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Candidate Gene Analysis Using Imputed Genotypes: Cell Cycle Single-Nucleotide Polymorphisms and Ovarian Cancer Risk

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Abstract

Polymorphisms in genes critical to cell cycle control are outstanding candidates for association with ovarian cancer risk; numerous genes have been interrogated by multiple research groups using differing tagging single-nucleotide polymorphism (SNP) sets. To maximize information gleaned from existing genotype data, we conducted a combined analysis of five independent studies of invasive epithelial ovarian cancer. Up to 2,120 cases and 3,382 controls were genotyped in the course of two collaborations at a variety of SNPs in 11 cell cycle genes (*CDKN2C*, *CDKN1A*, *CCND3*, *CCND1*, *CCND2*, *CDKN1B*, *CDK2*, *CDK4*, *RB1*, *CDKN2D*, and *CCNE1*) and one gene region (*CDKN2A-CDKN2B*). Because of the semi-overlapping nature of the 123 assayed tagging SNPs, we performed multiple imputation based on

fastPHASE using data from White non-Hispanic study participants and participants in the international Hap-Map Consortium and National Institute of Environmental Health Sciences SNPs Program. Logistic regression assuming a log-additive model was done on combined and imputed data. We observed strengthened signals in imputation-based analyses at several SNPs, particularly *CDKN2A-CDKN2B* rs3731239; *CCND1* rs602652, rs3212879, rs649392, and rs3212891; *CDK2* rs2069391, rs2069414, and rs17528736; and *CCNE1* rs3218036. These results exemplify the utility of imputation in candidate gene studies and lend evidence to a role of cell cycle genes in ovarian cancer etiology, suggest a reduced set of SNPs to target in additional cases and controls. (Cancer Epidemiol Biomarkers Prev 2009;18(3):935–44)

Introduction

Because genes regulating cell cycle control are excellent candidates for cancer risk, multiple groups have targeted these genes for etiologic investigation. Progression of cells from G₁ phase to S phase to G₂ phase is closely regulated by the retinoblastoma protein (pRb), cyclins, cyclin-dependent kinases (CDK), and CDK inhibitors (Fig. 1). Loss of growth control is a key trait of cancerous cells, resulting from abnormalities in cell replication, which is controlled by cell cycle genes (1, 2). Inhibitors

of the cyclin/CDK complexes also regulate cell cycle progression by controlling the activation of these complexes (3, 4). Studies of inherited variation in cell cycle genes suggest that genotypes in this pathway may be associated with risk of breast cancer (5, 6), prostate cancer (7), lung cancer (8), bladder cancer (9, 10), and oral cancer (11), but not necessarily colorectal cancer (12).

Evidence for a role of cell cycle variants in ovarian cancer comes from several lines of research. Overexpression of cyclins D1, D2, and E1 and deletion of CDK inhibitors 2A (p16) and 2B (p15) have been observed in ovarian cancers (13–15). In addition, ovarian cancers frequently have altered retinoblastoma protein (pRb), which regulates the G₁-to-S phase transition when cells either arrest development or proliferate (16). The complex interplay of cyclins D1, D2, D3, and E1; CDK2 and CDK4; CDK inhibitors 1A (p21) and 1B (p27); CDK4 inhibitors 2A (p16), 2B (p15), 2C (p18), and 2D (p19); and pRb suggests that perturbation of any of these molecules via germ-line variation may predispose a woman to ovarian carcinogenesis. Finally, previous reports of inherited variation and ovarian cancer survival have shown suggestive results (17, 18).

Improved precision of disease-risk estimates associated with single-nucleotide polymorphisms (SNP) in cell cycle control genes can be obtained with pooled analyses of several study populations. The use of tagSNPs and

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htSNPs has facilitated cost-savings in individual studies; however, tagging SNP sets often vary across studies due to the use of different algorithms, parameter values, data sources, and genotyping platforms (19). In the context of failed genotyping or multiple genome-wide platforms, several tools for imputation and analysis of missing genotypes have been developed (20-26). Although potentially informative, these tools have not been routinely applied to the candidate gene setting where multiple studies targeted differing but correlated SNPs.

Here, we analyze data from five ovarian cancer study populations that, as part of two collaborations, tagged common variation in 11 cell cycle genes and in one gene region (*CDKN2C*, *CDKN1A*, *CCND3*, the *CDKN2A-CDKN2B* region, *CCND1*, *CCND2*, *CDKN1B*, *CDK2*, *CDK4*, *RB1*, *CDKN2D*, and *CCNE1*). A total of 123 SNPs were genotyped but only 24 SNPs were genotyped in both collaborations; because SNPs sets were correlated but differed, we combined data using multiple imputation (17, 27). We present results of observed and imputed analyses of ovarian cancer risk, suggest SNPs worthy of additional genotyping, and provide guidance for the application of this and other imputation methods.

Materials and Methods

Study Populations. The first genotyping effort (28, 29) used subjects recruited into two case-control studies at Mayo Clinic in Rochester, Minnesota (MAY) and at Duke University in Durham, North Carolina (NCO). At Mayo Clinic, cases were women of ages >20 y with histologically confirmed epithelial ovarian cancer living in the Upper Midwest and enrolled within 1 y of diagnosis. Controls without ovarian cancer and who had at least one intact ovary were recruited from among those seen for general medical examinations and frequency matched to cases on age and region of residence. At Duke University, cases were women between ages of 20 and 74 y with histologically confirmed primary epithelial ovarian cancer identified using the North Carolina Central Cancer Registry rapid case ascertainment system. Controls without ovarian cancer and who had at least one intact ovary were identified from the same 48-county region as the cases using list-assisted random digit dialing and frequency matched to cases on race and age. DNA was extracted from blood using the Genra AutoPure LS Purgene salting out methods (Genra); for Duke University participants, DNA was whole-genome amplified with the REPLI-G protocol

Protein	Role	Gene
pRb	Tumor suppressor protein; inhibits cell cycle progression when active	<i>RB1</i>
cyclin D1	Regulatory protein that forms complex with cyclin-dependent kinase (CDK) 4; primary role in progression to G ₁ restriction point	<i>CCND1</i>
cyclin D2	Regulatory protein that forms complex with CDK 4	<i>CCND2</i>
cyclin D3	Regulatory protein that forms complex with CDK 4	<i>CCND3</i>
cyclin E1	Regulatory protein that forms complex with CDK 2; necessary for cell to transition from G ₁ to S phase	<i>CCNE1</i>
CDK 2	Forms complex with cyclin E1; cyclin E1/CDK 2 complex formation activates CDK 2	<i>CDK2</i>
CDK 4	Forms complex with cyclin Ds; cyclin D/CDK 4 complex formation activates CDK 4	<i>CDK4</i>
p21	Inhibits CDK 2 and CDK 4	<i>CDKN1A</i>
p27	Primary inhibitor of cyclin D/CDK 4 and cyclin E1/CDK 2 complexes	<i>CDKN1B</i>
p16-p15	Inhibit CDK 4	<i>CDKN2A-CDKN2B</i>
p18	Inhibits CDK 4	<i>CDKN2C</i>
p19	Inhibits CDK 4	<i>CDKN2D</i>

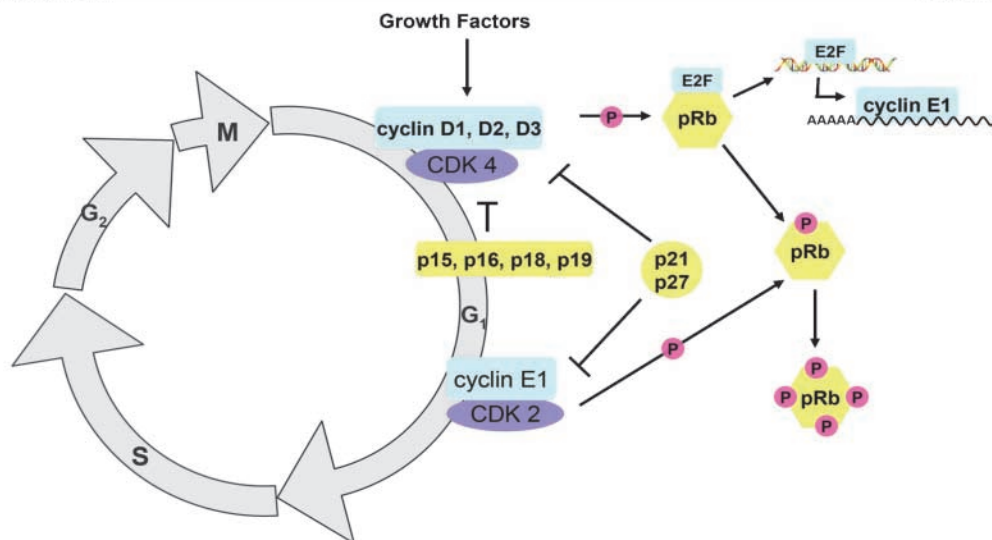


Figure 1. Cell cycle control. In response to growth factor signaling, cyclin D1 (also cyclins D2 and D3) forms a complex with CDK4. This active complex phosphorylates the tumor suppressor protein pRb, inactivating it and allowing the cell cycle to continue. Once freed from pRb, the transcription factor E2F initiates gene expression of cyclin E1. Cyclin E1 forms a complex with CDK2; this complex phosphorylates and inactivates p27 and phosphorylates pRb to continue its inactivation, which pushes the cell cycle past the G₁ restriction point into S phase. Progression of the cell cycle is regulated by inhibitors; p15, p16, p18, and p19 inhibit CDK4 and p21 inhibits both CDK2 and CDK4. These inhibitors must be phosphorylated and degraded before the cell cycle can progress to the next phase.

Table 1. Cell cycle genes

Gene	Chr.	Start (bp)	Size (kb)	No. of SNPs			
				MAY + NCO*	SEA + MAL + STA [†]	CEU [‡]	NIEHS SNPs [§]
<i>CDKN2C</i>	1	51,206,196	6.7	1	2	7	19
<i>CDKN1A</i>	6	36,754,465	8.6	4	11	40	42
<i>CCND3</i>	6	42,010,649	113.8	7	6	87	32
<i>CDKN2A-CDKN2B</i>	9	21,957,751	41.6	9	17	76	96
<i>CCND1</i>	11	69,165,054	13.4	5	7	16	53
<i>CCND2</i>	12	4,253,199	31.6	4	14	83	102
<i>CDKN1B</i>	12	12,761,576	5.0	8	8	17	15
<i>CDK2</i>	12	54,646,826	6.0	7	2	23	24
<i>CDK4</i>	12	56,428,270	4.1	3	2	11	26
<i>RB1</i>	13	47,775,884	178.1	8	11	171	197
<i>CDKN2D</i>	19	10,538,138	2.5	2	2	15	5
<i>CCNE1</i>	19	34,994,741	12.3	2	4	19	48
				60	86	565	659

*Source for tagging SNPs was HapMap release 20.

[†] Source for tagging SNPs was NIEHS SNPs, except *CCND2* and *CDKN1B*, which used HapMap, and *CDKN2C*, which used both HapMap and NIEHS SNPs (October 2005); *CCND3* rs1051130 was excluded due to Hardy-Weinberg equilibrium $P < 0.001$.

[‡] HapMap release 21a, within 10 kb of gene start or stop.

[§] September 2007.

(Qiagen, Inc.; ref. 29). Non-White and Hispanic participants and cases with borderline tumors were excluded from the analysis (one NCO case with unknown race was assumed to be White non-Hispanic, and one NCO case with unknown tumor behavior was assumed to be invasive); additional details are provided elsewhere (30).

The second genotyping collaboration (17, 27) used cases and controls from three different studies: the SEARCH ovarian cancer study from East Anglia, United Kingdom (SEA), the MALOVA cancer study from Denmark (MAL), and the GEOCS study from Stanford University in Palo Alto, California (STA). The SEARCH ovarian cancer study included invasive epithelial ovarian cancer cases collected from the East Anglian and West Midlands cancer registries and controls randomly selected from the European Prospective Investigation into Cancer and Nutrition (EPIC)-Norfolk cohort study. The MALOVA study contained invasive ovarian cancer cases and population controls randomly drawn from a defined study area in Denmark. The GEOCS study ascertained participants from six counties in northern California including invasive ovarian cancer cases and age-matched controls obtained using random-digit dialing. Non-White and Hispanic participants and cases with borderline tumors were excluded from the analysis (33 SEA cases and 1 SEA control with unknown race were assumed to be White non-Hispanic, and 75 SEA cases with unknown tumor behavior were assumed to be invasive); additional study participant details are provided elsewhere (31, 32).

SNP Selection. The first collaboration (MAY + NCO) identified tagSNPs within 5 kb of each gene using the algorithm of *ldselect* (33) to bin pairwise-correlated SNPs at $r^2 \geq 0.80$ with minor allele frequency (MAF) ≥ 0.05 among 60 unrelated Utah residents with Northern and Western European ancestry (CEU) genotyped as part of the international HapMap Consortium release 20 (HapMap, mapped to NCBI build 35; ref. 34). Within linkage disequilibrium bins, tagSNPs with the maximum Illumina-provided SNP_Score were selected. In addition to tagSNPs, putative-functional SNPs were included (within 1 kb upstream, 5' untranslated region, 3'

untranslated region, or nonsynonymous) with MAF ≥ 0.05 identified in Ensembl version 34 and Illumina-provided SNP_Score > 0.6 . Sixty SNPs were selected.

The second collaboration (SEA + MAL + STA) used the multimer tagging algorithm of *Tagger* (35) to bin SNPs pairwise-correlated or correlated with combinations of SNPs with MAF ≥ 0.05 and $R_s^2 \geq 0.80$ (36). CEU data from HapMap (October 2005) were used, as well as resequencing data when available (October 2005), from the National Institute of Environmental Health Sciences (NIEHS) SNPs Program (37). Analysis of NIEHS SNPs used 62 individuals thought to have the least amount of African ancestry from a panel of 90 individuals (PDR90); additional information is provided elsewhere (17). Eighty-seven SNPs were selected.

Genotyping. For MAY + NCO, genotyping of 1,086 genomic and 1,282 whole-genome amplified DNA samples (total of 2,368 including duplicates and laboratory controls) on 2,051 unique study participants was done at Mayo Clinic using the Illumina GoldenGate BeadArray assay and *BeadStudio* software for automated genotype clustering and calling according to standard protocol (38). Samples with call rates $< 90\%$ and SNPs with call rates $< 95\%$ were excluded. Of 2,051 participants genotyped, 10 were later found to be ineligible and were excluded, and 74 samples failed. Among SNPs with an overall call rate $\geq 95\%$, concordance was 99.99% between duplicates of genomic DNA, 99.97% between duplicates of whole-genome amplified DNA, and 99.16% between genomic and whole-genome amplified DNA, indicating adequate genotyping of whole-genome amplified DNA (29).

SEA + MAL + STA samples were genotyped using the TaqMan 7900HT Sequence Detection System according to the manufacturer's instructions. Each assay was carried out using 10 ng of DNA in a 5- μ L reaction with TaqMan Universal PCR Master Mix (Applied Biosystems), forward and reverse primers, and FAM- and VIC-labeled probes designed by Applied Biosystems (ABI Assay-by-Design). Primer and probe sequences and assay conditions used for each polymorphism analyzed are available on request. All assays were carried out in 384-well arrays with 12 duplicate samples in each

Table 2. Characteristics of 5,502 White non-Hispanic invasive cases and controls

	MAY		NCO		MAL		SEA		STA	
	Case (n = 287)	Control (n = 462)	Case (n = 382)	Control (n = 479)	Case (n = 447)	Control (n = 1,221)	Case (n = 717)	Control (n = 852)	Case (n = 287)	Control (n = 368)
Age, y										
Mean (SD)	61.6 (12.5)	60.0 (13.0)	56.8 (10.7)	54.8 (12.0)	59.9 (10.6)	56.8 (11.5)	55.7 (10.0)	52.7 (8.3)	51.2 (8.7)	48.3 (10.3)
Age quartile, n (%)										
≤46 y	30 (11)	69 (15)	68 (18)	123 (26)	53 (12)	243 (20)	112 (16)	202 (24)	82 (29)	137 (37)
47-53 y	52 (18)	78 (17)	82 (22)	118 (25)	83 (19)	293 (24)	178 (25)	323 (38)	76 (27)	105 (29)
54-62 y	70 (24)	113 (25)	108 (28)	80 (17)	114 (26)	247 (20)	236 (33)	205 (24)	110 (38)	104 (28)
63+ y	135 (47)	202 (44)	124 (33)	158 (33)	197 (44)	438 (36)	191 (27)	122 (14)	19 (7)	22 (6)
Age at menarche, y										
Mean (SD)	12.8 (1.53)	13.2 (4.47)	12.5 (1.45)	12.7 (1.41)	13.6 (1.69)	13.6 (1.63)	12.9 (1.95)	12.9 (1.52)	12.5 (1.44)	12.8 (1.54)
Menopause status, n (%)										
Pre/peri	70 (24)	108 (23)	73 (19)	154 (32)	74 (17)	373 (31)	240 (34)	557 (65)	14 (5)	182 (50)
Post	209 (73)	327 (71)	273 (72)	316 (66)	280 (63)	848 (70)	414 (58)	294 (35)	147 (51)	129 (35)
Unknown	8 (3)	27 (5.9)	36 (9)	9 (2)	93 (21)	0 (0)	63 (9)	1 (<1)	126 (44)	57 (16)
Ever used oral contraceptives, n (%)										
Yes	133 (46)	261 (57)	240 (63)	328 (69)	146 (33)	686 (56)	297 (41)	580 (68)	209 (73)	311 (85)
No	139 (48)	164 (36)	131 (34)	147 (31)	207 (46)	535 (44)	399 (56)	271 (32)	76 (27)	55 (15)
Unknown	15 (5)	37 (8)	11 (3)	4 (<1)	94 (21)	0 (0)	21 (3)	1 (<1)	2 (<1)	2 (<1)
No. of live births, n (%)										
0	47 (16)	64 (14)	77 (20)	62 (13)	54 (12)	72 (6)	117 (16)	111 (13)	70 (24)	59 (16)
1-2	97 (34)	153 (33)	185 (48)	268 (56)	157 (35)	510 (42)	343 (48)	387 (45)	111 (39)	156 (42)
3+	137 (48)	218 (47)	116 (31)	149 (31)	143 (32)	639 (52)	223 (31)	353 (4)	106 (37)	151 (41)
Unknown	6 (2)	27 (6)	4 (1)	0 (0)	93 (21)	0 (0)	34 (5)	1 (0.1)	0 (0)	2 (<1)
Histology, n (%)										
Serous	168 (59)	—	234 (61)	—	275 (62)	—	254 (35)	—	159 (55)	—
Endometrioid	58 (20)	—	58 (15)	—	56 (13)	—	129 (18)	—	44 (15)	—
Mucinous	9 (3)	—	22 (6)	—	43 (10)	—	97 (14)	—	24 (8)	—
Clear cell	18 (6)	—	32 (8)	—	33 (7)	—	62 (9)	—	22 (8)	—
Other/unknown epithelial*	43 (15)	—	67 (18)	—	40 (9)	—	175 (24)	—	38 (13)	—

*Includes papillary, undifferentiated, mixed histologies, and other unknown epithelial adenocarcinomas.

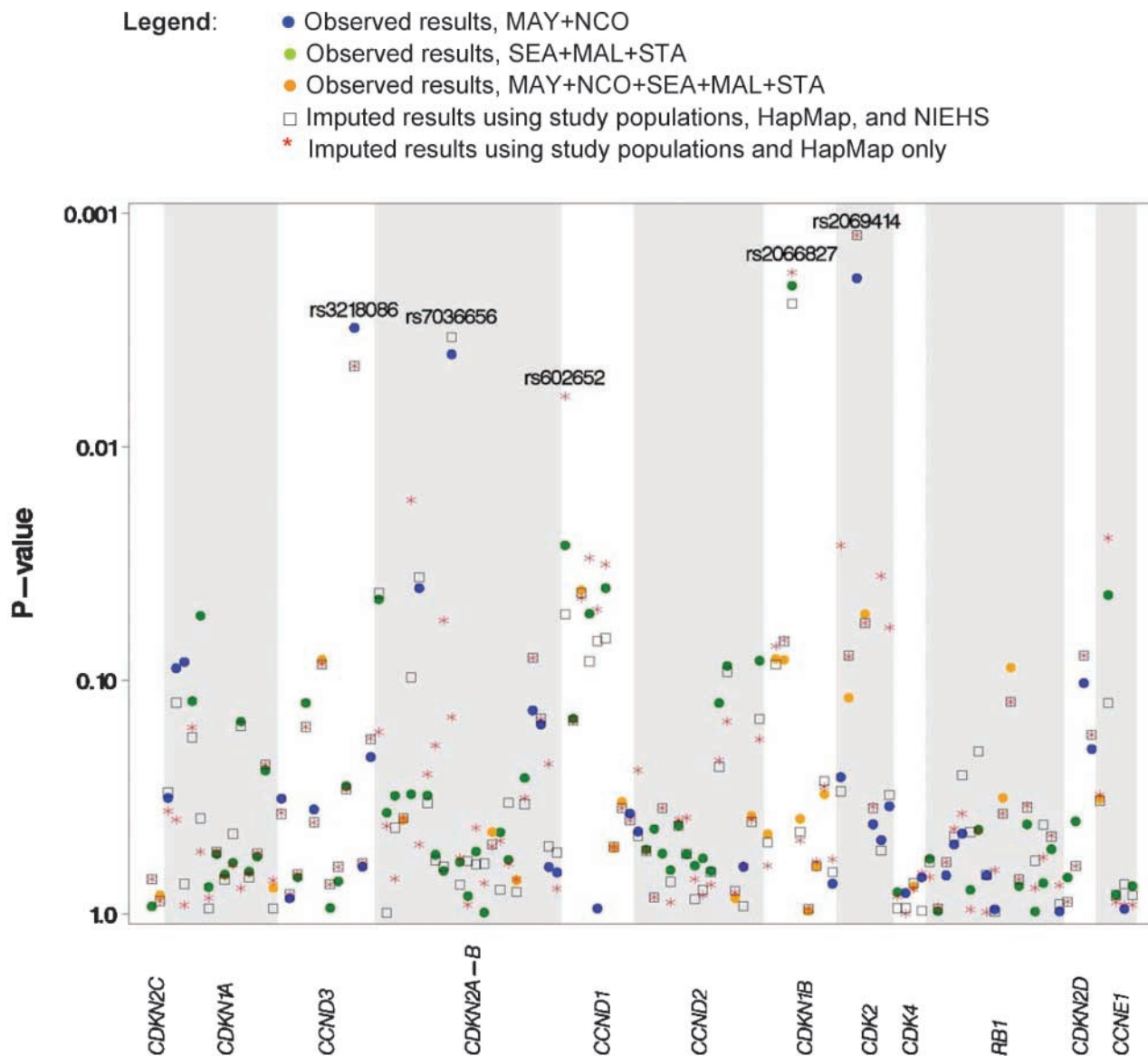


Figure 2. Significance of ordinal ORs using observed and imputed data. Ordinal P values at each SNP represented within gene in chromosome order; not to physical scale.

plate for quality control. Genotypes were determined using Allelic Discrimination Sequence Detection software (Applied Biosystems). Call rates ranged from 94.5% to 99.5% for all the studies and SNPs, and overall concordance between duplicate samples was >99% (17, 27).

Other Data Sources and Harmonization of Alleles.

To impute missing study participant genotypes, we used data from study participants as well as updated data from the sources originally used to identify tagging SNPs: 60 unrelated CEU individuals in HapMap version 21a⁹ (SNPs within 10 kb of each gene using genome build 36.3) and 62 individuals with minimal evidence of African ancestry from NIEHS SNPs¹⁰ (resequenced

regions September 2007). A total of 911 SNPs were identified from MAY + NCO, SEA + MAL + STA, HapMap CEU, and NIEHS SNPs including 395 SNPs with genotype data from two or more sources.

To verify allele consistency for 395 SNPs with genotype data from two or more sources, we reviewed study-designated allele names and MAFs across sources. We found that, for 270 SNPs (68%), genotypes were easily combined across studies (similar MAF, identical nomenclature); for 112 SNPs (28%), genotypes were combined following an obvious strand reversal for at least one data source (similar MAF, reverse strand nomenclature); and for 4 SNPs (1%), genotypes were clearly inconsistent or of a nonobvious nature (e.g., C>G, A>T, and MAF >0.40) and excluded for at least one source (and data remained for two or more sources). For 5 SNPs (1%), genotypes that were clearly inconsistent

⁹ <http://hapmap.org/downloads/genotypes/2007-01/>

¹⁰ http://egp.gs.washington.edu/finished_genes.html

Table 3. Cell cycle SNPs and risk of ovarian cancer

Gene	SNP	Distance (bp)	OR (95% CI)					
			MAY + NCO	SEA + MAL + STA	Combined	Combined imputed: HapMap + NIEHS	Combined imputed: HapMap only	
CDKN2C	rs3176459	—	—	1.00 (0.90-1.10)	1.00 (0.90-1.10)	0.98 (0.89-1.08)	0.98 (0.89-1.08)	
	rs12855	2,846	1.02 (0.80-1.30)	1.01 (0.86-1.19)	1.01 (0.89-1.16)	1.01 (0.88-1.15)	1.01 (0.88-1.15)	
CDKN1A	rs1977172	—	1.11 (0.90-1.38)	—	1.11 (0.90-1.38)	1.12 (0.90-1.39)	1.09 (0.91-1.31)	
	rs3829963	2,632	1.19 (0.97-1.46)	—	1.19 (0.97-1.46)	1.17 (0.96-1.42)	1.06 (0.92-1.22)	
	rs733590	817	0.87 (0.75-1.02)	—	0.87 (0.75-1.02)	0.98 (0.87-1.10)	1.00 (0.93-1.08)	
	rs762624	385	—	1.09 (0.98-1.21)	1.09 (0.98-1.21)	1.08 (0.97-1.20)	1.08 (0.97-1.20)	
	rs2395655	108	—	1.10 (1.00-1.21)	1.10 (1.00-1.21)	1.04 (0.95-1.13)	1.03 (0.95-1.11)	
	rs3176331	1,829	—	1.02 (0.89-1.18)	1.02 (0.89-1.18)	1.00 (0.87-1.16)	1.01 (0.88-1.17)	
	rs3176336	1,291	—	0.97 (0.88-1.07)	0.97 (0.88-1.07)	0.97 (0.88-1.07)	0.97 (0.88-1.07)	
	rs3176343	1,451	—	1.05 (0.84-1.30)	1.05 (0.84-1.30)	1.04 (0.84-1.29)	1.04 (0.84-1.29)	
	rs1801270	1,704	—	0.95 (0.78-1.15)	0.95 (0.78-1.15)	0.93 (0.77-1.13)	0.95 (0.79-1.15)	
	rs3176352	368	—	1.08 (0.97-1.20)	1.08 (0.97-1.20)	1.08 (0.97-1.20)	1.01 (0.92-1.11)	
	rs1059234	1,258	—	0.96 (0.79-1.16)	0.96 (0.79-1.16)	0.96 (0.79-1.17)	0.96 (0.79-1.16)	
	rs6457937	754	—	0.92 (0.68-1.24)	0.92 (0.68-1.24)	0.91 (0.68-1.23)	0.91 (0.68-1.23)	
	rs3176359	391	—	1.61 (0.72-3.56)	1.61 (0.72-3.56)	1.63 (0.73-3.61)	1.63 (0.73-3.61)	
	rs7767246	4,473	0.82 (0.69-0.99)	1.11 (0.98-1.25)	1.01 (0.92-1.12)	1.00 (0.91-1.11)	1.02 (0.92-1.12)	
	CCND3	rs2479726	—	0.92 (0.79-1.08)	—	0.92 (0.79-1.08)	0.93 (0.80-1.09)	0.93 (0.80-1.09)
		rs3828855	2,310	0.98 (0.76-1.25)	—	0.98 (0.76-1.25)	0.97 (0.76-1.25)	0.97 (0.76-1.25)
		rs3218114	1,946	—	0.98 (0.86-1.10)	0.98 (0.86-1.10)	0.97 (0.86-1.10)	0.97 (0.86-1.10)
rs3218110		157	—	1.09 (0.98-1.22)	1.09 (0.98-1.22)	1.08 (0.97-1.21)	1.08 (0.97-1.21)	
rs3218108		146	0.93 (0.79-1.09)	—	0.93 (0.79-1.09)	0.93 (0.79-1.10)	0.93 (0.79-1.10)	
rs9529		352	0.91 (0.78-1.07)	0.93 (0.84-1.03)	0.93 (0.85-1.01)	0.93 (0.85-1.01)	0.93 (0.85-1.01)	
rs2479717		2,167	—	1.00 (0.90-1.12)	1.00 (0.90-1.12)	1.02 (0.92-1.13)	1.02 (0.92-1.13)	
rs3218092		1,487	—	0.98 (0.87-1.11)	0.98 (0.87-1.11)	0.97 (0.86-1.10)	0.97 (0.86-1.10)	
rs1410492		1,194	—	1.06 (0.95-1.18)	1.06 (0.95-1.18)	1.06 (0.95-1.18)	1.06 (0.95-1.18)	
rs3218086		2,209	1.32 (1.10-1.58)	—	1.32 (1.10-1.58)	1.31 (1.09-1.57)	1.31 (1.09-1.57)	
rs3218085		115	1.17 (0.63-2.17)	—	1.17 (0.63-2.17)	1.18 (0.63-2.20)	1.18 (0.63-2.20)	
rs9381100		1,006	0.90 (0.76-1.06)	—	0.90 (0.76-1.06)	0.89 (0.75-1.05)	0.89 (0.75-1.05)	
CDKN2A—B		rs3731257	—	—	0.89 (0.80-1.00)	0.89 (0.80-1.00)	0.89 (0.80-1.00)	0.93 (0.85-1.03)
		rs3088440	1,938	—	1.08 (0.91-1.28)	1.08 (0.91-1.28)	1.00 (0.86-1.16)	1.07 (0.91-1.26)
		rs11515	40	—	1.07 (0.94-1.23)	1.07 (0.94-1.23)	1.06 (0.92-1.21)	0.98 (0.87-1.10)
		rs3731249	2,717	0.92 (0.62-1.35)	0.89 (0.67-1.19)	0.91 (0.72-1.14)	0.90 (0.72-1.14)	0.90 (0.72-1.14)
		rs3731239	3,302	—	1.05 (0.95-1.16)	1.05 (0.95-1.16)	1.08 (0.99-1.18)	1.11 (1.02-1.20)
	rs2811709	5,933	0.80 (0.64-0.99)	—	0.80 (0.64-0.99)	0.79 (0.64-0.99)	0.96 (0.85-1.08)	
	rs4074785	1,432	—	1.09 (0.92-1.28)	1.09 (0.92-1.28)	1.08 (0.92-1.28)	1.10 (0.93-1.29)	
	rs3731222	2,331	—	0.96 (0.83-1.10)	0.96 (0.83-1.10)	0.96 (0.84-1.11)	0.92 (0.82-1.04)	
	rs3731211	2,933	—	0.98 (0.88-1.09)	0.98 (0.88-1.09)	0.97 (0.88-1.08)	0.92 (0.84-1.00)	
	rs7036656	3,610	0.79 (0.67-0.93)	—	0.79 (0.67-0.93)	0.79 (0.67-0.92)	0.94 (0.86-1.02)	
	rs3731197	914	—	1.03 (0.93-1.13)	1.03 (0.93-1.13)	1.02 (0.93-1.11)	1.03 (0.93-1.13)	
	rs3218020	6,501	—	0.99 (0.89-1.10)	0.99 (0.89-1.10)	0.98 (0.89-1.07)	1.00 (0.91-1.09)	
	rs2811712	163	—	1.05 (0.90-1.22)	1.05 (0.90-1.22)	1.04 (0.89-1.21)	0.95 (0.84-1.08)	
	rs3218012	625	—	1.00 (0.91-1.10)	1.00 (0.91-1.10)	0.98 (0.90-1.06)	0.98 (0.89-1.08)	
	rs3218009	97	1.19 (0.96-1.46)	0.99 (0.86-1.14)	1.05 (0.93-1.17)	1.04 (0.93-1.17)	1.04 (0.93-1.17)	
	rs3218005	1,490	—	1.06 (0.91-1.25)	1.06 (0.91-1.25)	1.02 (0.88-1.18)	0.95 (0.84-1.09)	
	rs3217992	2,976	—	0.97 (0.88-1.08)	0.97 (0.88-1.08)	0.96 (0.87-1.05)	0.98 (0.90-1.06)	
rs1063192	144	1.09 (0.94-1.25)	0.98 (0.89-1.08)	1.01 (0.94-1.10)	1.01 (0.93-1.09)	1.02 (0.94-1.10)		
rs3217986	1,963	—	1.10 (0.93-1.29)	1.10 (0.93-1.29)	1.08 (0.92-1.27)	1.09 (0.92-1.28)		
rs2069418	4,368	1.11 (0.97-1.28)	—	1.11 (0.97-1.28)	1.13 (0.98-1.31)	1.13 (0.98-1.31)		
rs575427	1,779	0.84 (0.66-1.07)	—	0.84 (0.66-1.07)	0.83 (0.65-1.07)	0.91 (0.79-1.04)		
rs13298881	574	0.94 (0.75-1.19)	—	0.94 (0.75-1.19)	0.92 (0.73-1.17)	1.09 (0.95-1.25)		
rs10811640	1,360	0.97 (0.84-1.12)	—	0.97 (0.84-1.12)	0.96 (0.83-1.10)	0.99 (0.91-1.07)		
CCND1	rs602652	—	—	1.11 (1.01-1.23)	1.11 (1.01-1.23)	1.08 (1.00-1.17)	1.14 (1.04-1.25)	
	rs3862792	214	—	0.79 (0.58-1.09)	0.79 (0.58-1.09)	0.79 (0.58-1.09)	0.79 (0.58-1.09)	
	rs603965	54	1.01 (0.88-1.17)	1.12 (1.02-1.23)	1.09 (1.00-1.17)	1.08 (1.00-1.17)	1.08 (1.00-1.17)	
	rs3212879	571	—	0.91 (0.83-1.00)	0.91 (0.83-1.00)	0.93 (0.86-1.01)	0.90 (0.82-0.99)	
	rs649392	1,312	1.00 (0.87-1.14)	—	1.00 (0.87-1.14)	0.93 (0.86-1.01)	0.92 (0.85-1.00)	
	rs3212891	714	—	0.90 (0.82-1.00)	0.90 (0.82-1.00)	0.93 (0.86-1.00)	0.90 (0.82-0.99)	
	rs678653	1,230	1.01 (0.87-1.17)	0.96 (0.87-1.06)	0.97 (0.90-1.06)	0.97 (0.90-1.06)	0.97 (0.90-1.06)	
	rs7178	2,293	0.83 (0.64-1.09)	1.21 (1.02-1.45)	1.08 (0.93-1.24)	1.07 (0.93-1.24)	1.07 (0.93-1.24)	
	rs11603541	3,343	0.90 (0.72-1.13)	—	0.90 (0.72-1.13)	0.90 (0.72-1.14)	0.91 (0.72-1.14)	
	rs1049606	—	1.06 (0.92-1.22)	—	1.06 (0.92-1.22)	1.04 (0.94-1.15)	1.08 (0.95-1.22)	
CCND2	rs3217795	3,028	—	0.95 (0.80-1.12)	0.95 (0.80-1.12)	0.95 (0.80-1.12)	0.95 (0.80-1.12)	
	rs3217805	2,020	—	0.96 (0.87-1.06)	0.96 (0.87-1.06)	1.01 (0.92-1.11)	1.01 (0.92-1.11)	
	rs3217820	2,288	—	1.03 (0.93-1.14)	1.03 (0.93-1.14)	0.95 (0.87-1.05)	0.95 (0.87-1.05)	
	rs3217852	7,394	—	0.97 (0.87-1.09)	0.97 (0.87-1.09)	0.98 (0.88-1.10)	0.99 (0.89-1.10)	
	rs3217862	1,321	—	0.95 (0.83-1.08)	0.95 (0.83-1.08)	0.95 (0.84-1.08)	0.95 (0.83-1.07)	
	rs3217863	409	—	1.05 (0.88-1.26)	1.05 (0.88-1.26)	1.05 (0.89-1.25)	1.08 (0.91-1.28)	

(Continued on the following page)

Table 3. Cell cycle SNPs and risk of ovarian cancer (Cont'd)

Gene	SNP	Distance (bp)	OR (95% CI)				
			MAY + NCO	SEA + MAL + STA	Combined	Combined imputed: HapMap + NIEHS	Combined imputed: HapMap only
	rs3217901	5,419	—	1.03 (0.93-1.13)	1.03 (0.93-1.13)	1.01 (0.93-1.10)	1.01 (0.93-1.10)
	rs3217906	415	—	1.03 (0.92-1.14)	1.03 (0.92-1.14)	1.02 (0.92-1.14)	1.02 (0.92-1.13)
	rs3217916	2,869	—	0.92 (0.83-1.02)	0.92 (0.83-1.02)	0.94 (0.85-1.04)	0.95 (0.87-1.03)
	rs3217925	2,966	—	0.91 (0.81-1.01)	0.91 (0.81-1.01)	0.92 (0.83-1.01)	0.94 (0.85-1.02)
	rs3217926	44	1.03 (0.89-1.20)	0.98 (0.89-1.08)	0.99 (0.92-1.08)	0.99 (0.91-1.07)	0.99 (0.91-1.07)
	rs1049612	1,079	1.04 (0.90-1.20)	—	1.04 (0.90-1.20)	0.99 (0.88-1.12)	1.03 (0.93-1.13)
	rs3217933	238	0.95 (0.80-1.12)	1.09 (0.97-1.21)	1.04 (0.95-1.14)	1.04 (0.95-1.14)	1.04 (0.95-1.14)
	rs3217936	1,952	—	0.91 (0.82-1.01)	0.91 (0.82-1.01)	0.94 (0.85-1.02)	0.94 (0.87-1.03)
CDKN1B	rs3759216	—	1.05 (0.91-1.21)	1.02 (0.93-1.13)	1.03 (0.95-1.12)	1.03 (0.95-1.11)	1.02 (0.94-1.10)
	rs3759217	366	1.06 (0.86-1.31)	1.14 (0.99-1.32)	1.11 (0.99-1.26)	1.11 (0.99-1.25)	1.12 (0.99-1.26)
	rs34330	2,243	0.94 (0.80-1.11)	0.91 (0.81-1.02)	0.92 (0.84-1.01)	0.92 (0.84-1.01)	0.92 (0.84-1.01)
	rs2066827	404	—	0.84 (0.75-0.94)	0.84 (0.75-0.94)	0.84 (0.76-0.94)	0.85 (0.77-0.94)
	rs34329	2,134	1.12 (0.96-1.30)	1.00 (0.90-1.11)	1.04 (0.95-1.13)	1.03 (0.95-1.13)	1.03 (0.95-1.12)
	rs3093736	68	1.07 (0.72-1.59)	0.97 (0.75-1.26)	1.00 (0.80-1.24)	0.99 (0.80-1.23)	0.99 (0.80-1.23)
	rs7330	1,616	1.05 (0.91-1.21)	1.01 (0.92-1.11)	1.02 (0.94-1.10)	1.02 (0.94-1.10)	1.02 (0.94-1.11)
	rs1420023	1,194	0.84 (0.68-1.05)	0.98 (0.84-1.14)	0.94 (0.83-1.06)	0.93 (0.82-1.06)	0.93 (0.82-1.06)
	rs34322	3,459	0.98 (0.85-1.13)	—	0.98 (0.85-1.13)	0.97 (0.84-1.12)	1.02 (0.94-1.11)
CDK2	rs2069391	—	1.16 (0.89-1.52)	—	1.16 (0.89-1.52)	1.15 (0.88-1.50)	1.21 (1.02-1.43)
	rs2069408	4,443	0.94 (0.80-1.09)	0.94 (0.84-1.04)	0.94 (0.86-1.02)	0.93 (0.85-1.01)	0.93 (0.85-1.01)
	rs2069414	1,378	1.55 (1.18-2.04)	—	1.55 (1.18-2.04)	1.58 (1.20-2.09)	1.58 (1.20-2.09)
	rs1045435	461	1.17 (0.90-1.53)	1.14 (0.97-1.34)	1.15 (1.00-1.32)	1.14 (1.00-1.31)	1.14 (1.00-1.31)
	rs11171710	1,918	1.06 (0.92-1.22)	—	1.06 (0.92-1.22)	1.07 (0.93-1.23)	1.05 (0.95-1.17)
	rs17528736	440	1.15 (0.78-1.70)	—	1.15 (0.78-1.70)	1.13 (0.77-1.67)	1.23 (1.01-1.48)
	rs773108	1,393	0.93 (0.80-1.08)	—	0.93 (0.80-1.08)	0.92 (0.79-1.08)	0.92 (0.85-1.00)
CDK4	rs2069506	—	—	0.99 (0.89-1.09)	0.99 (0.89-1.09)	1.00 (0.92-1.08)	0.99 (0.90-1.09)
	rs2069502	1,811	1.02 (0.88-1.18)	—	1.02 (0.88-1.18)	1.00 (0.92-1.08)	1.00 (0.89-1.12)
	rs2270777	491	0.90 (0.78-1.03)	1.03 (0.93-1.13)	0.99 (0.91-1.07)	0.99 (0.91-1.07)	0.99 (0.91-1.07)
	rs2072052	1,563	1.03 (0.89-1.19)	—	1.03 (0.89-1.19)	1.00 (0.92-1.08)	1.03 (0.89-1.20)
RB1	rs1981434	—	—	0.97 (0.87-1.08)	0.97 (0.87-1.08)	0.97 (0.88-1.08)	0.98 (0.90-1.07)
	rs2854345	10,082	—	1.00 (0.89-1.12)	1.00 (0.89-1.12)	1.00 (0.88-1.12)	1.00 (0.88-1.12)
	rs4151467	28,687	1.07 (0.78-1.45)	—	1.07 (0.78-1.45)	1.09 (0.80-1.48)	1.09 (0.80-1.48)
	rs7329938	12,056	1.08 (0.87-1.33)	—	1.08 (0.87-1.33)	1.08 (0.88-1.33)	1.09 (0.88-1.35)
	rs4151510	13,196	1.08 (0.88-1.34)	—	1.08 (0.88-1.34)	1.08 (0.95-1.24)	1.10 (0.89-1.36)
	rs399413	3,394	—	1.02 (0.91-1.14)	1.02 (0.91-1.14)	0.96 (0.88-1.06)	1.00 (0.91-1.09)
	rs4151540	7,091	—	0.96 (0.86-1.07)	0.96 (0.86-1.07)	0.94 (0.85-1.03)	0.96 (0.86-1.07)
	rs9568036	16,276	1.03 (0.89-1.19)	—	1.03 (0.89-1.19)	1.03 (0.89-1.19)	1.00 (0.91-1.10)
	rs198604	12,127	1.00 (0.84-1.17)	—	1.00 (0.84-1.17)	1.00 (0.85-1.18)	0.98 (0.89-1.07)
	rs4151551	1,376	1.10 (0.85-1.41)	1.06 (0.90-1.25)	1.07 (0.93-1.23)	1.06 (0.93-1.22)	1.06 (0.93-1.22)
	rs2854344	12,254	0.97 (0.73-1.28)	0.81 (0.66-0.99)	0.87 (0.74-1.02)	0.88 (0.75-1.04)	0.88 (0.75-1.04)
	rs425834	14,800	—	1.04 (0.80-1.36)	1.04 (0.80-1.36)	1.05 (0.81-1.37)	1.05 (0.81-1.37)
	rs4151611	35,438	—	0.91 (0.72-1.14)	0.91 (0.72-1.14)	0.90 (0.71-1.13)	0.89 (0.71-1.13)
	rs4151620	1,128	—	1.00 (0.85-1.17)	1.00 (0.85-1.17)	1.04 (0.90-1.20)	1.02 (0.87-1.20)
	rs3092904	2,422	—	0.98 (0.87-1.10)	0.98 (0.87-1.10)	0.96 (0.86-1.07)	0.97 (0.89-1.07)
	rs4151636	5,253	—	0.93 (0.74-1.17)	0.93 (0.74-1.17)	0.92 (0.73-1.16)	0.92 (0.73-1.15)
	rs990814	2,843	1.00 (0.85-1.17)	—	1.00 (0.85-1.17)	1.01 (0.86-1.18)	0.98 (0.90-1.08)
CDKN2D	rs3218222	—	—	1.02 (0.91-1.14)	1.02 (0.91-1.14)	1.01 (0.91-1.12)	1.01 (0.91-1.12)
	rs1465702	1,951	—	1.10 (0.88-1.39)	1.10 (0.88-1.39)	1.05 (0.86-1.29)	1.05 (0.86-1.29)
	rs1465701	210	1.15 (0.97-1.35)	—	1.15 (0.97-1.35)	1.16 (0.98-1.36)	1.16 (0.98-1.36)
	rs17677316	1,399	0.90 (0.76-1.06)	—	0.90 (0.76-1.06)	0.89 (0.75-1.05)	0.89 (0.75-1.05)
CCNE1	rs997669	—	0.97 (0.84-1.13)	1.07 (0.97-1.18)	1.04 (0.96-1.13)	1.04 (0.96-1.13)	1.04 (0.96-1.13)
	rs3218036	1,201	—	1.11 (1.00-1.23)	1.11 (1.00-1.23)	1.07 (0.98-1.17)	1.12 (1.01-1.24)
	rs3218038	211	—	1.03 (0.80-1.33)	1.03 (0.80-1.33)	1.02 (0.84-1.24)	1.02 (0.79-1.32)
	rs1406	9,217	0.99 (0.84-1.18)	—	0.99 (0.84-1.18)	0.98 (0.89-1.08)	1.00 (0.92-1.10)
	rs3218076	158	—	1.02 (0.91-1.13)	1.02 (0.91-1.13)	1.01 (0.92-1.11)	1.00 (0.92-1.10)

NOTE: Bold emphasis indicates $P < 0.05$.

or of a nonobvious nature were excluded for at least one source and only one source remained (thus not requiring allele harmonizing), and for 4 SNPs (1%), genotypes that were clearly inconsistent or of a nonobvious nature were excluded for all sources and not used in analyses. Thus, a resulting 391 harmonized SNPs were merged with 516 SNPs available from only one source. One SNP (CCND3 rs1051130) was then excluded due to Hardy-Weinberg equilibrium $P < 0.001$ in the SEA + MAL + STA controls, leaving 901 SNPs included in the final analytic data set

(122 SNPs genotyped by MAY + NCO or SEA + MAL + STA). SNPs are tallied per gene and per population in Table 1; a complete listing of analyzed SNPs, MAFs, and call rates is provided in Supplementary Table S1.

Statistical Analysis. We ran a series of association analyses using observed data from each collaboration (MAY + NCO and SEA + MAL + STA), observed data combined across both collaborations, and imputed data. To impute missing genotypes, we used a hidden Markov

model as implemented in fastPHASE (39), with 25 iterations and 20 random starts of the EM algorithm. Five runs of fastPHASE were conducted using different random seeds. A logistic regression model was then fit to each of the five imputed data sets for each SNP of interest and the resulting parameter and variance estimates were extracted. Results were combined across imputation runs using standard multiple imputation techniques computing both the within- and between-imputation variations (40). The use of multiple imputation methods allowed us to estimate the variance due to imputation and incorporate this into our overall SNP variance estimates. In general, this imputation-based variance component was small (mean, 1.7×10^{-5}). The variance component was largest for SNPs genotyped among MAY + NCO participants only (mean, 3.7×10^{-5}), slightly smaller for SNPs genotyped as SEA + MAL + STA only (mean, 1.3×10^{-5}), and practically equal to zero for SNPs genotyped in both collaborations (mean, 1.1×10^{-7}). Because of an observed slightly greater MAF discrepancy between study participants and NIEHS SNPs participants than between study participants and HapMap participants, imputations were also carried out excluding NIEHS SNPs data.

Associations between genotypes and ovarian cancer risk were assessed using logistic regression to estimate odds ratios (OR) and 95% confidence intervals (95% CI) assuming an ordinal (log-additive) genotypic effect. For imputation-based analyses, we modeled the observed number of copies of the minor allele for participants with nonmissing genotypes for a given SNP and the estimated most likely number of copies of the minor allele for subjects with imputed genotypes. Association tests were two-sided, adjusted for the potential confounding effects of age and study population (MAY, NCO, SEA, MAL, and STA), and carried out using the SAS software system (SAS Institute, Inc.). Adjustment of *P* values due to multiple testing was not conducted because interpretation was based on relative changes in results due to imputation.

Results

Characteristics of 5,502 study participants (2,210 cases and 3,382 controls) are shown in Table 2. The study populations were generally similar, although MAY participants were older (no upper age limit had been used), STA participants included more oral contraceptive users, and SEA included fewer known serous cases. Using observed data only, four SNPs were associated with risk of invasive ovarian cancer at *P* < 0.01 (Fig. 2): *CCND3* rs3218086 (MAY + NCO; increased risk), *CDKN2A-CDKN2B* rs7036656 (MAY + NCO; decreased risk), *CDKN1B* rs2066827 (SEA + MAL + STA; decreased risk), and *CDK2* rs2069414 (MAY + NCO; increased risk; Table 3). Risk was associated at *P* < 0.05 with two additional SNPs in the MAY + NCO population (*CDKN1A* rs7767246 and *CDKN2A-CDKN2B* rs2811709, both decreased risk) and seven additional SNPs in SEA + MAL + STA (*CDKN2A-CDKN2B* rs3731257, decreased risk; *CCND1* rs602652, rs603695, and rs7178, increased risk; *CCND1* rs321891, decreased risk; *CCNE1* rs3218036, increased risk; and *RB1* rs2854344, decreased risk; Table 3; refs. 17, 27). No SNPs typed in both populations were significant in combined analysis without reaching significance in one of the study populations. Of four

SNPs typed in both populations and significant in only one population, two yielded ORs in opposite directions for null combined ORs (*CDKN1A* rs7767246 and *CCND1* rs7178) and two were significant (*P* < 0.05) only in the larger SEA + MAL + STA study (when combined, *CCND1* rs603965 remained at *P* < 0.05 whereas *RB1* rs2854344 lost significance).

On the whole, imputed results did not differ from results of combined analysis of observed data; *P* values increased by a mean of 0.001 using HapMap + NIEHS and decreased by a mean of 0.001 using HapMap only. For SNPs genotyped in both collaborations, the effect of imputation on results was minimal because only those participants that failed genotyping were affected. For SNPs genotyped in only one collaboration, use of only HapMap led to slightly greater discrepancy between observed and imputed *P* values, more often leading to greater significance, than use of HapMap + NIEHS, which varied *P* values to a lesser degree (Supplementary Fig. S1). For SNPs selected using NIEHS and HapMap, genotyped in SEA + MAL + STA, and imputed for MAY + NCO samples, *P* values from imputation-based analysis increased by a mean of 0.004 using NIEHS + HapMap and a mean of 0.01 using HapMap only. For SNPs selected using HapMap, genotyped in MAY + NCO, and imputed for SEA + MAL + STA samples, *P* values decreased by a mean of 0.01 using NIEHS + HapMap and a mean of 0.03 using HapMap only. Thus, the largest overall difference in results occurred when imputation took place on the largest number of samples (SEA + MAL + STA), and generally results became more significant. For example, the OR for *CDK2* rs2069414 increased from 1.55 (95% CI, 1.18-2.04; *P* = 0.002) in MAY + NCO to an imputed OR of 1.58 (95% CI, 1.20-2.09; *P* = 0.001), and the OR for *CCND1* rs649392 decreased from 1.00 (95% CI, 0.87-1.14; *P* = 0.95) in MAY + NCO to an imputed OR of 0.92 (95% CI, 0.85-1.00; *P* = 0.05; Table 3). In addition, two correlated *CDK2* SNPs, rs2069391 and rs17528736 (MAY + NCO controls, $r^2 = 0.38$; CEU, $r^2 = 0.56$), became significant at *P* < 0.05 when imputed with HapMap data; the OR for rs17528736 increased from 1.15 (95% CI, 0.78-1.70; *P* = 0.48) in MAY + NCO to an imputed OR of 1.23 (95% CI, 1.01-1.48; *P* = 0.04). Both of these SNPs are uncorrelated with *CDK2* rs2069414 (r^2 values < 0.01 in MAY + NCO and HapMap) indicative of independent associations. These results suggest that additional risk alleles in the SEA + MAL + STA population were correlated with genotyped SNPs in MAY + NCO and imputation increased power to detect associations.

Novel SNPs of interest also came to light with imputation of MAY + NCO data, notably *CDKN2A-CDKN2B* rs3731239, which increased from an OR of 1.05 (95% CI, 0.95-1.16; *P* = 0.31) to an OR of 1.11 (95% CI, 1.02-1.20; *P* = 0.02) using HapMap, and *CCND1* rs602652, which increased from an OR of 1.11 (95% CI, 1.01-1.23; *P* = 0.03) to an OR of 1.14 (95% CI, 1.04-1.25; *P* = 0.001) using HapMap (Fig. 2). Additional SNPs in *CCND1*, *CDKN2A-CDKN2B*, *CDK2*, and *CCNE1* became more significant with HapMap-based imputation in MAY + NCO although point estimates remained similar (Table 3; Fig. 2). As above, these results suggest that additional risk alleles in the MAY + NCO population were correlated with genotyped SNPs in SEA + MAL + STA and imputation increased power to detect associations.

CCND1 results warrant particular attention. Linkage disequilibrium patterns were similar across populations (Supplementary Fig. S2). Using NIEHS SNPs data (the study with maximal coverage of the nine SNPs genotyped by either MAY + NCO or SEA + MAL + STA), there was strong correlation between rs3212879, rs649392, and rs3212891 (r^2 values >0.86), rs602652 (r^2 values >0.75), and rs603965 (r^2 values >0.64). Combined analysis of observed study participant data yielded three P values <0.05; two resulted from SEA + MAL + STA data alone and one used data from MAY + NCO as well. Use of imputation with study participant and HapMap data increased the number of significant results from three to five P values <0.05, although HapMap did not genotype two of these SNPs (rs602652 and insertion/deletion polymorphism rs321879; Supplementary Table S1). These findings remind us that the underlying haplotype structure used to impute genotypes relies on all available data, here, a total of 20 SNPs with data from MAY + NCO, SEA + MAL + STA, or HapMap. An additional 39 SNPs were covered by NIEHS SNPs; inclusion of these data attenuated ORs and resulted in only one P value <0.05 (rs603965). Whether these results are closer to the truth, given that NIEHS SNPs participants were only presumed to be White non-Hispanic, remains to be verified by additional genotyping and fine-mapping.

In summary, our imputation-based analysis of SNPs in key cell cycle genes did not reveal novel SNPs worthy of follow-up in *CDKN2C*, *CDKN1A*, *CCND3*, *CCND2*, *CDKN1B*, *CDK4*, *RB1*, or *CDKN2D*, but suggested a handful of SNPs in *CDKN2A-CDKN2B* (rs3731239), *CCND1* (the correlated SNPs rs602652, rs3212879, rs649392, and rs3212891), *CDK2* (rs2069414 and the correlated SNPs rs17528736 and rs2069391), and *CCNE1* (rs3218036), which merit genotyping in the unassayed sample populations.

Discussion

Here, we report on analysis of 2,120 invasive ovarian cancer cases and 3,382 controls of White non-Hispanic ethnicity successfully genotyped on up to 122 SNPs but only 24 SNPs with maximally genotyped participants. Using data on 901 regional SNPs genotyped in study, HapMap or NIEHS SNPs participants, we applied a hidden Markov model to estimate underlying haplotypes and imputed missing genotypes among study participants. Analysis of imputation-based data revealed additional evidence of association with risk of ovarian cancer for SNPs in several genes. In particular, we find that additional genotyping is warranted in the genes encoding p16 and p15 (*CDKN2A-CDKN2B*), shown to be overexpressed and methylated in ovarian cancer, respectively (1, 41); cyclin D1 (*CCND1*), shown to be abnormally expressed in ovarian cancer (42); *CDK2* (*CDK2*), shown to inhibit G_1 arrest in ovarian cancer cells (43); and cyclin E1 (*CCNE1*), which is overexpressed in ovarian cancer (15). Several of the SNPs associated with ovarian cancer risk here have been studied in relation to risk of breast, prostate, lung, bladder, and oral cancers (5-11). Of particular interest is a SNP in the region of *CDKN2A-CDKN1B* rs3731239 that was found to be associated with decreased breast cancer risk (6) and shows a protective association in the current analysis.

Our combined, imputation-based analysis strengthens existing interrogations in which tagging SNPs are typed in

one collaboration and the most suggestive single SNPs are brought to a consortium for replication. For example, based on results from the SEA + MAL + STA collaboration alone, four of the currently assessed SNPs were genotyped in more than 3,500 cases and 5,700 controls by the Ovarian Cancer Association Consortium (*CCND1* rs7178 and rs603965, *CDKN1B* rs2066827, and *CDKN2A-CDKN2B* rs3731257), and *CDKN1B* rs2066827 and *CDKN2A-CDKN2B* rs3731257 remained associated (17). More recently, one of the *RB1* SNPs genotyped in SEA + MAL + STA (rs2854344) was assessed by the Ovarian Cancer Association Consortium using more than 4,600 cases and 8,100 controls and found to replicate, despite null results in MAY + NCO; another SNP in *CDKN2A* (rs2811712) did not replicate (18). Combining multiple tagging SNP studies using imputation when necessary will assist preliminary candidate gene studies by (a) improving power of "phase I" analyses and (b) highlighting specific SNPs to do "fill-in" genotyping before consortium genotyping. Here, data suggest additional SNPs to interrogate in MAY + NCO or SEA + MAL + STA study populations for maximal discriminatory power before selection of SNPs in future large-scale genotyping efforts.

These analyses have the potential strengths of theoretically improved sample size at no additional genotyping cost. However, several caveats are warranted. As with non-imputation-based analysis, the benefit of larger sample size may increase the potential for study heterogeneity. In addition, analyses make similar assumptions as in tagging SNP selection including that the populations used to estimate underlying haplotypes are similar to study populations of interest, an assumption which is not always testable. In the current analysis, ethnicity of NIEHS SNPs participants was genetically inferred, which may be particularly problematic. Here, we also assumed that linkage disequilibrium is similar among cases and controls and across all studies. Analyses also assume that the densely typed population is of sufficient sample size. Violation of these assumptions can impair inference of results. Finally, it is worth noting that merging genotype data across multiple studies and publicly available data requires great effort to harmonize alleles; a conservative approach excluding genotypes that are not easily combined is recommended.

We make modest suggestions for future imputation-based analysis. Imputation of genotypes has typically relied on single imputation; however, this approach ignores the variation in estimation due to the imputation. An accepted alternative is the use of multiple imputation, in which a number of "imputed" data sets are created and then analyzed using standard statistical methods and models (40, 44-47) allowing one to estimate the amount of variation attributable to the imputation procedure. In general, the imputation variance components for our study were small. The tool that we used (fastPHASE) only provides the "most likely genotype" as the imputed value. Multiple imputation based on this most likely genotype may not capture the total amount of variation due to imputation (i.e., if the posterior probability is large for a particular genotype, one would not see as much variation in most likely genotype due to imputation). Allowing for imputation of a quantitative value, such as an allele "dosage" variable with possible values ranging from 0 to 2, may better capture the variation due to imputation. In the current analyses, use

of HapMap data only (without NIEHS SNPs data) strengthened many associations, suggesting that either (a) the NIEHS SNPs samples were not appropriate to use as reference (if associations are true) or (b) the NIEHS SNPs samples provided increased power to discriminate true from false associations (if associations are false). Additional genotyping is under way to examine the accuracy of imputed genotypes and the consistency of ovarian cancer association signals in *CDKN2A-CDKN2B*, *CCND1*, *CDK2*, and *CCNE1* seen with imputed data. Although developed primarily for genome-wide association studies, we conclude that pooling genotypes and using imputation techniques may also strengthen our understanding of key candidate ovarian cancer pathways.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

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