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Review Article

PHYTOFLAVONOIDS: ANTIEPILEPTICS FOR THE FUTURE

PARAMDEEP SINGH1*, DAMANPREET SINGH2, RAJESH KUMAR GOEL2

¹Department of Pharmacology, Chitkara College of Pharmacy, Chitkara University, Chandigarh Patiala National Highway, Rajpura, Patiala 147002, Punjab, India, ²Department of Pharmaceutical Sciences and Drug Research, Punjabi University, Patiala 147002, Punjab, India. Email: paramdeepsond87@gmail.com.

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ABSTRACT

Epilepsy has become second most common neurological disorder affecting 70 million people worldwide. Several shortcomings appeared with the use of conventional antiepileptic drugs (AEDs) like, inadequate seizure control, side effects, cost, potentiation of epilepsy-induced co-morbidities, etc., which limit their use as comprehensive therapy. Phytoflavonoids in this regard have been observed to be promising AEDs which can interact with most of the possible targets involved in the pathogenesis of epilepsy. They show modulatory effect on voltage gated sodium channels, calcium channels, potassium channels, GABAergic system, opioid receptors, NMDA receptors and posses a strong antioxidant effects through modulation of nitric oxide pathway, xanthine oxidase system and through leukocytic immobilization. Along with antiepileptic effect, flavanoids have also been observed to ameliorate memory deficit and mood disorders commonly associated with AEDs treatment. Based on the vast literature reports, an attempt has been made to collate the fragmented information available on flavonoids, to access there potential as antiepileptics for future. From this review it is cleared that, flavonoids open new encouraging perspectives for the antiepileptic treatment. A prospective randomized trial of flavonoids may be justified in humans.

Keywords: Flavonoids, Epilepsy, Epilepsy-induced comorbidities, Flavonoidal plants.

INTRODUCTION

The name epilepsy is derived from a Greek word "epilambanein", which means "to seize upon", "to attack". Epilepsy is characterized as a chronic brain disorder in which there is tendency of the patient to experience periodic, unpredictable recurring seizures with or without characteristic body movements, called convulsions [1]. Epilepsy is the second most common neurological disorder affecting around 70 million people worldwide, out of which approximately 80 % are in developing countries [2].

Around 60-70% of the total patients with epilepsy depend on the medical treatment with antiepileptic drugs (AEDs) to achieve control of their epileptic seizures. These AEDs are targeted on supressing an abnormal epileptic discharge, acting generally via one or more mechanism(s) through inactivation of Na⁺(hydantoins, etc) and Ca²⁺ channels (ethosuximide, etc), increasing GABA-mediated inhibitory functions (barbiturates, benzodiazepines, etc), inhibition of glutamate-mediated excitation (felbamate, etc), activation of K⁺ channels (retigabine) [3-5].

Despite, decades of advancements in AEDs, there are a lot of shortcomings associated with their use. One-third of the epileptic patients treated with the available AEDs do not obtain satisfactory seizure control. Moreover, chronic use of these AEDs is associated with several systemic and central nervous system adverse effects like, fatigue, agressivity, anxiety, headache, hair loss, skin reactions, diplopia or blurred vision, dyspepsia, gingival hypertrophy or bleeding, tremors, weight gain, dizziness, somnolence, memory impairment, sleep disturbance, lack of concentration, lack of libido, difficulty of erection in men [6, 7].

A majority of epileptic patients suffer from one or more psychiatric or somatic comorbid conditions, whose nature and prevalence vary with age and sociodemographic factors, these comorbid conditions include cognitive deficit, mood disorders, anxiety, etc [8]. In the developed countries where drugs are easily available only 70 % patients with epilepsy responds to antiepileptic drug therapy. However, in developing countries 75% of people with epilepsy do not receive the treatment they need and their epilepsy remains uncontrolled, rendering the patient unproductive in all spheres of life. All these issues direct a large proportion of the patients to move on to traditional healers and medicinal plants [9]. Traditional herbs due to presence of large amount of bioactive active ingredients have a great potential to treat epilepsy. Among all, phytoflavonoids are the most important bioactive product isolated from traditional herbs used in the treatment of epilepsy. "Phyto" is a Greek word that means plant, so phtoflavonoids means the flavonoids obtained from natural plant resources. Many of the traditional plants containing flavonoids have been experimentally explored for their anticonvulsant effect. Studies have shown that flavonoids possess remarkable anticonvulsant activity which can be exploited for the development of comprehensive AEDs of future [10-12]. Isolated phytoflavonoids like, hispidulin, rutin, hesperetin, naringenin, eriodictyol, chrysin, gossypin, apigenin, kaempferol, myricetin, nuercetin, hyperoside etc. posses GABAergic agonism, sodiumchannels blockade, calciumchannels blockade, glutamate receptors antagonism and antioxidant etc., thereby act as anticonvulsants [13].

Flavonoids also had been found to ameliorate cognition deficit and depression, both of which occur in epileptic patient as comorbidities. It has also been found that herbal antioxidant like, curcumin have synergistic antiepileptic effect with standard AEDS like phenytoin [14]. Curcumin when co-administered with the subeffective doses of phenytoin, increased the latency to myoclonic jerks and protected against seizures, which was absent in animals treated *per se* with the same dose of phenytoin. Thus, flavonoids are turning out to be very useful and indispensable in the struggle for seizure management and future AEDs development. The present review focuses on antiepileptic flavonoids, their mechanisms, toxicology and their possible efficacy in epilepsy-induced comorbidities.

FLAVONOIDS

Flavonoids, also known as Vitamin P, are a class of secondary metabolites that provide UV protection and color to the plant. Nijveldt *et al.* [15] revealed their protective effect on brain stroke and cardiovascular system diseases.

Chemically, flavonoids are polyphenolic components containing a core of 15 carbon atoms; two benzene rings joined by a linear three carbon chain. Based upon the position present on basic parent moiety flavonoids are generally classified into 6 subclasses flavones, flavonols, isoflavones, flavonones, flavon-3-ols and anthocyanidin (Fig. 1). Biologically they are synthesised via acetate and shikimic acid pathway into a $C_6 - C_3 - C_6$ system [17, 18].



A- Flavones, B- Flavonols, C- Isoflavones, D- Flavonones, E- Flavan-3-ols, F-Anthocyanidins

-O-Me= Methoxy, -O-Glu= Glucosyl, -Alkoxy= Alkoxy

Fig. 1: It shows the comprehensive list of all the chemical structures for the main classes of flavonoids.

Antiepileptic mechanisms of flavonoids

Flavonoids have been found to inhibit almost all the mechanisms involved in seizures generation in epilepsy. They have been found to modulate neuronal Na⁺ channels, Ca²⁺ channels, GABA ergic pathway, glutamatergic pathway and opioid pathway (Fig. 2).

- 1. Inhibition of voltage gated Na*channels resulting in decrease sodium ion influx into the neuronal cell
- 2. Activation of Ca⁺activated K⁺channels resulting in increased potassium ion outflow from thr neuronal cell

- 3. Activation of inhibitory GABAergic receptors through direct action on GABA or through benzodiazepine receptor resulting in increased chloride ion influx causing neuronal hyperpolarisation
- 4. Inhibition of opioid receptors leading to proconvulsant or anticonvulsant effects
- 5. Antioxidant effects of flavonoids leading to increased seizure threshold.
- 6. Inhibition of NMDA receptor, resulting in decrease entry of calcium into the neuronal cell



Fig. 2: Schematic antiepileptic mechanism of flavonoids.

Inhibition of voltage gated sodium channels

Abnormal increase in intracellular Na⁺ causes high neuronal excitability leading to excessive depolarization leading to seizures. Several clinically used drugs like, Phenytoin, Carbamazepine, Valproic acid, etc. act via blockade of sodium channels, inhibiting abnormal depolarization. Flavonoids such as quercetin and pinostrobin have been found to act through similar mechanism [19]. Quercetin also decreases the amplitudes of sodium currents at different membrane potentials in rat hippocampal CA1 pyramidal neurons [20].

Activation of Ca+activated K+ channels

Repetitive ictal and interictal activity causes increase in extracellular K⁺, leading to increased neuronal excitability. Neurons are very sensitive to changes in membrane K⁺ currents, e.g. pyramidal cells in CA1 region of the hippocampus. Mutation in these voltage-gated K⁺ channel causes benign familial neonatal convulsions [21]. Retigabin is a clinically used antiepileptic drug and act by opening K⁺ channels. Flavonoids like quercetin have been reported to activate Ca²⁺ activated K⁺ channels and decreases extracellular K⁺ leading to hyperpolarization amelioration of seizures [22].

GABAergic inhibition

GABA (γ -aminobutyric acid) is the principal inhibitory neurotransmitter in the mammalian CNS. GABA elicits inhibitory response in the brain via three classes of receptors, GABA_A, GABA_B and GABA_C. GABA_A is a GABA gated chloride channel which allows influx of chloride ions resulting in decrease excitatory stimulus [23].

The GABA_B receptor is a G-protein coupled metabotropic receptor which interacts with G_i to inhibit adenylyl cyclase, activate K^{*} channels and reduce Ca²⁺ conductance. The GABA_C receptor is a transmitter-gated Cl⁻ channel and is less widely distributed than the A and B subtypes [24].

The GABA hypothesis of epilepsy implies that a reduction of GABAergic inhibition results in epilepsy, whereas an enhancement of GABA-ergic functions results in an antiepileptic effect. Modulation of ionotropic GABA receptors by a group of natural and synthetic flavonoids has been extensively studied in several experimental studies. Literature survey showed that, quercetin, apigenin, morine, chrysin and flavone inhibit ionic currents mediated by GABA_A and GABA_C receptors expressed in *Xenopus laevis* oocytes leading to suppression of abnormal depolarization [21, 25]. Flavonoids like apigenin and (-) epigallocatechin gallate potentiates the therapeutic effect of diazepam [26], indicating their efficacious role in combined therapy. This synergistic effect of flavonoids is due to their binding affinity with GABA_A receptor through high-affinity flumazenilsensitive site or low-affinity, flumazenil-insensitive site [25].

Benzodiazepines (BDZ) also play a vital role in ameliorating epilepsy and act by enhancing GABAergic functions leading to reduction in sustained repetitive firing. But because of their depressant nature they show several side effects. Whereas, flavones like, 6,3dinitroflavone act by similar GABAergic action as that of BDZ sites but devoid of side effects like, ataxia, amnesia, etc [21, 27].

Opioid receptor interactions

Opioid receptors are found in various brain regions and spinal medulla, as well as in intramural nerve plexuses. These receptors are classified as μ , δ and κ , they mediate their physiological functions through G-protein coupled mechanism. Activation of these receptors causes increase in K⁺ conductance, decrease in Ca²⁺influx into nerve terminal leading hyperpolarization, decreased release of excitatory neurotransmitters and decreased synaptic activity. Depending on the cell population affected, the synaptic inhibition translates into depressant or excitatory effects. Studies have shown that opioids and opioid peptides had both convulsant and anticonvulsant properties. Several flavonoids have been evaluated for their opioid receptor modulatory effects. Amentoflavone and hyperoside present in *H. perforatum* have κ antagonist activity *in vitro* [21]. Katavic [28] also suggested flavonoid core as a future new scaffold for the development of opioid receptor ligands.

Inhibition of NMDA receptors

Excessive activation of glutamatergic N-Methyl-D-aspartate (NMDA) receptor leads to neuronal degeneration and death, due to increased Ca²⁺ concentration in the postsynaptic neurons leading to abnormal neuronal depolarisation. Binding of Ca2+to calmodulin also activates nitric oxide synthase, thereby increasing the formation of nitric oxide (NO) that contributes to the neurotoxic effects. Moreover, activation of NMDA receptors also leads to increased production of superoxide radicals, leading to the generation of hydroxyl radicals causing oxidative damage which is responsible for worsening of epileptic condition and initiates its associated comorbidites. NMDA receptor antagonists are powerful anticonvulsant and are clinically used. They are responsible for the inhibition of calcium influx into the neurons thereby preventing abnormal excitation. Several flavonoids like, morin, kaemferol, myricetin, quercetin, anghyperoside have been reported to block glutamatergic excitation mediated through NMDA receptors. Morin provides neuroprotection by inhibiting excessive activation of NMDA receptors, NMDA receptor mediated neurotoxicity [29]. Thus, flavonoids like morin can be helpful in developing NMDA antagonists which can be effectively used for the treatment of epilepsy and its associated comorbidities in future.

Antioxidant potential

Free radicals play a crucial role in the pathogenesis of epilepsy. Increase in number of free radicals is coupled with low level of antioxidants, which results in the neuronal damage. It has been reported that free radical generation induces seizure activity by inactivation of glutamine synthase, leading to an abnormal build-up of excitatory neurotransmitter, glutamate or by decreasing glutamate decarboxylase, thus causing decrease in GABA turnover [13]. Flavonoids in this regard prove out to be a bioactive metabolite of choice for the treatment of oxidative stress induced diseases like, epilepsy. Traditionally also plants containing flavonoids have been experimentally explored for their anticonvulsant effect [10-12]. Flavonoids possess high antioxidant activity and low IC₅₀ in *in vitro* antioxidant tests like 2,2-Diphenyl-1-picrylhydrazyl (DPPH) radical scavenging assay [30]. IC₅₀ is the concentration at which the substance shows 50% inhibitory activity. Ishige *et al.* [31] found three distinct mechanisms of antioxidant protection like, increasing intracellular GSH, directly lowering levels of ROS, and preventing the influx of Ca²⁺ despite high levels of ROS.

Not all flavonoids have equal antioxidant potential. Flavonols were found to be best antioxidants when compared with anthocyanidins, flavone and flavanone and other type of flavonoids [32, 33]. Flavonols like goodyerin, quercetin, rutin can be used to effectively control epilepsy [11, 34-36]. Flavonols like quercetin and kaempferol are also reported for reversing oxidative stress induced-decrease of hippocampal BDNF and pCREB expression and leading to hippocampal neurogenesis and thus, ameliorating depression and cognitive deficit [37, 38].

Direct radical scavenging

Flavonoids have a highly reactive hydroxyl group in their structure. It reacts with free radical resulting into more stable less-reactive radical, thus providing direct radical scavenging activity [30]. Hanasaki *et al.* [40] studied the abilities of 15 flavonoids as a scavenger of active oxygens (hydroxyl radical and superoxide anion). Hydroxyl radical (OH) which was generated by the Fenton system, and assayed by the amount of methanesulfinic acid (MSA) formed from the reaction of dimethyl sulfoxide (DMSO) with. OH (+)-Catechin, epicatechin, 7,8-dihydroxy flavone, and rutin showed the. OH scavenging effect almost 100-300 times superior to that of mannitol, a typical. OH scavenger.

Modulation of nitric oxide pathway

Increased levels of nitric-oxide synthase in macrophages enhance the production of both nitric oxide and superoxide anions. Nitric oxide when reacts with free radicals causes production of peroxinitrite, which are highly damaging to the cell. Flavonoids like silibin have been found to scavenge nitric oxide and free radicals directly (Fig. 3) [41, 42].





Xanthine oxidase

During oxidative injury to tissues xanthine dehydrogenase gets configurationally changed to xanthine oxidase which is an important source of oxygen free radicals. Flavonoids such as quercetin, silibin, luteolin etc. inhibit xanthine oxidase activity, thereby resulting in decreased oxidative damage [44, 44]. Luteolin (3,4_5,7- tetrahydroxyflavone) have been found to be the most potent inhibitor of xanthine oxidase [45].

Leukocyte immobilization

Oxidative endothelium damage, as a consequence of ischaemia results in generate of endothelium derived mediators and complement factors, which cause adhesion of leukocytes to the endothelial wall and stimulating degranulation of neutrophils. As a result, oxidants and inflammatory mediators are released, resulting in tissue injury. Flavonoids due to modulation of receptor-directed Ca²⁺ channels in the plasma membrane [46] have been found to decrease the generation of various mediators and ameliorate oxidative stress [47, 48].

A majority of the above mentioned studies have been carried out in in vitro assay. In future the antioxidant effect of flavonoids can be further studied in the *in vivo* models of epilepsy to further understand and explore the mechanism involved in their anticonvulsant action.

Effect of flavonoids on epilepsy-induced comorbidities

Majority of epileptic patients suffer from one or more psychiatric or somatic comorbid conditions, whose nature and prevalence vary with age and sociodemographic factors, the most common comorbid conditions include, cognitive deficit and mood disorder. In these patients, comorbid conditions have a major adverse effect on overall health and quality of life and substantially increase health care costs [8]. Moreover, majority of antiepileptic drugs are consumed life long, concomitant administration predisposes to high risk of adverseeffects and drug interaction. Treatment with these conventional AEDs is associated with the brain dysfunction such as hippocampal atrophy causing memory disorders and mood disorders with a cascade of pervasive psychosocial problems. Phenobarbital and topiramate has the greatest potential to affect cognitive defect while levetiracetam, topiramate, zonisamide and phenobarbital has potential to cause behavioral adverse-effects [7]. Therefore, satisfactory treatment of epilepsy not only demands the suppression of abnormal neuronal discharge but also control of epileptic comorbidities associated with it.

During seizures the blood flow to the brain increases due to immediate increase in the diameter of pial arterioles and major resistance vessels of the brain that determine the cerebral blood flow. This increased cerebral blood flow is required to match increased oxygen and other metabolic demands of the activated neurons during the epileptic seizures [49]. This increased blood flow increases the metabolic load on the brain causing abnormal generation of free radicals. Excessive generation of free radicals like reactive oxygen species (ROS) and reactive nitrogen species (RNS) are the key factors that react with endogenous substrates like, fatty acids, proteins and DNA, causing oxidative damage [50-52]. Oxidative and nitrosative stress also activates neutrophils and monocytes that cause nitration and nitrosylation of proteins to generate inactive autoepitopes to neoantigens which acquire immunogenicity and causes autoimmune responses [53]. It also induces decrease in hippocampal BDNF and pCREB expression leading to hippocampal cells degeneration. These pathways in turn generate neurodegenerative process mainly responsible for the induction of comorbid conditions. Oxidative stress is also established in the genesis of cognitive deficit associated with epilepsy via interaction with several pathways like, CREB, PI3K, PKG, ERK1/2 etc [54-58]. Flavonoids, due to their antiepileptic and neuroprotective effects have been found to ameliorate the comorbities. They interact with critical protein and lipid kinase signaling cascades, such as phosphatidylinositol-3 kinase (PI3K)/Akt, protein kinase C and mitogen-activated protein kinase in the brain, leading to inhibition of apoptosis triggered by neurotoxic species and promote neuronal survival and synaptic plasticity. Secondly, flavonoids induce beneficial effects on the vascular system leading to changes in cerebrovascular blood flow capable of causing angiogenesis, neurogenesis and changes in neuronal morphology [59].

Flavonoids and memory deficit

Flavonoids due to their remarkable antioxidant activity act to ameliorate cognitive deficit mainly by three common processes. Firstly, they interact with critical protein and lipid kinase signaling cascades such as phosphatidylinositol-3 kinase (PI3K)/Akt, protein kinase C and mitogen-activated protein kinase in the brain, leading to an inhibition of apoptosis triggered by neurotoxic species and to promote neuronal survival and synaptic plasticity. Secondly, flavonoids induce beneficial effects on the vascular system by decreasing reactive oxygen species and increasing nitric oxide bioavailability. It leads to changes in cerebrovascular blood flow capable of causing angiogenesis, neurogenesis and changes in neuronal morphology by activating PI3 kinase-Akt-eNOS system. Angiogenesis is followed by neurogenesis. Thirdly, flavonoid activate ERK and Akt pathway leading to activation of CREB pathway causing the release of neurotrophins such as BDNF and activate PI3 kinase-mTOR cascade and Arc/ Arg3.1 which results in changes in synaptic plasticity and Long Term Potentiation(LTP) [59, 60]. Flavonoids such as flavonols, anthocyanins and flavonones have much better potential to ameliorate cognitive process. Isoflavone supplementation has been observed to have a favorable effect on verbal memory and on postmenopausal women. Isoflavones have been to mimic the actions and functions of oestrogens in the brain. Pure flavonols such as quercetin, rutin or fisetin have provided further evidence that dietary flavonoids are beneficial in reversing the course of neuronal and behavioural aging. The flavonoid-rich plant extract, Ginkgo Biloba has also been shown to induce positive effects on memory, learning and concentration. Fisetin, a flavonoid isolated from strawberries, has been shown to improve long-term potentiation and to enhance object recognition in mice.

The central cholinergic system plays an important role in learning and memory [61-62]. AChE (acetylcholinesterase), the main enzyme that hydrolyses acetylcholine, increases in the experimental and clinical epilepsy [63-65] and elevated AChE level has been suggested as pathogenic factor for memory deficit [66]. Acetylcholinesterase inhibitors have been shown to function by increasing the level of acetylcholine within the synaptic region, thereby restoring deficient cholinergic neurotransmission [69]. It has been reported that flavonoids like flavonols galangin, kaempferol, quercetin, myricetin, fisetin; flavons - apigenin, luteolin, and flavonol-glycoside quercetin-3rutinoside (rutin) inhibits human AChE thereby effects memory functions. It has been found that AChE inhibition potency increase from rutin < apigenin < fisetin < kaempferol < galangin < luteolin < quercetin ≤ myricetin. It seems that larger number of OH groups on the side phenyl ring would result in the more pronounced AChE inhibition like in case of flavonols; quercetin and myricetin [67].

Since majority of aforesaid flavonoids have shown antiepileptic effects, their memory enhancing effect indicate that these flavonoids can be used for the treatment of epilepsy without the risk of memory impairment, provided their memory enhancing effect during epileptic condition is studied in future.

Flavonoids and mood disorders

Flavonoids extracted from plants are known to exert antidepressant like effects in animal models of depression [68, 69]. Naringenin a dietary flavonoid possessing neuroprotective actions in various central pathophysiological conditions including depression [70]. Xiaobuxin-Tang (XBXT), a traditional Chinese herbal decoction, has been used for the treatment of depressive disorder for centuries in China. It is reported to contain a large amount of flavonoids which on treatment reversed behavioral alterations including depression [37]. Similarly flavonoid-rich extract of Hypericum perforatum mainly containing quercetin and hyperoside flavonols [71] and flavonoids of Scutellariae radix were found to be responsible for antidepressant action [72]. Flavonoids, mainly flavonols like, quercetin and kaempferol are responsible to revert the oxidative stress induced-decrease of hippocampal BDNF and pCREB expression, leading to hippocampal neurogenesis and thus, ameliorating depression [37, 38]. These antidepressant effects of flavonoids can be further studied in epilepsy-induced depression to further expand their therapeutic potential in epilepsy.

Flavonoidal plants as antiepileptic

In several previous studies flavonoidal plants have been studied for their anticonvulsant activity in different animal models of convulsions. In this section, we have tried to summarize and discussed about the plants containing flavonoids that have been screened for their anticonvulsant activity (also summarized in Table 1). These plants can be further explored in future as in majority of cases the information regarding the particular type of flavonoid involved for the anticonvulsant activity, exact mechanism of action, safety, etc. is lacking.

Anisomeles malabarica (L) R. Br

Anisomeles malabarica is an herbaceous Australian plant belonging to family Lamiaceae, it is a plant of immense use and utility in folk medicine. It has been used for the treatment of epilepsy. Choudhary *et al.* [73] studied the anticonvulsant effect of flavonoidal-rich fraction from the leaves of *A. malabarica*. In the study the ethyl acetate fraction containing flavonoid showed antiepileptic activity against pentylenetetrazole (PTZ) and maximal electroshock (MES)-induced convulsions in wistar rats. Chronic treatment with the fraction at doses of 6.25 and 12.5 mg/kg *i.p.* for 1 week has showed significant anticonvulsant effect without any signs of neurotoxicity. Further research is required to determine the active responsible flavonoid(s) and their exact mechanism of action.

Glychrrhiza glabra L.

G. glabra (Febaceae) also known as liquorice is native to southern Europe and parts of Asia. It is commonly known as "Mulaithi" in Northen India. It has been found to contain antioxidant substances like, ascorbic acid (0.58 g %) and flavonoids (0.926 g %) such as glabridin (an isoflavan) and isoliquiritigenin (a flavonoid) [74]. Ambawade *et al.* [75] studied its anticonvulsant activity using MES, PTZ and lithium-pilocarpine-induced convulsion mice models. Its ethanolic extract of roots and rhizomes containing flavonoids at 10, 30, 100 and 500 mg/kg *i.p.* doses inhibited PTZ and lithium-pilocarpine-induced convulsions, but was found to be ineffective against MES-induced convulsions. The extract was found to be safe up to 1g/kg *i.p.* dose in mice. Since it is rich in flavonoids, hence it is expected that the observed anticonvulsant effect can be due to their presence, however more studies are required to fulfill these expectations.

Passiflora incarnate L.

P. incarnate commonly known as passion flower/maypop/apricot vine belongs to family Passifloraceae. It was discovered in 1569 by Spanish explorers in Peru. The local Indians used it to soothe their nerves and topically as a poultice, the crushed leaves being used on cuts and bruises [76]. Phytochemical research carried out on *Passiflora incarnata* had lead to the isolation of several flavonoids like, chrysin, apigenin, homoorientin, vitexin, luteolin, quercetin, kaempferol, isovitexin, orientin, isoorientin, etc. The anticonvulsant effect of the hydroethanolic extract of its aerial parts has been extensively explored and has been suggested to be due to presence of flavonoids like chrysin that act by agonizing benzodiazepine receptor [27, 77, 78].

Scutellaria lateriflora L.

It belongs to family Lamiaceae and is commonly known as American Skullcap. S. lateriflora has been used for the treatment of epilepsy and several other neurological disorders in traditional medicine. Zhang et al. [79] isolated and identified 10 flavonoids, viscidulin III-2-O-β-D-glucopyranoside, chryin-6-C-α-L-arabionopyranosyl-8-C-β-D-glucopyranoside, trans-verbascoside, viscidulin III, baicalin, transmartynoside, oroxylin A-7-O-β-D-glucopyranoside, wogonoside, baicalein, wogonin from whole extract of skull cap. In the same study, the anticonvulsant effect of the hydroalcoholic extract of whole skullcap at 90 mg/kg i.p. showed significant protection against PTZ-induced convulsions. The flavonoids present in the extract like wogonin which is known to have anticonvulsant activity [80] may be the active ingredients responsible for the anticonvulsant activity of American skullcap as they show high affinity for the benzodiazepine binding site at GABAA receptor [81] and have antioxidant effects [82, 83].

Drosera burmanni Vahl

The genus *Drosera* is popularly known as Sundew and is the largest genus of carnivorous plants with over 105 species belonging to the family Droseraceae. Flavonoids like kaempferol, myricetin, quercetin and hyperoside have been isolated from *D. burmanni* [84]. Hema *et al.* [85] studied the anticonvulsant activity of the alcoholic extract of

whole plant at a dose of 500 *mg/kg p.o.* The extract significantly delayed the onset of PTZ-induced convulsions in swiss albino mice. This anticonvulsant activity may be attributed because of flavonoids as the flavonoids like quercetin and kaempferol are known to be responsible for anticonvulsant activity [86] and kaempferol, myricetin, quercetin are also reported to have neuroprotective effects [87]. Flavonoids due to reduction of T-type calcium currents, enhancement of GABA_A-BZD receptor activity and blocking of glutametargic excitation mediated by NMDA receptors can effectively inhibit PTZinduced convulsions.

Cleome viscose L.

C. viscose also known as Dog mustard belongs to family Capparidaceae. Its fresh juice of crushed seeds has been used for treatment of infantile convulsions and other mental disorders. In a recent study, the anticonvulsant activity of its ethanol and aqueous seed extract containing flavonoids has been studied [88]. The extracts at 200, 400 and 600 mg/kg p.o. doses prevented MES and PTZ-induced convulsions in mice. The anticonvulsive effect might be because of flavonoids attributed due to inhibitory effect on voltage dependent sodium channels or due to blockage of glutaminergic excitation. However further studies are required to find out the exact role and types of flavonoids responsible for the activity.

Caesalpinia bonducella (L.) Roxb.

It is a prickly shrub belonging to family Caesalpiniaceae and is commonly known as Nata Karanja. It is found throughout the hot regions of India, Myanmar and Sri Lanka. Ali *et al.* [89] investigated the anticonvulsant activity of the petroleum ether extract of its seed kernel. Phytochemical screening showed the presence of flavonoids like homoisoflavone (bonducillin) in the petroleum ether extract. The petroleum ether extract at chronic doses of 600 and 800 *mg/kg p.o.* for 8 days prevented PTZ, MES, strychnine and picrotoxin-induced convulsions in swiss albino mice. Anticonvulsant activity might be because of flavonoids, but still it is not clear that the effect of extract is due to flavonoids or some other bioactive metabolite, more studies are required in this area.

Vitex negundo L.

V. negundu commonly known as five-leaved chaste tree or Monk's Pepper belongs to family Verbenaceae. It was used in Iranian ancient medical schools and Alkandi for the treatment of epilepsy and psychosis, in India it is also used as antiepileptic. Its methanolic leaf extract showed protection against strychnine and PTZ induced convulsions [90]. Tandon [91] reported the anticonvulsant activity of its petroleum and butanolic leaf extracts against MES-induced convulsion model. The extracts showed the presence of flavonoids like, flavones. In another study treatment with its ethanolic leaf extract not only showed anticonvulsant activity, but also potentiated the anticonvulsant effect of standard antiepileptic drugs when used in combination [92]. The study suggested that flavanoids can be used in combination with standard antiepileptic drugs to increase efficacy and mask side effects of latter, provided the safety of combined treatment is warrant in future research.

Ocimum gratissimum L.

O. gratissimum also known as African Basil belongs to family Lamiaceae. Vieira *et al.* [93] reported the presence of Eugenol, thymol, and geraniol as the major volatile oil constituents and Xantomicrol and cirsimaritin as the major external flavones. Okoli *et al.* [94] studied the anticonvulsant activity of its leaves containing eugenol (volatile oil) and cirsimaritin (flavones) using PTZ model in mice. Methanolic and petroleum ether extract of leaves were administered at the doses of 200 and 400 mg/kg p.o. and was found to increase the latency of seizures and death and elicited 50% protection against mortality. As the studies aimed at isolating the anticonvulsant constituents of this plant are ongoing but still the role of flavonoids as anticonvulsant cannot be ruled out.

Centella asiatica (L.) Urban

It is commonly known as Gotu Kola and belongs to family Umbelliferare. It has been used in Indian traditional medicine as a memory enhancer, treatment of neurological disorders (insomnia, hysteria, insanity, epilepsy, etc) and skin diseases. Its leaves contain flavonoids like, 3-glucosylquercetin, 3-glucosylkaempferol and 7glucosylkaempferol [95]. Visweswari *et al.* [96] studied the anticonvulsive activity of its *n*-hexane, chloroform, ethyl acetate, *n*-butanol leaf extracts. Treatment with all the extracts except aqueous extract at 200 mg/kg b.w. 1 week prior to dose of PTZ showed significant protection against PTZ-induced convulsions in male wistar rats. It was found that the activities of Na⁺, K⁺-ATPase, Mg²⁺-ATPase, and Ca²⁺-ATPase in brain (cerebral cortex, cerebellum, pons medulla and hippocampus) were significantly increased after treatment with different *C. asiatica* extracts which in general is decreased after PTZ induced epilepsy in all the above brain regions.

Cissus sicyoides L.

C. sicyoides (Vitaceae) has been evaluated for its anticonvulsant activity. Phytochemical exploration had led to the isolation of two flavonoids, kaempferol 3-rhamnoside and guercetin 3-rhamnoside from its aerial parts [97]. De Almeida et al. [98] studied the anticonvulsant activity of the hydroalcoholic extract of the aerial parts of C. sicyoides. Treatment with the extract at 600 and 1000 mg/kg i.p. doses inhibited PTZ-induced convulsions in mice. Since, kaempferol and quercetin have been reported to have anticonvulsant activity, and kaempferol 3-rhamnoside and quercetin 3-rhamnoside are their flavano-glycosidic forms, hence the activity of the extract can be due to these metabolites. The glycosidic part conjugated with the flavonoid will increase solubility thus showing better pharmacokinetic property and increasing the anticonvulsant effects of these flavonoids. More studies are required to confirm the exact role of these flavano-glycosides for the observed anticonvulsant effect of C. sicyoides extract.

Ficus hispida L.

F. hispida is a medicinally important member of family Moraceae. Dhanasekaran *et al.* [99] studied the anticonvulsant effect of its methanolic leaf extract containing flavonoids, using strychnine, picrotoxin, and PTZ mice models of convulsions. Treatment with the extract at 400 mg/kg *p.o.* dose completely abolished the seizures induced by picrotoxin and strychnine, but was found to be ineffective against PTZ-induced convulsions. Since, both PTZ and picrotoxin induces convulsions through similar mechanism, but the reason for the effectiveness of the extract in one model and not in other require more studies.

Ficus sycomorus L. Range

F. sycomorus (Moraceae) is generally found in the Arabian peninsuala, tropical Africa to the east and Nigeria. Naturally, the tree is usually found in rich soils along rivers and in mixed woodlands. Ibrahim *et al.* [16] studied the anticonvulsant activity of the flavonoids-rich fraction of its ethanolic stem bark extract. Treatment with the flavonoids-rich fraction at 20 mg/kg *i.p.* dose significantly protected MES-induced convulsion in 30 day old white ranger cockerels and showed a weak anticonvulsant activity against PTZ-induced convulsions in mice. The flavonoids-rich fraction of the ethanolic stem bark extract of *F. sycomorus* can be further fractionated in future to get the bioactive flavonoid(s).

Argyreia speciosa (Burm.f.) Bojer

Argyreia speciosa (Convolvulaceae) commonly known as *Vridha daraka* in Sanskrit and is one of the most important medicinal plant used in indigenous system of medicine. Its roots are regarded as an alternative tonic to cure nervine disorders. The major flavonoids isolated from the roots include, quercetin and kaempferol. Vyawahare *et al.* [100] studied the anticonvulsant activity of the hydroal-coholic extract of its roots in PTZ and MES mice models.

Treatment with the extract at 200 and 400 mg/kg doses significantly delayed the latency to the onset and reduced the duration of convulsion in PTZ and MES tests, respectively. Since the plant has also shown nootropic effects [101], as described memory deficit is a major complication in epilepsy, hence activity of the plant in both disorders indicate its possibility in suppression of epilepsy and associated memory impairment, which can be explored in future.

Pongamia pinnata (L.) Pierre.

It belongs to family Febaceae and is commonly known as Karanj. Phytochemical studies revealed the presence of flavonoids in its leaves, pongol (flavone derivative), glabrachalone and isopongachromene in its seeds [102]. Manigauha *et al.* [103] reported the anticonvulsant activity of its ethanolic leaf extract. The study showed that, treatment with the extract at 250 mg/kg, *i.p.* dose prevented PTZ induced convulsions in wistar albino rats. It also reported that flavonoids like quercetin, kaempferol present in *P. pinnata* are responsible for protection against PTZ induced convulsions by modulating GABAergic or glutamatergic receptor functions.

Abutilon indicum (L.) Sweet

Abutilon indicum (Malvaceae) commonly known as Thuthi and Atibala in Ayurveda is widely used as medicine in Ayurveda for antifungal, antibacterial, immunomodulator and analgesic activities [104]. It rich in luteolin, chrysoseriol, luteolin-7-O- β glucopyranoside, chrysoseriol 7-O- β glucopyranoside, apigenin 7-O- β -gluco pyranoside, quercetin 3-O- β -gluco pyranoside, quercetin 3-O- β -gluco pyranoside, quercetin3-O- α rhamnopyranosyl, β -gluco pyranoside [105]. Golwala *et al.* [106] studied the ethanolic (AIE) and aqueous (AIA) leaf extracts for anticonvulsive properties at 100 mg/kg and 400 mg/kg p.o. against PTZ and MES induced convulsions. Both the extracts provided protection but AIE showed better significant protection than AIA against the PTZ and MES induced convulsions. This study also indicated flavonoids for its anticonvulsant effect which might be true as flavonoids are extracted in the alcoholic extract and this study have shown that AIE as better anticonvulsant than AIE.

Hypericum perforatum L

H. perforatum commonly known as St. John's Wort, belongs to family Hypericaceae. It has been used for the treatment of epilepsy since antiquity. It contains high amount of flavonols (catechins) and flavonoids (hyperoside, quercetin, quercetrin, rutin, biapigenin, kaempferol) [107, 108]. Hosseinzadeh et al. [109] reported the anticonvulsant activity of its aqueous and ethanolic extracts of aerial part in PTZ and MES convulsion mice models. Both the extracts at 0.1-1g/kg i.p. dose were found to be effective in PTZ, but not in MES model. Further pretreatment with a nitric oxide synthase inhibitor reduced anticonvulsant effect of the extracts, indication the involvement of nitric oxide pathways for the activity of extracts. As the class of flavonoids present in H. perforatum have shown anticonvulsant activity in other previous studies, and the role of flavonoids in the modulation of nitric oxide pathway has been well established, therefore it can be correlated that the anticonvulsant activity of *H. perforatum* is due to flavonoids. However further studies are required to generate more scientific support for the safe clinical use of *H. perforatum* in epilepsy.

Mimosa pudica L

M. pudica (Missociaceae) commonly known as Lajvanti has been used for the treatment of numerous diseases related to nervous system. Bum *et al.* [110] investigated the anticonvulsant activity of the decoction of *M. pudica* leaves. Treatment with the leaf decoction at 1000 and 4000 mg/kg *i.p.* protected mice against PTZ and strychnine-induced convulsions. Since the leaves of *M. pudica* contains an appreciable amount of flavonoids [111], hence it is expected that the anticonvulsant effect of the leaves can be due to flavonoids. Therefore further studies are required to validate the role of flavonoids for the anticonvulsant effect of the leaves of *M. pudica*.

Butea monosperma (Lam.) Taub.

In the Ayurveda it is known as Bastard Teak, it belongs to family Fabaceae. Silambujanaki *et al.* [112] evaluated the anticonvulsant activity of the methanolic leaf extract of *B. monosperma* in mice. Preliminary phytochemical screening showed the presence of flavonoids. The extract at 100, 200, 400 mg/kg p.o. doses was used against MES, PTZ and strychnine induced seizure models. It was found that the extract at 400mg/kg dose was effective against MES and PTZ but not against strychnine induced seizures. The anticonvulsant activity exhibited by the extract might be due inhibitory effect on Na⁺, t-type Ca²⁺ channels, NMDA receptors and excitatory

effect on GABA aminergic mechanism. The study also suggested that the extract might be effective due to the presence of flavonoids in the extract.

Nauclea latifolia Sm.

It is commonly known as pin cushion tree belonging to family Rubiaceae. It has been experimentally studied for its anticonvulsant activity [113]. Aqueous extract of the bark of dried roots at 16, 40, 80, and 160 mg/kg *i.p.* was studied against MES, PTZ and Strychnine induced seizures in mice. Study concluded that the dose of 160 mg/kg provided significant protection against MES, PTZ and Strychnine induced seizures. Flavonoids present in the extract might be showing this activity through benzodiazepine, GABA receptors in the GABA receptor complex and probably by prolonging the inactivation of sodium channels.

Sphaeranthus indicus L.

It is commonly known as Gorakhmundi, which is an herb widely distributed in wet places of India belonging to family Compositae. Galani *et al.* [114] investigated anticonvulsant property of the hydroalcoholic extract of whole plant using PTZ, MES models in rats. Preliminary phytochemical screening revealed the presence of flavonoids in the extract. Study concluded that the extract at 500 mg/kg, p.o. dose possess CNS depressant action in addition to anticonvulsant action. Flavonoids present in the extract might be responsible for anticonvulsant effects but needs further investigation.

Solanum nigrum L.

S.nigrum commonly known as Black night shade in English and Kakamachi in Tamil belongs to family Solanaceae. Ravi *et al.* [115] investigated the anticonvulsant activity of aqueous and ethanolic extract prepared from shade dried berries powder in albino rats. Ethanolic extract at the doses of 100, 200, 300 mg/kg, p.o. was found to possess anticonvulsant activity against seizures induced by MES. Phytochemical studies have reported the presence of flavonoids in aqueous and ethanolic extract. Exact mechanism of the extract showing anticonvulsant activity is not known but anticonvulsant activity might be due to stimulatory effect on GABA mediated synaptic inhibition which might be due to presence of flavonoids.

Aegle marmelos (L.) Corr.

It is commonly known as Bael belonging to family Rutaceae. Sankari *et al.* [116] investigated the anticonvulsant effects of the ethanolic leaf extract using MES and PTZ induced seizure model in mice. It was found that the administration of the extract at 200 mg/kg p.o. significantly increased the onset, decreased the duration of MES induced tonic hind limb extension. The onset of tonic convulsion produced by PTZ was also significantly delayed. As the phytochemical estimations revealed the presence of flavonoids, the anticonvulsant effect might be due to their enhancing action on GABA-mediated inhibition in the brain which needs further studies.

Vitex agnus castus L.

V. agnus commonly known as Vitagnus or Chaste tree belongs to family Lamiaceae. This plant has been used to treat epilepsy and psychosis in 1200 A.D. and is known as 'Black remedy'. Saberi *et al.* [117] investigated the hydroethanolic extract from the fruits containing flavonoid such as casticin, vitexin and orientin [118] for its possible anticonvulsant activity in electrical kindling model of epilepsy in male albino wistar rats. The extract at 60, 120 or 180 mg/kg *i.p.* doses showed anticonvulsant property indicated by increment in after discharge threshold, reduction in after discharge duration and seizure severity in a dose dependent manner. Anticonvulsant activity might be due to flavonoids having agonist action on kappa opioid receptors.

Ginkgo biloba

G. biloba belonging to family Ginkgoaceae has various uses in traditional medicine and neurodegenerative diseases including the formation of kindling epilepsy & epileptic discharge. Many flavonoids have been isolated from ginkgo leaves like monomer flavonoids (quercetin and kaempferol), dimer flavonoids (biflavone, ginkgetin, sciadopitysin and bilobetin). *G. biloba* extract can be used to prevent the development of epilepsy due to ischemic attacks. This is probably due to Bilobalides which appear to act at sites in the chloride channel of GABA_A receptor and is thus a negative allosteric modulators [119, 120] and due to flavonoids from *G. biloba* extract, for which it has been established that they perform modulation of ionic currents mediated by GABA receptors [121].

Another experimental study reported by Ivetic et al. [122] showed an opposite effect of G. biloba. The experimental study reported that, administration of the extract had pro-epileptogenic effect, probably by conditioning the depolarization shift of the stimulated neurons of the hippocampus. They also established that some constituents of the extract have pro-epileptogenic effect and decrease the effect of antiepileptic (valproate and carbamazepine). These fully opposite observations, epileptogenic and anti-epileptogenic effects of G. biloba extract are mostly explained by the change of the GABA content, NMDA receptors, beta receptors etc. in various parts of the brain. These variations might be due to variation in experimental models, dose levels and difficulties in precise monitoring of epileptogenic effect, especially when the results of clinical studies are analyzed. Thus, whether the *G. biloba* has antiepileptic effect due to presence of flavonoids or it has pro-epileptogenic effect, it needs further investigation to establish the right connection and mechanism involved in epilepsy.

Albizia lebbeck (L.) Benth.

It is commonly known as woman's tongue belonging to family Fabaceae. Its ethanolic leaf extract has been reported to protect the mice from MES and PTZ induced convulsions [108] which contain flavonoids kaempferol and quercetin 3-O-alpha-rhamnopyranosyl(1-->6)-beta-glactopyranosides. Thus the anticonvulsant activity might be due to presence of flavonoids which needs further investigation.

Tanacetum parthenium (L.) Sch. Bip.

T. parthenium commonly known as Feverfew belongs to family Asteraceae. It is a traditional medicinal herb found in many old gardens. The extracts of *T. parthenium* have been used in Danish folk medicine for the treatment of epilepsy. Studies have shown that the ethanolic extract contain flavonoid apigenin which might be showing anticonvulsant effect due to its high affinity for the GABAA-benzodiazepine site [18]. Thus flavonoids could be the active ingredient in the extract for its anticonvulsant activity.

Brahmi ghrita

Brahmi ghrita, one of the panchgavya formulation clamed as antiepileptic in ayurveda contains *Bacopa moneri*. *Acorus calamus, Evolves alsinoids, Sausserea lappa* out of which *Bacopa moneri* and *Acorus calamus* contain flavonoids which might be responsible for their anticonvulsant effects.

Bacopa monniera L. Pennell

It is commonly known as brahmi belongs to family Scrophulariaceae. It is used to provide relief to patients with anxiety or epileptic disorders. Alcoholic leaf extract of *B. monniera* contains large amount of flavonoids [108, 123]. Kaushik *et al.* [124] studied the anticonvulsant activity of ethanolic leaf extract against PTZ, MES, strychnine, hypoxic stress and lithium pilocarpine induced convulsions in albino mice and wistar rats. It was found that the extract at 55 and 50 mg/kg p.o. doses to mice and rats respectively was effective against all the models of epilepsy. Mathewa *et al.* [125] also reported its use in epilepsy associated comorbities such as learning and memory impairment.

Acorus calamus L.

Also known as *Acorus odoratus* and commonly known as Sweet Flag belongs to the family Adoraceae. It contains flavonoid Galangin (5,7-dihdroxyflavonol) [126]. Jayaraman *et al.* [127] evaluated the anticonvulsant activity of this plant using methanolic root extract at 100 and 200 mg/kg p.o. doses against PTZ model of convulsion in swiss albino mice. It was found that the extract significantly increased the latency period and reduced the duration of seizures. The flavonoids present might be responsible for the above action by exhibiting potentiating effect on GABA_{\rm A} receptors.

Origanum vulgare

O. vulgare commonly known as Oregano is an erectly spreading plant having strong aromatic characteristics in leaves belongs to family Lamiaceae. Its leaves contain flavonoids in addition to triterpenoids, sterols, vitamin C, and vitamin A. In India, leaves are used traditionally for epilepsy, bronchitis, asthma, diarrhea, nephrocystolithiasi, fever, indigestion and cough [128]. Flavonoids present in Oregano might be responsible for their anticonvulsant action but further studies in this area need to be done to support this evidence.

Moringa oleifera

Moringa oleifera commonly known as Drumstick belongs to family Moringaceae. It contains flavonoids in addition to alkaloids, tannins, carbohydrates, amino acids, glycosides. It is reported to provide significant protection against strychnine and PTZ induced convulsions [108, 129, 130]. The exact mechanism of its anticonvulsant activity is not known which needs further investigation.

Nyctanthes arbortristis

Commonly known as Night Jasmine or Coral Jasmine belonging to family Oleaceae, it is an indigenous plant native of India. The ethanolic and aqueous leaf extract have shown to provide protected effect against MES and PTZ induced convulsions in mice [108, 131]. Phytochemical screening showed that the extract contain flavonoids & phenolic compounds. Flavonoids present in the extract might be responsible for anticonvulsant action but it needs further investigation.

Artemesia vulgaris L.

vulgaris commonly known as Mugwort belonging to family Asteraceae has a long history of medicinal use as a traditional herbal remedy for epilepsy. It contains twenty known flavonoids which were isolated and identified as tricine, jaceosidine, eupafolin, chrysoeriol, diosmetin, homoeriodictyol, isorhamnetin, apigenin, eriodictyol, luteolin, luteolin 7-glucoside, kaempferol 3-glucoside, kaempferol 7glucoside, kaempferol 3-rhamnoside, kaempferol 3-rutinoside, quercetin 3-glucoside, quercetin 3-galactoside, quercetrin, rutin, and vitexin [132]. The presence of large amount of flavonoids together with anticonvulsant properties highlights the involvement of flavonoids in anticonvulsant action, which needs further investigations.

Abrus precatorius L.

A.precatorius commonly known as Rosary Pea belonging to family Fabaceae is famous for its beautiful but deadly seeds containing toxin abrin. The chloroform soluble fraction of the concentrated 80% methanolic extract of the seeds contains flavonol glycosides which are reported to possess anticonvulsant activity [133]. Flavonol glycoside has been reported to be present in this plant [134]. Thus flavonol glycoside might be responsible for its anticonvulsant action but more studies need to be done to support this evidence.

Citrus aurantium L.

It is commonly known as sour orange belonging to family Rutaceae. It is one of the medicinal plants which have been grown all over the world because of its high medicinal values and as a traditional drug in Islamic medicine. The flowers of this plant have been used to control seizures. Electrospray mass spectrometer (HPLC-ES-MS) analyzed eight flavonoids i.e. isonaringin, naringin, hesperidin, neohesperidin, naringenin, hesperitin, nobiletin and tangeritin from the ethanolic extract of fruits showing anticonvulsant effects [135]. Hesperetin is able to ameliorate the response to tetraethylammonium and PTZ. These flavonoids might be responsible for anticonvulsant action through activation of large conductance calcium dependent potassium channels [136] or agonistic effect on benzodiazepine receptor [137].

Astragalus centralpinus

It is commonly known as milk-vetch which belongs to family Papilionaceae. It contain various flavonoids like 5,7,4'-trihydroxy3,3'dimethoxy flavone, 3,5,7,3'-tetramethoxy-4'-hydroxyflavone, 3,7dihydro-flavone and 4,5-dimethoxy-7- hydroxyflavan-7-ol. These flavonoids are reported to be effective against reversed convulsive action of barium chloride [9].

Hibiscus vitifolius L.

H. vitifolius (Malvaceae) has been reported to contain a bioflavonoid, gossypin. Rasilingam *et al.* [138] studied the anticonvulsant activity of gossypin using PTZ, strychnine and MES-induced convulsion models in mice. Treatment with gossypin (10 and 20 *mg/kg p.o.*) significantly increased the latency to PTZ-induced convulsion. Moreover, treatment with gossypin at 20 mg/kg dose showed significant protection against strychnine and MES-induced convulsions. However more studies are required to understand the exact anticonvulsant sant mechanism of gossypin.

Plant	Family	Reported flavon- oid(s)	Plant part	Type(s) of extract	Dose/ evaluat- ing model(s)/ animal used	Inference	Refer- ence
Abrus pre- catorius L.	Fabaceae	Flavonol glycosides are present. A new flavo- nol glycoside 7,3',5'- trimethoxy-4'-hydroxy flavone-3- O - β - d - galactosyl- $(1 \rightarrow 4)$ - α - l -xyloside has been reported to be present in this plant.	Seeds	Chloroform soluble frac- tion of the concentrated 80% metha- nolic extract.	TNA	Flavonol glycoside might be responsible for its anti- convulsant action but more studies need to be done to support this evidence.	[133, 134].
Abuti- lon indiciu m (L.) Sweet.	Malvaceae	Luteolin, chrysoseriol, luteolin-7-O- β gluco pyranoside, chrysose- riol 7-O- β gluco pyra- noside, apige- nin 7-O- β -gluco pyrano side, Quercetin 3-O- β -g luco pyranoside, Quer- cetin 3-O- α rhamnopyranos yl, β -gluco pyranoside.	Leaves	Ethanolic and aqueous ex- tracts.	100 mg/kg and 400 mg/kg p.o./ PTZ and MES model/ wistar rats.	Ethanolic extract showed better protection than aqueous extract. Ethanolic extract gave significant protection against the PTZ and MES induced convul- sions.	[104- 106]
Acorus	Adoraceae	Galangin (5,7-	Roots	Methanolicex-	100 and 200	Extract significantly in-	[126,

Table 1: It shows the comprehensive list of some herbal plants with anticonvulsant activity.

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calamus L.		dihdroxyflavonol).		tract	mg/kg p.o./ PTZ/ swiss albino mice.	creased the latency period and reduced the duration of seizures. The flavonoids present might be responsi- ble for the above action by exhibiting potentiating effect on GABAA receptors.	127]
Aegle mar- melos (L.) Corr.	Rutaceae	Uncharacterized flavo- noids.	Leaves	Ethanolic ex- tract.	100, 200 mg/kg p.o./ MES and PTZ models/ mice.	Dose of 200 mg/kg p.o. was found to be effective. The anticonvulsant activity might be due to GABA in- hibitory action in the brain.	[116]
Albizia lebbeck (L.) Benth.	Fabaceae	Kaempferol and quer- cetin 3-0-alpha- rhamnopyranosyl(1 >6)-beta- glucopyranosyl(1>6)- beta- galactopyrano- sides	Leaves	Ethanolic ex- tract	MES and PTZ model/ mice.	It have been shown to protect the mice from MES, PTZ induced convulsions.	[108]
Anisomeles malabarica (L.) R. Br	Lamiaceae	Uncharacterized flavo- noids	Leaves	Ethyl acetate.	6.25 and 12.5 mg/kg <i>i.p.</i> for 1 week/ PTZ and MES model/ wistar rats.	Ethyl acetate extract rich in flavonoids ameliorate epi- lepsy in chronic treatment without causing any neuro- toxicity.	[73]
<i>Argyreia speciosa</i> (Burm.f.) Bojer	Convolvu- laceae	Quercetin and kaemp- ferol.	Roots	Hydroal- coholic extrct.	100, 200, 400 mg/kg p.o./ PTZ and MES model/ mice.	The extract significantly delayed the latency to the onset of convulsions, re- duced the duration, death in both the models of con- vulsion and prevented cognitive deficit in convul- sions.	[100, 101]
Artemesia vulgaris L.	Asteraceae	Twenty known flavo- noids- tricine, jaceosi- dine, eupafolin, chry- soeriol, diosmetin, homoeriodictyol, isor- hamnetin, apigenin, eriodictyol, luteolin, luteolin 7-glucoside, kaempferol 3- glucoside, kaempferol 7-glucoside, kaempferol 7-glucoside, kaempferol 3-rhamnoside, kaempferol 3- rutinoside, quercetin 3-glucoside, quercetin 3-glactoside, querce- trin, rutin, and vitexin.	Aerial parts of the plant.	TNA	Further re- search on this topic need to be done.	TNA	[132]
Bacopa monniera L. Pennell	Scrophula- riaceae	Uncharacterized flavo- noids	Leaves	Ethanolic ex- tract	55 and 50 mg/kg p.o. / PTZ, MES, strychnine, hypoxic stress and lithium pilocarpine induced convul- sions/ albino mice and wistar rats respec- tively.	Ethanolic extract is effec- tive against all the models epilepsy as well as in epi- lepsy associated comor- bities such as learning and memory impairment.	[108, 123- 125]
Butea mo- nosperma (Lam.) Taub.	Fabaceae	Uncharacterized flavo- noids.	Leaves	Methanolic extract.	100, 200, 400 mg/kg p.o. / MES, PTZ and strychnine model/mice.	400 mg/kg dose was effec- tive against MES and PTZ but not against strychnine induced seizures.	[112]
Caesalpinia bonducella (L.) Roxb.	Caesalpi- niaceae	Homoisoflavone (bon- ducillin)	Seed ker- nels	Petroleum ether, ethanol, methanol and	600, 800 mg/kg p.o. for 8 days/ PTZ, MES,	The action might be due glycine inhibitory action or through activation of GABA	[89]

				aqueous ex- tract.	strychnine and picrotoxin con- vulsion model/	channels.	
Centella asiatica (L.) Urban.	Umbellife- rare	3-glucosylquercetin, 3- glucosylkaempferol and 7- glucosylkaempferol	Leaves	N-hexane, chloroform, ethyl acetate, n-butanol.	mice. 200 mg/kg b.w. 1 week prior to dose of PTZ/ PTZ model/ male wistar rats.	It was found that the activ- ity levels of Na+, K+- ATPase, Mg2+-ATPase, and Ca2+-ATPase which were decreased in PTZ-induced epilepsy were significantly increased on pre-treatment with different CA extracts.	[95, 96]
Cissus si- cyoides L.	Vitaceae	Kaempferol 3- rhamnoside and quer- cetin 3-rhamnoside.	Aerial parts of the plant.	Hydroalcoholic extract.	300, 600, and 1000 mg/kg <i>i.p.</i> / PTZ model / mice.	Extract showed significant action in protection against PTZ model. The flavanoglu- cosides present shows better pharmacokinetic property the anticonvul- sant effects than flavon- oids.	[97, 98]
Citrus au- rantium L.	Rutaceae	Isonaringin, naringin, hesperidin, neohespe- ridin, naringenin, hes- peritin, nobiletin and tangeritin.	Fruits	Ethanolic ex- tract.	Tetraethylam- monium (TEA) and PTZ model.	Flavonoids might be re- sponsible for anticonvul- sant action through activa- tion of large conductance calcium dependent potas- sium channels or agonistic effect on benzodiazepine receptor.	[135- 137]
Cleome viscosa L.	Capparida- ceae	Uncharacterized flavo- noids.	Seeds	Alcoholic and aqueous ex- tract.	200, 400 and 600mg/kg p.o./ MES and PTZ models/ swiss albino mice.	The anticonvulsive effect might be because of fla- vonoids attributed due to inhibitory effect on voltage dependent sodium chan- nels or due to blockage of glutaminergic excitation.	[88]
Drosera burmanni Vahl	Drosera- ceae	Kaempferol, myricetin, quercetin and hyper- oside.	Entire plant	Alcoholic ex- tract	500 mg/kg p.o./ PTZ model/ swiss albino mice.	Anticonvulsant activity may be attributed due to flavonoids causing reduc- tion of T-type calcium cur- rents, enhancement of GABA _A -BZD receptor activi- ty and blocking of glutame- targic excitation mediated by NMDA receptors.	[84- 87]
Ficus Hispi- da L.	Moraceae	Uncharacterized flavo- noids.	Leaves	Methnolic extract	200 and 400 mg/kg p.o./ Strychnine, picrotoxin, and PTZ models/ mice	400mg/kg dose completely abolished the seizures induced by picrotoxin and strychnine but not with PTZ.	[99]
<i>Ficus syco- morus</i> L. Range	Moraceae	Uncharacterized flavo- noids	Stem, bark	Crude flavono- id rich fraction partitioned from Ethanolic extract	5, 10 and 20 mg/kg <i>i.p.</i> / PTZ and MES mod- els/ mice and chicks respec- tively	Crude flavonoid rich ex- tract revealed a significant activity in MES test and a weak anticonvulsant activi- ty in the subcutaneous PTZ test.	[16]
Glychrrhiza glabra L.	Febaceae	Glabridin (an isofla- van) and isoliquiriti- genin (a flavonoid)	Roots and rhizomes.	Ethanolic ex- tract	10, 30, 100 and 500 mg/kg <i>i.p./</i> PTZ, MES and lithium and pilocarpine models / mice	G. glabra might be active against petitmal or absence seizure but not against generalised tonic-clonic seizures.	[74, 75]
Hibiscus vitifolius L.	Malvaceae	Gossypin	Flowers	Gossypin	10, 20 mg/kg p.o./ PTZ, strychnine and MES models/ mice.	It shows anticonvulsant activity probably by en- hancing GABA transmis- sion and has glycine inhibi- tory activity.	[138]

Hypericum perforatum L.	Hyperica- ceae	Catechin, hyperoside, quercetin, quercetrin, rutin, biapigenin, kaempferol.	Aerial parts	Aqueous and ethanolic ex- tract	.1-1g/kg <i>i.p./</i> PTZ and MES models/ mice.	Extract is effective against petit mal convulsions.	[107- 109]
Mimosa pudica L.	Missocia- ceae	Uncharacterized flavo- noids	Leaves	Decoction	1000–4000 mg/kg <i>i.p.</i> / PTZ and strychnine- model/ mice	The decoction showed protected effect in mice against PTZ and strych- nine-induced seizures.	[110, 11]
Moringa oleifera	Moringa- ceae	Uncharacterized flavo- noids	TNA	TNA	TNA	It is reported to provide significant protection against strychnine and PTZ induced convulsions.	[108, 129, 130]
Nauclea latifolia Sm.	Rubiaceae	Uncharacterized flavo- noids	Bark of dried roots	Aqueous ex- tract of the bark of dried roots	16, 40, 80, and 160 mg/kg <i>i.p./</i> MES, PTZ and strychnine models/ mice.	Flavonoids present in the extract might be showing activity against MES, PTZ and Strychnine induced seizures through benzodi- azepine, GABA or by pro- longing the inactivation of sodium channels.	[113]
Nyctanthes arbortristis	Oleaceae	Uncharacterized flavo- noids	TNA	Ethanolic and aqueous ex- tract of leaves.	TNA	Flavonoids present in the extract might be responsi- ble for anticonvulsant ac- tion	[108, 131]
Ocimum gratissi- mum L.	Lamiaceae	Cirsimaritin (flavone)	Leaves	Methanolic and petroleum ether extracts.	200 and 400 mg/kg p.o./ PTZ model/ mice.	Extracts increased the latency of seizures and death and elicited 50% protection against mortal- ity.	[93, 94]
Passiflora incarnata L.	Passiflora- ceae	Chrysin	Leaves	Hydo-alcoholic extract.	0.05, 0.1, 0.2, 0.4 mg/kg <i>i.p. /</i> PTZ model/ mice.	The active constituent can be flavonoid chrysin which might be acting as a partial agonist at benzodiazepine receptor with micromolar affinity or through opioid receptors. But it still re- mains controversial whether chrysin is actually that active component or not.	[27, 76-78]
Pongamia pinnata (L.) Pierre.	Febaceae	Flavonoids in its leaves. Flavone deriva- tive 'pongol', glabra- chalone and isopon- gachromene from its coods	Leaves	Ethanolic ex- tract	250 mg/kg <i>i.p./</i> PTZ model/ wistar albino rats.	Anticonvulsant action might be by modulating the function of GABA or gluta- mate receptors.	[102, 103]
Scutellaria lateriflora L.	Lamiaceae	Viscidulin III-2-Ο-β-D- glucopyranoside, chryin-6-C-α-L- arabionopyranosyl-8- C-β-D- glucopyranoside, trans-verbascoside, viscidulin III, baicalin, trans-martynoside, oroxylin A-7-O-β-D- glucopyrano- side, wogonoside, baicalein, wogonin.	Above ground parts.	Hydroalcoholic fraction	30, 60, 90, and 150 mg/kg <i>i.p. /</i> Pilocarpine and PTZ model/ rats	Anticonvulsant effect might be due to high affinity for the benzodiazepine binding site of GABA _A receptor.	[79- 83]
Solanum nigrum L.	Solanaceae	Uncharacterized flavo- noids.	Berries	Aqueous and ethanolic ex- tract	100, 200, 300 mg/kg p.o./ MES model/ albino rats.	Ethanolic extract was found to be effective at all the doses. Anticonvulsant activity present might be due to stimulatory effect on GABA mediated synaptic inhibition.	[115]

Sphaeran- thus indicus L.	Compositae	Uncharacterized flavo- noids	Whole plant ex- tract	Hydroalcoholic extract	100, 200 and 500 mg/kg p.o./ PTZ and MES model/ rats.	500 mg/kg p.o. possesses CNS depressant action in addition to anticonvulsant action.	[114]
<i>Tanacetum parthenium</i> (L.) Sch. Bip.	Asteraceae	Apigenin	Finely grounded aerial parts.	TNA	Further re- search on this topic need to be done.	Anticonvulsant effect might be due to high affinity of apigenin for the GABA _A - benzodiazepine site.	[18]
Vitex agnus castus L.	Lamiaceae	Flavonoids such as casticin, vitexin and orientin are present.	Fruits	Hydroalcoholic extrct	60, 120 or 180 mg/kg <i>i.p.</i> / MES model/ male albino wistar rats	Extract showed increment in after discharge thresh- old, reduction in after dis- charge duration and stage 5 duration in a dose de- pendent manner.	[117, 118]
Vitex ne- gundo L.	Lamiaceae	Flavonoids like fla- vones	Leaves	Petroleum, butanol and methanol leaf extracts	MES, Strychnine and PTZ models	<i>V. negundu</i> leaf extracts are not just anticonvulsant but also have synergistic activ- ity with standard anticon- vulsants.	[90- 92]
TNA- Text Not	t Available						

CONCLUSION

Flavonoids emerged as a very valuable class of secondary metabolite having potential for the comprehensive treatment of epilepsy. As future investigations continue, this class may prove to be a rich source of new molecules for the development of new therapeutic agents for the treatment of epilepsy. During our literature review we observed the modulatory role of flavonoids in almost all the neuronal pathways involved in the pathogenesis of epilepsy. Studies carried out till date suggests that flavonoids inhibit voltage gated sodium channels, activate Ca+activated K+ channels, stimulate GABAergic inhibition, interact with opioid receptors, inhibit NMDA receptors and exhibit antioxidant actions via modulation of nitric oxide and xanthine oxidase pathways and by leukocytic immobilization, one or more of these mechanisms are involved in suppression of epileptic seizures. Hispidulin, rutin, hesperetin, naringenin, eriodictyol, chrysin, gossypin, apigenin, kaempferol, myricetin, quercetin, hoslundin, hoslundal, hoslunddiol, morine, amentoflavone, hyperoside, anghyperoside, epicatechin, rutin, silibin and luteolin are some of the important flavonoids that possess one or more of the above mentioned mechanisms.

Copious studies pertaining to the anticonvulsant potential crude extracts containing flavonoids and flavonoids-rich fraction containing uncharacterized flavonoids have been carried out. Thus, it is difficult to reproduce the results of these studies and pinpoint the bioactive flavonoids. Hence, there is a need of phytochemical standardization and bioactivity-guided identification of bioactive flavonoids. Moreover it is expected that bioactivity-guided fractionation of the extracts and the rich fractions will yield new flavonoidal molecules. In several instances even where the active anticonvulsant flavonoids is known, their mechanism of action is missing, which needs further studies.

Since oxidative injury play a vital role in the initiation and progression of epilepsy, current treatment strategies are aimed on reducing oxidative stress to ameliorate tissue damage and to alter the clinical course of the disease [13, 139]. It has been reported that free radical generation induces seizure activity by inactivation of glutamine synthase, thereby permitting an abnormal build-up of excitatory neurotransmitter, glutamate [140] or by inhibition of glutamate decarboxylase, leading to decrease in GABA turnover in turn facilitating abnormal excitation [141]. Oxidative stress also plays an important role in the etiology of seizure induced neuronal death [142]. Since flavonoids shows a strong antioxidant effects as they have a highly reactive hydroxyl group in their structure, therefore provide a direct radical scavenging activity [39], hence ameliorate epileptic condition [13]. In several studies the antioxidant effect of the active flavonoids, flavonoids-rich fraction and flavonoids containing crude extracts was studied in in vitro studies, therefore further in vivo studies are required to further explore their clinical effectiveness and therapeutic potential.

Treatment of epilepsy includes the use of multiple dose regimens for prolonged periods, making the side and adverse effects inevitable to the patients. It has been suggested that these ill effects can be decreased by reducing the dose of antiepileptics and/or prescribing substances that can potentiate the effect of former and decrease unwanted effects [143]. Some studies depicted the synergistic antiepileptic effect of flavonoids with clinically used antiepileptic drugs. For example, apigenin and (-) epigallocatechin potentiates the therapeutic effect of diazepam [26]. As flavonoids are devoid of side effects which are commonly associated with clinically used antiepileptic drugs like, ataxia, amnesia, etc, therefore in future these flavonoids can be supplemented antiepileptic drugs to reduce the dose of later to limit side effects. Flavonoids not only suppress epilepsy but also found to be effective against epilepsy-induced comorbidities. Activation of ERK and CREB pathway helps in treating amnesia, while its antioxidant property helps to mitigate depression (Fig. 4). Flavonoids extracted from plants are known to exert antidepressant like effects in animal models of depression [68, 69].

Flavonoids, mainly flavonols like quercetin and kaempferol are responsible for reversing oxidative stress induced-decrease of hippocampal BDNF and pCREB expression leading to hippocampal neurogenesis and thus, ameliorating depression [37, 38]. Similar improvement is also reported in the learning and memory with the use of flavonoids possibly via prohibiting oxidative stress [144-146].

With few exceptions, in majority of studies flavonoids have been found to cross blood brain barrier, indicated by there observed neurological effects. Youdim *et al.* [147] studied citrus flavonoids, hesperetin, naringenin and their relevant *in vivo* metabolites, as well as the dietary anthocyanins and *in vivo* forms, cyanidin-3-rutinoside and pelargonidin-3-glucoside *in vitro* models (brain endothelial cell lines from mouse (b.END5), rat (RBE4) and ECV304 monolayersco-cultured with C6 glioma cells). The study reported that flavonoids and its metabolites are able to travel through the blood brain barrier which is consistent with compound lipophilicity.

However, there are reports of toxic flavonoid-drug interactions, liver failure, contact dermatitis, hemolytic anemia, and estrogenic-related concerns such as male reproductive health and breast cancer associated with dietary flavonoid consumption [148]. Controlled clinical trials of flavonoids and their potential for toxicity is an understudied aspect till now, therefore it needs due attention and research before it gets too late. It is expected that systematic, preclinical and clinical studies will be undertaken in future in the above mentioned areas, so that the great potential of flavonoids to assist people living with epilepsy and other related ailments can be explored.



Fig. 4: Role of flavonoids in treating epilepsy-induced depression and cognition deficit via ameliorating oxidative stress.

REFERENCES

- 1. Ojewole JAO. Indigenous remedies and epilepsy:evaluation of some South African medicinal plants used as anticonvulsant remedies in Zulu folk medicine. J Blacpma Enero De 2005;4:1-21.
- 2. Yemadje LP, Houinato D, Quet F, Druet-Cabanac M, Preux PM. Understanding the differences in prevalence of epilepsy in tropical regions. J Epilepsia 2011;52:1376-81.
- Macdonald RL, Kelly KM. Antiepileptic drug mechanisms of action. J Epilepsia 1995;36:S2-S12.
- 4. Czapinski P, Blaszczyk B, Czuczwar S J. Mechanisms of action of antiepileptic drugs. J Curr Top Med Chem 2005;5:3-14.
- 5. Rogawski MA. Diverse mechanisms of antiepileptic drugs in the development pipeline. J Epilepsy Res 2006;69:273-94.
- Paulo AM, Maria AM, Marilisa MG, Carlos AM, Guerreiro. Pharmacovigilance in epileptic patients using antiepileptic drugs. J Arquivos de Neuro-Psiquiatria 2006;64:198-201.
- Cramer JA, Mintzer S, Wheless J, Mattson RH. Adverse effects of antiepileptic drugs:a brief overview of important issues. J Expert Rev Neurother 2010;10:885-91.
- 8. Wiebe S, Hesdorffer DC. Epilepsy:being ill in moreways than one. J Epilepsy Curr 2007;7:145-8.
- Moshi MJ, Kagashe GA, Mbwambo ZH. Plants used to treat epilepsy by Tanzanian traditional healers. J Ethnopharmacol 2005;97:327-36.
- Griebel G, Perrault G, Tan S, Shoemaker H, Sanger D. Pharmacological studies on synthetic flavonoids:comparison with diazepam. J Neuropharmacol 1999;38:965-77.
- 11. Du XM, Sun NY, Takizawa N, Guo YT, Shoyama Y. Sedative and anticonvulsant activity of goodyerin, a flavonol glycoside from *Goodyera schlechtendaliana*. J Phytother Res 2002;16:261-63.
- Kavvadias D, Sand P, Youdim KA, Qaiser MZ, Rice-Evans C, Baur R, *et al.* The flavone hispidulin, a benzodiazepine receptor ligand with positive allosteric properties, traverses blood brain barrier and exhibits anticonvulsive effect. Br J Pharmacol 2004;142:811-20.
- 13. Devi PU, Manocha A, Vohora D. Seizures, antiepileptics, antioxidants and oxidative stress:an insight for researchers. J Expert Opin Pharmacother 2008;9:3169-77.

- Fernandez SP, Wasowski C, Paladini AC, Marder M. Synergistic interaction between hesperidin, a natural flavonoid and diazepam. Eur J Pharmacol 2005;512:189-98.
- 15. Nijveldt RJ, Nood EV, Hoorn, Danny ECV, Boelens PG, Norren KV *et al.* Flavonoids:a review of probable mechanisms of action and potential applications. Am J Clin Nutr 2001;74:418-25.
- 16. Ibrahim G, Abdulmumin S, Musa KY, Yaro AH. Anticonvulsant activities of the crude flavonoid fraction of the stem bark of *Ficus sycomorus* (Moraceae). J Pharmacol Toxocol 2008;3:351-6.
- 17. Narayana KR, Reddy MS, Chaluvadi MR, Krishna DR. Bioflavonoids Classification, Pharmacological, Biochemical effects & Therapeutic potential. Indian J Pharmacol 2001;33:2-16.
- 18. Jager AK, Krydsfeldt K, Rasmussen HB. Bioassay-guided isolation of apigenin with GABA-benzodiazepine activity from *Tanacetum parthenium*. J Phytother Res 2009;23:1642-4.
- Nicholson RA, David LS, Pan RL, Liu XM. Pinostrobin from *Cajanus cajan* (L.) Millsp. inhibits sodium channel-activated depolarization of mouse brain synaptoneurosomes. J Fitoterapia 2010;81:826-9.
- Yao Y, Han DD, Zhang T, Yang Z. Quercetin Improves Cognitive Deficits in Rats with Chronic Cerebral Ischemia and Inhibits Voltage-dependent Sodium Channels in Hippocampal CA1 Pyramidal Neurons. J Phytother Res 2010;24:136-40.
- Engelborghs S, Hooge RD, Deyn PPD. Pathophysiology of epilepsy. J Acta Neurol Belg 2000;100:201-13.
- Cogolludo A, Frazziano G, Briones AM, Cobeno L, Moreno L, Federica L, *et al.* The dietary flavonoid quercetin activates BKCa currents in coronary arteries via production of H₂O₂;Role in vasodilatation. J Cardiovasc Res 2007;73:424-31.
- 23. Hanrahan JR, Chebib M, Johnston GA. Flavonoid modulation of GABAA receptors. Br J Pharmacol 2011;163(2):234-45.
- Chebib M, Hinton T, Schmid KL, Brinkworth D, Qian H, Matos S, et al. Novel, potent, and selective GABAC antagonists inhibit myopia development and facilitate learning and memory. J Pharmacol Exp Ther 2009;328:448-57.
- Goutman JD, Waxemberg MD, Oliver FD, Pomata PE, Calvo DJ. Flavonoid modulation of ionic currents mediated by GABA_A and GABA_C receptors. Eur J Pharmacol 2003;461:79-87.

- Campbell EL, Chebib M, Johnston GAR. The dietary flavonoids apigenin and (-)-epigallocatechingallate enhance the positive modulation by diazepam of the activation by GABA of recombinant GA-BA(A) receptors. J Biochem Pharmacol 2004;68:1631-38.
- 27. Medina JH, Viola H, Wolfman C, Marder M, Wasowski C, Calvo D, *et al.* Overview-flavonoids:a new family of benzodiazepine receptor ligands. J Neurochem Res 1997;22:419-25.
- Katavic PL, Lamb K, Navarro H, Prisinzano TE. Flavonoids as opioid receptor ligands:identification and preliminary structure-activity relationships. J Nat Prod 2007;70:1278-82.
- Subash S, Subramanian P. Morin a flavonoid exerts antioxidant potential in chronic hyperammonemic rats:a biochemical and histopathological study. J Mol Cell Biochem 2009;327:153-61.
- Majewska M, Skrzycki M, Podsiad M, Czeczot H. Evaluation of antioxidant potential of flavonoids:an *in vitro* study. J Acta Pol Pharm 2011;68:611-15.
- Ishige K, Schubert D, Sagara Y. Flavonoids protect neuronal cells from oxidative stress by three distinct mechanisms. J Free Radical Bio Med 2001;30:433-46.
- Rice-Evans CA, Miller NJ, Paganga G. Structure-antioxidant activity relationships of flavonoids and phenolic acids. J Free Radic Biol Med 1996;20:933-56.
- Lien EJ, Ren S, Bui HH, Wang R. Quantitative structure-activity relationship analysis of phenolic antioxidants. J Free Radic Biol Med 1999;26:285-94.
- Joshi D, Naidu PS, Singh A, Kulkarni SK. Protective effect of quercetin on alcohol abstinence-induced anxiety and convulsions. J Med Food 2005;8:392-96.
- Nassiri-Asl M, Shariati-Rad S, Zamansoltani F. Anticonvulsive effects of intracerebroventricular administration of rutin in rats. J Prog Neuropsychopharmacol Biol Psychiatry 2008;32:989-93.
- 36. Nassiri-Asl M, Mortazavi SR, Samiee-Rad F, Zangivand AA, Safdari F, Saroukhani S, *et al.* The effects of rutin on the development of pentylenetetrazole kindling and memory retrieval in rats. J Epilepsy Behav 2010;18:50-3.
- An L, Zhang YZ, Yu NJ, Liu XM, Zhao N, Yuan L, et al. Role for serotonin in the antidepressant-like effect of a flavonoid extract of Xiaobuxin-Tang. J Pharmacol Biochem Behav 2008;89:572-80.
- Hou Y, Aboukhatwa MA, Lei DL, Manaye K, Khan I, Luo Y. Antidepressant natural flavonols modulate BDNF and beta amyloid in neurons and hippocampus of double TgAD mice. J Neuropharmacology 2010;58:911-20.
- Middleton EJ. Effect of plant flavonoids on immune and inflammatory cell function. J Adv Exp Med Biol 1998;439:175-82.
- Hanasaki Y, Ogawa S, Fukui S. The correlation between active oxygens scavenging and antioxidative effects of flavonoids. J Free Radical Bio Med 1994;16:845-50.
- 41. Dehmlow C, Erhard J, de Groot H. Inhibition of Kupffer cell functions as an explanation for the hepatoprotective properties of silibinin. J Hepatology 1996;23:749-54.
- 42. Shutenko Z, Henry Y, Pinard E. Influence of the antioxidant quercetin *in vivo* on the level of nitric oxide determined by electron paramagnetic resonance in rat brain during global ischemia and reperfusion. J Biochem Pharmacol 1999;57:199-208.
- Chang WS, Lee YJ, Lu FJ, Chiang HC. Inhibitory effects of flavonoids on xanthine oxidase. J Anticancer Res 1993;13:2165-70.
- 44. Shoskes DA. Effect of bioflavonoids quercetin and curcumin on ischemic renal injury:a new class of renoprotective agents. J Transplantation 1998;66:147-52.
- 45. Cos P, Ying L, Calomme M. Structure-activity relationship and classification of flavonoids as inhibitors of xanthine oxidase and superoxide scavengers. J Nat Prod 1998;61:71-6.
- Bennett JP, Gomperts BD, Wollenweber E. Inhibitory effects of natural flavonoids on secretion from mast cells and neutrophils. J Arzneimittelforschung 1981;31:433-37.
- 47. Friesenecker B, Tsai AG, Allegra C, Intaglietta M. Oral administration of purified micronized flavonoid fraction suppresses leukocyte adhesion in ischemia-reperfusion injury:in vivo observations in the hamster skin fold. Int J Microcirc Clin Exp 1994;14:50-5.
- Friesenecker B, Tsai AG, Allegra C, Intaglietta M. Cellular basis of inflammation, edema and the activity of Daflon 500 mg. Int J Microcirc Clin Exp 1995;15:17-21.

- 49. Parfenova H, Carratu P, Tcheranova D, Fedinec A, Pourcyrous M, Leffler CW. Epileptic seizures cause extended postictal cerebral vascular dysfunction that is prevented by HO-1 overexpression. Am J Physiol Heart Circ Physiol 2005;288:H2843-H50.
- 50. Maes M, Mihaylova I, Kubera M, Uytterhoeven M, Vrydags N, Bosmans E. Coenzyme Q10 deficiency in myalgic encephalomyelitis/chronic fatigue syndrome (ME/CFS) is related to fatigue, autonomic and neurocognitive symptoms and is another risk factor explaining the early mortality in ME/CFS due to cardiovascular disorder. J Neuro Endocrinol Lett 2009a;30:470-76.
- Maes M, Mihaylova I, Kubera M, Uytterhoeven M, Vrydags N, Bosmans E. Increased 8-hydroxy-deoxyguanosine, amarker of oxidative damage to DNA, in major depression and myalgic encephalomyelitis / chronic fatigue syndrome. J Neuro Endocrinol Lett 2009b;30:715-22.
- 52. Maes M, Mihaylova I, Kubera M, Uytterhoeven M, Vrydags N, Bosmans E. Lower plasma coenzyme Q10 in depression:a marker for treatment resistance and chronic fatigue in depression and a risk factor to cardiovascular disorder in that illness. J Neuro Endocinol Lett 2009c;30:462-69.
- Ohmori H, Kanayama N. Immunogenicity of an inflammationassociated product, tyrosine nitrated self-proteins. J Autoimmun Rev 2005;4:224-29.
- 54. Rodriguez-Mora OG, Howe CJ, Lahair MM, McCubrey JA, Franklin RA. Inhibition of CREB transcriptional activity in human T lymphocytes by oxidative stress. J Free Radical Bio Med 2005;38:1653-61.
- 55. Kolosova NG, Shcheglova TV, Sergeeva SV, Loskutova LV. Long-Term antioxidant supplementation attenuates oxidative stress markers and cognitive deficits in senescent accelerated OXYS rats. J Neurobiol Aging 2006;27:1289-97.
- 56. Abdul HM, Butterfield DA. Involvement of PI3K/PKG/ERK1/2 signaling pathways in cortical neurons to trigger protection by cotreatment of acetyl-L-carnitine and α -lipoic acid against HNE-mediated oxidative stress and neurotoxicity:Implications for Alzheimer's disease. J Free Radical Bio Med 2007;42:371-84.
- Clausen A, Doctrow S, Baudry M. Prevention of cognitive deficits and brain oxidative stress with superoxide dismutase/ catalase mimetics in aged mice. J Neurobiol Aging 2010;31:425-33.
- 58. Michael J. Oxidative stress and cognitive longevity. J Nutrition 2010;26:595-603.
- Vauzour D, Vafeiadou K, Mateos AR, Rendeiro C, Spencer JPE. The neuroprotective potential of flavonoids:a multiplicity of effects. J Genes Nutr 2008;3:115-26.
- 60. Spencer JP. The interactions of flavonoids within neuronal signalling pathways. J Genes Nutr 2007;2:257-73.
- Biegon A, Greenberger V, Segal M. Quantitative histochemistry of brain acetylcholineesterase and learning in the aged rat. J Neurobiol Aging 1986;7:215-7.
- 62. Perry EK. Cholinergic component of cognitive impairment in dementia. In:Burns A, Levy R, editors. London:Dementia, Chapman and Hall;1994.p.36-48.
- 63. Kish SJ, Olivier A, Dubeau F, Robitaille Y, Sherwin AL. Increased activity of choline acetyltransferase and acetylcholinesterase in actively epileptic human cerebral cortex. J Epilepsy Res 1988;2:227-23.
- 64. Serra M, Dazzi L, Cagetti E, Chessa MF, Pisu MG, Sanna A, *et al.* Effect of pentylenetetrazole-induced kindling on acetylcholine release in the hippocampus of freely moving rats. J Neurochem 1997;68:313-18.
- 65. Pepeu G, Giovannini MG. Cholinesterase inhibitors and memory. J Chem Biol Interact 2010;187:403-08.
- 66. Adewusi EA, Moodley N, Steenkamp V. Antioxidant and acetylcholinesterase inhibitory activity of selected southern African medicinal plants. S Afr J Bot 2011;77:638-44.
- 67. Katalinic M, Rusak G, Domacinovic Barovic J, Sinko G, Jelic D, Antolovic R, et al. Structural aspects of flavonoids as inhibitors of human butyrylcholinesterase. Eur J Med Chem 2010;45:186-92.
- Messaoudi M, Bisson JF, Nejdi A, Rozan P, Javelot H. Antidepressant-like effects of a cocoa polyphenolic extract in Wistare Unilever rats. J Nutr Neurosci 2008;11:269-76.
- 69. Zhao Z, Wang W, Guo H, Zhou D. Antidepressant-like effect of liquiritin from *Glycyrrhiza uralensis* in chronic variable stress

induced depression model rats. J Behav Brain Res 2008;194:108-13.

- Yi LT, Li CF, Zhan X, Cui CC, Xiao F, Zhou LP, *et al.* Involvement of monoaminergic system in the antidepressant-like effect of the flavonoid naringenin in mice. J Prog Neuropsychopharmacol Biol Psychiatry 2010;34:1223-28.
- Luo L, Sun Q, Mao YY, Lu YH, Tan RX. Inhibitory effects of flavonoids from Hypericum perforatum on nitric oxide synthase. J Ethnopharmacol 2004;93:221-25.
- Xiukun W, Lujun Z, Lei H, Dongming X, Lijun DU. Effect of flavonoids in *Scutellariae Radix* on depression-like behavior and brain rewards:possible in dopamine system. J Tsing Sci and Tech 2010;15:460-66.
- Choudhary N, Bijjem KR, Kalia AN. Antiepileptic potential of flavonoids fraction from the leaves of *Anisomeles malabarica*. J Ethnopharmacol 2011;135:238-42.
- Maurya SK, Raj K, Srivastava AK. Antidyslipidaemic activity of *Glycyrrhiza glabra* in high fructose diet induced dyslipidaemic Syrian golden hamsters. Indian J Clin Biochem 2009;24:404-40.
- 75. Ambawade SD, Kasture VS, Kasture SB. Anticonvulsant activity of roots and rhizomes of *Glycyrrhiza glabra*. Indian J Pharmacol 2002;34:251-55.
- 76. Speroni E, Minghetti A. Neuropharmacological activity of extracts from *Passiflora incarnata*. J Planta Med 1988;54:488-91.
- Wolfman C, Viola H, Paladini A, Dajas F, Medina JH. Possible anxiolytic effects of chrysin, a central benzodiazepine receptor ligand isolated from *Passiflora coerulea*. J Pharmacol Biochem Behav 1994;47:1-4.
- Dhawan K, Dhawan S, Sharma A. Passiflora:a review update. J Ethnopharmacol 2004;97:1-23.
- 79. Zhang Z, Lian XY, Li S, Stringer JL. Characterization of chemical ingredients and anticonvulsant activity of American skullcap (*Scutellaria lateriflora*). J Phytomedicine 2009;16:485-93.
- Park HG, Yoon SY, Choi JY, Lee GS, Choi JH, Shin CY, et al. Anticonvulsant effect of wogonin isolated from *Scutellaria baicalen*sis. Eur J Pharmacol 2007;574:112-9.
- Wang H, Hui KM, Chen Y, Xu S, Wong JT, Xue H. Structureactivity relationships of flavonoids, isolated from *Scutellaria baicalensis*, binding to benzodiazepine site of GABA_A receptor complex. J Planta Med 2002;68:1059-62.
- Cho J, Lee HK. Wogonin inhibits excitotoxic and oxidative neuronal damage in primary cultured rat cortical cells. Eur J Pharmacol 2004;485:105-10.
- 83. Liu LY, Wei EQ, Zhao YM, Chen FX, Wang ML, WP Zhang, *et al.* Protective effects of baicalin on oxygen/glucose deprivation and NMDA-induced injuries in rat hippocampal slices. J Pharm Pharmacol 2005;57:1019-26.
- Ayuga C. Contribution to study of flavonoids in *Drosera rotundifolia*. J L An R Acad Farm 1985;51:321-6.
- Hema B, Bhupendra S, Mohamed STS, Gauthaman K. Anticonvulsant effect of *Drosera burmannii* Vahl. Int J Appl Res Nat Prod 2009;2:1-4.
- 86. Manigauha A, Patel S. Anticonvulsant study of *Pongamia pinnata* Linn against PTZ induced convulsion in rats. Int J of Pharm and Bio Sci 2010;1:1-4.
- Tan Z. Neural protection by naturopathic compounds-an example of tetramethylpyrazine from retina to brain. J Ocul Boil Dis Inform 2009;2:57-64.
- 88. Mishra A, Bhatti R, Singh A, Ishar MPS. Ameliorative effect of cinnamon oil from *Cinnamonum zeylanicum* upon early stage of diabetic nephropathy. J Planta Med 2010;76:412-7.
- 89. Ali A, Rao NV, Shalam MD, Gouda TS, Shantakumar SM. Anticonvulsive Effect of Seed Extract of *Caesalpinia bonducella* (Roxb.). Iran J Pharmacol Therp 2009;8:51-5.
- 90. Gupta M, Mazumdar UK, Bhawal SR. CNS activity of *Vitex negundo* Linn. in mice. Indian | Exp Biol 1997;37:143-6.
- 91. Tandon VR. Medicinal uses and biological activities of *Vitex negundo*. J Nat Prod Rad 2005;4:162-5.
- Gupta RK, Tandon VR. An experimental evaluation of anticonvulsant activity of *Vitex negundo*. Indian J Physiol Pharmacol 2005;49:199-205.

- Vieira RF, Grayer RJ, Paton A, Simon JE. Genetic diversity of *Ocimum gratissimum* L. based on volatile oil constituents, flavonoids and RAPD markers. J Biochem Syst Ecol 2001;29:287-304.
- 94. Okoli CO, Ezike AC, Agwagah OC, Akah PA. Anticonvulsant and anxiolytic evaluation of leaf extracts of *Ocimum gratissimum*, a culinary herb. J Phcog Res 2010;2:36-40.
- 95. Jamil SS, Nizami Q, Salam M. *Centella asiatica* linn. Urban oA Review. J Nat Prod Rad 2007;6:158-70.
- 96. Visweswari G, Siva PK, Lokanatha V, Rajendra W. The antiepileptic effect of *Centella asiatica* on the activities of Na⁺, K⁺-ATPase, Mg²⁺-ATPase, and Ca²⁺-ATPase in rat brain during pentylenetetrazol-induced epilepsy. Indian J Pharmacol 2010;42:82-6.
- 97. Beltrame F, Ferreira A, Cortez D. Coumarin glycoside from *Cissus sicyoides*. J Nat Prod Lett 2002;16:213-6.
- 98. de Almeida ER, Rafael KR, Couto GB, Ishigami AB. Anxiolytic and anticonvulsant effects on mice of flavonoids, linalool, and alpha-tocopherol presents in the extract of leaves of Cissus sicyoides L. (Vitaceae). J Biomed Biotechnol 2009;2009:1-6.
- 99. Dhanasekaran S, Palayan M. Sedative and anticonvulsant activities of the methanol leaf extract of *Ficus hispida* Linn. J Drug Inven Today 2009;1:23-7.
- 100. Vyawahare NS, Bodhankar SL. Anticonvulsant activity of *Argyreia speciosa* in mice. Indian J Pharm Sci 2009;71:131-4.
- 101. Joshi H, Kaur N, Chauban J. Evaluation of Nootropic effect of *Argyreia speciosa* in mice. J Health Sci 2007;53:382-8.
- 102. Arote SR, Yeole PG. *Pongamia pinnata* L:A Comprehensive Review. Int J Pharm Tech Res 2010;2:2283-90.
- 103. Manigauha A, Patel S. Anticonvulsant study of *Pongamia pinna-ta* Linn against PTZ induced convulsion in rats. Int J Pharm Bio Sci 2010;1:1-4.
- 104. Gupta M, Shaw BP. Uses of medicinal plants in Panchakarma ayurvedic therapy. IJTK 2009;8:372-8.
- Matlawska I, Silkorska M. Flavonoid compounds in the flowers of *Abutilon indicum* (Linn.) Sweet. Acta Pol Pharm 2002;59:227-9.
- 106. Golwala DK, Patel LD, Vaidya SK, Bothara SB, Mani M, Patel P. Anticonvulsant activity of *Abutilon indicum* leaf. Int J Pharmacy Pharm Sci 2010;2:66-71.
- 107. Vattikuti UMR, Ciddi V. An overview on *Hypericum perforatum* Linn. J Nat Prod Rad 2005;4:368-81.
- 108. Vyawahare NS, Khandelwal AR, Batra VR, Nikam AP. Herbal anticonvulsants. J Herb Med Toxicol 2007;1:9-14.
- Hosseinzadeh H, Karimi GR, Rakhshanizadeh M. Anticonvulsant effect of *Hypericum perforatum*:role of nitric oxide. J Ethnopharmacol 2005;98:207-8.
- 110. Bum EN, Dawacka DL, Schmutzb M, Rakotonirinac A, Rakotonirinac SV, Portetb C, *et al*. Anticonvulsant activity of *Mimosa pudica* decoction. J Fitoterapia 2004;75:309-14.
- 111. Sutar NG, Sutar UN, Behera. Antidiabetic activity of the leaves of *Mimisa pudica* Linn in albino mice. J Herb Med Toxicol 2009;3:123-6.
- 112. Silambujanaki P, Chitra V, Kumari S, Sankari M, Raju D, Tejo B, et al. Anti-convulsant activity of methanolic extract of *Butea* monosperma leaves. Res J Pharm Bio Chem Sci 2010;1:431-5.
- 113. Bum EN, Taiwe GS, Moto FCO, Ngoupaye GT, Nkantchoua GCN, Pelanken MM, *et al.* Anticonvulsant, anxiolytic, and sedative properties of the roots of *Nauclea latifolia* Smith in mice. J Epilepsy Behav 2009;15:434-40.
- 114. Galani VJ, Patel BG. Effect of hydoalcoholic extract of *Sphaeranthus indicus* against experimentally induced anxiety, depression, convulsions in rodents. Int J Ayurveda Res 2010;1:87-92.
- 115. Ravi V, Saleem TSM, Maiti PP, Gauthaman K, Ramamurthy J. Phytochemical and pharmacological evaluation of *Solanum nigrum*. Afr J Pharm Pharacol 2009;3:454-7.
- 116. Sankari M, Chitra V, Silambujanaki P, Raju D. Anticonvulsant activity of ethanolic extract of *Aegle marmelos* (Leaves) in mice. Int J Pharm Tech Res 2010;2:640-3.
- 117. Saberi M, Rezvanizadeh A, Bakhtiarian A. The antiepileptic activity of *Vitex agnus* castus extract on amygdala kindled seizures in male rats. J Neurosci Lett 2008;441:193-6.
- 118. Hajdu Z, Hohmann J, Forgo P, Martinek T, Dervarics M, Zupko I, *et al.* Diterpenoids and flavonoids from the fruits of *Vitex ag*-

nus-castus and antioxidant activity of the fruit extracts and their constituents. J Phytother Res 2007;21:391-4.

- 119. Sasaki K, Hatta S, Wada K, Ohshika H. Bilobalide prevents reduction of γ-aminobutyric acid levels and glutamic acid decarboxylase activity induced by 4-0-methylpyridoxine in mouse hippocampus. J Life Sci 2000;67:709-15.
- 120. Sasaki K, Wada K, Hatta S, Ohshika H, Haga M. Bilobalide, a constituent of Ginkgo biloba L., potentiates drug-metabolizing enzyme activities in mice:possible mechanism for anticonvulsant activity against 4-0-methylpyridoxine-induced convulsions. J Res Commun Mol Pathol Pharmacol 1997;96:45-56.
- 121. Heim KE, Tagliaferro AR, Bobilzya DJ. Flavonoid antioxidants:chemistry, metabolism and structure-activity relationships. J Nutr Biochem 2002;13:572-84.
- 122. Ivetic V, Popovic M, Naumovic N, Radenkovic M, Vasic V. The effect of *Ginkgo biloba* (EGb 761) on epileptic activity in rabbits. J Molecules 2008;13:2509-20.
- 123. Bandyopadhyay SK. Ischaemia induced oxidative stress in brain and its management with natural anti-oxidants. Bomb Hosp J 2009;51:460-71.
- 124. Kaushik D, Tripathi A, Tripathi R, Ganachari M, Khan SA. Anticonvulsant activity of *Bacopa monniera* in rodents. Braz J Pharm Sci 2009;45:643-9.
- 125. Mathewa J, Paula J, Nandhua MS, Paulose CS. *Bacopa monnieri* and Bacoside-A for ameliorating epilepsy associated behavioral deficits. J Fitoterapia 2010;81:315-22.
- 126. Raja AE, Vijayalaksshmi M, Devalarao G. *Acorus calamus* Linn:Chemistry and Biology. Res J Pharm and Tech 2009;2:256-61.
- 127. Jayaraman R, Anitha T, Joshi VD. Analgesic and anticonvulsant effects of *Acorus calamus* roots in mice. Int J of Pharm Tech Res 2010;2:552-5.
- Dragoeva AP, Nanova ZD, Kalcheva VK. Allelopathic activity of micropropagated *Origanum vulgare* ssp. hirtum and its effect on mitotic activity. J Allelopathy 2008;22:131-42.
- 129. Gupta M, Mazumdar UK, Chakrabrati S. CNS activities of methanolic extract of *Moringa oleifera* root in mice. J Fitoterapia 1999;70:244-50.
- 130. Rastogi T, Bhutda V, Moon K, Aswar PB, Khadabadi SS. Comparative studies on anthelmintic activity of *Moringa oleifera* and *Vitex negundo*. Asian J Res Chem 2009;2:181-2.
- 131. Singh D, Singh SK, Maurya VB, Prajapati K, Kumar H, Niranjain PS, *et al.* Evaluation of anticonvulsant activity of the leaves ethanolic and aqueous extracts of *Nyctanthes arbortristis* Linn. against seizures induced by PTZ and electroconvulsive shock in mice. Int J Pharm Sci Res 2010;1:63-71.
- 132. Lee SJ, Chung HY, Maier CGA, Wood AR, Dixon RA, Mabry TJ. Estrogenic Flavonoids from *Artemisia vulgaris* L. J Agric Food Chem 1998;46:3325-29.

- 133. Yadava RN, Madhu S, Reddy V. A new biologically active flavonol glycoside from the seeds of *Abrus precatorius* Linn. J Asian Nat Prod Res 2002;4:103-7.
- 134. He X, Lian L, Lin L, Bernart MW. High-performance liquid chromatography-electrospray mass spectrometry in phytochemical analysis of sour orange (*Citrus aurantium* L.). J Chromatogr A 1997;791:127-34.
- 135. Dimpfel W. Different anticonvulsive effects of hesperidin and its aglycone hesperetin on electrical activity in the rat hippocampus *in vitro*. J Pharm Pharmacol 2006;58:375-79.
- 136. Mahmoodi M, Zohoor A, Asadi M. Anticonvulsant effect of sour orange flowers extract in experimental pentylenetetrazolinduced seizures in rat. J Arch Iranian Med 2003;6:212-3.
- 137. Chauhan AK, Dobhal MP, Joshi BC. A review of medicinal plants showing anticonvulsant activity. J Ethnopharmacol 1988; 22:11-23.
- 138. Rasilingam D, Duraisamy S, Subramanian R. Anticonvulsant activity of bioflavonoid gossypin. Bangladesh J Pharmacol 2009;4:51-4.
- 139. Costello DJ, Delanty N. Oxidative injury in epilepsy:potential for antioxidant therapy? J Expert Rev Neurother 2004;4:541-53.
- 140. Oliver CN, Starve-Reed PE, Stadtman ER, Floyd RA. Oxidative damage to brain proteins, loss of free radicals during ischemia/ reperfusion induced injury to gerbil brain. J Proct Natt Acad Sci USA 1990;87:5144-47.
- 141. Halliwell RF, Davey PG, Lambert JJ. The effects of quinolones and NSAIDs upon GABA-evoked currents recorded from dorsal root ganglion neurones. J Antimicrob Chemother 1991;27:209-18.
- 142. Patel MN. Oxidative stress, mitochondrial dysfunction and epilepsy. J Free Radic Res 2002;6:1139-46.
- 143. Chimakurthy J, Murthy TEGK, Upadhyay L. Effect of Curcumin on sub-therapeutic doses of AED's and long term memory in mice induced GTC type of seizures in rats. Res J Pharm and Tech 2008;1:401-4.
- 144. Kumar A, Dogra S, Prakash A. Protective effect of curcumin (*Curcuma longa*), against aluminium toxicity:Possible behavioral and biochemical alterations in rats Behavioural. J Brain Res 2009;205:384-90.
- 145. Mehla J, Reeta KH, Gupta P, Gupta YK. Protective effect of curcumin against seizures and cognitive impairment in a pentylenetetrazole-kindled epileptic rat model. J Life Sci 2010;87:596-603.
- 146. Sharma V, Nehru B, Munshi A, Sharma S, Khanna P. Protective effect of curcumin in behavioral impairment induced by Pentylenetetrazole in rats. J Pharm Res 2011;4:11-4.
- 147. Youdim KA, Dobbie MS, Kuhnle G, Proteggente AR, Abbott NJ, Rice-Evans C. Interaction between flavonoids and the bloodbrain barrier:in vitro studies. J Neurochem 2003;85:180-92.
- 148. Galati G, O'Brien PJ. Potential toxicity of favonoids and other dietary phenolics:significance for their chemopreventive and anticancer properties. J Free Radic Biol Med 2004;37:287-303.