- κ B) through mitogen-activated protein kinase (MAPK) signal pathway. Finally, the rat arthritis model established by Freund's complete adjuvant further proved that compound **5e** had anti-RA activity *in vivo*. To sum up, compound **5e** may be a potential protective compound for the treatment of RA.

Experimental

Materials and methods

Materials

Methotrexate, manufacturer: MCE. Bio Legend LEGEND plex TM multiplex bead-based assay (# 740007), brand: Biolegend; manufacturer: Biolegend Corporation of the United States. Saline, manufacturer: Guangdong Litai Pharmaceutical Co., Ltd. PBS powder, manufacturer: American SIGMA Company.

Cells and cell culture

RA-FLS cells were obtained from Beijing Luyuan Byrd Biotechnology Co., Ltd., and the 3rd–5th generations were used in the experiments. The cells were cultured with high glucose Dulbecco's Modified Eagle Medium (DMEM) supplemented with 10% foetal bovine serum (FBS), 100 units/mL penicillin, 100 μ g/mL streptomycin, and 2 mM glutamine. All cells were incubated in a humidified atmosphere of 95% air and 5% CO₂ at 37 °C.

MTT assay

RA-FLS cells were inoculated in a 96-well plate (5×10^{-3} /well/100 μ L), incubated for 24 h, and then treated with different concentrations of synthesised derivatives (0.1, 0.3, 1, 3, 10, 30, 100, and 300 μ M) or 1.0 μ g/mL MTX for 72 h. The culture medium, was then removed and the cells were incubated with 5 μ g/mL MTT at 37 °C for 4 h, and 100 μ L DMSO was added to each well to dissolve the formed methylazan. The absorbance (OD value) was measured at 570 nm using a plate reader. The cell proliferation inhibition rate was calculated by following the formula: inhibition rate = (OD value of the control group - OD value of the treated groups) / OD value of the control group \times 100%. The IC₅₀ value (μ M) was calculated according to the curve.

Test of cytokines using enzyme-linked immunosorbent assay (ELISA)

The 3rd–5th passage RA-FLS cells were inoculated in 48-well plates (1.5×10^6 /well) and cultured at 37 °C for 24 h, and then cells were divided into four groups: blank control group, model group (LPS), LPS + compound group and LPS + MTX group. Different concentrations of compounds and 1.0 μ g/mL MTX were added and incubated for 48 h, and then LPS was added at a final concentration of 5 μ g/mL to stimulate the secretion of

proinflammatory cytokines. After a 24 h incubation, the cell culture medium of each group was collected, and the cell fragments were removed by centrifugation at $10,000 \times g$ for 5 min at 4 °C. The proinflammatory cytokines including IL-1 α , IL-1 β , TNF- α , and IL-10 were tested using ELISA assay by following the manufacturer's instructions.

Test of luciferase activity of NF- κ B

RA-FLS cells were seeded in 24-well plate (1×10^5 /well) and cultured at 37 °C for 24 h, then NF- κ B luciferase plasmid and control renilla plasmid were transfected for 48 h. Different concentrations of compound **5e** and LPS (final concentration: 5 μ g/mL) were added to stimulate the secretion of proinflammatory cytokines. After a 48-h incubation later, the cells were collected and the luciferase activity of NF- κ B pathway was detected using the double luciferase system.

Test of MAPK signal pathway

RA-FLS cells (3×10^5 /well/2 mL) were seed in 6-well plate cultured at 37 °C and for 24 h. The cells were pre-incubated with different concentrations of compound **5e** for 1 h, and then were treated with LPS (final concentration: 5 μ g/mL) for 72 h. The cells were collected for Western blotting analysis using anti-ERK1/2, anti-pho-ERK1/2, anti-p38, and anti-pho-p38 antibodies.

Western blotting

Cells were lysed in 20 mM HEPES, pH 7.5, 150 mM NaCl, 1% NP-40, 10 mM tetrasodium pyrophosphate, 100 mM NaF, 17.5 mM 1-glycerophosphate buffer supplemented with Complete Mini Protease Inhibitor Cocktail tablets (Roche Applied Sciences, Indianapolis, IN). Samples separated by SDS-PAGE were transferred to nitrocellulose membranes, blocked with 5% bovine serum albumin (w/v) at room temperature for 1 h, and incubated with primary antibodies (1:1000 dilution) at 4 °C overnight. After incubation with secondary antibody (1:3000 dilution) at room temperature for 1 h, the membranes were developed with chemiluminescence ECL reagent (Lumi Gold, Signa Gen Laboratories, Rockville, MD) and exposed to Hyper film MP (GE Healthcare Life Science, Pittsburgh, PA).

Animals

SD rats weighing 180–220 g were obtained from Jinan Peng Yue Experimental Animal Co., Ltd. (license No.: SCXK (Lu) 2014 Mel 0007), Co., Ltd. The animals were kept in captivity under standard laboratory conditions and ingested standard pellet feed and water at will. All experiments involving live animals and their care are conducted in strict accordance with the "National Care and use of Experimental Animals" of the State Animal Research Bureau (China) and the "regulations on the Administration of Experimental Animals" of the Life Science Research Centre of Shandong first Medical University. The experiment was approved by the Animal Protection and use Committee of Shandong first Medical University. Make every effort to reduce the suffering of animals and reduce the number of animals used.

Establishment and treatment of adjuvant arthritis in rats

The rat adjuvant arthritis model was first established by Freund in the 1950s. Which is simple and selective and is a widely used immune inflammation model^{56,57}. 8-week-old Sprague–Dawley (SD) rats, weighing 180–220 g, were purchased from Jinan Peng

Yue Experimental Animal Breeding Co., Ltd., Jinan, China. The Freund's complete adjuvant (CFA) was purchased from Sigma-Aldrich (item number and specification: F5881-10 mL) and 0.1 mL CFA was injected into the left hind paw of rats to induce inflammation. Arthritis was evaluated by observing the changes in paw swelling every week. After the rat adjuvant arthritis model was successfully established, SD rats were divided into the following groups: normal control group (Ctrl, saline), model group (Model, saline), high dose compound **5e** (60 mg/kg) group, low dose compound **5e** (30 mg/kg) group, positive control MTX (30 mg/kg) group, ($n=5$ per group). The compounds solutions (1 mL) or saline were orally administered to rats once a day for four weeks. Then the mice were euthanized using carbon dioxide (CO₂) and the knee joints were harvested and analysed.

Heat pain threshold determination

The thermal pain threshold of rats was measured by rat claw thermal radiation stimulation pain metre (7371 plantar test, Ugo Basile, Italy). The rats are in a state of free activity, adjust the instrument stimulation intensity (IR = 72), put the left hindlimb on the instrument, and measure the paw recovery time of rats. The reflex latency (paw withdrawal thermal latency, PWTL) of foot contraction induced by thermal stimulation at the bottom of rat foot is the thermal pain threshold.

Rat motor function test

The automatic gait analysis system (DigiGait[®], Mouse Specific, Inc.) was used to test the rats. The rats were placed in the plexi-glass chamber of the treadmill (25 cm long and 5 cm wide). The plastic rear bumper of the rats was used to tap the rats to keep them in the video shooting range. The treadmill speed was set to 18 cm/s. The shutter speed was 40 ms, and each animal recorded 5 s. DigiGait software automatically recognises paw prints and analyses gait parameters such as step size and step width.

Immunohistochemistry (IH)

The rat knee joints were fixed in phosphate-buffered formalin, embed in paraffin, cut in 4 μ m thickness, and applied to slides. The slides were deparaffinized in xylenes using three changes for 5 min each and hydrated gradually through graded alcohols: 100% ethanol twice for 10 min each, 95% ethanol twice for 10 min each, and then deionised water for 1 min with stirring. For antigen unmasking, slides were placed in a container, covered with 10 mM sodium citrate buffer, pH 6.0, and heated in a convection steamer for 1 h. The slides were washed in deionised water three times for 2 min each, blocked with 5% normal goat blocking serum for 30 min, and then stained with haematoxylin and eosin (HE). The slides were analysed and photographed using a micromirror.

Statistics

t-Test and analysis of variance (ANOVA) were performed using StatView (SAS Institute, Cary, NC). The data shown are representatives of at least two independent experiments with similar results, and the data points represent the mean of at least triplicate measurements with error bars corresponding to standard deviation. $p < 0.05$ was considered significantly different.

Disclosure statement

No potential conflict of interest was reported by the author(s).

Funding

This work was financially supported by Science and Shandong Provincial Natural Science Foundation [ZR2018LH021], the Innovation Project of Shandong Academy of Medical Sciences and Academic promotion programmer of Shandong First Medical University [2019LJ003].

ORCID

Jie Sun  <http://orcid.org/0000-0003-0552-9061>

References

- Smolen JS, Aletaha D, Koeller M, et al. New therapies for treatment of rheumatoid arthritis. *Lancet* 2007;370: 1861–74.
- McInnes IB, Schett G. Pathogenetic insights from the treatment of rheumatoid arthritis. *Lancet* 2017;389: 2328–37.
- Firestein GS. Evolving concepts of rheumatoid arthritis. *Nature* 2003;423:356–61.
- Schett G, Gravallesse E. Bone erosion in rheumatoid arthritis: mechanisms, diagnosis and treatment. *Nat Rev Rheumatol* 2012;8:656–64.
- Bartok B, Firestein GS. Fibroblast-like synoviocytes: key effector cells in rheumatoid arthritis. *Immunol Rev* 2010;233: 233–55.
- Bugatti S, Codullo V, Caporali R, Montecucco C. B cells in rheumatoid arthritis. *Autoimmun Rev* 2007;7:137–42.
- Marston B, Palanichamy A, Anolik JH. B cells in the pathogenesis and treatment of rheumatoid arthritis. *Curr Opin Rheumatol* 2010;22:307–15.
- Weyand CM, Seyler TM, Goronzy JJ. B cells in rheumatoid synovitis. *Arthritis Res Ther* 2005;7:S9.
- Noss EH, Brenner MB. The role and therapeutic implications of fibroblast-like synoviocytes in inflammation and cartilage erosion in rheumatoid arthritis. *Immunol Rev* 2008;223: 252–70.
- Wienecke T. Paracetamol versus nonsteroidal anti-inflammatory drugs for rheumatoid arthritis. *Cochrane DB Syst Rev* 2004;1:CD003789.
- Spies CM, Bijlsma JWJ, Burmester G-R, Buttgerit F. Pharmacology of glucocorticoids in rheumatoid arthritis. *Curr Opin Pharmacol* 2010;10:302–7.
- Kremer JM, Alarcón GS, Lightfoot RW, et al. Methotrexate for rheumatoid arthritis. Suggested guidelines for monitoring liver toxicity. *American College of Rheumatology. Arthritis Rheum* 1994;37:316–28.
- Atzeni F, Benucci M, Sallì S, et al. Different effects of biological drugs in rheumatoid arthritis. *Autoimmun Rev* 2013; 12:575–9.
- Feldmann M, Maini RN. Anti-TNF alpha therapy of rheumatoid arthritis: what have we learned? *Annu Rev Immunol* 2001;19:163–96.
- Peng X-M, Damu GLV, Zhou C-H. Current developments of coumarin compounds in medicinal chemistry. *Curr Pharm Des* 2013;19:3884–930.

16. Fylaktakidou KC, Hadjipavlou-Litina DJ, Litinas KE, Nicolaidis DN. Natural and synthetic coumarin derivatives with anti-inflammatory/ antioxidant activities. *Curr Pharm Des* 2004;10:3813–33.
17. Menezes JC, Diederich M. Translational role of natural coumarins and their derivatives as anticancer agents. *Future Med Chem* 2019;11:1057–82.
18. Wei Y, Peng W, Wang D, Hao SH, et al. Design, synthesis, antifungal activity, and 3D-QSAR of coumarin derivatives. *J Pestic Sci* 2018;43:88–95.
19. Kostova I, Bhatia S, Grigorov P, et al. Coumarins as antioxidants. *Curr Med Chem* 2011;18:3929–51.
20. Najafi Z, Mahdavi M, Saeedi M, et al. Novel tacrine-coumarin hybrids linked to 1,2,3-triazole as anti-Alzheimer's compounds: in vitro and in vivo biological evaluation and docking study. *Bioorg Chem* 2019;83:303–16.
21. Kontogiorgis CA, Hadjipavlou-Litina DJ. Synthesis and anti-inflammatory activity of coumarin derivatives. *J Med Chem* 2005;48:6400–8.
22. Arora RK, Kaur N, Bansal Y, Bansal G. Novel coumarin-benzimidazole derivatives as antioxidants and safer anti-inflammatory agents. *Acta Pharm Sin B* 2014;4:368–75.
23. Bansal Y, Sethi P, Bansal G. Coumarin: a potential nucleus for anti-inflammatory molecules. *Med Chem Res* 2013;22:3049–60.
24. Pan R, Gao XH, Li Y, et al. Anti-arthritis effect of scopoletin, a coumarin compound occurring in *Erycibe obtusifolia* Benth stems, is associated with decreased angiogenesis in synovium. *Fundam Clin Pharmacol* 2010;24:477–90.
25. Yao R, Fu Y, Li S, et al. Regulatory effect of daphnetin, a coumarin extracted from *Daphne odora*, on the balance of Treg and Th17 in collagen-induced arthritis. *Eur J Pharmacol* 2011;670:286–94.
26. Cheng JF, Chen M, Wallace D, et al. Discovery and structure-activity relationship of coumarin derivatives as TNF-alpha inhibitors. *Bioorg Med Chem Lett* 2004;14:2411–5.
27. Sahu D, Raghav SK, Gautam H, Das HR. A novel coumarin derivative, 8-methoxy chromen-2-one alleviates collagen induced arthritis by down regulating nitric oxide, NF- κ B and proinflammatory cytokines. *Int Immunopharmacol* 2015;29:891–900.
28. Berrino E, Milazzo L, Micheli L, et al. Synthesis and evaluation of carbonic anhydrase inhibitors with carbon monoxide releasing properties for the management of rheumatoid arthritis. *J Med Chem* 2019;62:7233–49.
29. Bua S, Lucarini L, Micheli L, et al. Bioisosteric development of multitarget nonsteroidal anti-inflammatory drug-carbonic anhydrases inhibitor hybrids for the management of rheumatoid arthritis. *J Med Chem* 2020;63:2325–42.
30. Menicatti M, Pallecchi M, Bua S, et al. Resolution of co-eluting isomers of anti-inflammatory drugs conjugated to carbonic anhydrase inhibitors from plasma in liquid chromatography by energy-resolved tandem mass spectrometry. *J Enzyme Inhib Med Chem* 2018;33:671–9.
31. Supuran CT. Coumarin carbonic anhydrase inhibitors from natural sources. *J Enzyme Inhib Med Chem* 2020;35:1462–70.
32. Bua S, Di Cesare Mannelli L, Vullo D, et al. Design and synthesis of novel nonsteroidal anti-inflammatory drugs and carbonic anhydrase inhibitors hybrids (NSAIDs-CAIs) for the treatment of rheumatoid arthritis. *J Med Chem* 2017;60:1159–70.
33. Di Cesare Mannelli L, Bani D, Bencini A, et al. Therapeutic effects of the superoxide dismutase mimetic compound MnlIme2DO2A on experimental articular pain in rats. *Mediators Inflammation* 2013;2013:905360.
34. Hu Y-H, Yang J, Zhang Y, et al. Synthesis and biological evaluation of 3-(4-aminophenyl)-coumarin derivatives as potential anti-Alzheimer's disease agents. *J Enzyme Inhib Med Chem* 2019;34:1083–92.
35. Liu Y, Pan YF, Xue Y-Q, et al. uPAR promotes tumor-like biological behaviors of fibroblast-like synoviocytes through PI3K/Akt signaling pathway in patients with rheumatoid arthritis. *Cell Mol Immunol* 2018;15:171–81.
36. Firestein GS, Zvaifler NJ. How important are T cells in chronic rheumatoid synovitis?: II. T cell-independent mechanisms from beginning to end. *Arthritis Rheum* 2002;46:298–308.
37. Feldmann M, Brennan FM, Maini RN. Role of cytokines in rheumatoid arthritis. *Annu Rev Immunol* 1996;14:397–440.
38. McInnes IB, Schett G. Cytokines in the pathogenesis of rheumatoid arthritis. *Nat Rev Immunol* 2007;7:429–42.
39. Maini RN, Taylor PC. Anti-cytokine therapy for rheumatoid arthritis. *Annu Rev Med* 2000;51:207–29.
40. Dinarello CA. Interleukin-1. *Cytokine Growth Factor Rev* 1997;8:253–65.
41. Dinarello CA. Biologic basis for interleukin-1 in disease. *Blood* 1996;87:2095–147.
42. Schett G, Dayer J-M, Manger B. Interleukin-1 function and role in rheumatic disease. *Nat Rev Rheumatol* 2016;12:14–24.
43. Genovese MC, Becker J-C, Schiff M, et al. Abatacept for rheumatoid arthritis refractory to tumor necrosis factor alpha inhibition. *N Engl J Med* 2005;353:1114–23.
44. Kollias G, Douni E, Kassiotis G, Kontoyiannis D. On the role of tumor necrosis factor and receptors in models of multiorgan failure, rheumatoid arthritis, multiple sclerosis and inflammatory bowel disease. *Immunol Rev* 1999;169:175–94.
45. Palladino MA, Bahjat FR, Theodorakis EA, Moldawer LL. Anti-TNF-alpha therapies: the next generation. *Nat Rev Drug Discov* 2003;2:736–46.
46. Katsikis PD, Chu CQ, Brennan FM, et al. Immunoregulatory role of interleukin 10 in rheumatoid arthritis. *J Exp Med* 1994;179:1517–27.
47. Moore KW, O'Garra A, de Waal Malefyt R, et al. Interleukin-10. *Annu Rev Immunol* 1993;11:165–90.
48. Bissantz C, Kuhn B, Stahl M. A medicinal chemist's guide to molecular interactions. *J Med Chem* 2010;53:5061–84.
49. Baker RG, Hayden MS, Ghosh S. NF- κ B, inflammation, and metabolic disease. *Cell Metab* 2011;13:11–22.
50. Lin X, Wang M, Zhang J, Xu R. p38 MAPK: a potential target of chronic pain. *Curr Med Chem* 2014;21:4405–18.
51. Tian W, Zhang Z, Cohen DM. MAPK signaling and the kidney. *Am J Physiol Renal Physiol* 2000;279:F593–604.
52. Dent P, Yacoub A, Fisher PB, et al. MAPK pathways in radiation responses. *Oncogene* 2003;22:5885–96.
53. He Y-H, Zhou J, Wang Y-S, et al. Anti-inflammatory and anti-oxidative effects of cherries on Freund's adjuvant-induced arthritis in rats. *Scandinavian J Rheumatol* 2006;35:356–8.
54. Guo W-Y, Chen L-Z, Shen B-N, et al. Synthesis and in vitro and in vivo anti-inflammatory activity of novel

- 4-ferrocenylchroman-2-one derivatives. *J Enzyme Inhib Med Chem* 2019;34:1678–89.
55. Maresca M, Micheli L, Cinci L, et al. Pain relieving and protective effects of *Astragalus* hydroalcoholic extract in rat arthritis models. *J Pharm Pharmacol* 2017;69:1858–70.
56. Wang S, Wang Y, Liu X, et al. Anti-inflammatory and anti-arthritic effects of taraxasterol on adjuvant-induced arthritis in rats. *J Ethnopharmacol* 2016;187:42–8.
57. Wang X, He X, Zhang C-F, et al. Anti-arthritic effect of berberine on adjuvant-induced rheumatoid arthritis in rats. *Biomed Pharmacother* 2017;89:887–93.