

# Persistence of fear memory across time requires the basolateral amygdala complex

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Mammals evolved a potent fear-motivated defensive system capable of single-trial fear learning that shows no forgetting over the lifespan of the animal. The basolateral amygdala complex (BLA) is considered an essential component of this conditional fear learning system. However, recent studies challenge this view and suggest that plasticity within other brain regions (i.e., central nucleus of the amygdala) may be crucial for fear conditioning. In the present study, we examine the mnemonic limits of contextual fear conditioning in the absence of the BLA using overtraining and by measuring remote fear memories. After excitotoxic lesions of the BLA were created, animals underwent overtraining and were tested at recent and remote memory intervals. Here we show that animals with BLA lesions can learn normal levels of fear. However, this fear memory loses its adaptive features: it is acquired slowly and shows substantial forgetting when remote memory is tested. Collectively, these findings suggest that fear-related plasticity acquired by brain regions outside of the BLA, unlike those acquired in the intact animals, do so for a relatively time-limited period.

fear conditioning | forgetting | remote memory | savings

Failure to defend against an environmental threat such as predation exacts an extreme cost on adaptive fitness (1). Unlike a single missed feeding or mating opportunity, a single failure to defend means you will have no future opportunities to pass on your genes. As a consequence, mammals have evolved a potent fear-motivated defensive system that is capable of single-trial learning (2) and shows no forgetting over the lifespan of the animal (3). Prior studies suggest that such memories are normally established and permanently maintained within the basolateral complex of the amygdala (BLA) (3–7). Typically, damage to the BLA eliminates the acquisition and expression of Pavlovian fear memories across a wide spectrum of mammals, including humans (8) and rodents (5, 6, 9–13). Both electrophysiological and molecular markers of neural activity within the BLA reveal a learning- and retrieval-specific pattern of activation (14–18). Moreover, blockade of NMDA receptors or de novo protein synthesis within the BLA disrupts the acquisition and consolidation of fear memories (19–23). In addition, regardless of whether fear memories are 1 day or 1.5 years old, posttraining BLA lesions completely abolish the expression of fear (3). Collectively, such findings are consistent with a mnemonic role of the BLA in fear learning.

However, recent studies have challenged this view and suggest that plasticity in other brain regions are capable of supporting fear conditioning (24, 25). Indeed, there is growing evidence that fear conditioning can be established in the absence of the BLA. In particular, deficits resulting from either lesions or inactivations of the BLA can be overcome with extensive overtraining (7, 25–27). Furthermore, both Zimmerman et al. (25) and Wilensky et al. (24) have recently shown that disruption of central nucleus of the amygdala (CEA) function can prevent fear conditioning.

In light of these recent findings that suggest a role of BLA independent memory systems in fear conditioning, we wanted to investigate the mnemonic limits of contextual fear conditioning established in the absence of the BLA by measuring memory retention at recent (1 day) and remote (7 and 30 days) long-term

memory intervals. In addition, we examined whether the absence of fear expression at these later retention intervals was due to either a retrieval or storage failure by testing for savings (experiment 2). Here we show that fear learning that occurs in the absence of the BLA, unlike BLA-based learning, reveals extreme deficits in the retrieval of remote memory and suggests that the storage of such memories rapidly decays as a function of time.

## Results

**Experiment 1: Context Fear Acquisition and the Retention of Recent and Remote Long-Term Memory in BLA Lesioned Rats.** The first experiment examined the acquisition of contextual fear conditioning and the retention of remote memory in BLA-lesioned animals. During surgery, animals received either pretraining excitotoxic (NMDA 20 mg/1 mL PBS;  $n = 20$ ) or sham lesions ( $n = 14$ ) of the BLA (Fig. 1). Seven animals from the BLA-lesioned group were excluded from the present analysis because of either unilateral damage to the BLA or bilateral damage that included the CEA. To examine the acquisition of fear, rats were overtrained with 76 presentations of a footshock (1 mA) in a novel context (26–28), during which time we used an online assessment of context conditioning as measured by postshock freezing (28). Animals were returned to the original training context (8 min) at either 1 or 30 days after acquisition to assess recent and remote memory retention, respectively (Fig. 1). Next, to examine the extent of the lesions, animals were overdosed (sodium pentobarbital), and their brains were fixed, extracted, and prepared for NeuN staining procedures (Fig. 2A).

Sham rats showed significant learning after one trial, and learning was asymptotic after the second trial (Fig. 3A). However, rats with excitotoxic lesions of the BLA acquired the freezing response at a significantly slower rate, reaching asymptotic levels close to those of the sham animals by the 20th trial. Such impressions were confirmed by a reliable 2-factor ANOVA that indicated a between group effect of lesion [ $F(1, 150) = 7.43$ ;  $P = 0.01$ ] and a repeated measure factor for trials [ $F(6, 150) = 21.10$ ;  $P < 0.0001$ ]. There was also a reliable lesion by trial interaction [ $F(6, 150) = 2.28$ ;  $P < 0.05$ ]. When memory was tested 24 h later, BLA- and sham-lesioned rats expressed nearly identical levels of fear (Fig. 3B). A separate group of sham-lesioned animals showed no evidence of forgetting, when tested 30 days later. However, BLA-lesioned animals expressed robust levels of freezing during overtraining and failed to express a significant level of contextual fear at this 30-day interval. This was confirmed by  $2 \times 2$  factorial ANOVA revealing a reliable interaction between lesion and retention interval [ $F(1, 24) = 6.20$ ;  $P < 0.05$ ].

## Experiment 2: Savings of Fear at Remote Memory Retention Intervals.

Fear memories are relatively stable across time and show virtually no forgetting. However, animals that learned fear

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structures outside of the BLA, as required by memory modulation of BLA function.

Given the present findings, recent studies have implicated a mnemonic role for the CEA similar to the BLA in the development and storage of fear memories. In both studies by Zimmerman et al. (25) and Wilensky et al. (24), either reversible inactivations or protein synthesis inhibition of the CEA disrupted the acquisition of auditory fear conditioning. In addition, Maren and colleagues (25) demonstrated that permanent or reversible lesions of the CEA disrupted the development of contextual fear conditioning, despite use of an overtraining procedure similar to the one used in the present study. Moreover, a recent paper by Kolber et al. (32) showed that the specific deletion CEA glucocorticoid receptors disrupted fear acquisition and could be subsequently rescued by intracranial ventricular infusions of corticotrophin releasing hormone (CRH). The findings presented here, and previous studies indicating a role of the CEA, suggest a reevaluation of the present serial model of fear conditioning and a possible dissociation of the contributions of the CEA and BLA in the acquisition and storage of fear memories. Further investigations of interactions between the BLA and CEA as well as other fear-related circuitry, such as the bed nuclei of the stria terminalis and ventral medial hypothalamus, could provide a more detailed analysis of neural circuits underlying fear conditioning. Interestingly, all 4 of these regions, which receive afferents from the hippocampus and send projections to the ventral periaqueductal gray (the final output in the expression of freezing), express CRH receptors and, if lesioned, attenuate fear responding (32–36). Our data are consistent with the suggestion that plasticity in regions outside of the BLA can support the acquisition of fear (24, 25). However, these forms of plasticity do not appear to be able to sustain a memory for the extraordinary long periods that plasticity in the BLA can support (3).

The present data indicate that the BLA is a crucial component of remote contextual fear memory. Recently there has been much attention to prefrontal cortical structures that are involved in remote contextual fear memories (37–39). The genetic modification of the alpha-CaMKII that affected remote memory was also not only present in the cortex but in the BLA as well (38). The data presented here indicates that the BLA is necessary for the establishment and/or maintenance of remote memory. Knowledge of the molecular events that mediate permanent memory remains elusive. Contrasting differences in the mechanisms required for the plasticity supporting similar fear memories established with and without the BLA may provide insight into these mechanisms. In conclusion, the present findings suggest that an important adaptive quality of fear memory—its persistence across time—may be an exclusive property of the BLA.

## Methods

**Subjects.** Subjects were 141 male Long-Evans hooded rats (300–450 g; Harlan) were individually housed on a 12-h light/dark cycle in hanging cages and provided ad lib access to food and water. All rats were cared for in accordance with guidelines established by the University of California, Los Angeles, and approved by the UCLA Chancellor's Animal Research Committee.

**Surgery.** Rats were anesthetized with an i.p. injection of sodium pentobarbital (65 mg/kg) and atropine methyl nitrate 0.04 mL (0.1 mg/kg). An incision was made across the midline of the scalp in which the tissue and skin were retracted, exposing the skull. A craniotomy was performed over the site of cannula insertion. The orientation of the skull was adjusted so that lambda and bregma aligned in the same horizontal plane. Infusion cannulae (33 gauge) were positioned 2.6 and 3.3 mm posterior, 5.0 mm lateral, and 8.0 and 8.2 mm ventral to bregma. Infusions of NMDA (20 mg/mL of PBS) were made via 5- $\mu$ L microsyringe (Hamilton Instruments) connected to polyethylene (PE20) tubing attached to infusion cannula. Compression of the syringe was produced by a syringe pump (Harvard Apparatus) at a rate of 0.1  $\mu$ L/min. Total volume infused at each of the 4 sites was 0.2  $\mu$ L. At the termination of the infusion, cannulae were maintained at the site of infusion for an additional 2 min to allow for the diffusion of NMDA from the tip of the cannulae. Stainless steel wound clips were applied along the extent of the incision. Thereafter, all animals were given the antibiotic baytril in

their drinking water for the next 5 days, and i.p. injections of the analgesic/anti-inflammatory ketoprofen (2 mg/kg) for 2 days after surgery. Rats were allowed a total of 10 days to recover from surgery.

**Apparatus.** Context A consisted of 6 clear plastic observation chambers (28  $\times$  21  $\times$  21 cm; Lafayette Instruments). The floor of each chamber consisted of 18 stainless steel rods spaced 1.5 cm apart. The rods were wired to a shock generator and scrambler, which were controlled by a programmable stimulus controller (MED Associates). An electrical fan provided background noise during all training sessions. Before and after each training session a 5% solution of sodium hydroxide was used to wipe clean the interior of each chamber. In addition, below the rod floor of each chamber, a stainless steel pan was coated with benzaldehyde solution (5  $\mu$ L/2 mL of 100% ethanol) as a background odor before each session.

Context B consisted of 6 distinct plastic observation chambers with the sidewalls collapsed at the top to form an A-shaped interior. The floor of context B chamber consisted of 18 stainless rods vertically staggered to be spaced 1.5 cm apart. A white noise generator provided background noise during the use of context B. Before and after each training session a 2% solution of acetic acid was used to wipe clean the interior of each chamber. In addition, below the rod floor of each chamber a stainless steel pan was coated with this same solution as a background odor before each session. The chamber activities were recorded via video cameras positioned on the wall opposite to each pair of chambers. Rats were recorded continuously during overtraining and test sessions.

**Behavioral Training.** All animals were transported to observation chambers in their home cages. Overtraining consisted of the delivery of 76 footshocks (1 mA; 2 s) spaced 64 s apart. Two minutes following placement in the observation chamber, the first footshock was delivered. Rats in experiment 1 were then assigned to 1 of 4 groups: sham/1 day, sham/30 day, lesion/1 day, and lesion/30 day. Sham/1-day and lesion/1-day animals were returned to the observation chamber to measure context fear at 24 h, and sham/30-day and lesion/30-day animals were returned to observation chambers at 30 days following the overtraining session. Rats in experiment 2 were assigned to 1 of 8 groups: savings (sham/7 day, sham/30 day, lesion/7 day, lesion/30 day), sensitization (lesion/7 day, lesion/30 day), and single trial (lesion/1 day, lesion/30 day). Savings and sensitization groups received overtraining followed by 7- and 30-day test of context fear as described in experiment 1.

Twenty-four hours following context testing, savings animals were returned to the original observation chamber, and sensitization animals were placed in a novel observation chamber (odor: acetic acid; background noise: white noise; background illumination: red fluorescent lighting; chamber floor: vertically staggered rods; and chamber walls: A-shaped white opaque plastic) and after 2 min received a single footshock (1 mA; 2 sec). The next day, all animals were returned to the same observation chamber for a savings or sensitization test. Single-trial animals, instead of receiving overtraining, were placed in observation chambers and, following 2 min, received a single footshock and were returned 1 day or 30 days later to measure context fear. Twenty-four hours following context testing, animals were returned to the original observation chamber and, following 2 min, received a single footshock (1 mA; 2 sec). The next day, all animals were returned to the same observation chamber for a savings test. All test sessions consisted of animals being placed in the observation for a total of 8 min and 32 s.

**Behavioral Measures.** Observation of freezing behavior was used as an index of conditional fear. Freezing was defined as the lack of movement, with the exception of those related to respiration. Measurements of freezing were taken from videotaped recording of each overtraining and test session by an observer blind to the experimental condition of each rat. Freezing responses were transcribed in 64-s epochs every 8 s for each rat.

During overtraining, postshock freezing behaviors were scored after the first, second, fifth, 10th, 20th, 30th, 40th, 50th, 60th, 70th, and 76th footshocks, whereas during test sessions, freezing was scored throughout the entire session.

**Histology and Immunohistochemistry.** Following training procedures, all rats were deeply anesthetized with sodium pentobarbital and intracardially perfused with 0.9% saline solution followed by 4% paraformaldehyde solution. The brain was extracted from the skull, placed in a 4% solution of paraformaldehyde for 1–2 days, and then placed in a 30% solution of sucrose for another 1–2 days. Brains were then placed in a cryostat at  $-18^{\circ}\text{C}$  and cut into 50- $\mu$ m coronal sections. Sections that included the amygdala were placed in holding wells that contained 1  $\times$  PBS.

**Immunohistochemistry.** Brain sections were washed in PBS for 5 min (3 times) on a mechanical shaker and placed in a blocking solution (10% goat serum, PBS, and 0.5% Triton) for 1 h. Sections were then placed in NeuN antibody (1:4,000) prepared in 2% goat serum solution (PBS and 0.25% Triton) at 4 °C for 24 h. Tissues were then rinsed in PBS (0.25% Triton) for 15 min (4 times) and transferred to mouse antibody IgG at room temperature for 1 h and 15 min. After the sections were rinsed in PBS for 10 min (4×), they were placed in a solution of avidin-biotin-peroxidase complex on a shaker for 1 h. Sections were rinsed once more in PBS (6 times) and transferred into wells containing 3–3' diaminobenzidine tetrahydrochloride (Sigma) as a

chromagen. Sections were free floated onto subbed microscope slide and coverslipped.

Verification of lesions was made using NeuN-stained tissue viewed under a microscope (Carl Zeiss Inc.) magnified at 1.25× and 4×. The region of the amygdala was outlined and examined for NeuN staining. The lack of staining in this region was used as an indication of neuronal loss due to lesions performed during surgery.

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