

Targeting hypoxia in cancer therapy

William R. Wilson and Michael P. Hay

Abstract | Hypoxia is a feature of most tumours, albeit with variable incidence and severity within a given patient population. It is a negative prognostic and predictive factor owing to its multiple contributions to chemoresistance, radioresistance, angiogenesis, vasculogenesis, invasiveness, metastasis, resistance to cell death, altered metabolism and genomic instability. Given its central role in tumour progression and resistance to therapy, tumour hypoxia might well be considered the best validated target that has yet to be exploited in oncology. However, despite an explosion of information on hypoxia, there are still major questions to be addressed if the long-standing goal of exploiting tumour hypoxia is to be realized. Here, we review the two main approaches, namely bioreductive prodrugs and inhibitors of molecular targets upon which hypoxic cell survival depends. We address the particular challenges and opportunities these overlapping strategies present, and discuss the central importance of emerging diagnostic tools for patient stratification in targeting hypoxia.

Hypoxia influences many aspects of the biology of tumours and their responses to therapy. Initially, hypoxia arises because of oxygen diffusion limitations in avascular primary tumours or their metastases, but the tumour microvasculature (induced in part as a response to this hypoxia) is highly abnormal^{1,2} and often fails to rectify the oxygen deficit. This persistent hypoxia reflects the spatial disorganization of tumour vascular networks, leading to intercapillary distances that are often beyond the diffusion range of oxygen (which is up to ~200 µm, depending on the local oxygen concentration in blood plasma). In addition to this diffusion-limited hypoxia, temporally unstable blood flow in tumour microvascular networks also leads to fluctuating perfusion-limited hypoxia³.

The many effects of hypoxia on tumour biology include: selection of genotypes favouring survival under hypoxia–re-oxygenation injury (such as *TP53* mutations⁴); pro-survival changes in gene expression that suppress apoptosis⁵ and support autophagy⁶; and the anabolic switch in central metabolism⁷. Hypoxia also enhances receptor tyrosine kinase-mediated signalling⁸, tumour angiogenesis⁹, vasculogenesis¹⁰, the epithelial-to-mesenchymal transition¹¹, invasiveness¹² and metastasis¹³, as well as suppressing immune reactivity¹⁴. In addition, hypoxia contributes to loss of genomic stability through the increased generation of reactive oxygen species (ROS)¹⁵ and the downregulation of DNA repair pathways¹⁶.

In part because of these effects on tumour development, hypoxia is implicated in resistance to therapy through multiple mechanisms (shown for cytotoxic

agents in TABLE 1; see also [Supplementary information S1](#) (tables)). Reflecting these major roles in cancer biology and therapy, there is compelling evidence that hypoxia can compromise clinical outcomes in human cancer (TABLE 2). However, as noted in TABLE 1, some changes in hypoxic cells can result in increased drug sensitivity; these exceptions caution against the frequent generalization in the literature that hypoxic cells are invariably chemoresistant.

The apparent extent of hypoxia in human tumours depends on the methods used to detect it; the most widely used methods are indicated in TABLE 2. Invasive oxygen electrodes provide the most direct measure and demonstrate extreme heterogeneity of oxygenation within and between tumours in every tumour type evaluated in patients¹⁷. Increasingly, evaluation of hypoxia in the clinic is shifting to the monitoring of endogenous markers, especially the transcriptional targets of the hypoxia-inducible factors (HIFs), and exogenous 2-nitroimidazole probes, such as [pimonidazole](#), that bind covalently to SH-containing molecules (thiols) in hypoxic tissue^{18,19}. The use of these markers to image hypoxia in a human tumour is illustrated in FIG. 1a, which shows the typically more restricted distribution of bound pimonidazole than the HIF1 target carbonic anhydrase 9 (CA9). This and other evidence indicates that metabolic activation of 2-nitroimidazole probes requires more severe hypoxia than does the HIF1 response. Quantitative understanding of hypoxia in tumours (and physiological hypoxia in some normal tissues) is far from complete, but the oxygen concentration dependencies

Auckland Cancer Society
Research Centre, The
University of Auckland,
Auckland, New Zealand.
Correspondence to W.R.W.
e-mail:
wr.wilson@auckland.ac.nz
doi:10.1038/nrc3064

At a glance

- Hypoxia represents a compelling therapeutic target, given that it has a major role in tumour development and resistance to therapy, and that the levels of hypoxia are more severe in most tumours than normal tissues.
- One approach to targeting hypoxia seeks to develop bioreductive prodrugs that are activated by enzymatic reduction in hypoxic tissue. These prodrugs are chemically diverse and represent two distinct strategies: activation under moderate hypoxia (as exemplified by tirapazamine) or only under severe hypoxia (as exemplified by PR-104). In the latter case, diffusion of the active drug to less hypoxic cells is essential.
- A second approach seeks small molecule inhibitors against molecular targets involved in the survival of hypoxic cells. Current interest focuses on the inhibition of the hypoxia-inducible factor 1 (HIF1), the unfolded protein response (UPR) and mTOR pathways, but the most important vulnerabilities in hypoxic cells are not well defined. Most molecularly targeted agents have been 'repurposed' from other applications, and have low selectivity as hypoxic cytotoxins.
- Both approaches face substantial challenges in relation to off-target effects, which, ironically, also present opportunities. For bioreductive prodrugs, activation by aerobic reductases can contribute to normal tissue toxicity, but this is exploitable in tumours that highly express these enzymes. For molecularly targeted agents, hypoxia-independent signalling through the same pathways may provide opportunities for additional antitumour activity.
- Both bioreductive prodrugs and molecularly targeted agents also need to overcome the problem of drug penetration through poorly perfused hypoxic tissue; strategies for addressing this requirement are being developed.
- The current generation of bioreductive prodrugs generate DNA-reactive cytotoxins, making them difficult to combine with conventional chemotherapy because of overlapping toxicity. This challenge is stimulating the development of bioreductive prodrugs that release molecularly targeted agents as their effectors, potentially combining the best features of both approaches.
- Given the marked heterogeneity in hypoxia between tumours of the same type, the clinical exploitation of hypoxia using all of these approaches will require their co-development with companion diagnostics for hypoxia (and for other determinants of sensitivity).

for some of the critical biological processes considered in this Review are illustrated schematically in FIG. 1b. These differences in oxygen concentration thresholds have important implications for targeting hypoxic cells, as have differences in the spatial distribution and duration of hypoxia and the genetic and environmental context in which hypoxia occurs. In particular, these factors will dictate the choice of hypoxia-targeted therapy that best complements existing agents used to treat the oxic cell population in tumours.

The compelling evidence for hypoxia in tumour tissue and its therapeutic importance makes hypoxia a high priority target for cancer therapy. In this Review we describe recent progress in developing small molecule drugs to kill hypoxic cells, including bioreductive prodrugs that are activated selectively under hypoxia, and drugs that inhibit molecular targets in hypoxic cells. We focus here on agents that kill hypoxic cells directly, rather than inhibitors of hypoxia-dependent processes such as angiogenesis.

Bioreductive prodrugs

Chemical classes and mechanisms of action. The concept of activating prodrugs selectively in tumours, to achieve targeted delivery of cytotoxins, has a long history. The first clear demonstration was the reactivation of β -glucuronide metabolites of an aniline nitrogen

mustard in tumours with high β -glucuronidase activity²⁰, but such approaches have struggled with the challenge of finding tumours with high enough expression of the activating enzymes to achieve useful selectivity. Hypoxia is potentially a more generic feature, with a clear basis for tumour selectivity, although expression of the activating enzymes is also critically important in this context.

Five different chemical moieties (nitro groups, quinones, aromatic *N*-oxides, aliphatic *N*-oxides and transition metals) have the potential to be metabolized by enzymatic reduction under hypoxic conditions, and thus provide the basis for the design of bioreductive prodrugs for exploiting tumour hypoxia. The mechanisms by which bioreductive prodrugs are selective for hypoxic cells are summarized in FIG. 2A; most often these mechanisms involve the re-oxidation by oxygen of the initial free radical intermediate formed by a one-electron reduction of the prodrug, thus generating superoxide. This futile redox cycling ensures that steady-state concentrations of the prodrug radical are kept low in oxic cells, resulting in hypoxia-selective cell killing provided that the prodrug radical (or its downstream products) is more cytotoxic than superoxide or the unreduced prodrug.

Inhibition of drug reduction by oxygen through this redox cycling mechanism was first demonstrated for nitro compounds²¹ and was subsequently shown to be responsible for the hypoxia-selective cytotoxicity of nitroimidazoles²². This bioreductive mechanism is distinct from hypoxic cell radiosensitization by the same compounds²³, which is due to the ability of these compounds to replace oxygen in oxidizing ionizing radiation-induced DNA free radicals to generate cytotoxic DNA strand breaks²⁴. This first proof-of-principle demonstration of the hypoxia-selective cytotoxicity of bioreductive prodrug activity stimulated the search for ways of linking nitroreduction to the formation of more potent cytotoxins, illustrated by PR-104 and TH-302 (FIG. 2B), and for other redox moieties capable of hypoxia-selective metabolic activation.

The potential for using quinones in this context can be traced to the discovery in the 1960s that the DNA-crosslinking anticancer antibiotic *mitomycin C* is activated by reduction of its indoloquinone moiety^{25,26}. Sartorelli's group subsequently designed simpler quinone bioreductive alkylating agents²⁷, which were proposed to exploit the more reducing environment in tumours relative to normal tissues²⁸. It was later shown that the bioreductive activation of quinones occurs selectively under hypoxia²⁹ through a redox cycling mechanism³⁰ analogous to that for nitro compounds, but with two sequential one-electron reductions (first to the semiquinone and then to the hydroquinone).

Subsequently, three other chemical moieties capable of hypoxia-selective metabolic reduction by tumour cells have been discovered. Martin Brown³¹ showed that the aromatic *N*-oxide *tirapazamine* (TPZ; FIG. 2B) is 50–200-fold more toxic to hypoxic than oxic cells in culture³¹ owing to one-electron reduction to a DNA-damaging free radical (originally thought to be the TPZ radical itself, but now considered to be an

Bioreductive prodrugs

Biologically inactive molecules that are converted to an active drug by enzymatic reduction.

Superoxide

A free radical formed by a one-electron reduction of oxygen, including by electron transfer from a prodrug free radical. Despite its name, superoxide itself is not highly reactive and is generally less toxic than the reduced prodrug, so its generation represents a detoxification mechanism in aerobic cells.

Table 1 | Mechanisms of resistance (and sensitivity) of hypoxic cells to cytotoxic therapy*

Effect of hypoxia	Resistance or sensitivity?	Mechanism	Agents affected	Example
Lack of oxidation of DNA free radicals by O ₂	Resistance	Failure to induce DNA breaks	Ionizing radiation	2–3-fold increase in ionizing radiation dose required for equivalent cell kill
			Antibiotics that induce DNA breaks	Bleomycin
Cell cycle arrest in G1 or G2 phase	Resistance	Repair before progression to S or M phase	Cycle-selective chemotherapy drugs	5-Fluorouracil
Cell cycle arrest in S phase	Sensitivity	Collapse of stalled replication forks	PARP inhibitors [†]	Veliparib (ABT-888)
Distance from vasculature (indirect)	Resistance	Compromised drug exposure	Drugs extensively bound in tumour cells	Taxanes
Extracellular acidification (indirect)	Resistance	Decreased uptake	Basic drugs	Doxorubicin
	Sensitivity	Increased uptake	Acidic drugs	Chlorambucil
Resistance to apoptosis	Resistance	Genetic selection of TP53 mutants	Multiple	
		Downregulation of BID and BAX	Multiple	Etoposide
Genomic instability	Resistance	Mutagenesis	Multiple	DHFR amplification and methotrexate
Suppression of DNA repair	Resistance	Downregulation of MMR	DNA methylating agents	
	Sensitivity	Downregulation of NER	Bulky DNA monoalkylating and crosslinking agents	
		Downregulation of HR	DNA crosslinking agents	Cisplatin
HIF1 stabilization	Resistance	Expression of ABC transporters	ABC transporter substrates	MDR1 and doxorubicin
		Downregulation of NHEJ	Agents that induce DSBs	Etoposide

BAX, BCL2-associated X protein; BID, BH3 interacting domain death agonist; DHFR, dihydrofolate reductase; DSB, double strand break; HIF1, hypoxia-inducible factor 1; HR, homologous recombination; MDR1, multidrug resistance protein 1; MMR, mismatch repair; NER, nucleotide excision repair; NHEJ, non-homologous end joining; PARP, poly(ADP-ribose) polymerase. *See also [Supplementary information S1](#) (tables) for tables with references. [†]Also sensitized by downregulation of HR under hypoxia.

oxidizing hydroxyl³² or benzotriazinyl³³ radical arising spontaneously from the TPZ radical) (FIG. 2B). Later, Laurence Patterson³⁴ and ourselves³⁵ independently demonstrated that inhibition by oxygen of the bio-reduction of aliphatic *N*-oxides to the corresponding tertiary amines can also be used as a basis for hypoxia-activated prodrugs, in these examples through increasing DNA binding affinity of intercalators (illustrated for [banoxantone](#) (also known as AQ4N) in FIG. 2B). For the aliphatic *N*-oxides, hypoxic selectivity stems from inhibition of two-electron reductases by oxygen (FIG. 2A), rather than redox cycling. Examples of the fifth class (transition metals) include cobalt(III)^{36,37} and copper(II)³⁸ complexes capable of hypoxia-selective bioreductive activation through one-electron reductions of the metal centres to unstable cobalt(II) or copper(I) complexes that then dissociate to release cytotoxic ligands.

Bioreductive prodrugs under recent or ongoing clinical development (FIG. 3; TABLE 3) include examples of each of these chemotypes (except transition metal complexes, for which hypoxic cell killing has only been reported in cell culture). Other than TPZ and [apaziquone](#) (also known as E09), for which Phase III clinical trial results are pending, the compound currently most advanced in clinical testing is TH-302 (FIG. 2B).

This 2-nitroimidazole-based nitrogen mustard prodrug has shown promising activity in a Phase I study³⁹ and is being evaluated in multiple Phase I and II trials, including a randomized Phase II trial with [gemcitabine](#) in pancreatic cancer (www.ClinicalTrials.gov identifier [NCT01144455](#)). The clinical status of the other compounds is discussed below in relation to unique features of their mechanisms of action. These prodrugs illustrate diverse strategies for exploiting oxygen-sensitive biotransformations to achieve cytotoxic activation (FIG. 2B), and are representative of other prodrugs reviewed previously^{40–43}. The prodrugs also differ in their quantitative oxygen dependence (K_{O_2} , the K_i for inhibition by oxygen), the activating reductases and the nature of the resulting DNA lesions (TABLE 3). A recent addition is a chloromethylbenzindoline prodrug, SN29730, which generates a potent DNA minor groove alkylator on nitroreduction and has high hypoxic potency and selectivity *in vitro* and *in vivo*⁴⁴. A common feature of all these prodrugs is that interference with the DNA replication fork appears to be the main mechanism of cytotoxicity, as illustrated by the dependence of the hypoxic cytotoxicity of TPZ⁴⁵ — and the alcohol metabolite of PR-104, PR-104A⁴⁶ — on homologous recombination (HR) repair, which is required for the resolution of damage at the replication fork⁴⁷.

Replication fork

The branch-point structure that forms between two DNA template strands during DNA replication at which nascent DNA synthesis is ongoing.

Homologous recombination

(HR). High-fidelity repair of DNA lesions, including double-strand breaks, in S and G2 phases of the cell cycle, using a sister chromatid as a template.

Identifying and exploiting the activating reductases. Targeting hypoxia with bioreductive prodrugs depends on tumour expression of the appropriate activating reductases. Most of the one-electron reductases responsible for the redox cycling (and hence the hypoxic selectivity) of prodrugs appear to be NAD(P)H-dependent flavoproteins with low substrate affinities and specificities as xenobiotic metabolizing enzymes; their identification represents an important ongoing challenge (BOX 1).

Reductases that catalyse concerted two-electron reductions provide an alternative pathway for bioreductive prodrug activation (FIG. 2A) and represent both an opportunity and challenge for tumour targeting. These enzymes fall into two broad groups. Haemoproteins, such as cytochrome P450s (CYPs), especially CYP3A4, can catalyse the two-electron reduction of AQ4N⁴⁸. A recently identified extrahepatic CYP, CYP2S1, also reduces AQ4N⁴⁹, which is notable given that this enzyme is upregulated by HIF1 (REF. 50). The one-electron reductase inducible nitric oxide synthase (iNOS; also known as NOS2) is also upregulated under hypoxia (BOX 1), and can similarly catalyse the two-electron reduction of AQ4N through its CYP-like haem domain⁵¹. Importantly, although these haem-dependent reductions of *N*-oxides do not generate an oxygen-sensitive radical intermediate, they are nonetheless inhibited by oxygen^{49,51}, presumably through competitive binding of O₂ and the *N*-oxide to the haem prosthetic group. This process is therefore potentially exploitable for targeting hypoxia, although the *K*_{O₂} is not well defined, and whether this pathway is fully suppressed under oxic conditions is unclear.

A second group of two-electron reductases catalyse hydride (H⁻) transfer from NAD(P)H and are not inhibited by oxygen. These can bypass the oxygen-sensitive free radical intermediate during reduction of quinones, nitro compounds and aromatic *N*-oxides. The best studied enzyme of this class is NAD(P)H dehydrogenase [quinone] 1 (NQO1; also known as DT-diaphorase), which catalyses the facile two-electron reduction of quinones including apaziquone and the aziridinylbenzoquinone RH1 to their hydroquinones⁵². NQO1 also reduces the dinitrobenzamide CB 1954 (tretazicar) to its active 4-hydroxylamine metabolite⁵³. Although CB 1954 is a poor substrate for human NQO1, it is efficiently reduced by its paralogue NQO2 using dihydronicotinamide riboside (NRH) as a cofactor⁵⁴. NQO2 also catalyses aerobic reduction of RH1 (REF. 55). In addition, the NADH-dependent two-electron reductase aldo-keto reductase 1C3 (AKR1C3) has recently been shown to reduce PR-104A (but not other bioreductive prodrugs) in some human tumour cell lines under aerobic conditions⁵⁶.

Aerobic two electron reductions by these enzymes represent 'off-target' activation in the context of hypoxia and are likely to contribute to the normal tissue toxicity of some quinones and nitro compounds, as illustrated by the resistance of *Nqo1* knockout mice to mitomycin C-induced myelotoxicity⁵⁷ and the expression of NQO1 in many normal human tissues⁵⁸. However, this activation may also be therapeutically exploitable in tumours that highly express these enzymes. *NQO1*, *NQO2* (REF. 59) and *AKR1C3* (REFS 56,60) are each transcriptionally regulated, through their antioxidant response elements (AREs), by the transcription factor nuclear

Table 2 | Representative examples of the prognostic and predictive significance of hypoxia in human cancer*

Measure of hypoxia	Probe	Clinical setting	Outcome for hypoxic tumours
Oxygen concentration	Eppendorf oxygen electrode	Chemoradiation of advanced HNSCC	Worse OS
		Radiotherapy of soft tissue sarcomas before surgery	Worse DFS owing to a higher rate of distant metastasis
		Brachytherapy of localized prostate cancer	Decreased biochemical control (shown by PSA levels)
		Cervical carcinoma	Worse DFS in node-negative patients owing to a higher rate of distant metastases
Endogenous markers	HIF1 α	Node-negative breast cancer	Worse OS
	HIF1 α	BRCA1 mutant breast cancer	Worse DFS
	HIF2 α , CA9	CHART trial in HNSCC	Worse local control and OS
	CA9	Adjuvant chemotherapy of breast cancer	Worse OS
	Osteopontin	Radiotherapy for HNSCC	Nimorazole (hypoxic radiosensitizer) improved local control and OS
	Lysyl oxidase	Breast cancer	Worse metastasis-free survival
	Hypoxic gene signature	HNSCC and breast cancer	Worse outcome, multiple end points
	Hypoxic gene signature	Hepatocellular carcinoma	Worse OS
Exogenous probes	Pimonidazole	Radiotherapy for advanced HNSCC	Worse local control
	EF5	Post-surgical radiotherapy of HNSCC	Worse DFS

CA9, carbonic anhydrase 9; CHART, continuous hyperfractionated accelerated radiotherapy; DFS, disease-free survival; EF5, etanidazole pentafluoride; HIF, hypoxia-inducible factor; HNSCC, head and neck squamous cell carcinoma; OS, overall survival; PSA, prostate specific antigen. *See also [Supplementary information S1](#) (tables) for tables with references.

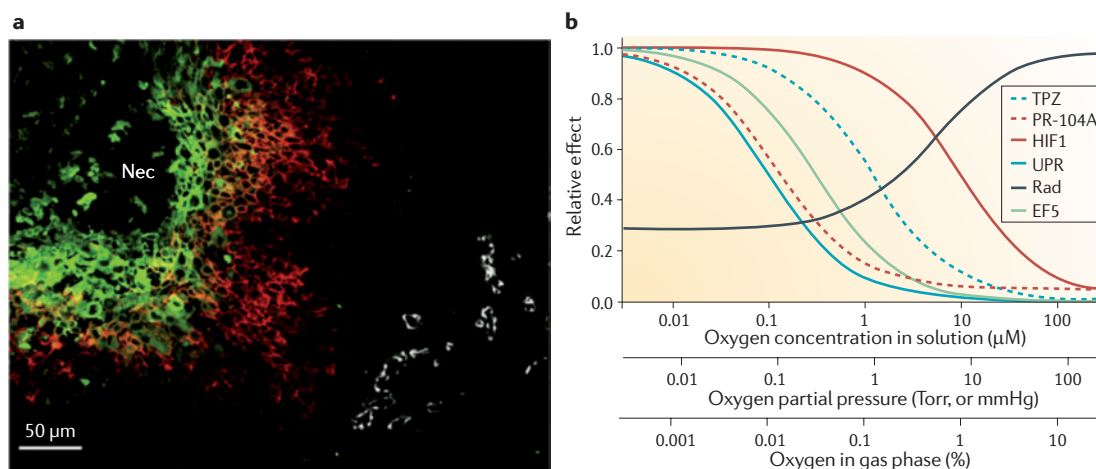


Figure 1 | Oxygen dependence of hypoxia-responsive processes in tumours. **a** | Pseudocolour immunofluorescence showing the difference in distribution of covalently bound pimonidazole (green), an exogenous 2-nitroimidazole hypoxia marker, and hypoxia-inducible factor 1 (HIF1)-regulated carbonic anhydrase 9 (CA9; red), an endogenous marker of hypoxia. This distribution is shown relative to blood vessels (white) and necrosis (Nec) in a representative region of a human squamous cell carcinoma of the larynx. **b** | Schematic representation of quantitative oxygen dependencies for ionizing radiation, bioreductive activation of prodrugs and imaging agents, and biological responses to hypoxia. Three commonly used units for oxygen concentration are shown on the x axis, assuming that the culture medium is in equilibrium with humidified gas mixtures at atmospheric pressure⁷⁷. The curves are based on representative oxygen sensitivity parameters for clonogenic cell killing by: ionizing radiation (Rad)¹⁸⁵, tirapazamine (TPZ)⁷⁸ and PR-104A⁸³. Also shown is binding of the 2-nitroimidazole etanidazole pentafluoride (EF5) to intracellular proteins¹⁸⁶. Biological responses to hypoxia are time- and cell-type-dependent; the indicative relationships shown here are based on acute stabilization of HIF1 in HT1080 cells¹⁸⁶ and evidence that the unfolded protein response (UPR) is rapidly induced only under severe hypoxia^{110,187}. Part **a** is reproduced, with permission, from REF. 150 © (2009) Elsevier Science.

factor erythroid 2-related factor 2 (NRF2; also known as NFE2L2). NRF2, in turn, is controlled by a redox-sensitive cytoplasmic repressor Kelch-like ECH-associated protein 1 (KEAP1), and independently by PRKR-like endoplasmic reticulum kinase (PERK; also known as eIF2AK3)⁶¹. Both of these signalling pathways provide the potential for indirect upregulation of NRF2-regulated reductases under hypoxia through increased ROS (especially under conditions of fluctuating hypoxia), leading to KEAP1 inactivation or activation of unfolded protein response (UPR) signalling through PERK (see below). High expression of NQO1 is the major driver for clinical development of apaziquone as an intravesicular (topical) therapy for non-invasive bladder cancer⁶², and RH1 is also being explored for treatment of tumours with high NQO1 expression⁶³. The combination of CB 1954 with the synthetic reducing cofactor caricotamide (also known as EP-0152R), an NRH analogue, has recently been explored for the treatment of NQO2-expressing hepatocellular carcinomas (HCCs). Similarly, high expression of AKR1C3 in some non-small-cell lung cancers and HCCs⁵⁶ has led to pilot clinical studies of PR-104 in these cancers, and evaluation is ongoing for acute myeloid leukaemia (AML), based on the high expression of AKR1C3 mRNA in leukaemic cells from some patients with AML⁶⁴. In each case, the additional hypoxia-selective activation by one-electron reductases is potentially beneficial, including in leukaemias and multiple myeloma, given recent evidence for hypoxia secondary to their expansion in the bone marrow^{65,66}.

TPZ is also a substrate for NQO1, but uniquely sidesteps the complications of two-electron reduction in that its mono-oxide and non-oxide reduction products (X and Y in FIG. 2A) are relatively non-toxic⁶⁷. This attractive feature of the aromatic *N*-oxides is retained in second-generation TPZ analogues such as SN30000 (REF. 68).

Bioreductive prodrug micropharmacokinetics: the extravascular transport problem. Limited extravascular penetration of drugs, an important contributor to the chemoresistance of solid tumours⁶⁹, becomes more crucial when the target cells are confined to hypoxic zones distant from functional blood vessels. The problem is particularly severe for bioreductive prodrugs, given that they are designed to be metabolized as they diffuse into hypoxic zones; if this metabolism is too facile, exposure of the most hypoxic cells will inevitably be compromised. This probably underlies the much lower hypoxic selectivity of TPZ in tumours than in low-density cell cultures⁷⁰. The first suggestion that metabolic consumption of TPZ compromises its tissue penetration came from studies showing loss of activity in hypoxic multicellular spheroids⁷¹. This was confirmed in more quantitative studies^{72,73} using another three-dimensional cell culture model, multicellular layers (MCLs), a model that is more amenable to the direct measurement of drug diffusion.

The importance of prodrug penetration in determining hypoxic cell killing in tumours is illustrated by a comparison of 15 TPZ analogues with widely different extravascular transport properties⁷⁴. In this study the

Multicellular spheroids

Spherical clusters of cells that grow large enough to become diffusion-limited, and thus model some features of the tumour microenvironment.

Multicellular layers

(MCLs). Three-dimensional cell cultures that model the extravascular compartment of tumours. Grown on collagen-coated micro-porous membranes, they allow measurement of drug diffusion and metabolism in tumour-like tissue.

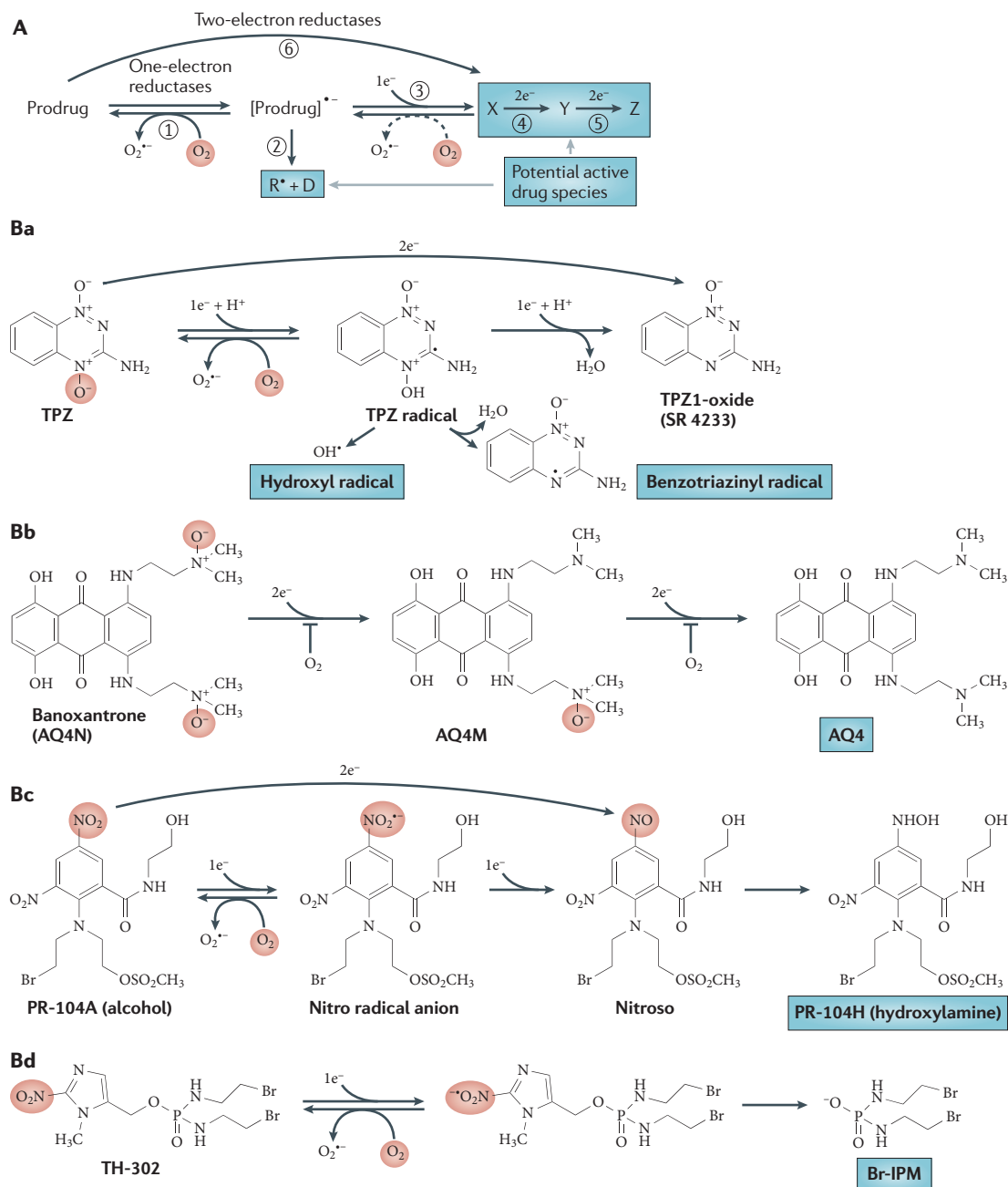


Figure 2 | Mechanisms of metabolic activation of bioreductive prodrugs. The cytotoxic metabolites are shown in blue. **A** | Generalized scheme showing competing one-electron and two-electron reductions of prodrugs. One-electron reduction generates a prodrug radical that can be re-oxidized by oxygen (reaction 1) in oxic cells, but generates active drug (blue boxes) in hypoxic cells, either by fragmentation of the prodrug radical (reaction 2) or by its further reduction, usually by disproportionation (reaction 3) and subsequent reduction of the two electron reduction product, X (reactions 4 and 5). Some prodrugs are also reduced by a concerted two-electron reduction (reaction 6), thus bypassing the oxygen-sensitive prodrug radical. Two-electron reduction is typically insensitive to oxygen, with important exceptions (see main text). **B** | Examples of well-studied prodrugs that exploit bioreduction in different ways to elicit selective killing of hypoxic cells. **Ba** | Reduction of an aromatic N-oxide to generate a DNA-reactive free radical; **Bb** | reduction of an aliphatic N-oxide to unmask a DNA intercalator; **Bc** | nitroreduction as an electronic switch to activate a reactive centre, thus generating an activated nitrogen mustard; and **Bd** | nitroreduction to initiate fragmentation to a non-radical cytotoxin, such as a nitrogen mustard.

tissue diffusion coefficient and bioreductive metabolism kinetics of each prodrug was measured using MCLs grown from HT29 human colon adenocarcinoma cells. These measurements were used to develop a spatially resolved

pharmacokinetic and pharmacodynamic model describing pharmacokinetics (concentration–time profiles) and pharmacodynamics (cell killing probability) as a function of position in a tumour microvascular network. Hypoxic

Bystander effect

In the context of bioreductive prodrugs, the killing of adjacent cells that lack prodrug-activating ability through local diffusion of the active drug.

cell killing in HT29 tumour xenografts was well predicted by the model, but only when extravascular transport was included explicitly. This study demonstrated that prodrug reduction kinetics need to be optimized to balance the competing requirements of metabolic stability (for maximal tissue penetration) and metabolism to the cytotoxic metabolite (for maximal cytotoxicity in hypoxic cells).

Until recently the penetration problem has largely been ignored during the development of bioreductive prodrugs, many of which have been found to lack activity as hypoxic cytotoxins in xenograft models despite marked hypoxic selectivity in low-density cell cultures. Some progress has been made in defining the physico-chemical properties (such as lipophilicity, molecular weight and hydrogen bond donors and acceptors) that determine diffusion coefficients using MCLs, at least for TPZ analogues⁷⁵. This has assisted the design of new analogues with higher tissue diffusion coefficients, making it possible to accommodate higher rates of bioreductive metabolism without compromising penetration⁷⁶. These features are illustrated by SN30000 (TABLE 3), which has higher activity than TPZ against hypoxic cells in multiple xenograft models⁶⁸.

Finessing bioreductive prodrug activation: K values and bystander effects. Bioreductive prodrugs can act as direct oxygen sensors through redox cycling or other mechanisms of reductase inhibition by oxygen, as outlined above. However, their quantitative oxygen dependence is crucially important for their ability to complement other anticancer agents such as ionizing radiation (FIG. 1b), and differs among prodrugs.

The elimination of hypoxic tumour cells at 'intermediate' oxygen concentrations (~1–10 μM oxygen) is arguably more important than the most severely hypoxic or anoxic cells, which are less frequent and probably less likely to contribute to tumour regrowth after therapy. Two different bioreductive prodrug strategies are being explored for targeting these moderately hypoxic cells, each with different strengths and weaknesses. One strategy is to use prodrugs with relatively high K_{O_2} to provide activation under moderate hypoxia. The only bioreductive prodrugs demonstrated to be activated under such conditions are TPZ^{77,78} and its analogues, such as SN30000 (REF. 68), which have K_{O_2} values of ~1 μM in cell culture (TABLE 3).

The other strategy is to confine prodrug activation to more severely hypoxic cells (K_{O_2} ~0.1 μM), which has the advantage of restricting activation to pathologically hypoxic regions in tumours and thus avoiding activation under physiological hypoxia in normal tissues. This also limits the metabolic loss of prodrugs during diffusion into hypoxic zones. These very low K_{O_2} values — although difficult to measure experimentally because of technical limitations in controlling and quantifying low oxygen concentrations in respiring cell cultures — seem to be typical of quinones⁷⁹, nitro compounds⁸⁰ and cobalt complexes⁸¹. These bioreductive prodrugs can be expected to spare many radioresistant and chemoresistant hypoxic cells at oxygen concentrations above the drugs' K_{O_2} . In this case it may be crucially

important that the active bioreductive metabolites can diffuse to cells at higher $p\text{O}_2$ (known as the bystander effect). Such local diffusion has been demonstrated for CB 1954 and dinitrobenzamide mustards using anoxic MCL co-cultures in which 'activator' cells overexpressing NADPH-cytochrome P450 reductase (CYPOR; also known as POR) facilitate the killing of 'target' cells that are less able to activate the prodrugs⁸². PR-104A provides an example of a bioreductive prodrug with this profile (a low K_{O_2} and efficient bystander killing)⁸³. Which of these strategies (high K_{O_2} versus low K_{O_2} plus bystander effect) is preferable may depend on tumour-specific features such as the depth and spatial distribution of hypoxia (for example, whether most moderately hypoxic cells are contiguous with more severely hypoxic cells) and on treatment-specific features such as the oxygen dependence and extravascular penetration of any other agents used in combination.

Beyond DNA-reactive cytotoxins as effectors for bioreductive prodrugs. A common feature of all bioreductive prodrugs currently in development (TABLE 3) is that their active metabolites are DNA-reactive cytotoxins that damage the replication fork. Although the DNA replication fork can be considered the most successful chemotherapy target to date⁸⁴, toxicity to proliferating normal tissues is an inescapable consequence. Existing chemotherapy and chemoradiation protocols are already titrated to maximal myelotoxicity, which limits the opportunities to add the current generation of bioreductive prodrugs to standard therapies. This makes it attractive to consider adapting bioreductive prodrug design to release a broader range of active metabolites, including non-genotoxic inhibitors of molecular targets. Early examples were 2-nitroimidazole prodrugs that, on chemical reduction, release the poly(ADP-ribose) polymerase 1 (PARP1) inhibitor 5-bromoisoquinolone⁸⁵ and the prototypical cyclo-oxygenase inhibitor aspirin⁸⁶. More recently a similar approach has been used to release the tubulin-stabilizing drug *combretastatin A4* (REF. 87) and the lysyl oxidase inhibitor β -aminopropionitrile by bioreduction of prodrugs under hypoxia⁸⁸. In addition, quaternary ammonium nitroheterocyclic bioreductive triggers⁸⁹ have been used to release non-myelotoxic, irreversible pan-ERBB inhibitors under hypoxia⁹⁰. The prototype of this new class, SN29966, provides marked activity as a monotherapy against human tumour xenografts, a result that is suggested to reflect the ability of this prodrug to exploit fluctuating hypoxia because of its long residence time in tumours⁹⁰.

Molecular targets in hypoxic cells

The identification of molecular mechanisms that mediate cellular responses to hypoxia has stimulated interest in targets that might compromise the survival of hypoxic cells if inhibited. The two main oxygen-responsive signalling pathways that mediate adaptation to hypoxia are centred on the HIF family of transcription factors^{3,91,92} and the UPR⁹³, whereas mTOR presents a less well-defined opportunity to target hypoxic cell survival (FIG. 4).

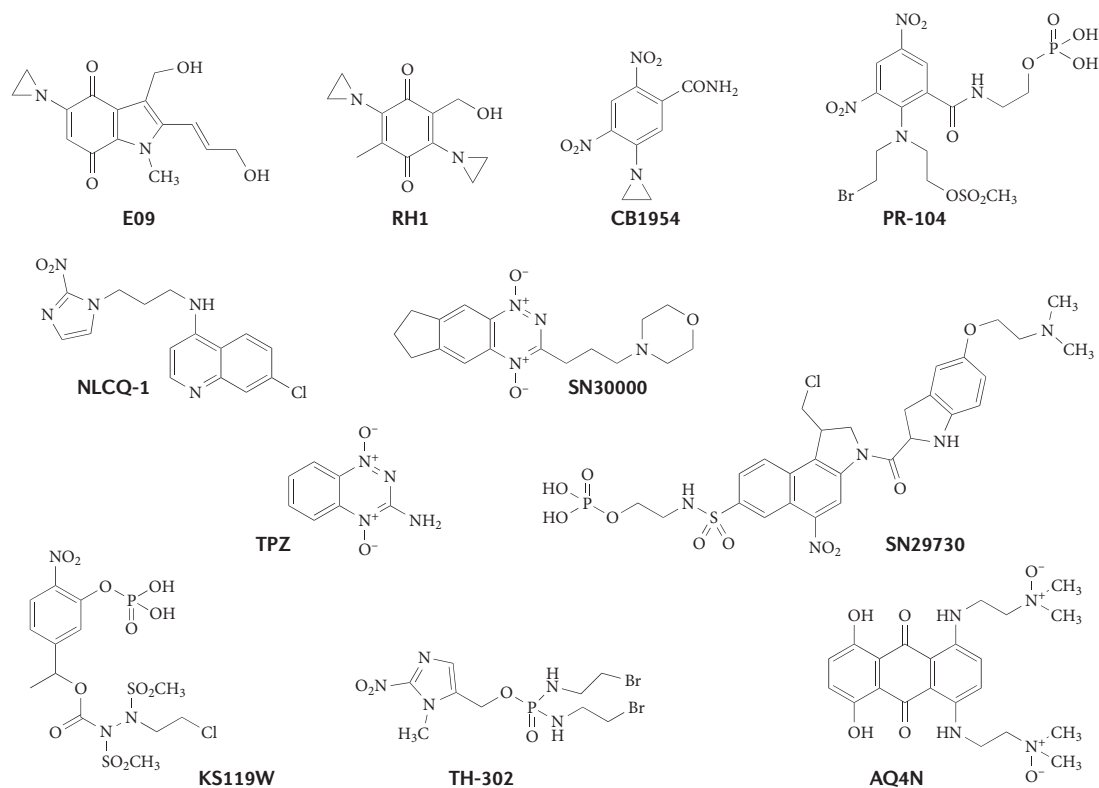


Figure 3 | **Structures of bio-reductive prodrugs.** Structures of the prodrugs presented in TABLE 3 and in the main text are shown.

HIFs. Regulation of HIF1 α and HIF2 α (also known as EPAS1) by oxygen-dependent dioxygenases such as prolyl hydroxylase domain (PHD) enzymes, the primary oxygen sensors, leads to a broad, adaptive response to hypoxia. This response includes the transcription of genes involved in angiogenesis (such as vascular endothelial growth factor A (VEGFA)), metabolic adaption (such as SLC2A1, which encodes the glucose transporter GLUT1), tolerance of acidosis (CA9), cell survival (for example, insulin-like growth factor 1 (IGF1)) and metastasis (such as lysyl oxidase (LOX))⁹². HIF1 α activity may also be influenced by many factors in addition to hypoxia⁹², hence targeting HIF1 α or its downstream products may additionally kill pseudo-hypoxic tumour cells. Nonetheless, even if not strictly specific to hypoxia, HIF1 inhibitors clearly have considerable potential to suppress resistance to therapy through multiple mechanisms, including the prevention of HIF1-dependent enhancement of endothelial cell radioresistance through cycling hypoxia⁹⁴ and blocking of the vasculogenic response to ionizing radiation-induced hypoxia¹⁰.

HIF1 α overexpression and its association with poor treatment response and outcome has been demonstrated in an extensive range of human tumours^{19,95} (TABLE 2). Multiple components of the HIF1 signalling pathway have been identified as candidate drug targets^{96,97} and a wide range of pharmacological approaches have been proposed; surveys of these have been published recently^{92,95} (TABLE 4). Several novel agents have

undergone Phase I evaluation (such as EZN-2968 (www.ClinicalTrials.gov identifier [NCT00466583](#)) and PX-478 (www.ClinicalTrials.gov identifier [NCT00522652](#))), but currently there is no clear clinical evidence of antitumour activity due to HIF1 inhibition. Other agents have been 'repurposed' from their original applications (such as the antibiotic geldanamycin⁹⁸), and have limited specificity for HIF1 α . In addition, many new agents have been discovered through phenotypic screens (inhibition of HIF1 α signalling) but their direct molecular targets and ability to selectively kill hypoxic cells are not yet well defined. A further interesting strategy for the selective killing of HIF1-expressing cells is the incorporation of a PHD-sensitive oxygen degradation domain (ODD) from HIF1 α into cytotoxic proteins, such as a procaspase 3 fusion protein containing both an ODD and a protein transduction domain⁹⁹.

The UPR. The elucidation of the role of the UPR in oxygen sensing and hypoxic cell survival has extended the potential molecular targets for drugging hypoxic cells¹⁰⁰. Oxygen is the preferred terminal electron acceptor in the redox relay required for disulphide bond formation in protein folding¹⁰¹. Severe hypoxia leads to increased levels of unfolded proteins in the endoplasmic reticulum (ER), leading to the induction of the UPR (FIG. 4). The UPR is mediated by three signalling pathways: the PERK–eukaryotic translation initiation factor 2A (eIF2A)–activating transcription factor 4 (ATF4) pathway, the inositol-requiring

Pseudo-hypoxia

The induction of molecular responses analogous to those caused by hypoxia but triggered by other conditions.

Table 3 | Bioreductive prodrugs of DNA-reactive cytotoxins recently or currently in clinical development

Prodrug	Current clinical status	Company or institution	Chemical class	Mechanism of activation*	Mechanism of cytotoxicity	One-electron reductases	Two-electron reductases	K _{O₂} (μM)
Tirapazamine (SR 4233)	Phase III, cervix (closed)	SRI International/ NCI	Aromatic N-oxide	1, 3 [R*]	Complex DNA damage	CYPOR, iNOS	NOO1 [†]	~1
Apaziquone (E09)	Phase III, bladder (closed)	Spectrum	Quinone	1, 4 [X,Y]	ICL	CYPOR	NOO1	
TH-302	Phase I/II, multiple (active)	Threshold	Nitro	1, 3 [D]	ICL	CYPOR		~10 [§]
PR-104	Phase I/II, leukaemia (active)	Proacta and University of Auckland	Nitro	1/2, 4, 5, 6 [Y,Z]	ICL	CYPOR, iNOS, MTRR, NDOR1	AKR1C3	~0.1
Banaxtrone (AQ4N)	Recent Phase I/II	Novacea	Aliphatic N-oxide	2, 5 [Y]	TOPOII	iNOS	CYP3A4, CYP2S1	
Caricotamide (EP-0152R) plus tretazicar (CB1954)	Phase II, HCC (discontinued)	BTG	Nitro	1/2, 4, 5, 6 [Y,Z]	ICL	CYPOR, iNOS	NOO1, NOO2	
RH1	Recent Phase I	CRUK	Quinone	1, 4 [X,Y]	ICL		NOO1, NOO2	
NLCQ-1	Preclinical	Evanston Hospital	Nitro	1, 4, 5	TOPOII or multiple?	CYPOR		~1 [§]
SN30000 (CEN-209)	Preclinical	Centella and University of Auckland	Aromatic N-oxide	1, 3 [R*]	Complex DNA damage	CYPOR		~1
SN29730	Preclinical	University of Auckland	Nitro	1, 4, 5, 6 [Z]	Adenine N3 alkylation	CYPOR		
KS119W	Preclinical	Yale University	Nitro	1, 4, 5, 6 [D]	Guanine O6 ICL	B5R, CYPOR		

See FIG. 3 for chemical structures. AKR1C3, aldo-keto reductase 1C3; B5R, NADH-cytochrome b5 reductase, CRUK, Cancer Research UK; CYP, cytochrome P450; CYPOR, NADPH-cytochrome P450 reductase; HCC, hepatocellular carcinoma; ICL, DNA interstrand crosslink; iNOS, inducible nitric oxide synthase; MTRR, methionine synthase reductase; NCI, US National Cancer Institute; NDOR1, NADPH-dependent diflavin oxidoreductase 1; NOO, NAD(P)H dehydrogenase [quinone]; TOPOII, topoisomerase II. *Reaction numbers refer to FIG. 2A. Active cytotoxins (X,Y etc in FIG. 2A) are shown in square brackets. [†]Detoxifying. [§]Gas phase O₂ concentration⁶⁶ (K_{O₂} values of 2-nitroimidazoles are typically much lower based on solution oxygen concentrations). See also [Supplementary information S1](#) (tables) for tables with references.

enzyme 1 (IRE1; also known as ERN1)–X-box binding protein 1 (XBP1) pathway and the ATF6 pathway. These pathways activate responses to suppress protein synthesis, stimulate protein degradation in the ER, and activate apoptosis and autophagy to resolve ER stress⁹³. An additional mechanism of activation of UPR by hypoxia is the stabilization of ATF4 through loss of its oxygen-dependent PHD3-mediated degradation¹⁰². Gene knockout and RNA interference studies have demonstrated that the PERK–eIF2A–ATF4 and IRE1–XBP1 pathways contribute to hypoxic cell survival^{102–104}.

Two drug strategies are being pursued to kill hypoxic cells selectively through UPR targets (TABLE 4). One approach seeks to inhibit the UPR by targeting PERK, ATF4 and IRE1. High-throughput screens and *in vivo* luminescence-based assays for UPR inhibitors have been reported¹⁰⁵, as have first-generation inhibitors of the endonuclease domain of IRE1 (REFS 106,107). Further drug discovery will be facilitated by the availability of crystal structures of the endonuclease domain of yeast IRE1 (REF. 108). A second approach seeks to exacerbate ER stress in order to overwhelm the UPR on the assumption that the UPR is near its capacity in hypoxic cells. Evidence that the ER stressors [thapsigargin](#) and [bortezomib](#) elicit hypoxia-selective cytotoxicity *in vitro* supports this approach¹⁰⁹.

mTOR. As a key node for the integration of the signals regulating cellular energy and nutrient status, mTOR presents a potential target for hypoxic cell killing. Under hypoxia, mTOR complex 1 (mTORC1) kinase activity is restricted through multiple mechanisms (FIG. 4), resulting in the suppression of protein synthesis to an extent that depends on the severity and duration of hypoxia¹¹⁰. The mechanisms include activation of the tuberous sclerosis 1 (TSC1)–TSC2 complex through the HIF1 target gene DNA-damage-inducible transcript 4 (*DDIT4*; also known as *REDD1*)¹¹¹ and through increased AMP-activated protein kinase (AMPK) activity under hypoxia^{110,112}. In addition, hypoxia induces the HIF1 target gene *BNIP3*, which inhibits mTORC1 through RAS homologue enriched in brain (RHEB)¹¹³. The resulting suppression of mTORC1 has multiple effects on transcription and translation, the latter in part owing to hypophosphorylation of eIF4EBP1, which leads to sequestration of eIF4E and thus inhibition of cap-dependent translation. This results in preferential cap-independent translation of a subset of mRNAs including *HIF1A* and *VEGFA*. Hypoxia has been proposed to have a dual role in tumour cell survival through modulation of mTORC1 (REF. 93). In small, early stage tumours, moderate hypoxia inhibits tumour growth through mTORC1 suppression, providing a selective pressure for abrogation of the pathway. In larger, late stage tumours, mTORC1 suppression by hypoxia may be an

Cap-dependent translation
Translation initiated by binding of the eIF4F complex to the methyl-7-G(5')pppN structure (cap) at the 5' end of the mRNA.

adaptive response in the face of energy limitations, thus favouring hypoxic cell survival. If so, the consequences of further inhibiting mTORC1 in hypoxic cells are difficult to predict. Several studies have explored the activity of mTOR inhibitors in hypoxic cells (TABLE 4). *Rapamycin* provided hypoxia-selective antiproliferative effects on HT29 cells and, when combined with low dose *irinotecan*, gave increased hypoxic cell killing *in vitro* and increased tumour control *in vivo*¹¹⁴. Treatment with WYE 125132, a potent and specific mTOR kinase inhibitor, gave substantial tumour control in a range of models and blocked HIF1 α and HIF2 α accumulation under hypoxic conditions, leading to reduced hypoxic adaptation¹¹⁵.

Targets downstream of the primary hypoxia-sensing pathways. The hypoxia-induced HIF, UPR and mTOR signalling pathways are highly interactive networks that influence many downstream gene products and processes that have potential as therapeutic targets. Here we outline some of the downstream targets under consideration for selective killing of hypoxic cells.

Recent studies have shown that the UPR activates autophagy to ameliorate hypoxic stress^{6,116}, and that inhibition of autophagy with *chloroquine* or 3-methyladenine causes selective hypoxic cell killing⁶.

Metabolic reprogramming in tumour cells, most famously demonstrated by the shift to aerobic glycolysis (known as the Warburg effect), is in part mediated by

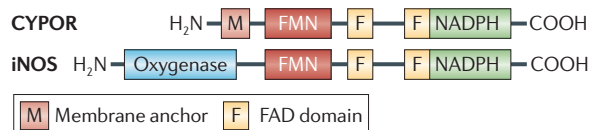
HIF1 (REF. 117) and mTOR⁷, and is therefore linked to hypoxia. This metabolic switch is also regulated by many other signalling nodes (especially by MYC, p53 and the PI3K–AKT pathway) and reflects the re-gearing of metabolism to support biosynthetic programmes and antioxidant defences to drive tumour cell growth^{7,118}. Although the shift from oxidative phosphorylation is not confined to hypoxic cells, the dependence on glycolytic ATP generation creates a vulnerability for these cells because they can no longer call on the residual mitochondrial oxidative phosphorylation, which still contributes significant ATP generation in aerobic tumour cells¹¹⁹. This reliance on glycolysis makes hypoxic tumour cells highly sensitive to suppression of glycolytic flux, hence glucose analogues that inhibit glycolysis (TABLE 4) produce striking hypoxia-selective cytotoxicity *in vitro*¹²⁰. The most widely studied compound of this class, 2-deoxy-D-glucose (2DG), is phosphorylated by hexokinases to the corresponding 6-phosphate. This phosphorylated analogue inhibits both hexokinases and phosphoglucose isomerase (GPI), which catalyses the next step in glycolysis¹¹⁹. The 2-fluoro analogue of 2DG is a more potent glycolytic inhibitor and hypoxic cytotoxin¹²¹. 2DG has been evaluated in clinical trials, but the results have not been reported; toxicity to other highly glucose-dependent tissues (such as the brain, retina and testes) represents a potential challenge in the further clinical development of this approach.

Box 1 | Identity of prodrug-activating one-electron reductases

Enzymes that catalyse one-electron transfer to prodrugs are central players in hypoxia-selective bioreduction (FIG. 2A). Their identification is an urgent priority to enable profiling of individual tumours, but has proven challenging. The best characterized enzyme is the diflavin reductase NADPH–cytochrome P450 reductase (CYPOR; also known as POR), which catalyses an intramolecular redox shuttle in which a hydride ion (H⁻) is transferred from the NADPH domain to the FAD domain, which then transfers electrons to the terminal one-electron donor flavin mononucleotide (FMN) domain (see the figure). CYPOR reduces non-mitochondrial cytochrome P450s (CYPs) and has broad substrate specificity for xenobiotics with one-electron reduction potentials that are similar to or higher than its FMN and FAD redox centres, including many bioreductive prodrugs (TABLE 3).

The nitric oxide synthases (NOSs) have diflavin (FMN and FAD) reductase domains that are homologous to CYPOR, but NOSs reduce an intramolecular haem prosthetic group in the oxygenase domain, which is responsible for nitric oxide synthesis. As for CYPOR, the transferred electron can be intercepted by small molecule electron acceptors such as tirapazamine (TPZ) and quinones^{163,164}. Interest has focused on the inducible NOS (iNOS; also known as NOS2) isoform because it is highly expressed in some tumours^{165,166} including by macrophages that accumulate in hypoxic zones¹⁶⁷. Notably, iNOS is upregulated under hypoxia through the binding of hypoxia inducible factor 1 (HIF1) to the transcription factor interferon regulatory factor 1 (IRF1)^{168,169}. This leads to localized iNOS expression in hypoxic regions of tumours¹⁷⁰, which provides an additional mechanism of hypoxic selectivity for its substrates. However, given that iNOS expression in tumours is often predominantly stromal¹⁶⁶, this enzyme will be best exploited by bioreductive prodrugs that generate cytotoxic metabolites with an efficient bystander effect. In this regard it is notable that the prodrugs AQ4N¹⁷¹, CB 1954 (REF. 172) and PR-104A¹⁷³ are activated by iNOS under hypoxia; each provides efficient bystander effects and thus has potential for exploiting hypoxic expression of iNOS in the tumour stroma. The tropism of macrophages for hypoxic regions of tumours is also being exploited for the delivery of prodrug-activating enzymes, using adenoviral transduction of CYPOR and hypoxia response element (HRE)-regulated CYP2B6 to activate cyclophosphamide¹⁷⁴. Increased hypoxic activation of TPZ has previously been demonstrated by transduction of tumour cells with HRE-driven CYPOR¹⁷⁵, suggesting the potential for further enhancing hypoxic targeting by bioreductive prodrugs by combining these approaches.

PR-104A can also be activated under hypoxia by the other members of the diflavin reductase family, NADPH-dependent diflavin oxidoreductase 1 (NDOR1) and methionine synthase reductase (MTRR)¹⁷³. Other flavoproteins capable of one-electron prodrug activation include NADH-cytochrome b5 reductases¹⁷⁶, ferredoxin reductase (FDXR)¹⁷⁷, xanthine oxidase⁵⁵ and xanthine dehydrogenase, which is also capable of two-electron reduction¹⁷⁸. However, much needs to be learned about the relative activity of these and other reductases in hypoxic regions of human tumours.



There is much interest in inhibiting other targets that can be rate-limiting for glycolysis, and which might offer greater tumour selectivity, including the HIF1-regulated facultative glucose transporter GLUT1, 6-phosphofructo-2-kinase/fructose-2,6-bisphosphatases (PFKFBs) and the tumour-specific pyruvate

kinase M2 (PKM2) isoform. Elevated GLUT1 levels has been described in a wide range of tumour types and has been demonstrated to be a negative prognostic indicator¹²². Many experimental GLUT1 inhibitors, such as phloretin, have multiple molecular targets or act indirectly, but recent examples (fasentin¹²³ and

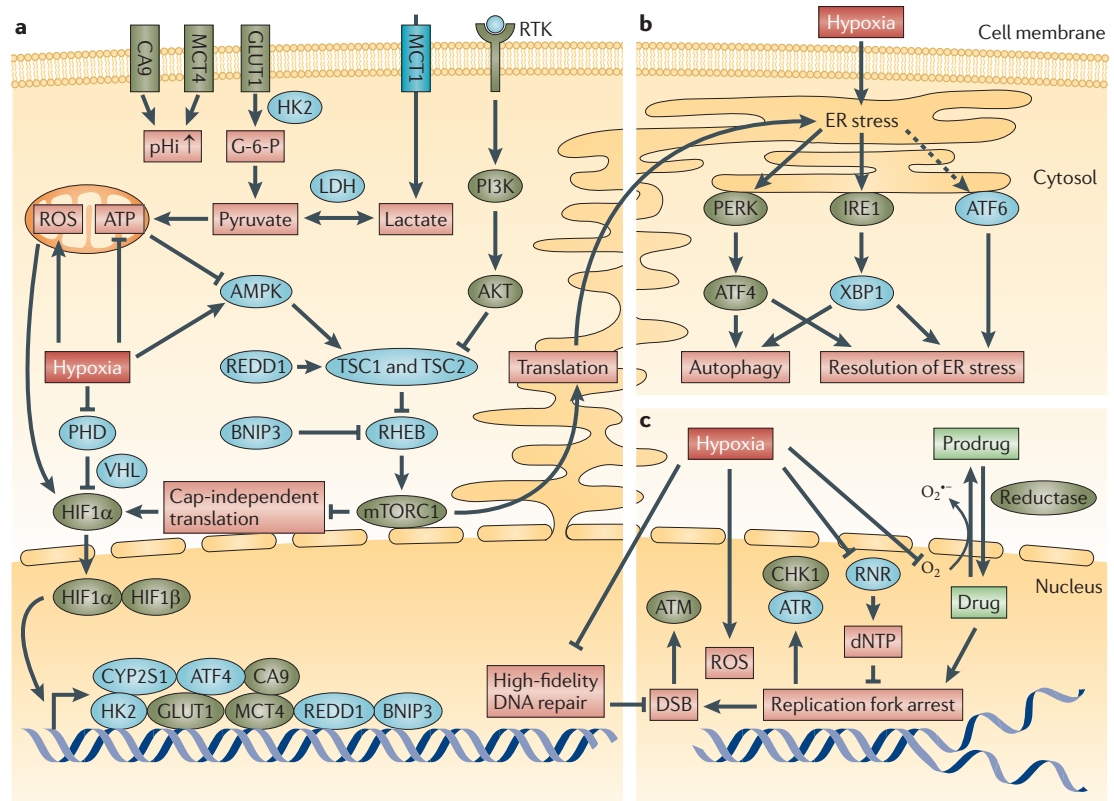


Figure 4 | Potential molecular targets for killing hypoxic cells in the oxygen-responsive signalling pathways that mediate adaptation to hypoxia. **a** | The hypoxia-inducible factor (HIF)-mTOR central metabolism module. Hypoxia inhibits prolyl hydroxylase domain (PHD)-mediated degradation of HIF1 α , which allows its dimerization with HIF1 β (also known as ARNT) and transcription of a range of genes associated with metabolic reprogramming (including hexokinase 2 (HK2) and the glucose transporter GLUT1 (encoded by *SLC2A1*)) and control of intracellular pH (pHi), such as monocarboxylate transporter 4 (MCT4) and carbonic anhydrase 9 (CA9). Also, the ability of aerobic tumour cells to use lactate in place of glucose for oxidative phosphorylation has been suggested to allow glucose to diffuse to hypoxic cells, which are highly glucose-dependent, defining the lactate transporter MCT1 as a potential target (potential target proteins are shown in green). Hypoxia induces the formation of reactive oxygen species (ROS), which stabilize HIF1 α . Hypoxia also inhibits mTOR complex 1 (mTORC1) through the HIF1-dependent transcription of DNA damage-inducible transcript 4 (*DDIT4*, which encodes REDD1) and *BNIP3* and through AMP-activated protein kinase (AMPK) signalling. This inhibition results in the hypophosphorylation of eukaryotic translation initiation factor 4E-binding protein 1 (eIF4EBP1), which favours cap-independent translation of a subset of transcripts, including *HIF1A* and provides an mTOR-HIF1 regulatory loop. Receptor tyrosine kinases (RTKs) also modulate HIF1 α translation through mTOR and other pathways in some cell lines and can also influence hypoxic survival responses. **b** | The unfolded protein response (UPR) module. Hypoxia, through the lack of oxygen to act as the ultimate electron acceptor in disulphide bond formation, impairs protein folding in the endoplasmic reticulum (ER). This leads to activation of the UPR, through PRKR-like endoplasmic reticulum kinase (PERK; also known as eIF2AK3), inositol-requiring enzyme 1 (IRE1; also known as ERN1) and potentially activating transcription factor 6 (ATF6), which supports hypoxic cell survival. **c** | DNA damage response module. Severe hypoxia inhibits ribonucleotide reductase (RNR), leading to replication fork arrest and protective ataxia telangiectasia and Rad3-related (ATR) signalling. Production of ROS in hypoxic cells, and especially on re-oxygenation, leads to DNA double-strand breaks (DSBs), which activate ataxia telangiectasia mutated (ATM) signalling. Thus, DNA damage signalling pathways provide potential targets for hypoxia-selective cell killing. Hypoxia also reduces high fidelity DNA repair (by, for example, homologous recombination (HR), which leads to sensitivity to poly(ADP-ribose) polymerase (PARP) inhibitors). In addition, hypoxia permits activation of bioreductive prodrugs, mainly by preventing redox cycling of the prodrug radical anions generated by one-electron reductases. The resulting cytotoxic drugs typically induce DNA replication fork damage, exacerbated by suppression of HR in hypoxic cells, leading to cell death. CYP2S1, cytochrome P450 2S1; G-6-P, glucose-6-phosphate; LDH, lactate dehydrogenase; RHEB, RAS homologue enriched in brain; TSC, tuberous sclerosis; VHL, von Hippel-Lindau tumour suppressor; XBP1, X-box binding protein 1.

Table 4 | Representative examples of pharmacological approaches to molecular targets in hypoxic cells*

Pathway	Target	Agent	Class
HIF1 α expression	HIF antisense mRNA	EZN-2968	RNA oligonucleotide
	Topoisomerase I	Topotecan	Camptothecin analogues
	Multiple	PX-478	Melphalan N-oxide
	Translation	Digoxin	Cardiac glycoside
	HSP90	Geldanamycin and tanespimycin (17-AAG)	Benzoquinone ansamycin antibiotics
HIF1 transcription	HIF-p300 binding	Chetomin and analogues	Dithiodiketopiperazine
	Thioredoxin 1	PX12	Imidazole disulphide
		PMX290	Indoloquinol
HIF1 target gene products	DNA binding	Echinomycin	DNA intercalator
	CA9 and CA12	Aryl sulphonamides	Sulphonamide zinc binders
	GLUT1	Glufosfamide	Glucose isophosphoramidate mustard
		2-GLU-SNAP	Glucose SNAP conjugate
		Fasentin	Oxobutanilide
		STF-31154	Unknown
HK2	5TDG, 2DG, 2FDG	Glycolysis inhibitors	
MCT1	α -cyano-4-hydroxycinnamate	Lactate transport inhibitor	
Receptor tyrosine kinases	VEGFR	Bevacizumab	Monoclonal antibody
	EGFR	Gefitinib and erlotinib	ATP competitive kinase inhibitors
		Cetuximab	Monoclonal antibody
RAS-MAPK signalling	BRAF	Sorafenib	ATP competitive kinase inhibitor
mTOR	mTORC1	Rapamycin and everolimus	Allosteric binders of FKBP12-rapamycin binding domain
		WYE-125132	ATP-competitive mTOR kinase inhibitor
	Autophagy	Chloroquine	Lysosomal pH
UPR	HSP90	Geldanamycin and 17-AAG	Benzoquinone ansamycin antibiotic
	IRE1	Salicaldehydes	IRE1 inhibitor
	26S proteasome	Bortezomib	Boronic acid tripeptide
		Nelfinavir and ritonavir	HIV protease inhibitors
SERCA	2,5-Dimethyl celecoxib	Celecoxib analogue	

CA, carbonic anhydrase; DG, deoxy-D-glucose; EGFR, epidermal growth factor receptor; FDG, fluorodeoxyglucose; FKBP12, FK506 binding protein 12; GLUT1, glucose transporter 1; HIF, hypoxia-inducible factor; HK2, hexokinase 2; HSP90, heat shock protein 90; IRE1, inositol-requiring enzyme 1 (also known as ERN1); MCT1, monocarboxylate transporter 1; mTORC1, mTOR complex 1; SERCA, sarco/endoplasmic reticulum Ca²⁺-ATPase; SNAP, S-nitroso-acetyl-penicillamine; UPR, unfolded protein response; VEGFR, vascular endothelial growth factor receptor. *See also [Supplementary information S1](#) (tables) for tables with references.

STF-31154 (REF. 124)) target GLUT1 directly. The shift to glycolysis is accompanied by increased generation of pyruvate and its conversion to lactate by lactate dehydrogenase A (LDHA). The lactate transporter monocarboxylate transporter 1 (MCT1) has been suggested as a target for killing hypoxic cells by glucose starvation, through a novel mechanism of metabolic symbiosis¹²⁵. This study showed that aerobic tumour cells expressing MCT1 can use lactate as a preferred substrate for respiration, and further demonstrated that inhibition of MCT1 by α -cyano-4-hydroxycinnamate increases glucose consumption *in vitro* and tumour radiosensitivity¹²⁵. The proposed model is that the stimulation of glucose consumption in aerobic tumour cells compromises glucose penetration into hypoxic regions, leading to the selective death of hypoxic cells in tumours. However, laboratory tools such as α -cyano-4-hydroxycinnamate are not particularly selective for the MCTs¹²⁶ and one class of selective

MCT1 inhibitors has been identified as an immunomodulator¹²⁷, raising concerns about the selectivity of such an approach for targeting hypoxic cells.

One of the consequences of the glycolytic shift, driven in part by hypoxia, is that increased generation of metabolic acids further compromises hypoxic cell survival. Disruption of pH homeostasis by targeting MCTs (such as MCT1 and MCT4) and carbonic anhydrases in hypoxic tumour cells has been proposed as a tumour-selective approach¹²⁸. MCT4 is upregulated in a HIF1 α -dependent manner¹²⁹ and increased expression of MCT4 in tumour cells has been demonstrated¹³⁰. MCT4 export of lactate and H⁺ prevents intracellular acidification and assists in the remodelling of the extracellular milieu, but specific inhibitors of MCT4 have yet to be reported.

Carbonic anhydrases are metalloenzymes that catalyse the reversible hydration of carbon dioxide to carbonic acid. The expression of CA9 and CA12 is

controlled by HIF1 (REF. 131) and CA9 is also regulated through the UPR by ATF4 (REF. 132). Despite generating H^+ and HCO_3^- with equivalent stoichiometry at the extracellular catalytic domain of these transmembrane proteins, linked bicarbonate transporters raise the intracellular pH to protect hypoxic cells¹²⁸. Silencing both CA9 and CA12 resulted in marked inhibition of the growth of LS174 human colon carcinoma cell xenograft tumours¹³¹. Extensive drug development efforts have identified a range of compounds with varying selectivity for CA9 and CA12; several compounds inhibited tumour growth and metastasis selectively in CA9-positive tumour models¹³³.

Molecular targets in DNA damage response and repair pathways. Inhibitors of DNA damage signalling and DNA repair have the potential to exploit changes in these pathways in hypoxic cells^{134–136}. Three approaches have recently been considered. The first is to exploit activation of the DNA damage response in hypoxic cells. Severe hypoxia rapidly induces replication arrest through a HIF1- and p53-independent mechanism¹³⁷. Recent evidence indicates this is due to depletion of dCTP, dGTP and dATP pools¹³⁸, reflecting the requirement of class 1a (eukaryotic) ribonucleotide reductases for molecular oxygen¹³⁹. Single-stranded DNA at stalled replication forks then induces ataxia telangiectasia and Rad3-related (ATR)–CHK1 signalling, which is required to maintain replication fork integrity. Consistent with this, knockdown of CHK1 is selectively toxic to hypoxic cells¹⁴⁰. This ATR-mediated replication arrest is reversible if cells are re-oxygenated within a few hours, but re-oxygenation then induces ROS-mediated DNA damage, including double-strand breaks that activate the kinase ataxia-telangiectasia mutated (ATM)¹⁴¹, potentially providing sensitivity to inhibitors of ATM signalling.

A second strategy is to exploit defects in DNA repair in hypoxic cells. ATR- and ATM-mediated signalling in hypoxic cells can help to facilitate DNA repair. For example, hypoxia stimulates CHK2-mediated Ser988 phosphorylation of BRCA1¹⁴², which stimulates its activity in HR. However, hypoxia also downregulates expression of key HR proteins such as RAD51 and BRCA1 through HIF1-independent repression of transcription and translation¹³⁶. In addition, hypoxia suppresses RAD51 expression in breast cancer initiating cells through HIF1-dependent upregulation of the Polycomb protein enhancer of zeste homologue 2 (EZH2)¹⁴³; *RAD51* mRNA has also recently been shown to be downregulated in hypoxic regions of 9L gliomas by laser-capture microdissection of etanidazole pentafluoride (EF5)-stained tissue¹⁴⁴. Hypoxia-mediated suppression of HR in chronically hypoxic cells^{145,146} confers an increased sensitivity to DNA-damaging cytotoxins¹⁴⁶, which may make a significant contribution to the activity of bio-reductive prodrugs that deliver such cytotoxins to hypoxic cells. Notably, hypoxia-induced downregulation of HR creates the same phenotype that sensitizes BRCA1 or BRCA2 homozygous mutant cells to PARP1 inhibition. Recently a synthetic

lethal interaction has been demonstrated for hypoxia and genetic deletion or chemical inhibition of PARP1, analogous to that for BRCA1 or BRCA2 mutations, and the PARP1 inhibitor *veliparib* (also known as ABT-888) has been shown to selectively reduce the proportion of radioresistant (that is, hypoxic) cells in RKO colon carcinoma xenografts¹⁴⁷. The authors point to the potential for synthetic lethal interactions between hypoxia and inhibitors of other repair pathways downregulated by hypoxia.

A third strategy is to pharmacologically reactivate p53 to restore hypoxia-mediated apoptosis¹³⁵. Small molecules that are in development for p53 reactivation include *APR-246* (also known as PRIMA-1), which restores transcriptional activity of mutant p53, and Nutlin-3 and RITA, which interfere with MDM2-mediated p53 degradation¹⁴⁸. RITA also induced a DNA damage response that appears to contribute to its stimulation of p53-dependent apoptosis, but cell killing was similar in hypoxic and aerobic cells¹⁴⁹.

Hypoxia and personalized cancer medicine

As in other aspects of cancer medicine, emerging technologies for profiling individual tumours have the potential to revolutionize the development of hypoxia-targeted agents. Indeed, the heterogeneity in tumour hypoxia at the broader human population level, even within a single disease subtype, means that successful development of hypoxia-targeted agents is probably a forlorn hope unless hypoxic tumours can be identified prospectively. Studies with advanced head and neck squamous cell carcinomas (HNSCCs), in which hypoxia has been demonstrated to be a negative prognostic factor using every type of diagnostic tool available (TABLE 2), are instructive in this regard. A large, relatively homogenous (stage T2–T4 laryngeal) series of HNSCC samples showed evidence of hypoxia by both pimonidazole and CA9 immunostaining in the majority of tumours, but with extreme variability¹⁵⁰. The need to quantify (not just to detect) hypoxia is illustrated by a meta-analysis of oxygen-electrode studies, which suggested that hypoxia compromised overall survival in patients with advanced HNSCC undergoing chemoradiation treatment but only in the subset of patients with the most extensive hypoxia¹⁵¹. This situation is different from the subcutaneous xenograft models widely used in preclinical studies, in which essentially all tumours display extensive hypoxia; these models thus tend to over-represent the target (and will over-predict activity) relative to autochthonous tumours in humans.

Thus there is currently much interest in the further development of hypoxia diagnostics as predictive biomarkers^{18,19,152,153}. Although studies using invasive methods (TABLE 2) have been important in establishing the significance of tumour hypoxia at the population level, broader clinical application for stratifying patients will require less-invasive tools such as positron emission tomography (PET) imaging (BOX 2). There is also great potential for minimally invasive serum-based diagnostics and global gene expression signatures for the identification of hypoxia (TABLE 2).

Synthetic lethal interaction

In genetics, an interaction between two non-lethal mutations that, in combination, confer lethality. In chemical genetics, this term can refer to interaction between a drug and mutation that confers greater drug-sensitivity than with the wild type.

Autochthonous tumours

Tumours that arise in the host being studied, as distinct from tumours introduced by transplantation.

The presence of hypoxia is a necessary but not sufficient condition for hypoxia-targeting, given that there are other crucially important determinants of sensitivity to such agents. For bioreductive prodrugs, the molecular targets are in effect the specific reductases in hypoxic cells for which these compounds are substrates. Although identification of these enzymes is incomplete (BOX 1), their activity clearly varies widely between tumours. The need for reductase profiling to identify tumours potentially responsive to bioreductive prodrugs has long been recognized¹⁵⁴, but only now are the tools becoming available to address this requirement. In addition, there is a further set of molecular targets, for the active drug metabolites, which brings into play many potential mechanisms of drug resistance. Given that most bioreductive prodrugs generate DNA damage that is repaired by HR, the validation of biomarkers for this repair pathway (currently driven by

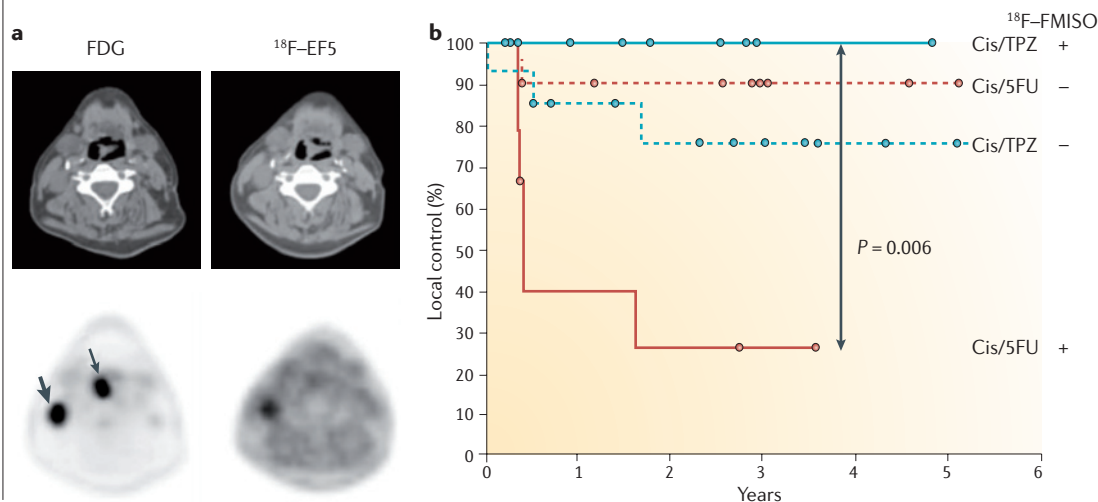
predicting the sensitivity to PARP inhibitors and cytotoxic chemotherapy^{155–157}) has strong potential to affect their development.

Clearly, the diagnostic tools for selecting patients for treatment with hypoxia-targeted drugs need to be matched to the specific therapeutic agent. Thus, one would expect the preferred diagnostic for a bioreductive prodrug to be an exogenous probe that is activated through bioreductive metabolism (by similar enzymes and with similar oxygen-dependence to the therapeutic agent). As an example, binding of the 2-nitroimidazole probe EF5 reports activity of the one-electron reductases that activate SN30000, as well as reporting hypoxia, making it a potential dual probe for both of these stratification biomarkers¹⁵⁸. By contrast, endogenous markers of hypoxia-responsive signalling pathways will be more appropriate for agents that target such pathways. It is noteworthy that there tends to be poor correlation between different hypoxia markers in both

Box 2 | PET imaging for tumour hypoxia

The variability in levels of hypoxia among individual tumours, even within a single disease subtype, calls for tools that can be used to quantify tumour hypoxia in a clinical setting. Positron emission tomography (PET) methods are undergoing active development in this context¹⁵². One strategy depends on radiolabelled antibodies against carbonic anhydrase 9 (CA9)^{179,180}, which would be of value for the selection of patients for treatment with CA9-targeted therapeutics¹³³. To the extent that CA9 can be considered a specific hypoxia-inducible factor 1 (HIF1) reporter^{132,181}, and that HIF1 activity is regulated by hypoxia⁹², this approach also has potential for monitoring hypoxia.

The most widely studied PET strategy depends on entrapment of 2-nitroimidazole probes — such as fluoromisonidazole (FMISO), fluoroazomycinara binofuranoside (FAZA) and etanidazole pentafluoride (EF5) — in hypoxic cells as a result of their bioreductive metabolism¹⁵². The mechanism is analogous to that for one-electron (oxygen-inhibited) metabolic activation of bioreductive prodrugs, subsequently generating nitroso and hydroxylamine metabolites (X and Y in FIG. 2A), which react covalently with intracellular thiols. The resulting protein adducts can be detected by immunohistochemistry (FIG. 1a), which requires a tumour biopsy, but ¹⁸F-labelled versions of the same compounds have been adapted for non-invasive PET imaging. The PET-computerized tomography (CT) scan shown in part a of the figure demonstrates a difference in ¹⁸F-EF5 entrapment in two lesions in the same patient that both rapidly metabolize ¹⁸F-fluorodeoxyglucose (FDG), suggesting that the lesion marked with the wide arrow is more hypoxic than that marked with the thin arrow. The related 2-nitroimidazole probe ¹⁸F-FMISO has been used to evaluate hypoxia in a small subset of patients in clinical trials of the bioreductive prodrug tirapazamine (TPZ) combined with cisplatin (cis) and radiotherapy, versus 5-fluorouracil (5FU) combined with cisplatin and radiotherapy for advanced head and neck squamous cell carcinoma (HNSCC). As shown in part b of the figure, a retrospective analysis demonstrated a marked advantage of the TPZ-containing regimen compared to the 5FU-containing regimen in patients with hypoxic tumours (solid lines, ¹⁸F-FMISO-negative)¹⁸³. This notable result points the way for future trials of hypoxia-targeted agents, but, regrettably, stratification for hypoxia was not used in subsequent unsuccessful Phase III trials of TPZ in this same setting¹⁸⁴. Part a of the figure is reproduced, with permission, from REF. 182 © (2008) Society of Nuclear Medicine, Inc. Part b of the figure is modified, with permission, from REF. 183 © (2006) The American Society of Clinical Oncology.



Network medicine

Analysis of biological networks to derive understanding of disease and therapy.

preclinical and clinical studies^{150,159}. Ultimately, paired diagnostics and therapeutics will need to be validated in prospective clinical trials, despite the logistical and regulatory challenges that this presents.

Conclusions and perspective

This Review has considered the two main approaches to the selective killing of hypoxic cells in tumours, with different strengths and weaknesses. Bioreductive prodrugs achieve striking selectivity between aerobic and severely hypoxic cells in culture, typically with potency differentials in the order 10–1,000-fold. By contrast, inhibition of molecular targets in hypoxic cells typically gives much more modest cytotoxicity differentials. However, these targeted inhibitors offer a more benign toxicity profile, which is distinctly different from that of cytotoxic therapy, and therefore have greater opportunity for combination with current standards of care. Compatibility with existing therapy is fundamentally important for the clinical translation of these targeted drugs, given that hypoxic cells represent only a minority subpopulation in most tumours (although a critically important one). Therefore, monotherapy activity is not a realistic expectation for hypoxia-selective agents that are strictly on-mechanism unless exceptional requirements can be met, such as a very long residence time in tumours (to exploit fluctuating hypoxia) or efficient, long-range bystander killing. Bioreductive prodrugs that generate molecularly targeted drugs as effectors, rather than DNA-damaging cytotoxins, arguably offer an opportunity to combine the best features of both classes of drug (high hypoxic selectivity and more benign toxicity), but are at an early stage of development.

Although much has already been learned about the molecular responses to hypoxia, the identification of the most useful molecular targets in hypoxic cells is far from complete. While new targets with roles in hypoxic cell survival continue to be identified, the highly interactive nature of the PHD–HIF, mTOR, UPR–autophagy and DNA damage response modules (FIG. 4) makes it difficult to identify the vulnerabilities of hypoxic cells that can best be exploited as drug targets. The results of unbiased whole-genome screens, analogous to the RNA interference screens used to identify synthetic lethal interactions with chemotherapy¹⁶⁰, are eagerly awaited. Ideally, these screens will compare multiple cancer cell lines with normal cells, under hypoxia, to reveal targets that provide selectivity for hypoxia in the context of cancer genomes, and will be interpreted in a network medicine framework¹⁶¹.

A better definition of the preferred molecular targets will make it feasible to design small molecules of greater specificity, and to move beyond the repurposing of drugs that have been developed for other applications, an approach that currently characterizes this field (TABLE 4). In a similar fashion, improved understanding of the human reductases that activate prodrugs will provide opportunities for structure-based design to improve specificity for enzymes that confer tumour selectivity.

Many of the challenges in targeting hypoxic cells are similar for both bioreductive prodrugs and molecularly targeted inhibitors; both need to be designed to address the stringent micropharmacokinetic requirements for efficient penetration to cells distant from blood vessels. This critical issue is still rarely addressed explicitly. Both classes of drugs also need to address, and where possible exploit, off-target effects (such as the aerobic reduction of bioreductive drugs, and the inhibition of hypoxia-independent HIF1 responses to ionizing radiation¹⁶²). An associated challenge is the potential toxicity resulting from physiological hypoxia in normal tissues; there is still little understanding of the contribution of such hypoxia to the dose-limited toxicities of bioreductive prodrugs. In addition, clinical development of all hypoxia-targeted agents suffers from a lack of information about the clinical settings in which hypoxic cells contribute to treatment failure. The notable exception is in chemoradiation treatment of HNSCC, for which there is overwhelming evidence from multiple hypoxic biomarkers that hypoxia compromises outcome (TABLE 3). An additional challenge is the lack of a drug-development culture in the field of radiation oncology, which is the setting in which the impact of hypoxia is most clearly understood.

Perhaps the most crucial requirement for hypoxia-targeting strategies is the development of improved predictive tools for patient stratification. These tools need to evaluate not only hypoxia, but also many other determinants of sensitivity, as discussed above. Ultimately, tumour and host genomic analyses will revolutionize the matching of hypoxia-targeted therapeutics to individual patients. However, extracting information on physiological features such as the severity of hypoxia from genomic data will be challenging, so functional assays such as PET imaging are likely to play a major part in the foreseeable future. Together, this individualized phenotyping has the potential to identify clinical niches for the diverse types of cytotoxins that are already identified as hypoxia-selective, and provide a rational basis for their clinical development.

- Jain, R. K. Normalization of tumor vasculature: an emerging concept in antiangiogenic therapy. *Science* **307**, 58–62 (2005).
- Pries, A. R. *et al.* Structural adaptation and heterogeneity of normal and tumor microvascular networks. *PLoS Comput. Biol.* **5**, e1000394 (2009).
- Dewhirst, M. W., Cao, Y. & Moeller, B. Cycling hypoxia and free radicals regulate angiogenesis and radiotherapy response. *Nature Rev. Cancer* **8**, 425–437 (2008).
- Graeber, T. G. *et al.* Hypoxia-mediated selection of cells with diminished apoptotic potential in solid tumours. *Nature* **379**, 88–91 (1996).
- Erler, J. T. *et al.* Hypoxia-mediated down-regulation of Bid and Bax in tumors occurs via hypoxia-inducible factor 1-dependent and -independent mechanisms and contributes to drug resistance. *Mol. Cell. Biol.* **24**, 2875–2889 (2004).
- Rouschop, K. M. *et al.* The unfolded protein response protects human tumor cells during hypoxia through regulation of the autophagy genes *MAP1LC3B* and *ATG5*. *J. Clin. Invest.* **120**, 127–141 (2010). **This study demonstrates a mechanism by which the UPR enhances the survival of tumour cells under severe hypoxia and that inhibition of the UPR by a small molecule (chloroquine) selectively kills hypoxic cells.**
- Cairns, R. A., Harris, I. S. & Mak, T. W. Regulation of cancer cell metabolism. *Nature Rev. Cancer* **11**, 85–95 (2011).
- Wang, Y. & Ohh, M. Oxygen-mediated endocytosis in cancer. *J. Cell. Mol. Med.* **14**, 496–503 (2010).
- Semenza, G. L. Hypoxia, clonal selection, and the role of HIF-1 in tumor progression. *Crit. Rev. Biochem. Mol. Biol.* **35**, 71–103 (2000).
- Kioi, M. *et al.* Inhibition of vasculogenesis, but not angiogenesis, prevents the recurrence of glioblastoma after irradiation in mice. *J. Clin. Invest.* **120**, 694–705 (2010).
- Hill, R. P., Marie-Egyptienne, D. T. & Hedley, D. W. Cancer stem cells, hypoxia and metastasis. *Semin. Radiat. Oncol.* **19**, 106–111 (2009).
- Pennacchietti, S. *et al.* Hypoxia promotes invasive growth by transcriptional activation of the met protooncogene. *Cancer Cell.* **3**, 347–361 (2003).

13. Chang, Q., Jurisica, I., Do, T. & Hedley, D. W. Hypoxia predicts aggressive growth and spontaneous metastasis formation from orthotopically grown primary xenografts of human pancreatic cancer. *Cancer Res.* **78**, 3110–3120 (2011).
14. Yotnda, P., Wu, D. & Swanson, A. M. Hypoxic tumours and their effect on immune cells and cancer therapy. *Methods Mol. Biol.* **651**, 1–29 (2010).
15. Guzy, R. D. *et al.* Mitochondrial complex III is required for hypoxia-induced ROS production and cellular oxygen sensing. *Cell. Metab.* **1**, 401–408 (2005).
16. Bristow, R. G. & Hill, R. P. Hypoxia, DNA repair and genetic instability. *Nature Rev. Cancer* **8**, 180–192 (2008).
17. Vaupel, P., Hockel, M. & Mayer, A. Detection and characterization of tumor hypoxia using pO₂ histography. *Antioxid. Redox Signal.* **9**, 1221–1235 (2007).
18. Tatum, J. L. *et al.* Hypoxia: importance in tumor biology, noninvasive measurement by imaging, and value of its measurement in the management of cancer therapy. *Int. J. Radiat. Biol.* **82**, 699–757 (2006).
19. Jubb, A. M., Buffa, F. M. & Harris, A. L. Assessment of tumour hypoxia for prediction of response to therapy and cancer prognosis. *J. Cell. Mol. Med.* **14**, 18–29 (2010).
20. Connors, T. A. & Whisson, M. E. Cure of mice bearing advanced plasma cell tumours with aniline mustard: the relationship between glucuronidase activity and tumour sensitivity. *Nature* **210**, 866–867 (1966).
21. Mason, R. P. & Holtzman, J. L. The role of catalytic superoxide formation in the O₂ inhibition of nitroreductase. *Biochem. Biophys. Res. Commun.* **67**, 1267–1274 (1975).
- The discovery of the mechanism by which oxygen inhibits the reduction of substrates by one-electron reductases.**
22. Mohindra, J. K. & Rauth, A. M. Increased cell killing by metronidazole and nitrofurazone of hypoxic compared to aerobic mammalian cells. *Cancer Res.* **36**, 930–936 (1976).
23. Adams, G. E., Dische, S., Fowler, J. F. & Thomlinson, R. H. Hypoxic cell sensitizers in radiotherapy. *Lancet* **1**, 186–188 (1976).
24. Wardman, P. Chemical radiosensitizers for use in radiotherapy. *Clin. Oncol.* **19**, 397–417 (2007).
25. Schwartz, H. S., Sodergren, J. E. & Phillips, F. S. Mitomycin C: chemical and biological studies on alkylation. *Science* **142**, 1181–1183 (1963).
26. Iyer, V. N. & Szybalski, W. Mitomycins and porfirromycins: chemical mechanism of activation and cross-linking of DNA. *Science* **145**, 55–58 (1964).
27. Lin, A. J., Cosby, L. A., Shanky, C. W. & Sartorelli, A. C. Potential bioreductive alkylating agents. I. Benzoquinone derivatives. *J. Med. Chem.* **15**, 1247–1252 (1972).
28. Carter, D. B. & Phillips, A. F. Measurement of electrode potentials in living and dead tissues. *Nature* **174**, 121–123 (1954).
29. Kennedy, K. A., Rockwell, S. & Sartorelli, A. C. Preferential activation of mitomycin C to cytotoxic metabolites by hypoxic tumor cells. *Cancer Res.* **40**, 2356–2360 (1980).
30. Bachur, N. R., Gordon, S. L. & Gee, M. V. A general mechanism for microsomal activation of quinone anticancer agents to free radicals. *Cancer Res.* **38**, 1745–1750 (1978).
31. Brown, J. M. SR 4233 (tirapazamine): a new anticancer drug exploiting hypoxia in solid tumours. *Br. J. Cancer* **67**, 1163–1170 (1993).
32. Chowdhury, G., Junnotula, V., Daniels, J. S., Greenberg, M. M. & Gates, K. S. DNA strand damage product analysis provides evidence that the tumor cell-specific cytotoxin tirapazamine produces hydroxyl radical and acts as a surrogate for O₂. *J. Am. Chem. Soc.* **129**, 12870–12877 (2007).
33. Shinde, S. S., Hay, M. P., Patterson, A. V., Denny, W. A. & Anderson, R. F. Spin trapping of radicals other than the *OH radical upon reduction of the anticancer agent tirapazamine by cytochrome P450 reductase. *J. Am. Chem. Soc.* **131**, 14220–14221 (2009).
34. Patterson, L. H. Rationale for the use of aliphatic N-oxides of cytotoxic anthraquinones as prodrug DNA binding agents: a new class of bioreductive agent. *Cancer Metastasis Rev.* **12**, 119–134 (1993).
35. Wilson, W. R., van Zijl, P. & Denny, W. A. Bis-bioreductive agents as hypoxia-selective cytotoxins: nitracrine N-oxide. *Int. J. Radiat. Oncol. Biol. Phys.* **22**, 693–696 (1992).
36. Ware, D. C., Palmer, B. D., Wilson, W. R. & Denny, W. A. Hypoxia-selective antitumor agents. 7. Metal complexes of aliphatic mustards as a new class of hypoxia-selective cytotoxins. Synthesis and evaluation of cobalt(III) complexes of bidentate mustards. *J. Med. Chem.* **36**, 1839–1846 (1993).
- The first use of transitional metal complexes as the basis for the design of hypoxia-activated prodrugs.**
37. Ahn, G. O. *et al.* Radiolytic and cellular reduction of a novel hypoxia-activated cobalt(III) prodrug of a chloromethylbenzindoline DNA minor groove alkylator. *Biochem. Pharmacol.* **71**, 1683–1694 (2006).
38. Parker, L. L. *et al.* A novel design strategy for stable metal complexes of nitrogen mustards as bioreductive prodrugs. *J. Med. Chem.* **47**, 5683–5689 (2004).
39. Weiss, G. J. *et al.* Phase I study of the safety, tolerability and pharmacokinetics of TH-302, a hypoxia-activated prodrug, in patients with advanced solid malignancies. *Clin. Cancer Res.* **17**, 2997–3004 (2011).
- The first clinical data for this novel hypoxia-targeted prodrug, showing evidence of tumour responses when used as a monotherapy.**
40. Wardman, P. Electron transfer and oxidative stress as key factors in the design of drugs selectively active in hypoxia. *Curr. Med. Chem.* **8**, 739–761 (2001).
41. Stratford, I. J., Williams, K. J., Cowen, R. L. & Jaffar, M. Combining bioreductive drugs and radiation for the treatment of solid tumors. *Semin. Radiat. Oncol.* **13**, 42–52 (2003).
42. Ahn, G. O. & Brown, M. Targeting tumors with hypoxia-activated cytotoxins. *Front. Biosci.* **12**, 3483–3501 (2007).
43. Chen, Y. & Hu, L. Design of anticancer prodrugs for reductive activation. *Med. Res. Rev.* **29**, 29–64 (2009).
44. Tercel, M. *et al.* Selective treatment of hypoxic tumor cells *in vivo*: phosphate pre-prodrugs of nitro analogues of the duocarmycins. *Angew. Chem. Int. Ed.* **50**, 2606–2609 (2011).
- A report of a novel class of hypoxia-activated prodrugs of DNA minor groove alkylators showing potent and selective killing of hypoxic cells in xenograft models.**
45. Evans, J. W. *et al.* Homologous recombination is the principal pathway for the repair of DNA damage induced by tirapazamine in mammalian cells. *Cancer Res.* **68**, 257–265 (2008).
- Evidence for the critical importance of HR-mediated DNA repair in determining the sensitivity of hypoxic cells to TPZ.**
46. Gu, Y. *et al.* Roles of DNA repair and reductase activity in the cytotoxicity of the hypoxia-activated dinitrobenzamide mustard PR-104A. *Mol. Cancer Ther.* **8**, 1714–1723 (2009).
47. Branzei, D. & Foiani, M. Maintaining genome stability at the replication fork. *Nature Rev. Mol. Cell Biol.* **11**, 208–219 (2010).
48. Raleigh, S. M., Wanogho, E., Burke, M. D., McKeown, S. R. & Patterson, L. H. Involvement of human cytochromes P450 (CYP) in the reductive metabolism of AQ4N, a hypoxia activated anthraquinone di-N-oxide prodrug. *Int. J. Radiat. Oncol. Biol. Phys.* **42**, 763–767 (1998).
49. Nishida, C. R., Lee, M. & Ortiz de Montellano, P. R. Efficient hypoxic activation of the anticancer agent AQ4N by CYP2S1 and CYP2W1. *Mol. Pharmacol.* **78**, 497–502 (2010).
- A demonstration that the 'orphan' cytochrome P450 CYP2S1, which is upregulated under hypoxia, catalyses hypoxic activation of the bioreductive prodrug AQ4N.**
50. Rivera, S. P. *et al.* A novel promoter element containing multiple overlapping xenobiotic and hypoxia response elements mediates induction of cytochrome P4502S1 by both dioxin and hypoxia. *J. Biol. Chem.* **282**, 10881–10893 (2007).
51. Nishida, C. R. & Ortiz de Montellano, P. R. Reductive heme-dependent activation of the N-oxide prodrug AQ4N by nitric oxide synthase. *J. Med. Chem.* **51**, 5118–5120 (2008).
52. Colucci, M. A., Moody, C. J. & Couch, G. D. Natural and synthetic quinones and their reduction by the quinone reductase enzyme NQO1: from synthetic organic chemistry to compounds with anticancer potential. *Org. Biomol. Chem.* **6**, 637–656 (2008).
53. Knox, R. J. *et al.* The nitroreductase enzyme in Walker cells that activates 5-(aziridin-1-yl)-2,4-dinitrobenzamide (CB 1954) to 5-(aziridin-1-yl)-4-hydroxylamino-2-nitrobenzamide is a form of NAD(P)H dehydrogenase (quinone) (EC 1.6.99.2). *Biochem. Pharmacol.* **37**, 4671–4677 (1988).
- The first characterization of an oxygen-independent two-electron reductase responsible for the activation of a bioreductive prodrug.**
54. Celli, C. M., Tran, N., Knox, R. & Jaiswal, A. K. NRH:quinone oxidoreductase 2 (NQO2) catalyzes metabolic activation of quinones and anti-tumor drugs. *Biochem. Pharmacol.* **72**, 366–376 (2006).
55. Yan, C., Kopa, J. K., Siegel, D., Stratford, I. J. & Ross, D. Dissecting the role of multiple reductases in bioactivation and cytotoxicity of the anticancer agent 2, 5-diaziridinyl-3-(hydroxymethyl)-6-methyl-1,4-benzoquinone (RH1). *Mol. Pharmacol.* **74**, 1657–1665 (2008).
56. Guise, C. P. *et al.* The bioreductive prodrug PR-104A is activated under aerobic conditions by human aldo-keto reductase 1C3. *Cancer Res.* **70**, 1573–1584 (2010).
- The surprising observation that AKR1C3, which is highly expressed in some tumours, activates PR-104A (but not other bioreductive prodrugs) by a two-electron reduction.**
57. Adikesavan, A. K., Barrios, R. & Jaiswal, A. K. *In vivo* role of NAD(P)H:quinone oxidoreductase 1 in metabolic activation of mitomycin C and bone marrow cytotoxicity. *Cancer Res.* **67**, 7966–7971 (2007).
58. Siegel, D. & Ross, D. Immunodetection of NAD(P)H:quinone oxidoreductase 1 (NQO1) in human tissues. *Free Rad. Biol. Med.* **29**, 246–253 (2000).
59. Wang, W. & Jaiswal, A. K. Nuclear factor Nrf2 and antioxidant response element regulate NRH:quinone oxidoreductase 2 (NQO2) gene expression and antioxidant induction. *Free Rad. Biol. Med.* **40**, 1119–1130 (2006).
60. MacLeod, A. K. *et al.* Characterization of the cancer chemopreventive NRF2-dependent gene battery in human keratinocytes: demonstration that the KEAP1-NRF2 pathway, and not the BACH1-NRF2 pathway, controls cytoprotection against electrophiles as well as redox-cycling compounds. *Carcinogenesis* **30**, 1571–1580 (2009).
61. Cullinan, S. B. & Diehl, J. A. Coordination of ER and oxidative stress signaling: the PERK/Nrf2 signaling pathway. *Int. J. Biochem. Cell. Biol.* **38**, 317–332 (2006).
62. Jain, A. *et al.* Response of multiple recurrent TaT1 bladder cancer to intravesical apaziquone (EO9): comparative analysis of tumor recurrence rates. *Urology* **73**, 1083–1086 (2009).
63. Danson, S. J. *et al.* Phase I pharmacokinetic and pharmacodynamic study of the bioreductive drug RH1. *Ann. Oncol.* 4 Mar 2011 (doi:10.1093/annonc/mdq638).
64. Birtwistle, J. *et al.* The aldo-keto reductase AKR1C3 contributes to 7,12-dimethylbenz(a)anthracene-3,4-dihydrodiol mediated oxidative DNA damage in myeloid cells: implications for leukemogenesis. *Mutat. Res.* **662**, 67–74 (2009).
65. Konopleva, M. *et al.* Therapeutic targeting of the hypoxic microenvironment in acute lymphocytic leukemia. *Blood (ASH Annual Meeting Abstracts)* **114**, Abstract 2040 [online], <http://ash.confex.com/ash/2009/webprogram/Paper24327.html> (2009).
66. Hu, J. *et al.* Targeting the multiple myeloma hypoxic niche with TH-302, a hypoxia-activated prodrug. *Blood* **116**, 1524–1527 (2010).
67. Baker, M. A., Zeman, E. M., Hirst, V. K. & Brown, J. M. Metabolism of SR 4233 by Chinese hamster ovary cells: basis of selective hypoxic cytotoxicity. *Cancer Res.* **48**, 5947–5952 (1988).
68. Hicks, K. O. *et al.* Pharmacokinetic/pharmacodynamic modeling identifies SN30000 and SN29751 as tirapazamine analogues with improved tissue penetration and hypoxic cell killing in tumors. *Clin. Cancer Res.* **16**, 4946–4957 (2010).
- The development of a second-generation benzotriazine dioxide analogue of TPZ using a novel testing algorithm based on spatially resolved pharmacokinetic and pharmacodynamic modelling.**
69. Minchinton, A. I. & Tannock, I. F. Drug penetration in solid tumours. *Nature Rev. Cancer* **6**, 583–592 (2006).
70. Durand, R. E. & Olive, P. L. Physiologic and cytotoxic effects of tirapazamine in tumor-bearing mice. *Radiat. Oncol. Invest.* **5**, 213–219 (1997).
71. Durand, R. E. & Olive, P. L. Evaluation of bioreductive drugs in multicell spheroids. *Int. J. Radiat. Oncol. Biol. Phys.* **22**, 689–692 (1992).

72. Hicks, K. O., Fleming, Y., Siim, B. G., Koch, C. J. & Wilson, W. R. Extravascular diffusion of tirapazamine: effect of metabolic consumption assessed using the multicellular layer model. *Int. J. Radiat. Oncol. Biol. Phys.* **42**, 641–649 (1998).
73. Kyle, A. H. & Minchinton, A. I. Measurement of delivery and metabolism of tirapazamine to tumour tissue using the multilayered cell culture model. *Cancer Chemother. Pharmacol.* **43**, 213–220 (1999).
74. Hicks, K. O. *et al.* Use of three-dimensional tissue cultures to model extravascular transport and predict *in vivo* activity of hypoxia-targeted anticancer drugs. *J. Natl Cancer Inst.* **98**, 1118–1128 (2006). **Conclusive evidence that hypoxic cell killing by TPZ and its analogues, in xenografts, is limited by the ability of these drugs to penetrate hypoxic tumour tissue.**
75. Puijn, F. B., Patel, K., Hay, M. P., Wilson, W. R. & Hicks, K. O. Prediction of tumour tissue diffusion coefficients of hypoxia-activated prodrugs from physicochemical parameters. *Aust. J. Chem.* **61**, 687–693 (2008).
76. Hay, M. P. *et al.* Hypoxia-selective 3-alkyl 1,2,4-benzotriazine 1,4-dioxides: the influence of hydrogen bond donors on extravascular transport and antitumor activity. *J. Med. Chem.* **50**, 6654–6664 (2007).
77. Koch, C. J. Unusual oxygen concentration dependence of toxicity of SR-4233, a hypoxic cell toxin. *Cancer Res.* **53**, 3992–3997 (1993). **The first report that the inhibition of TPZ cytotoxicity requires higher oxygen concentrations than for nitro compounds and quinones.**
78. Hicks, K. O., Siim, B. G., Puijn, F. B. & Wilson, W. R. Oxygen dependence of the metabolic activation and cytotoxicity of tirapazamine: implications for extravascular transport and activity in tumors. *Radiat. Res.* **161**, 656–666 (2004).
79. Marshall, R. S. & Rauth, A. M. Oxygen and exposure kinetics as factors influencing the cytotoxicity of porfomycin, a mitomycin C analogue, in Chinese hamster ovary cells. *Cancer Res.* **48**, 5655–5659 (1988).
80. Siim, B. G., Atwell, G. J. & Wilson, W. R. Oxygen dependence of the cytotoxicity and metabolic activation of 4-alkylamino-5-nitroquinoline bioreductive drugs. *Br. J. Cancer* **70**, 596–603 (1994).
81. Wilson, W. R., Moselen, J. W., Cliffe, S., Denny, W. A. & Ware, D. C. Exploiting tumor hypoxia through bioreductive release of diffusible cytotoxins: the cobalt(III)-nitrogen mustard complex SN 24771. *Int. J. Radiat. Oncol. Biol. Phys.* **29**, 323–327 (1994).
82. Wilson, W. R. *et al.* Bystander effects of bioreductive drugs: potential for exploiting pathological tumor hypoxia with dinitrobenzamide mustards. *Radiat. Res.* **167**, 625–636 (2007).
83. Hicks, K. O. *et al.* Oxygen dependence and extravascular transport of hypoxia-activated prodrugs: comparison of the dinitrobenzamide mustard PR-104A and tirapazamine. *Int. J. Radiat. Oncol. Biol. Phys.* **69**, 560–571 (2007).
84. Helleday, T., Petermann, E., Lundin, C., Hodgson, B. & Sharma, R. A. DNA repair pathways as targets for cancer therapy. *Nature Rev. Cancer* **8**, 193–204 (2008).
85. Parveen, I., Naughton, D. P., Whish, W. J. & Threadgill, M. D. 2-nitroimidazol-5-ylmethyl as a potential bioreductively activated prodrug system: reductively triggered release of the PARP inhibitor 5-bromoisoquinolinone. *Bioorg. Med. Chem. Lett.* **9**, 2031–2036 (1999).
86. Everett, S. A., Naylor, M. A., Patel, K. B., Stratford, M. R. L. & Wardman, P. Bioreductively-activated prodrugs for targeting hypoxic tissues: elimination of aspirin from 2-nitroimidazole derivatives. *Bioorg. Med. Chem. Lett.* **9**, 1267–1272 (1999).
87. Thomson, P. *et al.* Synthesis and biological properties of bioreductively targeted nitrothienyl prodrugs of combretastatin A-4. *Mol. Cancer Ther.* **5**, 2886–2894 (2006).
88. Granchi, C. *et al.* Bioreductively activated lysyl oxidase inhibitors against hypoxic tumours. *ChemMedChem* **4**, 1590–1594 (2009).
89. Tercel, M., Wilson, W. R., Anderson, R. F. & Denny, W. A. Hypoxia-selective antitumor agents. 12. Nitrobenzyl quaternary salts as bioreductive prodrugs of the alkylating agent mechlorethamine. *J. Med. Chem.* **39**, 1084–1094 (1996).
90. Patterson, A. V. *et al.* Cellular metabolism, murine pharmacokinetics and preclinical antitumor activity of SN29966, a novel hypoxia-activated irreversible pan-HER inhibitor. *Mol. Cancer Ther.* **8** (Suppl. 1), B76 (2009).
91. Semenza, G. L. Targeting HIF-1 for cancer therapy. *Nature Rev. Cancer* **3**, 721–732 (2003).
92. Poon, E., Harris, A. L. & Ashcroft, M. Targeting the hypoxia-inducible factor (HIF) pathway in cancer. *Expert Rev. Mol. Med.* **11**, e26 (2009).
93. Wouters, B. G. & Koritzinsky, M. Hypoxia signalling through mTOR and the unfolded protein response in cancer. *Nature Rev. Cancer* **8**, 851–864 (2008).
94. Martinive, P. *et al.* Preconditioning of the tumor vasculature and tumor cells by intermittent hypoxia: implications for cancer therapies. *Cancer Res.* **66**, 11736–11744 (2006).
95. Semenza, G. L. Evaluation of HIF-1 inhibitors as anticancer agents. *Drug Discov. Today* **12**, 853–859 (2007).
96. Giaccia, A., Siim, B. G. & Johnson, R. S. HIF-1 as a target for drug development. *Nature Rev. Drug Discov.* **2**, 803–811 (2003).
97. Melillo, G. Targeting hypoxia cell signalling for cancer therapy. *Cancer Metastasis Rev.* **26**, 341–352 (2007).
98. Isaacs, J. S. *et al.* Hsp90 regulates a von Hippel Lindau-independent hypoxia-inducible factor-1 alpha-degradative pathway. *J. Biol. Chem.* **277**, 29936–29944 (2002).
99. Kizaka-Kondoh, S., Tanaka, S., Harada, H. & Hiraoka, M. The HIF-1-active microenvironment: an environmental target for cancer therapy. *Advanced Drug Deliv. Rev.* **61**, 623–632 (2009).
100. Rouschop, K. M. & Wouters, B. G. Regulation of autophagy through multiple independent hypoxic signaling pathways. *Curr. Mol. Med.* **9**, 417–424 (2009).
101. Tu, B. P. & Weissman, J. S. The FAD- and O₂-dependent reaction cycle of Ero1-mediated oxidative protein folding in the endoplasmic reticulum. *Mol. Cell* **10**, 983–994 (2002).
102. Koditz, J. *et al.* Oxygen-dependent ATF-4 stability is mediated by the PHD3 oxygen sensor. *Blood* **110**, 3610–3617 (2007).
103. Bi, M. *et al.* ER stress-regulated translation increases tolerance to extreme hypoxia and promotes tumor growth. *EMBO J.* **24**, 3470–3481 (2005).
104. Romero-Ramirez, L. *et al.* XBP1 is essential for survival under hypoxic conditions and is required for tumor growth. *Cancer Res.* **64**, 5943–5947 (2004). **The identification of IRE1 as a target for killing hypoxic cells.**
105. Spiotto, M. T. *et al.* Imaging the unfolded protein response in primary tumors reveals microenvironments with metabolic variations that predict tumor growth. *Cancer Res.* **70**, 78–88 (2010).
106. Papanadrou, I. *et al.* Identification of an Ire1α endonuclease specific inhibitor with cytotoxic activity against human multiple myeloma. *Blood* **117**, 1311–1314 (2011).
107. Volkman, K. *et al.* Potent and selective inhibitors of the inositol-requiring enzyme 1 endoribonuclease. *J. Biol. Chem.* **286**, 12743–12755 (2011). **The first demonstration of salicylaldehydes as novel IRE1 inhibitors in vitro and in vivo.**
108. Lee, K. P. *et al.* Structure of the dual enzyme Ire1 reveals the basis for catalysis and regulation in nonconventional RNA splicing. *Cell* **132**, 89–100 (2008).
109. Fels, D. R. *et al.* Preferential cytotoxicity of bortezomib toward hypoxic tumor cells via overactivation of endoplasmic reticulum stress pathways. *Cancer Res.* **68**, 9323–9330 (2008).
110. Koumenis, C. & Wouters, B. G. “Translating” tumor hypoxia: unfolded protein response (UPR)-dependent and UPR-independent pathways. *Mol. Cancer Res.* **4**, 423–436 (2006).
111. Brugarolas, J. *et al.* Regulation of mTOR function in response to hypoxia by REDD1 and the TSC1/TSC2 tumor suppressor complex. *Genes Dev.* **18**, 2893–2904 (2004).
112. Liu, L. *et al.* Hypoxia-induced energy stress regulates mRNA translation and cell growth. *Mol. Cell* **21**, 521–531 (2006).
113. Li, Y. *et al.* BNIP3 mediates the hypoxia-induced inhibition on mammalian target of rapamycin by interacting with Rheb. *J. Biol. Chem.* **282**, 35803–35813 (2007).
114. Pencreach, E. *et al.* Marked activity of irinotecan and rapamycin combination toward colon cancer cells *in vivo* and *in vitro* is mediated through cooperative modulation of the mammalian target of rapamycin/hypoxia-inducible factor-1α axis. *Clin. Cancer Res.* **15**, 1297–1307 (2009).
115. Yu, K. *et al.* Beyond rapalog therapy: preclinical pharmacology and antitumor activity of WYE-125132, an ATP-competitive and specific inhibitor of mTORC1 and mTORC2. *Cancer Res.* **70**, 621–631 (2010).
116. Rzymiski, T. *et al.* Regulation of autophagy by ATF4 in response to severe hypoxia. *Oncogene* **29**, 4424–4435 (2010).
117. Marin-Hernandez, A., Gallardo-Perez, J. C., Ralph, S. J., Rodriguez-Enriquez, S. & Moreno-Sanchez, R. HIF-1α modulates energy metabolism in cancer cells by inducing over-expression of specific glycolytic isoforms. *Mini-Rev. Med. Chem.* **9**, 1084–1101 (2009).
118. Levine, A. J. & Puzio-Kuter, A. M. The control of the metabolic switch in cancers by oncogenes and tumor suppressor genes. *Science* **330**, 1340–1344 (2010).
119. Kurtoglu, M., Maher, J. C. & Lampidis, T. J. Differential toxic mechanisms of 2-deoxy-D-glucose versus 2-fluorodeoxy-D-glucose in hypoxic and normoxic tumor cells. *Antiox. Redox Signal.* **9**, 1383–1390 (2007). **A critical review addressing the mechanism of hypoxia-selective cytotoxicity of glucose analogs.**
120. Song, C. W., Clement, J. J. & Levitt, S. H. Preferential cytotoxicity of 5-thio-D-glucose against hypoxic tumor cells. *J. Natl Cancer Inst.* **57**, 603–605 (1976).
121. Lampidis, T. J. *et al.* Efficacy of 2-halogen substituted D-glucose analogs in blocking glycolysis and killing “hypoxic tumor cells”. *Cancer Chemother. Pharmacol.* **58**, 725–734 (2006).
122. Macheda, M. L., Rogers, S. & Best, J. D. Molecular and cellular regulation of glucose transporter (GLUT) proteins in cancer. *J. Cell. Physiol.* **202**, 654–662 (2005).
123. Wood, T. E. *et al.* A novel inhibitor of glucose uptake sensitizes cells to FAS-induced cell death. *Mol. Cancer Ther.* **7**, 3546–3555 (2008).
124. Lai, E. W., Chan, D. A., Hay, M. P. & Giaccia, A. J. Selective cytotoxic targeting of von Hippel-Lindau-deficient renal carcinoma cells. *Proc. Am. Assoc. Cancer Res.* **51**, Abstract 67 (2010).
125. Sonveaux, P. *et al.* Targeting lactate-fueled respiration selectively kills hypoxic tumor cells in mice. *J. Clin. Invest.* **118**, 3930–3942 (2008). **A demonstration that inhibition of MCT1 blocks the use of lactate for respiration in aerobic cells, eliciting rapid glucose consumption and death of hypoxic cells owing to glucose starvation.**
126. Owens, M. J., Davies, A. J., Wilson, M. C., Murray, C. M. & Halestrap, A. P. AR-C155858 is a potent inhibitor of monocarboxylate transporters MCT1 and MCT2 that binds to an intracellular site involving transmembrane helices 7–10. *Biochem. J.* **425**, 523–530 (2010).
127. Murray, C. M. *et al.* Monocarboxylate transporter MCT1 is a target for immunosuppression. *Nature Chem. Biol.* **1**, 371–376 (2005).
128. Chiche, J., Brahimi-Horn, M. C. & Pouyssegur, J. Tumor hypoxia induces a metabolic shift causing acidosis: a common feature in cancer. *J. Cell. Mol. Med.* **14**, 771–794 (2010).
129. Ullah, M. S., Davies, A. J. & Halestrap, A. P. The plasma membrane lactate transporter MCT4, but not MCT1, is up-regulated by hypoxia through a HIF-1α-dependent mechanism. *J. Biol. Chem.* **281**, 9030–9037 (2006).
130. Gallagher, S. M., Castorino, J. J., Wang, D. & Philp, N. J. Monocarboxylate transporter 4 regulates maturation and trafficking of CD147 to the plasma membrane in the metastatic breast cancer cell line MDA-MB-231. *Cancer Res.* **67**, 4182–4189 (2007).
131. Chiche, J. *et al.* Hypoxia-inducible carbonic anhydrase IX and XII promote tumor cell growth by counteracting acidosis through the regulation of the intracellular pH. *Cancer Res.* **69**, 358–368 (2009).
132. van den Beucken, T. *et al.* Hypoxia-induced expression of carbonic anhydrase 9 is dependent on the unfolded protein response. *J. Biol. Chem.* **284**, 24204–24212 (2009).
133. Lou, Y. *et al.* Targeting tumor hypoxia: suppression of breast tumor growth and metastasis by novel carbonic anhydrase IX inhibitors. *Cancer Res.* **71**, 3364–3376 (2011).
134. Huang, L. E., Bindra, R. S., Glazer, P. M. & Harris, A. L. Hypoxia-induced genetic instability—a calculated mechanism underlying tumor progression. *J. Mol. Med.* **85**, 139–148 (2007).

135. Olcina, M., Lecane, P. S. & Hammond, E. M. Targeting hypoxic cells through the DNA damage response. *Clin. Cancer Res.* **16**, 5620–5629 (2010).
136. Chan, N., Koch, C. J. & Bristow, R. G. Tumor hypoxia as a modifier of DNA strand break and cross-link repair. *Curr. Mol. Med.* **9**, 401–410 (2009).
137. Hammond, E. M., Green, S. L. & Giaccia, A. J. Comparison of hypoxia-induced replication arrest with hydroxyurea and aphidicolin-induced arrest. *Mutat. Res.* **532**, 205–213 (2003).
138. Pires, I. M. *et al.* Effects of acute versus chronic hypoxia on DNA damage responses and genomic instability. *Cancer Res.* **70**, 925–935 (2010). **A demonstration that severe hypoxia induces DNA replication arrest through loss of ribonucleotide reductase activity and suppression of dNTP pools.**
139. Norlund, P. & Reichard, P. Ribonucleotide reductases. *Ann. Rev. Biochem.* **75**, 681–706 (2006).
140. Hammond, E. M., Dorie, M. J. & Giaccia, A. J. Inhibition of ATR leads to increased sensitivity to hypoxia/reoxygenation. *Cancer Res.* **64**, 6556–6562 (2004).
141. Hammond, E. M., Dorie, M. J. & Giaccia, A. J. ATR/ATM targets are phosphorylated by ATR in response to hypoxia and ATM in response to reoxygenation. *J. Biol. Chem.* **278**, 12207–12213 (2003).
142. Gibson, S. L., Bindra, R. S. & Glazer, P. M. CHK2-dependent phosphorylation of BRCA1 in hypoxia. *Radiat. Res.* **166**, 646–651 (2006).
143. Chang, C.-J. *et al.* EZH2 promotes expansion of breast tumor initiating cells through activation of RAF1- β -catenin signaling. *Cancer Cell* **19**, 86–100 (2011).
144. Marotta, D. *et al.* *In vivo* profiling of hypoxic gene expression in gliomas using the hypoxia marker EF5 and laser-capture microdissection. *Cancer Res.* **71**, 779–789 (2011).
145. Bindra, R. S. *et al.* Down-regulation of Rad51 and decreased homologous recombination in hypoxic cancer cells. *Mol. Cell. Biol.* **24**, 8504–8518 (2004). **An early demonstration of the effects of hypoxia on HR repair and the implications of hypoxia for genomic instability and therapy.**
146. Chan, N. *et al.* Chronic hypoxia decreases synthesis of homologous recombination proteins to offset chemoresistance and radioresistance. *Cancer Res.* **68**, 605–614 (2008). **A demonstration of selective killing of hypoxic cells by a PARP inhibitor in cell culture and in xenografts.**
147. Chan, N. *et al.* Contextual synthetic lethality of cancer cell kill based on the tumor microenvironment. *Cancer Res.* **70**, 8045–8054 (2010).
148. Di Cintio, A., Di Gennaro, E. & Budillon, A. Restoring p53 function in cancer: novel therapeutic approaches for applying the brakes to tumorigenesis. *Recent Pat. Anti-Cancer Drug Discov.* **5**, 1–13 (2010).
149. Yang, J. *et al.* Small-molecule activation of p53 blocks hypoxia-inducible factor 1 α and vascular endothelial growth factor expression *in vivo* and leads to tumor cell apoptosis in normoxia and hypoxia. *Mol. Cell. Biol.* **29**, 2243–2253 (2009).
150. Hoogsteen, I. J. *et al.* Hypoxia in larynx carcinomas assessed by pimonidazole binding and the value of CA-IX and vascularity as surrogate markers of hypoxia. *Eur. J. Cancer* **45**, 2906–2914 (2009).
151. Nordmark, M. *et al.* Prognostic value of tumor oxygenation in 397 head and neck tumors after primary radiation therapy. An international multicenter study. *Radiother. Oncol.* **77**, 18–24 (2005). **A meta-analysis of the prognostic significance of tumour hypoxia in head and neck tumours, assessed with oxygen electrodes, demonstrating a major impact on outcome after radiotherapy.**
152. Mees, G., Dierckx, R., Vangestel, C. & Van De, W. C. Molecular imaging of hypoxia with radiolabelled agents. *Eur. J. Nucl. Med. Mol. Imag.* **36**, 1674–1686 (2009).
153. Mason, R. P. *et al.* Multimodality imaging of hypoxia in preclinical settings. *Quart. J. Nucl. Med. Mol. Imag.* **54**, 259–280 (2010).
154. Workman, P. & Stratford, I. J. The experimental development of bioreductive drugs and their role in cancer therapy. *Cancer Metastasis Rev.* **12**, 73–82 (1993).
155. Willers, H. *et al.* Utility of DNA repair protein foci for the detection of putative BRCA1 pathway defects in breast cancer biopsies. *Mol. Cancer Res.* **7**, 1304–1309 (2009).
156. Konstantinopoulos, P. A. *et al.* Gene expression profile of BRCAness that correlates with responsiveness to chemotherapy and with outcome in patients with epithelial ovarian cancer. *J. Clin. Oncol.* **28**, 3555–3561 (2010).
157. Graeser, M. *et al.* A marker of homologous recombination predicts pathological complete response to neoadjuvant chemotherapy in primary breast cancer. *Clin. Cancer Res.* **16**, 6159–6158 (2010).
158. Wang, J. *et al.* EF5 as a predictive biomarker for activation of the new hypoxia targeting prodrug SN30000. *Am. Soc. Clin. Oncol. Abstract e13597*, (2011).
159. Hedley, D. *et al.* Carbonic anhydrase IX expression, hypoxia, and prognosis in patients with uterine cervical carcinoma. *Clin. Cancer Res.* **9**, 5666–5674 (2003).
160. Whitehurst, A. W. *et al.* Synthetic lethal screen identification of chemosensitizer loci in cancer cells. *Nature* **446**, 815–819 (2007).
161. Barabasi, A.-L., Gulbache, N. & Loscalzo, J. Network medicine: a network-based approach to human disease. *Nature Rev. Genet.* **12**, 56–68 (2011).
162. Moeller, B. J., Cao, Y., Li, C. Y. & Dewhirst, M. W. Radiation activates HIF-1 to regulate vascular radiosensitivity in tumors: role of reoxygenation, free radicals, and stress granules. *Cancer Cell* **5**, 429–441 (2004).
163. Garner, A. P. *et al.* Nitric oxide synthases catalyze the activation of redox cycling and bioreductive anticancer agents. *Cancer Res.* **59**, 1929–1934 (1999).
164. Chinje, E. C. *et al.* Non-nuclear localized human NOSII enhances the bioactivation and toxicity of tirapazamine (SR4233) *in vitro*. *Mol. Pharmacol.* **63**, 1248–1255 (2003).
165. Thomsen, L. L. *et al.* Nitric oxide synthase activity in human breast cancer. *Br. J. Cancer* **72**, 41–44 (1995).
166. Swana, H. S. *et al.* Inducible nitric oxide synthase with transitional cell carcinoma of the bladder. *J. Urol.* **161**, 630–634 (1999).
167. Lewis, C. & Murdoch, C. Macrophage responses to hypoxia: implications for tumor progression and anticancer therapies. *Am. J. Pathol.* **167**, 627–635 (2005).
168. Melillo, G. *et al.* A hypoxia-responsive element mediates a novel pathway of activation of the inducible nitric oxide synthase promoter. *J. Exp. Med.* **182**, 1683–1693 (1995).
169. Tendler, D. S. *et al.* Intersection of interferon and hypoxia signal transduction pathways in nitric oxide-induced tumor apoptosis. *Cancer Res.* **61**, 3682–3688 (2001).
170. Chinje, E. C. *et al.* 17 β -Oestradiol treatment modulates nitric oxide synthase activity in MDA231 tumour with implications on growth and radiation response. *Br. J. Cancer* **86**, 136–142 (2002).
171. Patterson, L. H. & McKeown, S. R. AQ4N: a new approach to hypoxia-activated cancer chemotherapy. *Br. J. Cancer* **83**, 1589–1593 (2000).
172. Chandor, A. *et al.* Metabolic activation of the antitumor drug 5-(Aziridin-1-yl)-2,4-dinitrobenzamide (CB1954) by NO synthases. *Chem. Res. Toxicol.* **21**, 836–843 (2008).
173. Guise, C. P. *et al.* Identification of human oxidoreductases involved in the hypoxia-dependent activation of bioreductive prodrugs. *Proc. Am. Assoc. Cancer Res.* **51**, Abstract 453 (2010).
174. Kan, O. *et al.* Genetically modified macrophages expressing hypoxia regulated cytochrome P450 and P450 reductase for the treatment of cancer. *Int. J. Mol. Med.* **27**, 173–180 (2011).
175. Cowen, R. L. *et al.* Hypoxia targeted gene therapy to increase the efficacy of tirapazamine as an adjuvant to radiotherapy: reversing tumor radioresistance and effecting cure. *Cancer Res.* **64**, 1396–1402 (2004).
176. Hodnick, W. F. & Sartorelli, A. C. Reductive activation of mitomycin C by NADH:cytochrome b_5 reductase. *Cancer Res.* **53**, 4907–4912 (1993).
177. Miskiniene, V., Dickanaitė, E., Nemeikaitė, A. & Cenā, N. Nitroaromatic betulin derivatives as redox cycling agents. *Biochem. Mol. Biol. Int.* **42**, 391–397 (1997).
178. Kutcher, W. W. & McCalla, D. R. Aerobic reduction of 5-nitro-2-furaldehyde semicarbazone by rat liver xanthine dehydrogenase. *Biochem. Pharmacol.* **33**, 799–805 (1984).
179. Ahlskog, J. K. *et al.* Human monoclonal antibodies targeting carbonic anhydrase IX for the molecular imaging of hypoxic regions in solid tumours. *Br. J. Cancer* **101**, 645–657 (2009).
180. Hoeben, B. A. *et al.* PET of hypoxia with 89Zr-labeled cG250-F(ab) $_2$ in head and neck tumors. *J. Nucl. Med.* **51**, 1076–1083 (2010).
181. Kaluz, S., Kaluzova, M., Liao, S. Y., Lerman, M. & Stanbridge, E. J. Transcriptional control of the tumor- and hypoxia-marker carbonic anhydrase 9: a one transcription factor (HIF-1) show? *Biochim. Biophys. Acta* **1795**, 162–172 (2009).
182. Komar, G. *et al.* 18F-EF5: a new PET tracer for imaging hypoxia in head and neck cancer. *J. Nucl. Med.* **49**, 1944–1951 (2008).
183. Rischin, D. *et al.* Prognostic significance of [18F]-misonidazole positron emission tomography-detected tumor hypoxia in patients with advanced head and neck cancer randomly assigned to chemoradiation with or without tirapazamine: a substudy of Trans-Tasman Radiation Oncology Group Study 98.02. *J. Clin. Oncol.* **24**, 2098–2104 (2006). **A provocative study, from a single clinical centre, demonstrating that TPZ-responsive patients can be imaged by PET imaging of hypoxia.**
184. Rischin, D. *et al.* Tirapazamine, cisplatin, and radiation versus cisplatin and radiation for advanced squamous cell carcinoma of the head and neck (TROG 02.02, HeadSTART): a phase III trial of the Trans-Tasman Radiation Oncology Group. *J. Clin. Oncol.* **28**, 2989–2995 (2010).
185. Wouters, B. G. & Brown, J. M. Cells at intermediate oxygen levels can be more important than the “hypoxic fraction” in determining tumor response to fractionated radiotherapy. *Radiat. Res.* **147**, 541–550 (1997). **An important theoretical study that makes the case that moderately hypoxic (partially radioresistant) cells are more important to radiotherapy outcome than the most hypoxic cells in tumours.**
186. Tuttle, S. W. *et al.* Detection of reactive oxygen species via endogenous oxidative pentose phosphate cycle activity in response to oxygen concentration. *J. Biol. Chem.* **282**, 36790–36796 (2007).
187. Rzymiski, T. & Harris, A. L. The unfolded protein response and integrated stress response to anoxia. *Clin. Cancer Res.* **13**, 2537–2540 (2007).

Acknowledgements

We thank our collaborators (R. Anderson, W. Denny, K. Hicks, C. Guise, A. Patterson, F. Puijui, J. Smail, M. Tercel and J. Wang) for many fruitful discussions that have helped to inform the views expressed here. The authors acknowledge financial support from the Health Research Council of New Zealand (W.R.W.) and the Maurice Wilkins Centre for Biodiscovery (M.P.H.).

Competing interests statement

The authors declare competing financial interests: see Web version for details.

DATABASES

ClinicalTrials.gov: <http://www.clinicaltrials.gov>
 NCT00466583 | NCT00522652 | NCT01144455
 US National Cancer Institute Drug Dictionary:
<http://www.cancer.gov/drugdictionary>
 apaziquone | APR-246 | banoxantrone | borteozomib |
 CB1954/caricotamide combination | chloroquine |
 combretastatin A4 | EZN-2968 | geldanamycin | gemcitabine |
 irinotecan | mitomycin C | pimonidazole | PR-104 | PX-478 |
 rapamycin | RH1 | TH-302 | thapsigargin | tirapazamine |
 veliparib

FURTHER INFORMATION

William R. Wilson's homepage: <http://www.fmhs.auckland.ac.nz/sms/acsrc/etg/default.aspx>
 Michael P. Hay's homepage: http://www.fmhs.auckland.ac.nz/sms/acsrc/medicinal_chemistry/hay/default.aspx

SUPPLEMENTARY INFORMATION

See online article: S1 (tables)
 ALL LINKS ARE ACTIVE IN THE ONLINE PDF