REVIEW ARTICLE

Unravelling the role of arbuscular mycorrhizal fungi in mitigating the oxidative burst of plants under drought stress

Y.-N. Zou¹, Q.-S. Wu^{1,2} 🕞 & K. Kuča²

1 College of Horticulture and Gardening, Yangtze University, Jingzhou, China

2 Department of Chemistry, Faculty of Science, University of Hradec Kralove, Hradec Kralove, Czech Republic

Keywords

Abiotic stress; fatty acids; H₂O₂; mycorrhiza; polyamines; water relations.

Correspondence

Q.-S. Wu, College of Horticulture and Gardening, Yangtze University, Jingzhou 434025, China. E-mail: wuqiangsh@163.com

Editor

Y. Hu

Received: 26 June 2020; Accepted: 22 July 2020

doi:10.1111/plb.13161

INTRODUCTION

Plants are continuously exposed to the external environment, including abiotic and biotic stresses (Nath *et al.* 2016). Abiotic stress produces adverse effects on crop growth and production (Millar & Bennett 2016). Among the different forms of abiotic stress, drought stress is one of most devastating abiotic factors threatening crop growth and productivity worldwide (Guo *et al.* 2020). With climate change and the expansion of human activities, drought stress is becoming more severe and frequent in arid and semi-arid regions. Drought stress triggers several destructive effects in plants, including osmotic imbalance, membrane system damage, cellular metabolic disturbance and abnormal water distribution (Guo *et al.* 2020; Yang *et al.* 2020b).

In plants, drought stress triggers the generation of toxic reactive oxygen species (ROS), such as the superoxide anion free radical $(O_2^{\bullet-})$, singlet oxygen $(^1O_2)$, hydrogen peroxide (H₂O₂) and the hydroxyl radical (OH•) because of the overflow of electrons from chloroplasts, mitochondria, peroxisomes and plasma membranes (Tiwari et al. 2018; Bhattacharjee 2019). Accumulation of ROS induces an oxidative burst in plants subjected to drought stress, and thus causes oxidative damage, including lipid peroxidation, nucleic acid damage, protein oxidation and programmed cell death (Choudhary et al. 2018). Therefore, the adverse effects of drought on plant growth are partly attributed to ROS accumulation (Yang et al. 2020a). Plants also develop enzymatic and non-enzymatic antioxidant defence systems to reduce their oxidative stress (Choudhary et al. 2018). An ascorbic acid-glutathione (AsA-GSH) cycle in plants is utilized to detoxify H2O2 (Bashri & Prasad 2016). It is

ABSTRACT

With continued climate changes, soil drought stress has become the main limiting factor for crop growth in arid and semi-arid regions. A typical characteristic of drought stress is the burst of reactive oxygen species (ROS), causing oxidative damage. Plant-associated microbes, such as arbuscular mycorrhizal fungi (AMF), can regulate physiological and molecular responses to tolerate drought stress, and they have a strong ability to cope with drought-induced oxidative damage via enhanced antioxidant defence systems. AMF produce a limited oxidative burst in the arbuscule-containing root cortical cells. Similar to plants, AMF modulate a fungal network in enzymatic (e.g. GmarCuZnSOD and GintSOD1) and non-enzymatic (e.g. GintMT1, GinPDX1 and GintGRX1) antioxidant defence systems to scavenge ROS. Plants also respond to mycorrhization to enhance stress tolerance via metabolites and the induction of genes. The present review provides an overview of the network of plant – arbuscular mycorrhizal fungus dialogue in mitigating oxidative stress. Future studies should involve identifying genes and transcription factors from both AMF and host plants in response to drought stress, and utilize transcriptomics, proteomics and metabolomics to clarify a clear dialogue mechanism between plants and AMF in mitigating oxidative burst.

> already recognized that ROS levels generate two distinct physiological responses: high concentrations of ROS trigger cell death while low levels of ROS act as a signal to initiate the protective responses of G-proteins and redox-sensitive proteins (Das & Roychoudhury 2014).

> Symbiotic arbuscular mycorrhizal fungi (AMF) are involved in many plant processes, such as nutrient acquisition, photosynthesis, root formation and stress resistance (Humphreys et al. 2010; Chiu et al. 2018; Lanfranco et al. 2018; Liu et al. 2018; Rivero et al. 2018; Gavito et al. 2019; Qiao et al. 2019). AMF from the phylum Mucoromycota, subphylum Glomeromycotina, establish the most common arbuscular mycorrhiza (AM) with the roots of 72% of vascular plants (Bonfante 2018). AM is characterized by the presence of arbuscules inside root cortical cells (Strullu-Derrien et al. 2017). Arbuscules are the site of nutrient exchange from the AMF to the host. Symbiotic associations are characterized by the stimulation of host nutrient uptake by AMF in return for photosynthetic products and lipids form the plants for AM fungal growth. AMF act as protection for host plants against drought stress via a series of underlying mechanisms, including (i) direct water absorption of the mycorrhizal extraradical mycelium; (ii) physiological improvements in nutrient acquisition, osmotic adjustment, hormone balance, antioxidant protective systems, fatty acids (FAs) and polyamines (PAs); (iii) host morphological adaptation; (iv) soil structure improvement by the mycorrhizal extraradical mycelium; and (v) molecular changes (Blee & Anderson, 2000; Ruiz-Lozano et al. 2012; Wu et al. 2013, 2019; Millar & Bennett 2016; Nath et al. 2016; Wu & Zou 2017; Liu et al. 2018; Zhang et al. 2020).

Earlier studies have shown that AM roots are associated with ROS generation and scavenging; on the other hand, AMF also trigger ROS generation. Moreover, H_2O_2 accumulation in arbuscule-containing cortical cells of roots implies the arbuscule degradation during their life cycle (Wu *et al.* 2014). AMF equipped with antioxidant enzymes and non-enzymatic antioxidants respond to ROS accumulation, thus mitigating the oxidative burst (Lanfranco *et al.* 2005; González-Guerrero *et al.* 2007, 2010; Benabdellah *et al.* 2009a, 2009b). In addition, AMF activate the protective systems of host plants to alleviate oxidative damage under abiotic stress (Wu *et al.* 2014; Kapoor & Singh 2017). The network of plant–AM fungus dialogue for mitigating a ROS burst under drought stress is extremely complex (Fig. 1).

ACCUMULATION OF ROS IS ASSOCIATED WITH THE ESTABLISHMENT OF ARBUSCULAR MYCORRHIZAS

Earlier studies paid great attention to the ROS burst of plants, whereas there is limited information regarding ROS accumulation of mycorrhizal plants, especially in mycorrhiza-containing

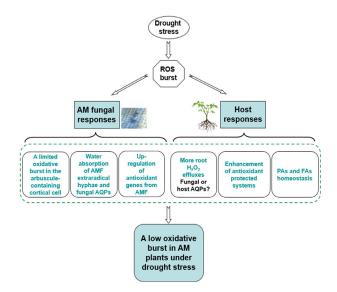


Fig. 1. Schematic diagram of the role of arbuscular mycorrhizal fungi (AMF) in mitigating drought-induced oxidative burst in host plants. Drought stress seriously induces the accumulation of reactive oxygen species (ROS) in plants, thus resulting in an oxidative burst. Mycorrhizal plants have two response patterns to deal with the ROS burst. The first pathway is arbuscular mycorrhizal (AM) fungal responses. AMF produce a limited oxidative burst in the arbuscule-containing root cortical cell. In addition, expression of antioxidant genes from AMF are induced, and AM fungal extraradical hyphae and fungal aquaporins (AQPs) participate in water absorption and transport to hosts, collectively alleviating the drought-induced oxidative burst. The second pathway is host responses, in which antioxidant protective systems, especially antioxidant enzyme gene expression patterns, are enhanced; polyamine (PAs) and fatty acid (FAs) homeostasis is optimized; more root H₂O₂ is effluxed to the rhizosphere. The two approaches sometimes act together. For example, PAs in AMF can be transferred to the host, thus affecting PA homeostasis in hosts, fungal AQPs may participate in root H₂O₂ effluxes, fungal antioxidant genes interact with host antioxidant enzyme genes, collectively enhancing the antioxidant protective system. Thus, the dialogue between both AMF and host plants in mitigating a drought-induced oxidative burst involves a complex network.

root cells. ROS production by AMF is a common phenomenon (Fester & Hause 2005) and is central to the fungal colonization process. ROS levels are dynamic during fungal colonization, symbiosis formation and degradation (Segal & Wilson 2017). Hydrogen peroxide generation in mycorrhiza-containing cortical cells ensures the initial AM fungal colonization in roots, although the generation is transient and quickly eliminated by superoxide dismutase (SOD), catalase (CAT) and carotenoids (Kapoor & Singh 2017). In Medicago truncatula plants, H₂O₂ has been found to accumulate in the arbuscule-containing root cortical cells, where the arbuscules are clumped and less branched, corresponding to the later stage of the arbuscule life cycle (Salzer et al. 1999). When H2O2 is diffused across the hyphal wall of the arbuscule branches, initiation of the fungal programme for senescence occurs (Fester & Hause 2005). Hence, ROS accumulation precedes the degradation of arbuscules. The accumulation of H₂O₂ may be derived from the activation of carotenoid biosynthesis or plant stress responses. In addition to arbuscules, H2O2 has also been found in AMF extraradical hyphal tips, whereas arbuscules were not observed in vesicles and hyphopodia (Salzer et al. 1999). The fungal hyphal cytosol and surface of the intercellular hyphae also accumulated H₂O₂ (Fester & Hause 2005), implying a mycorrhizal protection system in response to stress. Therefore, the oxidative burst would be partly restricted to arbuscule-containing root cells, thereby mitigating the oxidative damage to non-AM cells.

In addition, H₂O₂ was found to be mainly accumulated during the later stage of the development of arbuscules; therefore, H₂O₂ might be a molecule that signals programmed cell death in arbuscules of AM and acts as an indicator of arbuscule degradation (Lanfranco et al. 2005; Wu et al. 2014). Belmondo et al. (2016b) analysed the expression of five respiratory burst oxidase homologue (Rboh, a NADPH oxidase involved in the generation of H_2O_2) genes in the arbuscule-containing root cells of M. truncatula colonized by Gigaspora margarita. Among these five MtRboh genes, only MtRbohE expression was induced in the cell, which was related to an event in the growth of arbuscules. RNAi silencing of MtRbohE further indicated plentiful intercellular hyphae and infrequent arbuscules in the mycorrhizal plants (Belmondo et al. 2016b). In bean (Phaseolus vulgaris), RbohB silencing had a positive effect during the early stages of AM, and PvRbohB was involved in ROS scavenging and hormone homeostasis (Fonseca-García et al. 2019). Under Pb stress, the MtRbohC-G gene of M. truncatula roots was down-regulated by inoculation with Rhizophagus irregularis, which was accompanied by lower H₂O₂ accumulation (Zhang et al. 2019b). These authors conclude that the Rboh gene could mediate ROS homeostasis and thus is associated with arbuscule formation and development (Belmondo et al. 2016a).

ANTIOXIDANT GENES OF AMF PARTICIPATE IN MITIGATING ROS ACCUMULATION

The antioxidant systems in plants can be optimized to remove ROS under drought stress conditions (Ruiz-Lozano *et al.* 2012; Wu & Zou 2017; Wu *et al.* 2019). There is a small amount of evidence that the antioxidant system of AMF contributes to the mitigation of oxidative stress. Several studies have cloned antioxidant genes from AMF, which participate in mitigating ROS accumulation (Table 1). In *Glomus intraradices*, a

AM fungal species	antioxidant genes from AMF	location of AMF	mycorrhizal fungal gene roles	references
Gigaspora margarita	GmarCuZnSOD	Germinated spores and fungal structure in root cells	ROS-inactivating system	Lanfranco <i>et al.</i> (2005)
Glomus intraradices	GintGRX1	Extraradical mycelium	Keeping cytoplasmic proteins in a reduced state, in which H ₂ O ₂ and/or O ₂ are signal molecules	Benabdellah <i>et al.</i> (2009b)
	GintMT1	Extraradical mycelium	Regulation of redox status and protection from ROS	González- Guerrero <i>et al.</i> (2007)
	GintPDX1	Extraradical mycelium	Stimulating vitamin B6 biosynthesis and thus acting as antioxidant of ROS	Benabdellah <i>et al.</i> (2009a)
	GintSOD1	Extraradical mycelium and mycorrhizal roots of Ri-T DNA transformed carrot	Scavenging ROS and a factor for mycorrhizal establishment	González- Guerrero <i>et al.</i> (2010)

Table 1. Antioxidant genes identified from arbuscular mycorrhizal fungi (AMF) and its roles.

metallothionein (MT) gene, GintMT1, was isolated from the extraradical mycelium (González-Guerrero *et al.* 2007). The expression levels of GintMT1 in response to 500 µM paraquat (an intracellular superoxide generator) was up-regulated nine-fold after 7 days of exposure, indicating that GintMT1 has a role as a protectant from oxidative damage. Therefore, GintMT1 may be involved in regulating the redox status of the AMF extraradical hyphae *via* its metal chelation or thiol groups, resulting in the mitigation of ROS accumulation in AMF and their hosts.

In G. margarita, a Cu/Zn SOD gene, named GmarCuZn-SOD, has been identified from germinated spores (Lanfranco et al. 2005). This gene was highly expressed in mycorrhizal root cells, and thus was responsible for localized H₂O₂ accumulation in the intracellular fungal structure, such as collapsing arbuscules. Hence, the fungal SOD possibly controlled plant defence responses to maintain the function of the arbuscules. According to the transcriptome data for G. margarita, ROS scavenger genes in the fungus were identified as associated with the detoxification of ROS, these included glutathione peroxidase, Cu/Zn-SOD, thioredoxin reductase, peroxiredoxins and selenocompound metabolism (Salvioli et al. 2016). Based on proteomic and transcriptomic data, G. margarita with an endobacterium had lower levels of both protein carbonylation and H₂O₂ and higher levels of fungal respiration and total glutathione than those without an endobacterium, indicating that endobacteria elicit mycorrhizal fungal antioxidative activity (Vannini et al. 2016). Further, the enhancement of antioxidant activity in AMF by endobacteria is subsequently transmitted to the host plant, according to analysis of protein carbonylation profiles in the fungal partner. However, it is not clear how AMF transmit this cellular signalling to the host plant.

In the model AM fungus *G. intraradices*, a *GintSOD1* encoding Cu/Zn-SOD maintained this function in scavenging ROS because up-regulation of *GintSOD1* was found after ROS-inducing agents were included (González-Guerrero *et al.* 2010). According to Corradi *et al.* (2009), fungal SOD possesses directional selection for diversity and provides evolutionary advantages in response to stress. Root exudates of *Lotus japonicus* have been found to slightly induce the expression of *Gmar*-CuZnSOD (Lanfranco *et al.* 2005). In brief, fungal SOD acts as an antioxidant enzyme to scavenge ROS, as well as being a component in AMF – plant dialogue for mycorrhizal establishment and plant defence responses.

Benabdellah *et al.* (2009a, 2009b) isolated a *GintPDX1* that encodes a protein involved in vitamin B6 biosynthesis, and that a *GintGRX1* encodes the glutaredoxin from *G. intraradices.* Moreover, *GintPDX1* is expressed in the fungal life cycle, with the highest expression levels observed in intraradical fungal structure, and is induced by exogenous H_2O_2 and ROS-inducing agents (Benabdellah *et al.* 2009a); hence, *GintGRX1* plays a role in alleviating ROS (Benabdellah *et al.* 2009b). Both *GintGRX1* and *GintPDX1* act as antioxidants in protecting the AMF and/or the plants against oxidative damage.

In short, AMF, similar to plants, possess a regulated network of antioxidant enzyme systems (*e.g. GmarCuZnSOD* and *GintSOD1*) and antioxidant systems (*e.g. GintMT1*, *GinPDX1* and *GintGRX1*) for scavenging ROS.

PLANT RESPONSE MECHANISMS ON AMF-MITIGATING ROS BURST UNDER DROUGHT STRESS

Drought stress can induce adverse effects in plants, while plants trigger response mechanisms at morphological (escape and avoidance), physiological (compatible solute accumulation, antioxidant defence systems, phytohormonal homeostasis, etc.) and molecular levels to cope with drought stress (Jangra et al. 2019). Drought escape is a primary adaptation mechanism to avoid stress by improving water uptake and reducing water loss (Nadeem et al. 2019). Plants also accumulate compatible solutes, including soluble sugars, proline, glycine betaine, organic acids, potassium, calcium, etc., to facilitate water uptake by decreasing cytoplasmic osmotic potential (Tani et al. 2019). Antioxidant defence systems in plants are activated to remove further ROS accumulation, thus mitigating the oxidative burst induced by drought stress (Choudhury et al. 2017). As an endophytic fungus, AMF colonize the root system of their hosts and further enhance plant response mechanisms to drought. Here, we only review recent advances in mitigating the ROS burst of host plants under drought stress.

Antioxidant defence systems

A low oxidative burst in AMF-colonized host plants is associated with enhancement of antioxidant enzymes (SOD, CAT, guaiacol peroxidase, glutathione reductase, monodehydroascorbate reductase, dehydroascorbate reductase and ascorbate peroxidase) and non-enzymatic antioxidants (AsA, carotenoids, GSH and α -tocopherol) in the defence system activated by AMF, as reviewed in detail by Bahadur et al. (2019), Lenoir et al. (2016), Kapoor & Singh (2017), Wu et al. (2013, 2014) and Wu & Zou (2017). It has been estimated that AMF increase production of antioxidant enzymes in plants by approximate 16% (Lokhandwala & Hoeksema 2019). In sesame plants exposed to severe drought stress, Funneliformis mosseae and Rhizophagus irregularis collectively enhanced ascorbate peroxidase, polyphenol oxidase, guaiacol peroxidase (G-POD), CAT and phenylalanine ammonia lyase (PAL) activity, followed by an increase in total soluble sugar content (Gholinezhad et al. 2020). Similarly, Langeroodi et al. (2020) showed an enhanced activity of antioxidant defence systems in chicory with R. irregularis, such as SOD, POD, AsA and GSH, accompanied by lower H2O2 accumulation and reduced oxidative damage. Baslam & Giocoechea (2012) further found that AMF improved the accumulation of antioxidant compounds in lettuce (mainly carotenoids and anthocyanins) and that the improvement was larger under drought stress than under wellwatered conditions. Elevation of antioxidant defence systems in mycorrhizal plants under drought stress removes further ROS accumulation, thus mitigating toxicity effects on lipids, proteins and DNA, with significant consequences for plant functioning under stress (Acuña-rodríguez et al. 2020). Thus, mycorrhizal plants have higher drought tolerance than nonmycorrhizal plants through the activation of antioxidant defence systems.

Mycorrhizal Triticum aestivum plants colonized by Glomus mosseae had a higher content of antioxidant secondary metabolites, such as anthocyanins, flavonoids and total phenolic compounds than control plants not inoculated with mycorrhiza when exposed to soil water deficit (Bitaraf et al. 2020). The increase in flavonoids under mycorrhization is due to an increase in PAL activity (Mirjani et al. 2019) and the up-regulated expression of genes involved in the flavonoid biosynthetic pathway (Aseel et al. 2019). The phenylpropanoid pathway was also elicited in mycorrhizal tomato plants (Chialva et al. 2018). It is documented that flavonoids, as antioxidants, directly scavenge ROS, and that flavonoids also stimulate spore germination, mycorrhizal hyphal growth and arbuscule formation within the host roots to establish mycorrhizal fungal activity (Salloum et al. 2018; Mirjani et al. 2019). Total content of phenols was increased by Glomus versiforme in maize under two drought regimes, indicating that AMF-induced phenols, as active scavengers of ROS, protect cellular structures and the functioning of their hosts (Begum et al. 2019). Numerous studies have confirmed that AMF inoculation enhances concentrations of carotenoids in Aloe barbadensis, Citrus aurantifolia, Ricinus communis and Zinnia elegans under drought stress (Zhang et al. 2018b; Bagheri et al. 2019; Khajeeyan et al. 2019; Nejad et al. 2019). An increase in carotenoids in droughtstressed host plants by AMF inoculation could quench more ¹O₂ from excess light energy, thereby protecting the photosynthetic machinery from photo-oxidative damage (Shi et al.

2015). Baslam *et al.* (2013) reported that inoculation with *Glomus fasciculatum* and a commercially mixed mycorrhizal fungal inoculum increased concentrations of major carotenoids, chlorophylls and tocopherols in lettuce, which further eliminated ROS. Under arsenic toxicity, AMF also increased antioxidant (carotenoids and α -tocopherol) concentrations in wheat in response to oxidative stress (Sharma *et al.* 2017). Based on RNA sequence analysis, the AMF cope with oxidative stress *via* induction of multiple metabolic processes, including antioxidant enzyme genes, increased methionine content, sulphur uptake and the pentose phosphate pathway (Venice *et al.* 2017).

Water absorption of mycorrhizal extraradical hyphae and aquaporins

The degree of oxidative burst is closely related to the water status of plants under drought (Bhattacharjee 2005). Under drought stress, the water content in plants decreases, indicating the occurrence of oxidative stress (Arora et al. 2002). AM can form an extraradical mycelium of >100 cm in the plant rhizosphere to enlarge the root surface area for better water acquisition and allow the plant to survive at extremely low soil moisture content (Allen 2006). However, the AMF extraradical mycelium directly takes up water from the soil matrix through the hyphal tips and then transfers this water through the fungal cytoplasm or inner wall layers into root cortical cells, without any obstruction from the plant membrane system (Allen 2007). The water within the hyphae is not lost because of the hydrophobic cell walls. Such mycorrhizal mycelium provides a 'highway' for water extraction from arid soils, and thus plays a critical role in the uptake and transfer of unavailable plant water sources under adverse conditions (Allen 2007). Zhang et al. (2018a) calculated that the water absorption rate of the AMF extraradical hyphae ranged from 0.13 to 1.97 mg $H_2O \cdot h^{-1}$, depending on the AM fungal species, and that the hyphal water absorption rate was dramatically higher under a soil water deficit than in saturated soils. A reduction in hyphal water redistribution (water flows out of the hyphae into the rhizosphere) in the mycorrhizosphere versus the non-mycorrhizosphere was found under a soil water deficit, indicating more water is maintained by mycorrhizal plants (Zou et al. 2018).

Six aquaporin (AQP) genes were identified in G. intraradices and Rhizophagus clarus, where GintAQPF2 and RcAQP3, respectively, were involved in water absorption (Aroca et al. 2009; Li et al. 2013; Kikuchi et al. 2016). Host AQP gene expression can be down-regulated, up-regulated or unchanged by AMF under drought stress, implying a diverse response pattern (He et al. 2019; Zou et al. 2019; Cheng et al. 2020). In Lotus japonicus, Giovannetti et al. (2012) identified two putative AQP genes, LjNIPs and LjXIP1; LiNIP1 was expressed in arbuscule-containing cortical cells of roots, accumulated in the inner membrane system of arbusculated plant cells and could transport water. However, fungal AQP expression patterns are a compensatory method for host AQP expression under drought stress (Aroca et al. 2009; Ding et al. 2020). Therefore, plant and fungal AQP synergistically help roots to absorb water under drought stress, and AM plants are subject to a lower degree of soil drought than non-AM plants, thus having a lower oxidative burst. Future work will revolve around the functional role of host AQP induced by AMF under drought stress in arbusculated plant root cells.

Root H₂O₂ efflux

In plants, root H₂O₂ efflux is an indicator of the degree of oxidative stress (Sousa-Lopes et al. 2004; Huang et al. 2017). In trifoliate orange, inoculation with Funneliformis mosseae caused an increase in H₂O₂ efflux in plant roots under drought stress, which was dependent on root class and region (Zou et al. 2015; Huang et al. 2017). The negative correlation between root net H2O2 efflux and root H2O2 or malondialdehyde concentration suggests a low oxidative burst in mycorrhizal plants exposed to drought stress (Fig. 1) (Huang et al. 2017). AMF inoculation under soil drought induced an increase in root calmodulin (CaM) and Ca²⁺ influx (Huang et al. 2014; Zou et al. 2015), and Ca²⁺ influx was positively correlated with H₂O₂ efflux (Zou et al. 2015), indicating that Ca^{2+}/CaM may initiate H_2O_2 signal transduction to respond to H₂O₂ efflux. Because of the similar electrochemical properties of H₂O and H₂O₂, some AQP proteins, e.g. an aquaporin gene (So-aqpA) isolated from Streptococcus oligofermentans, was a bidirectional H₂O₂-transferring AQP, and thus AQP-modulated H₂O₂ efflux is an important mechanism in ROS detoxification (Tong et al. 2019). It is not clear whether the AQP genes of both the AMF and host are involved in root H₂O₂ efflux.

Up-regulation of antioxidant enzyme gene expression

Antioxidant enzyme gene (e.g. SOD, POD and CAT) expression in lettuce, trifoliate orange and wheat is activated under a combination of the presence of AMF and plant drought stress (Ruiz-Lozano et al. 2001; Mustafa et al. 2017; Zhang et al. 2019a; He et al. 2020). The expression of CAT and POD in arbuscule-containing cells colonized by G. intraradices was upregulated (Blee & Anderson 2000), suggesting that mycorrhizas stimulated localized defence responses in mycorrhizal plant roots. Under drought stress, Mn-sod II expression in lettuce plants was induced by G. mosseae and G. intraradices, which protected mycorrhizal plants against oxidative stress (Ruiz-Lozano et al. 2001). Similarly, AMF up-regulated expression levels of PtFe-SOD, PtMn-SOD, PtPOD and PtCAT1 in trifoliate orange exposed to drought stress, thus maintaining low ROS levels (e.g. H₂O₂) (Zhang et al. 2019a; He et al. 2020). Relative expression of the phenylalanine ammonia-lyase gene was higher in roots of micropropagated plantlets of Satureja khuzistanica inoculated with Rhizoglomus fasciculatum than that in non-AMF inoculated control plantlets on days 7 and 42; thus monitoring flavonoid biosynthesis for low oxidative damage (Mirjani et al. 2019). The improved tolerance of AMF-inoculated sunflower under high temperatures was due to up-regulated expression of the glutathione S-transferase gene (Mayer et al. 2017). Hence, the above results suggest that AMF trigger the overexpression of host antioxidant enzyme genes in response to drought stress (Fig. 1), thereby leading to reduced accumulation of ROS in host plants (He et al., 2020).

Polyamines and fatty acids

Polyamines (PAs) are low-molecular weight aliphatic polycations, and three main types are found in plants: spermine (Spm), spermidine (Spd) and putrescine (Put) (Liu et al. 2015). PAs can bind anionic macromolecules (e.g. nucleic acids and proteins) to regulate transcription and translation, as well as maintain membrane stability and modulate antioxidant systems (Liu et al. 2015). PA degradation can induce H₂O₂ generation (Pottosin et al. 2014); hence, the relationship between PAs and ROS is complex. Zhang et al. (2020) found higher Put concentrations and lower Spd and Spm concentrations in drought-stressed trifoliate orange after inoculation with F. mosseae, accompanied by the up-regulation of PA synthase genes and PA catabolic enzyme genes. Changes in PAs through mycorrhization trigger ROS signals for stress responses via low H₂O₂ accumulation by up-regulating PtMn-SOD, PtCu/Zn-SOD and PtCAT1 (Fig. 1). In addition, Put catabolism, together with an improvement in N assimilation and the accumulation of γ -aminobutyric acid, was found in mycorrhizal maize plants under drought stress (Hu & Chen 2020). AM plants contain higher PA levels under stress conditions, which further improve root growth, thereby better responding to stress (Sannazzaro et al. 2007). Exogenous Put in combination with AMF is an effective strategy for coping with nodule senescence in leguminous plants under stress (Garg & Sharma 2019) and cadmium toxicity in sorghum (Kumar & Pathak 2019) because Put is an important stimulus for arbuscule formation and mycorrhizal colonization (Salloum et al. 2018). Therefore, PAs, especially Put, may inhibit the auto-oxidation of metals, impairing the supply of electrons for ROS generation, and directly acting as ROS scavengers or inducing the initiation of antioxidant systems (Liu et al. 2015).

In addition, fatty acids (FAs) located in the plant cell membrane are a major determinant of membrane integrity and function (Wu et al. 2018). Lipoxygenase located in the cell walls can lead to hydroperoxidation of polyunsaturated FAs for ROS generation. FAs play an important role in maintaining AM growth (Luginbuehl et al. 2017). After 4 days of AMF inoculation, FA desaturation of host plants was found in response to AMF colonization (Vangelisti et al. 2020). Wu et al. (2019) reported that under drought stress, FA components in the roots of trifoliate orange had a decreased content of saturated FAs and an increased content of unsaturated FAs with F. mosseae colonization, accompanied by up-regulation of FA desaturase genes (e.g. PtFAD2, PtFAD6 and Pt Δ 9). Therefore, mycorrhizal plants had a relatively higher unsaturation index for FAs than non-mycorrhizal plants grown under soil water deficit (Wu et al. 2019). RNA sequencing further revealed that the most differentially expressed genes in mycorrhizal plants were associated with lipid metabolic processes under drought stress (Tarnabi et al. 2020). AMF colonization induced high cell membrane lipid fluidity, improved membrane integrity and cell wall rigidity of host plants (Essahibi et al. 2018), which could then cope with an oxidative burst. Mycorrhiza-mediated FA changes in components and their unsaturation levels are a precondition for reducing the oxidative burst in host plants under drought stress (Fig. 1).

CONCLUSIONS AND OUTLOOKS

Arbuscular mycorrhizal fungi can potentially mitigate effects of drought stress on their host plants, and the proposed mechanisms involved have been reviewed previously (Wu *et al.* 2013; Lenoir *et al.* 2016). Understanding these mechanisms will enable better use of AMF to protect crops against drought stress. Previous studies have revealed the role of antioxidant defence systems in physiological and molecular responses to AM under drought stress, whereas the regulative network for ROS in antioxidant defence systems still needs to be elucidated based on 'omics' techniques. For example, an endobacterium of G. margarita elicited fungal antioxidant activity, and then the fungus transmitted this to the host plant through signalling pathways in the system plant-AM and fungus-endobacterium (Vannini et al. 2016). It is not clear whether ROS scavenging enzyme genes from both AMF and hosts synergistically enhance drought tolerance of host plants. However, arbuscule-containing plant cells modulate the synthesis of pathogenesis-related protein PR-b1 and the distribution of hydroxyproline-rich glycoprotein, and thus the existence of arbuscules in root cortical cells is associated with plant defence responses (Song et al. 2011). Future in-depth studies on changes in ROS production and antioxidant defence systems in arbuscule-containing plant root cortical cells will help to understand the regulatory network of mycorrhizas in tolerating drought stress.

In AMF, *GintMT1*, *GintSOD1* and *GmarCuZnSOD* genes were identified and their functions partly clarified in AMF and/ or plants. In scavenging ROS, SOD are the first line of defence for removing $O_2^{\bullet-}$ in H_2O_2 generation. Therefore, more antioxidant enzyme genes must be identified from AMF

REFERENCES

- Acuña-Rodríguez I.S., Newsham K.K., Gundel P.E., Torres-Díaz C., Molina-Montenegro M.A. (2020) Functional roles of microbial symbionts in plant cold tolerance. *Ecology Letters*, 23, 1034–1048.
- Allen M.F. (2006) Water dynamics of mycorrhizas in arid soils. In: Gadd G. M. (Ed), *Fungi in biogeochemical cycles*. Cambridge University Press, Cambridge, UK, pp 74–97.
- Allen M.F. (2007) Mycorrhizal fungi: highways for water and nutrients in arid soils. Vadose Zone Journal, 6, 291–297.
- Aroca R., Bago A., Sutka M., Paz J.A., Cano C., Amodeo G., Ruiz-Lozano J.M. (2009) Expression analysis of the first arbuscular mycorrhizal fungi aquaporin described reveals concerted gene expression between salt-stressed and non-stressed mycelium. *Molecular Plant-Microbe Interactions*, 22, 1169–1178.
- Arora A., Sairam R.K., Srivastava G.C. (2002) Oxidative stress and antioxidative system in plants. *Current Science*, **82**, 1227–1238.
- Aseel D.G., Rashad Y.M., Hammad S.M. (2019) Arbuscular mycorrhizal fungi trigger transcriptional expression of flavonoid and chlorogenic acid biosynthetic pathway genes in tomato against *Tomato Mosaic Virus. Scientific Reports*, **9**, 9692.
- Bagheri V., Shamshiri M.H., Alaei H., Salehi H. (2019) The role of inoculum identity for growth, photosynthesis, and chlorophyll fluorescence of zinnia plants by arbuscular mycorrhizal fungi under varying water regimes. *Photosynthetica*, 57, 409–419.
- Bahadur A., Batool A., Nasir F., Jiang S.J., Qin M.S., Zhang Q., Pan J.B., Liu Y.J., Feng F.Y. (2019) Mechanistic insights into arbuscular mycorrhizal fungimediated drought stress tolerance in plants. *International Journal of Molecular Sciences*, 20, 4199.
- Bashri G., Prasad S.M. (2016) Exogenous IAA differentially affects growth, oxidative stress and antioxidant

and analysed to reveal their functions. The gene *RbohE* from *M. truncatula* is involved in AM development, whereas the role of *RbohE* in AMF is unknown. Further studies should be undertaken on the role of *RbohE* from AMF and fungal partners to clarify the function of this gene in AM.

The H_2O_2 derived from polyphenol oxidase-mediated oxidation is a signal for stomatal closure, and thus polyphenol oxidase must be part of the signalling mechanism that induces plant defence responses (Sequera-Mutiozabal *et al.* 2016). It is necessary to clarify the relationships among polyphenol oxidase, H_2O_2 and stomatal closure.

The AMF enhance plant tolerance to drought stress, depending on the ecotype of the AMF strain used. AMF strains isolated from arid land confer different metabolic pathways and compounds in response to stress (Rivero *et al.* 2018). Hence, additional studies are required to compare the mechanisms of the plasticity in ROS generation and elimination in host plants inoculated with different AMF species.

ACKNOWLEDGEMENTS

This study was supported by the National Key Research and Development Program of China (2018YFD1000300) and the Plan in Scientific and Technological Innovation Team of Outstanding Young Scientists, Hubei Provincial Department of Education (T201604).

system in Cd stressed *Trigonella foenum-graceum* L. seedlings: toxicity alleviation by up-regulation of ascorbate-glutathione cycle. *Ecotoxicology and Environmental Safety*, **132**, 329–338.

- Baslam M., Giocoechea N. (2012) Water deficit improved the capacity of arbuscular mycorrhizal fungi (AMF) for inducing the accumulation of antioxidant compounds in lettuce leaves. *Mycorrhiza*, 22, 347–359.
- Baslam M., Esteban R., Garcia-Plazaola J.I., Goicoechea N. (2013) Effectiveness of arbuscular mycorrhizal fungi (AMF) for inducing the accumulation of major carotenoids, chlorophylls and tocopherol in green and red leaf lettuces. Applied Microbiology and Biotechnology, 97, 3119–3128.
- Begum N., Ahanger M.A., Su Y.Y., Lei Y.F., Mustafa N.S.A., Ahmad P., Zhang L.X. (2019) Improved drought tolerance by AMF inoculation in maize (*Zea mays*) involves physiological and biochemical implications. *Plants*, 8, 579.
- Belmondo S., Calcagno C., Genre A., Puppo A., Pauly N., Lanfranco L. (2016a) NADPH oxidases in the arbuscular mycorrhizal symbiosis. *Plant Signaling* and Behavior, 11, e1165379.
- Belmondo S., Calcagno C., Genre A., Puppo A., Pauly N., Lanfranco L. (2016b) The *Medicago truncatula MtRbohE* gene is activated in arbusculated cells and is involved in root cortex colonization. *Planta*, 243, 251–262.
- Benabdellah K., Azcón-Aguilar C., Valderas A., Speziga D. (2009a) *GintPDX1* encodes a protein involved in vitamin B6 biosynthesis that is up-regulated by oxidative stress in the arbuscular mycorrhizal fungus *Glomus intraradices. New Phytologist*, **184**, 682–693.
- Benabdellah K., Merlos M.A., Azcón-Aguilar C., Ferrol N. (2009b) GintGRX1, the first characterized glomeromycotan glutaredoxin, is a multifunctional enzyme that responds to oxidative stress. Fungal Genetics and Biology, 46, 94–103.

- Bhattacharjee S. (2005) Reactive oxygen species and oxidative burst: Roles in stress, senescence and signal transduction in plants. *Current Science*, **89**, 1113– 1121.
- Bhattacharjee S. (2019) ROS and oxidative stress: origin and implication. In: Bhattacharjee S. (Ed), *Reactive oxygen species in plant biology*. Springer, New Delhi, India, pp 1–32.
- Bitaraf N., Saadatmand S., Mehregan I., Ahmadvand R., Ebadi M. (2020) Evaluation of mitigation effects of *Glomus mosseae* on *Triticum aestivum* L., cv. Chamran under drought stress. *Periodico Tche Quimica*, **17**, 1033–1045.
- Blee K.A., Anderson A.J. (2000) Defence responses in plants to arbuscular mycorrhizal fungi. In: Podila G. K., Douds D. D. (Eds), *Current advances in mycorrhizae research*. The American Phytopathological Society, St Paul, MN, USA, pp 27–44.
- Bonfante P. (2018) The future has roots in the past: the ideas and scientists that shaped mycorrhizal research. New Phytologist, 220, 982–995.
- Cheng H.Q., Ding Y.E., Shu B., Zou Y.N., Wu Q.S., Kuča K. (2020) Plant aquaporin responses to mycorrhizal symbiosis under abiotic stress. *International Journal of Agriculture and Biology*, 23, 786–794.
- Chialva M., Salvioli di Fossalunga A., Daghino S., Ghignone S., Bagnaresi P., Chiapello M., Novero M., Spadaro D., Perotto S., Bonfante P. (2018) Native soils with their microbiotas elicit a state of alert in tomato plants. *New Phytologist*, **220**, 1296–1308.
- Chiu C.H., Choi J., Paszkowski U. (2018) Independent signalling cues underpin arbuscular mycorrhizal symbiosis and large lateral root induction in rice. *New Phytologist*, 217, 552–557.
- Choudhary K.K., Chaudhary N., Agrawal S.B., Agrawal M. (2018) Reactive oxygen species: generation, damage, and quenching in plants during stress. In: Singh V. P., Singh S., Tripathi D. K., Prasad S. M., Chauhan D. K. (Eds), *Reactive oxygen species in plants:*

boon or bane – revisiting the role of ROS. Wiley, Hoboken, NJ, USA, pp 89–116.

- Choudhury F.K., Rivero R.M., Blumwald E., Mittler R. (2017) Reactive oxygen species, abiotic stress and stress combination. *The Plant Journal*, **90**, 856–867.
- Corradi N., Buffner B., Croll D., Colard A., Horak A., Sanders I.R. (2009) High-level molecular diversity of copper-zinc superoxide dismutase genes among and within species of arbuscular mycorrhizal fungi. *Applied and Environmental Microbiology*, 75, 1970– 1978.
- Das K., Roychoudhury A. (2014) Reactive oxygen species (ROS) and response of antioxidants as ROSscavengers during environmental stress in plants. *Frontiers in Environmental Science*, 2, 53.
- Ding Y.E., Fan Q.F., He J.D., Wu H.H., Zou Y.N., Wu Q.S., Kuča K. (2020) Effects of mycorrhizas on physiological performance and root TIPs expression in trifoliate orange under salt stress. Archives of Agronomy and Soil Science, 66, 182–192.
- Essahibi A., Benhiba L., Babram M.A., Ghoulam C., Qaddoury A. (2018) Influence of arbuscular mycorrhizal fungi on the functional mechanisms associated with drought tolerance in carob (*Ceratonia siliqua* L.). *Trees – Structure and Function*, **32**, 87–97.
- Fester T., Hause G. (2005) Accumulation of reactive oxygen species in arbuscular mycorrhizal roots. *Mycorrhiza*, 15, 373–379.
- Fonseca-García C., Zayas A.E., Montiel J., Nava N., Sánchez F., Quinto C. (2019) Transcriptome analysis of the differential effect of the NADPH oxidase gene *RbohB* in *Phaseolus vulgaris* roots following *Rhizobium tropici* and *Rhizophagus irregularis* inoculation. *BMC Genomics*, 20, 800.
- Garg N., Sharma A. (2019) Role of putrescine (Put) in imparting salt tolerance through modulation of Put metabolism, mycorrhizal and rhizobial symbioses in *Cajanus cajan* (L.) Millsp. *Symbiosis*, **79**, 59–74.
- Gavito M.E., Jakobsen I., Mikkelsen T.N., Mora F. (2019) Direct evidence for modulation of photosynthesis by an arbuscular mycorrhiza-induced carbon sink strength. *New Phytologist*, 223, 896–907.
- Gholinezhad E., Darvishzadeh R., Moghaddam S.S., Popovic-Djordjevic J. (2020) Effect of mycorrhizal inoculation in reducing water stress in sesame (Sesamum indicum L.): the assessment of agrobiochemical traits and enzymatic antioxidant activity. Agricultural Water Management, 238, 106234.
- Giovannetti M., Balestrini R., Volpe V., Guether M., Straub D., Costa A., Ludewig U., Bonfante P. (2012) Two putative aquaporin genes are differentially expressed during arbuscular mycorrhizal symbiosis in *Lotus japonicus. BMC Plant Biology*, **12**, 186.
- González-Guerrero M., Cano C., Azcón-Aguilar C., Ferrol N. (2007) *GintMT1* encodes a functional metallothionein in *Glomus intraradices* that responds to oxidative stress. *Mycorrhiza*, 17, 327–335.
- González-Guerrero M., Oger E., Benabdellah K., Azcon-Aguilar C., Lanfranco L., Ferrol N. (2010) Characterization of a CuZn superoxide dismutase gene in the arbuscular mycorrhizal fungus *Glomus intraradices*. *Current Genetics*, **56**, 265–274.
- Guo H., Cui Y.N., Pan Y.Q., Wang S.M., Bao A.K. (2020) Sodium chloride facilitates the secretohalophyte Atriplex canescens adaptation to drought stress. Plant Physiology and Biochemistry, 150, 99– 108.
- He J.D., Dong T., Wu H.H., Zou Y.N., Wu Q.S., Kuča K. (2019) Mycorrhizas induce diverse responses of root *TIP* aquaporin gene expression to drought

stress in trifoliate orange. *Scientia Horticulturae*, **243**, 64–69.

- He J.D., Zou Y.N., Wu Q.S., Kuča K. (2020) Mycorrhizas enhance drought tolerance of trifoliate orange by enhancing activities and gene expression of antioxidant enzymes. *Scientia Horticulturae*, 262, 108745.
- Hu Y.B., Chen B.D. (2020) Arbuscular mycorrhiza induced putrescine degradation into γ-aminobutyric acid, malic acid accumulation, and improvement of nitrogen assimilation in roots of water-stressed maize plants. *Mycorrhiza*, **30**, 329–339.
- Huang Y.M., Srivastava A.K., Zou Y.N., Ni Q.D., Han Y., Wu Q.S. (2014) Mycorrhizal-induced calmodulin mediated changes in antioxidant enzymes and growth response of drought-stressed trifoliate orange. *Frontiers in Microbiology*, 5, 682.
- Huang Y.M., Zou Y.N., Wu Q.S. (2017) Alleviation of drought stress by mycorrhizas is related to increased root H₂O₂ efflux in trifoliate orange. *Scientific Reports*, 7, 42335.
- Humphreys C.P., Franks P.J., Rees M., Bidartondo M.I., Leake J.P., Beerling D.J. (2010) Mutualistic mycorrhiza-like symbiosis in the most ancient group of land plants. *Nature Communications*, 1, 103.
- Jangra S., Mishra A., Priti Kamboj D., Yadav N.R., Yadav R.C. (2019) Plant responses and tolerance to drought. In: Hasanuzzaman M., Nahar K., Fujita M., Oku H., Tofazzal Islam M. (Eds), Approaches for enhancing abiotic stress tolerance in plants. CRC Press, Boca Raton, FL, USA, pp 79–98.
- Kapoor R., Singh N. (2017) Arbuscular mycorrhiza and reactive oxygen species. In: Wu Q. S. (Ed), Arbuscular mycorrhizas and stress tolerance of plants. Springer, Singapore, pp 225–243.
- Khajeeyan R., Salehi A., Dehnavi M.M., Farajee H., Kohanmoo M.A. (2019) Physiological and yield responses of *Aloe vera* plant to biofertilizers under different irrigation regimes. *Agricultural Water Management*, 225, 105768.
- Kikuchi Y., Hijikata N., Ohtomo R., Handa Y., Kawaguchi M., Saito K., Masuta C., Ezawa T. (2016) Aquaporin-mediated long-distance polyphosphate translocation directed towards the host in arbuscular mycorrhizal symbiosis: application of virus-induced gene silencing. *New Phytologist*, **211**, 1202–1208.
- Kumar P., Pathak S. (2019) Responsiveness index of sorghum (Sorghum bicolor (L.) Moench) grown under cadmium contaminated soil treated with putrescine and mycorrhiza. Bangladesh Journal of Botany, 48, 139–143.
- Lanfranco L., Novero M., Bonfante P. (2005) The mycorrhizal fungus Gigaspora margarita possesses a CuZn superoxide dismutase that is up-regulated during symbiosis with legume hosts. Plant Physiology, 137, 1319–1330.
- Lanfranco L., Fiorilli V., Gutjahr C. (2018) Partner communication and role of nutrients in the arbuscular mycorrhizal symbiosis. *New Phytologist*, 220, 1031–1046.
- Langeroodi A.R.S., Osipitan O.A., Radicetti E., Mancinelli R. (2020) To what extent arbuscular mycorrhiza can protect chicory (*Cichorium intybus* L.) against drought stress. *Scientia Horticulturae*, 263, 109109.
- Lenoir I., Fontaine J., Sahraoui A.L.H. (2016) Arbuscular mycorrhizal fungal responses to abiotic stresses: a review. *Phytochemistry*, **123**, 4–15.
- Li T., Hu Y.J., Hao Z.P., Li H., Wang Y.S., Chen B.D. (2013) First cloning and characterization of two

functional aquaporin genes from an arbuscular mycorrhizal fungus *Glomus intraradices*. *New Phytologist*, **197**, 617–630.

- Liu J.H., Wang W., Wu H., Gong X., Moriguchi T. (2015) Polyamines function in stress tolerance: from synthesis to regulation. *Frontiers in Plant Science*, 6, 827.
- Liu C.Y., Zhang F., Zhang D.J., Srivastava A.K., Wu Q.S., Zou Y.N. (2018) Mycorrhiza stimulates roothair growth and IAA synthesis and transport in trifoliate orange under drought stress. *Scientific Reports*, 8, 1978.
- Lokhandwala A., Hoeksema J.D. (2019) Priming by arbuscular mycorrhizal fungi of plant antioxidant enzyme production: a meta-analysis. *Annual Plant Reviews*, 2, 1069–1084.
- Luginbuehl L.H., Menard G.N., Kurup S., Van Erp H., Radhakrishnan G.V., Breakspear A., Oldroyd G.E.D., Eastmond P.J. (2017) Fatty acids in arbuscular mycorrhizal fungi are synthesized by the host plant. *Science*, **356**, 1175–1178.
- Mayer Z., Duc N.H., Sasvari Z., Posta K. (2017) How arbuscular mycorrhizal fungi influence the defense system of sunflower during different abiotic stresses. *Acta Biologica Hungarica*, **68**, 376–387.
- Millar N.S., Bennett A.E. (2016) Stressed out symbiotes: hypotheses for the influence of abiotic stress on arbuscular mycorrhizal fungi. *Oecologia*, 182, 625–641.
- Mirjani L., Salimi A., Matinizadeh M., Razavi K., Shahbazi M. (2019) The role of arbuscular mycorrhizal fungi on acclimatization of micropropagated plantlet *Satureja khuzistanica* Jam. by ameliorating of antioxidant activity and expression of *PAL* gene. *Scientia Horticulturae*, 253, 364–370.
- Mustafa G., Khong N.G., Tisserant B., Randoux B., Fontaine J., Magnin-Robert M., Reignault P., Sahraoui A.L.H. (2017) Defence mechanisms associated with mycorrhiza-induced resistance in wheat against powdery mildew. *Functional Plant Biology*, 44, 443– 454.
- Nadeem M., Li J.J., Yahya M., Sher A., Ma C.X., Wang X.B., Qiu L.J. (2019) Research progress and perspective on drought stress in legumes: a review. *International Journal of Molecular Sciences*, **10**, 2541.
- Nath M., Bhatt D., Prasad R., Gill S.S., Anjum N.A., Tuteja N. (2016) Reactive oxygen species generation scavenging and signaling during plant–arbuscular mycorrhizal and *Piriformospora indica* interaction under stress condition. *Frontiers in Plant Science*, 7, 1574.
- Nejad K.Z., Ghasemi M., Shamili M., Damizadeh G.R. (2019) Effect of mycorrhiza and vermicompost on drought tolerance of lime seedlings (*Citrus aurantifolia* cv. Mexican Lime). *International Journal of Fruit Science*. https://doi.org/10.1080/15538362. 2019.1678448
- Pottosin I., Velarde-Buendia A.M., Bose J., Zepeda-Jazo I., Shabala S., Dobrovinskaya O. (2014) Crosstalk between reactive oxygen species and polyamines in regulation of ion transport across the plasma membrane: implications for plant adaptive responses. *Journal of Experimental Botany*, 65, 1271– 1283.
- Qiao Y.G., Bai Y.X., Zhang Y.Q., She W.W., Lai Z.R., Qin S.G. (2019) Arbuscular mycorrhizal fungi shape the adaptive strategy of plants by mediating nutrient acquisition in a shrub-dominated community in the Mu Us Desert. *Plant and Soil*, **443**, 549–564.

AMF and oxidative burst of drought-stressed plants

- Rivero J., Alvarez D., Flors V., Azcon-Aguilar C., Pozo M.J. (2018) Root metabolic plasticity underlies functional diversity in mycorrhiza-enhanced stress tolerance in tomato. *New Phytologist*, **220**, 1322–1336.
- Ruiz-Lozano J.M., Collados C., Barea J.M., Azcón R. (2001) Cloning of cDNAs encoding SODs from lettuce plants which show differential regulation by arbuscular mycorrhizal symbiosis and by drought stress. *Journal of Experimental Botany*, 52, 2241–2242.
- Ruiz-Lozano J.M., Porcel R., Azcón C., Aroca R. (2012) Regulation by arbuscular mycorrhizae of the integrated physiological response to salinity in plants: new challenges in physiological and molecular studies. *Journal of Experimental Botany*, 63, 4033– 4044.
- Salloum M.S., Menduni M.F., Benavides M.P., Larrauri M., Luna C.M., Silvente S. (2018) Polyamines and flavonoids: key compounds in mycorrhizal colonization of improved and unimproved soybean genotypes. Symbiosis, 76, 265–275.
- Salvioli A., Ghignone S., Novero M., Navazio L., Venice F., Bagnaresi P., Bonfante P. (2016) Symbiosis with an endobacterium increases the fitness of a mycorrhizal fungus, raising its bioenergetic potential. *The ISME Journal*, **10**, 130–144.
- Salzer P., Corbiere H., Boller T. (1999) Hydrogen peroxide accumulation in *Medicago truncatula* roots colonized by the arbuscular mycorrhiza-forming fungus *Glomus intraradices*. *Planta*, **208**, 319–325.
- Sannazzaro A.I., Echeverria M., Alberto E.O., Ruiz O.A., Menendez A.B. (2007) Modulation of polyamine balance in *Lotus glaber* by salinity and arbuscular mycorrhiza. *Plant Physiology and Biochemistry*, 45, 39–46.
- Segal L.M., Wilson R.A. (2017) Reactive oxygen species metabolism and plant–fungal interactions. *Fungal Genetics and Biology*, **110**, 1–9.
- Sequera-Mutiozabal M., Tiburcio A.F., Alcázar R. (2016) Drought stress tolerance in relation to polyamine metabolism in plants. In: Hossain M., Wani S., Bhattacharjee S., Burritt D., Tran L. S. (Eds), Drought stress tolerance in plants, Vol 1. Springer, Cham, Switzerland, pp 267–286.
- Sharma S., Anand G., Singh N., Kapoor R. (2017) Arbuscular mycorrhiza augments arsenic tolerance in wheat (*Triticum aestivum* L.) by strengthening antioxidant defense system and thiol metabolism. *Frontiers in Plant Science*, 8, 906.
- Shi Y.M., Guo J.G., Zhang W., Jin L.F., Liu P.P., Chen X., Li F., Wei P., Li Z.F., Li W.Z., Wei C.Y., Zheng Q.X., Chen Q.S., Zhang J.F., Lin F.C., Qu L.B., Snyder J.H., Wang R. (2015) Cloning of the *lycopene* βcyclase gene in *Nicotiana tabacum* and its overexpression confers salt and drought tolerance. *International Journal of Molecular Sciences*, 16, 30438– 30457.
- Song F.Q., Song G., Dong A.R., Kong X.S. (2011) Regulatory mechanisms of host plant defense responses to arbuscular mycorrhiza. *Acta Ecologica Sinica*, 31, 322–327.
- Sousa-Lopes A., Antunes F., Cyrne L., Marinho H.S. (2004) Decreased cellular permeability to H₂O₂

protects *Saccharomyces cerevisiae* cells in stationary phase against oxidative stress. *FEBS Letters*, **587**, 152–156.

- Strullu-Derrien C., Kenrick P., Selosse M.A. (2017) Origins of the mycorrhizal symbioses. In: Martin F. (Ed), *Molecular mycorrhizal symbiosis*. Wiley, Hoboken, NJ, USA, pp 3–20.
- Tani E., Chronopoulou E.G., Labrou N.E., Sarri E., Goufa M., Vaharidi X., Tornesaki A., Psychogiou M., Bebeli P.J., Abraham E.M. (2019) Growth, physiological, biochemical, and transcriptional responses to drought stress in seedlings of *Medicago sativa* L., *Medicago arborea* L., and their hybird (alborea). *Agronomy*, 9, 38.
- Tarnabi Z.M., Iranbakhsh A., Mehregan I., Ahmadvand R. (2020) Impact of arbuscular mycorrhizal fungi (AMF) on gene expression of some cell wall and membrane elements of wheat (*Triticum aestivum* L.) under water deficit using transcriptome analysis. *Physiology and Molecular Biology of Plants*, 26, 143–162.
- Tiwari S., Tiwari S., Singh M., Singh A., Prasad S.M. (2018) Generation mechanisms of reactive oxygen species in the plant cell: an overview. In: Singh V. P., Singh S., Tripathi D. K., Prasad S. M., Chauhan D. K. (Eds), *Reactive oxygen species in plants: boon or bane – revisiting the role of ROS*. Wiley, Hoboken, NJ, USA, pp 1–22.
- Tong H.C., Wang X.H., Dong Y.Z., Hu Q.Q., Zhao Z.Y., Zhu Y., Dong L.X., Bai F., Dong X.Z. (2019) A *Streptococcus* aquaporin acts as peroxiporin for efflux of cellular hydrogen peroxide and alleviation of oxidative stress. *Journal of Biological Chemistry*, 294, 4583–4595.
- Vangelisti A., Turrini A., Sbrana C., Avio L., Giordani T., Natali L., Giovannetti M., Cavallini A. (2020) Gene expression in *Rhizoglomus irregulare* at two different time points of mycorrhiza establishment in *Helianthus annuus* roots, as revealed by RNA-seq analysis. *Mycorrhiza*, **30**, 373–387.
- Vannini C., Carpentieri A., Salvioli A., Novero M., Marsoni M., Testa L., de Pinto M.C., Amoresano A., Ortolani F., Bracale M., Bonfante P. (2016) An interdomain network: the endobacterium of a mycorrhizal fungus promotes antioxidative responses in both fungal and plant hosts. *New Phytologist*, 211, 265–275.
- Venice F., de Pinto M.C., Novero M., Ghignone S., Salvioli A., Bonfante P. (2017) *Gigaspora margarita* with and without its endobacterium shows adaptive responses to oxidative stress. *Mycorrhiza*, 27, 747– 759.
- Wu Q.S., Zou Y.N. (2017) Arbuscular mycorrhizal fungi and tolerance of drought stress in plants. In: Wu Q. S. (Ed), Arbuscular mycorrhizas and stress tolerance of plants. Springer, Singapore, pp 25–42.
- Wu Q.S., Srivastava A.K., Zou Y.N. (2013) AMF-induced tolerance to drought stress in citrus: a review. *Scientia Horticulturae*, 164, 77–87.
- Wu Q.S., Zou Y.N., Abd-Allah E.F. (2014) Mycorrhizal association and ROS in plants. In: Ahmad P. (Ed),

Oxidative damage to plants: antioxidant networks and signaling. Academic Press, London, UK, pp 453–475.

- Wu J.X., Chu Z.J., Ruan Z., Wang X.Y., Dai T.H., Hu X.Q. (2018) Changes of intracellular porphyrin, reactive oxygen species, and fatty acid profiles during inactivation of methicillin-resistant *Staphylococcus aureus* by antimicrobial blue light. *Frontiers in Physiology*, 9, 1658.
- Wu Q.S., He J.D., Srivastava A.K., Zou Y.N., Kuča K. (2019) Mycorrhizas enhance drought tolerance of citrus by altering root fatty acid compositions and their saturation levels. *Tree Physiology*, **39**, 1149– 1158.
- Yang Z., Chi X.Y., Guo F.F., Jin X.Y., Luo H.L., Hawar A., Chen Y.X., Feng K.K., Wang B., Qi J.L., Yang Y.H., Sun B. (2020a) SbWRKY30 enhances the drought tolerance of plants and regulates a drought stress-responsive gene, *SbRD19*, in sorghum. *Journal* of *Plant Physiology*, **246–247**, 153142.
- Yang C.F., Huang Y.Z., Lv W.H., Zhang Y.Y., Bhat J.A., Kong J.J., Xing H., Zhao J.M., Zhao T.J. (2020b) GmNAC8 acts as a positive regulator in soybean drought stress. *Plant Science*, **293**, 110442.
- Zhang F., Zou Y.N., Wu Q.S. (2018a) Quantitative estimation of water uptake by mycorrhizal extraradical hyphae in citrus under drought stress. *Scientia Horticulturae*, 229, 132–136.
- Zhang T., Hu Y.J., Zhang K., Tian C.Y., Guo J.X. (2018b) Arbuscular mycorrhizal fungi improve plant growth of *Ricinus communis* by altering photosynthetic properties and increasing pigments under drought and salt stress. *Industrial Crops and Products*, **117**, 13–19.
- Zhang F., Zou Y.N., Wu Q.S. (2019a) Effects of Funneliformis mosseae on the expression of antioxidant enzyme genes in trifoliate orange exposed to drought stress. Mycosystema, 38, 2043–2050. (in Chinese with English abstract)
- Zhang X., Zhang H., Lou X., Tang M. (2019b) Mycorrhizal and non-mycorrhizal *Medicago truncatula* roots exhibit differentially regulated NADPH oxidase and antioxidant response under Pb stress. *Envi*ronmental and Experimental Botany, **164**, 10–19.
- Zhang F., Zou Y.N., Wu Q.S., Kuča K. (2020) Arbuscular mycorrhizas modulate root polyamine metabolism to enhance drought tolerance of trifoliate orange. *Environmental and Experimental Botany*, 171, 103962.
- Zou Y.N., Huang Y.M., Wu Q.S., He X.H. (2015) Mycorrhiza-induced lower oxidative burst is related with higher antioxidant enzyme activities, net H_2O_2 effluxes, and Ca^{2+} influxes in trifoliate orange roots under drought stress. *Mycorrhiza*, **25**, 143–152.
- Zou Y.N., Srivastava A.K., Wu Q.S. (2018) Water redistribution in mycorrhizosphere of trifoliate orange. *Indian Journal of Agricultural Sciences*, 88, 1198–1201.
- Zou Y.N., Wu H.H., Giri B., Wu Q.S., Kuča K. (2019) Mycorrhizal symbiosis down-regulates or does not change root aquaporin expression in trifoliate orange under drought stress. *Plant Physiology and Biochemistry*, 144, 292–299.