

β -Glucans: An Important Bioactive Molecule of Edible and Medicinal Mushrooms

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Abstract

Edible mushrooms are important for their dietary value and their biologically active and health promoting compounds such as polysaccharides, polysaccharopeptides and polysaccharide-protein complexes. Mushroom β -glucans such as β -glucan, Schizophyllan, Ganoderan, Lentinan and Pleuran are the components of the cell wall. They consist of glucopyranose molecules linked through β (1 \rightarrow 3), β (1 \rightarrow 4) or β (1 \rightarrow 6) linkages. The mushroom β -glucans are not digested in human gastrointestinal tract and are thus considered as a potential source of prebiotics. β -glucans possess profound health promoting properties like speeding up the transit of bowel contents, increasing fecal bulk and frequency, consequently protecting the body from colon cancer, diverticular diseases and irritable bowel syndrome. They stimulate the immune system by having immunomodulatory, antitumour, antioxidant activities and are identified as biological response modifiers. Mushroom β -glucans differ in their nutraceutical effect due to the difference in their molecular masses, solubility, degree of polymerization, their structures and helical conformation. Various mushroom β -glucans are available as pure extracts in the market which are used as therapeutic agents, however, no commercialized functional products are available which have been enriched with mushroom β -glucans. Furthermore, it has a great potential to be used as an ingredient in the near future in various food industries, such as breakfast cereals, sport nutrition products, dairy products, bakery such as biscuits and breads, salad dressings and fat replacer. The aim of this review is to present information on β -glucans of edible and medicinal mushrooms, emphasize their benefits and the usage potential in the functional food and nutraceuticals.

Keywords: Mushrooms, β -glucan, bioactivity, functional, nutraceutical, food industry.

1. Introduction

β -glucans are polysaccharides of D-glucose monomers linked through β -glycosidic bonds. The structures of β -glucans and chemically modified β -glucans were given in Figure 1.

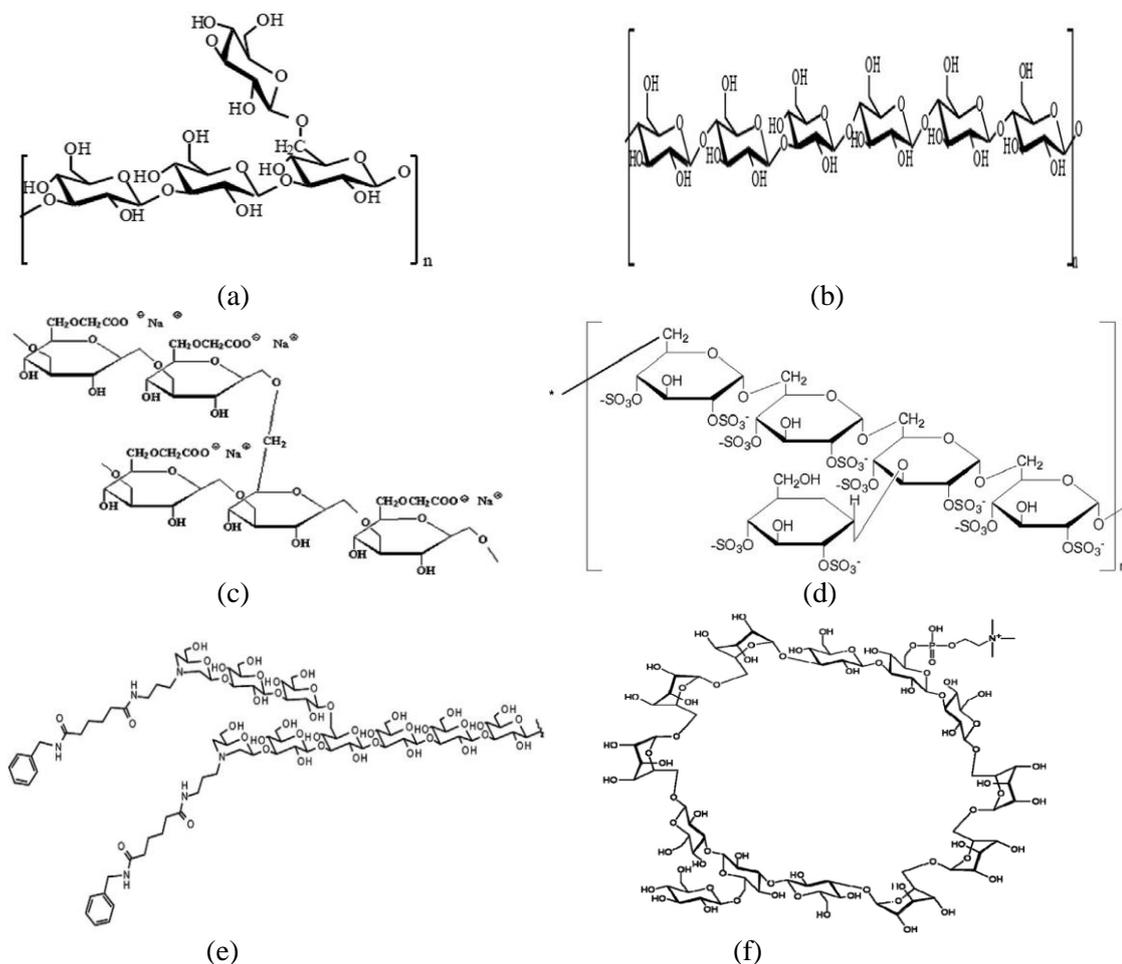


Figure 1. The structures of β -glucans and chemically modified β -glucans. (a) (1 \rightarrow 3) β -glucans with ramifications β (1 \rightarrow 6); (b) (1 \rightarrow 3) β -glucans with ramifications β (1 \rightarrow 4); (c) CM β -glucan; (d) Sulfated β -glucan; (e) Aminated β -glucan; (f) Cyclic glucan (Zhu et al., 2016).

As a kind of dietary fiber (DF), β -glucan could be found in a variety of natural sources such as yeast, mushrooms, bacteria, algae, barley and oat (Zhu et al., 2015). β -glucan exhibits a broad spectrum of biological activities including antitumour, immune-modulating (Rieder and Samuelsen, 2012), antiaging, antimicrobial, antioxidant and antiinflammatory properties. β -glucans have attracted attention because of their physical and chemical properties over the years. β -glucans from different sources and with different molecular weights have different biological activities (Du and Xu, 2014). Fungal β -glucan has been shown to be effective as an immune system booster and an antitumour substance (Du et al., 2015). The results of clinical

research indicate that the presence of β -glucan is linked to the production and activation of macrophages, NK-cells, T-cells, B-cells from the body's natural defense system (Lindequist et al., 2005).

2. Mushroom β -glucans

Mushroom β -glucan is a carbohydrate polymer derived from the cell wall of mushrooms. β -glucan is known as biological response modifier (BRM), which refers to the ability to up-regulate and down-regulate the response of biological systems (Brown and Gordon, 2003; Novak and Vetvicka, 2009).

Mushroom β -glucans such as Schizophyllan, Ganoderan, Lentinan and Pleuran are the components of the cell wall. They consist of glucopyranose molecules linked through β (1 \rightarrow 3), β (1 \rightarrow 4) or β (1 \rightarrow 6) linkages. Especially in Japan and China, Pleuran from *Pleurotus ostreatus*, Lentinan from *Lentinula edodes*, Schizophyllan from *Schizophyllum commune*, Grifolan (MD-fraction) from *Grifola frondosa* and Krestin from *Trametes versicolor* (PSK and PSP) in addition to the major cancer therapies like surgical operation, radiotherapy and chemotherapy are in clinical use for the adjuvant tumour therapy (immunotherapy) (Lindequist et al., 2005; Chan et al., 2009; Novak and Vetvicka, 2009).

β -glucans are also present in many other mushrooms such as *Auricularia auricula*, *Calocybe indica* (Calocyban), *Flammulina velutipes*, *Ganoderma lucidum* (Ganoderan/Ganopoly), *Grifola frondosa*, and *Pleurotus abalones* (Lindequist et al., 2005; Villares et al., 2012; Zhu et al., 2015).

2.1. β -glucan Amounts in Mushrooms

Important medical mushrooms containing β -glucan as bioactive compound are seen in Table 1. The β -glucan contents of the mushrooms vary between 0.22 and 0.53 g/100 g on dry weight basis. According to Manzi and Pizzoferrato (2000), *Pleurotus pulmonarius* seemed to be the richest source of fungal β -glucans and it has been reported that *L. edodes* contains high levels of β -glucans in the soluble fraction. Camelini et al. (2005) found that *Agaricus brasiliensis* had higher (1 \rightarrow 6)- β -glucan ratio and (1 \rightarrow 3)- β -glucan increased with the maturation of fruiting bodies.

Table 1. Important medicinal mushrooms with β -glucans as bioactive components (Chan et al., 2009).

Mushroom species	Common name	β -glucan structure	Type of β -glucan
<i>Agaricus blazei</i>	Brazilian sun-mushroom, Himematsutake mushroom	Protein bound β -1,6-glucan	<i>Agaricus</i> polysaccharides
<i>Coprinus comatus</i>	Shaggy ink cap, lawyer's wig, or shaggy mane	β -1,3-glucan	<i>Coprinus</i> polysaccharides
<i>Coriolus versicolor</i>	Yun Zhi	Protein bound β -1,3;1,6-glucan	PSP (polysaccharide peptide) PSK (polysaccharide-Kureha or polysaccharide-K, krestin)
<i>Ganoderma lucidum</i>	Lingzhi, Reishi	β -1,3;1,6-glucan	<i>Ganoderma</i> polysaccharides, Ganopoly
<i>Grifola frondosa</i>	Maitake mushroom	β -1,3;1,6-glucan with xylose and mannose	Maitake D-Fraction
<i>Lentinula edodes</i>	Shiitake mushroom	β -1,3;1,6-glucan	Lentinan
<i>Pleurotus ostreatus</i>	Oyster mushroom, píng gû	β -1,3-glucan with galactose and mannose	Pleuran
<i>Schizophyllum commune</i>	Brazilian mushroom	β -1,3;1,6-glucan	Schizophyllum (SPG) or sizofiran

It was determined that Bracket fungi *Trametes versicolor*, *Piptoporus betulinus* or *Phlebia tremellosa* contained more than 50% β -glucans and in *Boletus edulis* (Bull. ex Fr., stipe part) or *Piptoporus betulinus* (Bull. ex Fr.) Karst. the amount was more than 50 g/100 g dw. In most of the wild mushrooms analysed, the β -glucan contents were significantly higher in stipes than in caps (Sari et al., 2017). Özcan and Ertan (2018) have determined that *Boletus edulis* is the highest β -glucan containing wild mushroom (13.93%). It was followed by *Cantharellus cibarius* and *Hydnum repandum* with 12.89 and 12.84% contents, respectively. Synytsya et al. (2008) working with *Pleurotus* spp. mushrooms found that the β -glucan content of the pilei was between 20.4-39.2% and the content of the stems was between 35.5-50.0%. The results of a study comparing the β -glucan content of some wild mushrooms (Sari et al., 2017) is presented in Table 2.

Table 2. β -glucan content of some wild mushrooms (not dividable in cap and stipe parts).

Mushroom species	Dry matter (%)	β -glucans (g/100g dm)	% β -glucans/all glucans
<i>Auricularia auricula</i> (L.) Underwood	91.276	41.755±4.644	99.096
<i>Fomes fomentarius</i> (L.) Fr.	86.122	22.495±2.329	90.186
<i>Grifola frondosa</i> (Dicks. ex Fr.) Gray	92.783	25.991±3.643	83.761
<i>Laetiporus sulphureus</i> (Bull. ex Fr.) Murr.	89.627	47.006±6.517	89.345
<i>Phlebia tremellosa</i> (Schrad.) Nakasone & Burds.	96.547	53.555±2.452	98.572
<i>Piptoporus betulinus</i> (Bull. ex Fr.) Karst.	90.825	51.801±4.024	95.659
<i>Trametes versicolor</i> (L.) Lloyd.	87.892	60.788±11.795	99.337

2.2. Bioactive Properties of β -glucans in Mushrooms

Mushrooms polysaccharides having β -linkage have been demonstrated a boost in the human immune system and the modulation of the immunological response under certain conditions, thus they are commonly termed as biological response modifiers (BRM). As the result of the activation of the host's immune system, these polysaccharides show significant antitumour, antiviral and antimicrobial activities besides their other effects (Villares et al., 2012). A number of studies have been carried out on β -glucans that have a health-enhancing effect in various important ways such as antitumour and immunomodulatory (Table 3), antitumour, antiviral (Borchers et al., 2004; Moradali et al., 2007), cardiovascular (Wasser and Weis, 1999), liver protective, antiinflammatory (Lindequist et al., 2005), radioprotective (Pillai and Devi, 2013), antidiabetic (Kim et al., 2005), antioxidant (Deng et al., 2012), antibacterial (Beattie et al., 2010), and antiobesity activities (Zhang et al., 2013). Antitumour activity (Deng et al., 2012; Ren et al., 2012) and immunomodulating activity (Wasser, 2002) of mushroom β -glucans have been documented in the previous reviews. Basically, their health-promoting abilities are influenced by the molecular mass, branching configuration, conformation, and chemical modification of the polysaccharides (Ren et al., 2012). In terms of biological activity, β -1,3-D-glucans and β -1,6-D-glucans contained in oyster, shiitake, split gill, and himematsutake mushrooms, as well as other Basidiomycetes, are considered to be the most effective (Rop et al., 2009).

Table 3. Some mushroom β -glucans with antitumour and immunomodulatory activities (Zhang et al., 2007; Novak and Vetvicka, 2008; Kothari et al., 2018).

Mushroom species	Type of β -glucan	Character of polymer	Degree of branching
<i>Agaricus blazei</i>	β -glucomannan	Branched	-
<i>Dictyophora indusiata</i>	T-4-N, T-5-N	Branched	-
<i>Ganoderma lucidum</i>	Ganoderan	Branched	-
<i>Grifola frondosa</i>	Grifolan	Branched	0.31-0.36
<i>Inonotus obliquus</i>	Xylogalactoglucan	Branched	-
<i>Laminaria</i> spp.	Laminaran	Linear	-
<i>Lentinula edodes</i>	Lentinan	Branched	0.23-0.33
<i>Pleurotus ostreatus</i>	Pleuran (HA-glucan)	Branched	0.25
<i>Poria cocos</i>	Pachymaran	Linear	0.015-0.020
<i>Schizophyllum commune</i>	Schizophyllan	Branched	0.33
<i>Sclerotinia sclerotiorum</i>	Sclerotinan (SSG)	Branched	0.50

2.3. Extraction and Production of Mushroom β -glucans

The methods used to extract and produce β -glucans from various edible/medicinal mushrooms are given in Table 4.

Table 4. Production/extraction process of β -glucans from various edible/medicinal mushrooms.

Mushroom species	Production/Extraction process	References
<i>Agaricus bisporus</i>	Ultrasonic-assisted extraction, precipitation with ethanol, centrifugation	Tian et al., 2012
	Lyophilization, milled and submitted to successive cold and hot aqueous extraction	Smiderle et al., 2013
<i>Agaricus brasiliensis</i>	Sequentially extracted with 350 ml water, concentrated, dialyzed and DEAE-cellulose column chromatography	Camelini et al., 2005
	Lyophilization, milled and submitted to successive cold and hot aqueous extraction	Smiderle et al., 2013
<i>Astraeus hygrometricus</i>	Aqueous extraction, DEAE cellulose bag and Sepharose 6B column	Chakraborty et al., 2004
<i>Boletus edulis</i>	The estimation of non-starch glucans was based on the difference between glucose contents after total acidic hydrolysis of glucans and specific enzymatic hydrolysis of α -1,4-glucans	Özcan and Ertan, 2018
<i>Boletus erythropus</i>	Water extraction, centrifugation, DEAE Trisacryl M column and S 400 HR column	Chauveau et al., 1996
<i>Botryosphaeria rhodina</i>	β -glucan production were monitored in a stirred-tank bioreactor	Crognale et al., 2007
<i>Cantharellus cibarius</i>	The estimation of non-starch glucans was based on the difference between glucose contents after total acidic hydrolysis of glucans and specific enzymatic hydrolysis of α -1,4-glucans	Özcan and Ertan, 2018
<i>Flammulina velutipes</i>	Successive hot extraction with water and KOH and submitted to freeze-drying	Smiderle et al., 2006
<i>Ganoderma lucidum</i>	Extraction using dilute NaOH solution and Sephadex G-15 gel-filtration chromatography	Kao et al., 2012; Nie et al., 2013
<i>Paenibacillus polymyxa</i>	Seed culture was supplemented with carbon source to induce glucan production	Jung et al., 2007
<i>Pleurotus eryngii</i>	The estimation of non-starch glucans was based on the difference between glucose contents after total acidic hydrolysis of glucans and specific enzymatic hydrolysis of α -1,4-glucans	Synytsya et al., 2008
	Washing with ethanol and distilled water, extraction with boiling water, incubation with α -amylase, chemical deproteinization, dialization and lyophilization	Synytsya et al., 2009
<i>Pleurotus ostreatus</i>	The estimation of non-starch glucans was based on the difference between glucose contents after total acidic hydrolysis of glucans and specific enzymatic hydrolysis of α -1,4-glucans	Synytsya et al., 2008
	Washing with ethanol and distilled water, extraction with boiling water, incubation with α -amylase, chemical deproteinization, dialization and lyophilization	Synytsya et al., 2009
	Lyophilization, using of methanolic extraction, cold water, hot water, hot aqueous NaOH solutions, enzyme protease, and ethanol precipitation	Palacios et al., 2012
<i>Ramaria botrytis</i>	Hot water extraction followed by treating NaOH	Bhanja et al., 2014
<i>Schizophyllum commune</i>	Seed culture preparation, optimization of fermentation medium and schizophyllan production	Kumari et al., 2008
<i>Termitomyces eurhizus</i>	Hot alkaline extraction, centrifugation, DEAE cellulose bag and freeze dry	Chakraborty et al., 2006

The detection methods of β -glucan from mushrooms are summarized as: (1) enzymic method or McCleary method (Megazyme kit), (2) enzymelinked immunosorbent assay

(ELISA) method, (3) fluorimetric method with aniline blue, and (4) colorimetric method with Congo red (Zhu et al., 2015).

2.3.1 Extraction and Production of β -glucans from Fruiting Bodies of Mushrooms

Kim et al. (2005) extracted β -glucan from the fruiting bodies of *Agaricus blazei* using hot water for 3 h. Bhanja et al. (2014) extracted and isolated two water-insoluble glucans from fruiting bodies of *Ramaria botrytis*. Extraction in hot or boiling water is the most common and convenient method for extracting water-soluble fungal polysaccharides (Yan et al., 2014). Liu et al. (2014) obtained a purified β -glucan by precipitating a hot-water extract from fruiting bodies of *G. lucidum* with 20% (V/V) ethanol. The total carbohydrate content was 95.9% in prepared β -glucan.

2.3.2. Extraction and Production of β -glucans from the Mycelia of Mushrooms

Kim et al. (2009) provided a method for mass production of β -glucan from *S. commune*, comprising subjecting mycelia of *S. commune* to liquid culture with an addition of a synthetic adsorbent. In another study, a neutral polysaccharide, GLSA50-1B, was isolated from sporoderm-broken spores of *G. lucidum*, by hot-water extraction, graded ethanol precipitation, anion-exchange chromatography, and gel permeation chromatography (Dong et al., 2012). Kim et al. (2013) demonstrated generation of high β -glucan producing mutant strains of *Sparassis crispa*, additional culture optimization further increased β -glucan productivity of the mutant strains. Recently, Park et al. (2014) enhanced the β -glucan content in the sawdust-based cultivation of cauliflower mushroom (*Sparassis latifolia*) using three kinds of enzymes (chitinase, β -glucuronidase, and lysing enzyme complex) as elicitors.

2.4. Chemical Modification and Purification of Mushroom β -glucans

β -glucan is an important bioactive compound for human health, but its low solubility has led to the development of chemical modification technologies to improve bioavailability. Several methods to modify β -glucan are laid out to improve their functional and technological properties via physical and chemical crosslinking reactions (Ahmad et al., 2015). In this respect, β -glucans can be chemically modified to obtain various derivatives with potential industrial or medicinal importance (Synytsya and Novak, 2013).

Ion-exchange chromatography and gel filtration chromatograph are the most common and convenient methods for purifying polysaccharide. In general, the crude polysaccharide extracts were further applied to a Sephadex column and eluted with water (Zhu et al., 2015).

3. Studies on Mushroom β -glucan for Food and Nutraceutical Applications

3.1. Food Applications and Some Patents

There have been some studies previously conducted by enriching a product with the mushroom β -glucan agglomerated as food additive. β -glucans from *P. ostreatus* and *L. edodes* have been demonstrated satisfactory results when they were added to yogurt (Hozova et al., 2004) were used in the production of extruded snack products with low glycemic index (Brennan et al., 2013). Also chicken burgers were enriched with *P. sajor-caju*, fiber and β -glucans (Wan Rosli et al., 2011) as well as *P. ostreatus* was incorporated into sausages in an effort to lower their fat content (Chockchaisawasdee et al., 2010). In a study to produce a novel high-fibre and low-calorie functional food, Kim et al. (2011) used β -glucans from *L. edodes* as a wheat flour substitute in baked foods. These glucans improved the pasting properties of wheat flour and increased batter viscosity and shear-thinning elasticity without any adverse effect on air holding capacity or hardness. β -glucans of *Ganoderma amboinense*, *Agaricus* or *Fomes yucatensis*, or mixed mushrooms have also been tested for encapsulation of pickling liquid to be released in soups or sauces during cooking (Watanabe, 2005).

3.2. Nutraceutical Applications, Some Clinical Studies and Patents

“Mushroom nutraceuticals” is nowadays a relatively common term which refers to a refined polysaccharide, or a partially refined fruit body extract, or the dried biomass from mycelium or the fruiting body of a mushroom, which is consumed in the form of capsules, tablets, powder, syrups, solutions as a dietary supplement with some therapeutic properties (Giavasis, 2014). Camelini et al. (2005) investigated the β -glucans of *A. brasiliensis* in different stages of fruiting body maturity and their use in nutraceutical products. The results showed that because of their important glucan contents, mature fruiting bodies of *A. brasiliensis* should be used for nutraceutical products. Cap-opened, more fragile mature fruiting bodies of *A. brasiliensis* should be selected over immature ones for the production of

nutraceuticals. Synytsya et al. (2008) reported that the stems of *Pleurotus eryngii* and *P. ostreatus* could be used for the preparation of biologically active polysaccharide complexes as food supplements. Schizophyllan, produced by *S. commune* ATCC 38548 has attracted attention as immunomodulatory and anti-neoplastic agent in pharmaceutical industry in the recent years (Kumari et al., 2008). Akiyama et al. (2011) studied the effects of agaritine, a hydrazine-derivative from hot-water extract of *A. blazei* Murrill on human leukemic monocyte lymphoma (U937) cells. Agaritine induced DNA fragmentation, annexin V expression, and cytochrome C release. Caspase-3, 8 and 9 activities were gradually increased after agaritine treatment. *A. blazei* has been used as an adjuvant in cancer chemotherapy and various types of anti-leukemic bioactive components have been extracted from it (Patel and Goyal, 2012). It was proposed to mix β -glucan from mushroom with one or two substances such as ubiquinone Q10 and ferments leading to a biologically active additive for food with a wide range of action (Bragintseva et al., 2002). Suga et al. (2005) suggested converting lentinan into superfine particles, improving absorption through mucosa.

In animal experiments, β -glucans have been shown to have varying activity against sarcomas, mammary cancer, some chemically induced cancers, adenocarcinoma, colon cancer and some leukemias. Lentinan has already been shown effective in gastric carcinomas (Taguchi et al., 1985; Jeannin, et al., 1988). Furthermore, lentinan was reported to induce apoptosis in murine skin carcinoma cell-lines (Gu and Belury, 2005). Even if mushrooms and especially β -glucans have been used in Chinese medicine for decades, mechanisms need to be elucidated. However, lot of these substances have already been patented for antitumour treatments. Among them, β -glucan extracted from *Agaricus* mushroom was proposed, together with fucoidan (Hosokawa, 2003). The use of *Grifola frondosa* extract has also been patented, mixed with fucoidan and organic germanium (Sogabe, 1998).

Extracts of *L. edodes* markedly inhibited the growth of Sarcoma 180 (a retrovirus, similar to HIV which uses reverse transcriptase for its tumourpromoting activity) (Chihara et al., 1987). According to clinical studies, lentinan produces specific T-helper cell stimulation in healthy humans as well as animals. It has also been recognized to stimulate lymphokine activated killer activity in combination with Interleukin-2 (Suzuki et al., 1990). Other patents concerning direct utilisation of β -D-glucans such as *G. frondosa* extract (Sogabe, 1996) for treating AIDS have been rare.

In 2003, an original application has been patented, proposing to use β -glucan as a gene carrier (Sakurai et al., 2003). In this patent, a hydrogen-bonding polymer with a triple-helix structure (such as schizophyllan, curdlan, lentinan, scleroglucan) was used for binding to a

nucleic acid. Thus, a nucleic acid-polymer complex was obtained and could be applied as a vector. Moreover, this complex was also resistant to nuclease, allowing its use as a nucleic acid-protecting agent. PSP derived from *Coriolus versicolor* (syn. *Trametes versicolor*), a Chinese product commercially available since 1987 (Cui and Chisti, 2003), has been documented to improve the quality of life in cancer patients by providing substantial pain relief and enhancing immune status in 70-97% of patients with stomach, esophagus, lung, ovary and cervical cancers. Both PSK and PSP boosted immune cell production, ameliorated chemotherapy symptoms and enhanced tumour infiltration by dendritic and cytotoxic T-cells (Kidd, 2000). From a commercial standpoint, pleuran from oyster (*P. ostreatus*) mushrooms and lentinan from Shiitake (*L. edodes*) mushrooms are currently the most frequently used β -glucans. Both of them show positive effects on the intestines. They increase the resistance of intestinal mucosa to inflammation (Zeman et al., 2001) and inhibit the development of intestinal ulcers (Nosalova et al., 2001). Lentinan also shows a positive effect on peristalsis (Van Nevel et al., 2003).

3.3. Industrial Food Applications of Mushroom β -glucan in Functional Foods and Dietary Supplements

According to literature data, β -glucan has the potential to perform functions in the food industry such as thickening, water-holding, or oil-binding, gelling, film-making and encapsulation agent, and emulsifying stabilizer (Ahmad et al., 2012a, b; Giavasis, 2013; Zhu et al., 2016). Today, mushroom-glucans are found in the market more in the form of capsules or tablets as food supplements and to a lesser extent as ingredients in the food products (Eleftherios et al., 2014). In addition to food, β -glucans have potential applications in medicine and pharmacy, cosmetic and chemical industries, in veterinary medicine and feed production (Laroche and Michaud, 2007; Zhu et al., 2016). Besides, various mushroom β -glucans are available as pure extracts in the market which are used as therapeutic agents. *S. commune* glucan manufactured by Bioland Technology Co. Ltd. is commercially available in the market (Zhu et al., 2016). There have been two patents on production technology of β -glucan from *S. commune*, today (Kim et al., 2008; 2009). Polysachharides such as lentinan, schizophyllan from shiitake and Schizophyllan mushrooms, PSK and PSP, the protein bound polysachharides from turkey tail mushroom, have been developed as anticancer agents in Japan and are now available worldwide (Lull, et al., 2005). There has also been a patent on application on β -glucan process, additive and food product (Cahill et al., 2003). Although

some of the most studied polysaccharides produced by mushrooms (e.g. schizophyllan and lentinan) are already available and marketed as nutraceuticals (pharmaceutical formulation), their addition to food in their purified form has not been commercialized, yet (Giavasis, 2013). Nevertheless, β -glucan has a great potential to be used as an ingredient in the near future in various food industries, such as breakfast cereals, probiotic sausage formulations, sport nutrition products, dairy products such as yogurts, bakery products such as biscuits, breads, cakes and ready-to-eat snacks, beverages, salad dressings and fat replacer that have some functionalities such as noticeable effect on physical and sensory properties, calorie-reducing and cholesterol-lowering actions and faster proteolysis, lower release of large peptides and a higher proportion of free amino acids, the glycemic response manipulation, controlling food intake and reducing 24 h energy intake and having good quality characteristics (Zhu et al., 2016).

The polysaccharides extracted from *A. brasiliensis*, *C. sinensis*, *G. lucidum*, *G. frondosa*, *L. edodes*, and *T. versicolor* are used to produce tablets for inhibiting the growth of tumours and improving the immunity (Rai et al., 2005). Several mushroom products, mainly polysaccharides such as β -D-glucans, have also proceeded successfully through clinical trials and are used as drugs to treat cancer and chronic diseases (Morris et al., 2016).

Today, it is possible to find commercial dietary supplements originated from various mushroom β -glucans in the form of powdered extracts, tablets, capsules, teas and syrups on the market. Imunoglukan P4H® from *P. ostreatus*, LentinanXP in USA/Lentinex® in Europe and Shiitake Gold and Pure Shiitake™ from *L. edodes*, Ganopoly® and Immulink MBG® from *G. lucidum*, D-fraction, MD fraction, MaitakeGold 404® nutraceutical extract and Pure Maitake™ from *G. frondosa*, Pure Turkey Tail™ from *T. versicolor* and Immune Assist™ from *A. blazei*, *C. sinensis*, *G. lucidum*, *G. frondosa*, *L. edodes* and *T. versicolor* can be given as example (Point Institute, 2013; Morris et al., 2016; Reis et al., 2017; URL-1, 2018). McCleary and Draga (2016) developed a robust and reliable method for the measurement of β -glucan in mushroom and mycelial products. In the literature, there have also been some clinical studies on pharmacological benefits and safe doses of these mushroom β -glucan derived dietary supplements such as Lentinex® (Gauillier et al., 2011) and Imunoglukan P4H® (Jesenak et al., 2012). A scientific documentation was published to carry out the additional safety assessment for Lentinex®, an aqueous mycelial extract of *L. edodes*, as a novel food ingredient (EFSA, 2010). On the other hand, Gründemann et al. (2015) have reported that the standardisation of shiitake preparations is difficult because even preparations with similar polysaccharide and β -glucan contents have different immunological properties.

4. β -glucan Market by Food & Beverage Applications and Regions

The industries are adopting β -glucan to fortify foodstuff with high dietary fibres as consumer interests in the nutraceutical products is on escalation. Furthermore β -glucan actively impacts the metabolic parameters and help curing the chronic diseases. The worldwide development of policies for inclusion of functional ingredients in industrial products boosts the global β -glucan market. According to application segmentation, food and beverage segment was accounted more than 25% value share in 2016. Increasing demand for fibrous intake and concerns over blood cholesterol levels majorly drives the β -glucan market in food and beverage applications. In addition, β -glucan allows food product manufacturers to attract attention heart health claims in functional foods such as heart healthy biscuits, dairy products, snack bars etc., which in turn aids in driving the global β -glucan market (URL-2, 2018).

Geographically, the Europe accounted major share in the global β -glucan market in 2016. Approval of health claims by EU, related to heart health, blood glucose, cholesterol control and digestive health will be fueling the growth for β -glucan market in the region over the forecast period. In Asia Pacific, the government initiatives for awareness on cancer, women heart and maternal health are expected to drive the sales revenue of β -glucan during the forecast period of 2017-2025 (URL-2, 2018).

5. Conclusion

Although there are many findings related to the biological effects of β -glucans *in vitro* and *in vivo*, there are still some questions about structure activity and dose activity relationships. Moreover, β -glucan content of mushroom products has not been standardised, yet. To make better use of β -glucan, food manufacturers and processors must bring attention not only to ensure sufficient concentration of β -glucan in the raw material but also to the processing methods and physicochemical properties of β -glucan, decreasing mechanical and enzymatic breakdown of the β -glucans in end-product and optimizing processing conditions. Mushroom β -glucans have potential nutraceutical properties that could be explored in the food and the pharmaceutical fields and might present different functional properties upon modification through suitable means and continuity of detailed clinical studies for the convenience of consumers.

References

- Ahmad, A., Anjum, F. M., Zahoor, T., Nawaz, H., and Dilshad, S. M. (2012a). Beta glucan: A valuable functional ingredient in foods. *Critical Reviews in Food Science and Nutrition*, 52, 201-212.
- Ahmad, A., Munir, B., Abrar, M., Bashir, S., Adnan, M., and Tabassum, T. (2012b). Perspective of β -glucan as functional ingredient for food industry. *Journal of Nutrition and Food Sciences*, 2, 133.
- Ahmad, N. H., Mustafa, S., and Che Man, Y. B. (2015). Microbial polysaccharides and their modification approaches: A review. *International Journal of Food Properties*, 18, 332-347.
- Akiyama, H., Endo, M., Matsui, T., Katsuda, I., Emi, N., Kawamoto, Y., Koike, T., and Beppu, H. (2011). Agaritine from *Agaricus blazei* Murrill induces apoptosis in the leukemic cell line U937. *Biochimica et Biophysica Acta*, 1810, 519-525.
- Beattie, K. D., Rouf, R., Gander, L., May, T. W., Ratkowsky, D., Donner, C. D., Gill, M., Grice, I. D., and Tiralongo, G. (2010). Antibacterial metabolites from Australian macrofungi from the genus *Cortinarius*. *Phytochemistry*, 71, 948-955.
- Bhanja, S. K., Rout, D., Patra, P., Sena, I. K., Nandan, C. I. K., and Islam, S. S. (2014). Water-insoluble glucans from the edible fungus *Ramaria botrytis*. *Bioactive Carbohydrates and Dietary Fibre*, 3(2), 52-58.
- Borchers, A., Keen, C. L., and Gershwin, M. E. (2004). Mushrooms, tumors, and immunity: an update. *Experimental Biology and Medicine*, 229, 393-406.
- Bragintseva, L. M., Grigorash, A. I., Kovalenko, V. A., Maklanov, A. I., and Ustynjuk, T. K. (2002). RU2177699.
- Brennan, M. A., Derbyshire, E., Tiwari, B. K., and Brennan, C. (2013). Integration of β -glucan fibre rich fractions from barley and mushrooms to form healthy extruded snacks. *Plant Foods for Human Nutrition*, 68(1), 78-82.
- Brown, G. D., and Gordon, S. (2003). Fungal beta-glucans and mammalian immunity. *Immunity*, 19, 311-315.
- Cahill, A. P., Fenske, D. J., Freeland, M., and Hartwig, G. W. (2003). Beta-glucan process, additive and food product. US patent 6749885 B2.
- Camelini, C. M., Maraschin, M., Matos de Mendonça, M., Zucco, C., Ferreira, A. G., and Tavares, L. A. (2005). Structural characterization of β -glucans of *Agaricus brasiliensis* in different stages of fruiting body maturity and their use in nutraceutical products. *Biotechnology Letters*, 27(17), 1295-1299.
- Chakraborty, I., Mondal, S., Pramanik, M., Rout, D., and Islam, S. S. (2004). Structural investigation of a water-soluble glucan from an edible mushroom, *Astraeus hygrometricus*. *Carbohydrate Research*, 339(13), 2249-2254.
- Chakraborty, I., Mondal, S., Rout, D., and Islam, S. S. (2006). A water-insoluble (1 \rightarrow 3)-beta-D-glucan from the alkaline extract of an edible mushroom *Termitomyces eurhizus*. *Carbohydrate Research*, 341(18), 2990-2993.
- Chan, G. C. F., Chan, W. K., and Sze, D., M. Y. (2009). The effects of β -glucan on human immune and cancer cells. *Journal of Hematology & Oncology*, 2, 25.
- Chauveau, C., Talaga, P., Wieruszkeski, J. M., Strecker, G., and Chavant, L. (1996). A water-soluble beta-D-glucan from *Boletus erythropus*. *Phytochemistry*, 43(2), 413-415.
- Chihara, G., Hamuro, J., Maeda, Y.Y., Arai, Y., and Fukuoka, F. (1987). Function and purification of the polysaccharides with marked antitumor activity, especially Lentinan, from *Lentinus edodes* (Berk.). *Cancer Research*, 30, 2776-2781.
- Chockchaisawasdee, S., Namjaidee, S., Pochana, S., and Stathopoulos, C. E. (2010). Development of fermented oyster mushroom sausage. *Asian Journal of Food & Agro-Industry*, 3, 35-43.
- Crognale, S., Bruno, M., Fidaleo, M., Moresi, M., and Petruccioli, M. (2007). Production of β -glucan and related glucan-hydrolases by *Botryosphaeria rhodina*. *Journal of Applied Microbiology*, 102, 860-871.
- Cui, J., and Chisti, Y. (2003). Polysaccharopeptides of *Coriolus versicolor*: physiological activity, uses, and production. *Biotechnology Advances*, 21(2), 109-22.

- Deng, C., Hu, Z., Fu, H. T., Hu, M. H., Xu, X., and Chen, J. H., (2012). Chemical analysis and antioxidant activity in vitro of a β -D-glucan isolated from *Dictyophora indusiata*. *International Journal of Biological Macromolecules*, 51, 70-75.
- Dong, Q., Wang, Y., Shi, L., Yao, J., Li, J., Ma, F., and Ding, K. (2012). A novel water-soluble β -D-glucan isolated from the spores of *Ganoderma lucidum*. *Carbohydrate Research*, 353, 100-105.
- Du, B., and Xu, B. J. (2014). Oxygen radical absorbance capacity (ORAC) and ferric reducing antioxidant power (FRAP) of β -glucans from different sources with various molecular weight. *Bioactive Carbohydrate and Dietary Fibre*, 3, 11-16.
- Du, B., Lin, C. Y., Bian, Z. X., and Xu, B. J. (2015). An insight into anti-inflammatory effects of fungal beta-glucans. *Trends in Food Science & Technology*, 41, 49-59.
- EFSA Panel on Dietetic Products, Nutrition and Allergies (NDA). (2010). Scientific opinion on the safety of "*Lentinus edodes* extract" as a novel food ingredient. *EFSA Journal*, 8(7), 1685.
- Eleftherios, E., Vassilis, M. G., and Cleanthes, I. (2014). The potential use of mushrooms β -glucans in the food industry. *International Journal of Biotechnology for Wellness Industries*, 3, 15-18.
- Gaullier, J. M., Sleboda, J., Øfjord, E. S., Ulvestad, E., Nurminiemi, M., Moe, C., Albrektsen, T., and Gudmundsen, O. (2011). Supplementation with a soluble beta-glucan exported from shiitake medicinal mushroom, *Lentinus edodes* (Berk.) Singer mycelium: A crossover, placebo-controlled study in healthy elderly. *International Journal of Medicinal Mushrooms*, 13(4), 319-326.
- Giavasis, I. (2013). Production of microbial polysaccharides for use in food. In: *Microbial production of food ingredients, enzymes and nutraceuticals*. United Kingdom: Woodhead Publishing.
- Giavasis, I. (2014). Bioactive fungal polysaccharides as potential functional ingredients in food and nutraceuticals. *Current Opinion in Biotechnology*, 26, 162-173.
- Gründemann, C., Garcia-Käufer, M., Sauer, B., Scheer, R., Merdivan, S., Bettin, P., Huber, R., and Lindequist, U. (2015). Comparative chemical and biological investigations of β -glucan-containing products from shiitake mushrooms. *Journal of Functional Foods*, 18, 692-702.
- Gu, Y. H., and Belury M. A. (2005). Selective induction of apoptosis in murine skin carcinoma cells (CH72) by an ethanol extract of *Lentinula edodes*. *Cancer Letters*, 220, 21-28.
- Hosokawa, J. (2003). JP2003055234.
- Hozova, B., Kuniak, L., and Kelemova, B. (2004). Application of β -glucans isolated from mushrooms *Pleurotus ostreatus* (pleuran) and *Lentinula edodes* (lentinan) for increasing the bioactivity of yoghurts. *Czech Journal of Food Sciences*, 22, 204-214.
- Jeannin, J. F., Lagadec, P., Pelletier, H., Reisser, D., Olsson, N. O., Chihara, G., and Martin, F. (1988). Regression induced by lentinan of peritoneal carcinomas in a model of colon cancer in rat. *International Journal of Immunopharmacology*, 10(7), 855-861.
- Jesenak, M., Majtan, J., Rennerova, Z., Kyselovic, J., Banovcin, P., and Hrubisko, M. (2012). Immunomodulatory effect of pleuran (β -glucan from *Pleurotus ostreatus*) in children with recurrent respiratory tract infections. *International Immunopharmacology*, 15, 395-399.
- Jung, H. K., Hong, J. H., Park, S. C., Park, B. K., Nam, D. H., and Kim, S. D. (2007). Production and physicochemical characterization of β -glucan produced by *Paenibacillus polymyxa* JB115. *Biotechnology and Bioprocess Engineering*, 12, 713-719.
- Kao, P. F., Wang, S. H., Hung, W. T., Liao, Y. H., Lin, C. M., and Yang, W. B. (2012). Structural characterization and antioxidative activity of low-molecular-weights beta-1, 3-glucan from the residue of extracted *Ganoderma lucidum* fruiting bodies. *BioMed Research International*, 1-8.
- Kidd, P. M. (2000). The use of mushroom glucans and proteoglycans in cancer treatment. *Alternative Medicine Review*, 5(1), 4-27.
- Kim, Y. W., Kim, K. H., Choi, H. J., and Lee, D. S. (2005). Anti-diabetic activity of beta-glucans and their enzymatically hydrolyzed oligosaccharides from *Agaricus blazei*. *Biotechnology Letters*, 27(7), 483-487.
- Kim, M. S., Park, Y. D., and Lee, S. R. (2008). Preparation method of beta-glucan from *Schizophyllum commune* and composition for experimental application comprising the same. US patent 0160043 A1.
- Kim, M. S., Park, Y. D., and Lee, S. R. (2009). Method of using beta-glucan from *Schizophyllum commune*. US patent 0023681 A1.

- Kim, J., Lee, S. M., Bae, I. Y., Park, H. G., Gyu Lee, H., and Lee, S. (2011). (1-3)(1-6)- β -glucan-enriched materials from *Lentinus edodes* mushroom as a high-fibre and low-calorie flour substitute for baked foods. *Journal of the Science of Food and Agriculture*, 91, 1915-1919.
- Kim, S. R., Kang, H. W., and Ro, H. S. (2013). Generation and evaluation of high β -glucan producing mutant strains of *Sparassis crispa*. *Mycobiology*, 41(3), 159-163.
- Kothari, D., Patel, S., and Kim, S. K. (2018). Anticancer and other therapeutic relevance of mushroom polysaccharides: A holistic appraisal. *Biomedicine & Pharmacotherapy*, 105, 377-394.
- Kumari, M., Survase, S. A., and Singhal, R. S. (2008). Production of schizophyllan using *Schizophyllum commune* NRCM. *Bioresource Technology*, 99, 1036-1043.
- Laroche, C., and Michaud, P. (2007). New developments and prospective applications for β -(1,3) glucans. *Recent Patents on Biotechnology*, 1, 59-73.
- Lindequist, U., Niedermeyer, T., and Jülich, W. D. (2005). The pharmacological potential of mushrooms. *Evidence-Based Complementary and Alternative Medicine*, 2(3), 285-299.
- Liu, Y., Zhang, J., Tang, Q., Yang, Y., Guo, Q., Wang, Q., Wu, D., and Cui, S. W. (2014). Physicochemical characterization of a high molecular weight bioactive β -D-glucan from the fruiting bodies of *Ganoderma lucidum*. *Carbohydrate Polymers*, 101, 968-974.
- Lull, C., Wichers, H. J., and Savelkoul, H. F. (2005). Antiinflammatory and immunomodulating properties of fungal metabolites. *Mediators of Inflammation*, 2, 63-80.
- Manzi, P., and Pizzoferrato, L. (2000). Beta-glucans in edible mushrooms. *Food Chemistry*, 68, 315-318.
- McClearly, B. V., and Draga, A. (2016). Measurement of β -glucan in mushrooms and mycelial products. *Journal of AOAC International*, 99(2), 364-373.
- Moradali, M. F., Mostafavi, H., Ghods, S., and Hedjaroude, G. A. (2007). Immunomodulating and anticancer agents in the realm of macromycetes fungi (macrofungi). *International Immunopharmacology*, 7, 701-724.
- Morris, H. J., Llaurodo, G., Beltran, Y., Lebeque, Y., Bermudez, R. C., Garcia, N., Gaime-Perraud, I., and Moukha, S. (2016). The use of mushrooms in the development of functional foods, drugs or nutraceuticals. In: *Wild Plants, Mushrooms and Nuts: Functional Food Properties and Applications*, pp. 123-157. John Wiley & Sons, Ltd.
- Nie, S., Zhang, H., Li, W., and Xie, M. (2013). Current development of polysaccharides from *Ganoderma*: Isolation, structure and bioactivities. *Bioactive Carbohydrates and Dietary Fiber*, 1, 10-20.
- Novak, M., and Vetvicka, V. (2008). β -glucans, history, and the present: Immunomodulatory aspects and mechanism of action. *Journal of Immunotoxicology*, 5, 47-57.
- Novak, M., and Vetvicka, V. (2009). Glucans as biological response modifiers. *Endocrine, Metabolic & Immune Disorders Drug Targets*, 9, 67-75.
- Nosalova, V., Bobek, P., Cerna, S., Galbavy, S., and Stvrtna, S. (2001). Effects of pleuran (beta-glucan isolated from *Pleurotus ostreatus*) on experimental colitis in rats. *Journal of the Physiological Research*, 50, 575-581.
- Özcan, Ö., and Ertan, F. (2018). Beta-glucan content, antioxidant and antimicrobial activities of some edible mushroom species. *Journal of Food Science and Technology*, 6(2), 47-55.
- Palacios, I., García-Lafuente, A., Guillamón, E., and Villares, A. (2012). Novel isolation of water-soluble polysaccharides from the fruiting bodies of *Pleurotus ostreatus* mushrooms. *Carbohydrate Research*, 358, 72-77.
- Patel, S., and Goyal, A. (2012). Recent developments in mushrooms as anti-cancer therapeutics: A review. *Biotechnology*, 2, 1-15.
- Park, H., Ka, K. H., and Ryu, S. R. (2014). Enhancement of β -glucan content in the cultivation of cauliflower mushroom (*Sparassis latifolia*) by elicitation. *Mycobiology*, 42, 41-45.
- Pillai, T. G., and Devi, U. P. (2013). Mushroom beta glucan: Potential candidate for post irradiation protection. *Mutation Research*, 751, 109-115.
- Point Institute. (2013). The use of mushroom-derived dietary supplements as immunomodulating agents: An overview of evidence-based clinical trials and the mechanisms and actions of mushroom constituents. Technical Report. Wisconsin: Stevens Point.
- Rai, M., Tidke, G., and Wasser, S. P. (2005). Therapeutic potential of mushrooms. *Natural Product Radiance*, 4(4), 246-257.

- Reis, F. S., Martins, A., Vasconcelos, M. H., Morales, P., and Ferreira, I. C. F. R. (2017). Functional foods based on extracts or compounds derived from mushrooms. *Trends in Food Science & Technology*, 66, 48-62.
- Ren, L., Perera, C., and Hemar, Y. (2012). Antitumor activity of mushroom polysaccharides: A review. *Food & Function*, 3, 1118-1130.
- Rop, O., Mlcek, J., and Jurikova, T. (2009). Beta-glucans in higher fungi and their health effects. *Nutrition Reviews*, 67(11), 624-631.
- Rieder, A., and Samuelson, A. B. (2012). Do cereal mixed-linked β -glucans possess immune-modulating activities? *Molecular Nutrition & Food Research*, 56, 536-547.
- Sakurai, K., Shinkai, S., Kimura, T., Tabata, K., Koumoto, K., and Gronwald, O. (2003). US2003216346.
- Sari, M., Prange, A., Lelley, J. I., and Hambitzer, R. (2017). Screening of beta-glucan contents in commercially cultivated and wild growing mushrooms. *Food Chemistry*, 216, 45-51.
- Smiderle, F. R., Carbonero, E. R., Mellinger, C. G., Sasaki, G. L., Gorin, P. A., and Iacomini, M. (2006). Structural characterization of a polysaccharide and a beta-glucan isolated from the edible mushroom *Flammulina velutipes*. *Phytochemistry*, 67(19), 2189-2196.
- Smiderle, F. R., Alquini, G., Tadra-Sfeir, M. Z., Iacomini, M., Wichers, H. J., and Van Griensven, L. J. L. D. (2013). *Agaricus bisporus* and *Agaricus brasiliensis* (1 \rightarrow 6)- β -D-glucans show immunostimulatory activity on human THP-1 derived macrophages. *Carbohydrate Polymers*, 94, 91-99.
- Sogabe, T. (1996). JP8119874.
- Sogabe, T. (1998). JP10033142.
- Suga, T., Ogasawara, Y., Kaneko, Y., Kajiura, M., and Suga, Y. (2005). JP2005097308.
- Suzuki, M., Higuchi, S., Taki, Y., Miwa, K., and Hamuro, J. (1990). Activity of Lentinan and Interleukin 2. *International Journal of Immunopharmacology*, 12(6), 613-623.
- Synytsya, A., Míčková, K., Jablonský, I., Sluková, M., and Čopíková, J. (2008). Mushrooms of genus *Pleurotus* as a source of dietary fibres and glucans for food supplements. *Czech Journal of Food Sciences*, 26, 441-446.
- Synytsya, A., Míčková, K., Synytsya, A., Jablonský, I., Spěvácěk, J., Erban, V., Kovarikova, E., and Čopíková, J. (2009). Glucans from fruit bodies of cultivated mushrooms *Pleurotus ostreatus* and *Pleurotus eryngii*: Structure and potential prebiotic activity. *Carbohydrate Polymers*, 76(4), 548-556.
- Synytsya, A., and Novak, M. (2013). Structural diversity of fungal glucans. *Carbohydrate Polymers*, 92, 792-809.
- Taguchi, T., Furue, H., Kimura, T., Kondo, T., Hattori, T., Itoh, T., and Osawa, N. (1985). End-point results of phase 111 study of Lentinan. *Japanese Journal of Cancer and Chemotherapy*, 12, 366-371.
- Tian, Y. T., Zeng, H. L., Xu, Z. B., Zheng, B. D., Lin, Y. X., Gand, C. J., and Lo, Y. M. (2012). Ultrasonic-assisted extraction and antioxidant activity of polysaccharides recovered from white button mushroom (*Agaricus bisporus*). *Carbohydrate Polymers*, 88, 522-529.
- Toyoda, S., and Kimura, M. (2004). US2004047949.
- URL-1: <https://alohamedicinals.com/?s=beta+glucan>, (Date of access: 30 September 2018).
- URL-2: <http://www.credenceresearch.com/report/beta-glucan-market>, Beta glucan market by source type (cereals, fungal & microbial), by application (food & beverage, pharmaceutical, cosmetics, animal feed) - growth, future prospects and competitive analysis, 2017-2025, (Date of access: 24 September 2018).
- Van Nevel, C. J., Decuyper, J. A., Dierick, N., and Molly, K. (2003). The influence of *Lentinus edodes* (Shiitake mushroom) preparations on bacteriological and morphological aspects of the small intestine piglets. *Archives of Animal Nutrition*, 57, 399-412.
- Villares, A., Mateo-Vivaracho, L., and Guillamon, E. (2012). Structural features and healthy properties of polysaccharides occurring in mushrooms. *Agriculture*, 2, 452-471.
- Wan Rosli, W. I., Solihah, M. A., Aishah, M., Fakurudin, N. A., and Mohsin, S. S. J. (2011). Colour, texture properties, cooking characteristics and fibre content of chicken patty added with oyster mushroom (*Pleurotus sajor-caju*). *International Food Research Journal*, 18, 621-627.

- Wasser, S. P., and Weis, A. L. (1999). Medicinal properties of substances occurring in higher Basidiomycetes mushrooms: Current perspectives (Review). *International Journal of Medicinal Mushrooms*, 1(1), 31-62.
- Wasser, S. P. (2002). Medicinal mushrooms as a source of antitumor and immunomodulating polysaccharides. *Applied Microbiology and Biotechnology*, 60, 258-274.
- Watanabe, M. (2005). JP2005160326.
- Yan, J. K., Wang, W. Q., and Wu, J. Y. (2014). Recent advances in *Cordyceps sinensis* polysaccharides: mycelial fermentation, isolation, structure, and bioactivities: A review. *Journal of Functional Foods*, 6, 33-47.
- Zeman, M., Nosalova, V., Bobek, P., Zakalova, M., and Cerna, S. (2001). Changes of endogenous melatonin and protective effect of diet containing pleuran and extract of black elder in colonic inflammation in rats. *Biologia*. 56, 659-701.
- Zhang, M., Cuia, S. W., Cheung, P. C. K and Wang, Q. (2007). Antitumor polysaccharides from mushrooms: A review on their isolation process, structural characteristics and antitumor activity. *Trends in Food Science & Technology*, 18, 4-19.
- Zhang, Y., Xia, L., Pang, W., Wang, T., Chen, P., Zhu, B., and Zhang, J. (2013). A novel soluble β -1, 3-D-glucan salean reduces adiposity and improves glucose tolerance in highfat diet-fed mice. *British Journal of Nutrition*, 109, 254-262.
- Zhu, F., Du, B., Bian, Z., and Xu, B. (2015). Beta-glucans from edible and medicinal mushrooms: Characteristics, physicochemical and biological activities. *Journal of Food Composition and Analysis*, 41, 165-173.
- Zhu, F., Du, B., and Xu, B. (2016). A critical review on production and industrial applications of beta-glucans. *Food Hydrocolloids*, 52, 275-288.