Actions of lead on transmitter release at mouse motor nerve terminals

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Abstract. The actions of lead (Pb²⁺) on transmitter release were studied at neuromuscular junctions in mouse diaphragm in vitro. The quantal content of end-plate potentials (EPPs) was reduced by Pb^{2+} in a dose-related manner consistent with inhibition of Ca^{2+} entry into nerve terminals, with a half-maximal effect at 1.4 μ M (in 0.5 mM Ca²⁺ and 2 mM Mg²⁺). Pb²⁺ also inhibited the increased frequency of MEPPs (f_{MEPP} where MEPPs denotes miniature EPPs) produced by Ba²⁺ in the presence of raised K^+ , blocking the calculated Ba^{2+} entry half-maximally at 170 μ M. However, at concentrations of 50-200 nM, Pb²⁺ often increased f_{MEPP} in 20 mM K⁺ in the presence of Ca²⁺ and acted to promote the irreversible effect of lanthanum (La³⁺) to raise f_{MEPP} . In nominally Ca²⁺-free solution with 20 mM K⁺, brief (1 min) application of Pb²⁺ (20-320 μ M) caused rapid dose-dependent reversible rises in f_{MEPP} . With prolonged exposure to Pb^{2+} , f_{MEPP} rose and then slowly declined; after removal of Pb^{2+} , once f_{MEPP} had fallen to low levels, f_{MEPP} responded nearly normally to Ca^{2+} or ethanol, but not to Pb^{2+} itself. In 5 mM K⁺, 0 mM Ca²⁺ and varied $[Pb^{2+}]$ (where [] denotes concentration), nerve stimulation caused no EPPs, but prolonged tetanic stimulation produced increases in f_{MEPP} graded with [Pb²⁺] that persisted as a "tail"; results were consistent with growth of f_{MEPP} with the 4th power of intracellular Pb²⁺ and removal of intracellular Pb^{2+} with a time constant of about 30 s. These results suggest that Pb^{2+} acts to block the entry of Ca^{2+} and Ba^{2+} into the terminal via voltage-gated Ca^{2+} channels through which Pb^{2+} , at higher concentrations, also penetrates and then acts as an agonist at intracellular sites that govern transmitter release.

Key words: Lead – Nerve terminal – Transmitter release – Calcium channels

Introduction

It is now well established that at the neuromuscular junction Ca²⁺ enters the nerve terminal via voltage-gated channels and functions to link transmitter release to nerve terminal depolarization [22]. However, many ions that block depolarization/Ca²⁺ transmitter release, apparently by blocking Ca²⁺ entry, themselves induce or enhance release at depolarized terminals. These include manganese (Mn²⁺, [3]), cobalt (Co²⁺, [24]), lanthanum (La³⁺, [6]), cadmium (Cd²⁺, [9, 15]) and zinc (Zn²⁺, [12, 14, 23]). Silbergeld et al. [20] found that Pb²⁺ decreased the force of contraction and increased the latency between nerve stimulation and contraction in mouse and rat in vitro; a blocking activity of Pb²⁺ was subsequently demonstrated at neuromuscular junctions of frog [5, 13] and of rat diaphragm [2]. In addition, Pb²⁺ can also act to increase "spontaneous" transmitter release [1, 2, 5].

The experiments described here were carried out to investigate further the interaction of Pb²⁺ and Ca²⁺ at both extracellular and intracellular sites. The results indicate that Pb²⁺ has several effects on transmitter release, acting, not only as a competitive blocker of Ca²⁺ (and Ba²⁺) entry at extracellular sites, but also as a substitute for Ca²⁺ in supporting depolarization-induced release. In addition, at a very low concentration range (nanomolar), Pb²⁺ enhances the effectiveness of Ca²⁺ or La³⁺ to raise the frequency of miniature end-plate potentials (f_{MEPP}) in high K⁺.

Materials and methods

Experiments were performed upon hemidiaphragms from anaesthetized mice. The techniques used for the mounting and superfusion of the preparation that allowed switching of the bathing solution in a few seconds have been described elsewhere [4]. Intracellular recording of MEPPs and EPPs at end-plates was conventional, using microelectrodes filled with 3 M KCl. Because Pb²⁺ precipitates in bicarbonate/phosphate buffered solutions, experiments were carried out in solutions buffered with 4-(2hydroxyethyl)-1-piperazineethanesulphonic acid (HEPES) and

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bubbled with 100% O₂. As in previous work [9, 23] the standard solution for most experiments using raised K⁺ had the following composition (mM): NaNO₃ 100, KCl 20, MgCl₂ 1, glucose 11, HEPES 3, pH 7.3. Sucrose (60 mM) provided isotonicity with solutions containing 150 mM Na⁺ and 5 mM K⁺. Lowered Na⁺ concentration raises the apparent potency of Ca²⁺ for extracellular sites so that half-maximal responses of f_{MEPP} to raised Ca²⁺ are obtained at lower concentrations [8]. Replacement of Cl⁻ by NO₃⁻ improves signal/noise ratio for recording MEPPs, which does not significantly affect presynaptic mechanisms [19]. In experiments using nerve stimulation Cl⁻ was used rather than NO₃⁻ and [K⁺] (where [] denotes concentration) was 5 mM. To remove Pb²⁺ the preparation was washed with solution containing Ca-EDTA [usually using 0.15 mM Ca²⁺ and 0.1 mM ethylenediaminetetraacetic acid (EDTA)]. Silen and Martell [21] quote the dissociation constant of the Pb-EDTA complex as $10^{-18.04}$, about 7 orders of magnitude smaller than that for Ca-EDTA ($10^{-10.6}$).

MEPPs were counted by a microcomputer using a program that provided sequential displays of 0.4-s portions of the recorded signal, with lines indicated each MEPP that was counted by the computer; this permitted continuous monitoring of the accuracy of the count. To follow the time course of changes in f_{MEPP} , MEPPs were either recorded continuously at individual junctions or muscle fibres were penetrated randomly and the time of penetration and the f_{MEPP} for each junction recorded. For assessment of quantal content of EPPs we used a computer program that "deconvolutes" the EPP into quantal components, and counts MEPPs at all points in the record. This program will be described elsewhere.

Results

Blockade by Pb^{2+} of Ca^{2+} -dependent release

Pb²⁺ has previously been reported to inhibit Ca²⁺-induced release of transmitter at neuromuscular junctions of frog [5] and rat [2]. The same phenomenon at a mouse junction is illustrated in Fig. 1a. In the presence of 0.5 mM Ca^{2+} and 2 mM Mg^{2+} , nerve stimulation produced EPPs with a quantal content of 1.5, which was reversibly diminished by $0.5-16 \mu M Pb^{2+}$ in a doserelated manner. By interpolation, the half-maximal effect was at 0.24 µM. If transmitter release is proportional to the 4th power of intracellular Ca^{2+} [10] inhibition of Ca^{2+} entry can be calculated from the reduction of the 4th root of the release rate corresponding to the quantal content of the EPP (cf. [23]). On this basis the halfmaximal effect of Pb^{2+} to reduce Ca^{2+} entry in this experiment was $1.4 \ \mu M \ Pb^{2+}$. The reduction in quantal content by Pb²⁺ was associated with a loss of "facilitation" in short trains, as described by Zengel et al. [25] for Cd^{2+} , the facilitation not being restored with addition of Ca^{2+} . This is illustrated in Fig. 1b which shows, in control, typical facilitation of quantal content in trains of 7 pulses at 100 Hz. Here 100 μ M Pb²⁺ reduced the quantal content of EPPs to very low levels. Increasing Ca^{2+} to 2 mM restored the original quantal content but facilitation remained absent. It is notable that the restoration of quantal content by only a quadrupling of $[Ca^{2+}]$, in the presence of 100 μ M Pb²⁺, would appear to contradict the previous value of 1.4 μ M for the apparent dissociation constant of Pb^{2+} for block of Ca^{2+} channels. Previously, with Cd^{2+} [10] and with Zn^{2+} [23] it was found that the apparent potency of these ions is diminished by Ca²⁺ to an extent greater than expected



Fig. 1. A Inhibition by Pb^{2+} of end-plate potentials (EPPs) induced by 80-Hz trains in 5 mM K⁺, 0.5 mM Ca²⁺ and 2 mM Mg²⁺. The *line* is a theoretical curve for single-site action of Pb^{2+} to block Ca²⁺ entry, with a half-maximal effect at 1.4 μ M, assuming transmitter release is proportional to the 4th power of internal Ca²⁺. **B** Blockade by Pb^{2+} of facilitation in 100-Hz trains of 7 stimuli, in 5 mM K⁺ and 2 mM Mg²⁺. Open circles, (controls) 0.5 mM Ca²⁺; solid triangles, 1 mM Ca²⁺ and 100 μ M Pb²⁺; solid squares, 2 mM Ca²⁺ and 100 μ M Pb²⁺



Fig. 2. Inhibition by Pb^{2+} of the increase in the frequency of miniature EPPs (f_{MEPP}) produced by 0.3 mM Ba²⁺ in the presence of 20 mM K⁺; data from one junction. The *ordinate* represents the difference between f_{MEPP} in the presence of Ba²⁺ (f_m) and f_{MEPP} in the absence of Ba²⁺ (f_o) . Open circle, control; filled circles, with Pb²⁺

simply from competition of Ca^{2+} with the blocking ion at a single site.

Effect of Pb^{2+} to increase f_{MEPP} in high K^+

From the above result it would be expected that Pb^{2+} would block the enhanced f_{MEPP} , produced by Ca^{2+} or Ba^{2+} at end-plates depolarized by raised K⁺ [10]; with Ba^{2+} , in 20 mM K⁺, this was indeed always the case. In the example in Fig. 2 the effect at 50 nM Pb²⁺ is less than



Fig. 3A, B. Effect of Pb^{2+} to increase neurotransmitter release produced by Ca^{2+} in the presence of 20 mM K⁺. A f_{MEPP} was about 21 s⁻¹ in 0.4 mM Ca²⁺ (*open circles*). Addition of 100 nM Pb²⁺ (*filled circles*) caused a transient reduction followed by a rise. f_{MEPP} fell nearly to control values upon withdrawal of Pb²⁺. B f_{MEPP} was about 1 s⁻¹ before (*open circles*) and with 100 nM Pb²⁺ (*filled circles*). With 100 nM Pb²⁺ and 1 mM Ca²⁺ (*filled squares*) f_{MEPP} rose to 250 s⁻¹; it returned to control with "wash" with 50 μ M Ca²⁺ and 100 μ M Ca-EDTA (*open triangles*). 1 mM Ca²⁺ in the absence of Pb²⁺ (*open squares*) raised f_{MEPP} to 150 s⁻¹ and subsequent addition of 100 nM Pb²⁺ (*filled squares*) raised f_{MEPP} to 250 s⁻¹

expected from the effects at a higher concentration, but the data otherwise fit half blockade of Ba^{2+} entry at 170 nM (see [23] for method of calculation), if release is proportional to the 4th power of Ba^{2+} entry [17]. The relatively high potency of Pb^{2+} as a blocker when tested versus Ba^{2+} rather than Ca^{2+} was also seen with Zn^{2+} [23] and with Cd^{2+} [10].

The raised f_{MEPP} induced by Ca²⁺ in 20 mM K⁺ was also usually inhibited by Pb²⁺. However, this inhibition was often transient, followed by a rise in f_{MEPP} despite the continued presence of Pb²⁺. At relatively high concentrations of Pb²⁺ the rise in f_{MEPP} with Pb²⁺ also occurred in the absence of Ca²⁺ (see below) but at low concentrations, e.g. 100 or 200 nM (Fig. 3) the rise did not occur in the absence of Ca²⁺, but did occur in 0.4 mM Ca²⁺, 1 mM Ca²⁺, and sometimes in 2 mM Ca²⁺ (Fig. 3a, b).

To test the possibility that this phenomenon might be related to an effect of Pb²⁺ to enhance entry of other ions into the nerve terminal we examined the effect of Pb²⁺ on the development of the irreversible action of La³⁺ to raise f_{MEPP} ; Curtis et al. [6] found that small amounts of Ca²⁺ accelerate the irreversible development of a high f_{MEPP} in the presence of La³⁺ which is attributable to entry of La³⁺ into nerve terminals via Ca²⁺ channels. Preparations were incubated with 0.5 μ M La³⁺ (in 10 mM K⁺) together with different concentrations of



Fig. 4. Pb^{2+} increases the irreversible action of lanthanum (La³⁺) in 10 mM K⁺ to raise f_{MEPP} . Each *point* represents the geometric mean from 22–71 junctions (\pm S.E.) in a segment of a diaphragm, in 50 µM Ca²⁺ and 100 µM Ca-EDTA, following incubation for 30 min with 0.5 µM La³⁺ alone (control, *open symbol*) or with various concentrations of Pb²⁺ (*filled circles*), or Ca²⁺ (*filled triangles*). The experiment with Ca²⁺ was on a different diaphragm from that with Pb²⁺. Incubation with Pb²⁺ alone (200 nM, not shown) had no effect to raise subsequent f_{MEPP}

 Pb^{2+} for 30 min and f_{MEPP} recorded subsequently in the absence of La³⁺ or Pb²⁺ (with 0.1 mM Ca-EDTA to exclude extracellular Pb^{2+} and La^{3+}). The results (Fig. 4) show that concurrent exposure to Pb^{2+} , in the range of 50-200 nM, increased the effect of exposure to 0.5 μ M La^{3+} , with much greater potency, and perhaps more efficacy than Ca^{2+} . In control experiments exposure of preparations for 30 min to Pb²⁺ (200 nM) alone caused no increase in the subsequent f_{MEPP} . Moreover, the effect of Pb²⁺ depended upon its simultaneous presence with La^{3+} ; exposure to 200 nM Pb²⁺ for 30 min, followed by "wash" with 0.1 mM Ca-EDTA and subsequent exposure to 0.5 μ M La³⁺ gave a maintained f_{MEPP} in the absence of Ca²⁺ [mean $\log_{10} f_{\text{MEPP}} = 0.37 \pm 0.09 \ (\pm \text{ S.E.}, n =$ 35)] the same as exposure to only $0.5 \,\mu\text{M} \,\text{La}^{3+}$ [mean $\log_{10} f_{\text{MEPP}} = 0.40 \pm 0.09, (\pm \text{S.E.}, n = 43)$]. Thus, Pb²⁺ at about 100 nM, apparently promotes the entry of La³⁺ into nerve terminals, in the same way as Ca²⁺ at about 100 µM, perhaps by promoting opening of channels. A similar action exerted on entry of Ca2+ would account for the enhancement by 100 nM Pb²⁺ on f_{MEPP} induced by Ca^{2+} in raised K⁺ (Fig. 3), but only if Ca^{2+} itself has less efficacy then Pb²⁺ for this action.

In 20 mM K⁺ in the absence of Ca²⁺, Pb²⁺ at sufficiently high concentrations produced an increase in f_{MEPP} that developed within about 30 s and subsided quickly upon removal of Pb²⁺. In the example in Fig. 5a, 160 μ M Pb²⁺ caused f_{MEPP} to rise from less than 1 s⁻¹ to 100 s⁻¹ with most of the rise complete in 30 s; upon "washing" with solution containing 0.1 mM Ca-EDTA f_{MEPP} returned to control values in about 1 min. In Fig. 5b average data from 6 junctions are plotted as $f_{\text{MEPP}}^{(1/4)}$. $f_o^{(1/4)}$ (i.e., the increment in the 4th root of f_{MEPP} , which, see below, should be proportional to internal Pb²⁺) versus external Pb²⁺. The half maximally effective [Pb²⁺] in terms of rise of the 4th root of f_{MEPP} was at 85 μ M; in terms of f_{MEPP} , the half maximally effective [Pb²⁺] was at 135 μ M. Thus, saturation of this effect of Pb²⁺ occurred at



Fig. 5A, B. Rapid and reversible effect of Pb²⁺ to raise f_{MEPP} in the presence of 20 mM K⁺ and 0 mM Ca²⁺. A Example of 1-min application of 160 μ M Pb²⁺ (*solid circles*) raising f_{MEPP} , with reversal upon removal of Pb²⁺ using 0.1 mM Ca-EDTA (*open triangles*). Open circles represent control (no Pb²⁺). B Dose/response curve, where the points represent mean $f_{\text{MEPP}}^{1/4}$ at 0–60 s after applying Pb²⁺, minus 4th root of spontaneous f_{MEPP} in 0 mM Ca²⁺ ($f_0^{1/4}$). Each point represents an average from 6 junctions \pm S.E.

concentrations very much higher than the concentrations that block Ca^{2+} entry.

With prolonged exposure to Pb^{2+} , at 10 µM or greater, f_{MEPP} rose to levels higher than obtained with 1-min exposures, followed by a slow decline despite the continued presence of Pb^{2+} . The effect of Pb^{2+} to raise f_{MEPP} remained reversible; f_{MEPP} rapidly fell to control levels or lower with removal of Pb^{2+} and "wash" with solution containing 0.1 mM Ca-EDTA, even during the declining phase (Fig. 6a). However, once f_{MEPP} had declined in the continued presence of Pb^{2+} , washing with Ca-EDTA and reexposure to Pb^{2+} did not restore the high f_{MEPP} found after the initial exposure to Pb^{2+} . Data obtained by multiple sampling in a diaphragm continuously exposed to 100 µM Pb²⁺ (in 20 mM K⁺ and 0 mM Ca²⁺), is shown in Fig. 6b illustrating the large variations in frequencies attained at different junctions.

Even in nominally Ca^{2+} -free solution, the possibility exists that the effect of Pb²⁺ of increasing f_{MEPP} might be secondary to increase in intracellular Ca²⁺. Therefore, the effect of Pb²⁺ in 20 mM K⁺ was tested in preparations incubated for 5 h in 20 mM K⁺ in very low buffered Ca²⁺ (0.1 mM EDTA plus 10 μ M Ca²⁺); responses to 100 μ M Pb²⁺ (Fig. 6c) were unaffected.

Although brief (1 min) exposure to 10 μ M Pb²⁺ had little effect to raise f_{MEPP} in 20 mM K⁺ (Fig. 5b) with more prolonged exposure there was generally a rise that was complete within about 10 min, to about 30 s⁻¹, followed by a fall over the next hour (Fig. 7a, b). In 5 mM



Fig. 6A – C. Response of f_{MEPP} to 100 µM Pb²⁺ in 20 mM K⁺ and 0 mM Ca²⁺. A Addition of Pb²⁺ (solid circles) caused a rapid increase in f_{MEPP} followed by a decline. f_{MEPP} fell to control or lower upon "wash" with Pb²⁺, 50 µM Ca²⁺ and 100 µM Ca-EDTA (open circles). Repeated applications of Pb²⁺ caused progressively less effect. Data from one junction. B Variation between junctions of the effect of 100 µM Pb²⁺ (filled circles) to raise f_{MEPP} in 20 mM K⁺ and 0 mM Ca²⁺. C Increase of f_{MEPP} by 100 µM Pb²⁺ (solid circles) at a junction in a preparation previously incubated for 5 h in 20 mM K⁺, 0.1 mM EDTA plus 10 µM Ca²⁺ (open circles)

K⁺, the rise was less and developed more slowly (Fig. 7a), in contrast to the observation of Anwyl et al. [1], suggesting that the effect of Pb²⁺ is secondary to entry into the nerve terminal cytoplasm via voltage-gated channels, presumably those that normally admit Ca²⁺. In support of this, the response to Pb²⁺ was inhibited by 4 mM Mg²⁺ (Fig. 7b). Following such long exposures to Pb²⁺, f_{MEPP} fell to control levels after withdrawal of Pb²⁺.

Effect of Pb^{2+} on f_{MEPP} induced by nerve stimulation

In the presence of Pb²⁺ and absence of Ca²⁺ (5 mM K⁺ and 1 mM Mg²⁺), nerve stimulation caused a rise in f_{MEPP} , much greater than occurred in the absence of Pb²⁺; this persisted after a train as a "tail" that declined over a



Fig. 7. A The rise in f_{MEPP} produced by 10 μ M Pb²⁺ in 0 mM Ca²⁺ and 20 mM K⁺ (solid triangles) or 5 mM K⁺ (solid circles). **B** Inhibition by 4 mM Mg²⁺ (solid triangles) of the rise of f_{MEPP} produced by 10 μ M Pb²⁺ (20 mM K⁺ and 0 mM Ca²⁺), compared with 1 mM Mg²⁺ (solid circles). In both graphs, each point represents the mean from about 20 junctions \pm S.E.

period of seconds. The maximum f_{MEPP} attained grew with the number of impulses applied.

An example of time course of development and decline of f_{MEPP} during and after nerve stimulation is shown in Fig. 8a. With nerve stimulation in the presence of Pb^{2+} , the rise and fall of f_{MEPP} were asymmetrical in time course when data were plotted either linearly or semilogarithmically versus time. Here, in 100 µM Pb²⁺, during a 40-Hz train f_{MEPP} rose from the control value (f_{o}) with a time constant (τ) of 64 s (by non-linear least squares fitting) but after that the train fell with a τ of 16 s. Plots of $f_{\rm m}^{1/n}$ versus time (where $f_{\rm m}$ denotes mean $f_{\rm MEPP}$) became increasingly symmetrical with increasing *n* up a value equal to 4, at which τ for growth of $f_m^{1/4}$ was 34 s, while τ for decay of $f_{\rm m}^{1/4}$ was 32 s; at *n* equal to 5, τ values were 32 and 33 s, respectively. This corresponds to what would be expected if f_{MEPP} is proportional to the 4th or 5th power of internal Pb²⁺, which rises with each "injection" of Pb^{2+} by a nerve impulse and is disposed of by a 1st order process with a τ value of 33 s. Graphs (not shown) of the \log_{10} of the absolute value of $(f_m^{1/4}-f_f^{1/4})$ versus time, $f_{\rm f}$ being maximum $f_{\rm m}$ value for the rising phase and f_0 for the falling phase, showed good fits of both phases to a single exponential with a time constant of 33 s.

An example of the relation of "tail" intensity to number of stimuli in 30-s trains, in 100 μ M Pb²⁺, is shown in Fig. 8b. Designating the f_m value over the period of 30 s after the last pulse in the train as f_t , $f_t^{1/4}$ (or $f_t^{1/5}$, not shown) varied linearly with the stimulation frequency, i.e. with the number of impulses in the train, in the same way as "tails" of raised f_{MEPP} produced by stimulation in



Fig. 8A – C. The rise of f_{MEPP} produced by nerve stimulation in the presence of 100 μ M Pb²⁺, 1 mM Mg²⁺, 0 mM Ca²⁺ and 5 mM K⁺. **A** Time course of the rise and fall of f_{MEPP} before (*open circle*) during (*filled circles*) and after (*open circle*) nerve stimulation for 2 min at 40 Hz. **B** Linearity with number of stimuli in 30-s trains of the 1/4 power of the "Pb²⁺ tail" intensity, estimated as the average f_{MEPP} in the 1st 30 s after the train. In the absence of Pb²⁺ such trains caused much smaller increases in f_{MEPP} . **C** Gradation of apparent Pb²⁺ entry with external [Pb²⁺]. Stimuli were given in 30-s trains at a [Pb²⁺] of 25-200 μ M in 0 mM Ca²⁺ and 1 mM Mg²⁺. "Pb²⁺ entry" per pulse was calculated as the increment of f_{MEPP} raised to the power 1/4 caused by each train of number k as $(f_t^{r1/4}-f_0^{r1/4})/k$, where f_0 is control value of f_{MEPP} and f_t is the f_{MEPP} just after the train by extrapolation (using a 30-s time constant) from the average f_{MEPP} at 0-30 s after the train

the presence of Ba^{2+} and attributable to accumulation of Ba^{2+} in the nerve terminal [17]. The calculated per pulse increment of $f_{MEPP}^{1/4}$ in this example is much the same as that found with 50 μ M Ba^{2+} [18] but, in contrast to what was found with Ba^{2+} , this could not be much increased by increase of Pb^{2+} above 100 μ M (Fig. 8c).

Corresponding to the low effectiveness (or low entry) of Pb^{2+} , nerve stimulation did not induce any noticeable EPP, even with $[Pb^{2+}]$ raised up to 1 mM; latency histograms of quanta released after nerve stimuli showed no consistent increase in frequency in the period of 0.8-2.5 ms, in which the EPP normally occurs.

Possible intracellular effect on release

After prolonged exposure to Zn^{2+} there occurs a complete blockade of the response of f_{MEPP} to ethanol (cf. [18]) and to Ca^{2+} in raised K⁺ [23]. To determine whether a similar effect might be exerted by Pb²⁺, as suggested by the decline in f_{MEPP} that occurred in the continued presence of Pb²⁺ (Fig. 6), f_{MEPP} was measured in 20 mM K⁺ with either added ethanol or Ca²⁺, before and after a 1-h exposure to 100 μ M Pb²⁺. To ensure the absence of extracellular Pb²⁺, 0.15 mM Ca²⁺ and 0.1 mM EDTA was added to all solutions except that containing Pb^{2+} . After Pb²⁺ exposure, f_{MEPP} (about 0.7 s⁻¹) was not significantly changed in 0 mM Ca²⁺ but there was a tendency to a lower f_{MEPP} than in the controls, with Ca²⁺ and ethanol. In 2 out of 3 preparations f_{MEPP} in 1 mM Ca²⁺ (normally about 100 s^{-1}) was reduced relative to the controls (to 45% and 54%) and in both preparations tested with 0.8 M ethanol (producing f_{MEPP} of about 50 s⁻¹ in controls) f_{MEPP} was reduced after exposure to Pb²⁺, to 70.5% and 52% of control values. These reductions are small compared to the effects of Ca^{2+} and ethanol to raise f_{MEPP} . This result suggests that there is relatively little (if any) long term accumulation of Pb²⁺ and "toxic" action, in contrast to Zn^{2+} .

Discussion

The present results concur with previous observations that Pb^{2+} , like some other divalent ions, not only inhibits Ca²⁺-mediated transmitter release, presumably by blocking entry of Ca²⁺ into nerve terminals via voltagesensitive channels, but also can itself promote release [1, 2, 5]. The effect to inhibit release resembles that shown by Cd^{2+} [10] and Zn^{2+} [23] in that the apparent potency of the blocking ion is much greater when tested versus Ba^{2+} in raised K⁺ than when tested versus Ca^{2+} (giving a similar f_{MEPP}) and apparent potency is reduced by Ca²⁺ much more than is compatible with simple competition of the ions at a single site. The action of Pb^{2+} rapidly to promote release in raised K^+ , in the absence of Ca^{2+} , resembles that of Zn^{2+} in that it is evident only at concentrations much higher than those that apparently block Ca^{2+} or Ba^{2+} entry by 50%, i.e. it appears that a concentration of Pb²⁺ which blocks entry of other ions does not block its own entry. This phenomenon may be the same as that seen in cardiac Ca^{2+} channels where Ca^{2+} blocks Na⁺ flux in the micromolar range but Ca²⁺ flux grades with $[Ca^{2+}]$ in the millimolar range, which is explicable if channels have more than one binding site [11]. Complexity of ion interaction with presynaptic Ca^{2+} channels is also indicated by the sensitivity to Ca^{2+} of the inhibi-tory effect of Pb²⁺, Zn²⁺, and Cd²⁺, on release mediated by Ca²⁺ or Ba²⁺. Pb²⁺ contrasts with Zn²⁺ in its much greater potency

Pb²⁺ contrasts with Zn²⁺ in its much greater potency and effectiveness in producing a rapid increase in f_{MEPP} in raised K⁺, and with Cd²⁺, for which no such effect is visible, and in the apparently complete reversibility of this effect. Moreover, the long term effects seen with Cd²⁺ and Zn²⁺, apparently due to irreversible sequelae

of ion entry into the nerve terminal, are nearly absent; there is little if any tendency for release to be reduced after prolonged exposure to Pb^{2+} . The effect to raise f_{MEPP} in raised K⁺ is consistent with Pb²⁺ acting within the nerve terminal, after entry via voltage-gated channels, in the same way as occurs with tetanic stimulation. The effect of Pb²⁺ to support the temporarily raised f_{MEPP} during and after tetanic stimulation closely resembles the effect seen with Ba²⁺ [16], the only differences being a more prolonged time course and apparent saturation at more than 100 µM or so. Notably, the Hill coefficient for cooperativity of Pb^{2+} to induce release, once inside the terminal, appears to be 4 (or 5) which is the same as for Ba^{2+} [16, 17] and for Ca^{2+} [10]. The lack of an EPP with Pb^{2+} (in contrast to Ba^{2+}) follows from the rather small effect of Pb^{2+} (per pulse) that is attainable in comparison to that with Ba^{2+} . However, we cannot rule out the possibility that the intracellular effect of Pb²⁺ to induce release might be secondary to intracellular release of Ca^{2+} .

The action of Pb²⁺ to block facilitation has been observed previously with Cd²⁺ and Zn²⁺ [25], and presumably reflects the same phenomenon for all three ions. It is not impossible that this action could be secondary to block of Ca²⁺ entry, although restoration of quantal content by added Ca^{2+} in the continued presence of Pb²⁺ does not restore facilitation, since the effect of Pb^{2+} to block Ca^{2+} entry might grow with repetitive stimulation. Alternatively, it is conceivable that Pb^{2+} is much more effective than Ca²⁺ in activating an intracellular mechanism that produces facilitation, and saturates at moderate intracellular Ca^{2+} [7], and therefore obviates the normal facilitation produced by Ca^{2+} entering with each impulse. The action of Pb^{2+} at very low concentrations to increase Ca²⁺-mediated release, and to promote the development of a high f_{MEPP} in the presence of La³⁺ presumably reflects an increase in the rate of opening of presynaptic Ca²⁺ channels, and supports the idea that the opening of these channels may normally be governed partially by Ca^{2+} [6].

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