

RESEARCH

Open Access

Short-term diesel exhaust inhalation in a controlled human crossover study is associated with changes in DNA methylation of circulating mononuclear cells in asthmatics

Ruiwei Jiang¹, Meaghan J Jones¹, Francesco Sava², Michael S Kobor^{1,3} and Christopher Carlsten^{2*}

Abstract

Background: Changes in DNA methylation have been associated with traffic-related air pollution in observational studies, but the specific mechanisms and temporal dynamics therein have not been explored in a controlled study of asthmatics. In this study, we investigate short-term effects of diesel exhaust inhalation on DNA methylation levels at CpG sites across the genome in circulating blood in asthmatics.

Methods: A double-blind crossover study of filtered air and diesel exhaust exposures was performed on sixteen non-smoking asthmatic subjects. Blood samples were collected pre-exposure, and then 6 and 30 hours post-exposure. Peripheral blood mononuclear cell DNA methylation was interrogated using the Illumina Infinium HumanMethylation450 Array. Exposure-related changes in DNA methylation were identified. In addition, CpG sites overlapping with Alu or LINE1 repetitive elements and candidate microRNA loci were also analyzed.

Results: DNA methylation at 2827 CpG sites were affected by exposure to diesel exhaust but not filtered air; these sites enriched for genes involved in protein kinase and *NFκB* pathways. CpG sites with significant changes in response to diesel exhaust exposure primarily became less methylated, with a site residing within *GSTP1* being among the significant hits. Diesel exhaust-associated change was also found for CpG sites overlapping with Alu and LINE1 elements as well as for a site within *miR-21*.

Conclusion: Short-term exposure to diesel exhaust resulted in DNA methylation changes at CpG sites residing in genes involved in inflammation and oxidative stress response, repetitive elements, and microRNA. This provides plausibility for the role of DNA methylation in pathways by which airborne particulate matter impacts gene expression and offers support for including DNA methylation analysis in future efforts to understand the interactions between environmental exposures and biological systems.

Keywords: Particulate matter, Air pollution, Epigenetics, Interspersed repetitive sequences, microRNAs

Background

Exposure to air pollutants is an increasing public health concern that has been associated with adverse health effects focused on cardiovascular and respiratory diseases [1-3]. Traffic-derived pollution appears particularly toxic, perhaps due to its abundance of fine particulate matter

(PM) [4]. Ambient PM is a heterogeneous mix of components varying in concentration, and chemistry [5]. PM is associated with both chronic and acute effects on health [5], including increased risk of lung cancer [6-8], increased hospital admission rate for respiratory and cardiovascular diseases in both adults and children [3,5,9], and increased risk for mortality [10]. In urban environments, the major contributor to fine PM (diameter between 0.1 μm and 2.5 μm) is diesel exhaust (DE) [11]. Due to its small size, fine PM can deposit deep in the lung, and its soluble components such as transition

* Correspondence: carlsten@mail.ubc.ca

²Air Pollution Exposure Laboratory, Chan-Yeung Centre for Occupational and Environmental Lung Disease, Department of Medicine, Division of Respiratory Medicine, University of British Columbia, 2775 Laurel Street, Vancouver, British Columbia V5Z1L9, Canada

Full list of author information is available at the end of the article

metals may cross the lung epithelium into systemic circulation and interact with internal organs [2]. On the cellular level, exposure to DE prompts the generation of reactive oxygen species, leading to oxidative stress and damage to cellular structures [12-14]. On the molecular scale, DE has been found to change microRNA expression, increase production of allergic antibodies, up-regulate mRNA expression of pro-inflammatory mediators and antioxidant enzymes, as well as decrease methylation of repetitive genomic elements [15-19]. The hazardous effects of DE is also associated with asthma susceptibility and severity; for example, evidence links exposure to diesel exhaust with worse lung functions and increased airway resistance [20,21]. However, the precise mechanism by which pollution exacerbates asthma is not yet fully understood [5]. It has also been hypothesized that exposure to DE may be partly responsible for the increase in allergic diseases in industrialized nations [22].

One possible mechanism through which air pollution impacts transcriptional pathways may be exposure-related epigenetic modifications. Epigenetics refers to persistent changes in gene regulation that do not involve changes in DNA sequence [23]. Arguably the most studied and best-understood epigenetic modification is DNA methylation, the covalent addition of a methyl group to a cytosine primarily in the context of a cytosine-guanine dinucleotide (CpG). DNA methylation modulates gene expression, and can vary in response to external stimuli [23-26]. DNA methylation is also commonly observed at repetitive elements. LINE1 and Alu generally have higher methylation than the rest of the genome, and their methylation level is negatively correlated with mobility of retrotransposons [27]. Increased mobility of these retrotransposons, especially at cancer-related genomic loci, is associated with mutation and higher tumorigenesis rates [27]. Many recent studies have shown that ambient particulate matter change methylation of LINE1 and Alu repetitive elements as well as that of pro-inflammatory and tumor suppressor genes [28-31]. Thus methylation at repeat elements is responsive to environmental influences; this could affect chromosomal arrangements and gene expression, relating environment to disease.

These epigenetic responses to environmental exposures are not limited to repetitive elements and particulate matter [23,25,32]. DNA methylation is an important mechanism through which the environment in general interacts with the genome, likely affecting phenotypic outcomes. Environmental effects with known DNA methylation associations include socio-economic status, pollution, stress, and personal habits such as diet and smoking [16,23,33]. Although the mechanisms behind these associations are still unknown, it is undeniable that there is a complex dynamic at play among environment, genes and epigenetics. As such, epigenetics presents an intriguing target for

understanding the mechanisms behind the adverse effects of DE.

Despite the numerous existing investigations regarding ambient particulate matter and DNA methylation, no controlled investigation of particulate matter on DNA methylation at sites distributed across the genome, an important starting point for unbiased mechanistic inquiry, has been reported. We were interested in understanding the systemic impact of air pollution from the perspective of DNA methylation, and thus focused on peripheral blood mononuclear cells (PBMCs). We hypothesized that short-term exposure to DE would lead to changes in DNA methylation status of PBMCs at CpG sites across the genome in asthmatic individuals, especially in genes relevant to the etiology of allergic diseases. Furthermore, we speculated that we would also observe changes in methylation of LINE1 and Alu repetitive elements, given that repetitive elements have been shown to be sensitive epigenetic indicators of environmental exposure [16,30]. Lastly, we investigated whether any methylation changes were linked to DE induced changes in microRNA expression that we have previously demonstrated in the same individuals tested here [19].

Results

Diesel exhaust was associated with changes in DNA methylation

The demographic characteristics of the sixteen subjects with physician-diagnosed asthma and/or baseline methacholine PC₂₀ below 8 mg/mL are in Table 1. Using PCA, a dimensionality reduction technique, we decomposed the data to a set of 95 principal components (PCs), each of which explained a dominant and independent pattern of variation across the samples; we then assessed the relative contribution of each principal component to the overall variance (see Additional file 1: Figure S1 in online data supplement). PCA facilitated linear decomposition of the data, allowing identification of data dimension that significantly captured the association between DNA methylation change and exposure to DE. By focusing our analysis on the CpG sites most representative of that dimension, we were able to analyze only CpG sites with higher chance of being DE-associated. Surveying the pattern of methylation within the data, we found that demographic variables were associated with a number of PCs, and many of the associations were driven by correlations inherent among these variables (see Additional file 2: Figure S2 in online data supplement; significant associations were captured after multiple testing correction by the q-value method with FDR = 10%). In particular, age was a major factor driving data variability in PCs 1, 4, and 5, and BMI was strongly associated with PC4 (Figure 1). Since PBMCs are heterogeneous, we considered the contribution of blood cell composition to methylation change. To achieve this, we tested for association between PCs and white blood cell counts. We did

Table 1 Subject demographics used with principal component scores to determine main drivers of variation in data

Variable	Measurements
Gender: Male	7 [44%] [†]
Age, years	28.7 ± 6.7 [19.2-46.5]
Ethnicity	
Asian	2 [13%] [†]
Caucasian	12 [75%] [†]
Middle Eastern	1 [6%] [†]
South Asian	1 [6%] [†]
Height, cm	170.0 ± 12.2 [151.0-185.5]
Mass, kg	71.9 ± 15.5 [53.0-104.6]
BMI, kg/m ²	24.8 ± 3.9 [19.8-34.7]
FEV1, % predicted	90.3 ± 14.1 [66.0-120.0]
Atopy: atopic	9 [56%]
Asthma diagnosis: yes	12 [75%]
Methacholine-responsive: yes	15 [94%]
GSTP1: G allele	11 [69%]
Lymphocytes, K/μL	1.9 ± 0.5 [1.2-2.6]
Monocytes, K/μL	0.43 ± 0.13 [0.20-0.70]
Basophils, K/μL	0.03 ± 0.05 [0.00-0.10]

Except where indicated, data are presented in terms of mean ± standard deviation with range or percentage of total in brackets.

[†]Count [percentage].

not find association between PCs and any granulocyte cell types, as expected since they are removed during PBMC isolation. However, monocyte and lymphocyte counts were positively associated with PCs 7 and 14, and PCs 5, 11, 15, respectively. This indicated that inter-individual difference in blood cell composition was also a source of variance in the dataset. To account for blood composition difference

and avoid confounding between diesel related methylation change and change resulting from blood cell population shifts, we identified CpG sites that were significantly associated with changes in blood count. First we calculated the probe-wise loading values of PCs 5, 7, 11, 14 and 15; for CpG site *i* with loading value *m* for PC *n*, the larger the magnitude of *m*, the closer the methylation pattern of CpG *i* approximates PC *n*. Using cutoffs of +/-3SD, we found 19250 sites that were potentially associated with blood cell counts. By regressing DNA methylation measurements at these sites against lymphocyte and monocyte cell counts, we identified 11378 sites that were significantly associated with blood cell counts (multiple testing correction by the q-value method; FDR =10%). All hits reported to be associated with DE in this study have been filtered to exclude these sites associated with blood cell composition.

A lingering concern with DNA methylation time-series data is that there would an influence of time, stochastic or biological, that affects measurements. We did not find association of time (0 hr to 30 hr) with any PC, indicating that DNA methylation did not vary as a result of measurements taken across the three time points.

Having controlled for potentially confounding variables, we next sought to identify CpG sites associated with DE exposure. We assessed three different scenarios of DE-induced changes assuming baseline measurements at FA-0 hr, FA-6 hr, FA-30 hr, and DE-0 hr (serving as pre-exposure baseline), and tested for association between these scenarios and principle component scores using Wilcoxon's ranked sum test. Scenario-1 hypothesized that methylation changes are detectable 6 hr post DE-exposure, but diminish at 30 hr post-exposure (thus, it compared DE6hr against non-DE6hr). Scenario-2 hypothesized that changes are detectable 30 hr post-exposure (thus, it compared DE30hr against non-

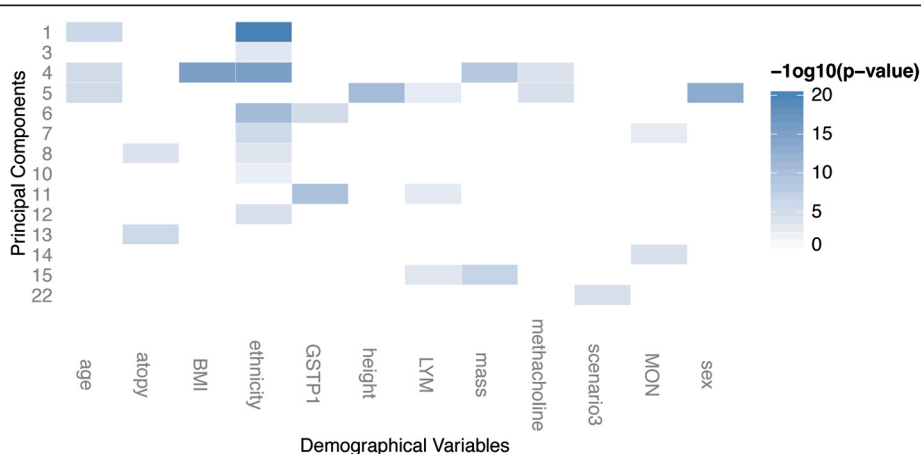


Figure 1 Principal components were associated with demographics and biological variables. For principal components 1-22, only those with associations were included. Colors correspond to p-values where darker colors indicate higher association, and lighter colors indicate lower association. Abbreviations are as follows: basophils (BAS), lymphocytes (LYM), monocytes (MON).

DE30hr). Scenario-3 hypothesized that changes are detectable at 6 hr and persist at 30 hr post-exposure (thus, it compared DE6hr&30 hr against non-DE6hr&30 hr). Scenario-3 was significantly correlated with PC22, which accounted for 0.6% of the total variance. Importantly, PC22 was not associated with any experimental confounds such as demographic factors and blood cell counts. To further examine the pattern of variation underlying PC22, we derived the PC22 scores, which could be used to interpret the correlation between that PC and variable Scenario-3. By ordering the PC22 scores into DE6hr&30 hr versus non-DE6hr&30 hr, we observed that DE6hr&30 hr samples exhibited mostly positive scores with a mean of 0.048, whereas baseline samples exhibited mostly negative scores with a mean

of -0.024. This observation was statistically substantiated by a two-sample t-test (p -value = 1.6×10^{-4}) (Figure 2A). Next, we identified DE-associated CpG sites by calculating the probe-wise loading values of PC22 (Figure 2B). Using cutoffs of ± 3 SD, we found 2827 such sites. Functional analysis of these probes using DAVID revealed enrichment of genes involved in regulation of protein kinase and *NFkB* pathways (enrichment score of 3.01; Additional file 3: Table S1) [34].

DE-associated changes were found in genes relevant to allergic disease

The deviation from random of the raw p -value distributions showed that the post-DE (6 hr&30 hr) versus pre-DE (0 hr) comparison demonstrated high association with

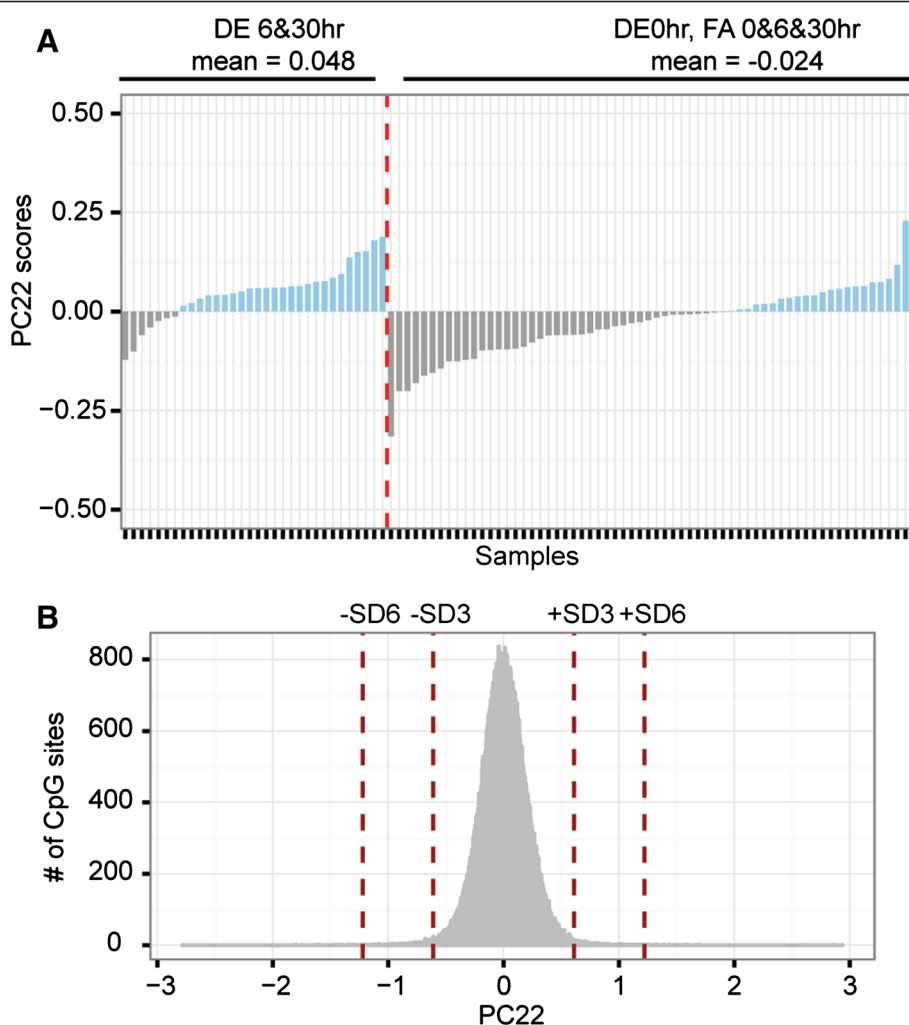
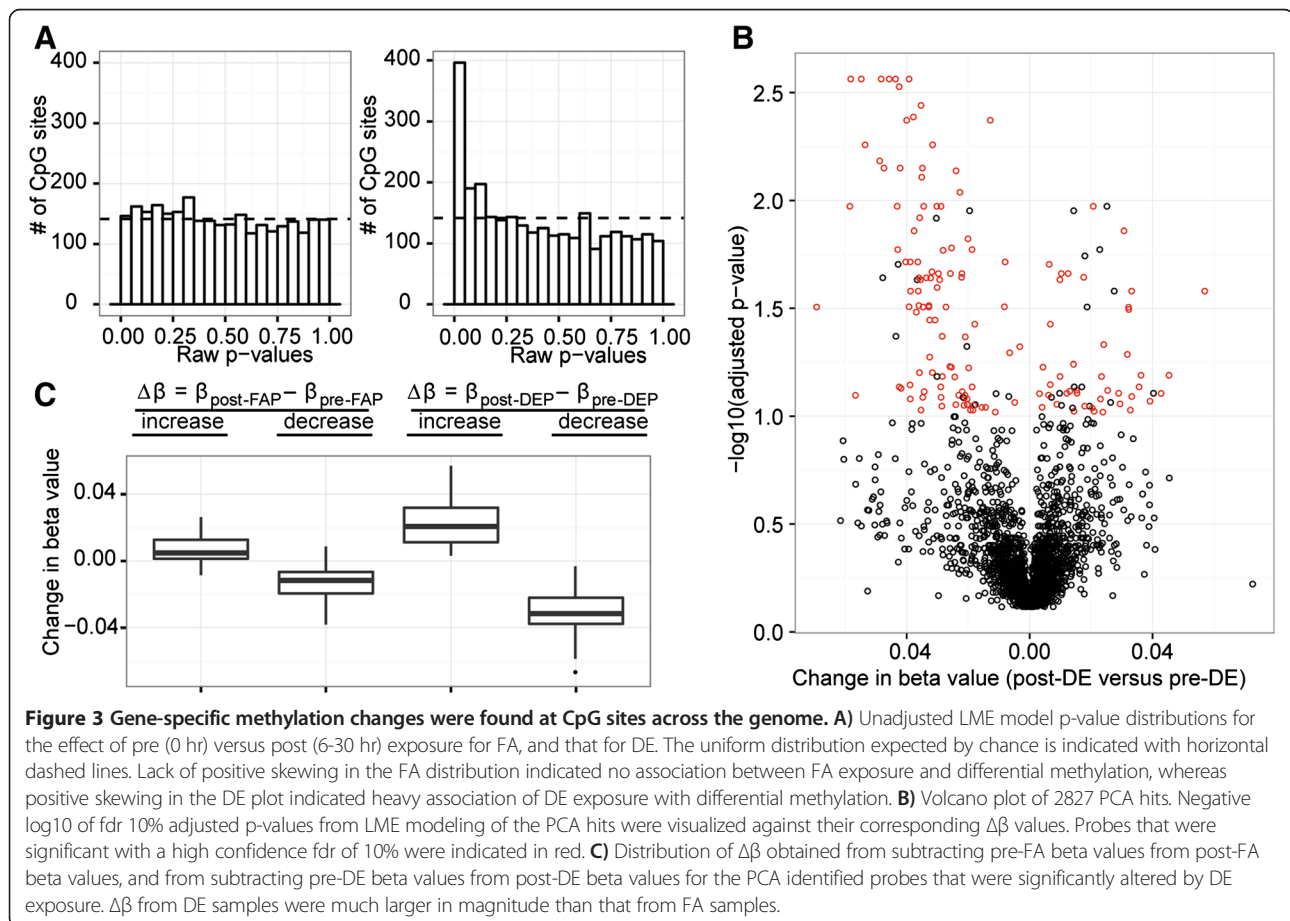


Figure 2 Diesel exhaust-associated exposure patterns were captured in principal component 22. A) PC22 was associated with model-3 (DE6&30 hr vs. non-DE6&30 hr). Samples were sorted according to DE6&30 hr samples versus non-DE6&30 hr baseline samples and their PC22 associated principal component scores were displayed. There was a visible pattern indicative of DE-exposure with the DE6&30 hr samples showing mostly positive scores, and the baseline samples showing mostly negative scores. **B)** For each probe in the dataset, its loading value in association with PC22 was calculated. The ± 3 SD and ± 6 SD positions are marked by vertical dashed lines. And the probes harboring the largest contribution to diesel exposure methylation pattern were selected for using the ± 3 SD cut off, resulting in 2827 hits.

DNA methylation change, suggesting that they were more correlated than that expected by chance, while the same association was not found for post-FA versus pre-FA (Figure 3A). Consistent with the pronounced skewing of the p-value distributions, 170 differentially methylated positions (DMPs) were significant for DE exposure (after multiple testing correction using high confidence FDR of 10%), but not FA exposure. Out of these 170 sites, 25 were previously identified to be significantly associated with blood cell counts; these sites were removed from proceeding analysis, leaving 145 DMPs that demonstrated significant change in methylation as a result of exposure to DE. Specifically, 102 out of the 145 DMPs decreased in methylation (Additional file 3: Table S2a), while 43 sites increased in methylation (Figure 3B and C, Additional file 3: Table S2b). We calculated the average beta value change ($\Delta\beta$) between post-exposure and pre-exposure for the DMPs, and we found 1 probe showing increase and 6 probes showing decrease in DNA methylation of greater than 5%. Interestingly, 1 out of 8 *GSTP1* sites (cg09038676) were among the significantly correlated loci. For *GSTP1*, we genotyped subjects for the A->G substitution at position 313 of the *GSTP1* gene, and we used this information to stratify the samples in an

attempt to determine whether the difference in magnitude of DNA methylation change was related to genotype. As expected, at baseline under the FA condition, DNA methylation levels at cg09038678 for subjects with the G allele were significantly different from that of subjects with the A allele (two-sample t-test p-value = 0.00024). Furthermore, individuals with the A allele showed significant change pre- and post- DE exposure, but not individuals with the G allele (see Additional file 4: Figure S3 in online data supplement).

Lastly, we tested whether DE-related effects carried over to the subsequent FA exposure. Hypothetically, if carryover effects were present then probes found to have increased methylation would show higher mean FA methylation in subjects exposed to diesel first (vice versa for probes found to have decreased methylation). We compared FA mean methylation values in participants who were exposed to DE before FA by first separating the 145 DMPs into ones with decreased and increased methylation in response to DE, then separating the groups further according to exposure order (see Additional file 5: Figure S4 in online data supplement). Neither visual inspection of the graphs nor Welch's two-sample t-test did support any carryover effect



(p -value > 0.05), consistent with our previous assessment of effective binding [35].

Alu and LINE1 CpG sites exhibited methylation change post DE exposure

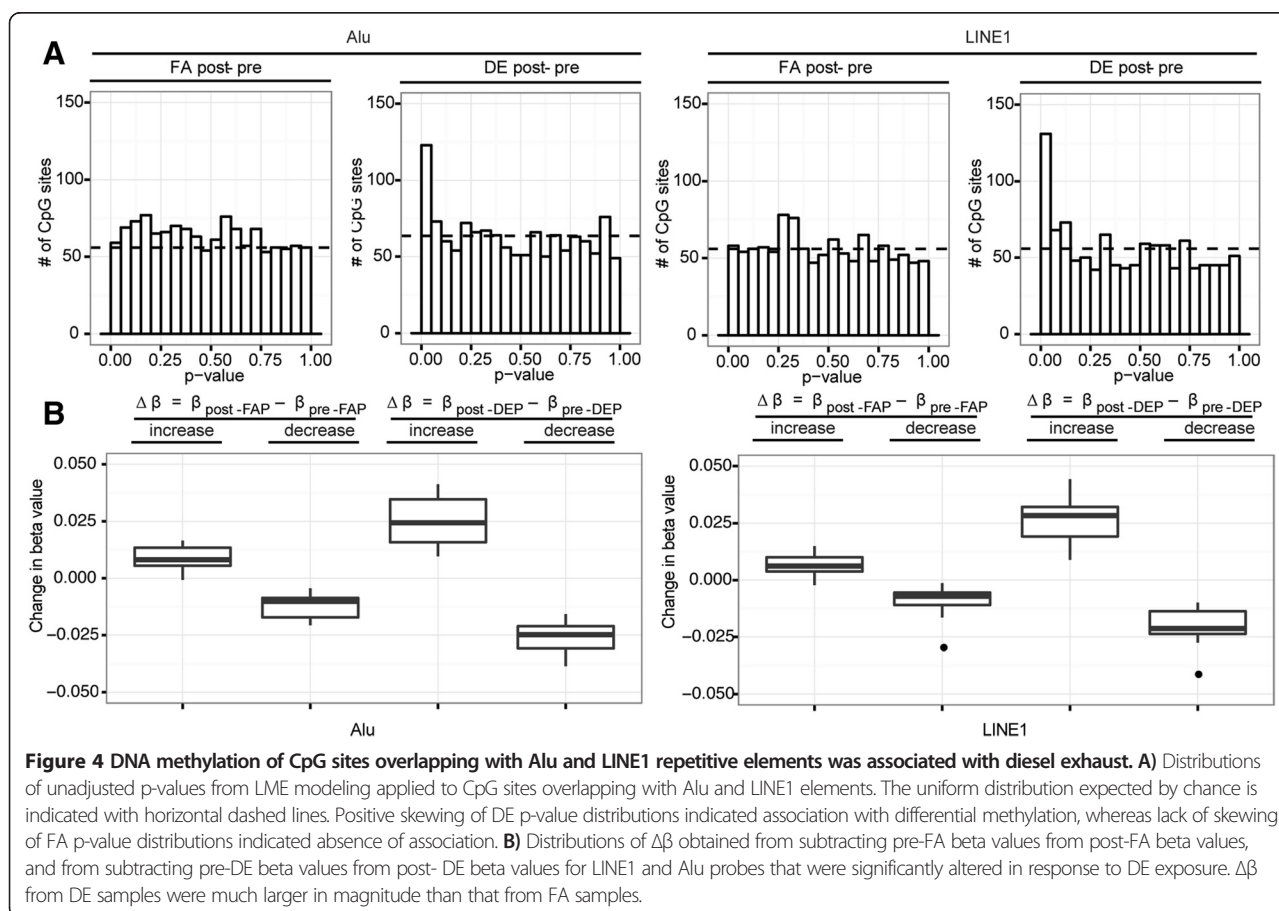
To investigate the impact of DE-exposure on the methylation of Alu and LINE1, we identified *in silico* all probe positions on the array that shared a larger than 15 base-pairs overlap with Alu or LINE1 repetitive elements in the genome. Applying LME modeling to these CpG sites, both Alu and LINE1 elements exhibited changes in DNA methylation as a result of DE exposure, and no changes as a result of FA exposure (Figure 4). 25/1271 (2%) and 31/1118 (3%) DMPs (CpG sites significantly associated with blood cell counts were not considered DMPs) were identified for Alu and LINE1, respectively (Additional file 3: Table S2c, Table S2d). Out of the 25 Alu DMPs, 12 increased in methylation while 13 decreased in methylation after DE exposure; similarly for the 31 LINE1 DMPs, 13 increased while 18 decreased in DNA methylation after exposure. The significant loci found for Alu and LINE1 elements did not overlap with the 145 PCA DMPs, since none of their PCA loading values made the ± 3 SD cutoff.

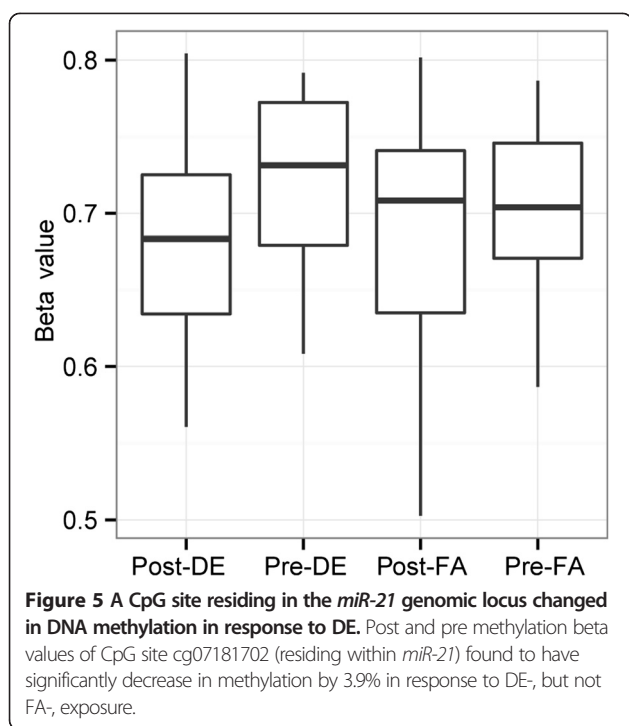
Mir21 showed decrease in methylation upon DE exposure

Recently we identified changes in expression in peripheral blood of four microRNAs (*miR-21*, *miR-30e*, *miR-215* and *miR-144*) in response to DE exposure in the same subjects that were involved in this study [19]. To determine whether differential methylation could be a mediator for the expression of these four microRNAs, we localized a total of 7 probes on the array that overlapped with the genomic positions of these microRNAs. A probe residing within *miR-21* (cg07181702), which demonstrated increase in expression in response to DE, showed significant decrease in methylation in this study (and absence of significant change with FA): cg07181702 decreased in methylation by 3.9% in response to DE exposure (significant with 10% FDR) (Figure 5)[19].

Discussion

Inhaled air pollution including emissions from diesel engines have been associated with a host of cardiovascular and respiratory diseases, imparting a significant strain on public health [3,9]. Data presented here on DNA methylation changes in response to short-term exposure to DE demonstrated a potential epigenetic mechanism





for biological responses to DE exposure. PCA enabled detection of DE signals from subject-level noise generated by demographic variables such as age, ethnicity, and BMI, and focused the investigation onto a smaller list of pertinent CpG sites. Furthermore, the DE-associated methylation changes in LINE1 and Alu CpG sites corroborated with previous research demonstrating sensitivity of repetitive elements to environmental exposure [16,29]. Lastly, methylation change was also observed for *miR-21*, a microRNA associated with oxidative stress and allergic inflammation. Collectively, these results were consistent with our hypothesis that epigenetics serves as a potential avenue through which exposure to air pollution impacts biological systems.

Air pollution has long been linked to diseases including asthma and cardiovascular issues [1,3,32,33]. What is still lacking in our understanding is defining the biological pathways that may be either a mediator or a consequence of the association. DE is a major source of fine particles that have been shown to impact gene expression, and change DNA methylation at inflammation associated sites [16,28,29,31]. Our results support previous studies that have found an association between PM exposure and decrease in DNA methylation of target promoter, candidate genes and repetitive elements [16,29,30]. We suggest that this could be explained through the effects of DE on the dynamics between DNA and DNA methyltransferases. First, ROS-induced oxidative damage to methyl-CpG binding protein recognition sequences could have inhibited the capacity for DNA methyltransferase to bind and

methylate DNA [36]. Secondly, decrease in saturation of the DNA methyltransferase enzymes could potentially come into play, since studies have shown a dependent relationship between exposure to particles and decrease in mRNA transcripts of the DNA methyltransferases *Dnmt1*, *Dnmt3a*, *Dnmt3b* in mouse macrophages [37]. Decreased mRNA transcripts could lower the cellular concentration of methyltransferases and as a result decrease the methylation level of CpG sites. Both of these scenarios could be part of the mechanism leading to loss of methylation that partially explains why the majority of the gene-specific sites found here showed decrease in methylation.

DAVID functional enrichment analysis of the 2827 DE-associated probes identified by PCA revealed enrichment of *NFkB*-related functions. *NFkB* is a redox-related transcription factor that activates a pro-inflammatory response to ROS induced oxidative stress [38-41]. *NFkB* has been shown to activate in response to DE exposure, increasing the downstream expression of inflammatory cytokines [42]. Besides *NFkB*, we also discovered enrichment of the MAPK protein kinase pathway. Previous studies have shown that MAP kinase is a stress-activated kinase pathway that can be induced by DE exposure [12,43]. Given the proposed role of DNA methylation as a mediator between environment and gene expression, the observations made here further demonstrated that a tight circuitry exists between transcriptional pathways known to be involved in response to DE and DNA methylation changes elicited by DE exposure.

Results from PCA followed by regression modeling in this study revealed that in agreement with our hypothesis, DE was associated with changes in DNA methylation at genes that are known to be associated with inflammation and oxidative stress, most notably *GSTP1* [44,45]. Studies investigating the effects of *GSTP1* gene polymorphism found that subjects homozygous for the *GSTP1* G allele have lower functional levels of the enzyme, and thus are at increased risk for oxidative stress, lung cancer, and asthma [44,46,47]. Since the *GSTP1* associated probe (cg09038676) was located in the body of the *GSTP1* gene, then its increase in DNA methylation would likely to be associated with increase in *GSTP1* transcription, resulting in higher levels of the enzyme to combat the effects of oxidative stress. Interestingly, this effect was most profound for subjects with at least one A allele, suggesting that these individuals are less susceptible to the negative effects of DE exposure than individuals homozygous for the G allele, consistent with the literature [48]. Non-asthmatic controls were not included in this study, thus we cannot conclude that the changes observed are specific to asthmatics, but our finding bolsters mechanistic plausibility nonetheless. Furthermore, the significant baseline (FA) methylation difference between subject with and without the *GSTP1*

G allele substitution that cg09038676 could have been a methylation associated SNP; however, due to the fact that only 5 of 16 subjects had an A allele, we were not able to quantitatively assess this possibility.

Besides *GSTP1*, we also found decreased methylation in a probe residing within the body of *HDAC9* (cg24458314), a class IIa histone deacetylase [49]. A study involving regulatory T (Treg) cells in mice showed that absence of *HDAC9* enhances the suppressive ability of Treg cells, resulting in decreased immune responsiveness and inflammation [49]. Changes in DNA methylation of *HDAC9* such as we observed could impact downstream gene expression that then modifies the allergic airway inflammation in response to DE exposure. Considering that Tregs constitute only a small fraction of PBMCs, we speculate this small change might indicate a profound effect on the Treg population; however, a more concrete conclusion could not be reached without independent examination of Treg cell-specific methylation.

Lastly, we discovered that cg05094429, which resides in the promoter region of *CCR6* gene, decreased in methylation after exposure to DE. *CCR6* is expressed by both Treg and Th17 cells, and it is a key regulator of the migration of these cells to sites of inflammation [50]. Lack of this protein in Th17 cells hampers the recruitment of both Th17 and Tregs [50]. Thus it is possible that decreased methylation of *CCR6* in the promoter region could have resulted in increased expression of *CCR6*, eventually leading to increased presence of Th17 and Treg cells responding to DE induced inflammation. It should be noted an effect of ambient particulate matter on DNA methylation patterns has been previously documented in numerous studies [16,28-31]. Therefore, despite the fact that most changes found here were small in magnitude, the overall findings reported here demonstrated an effect of DE that is concordant with existing observations and is impactful on the genomic scale.

Repetitive elements comprise of half of the genome and, under normal conditions, harbor higher DNA methylation in comparison to the rest of the genome [51]. Repetitive elements are activated during cellular stress, which in this case could be elicited by exposure to DE [52]. In accordance with past research on effects of particulate matter, we found both increase and decrease in Alu and LINE1 methylation after DE exposures [37]. In some cases, demethylation of Alu and LINE1 increases genomic instability, which could mean that DE exposure predisposes some cells to genomic rearrangements [53-55]. Furthermore, repeat elements also impact adjacent sequences; they can propagate the spread of DNA methylation to nearby sequences or serve as insulators [56,57]. A recent study has shown that sequences close to a 3' repeat element demonstrated better methylation stability [58]. Therefore, it is conceivable that disruption of repetitive element DNA

methylation due to DE exposure could be associated with genomic rearrangement as well as affecting methylation stability of nearby sequences causing changes in gene expression.

In this study we also examined the effects of DE on methylation of microRNAs, which are a class of small noncoding RNAs with functions in post-translational regulation of expressed genes that are important mediators of cellular processes [59]. Pollutants can cause microRNA dysregulation, which could lead to lung diseases and inflammation [19]. In particular, *miR-21* is involved in oxidative stress and allergic inflammation, and has been shown to be up-regulated in asthma [19,60-62]. Our results showing that DNA methylation was also altered after DE build upon a previous study on the same individuals that reported changes in *miR-21* expression upon DE exposure [19], though this observation was associative and we cannot conclude that the changes in methylation are directly responsible for the changes in expression.

Lastly we were able to demonstrate that DE-associated changes lasted at least 30 hr post-exposure. This observation was consistent with reports showing that methylation changes elicited by environmental exposure could persist for days in the absence of persistent triggers, though the precise dynamics of this in the context of air pollution requires further study. These results should be interpreted in light of the limited sample size, lack of another dataset sufficiently similar to attempt validation, and the systemic nature of the tissue investigated. Nevertheless, this investigation presents a novel approach to analyzing the association between PM and genome-wide DNA methylation, and although we did not observe residual effects in the subjects after 2 weeks, long-term or repeated exposures to DE may lead to accumulative effects. Future studies could build upon the approach presented here to investigate similar events in airway tissues.

Materials and methods

Study demographics

Sixteen participants were recruited at the Air Pollution Exposure Laboratory (APEL) in Vancouver, British Columbia, Canada. Written consent was obtained from all subjects, and the protocol was approved by the institutional review board for human studies at the University of British Columbia (H08-02288). Participants were 19- to 35-year-old nonsmokers who had had physician-diagnosed asthma for at least 1 year and/ or a methacholine challenge with ≤ 8 mg/mL in terms of the PC20, a provocative concentration of methacholine that induces a 20% fall in forced expiratory volume in 1 sec (FEV1). All participants were stable in terms of asthma symptoms [assessed by the asthma control questionnaire] and were free of respiratory infections for 4 weeks prior to and during the study period [63]. The participants were free from current use of inhaled

corticosteroids, regular use of bronchodilator, and use of vitamin A, C, or E supplements. Throughout the study, participants were asked to withhold long-acting β 2-agonists 48 hr prior to spirometry, short-acting β 2-agonists 8 hr prior to spirometry, and caffeine 4 hr prior to methacholine challenges. The participants maintained a stable diet, including intake of cruciferous vegetables, over the course of the study; thus, in the context of the crossover design, diet was not considered confounding.

Exposure design and procedures

Details of the exposure were previously documented [64]. This study followed a double-blind, crossover design in which each subject was subjected to either filtered air (FA) or diesel exhaust (nomimally, 300 $\mu\text{g}/\text{m}^3$ $\text{PM}_{2.5}$) for 2 hours on two separate occasions at least two weeks apart. The DE concentration was, on average, 301 μg $\text{PM}_{2.5}/\text{m}^3$ (SD of 16 $\mu\text{g}/\text{m}^3$). The exposure booth temperature was maintained at 20 degrees Celcius and the relative humidity at 40 percent for all exposures. The sequence of filtered air or diesel exhaust exposure was randomized and counterbalanced, and exposures took place within the same season for a given subject. During exposures, subjects alternated between light exercise (15 min) and rest (45 min) on a stationary bicycle. The wattage of the stationary bike was calibrated in each individual to a minute ventilation of 15 L \cdot min⁻¹ \cdot m² body surface area.

Blood was collected in EDTA and Vacutainer® CPT™ tubes (BD Biosciences, Franklin Lakes, NJ) immediately before exposure as well as 6 and 30 hours after exposure onset and processed within 4 hours. CBCs were performed on whole blood prior to PBMC isolation. PBMC separation was performed following the protocol suggested by the manufacturer, after which PBMCs were stored at -80°C for analysis. Each subject was assessed for *GSTP1* rs1695 SNP genotype by PCR-Restriction Fragment Length Polymorphism using DNA isolated from whole blood.

DNA methylation analysis

All procedures conducted using commercially available kits were done so following the manufacturers' protocol. Two μg of genomic DNA per sample was extracted from PBMCs using the DNeasy Blood & Tissue Kit (Qiagen, Valencia, CA, USA). One μg of the purified DNA was then bisulfite-converted using the EZ-DNA methylation kit (Zymo Research, Orange, CA, USA), which changed epigenetic data into sequence-based data by the selective conversion of unmethylated cytosines to thymidines. Bisulfite-converted DNA was assessed for concentration and quality using the NanoDrop, and 4 μL of the conversion product was used for genome-wide DNA methylation evaluation at over 485,000 CpG sites using the Illumina Infinium HumanMethylation450 BeadChip array in house as described before [65].

Array quality control and normalization

A total of 16 subjects were included in this study, distributed randomly across two chips. There were 6 samples for each subject (3 DE and 3 FA), making a total of 96 samples. Illumina GenomeStudio software was used to interpret array output, and signalA, signal, and probe intensity were exported into R for further processing and analysis. For each sample, probes with one or more samples that had undesirable detection p-values (p-value > 0.01) or with one or more missing measurements were removed (58482 sites). Then probes residing on the X or Y chromosome were also removed to control for gender-derived differences in the array (11648 sites). Lastly, probes that cross-hybridize to somatic sites or to sites on the X or Y chromosomes, as well as probes that possibly reside at SNP sites (as defined by Illumina annotation or independent re-annotation) were removed (52042 sites), regardless of their allele frequency in the population, leaving 363340 probes and 96 samples from 16 subjects for analysis [66]. Such stringent filtration method was applied to ensure that methylation measurements of the CpG sites investigated in this study were most representative of the larger population.

Chip to chip color bias correction was performed in R using the built-in color-correction and background subtraction, and quantile normalization functions of the lumi package with default settings. Finally, we applied peak-based correction method which has been reported to improve data accuracy and reproducibility. Furthermore, peak based correction improves detection rates of differentially methylation at CpG sites that would otherwise have been missed [67-69]. Two distinct values of DNA methylation were calculated, beta-values and M-values. Beta-value has a range of 0 to 1 and approximately represents percent methylation. M-values are log transformation of beta-values, and are more statistically robust. Thus all statistical analyses were performed using M values while visualization and discussion were presented using beta-values. The data obtained in this study has been deposited in the Gene Expression Omnibus repository under the accession number GSE56553.

Identification of Alu, LINE1, and microRNA CpG sites

To identify probes overlapping with Alu and LINE1 repetitive elements, the repeat element track was downloaded from the University of California Santa Cruz (UCSC) Genome Browser (<http://genome.ucsc.edu>), and probes having at least 15 base pairs (bp) overlap with Alu or LINE1 elements in the genome were identified; given that the average Illumina probe is 50 bp in length, this coverage would ensure that at least 30% of the probes resides within the repetitive elements. Furthermore, after testing multiple cutoffs, we found that a cutoff of 15 bp gave the desirable number of Alu or LINE1 associated probes to represent coverage of Alu and LINE1 elements

across the genome. A total of 7 probes overlapping with microRNAs *miR-21*, *miR-30e*, *miR-215* and *miR-144* were also identified and used in subsequent analysis.

Principal component analysis

Mean normalization was conducted across samples, and the `prcomp` function from the R stats package was used to perform principal component analysis on the dataset. The 0th principal component (PC) accounted for the difference in global probe intensity from one probe to the next and thus was not considered. To reduce the number of random variables under consideration, tests for association between experimental variable and PCs were carried out. Specifically ANOVA was used for nominal variables (time, subject, ethnicity, chip number, location on chip), spearman correlation was used for continuous variables (Height in cm, mass in kg, BMI, FEV1, age), and Wilcoxon-ranked sum test was used for dichotomous variables (FA vs. DE, DE hr6 vs. non-DEhr6, DEhr30 vs. non-DEhr30, DEhr6&30 vs. non-DEhr6&30, sex, atopy, asthma, methylation, *GSTP1* genotype). To control for type II error, Storey's *q*value method was used with false discovery cutoff of 0.1 (high confidence) [70].

DAVID functional enrichment analysis

UCSC refgene accession-IDs corresponding to the 363340 CpG sites involved in this analysis was used as background for DAVID GO analysis (<http://david.abcc.ncifcrf.gov/>) [34]. The 2827 CpG sites were tested for enrichment of GO sub-categories biological process and cellular component. Functional enrichment scores of larger than 1.3 were considered significant.

Linear regression modeling:

The following linear regression model was applied to the 11378 lymphocyte and monocyte-count associated CpG sites identified by principal component analysis.

$$Y_{ij} = \beta_0 + \beta_1(\text{Lymphocyte}) + \beta_2(\text{Monocyte}) + \varepsilon_{ij}$$

This model was used to identify sites that were significantly associated with blood cell counts. Y_{ij} represented the measured methylation value of individual i at CpG site j . β_0 represented the overall intercept, β_1 represented the effect of lymphocyte count on methylation value, β_2 represented the effect of monocyte count, and ε_{ij} represented the error term.

Linear mixed effects modeling

The following linear mixed effects model was applied to principal component analysis identified DE-associated CpG sites, as well as sites overlapping with Alu, LINE1, and microRNA sites of interest using the `lme` function from the `nlme` package [71]. Since PC22 was associated

with only diesel exposure, and it did not show confounding with any covariates, only exposure associated variables needed to be included in this model.

$$Y_{ijkl} = \beta_0 + \mu_i + \beta_1(\text{Exposure}) + \beta_2(\text{Time}) + \beta_3(\text{Exposure} \times \text{Time}) + \varepsilon_{ijkl}$$

The model was used to test for change in methylation due to diesel exhaust exposure, assuming random intercept according to subject. Y_{ijkl} represented the measured methylation value of individual i , at CpG site j , exposure k and time l . β_0 represented the overall intercept, μ_i represented the random intercept of individual i , β_1 represented the main effects of exposure, β_2 represented the main effects of time (pre-exposure 0 hr vs. post exposure 6&30 hr), and β_3 represented the interaction effect between exposure and time, and lastly, ε_{ijkl} represented the error term. This model was used to identify probes that changed in methylation as a result of exposure to DE but not FA.

Additional files

Additional file 1: Figure S1. Percent variance within the dataset accounted for by the first 22 principal components (0th principal component was disregarded since it is focused on probe offsets).

Additional file 2: Figure S2. Heatmap showing correlation among demographic variables and differential cell counts (please see Table 1 for units). Red indicates negative Pearson's correlation coefficient of -0.5. Green indicates positive Pearson's correlation coefficient of 1.0. Abbreviations are as follows: basophils (BAS), lymphocytes (LYM), monocytes (MON).

Additional file 3: Table S1. Top 4 DAVID functional annotation clusters. Enrichment of cellular component, biological process, and molecular function in the 2827 PCA filtered probes was calculated using all probes involved in this analysis as the background. **Table S2.** CpG sites found to have significant decrease in methylation as a result of DE exposure through LME modeling. For each probe, $\Delta\beta(\text{FA6\&30hr-FA0hr})$ represents the change in beta value from pre-FA exposure at 0hr and post-FA exposure at 6 and 30hr. $\Delta\beta(\text{DE6\&30hr-DE0hr})$ represents the change in beta value from pre-DE exposure to post-DE exposure. Closest TSS gene name indicates the gene name of the closest transcription start site. UCSC RefGene name indicates the name of the gene at which the probe is located. UCSC refgene group indicates the genomic region at which the probe is located. **a)** Sites with decrease in methylation were mined from the 2827 probes identified to be associated with DE exposure patterns. Sites were ordered from largest $\Delta\beta$ to smallest $\Delta\beta$. **b)** Sites with increase in methylation were mined from the 2827 probes identified to be associated with DE exposure patterns. Sites were ordered from largest $\Delta\beta$ to smallest $\Delta\beta$. **c)** Sites were mined from the 1118 probes with $\geq 15\text{bp}$ overlap with LINE1 elements in the genome. Sites were ordered from largest decrease in methylation to largest increase in methylation. **d)** Sites were mined from the 1271 probes with $\geq 15\text{bp}$ overlap with Alu elements in the genome. Sites were ordered from largest decrease in methylation to largest increase in methylation.

Additional file 4: Figure S3. Changes in methylation of the *GSTP1* probe cg09038676 in response to DE-exposure stratified between subjects with the A allele and those with the G allele.

Additional file 5: Figure S4. Mean beta value of exposure to FA at 0hr, 6hr, and 30hr for the 145 probes found to be significant for DE exposure but not for FA exposure. The beta value differences were first stratified between probes with decreased and increased methylation in response to DE, and then further divided between subjects who were exposed to DE first and those who were exposed to FA first.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

Acquisition of data: FS. Conception and design of data analysis and interpretation of results: RJ. Examination of analysis and interpretation of results: RJ, MJJ, MSK, CC. Drafting and revision of content: RJ. Revision of content and final approval: RJ, MJJ, FS, MSK, CC. All authors read and approved the final manuscript.

Acknowledgments

We thank Dr. Andrea Baccarelli for generous and helpful insight into our work during multiple discussions over recent years. We also thank Lucia Lam for excellent technical assistance.

This work was supported by grants from the AllerGen NCE National Centres of Excellence to MSK and CC. MJJ was supported by a Mining for Miracles fellowship from the Child and Family Research Institute. CC was supported by a New Investigator award from the Canadian Institutes of Health Research and by a Career Investigator award from the Michael Smith Foundation for Health Research. MSK is a Senior Fellow of the Canadian Institute for Advanced Research.

Author details

¹Centre for Molecular Medicine and Therapeutics, University of British Columbia, 950 west 28th Avenue, Vancouver V5Z4H4, Canada. ²Air Pollution Exposure Laboratory, Chan-Yeung Centre for Occupational and Environmental Lung Disease, Department of Medicine, Division of Respiratory Medicine, University of British Columbia, 2775 Laurel Street, Vancouver, British Columbia V5Z1L9, Canada. ³Human Early Learning Partnership, School of Population and Public Health, University of British Columbia, 2206 East Mall, Vancouver, British Columbia V6T1Z3, Canada.

Received: 18 July 2014 Accepted: 24 November 2014

Published online: 09 December 2014

References

- Zanobetti A, Schwartz J, Dockery DW: **Airborne particles are a risk factor for hospital admissions for heart and lung disease.** *Environ Health Perspect* 2000, **108**:1071–1077.
- Brook RD, Franklin B, Cascio W, Hong Y, Howard G, Lipsett M, Luepker R, Mittleman M, Samet J, Smith SC, Tager I: **Expert Panel on Population and Prevention Science of the American Heart Association: Air pollution and cardiovascular disease: a statement for healthcare professionals from the Expert Panel on Population and Prevention Science of the American Heart Association.** *Circulation* 2004, **109**:2655–2671.
- Peng RD, Bell ML, Geyh AS, McDermott A, Zeger SL, Samet JM, Dominici F: **Emergency admissions for cardiovascular and respiratory diseases and the chemical composition of fine particle air pollution.** *Environ Health Perspect* 2009, **117**:957–963.
- Hoffmann B, Moebus S, Mohlenkamp S, Stang A, Lehmann N, Dragano N, Schermund A, Memmesheimer M, Mann K, Erbel R, Jockel KH, Heinz Nixdorf Recall Study Investigative Group: **Residential exposure to traffic is associated with coronary atherosclerosis.** *Circulation* 2007, **116**:489–496.
- Sava F, Carlsten C: **Respiratory health effects of ambient air pollution: an update.** *Clin Chest Med* 2012, **33**:759–769.
- Dockery DW, Pope CA, Xu X, Spengler JD, Ware JH, Fay ME, Ferris BG, Speizer FE: **An association between air pollution and mortality in six U.S. cities.** *N Engl J Med* 1993, **329**:1753–1759.
- Vineis P, Husgafvel-Pursiainen K: **Air pollution and cancer: biomarker studies in human populations.** *Carcinogenesis* 2005, **26**:1846–1855.
- Lepeule J, Laden F, Dockery D, Schwartz J: **Chronic exposure to fine particles and mortality: an extended follow-up of the Harvard Six Cities study from 1974 to 2009.** *Environ Health Perspect* 2012, **120**:965–970.
- Brook RD: **Cardiovascular effects of air pollution.** *Clin Sci* 2008, **115**:175–187.
- Pope CA, Dockery DW: **Health effects of fine particulate air pollution: lines that connect.** *J Air Waste Manag Assoc* 2006, **56**:709–742.
- Laden F, Neas LM, Dockery DW, Schwartz J: **Association of fine particulate matter from different sources with daily mortality in six U.S. cities.** *Environ Health Perspect* 2000, **108**:941–947.
- Hiura TS, Kaszubowski MP, Li N, Nel AE: **Chemicals in diesel exhaust particles generate reactive oxygen radicals and induce apoptosis in macrophages.** *J Immunol* 1999, **163**:5582–5591.
- Kumagai Y, Arimoto T, Shinyashiki M, Shimajo N, Nakai Y, Yoshikawa T, Sagai M: **Generation of reactive oxygen species during interaction of diesel exhaust particle components with NADPH-cytochrome P450 reductase and involvement of the bioactivation in the DNA damage.** *Free Radic Biol Med* 1997, **22**:479–487.
- Li N, Kim S, Wang M, Froines J, Sioutas C, Nel A: **Use of a stratified oxidative stress model to study the biological effects of ambient concentrated and diesel exhaust particulate matter.** *Inhal Toxicol* 2002, **14**:459–486.
- Al-Humadi NH, Siegel PD, Lewis DM, Barger MW, Ma JYC, Weissman DN, Ma JKH: **Alteration of intracellular cysteine and glutathione levels in alveolar macrophages and lymphocytes by diesel exhaust particle exposure.** *Environ Health Perspect* 2002, **110**:349–353.
- Baccarelli A, Wright RO, Bollati V, Tarantini L, Litonjua AA, Suh HH, Zanobetti A, Sparrow D, Vokonas PS, Schwartz J: **Rapid DNA methylation changes after exposure to traffic particles.** *Am J Respir Crit Care Med* 2009, **179**:572–578.
- Takizawa H, Ohtoshi T, Kawasaki S, Abe S, Sugawara I, Nakahara K, Matsushima K, Kudoh S: **Diesel exhaust particles activate human bronchial epithelial cells to express inflammatory mediators in the airways: a review.** *Respirology* 2000, **5**:197–203.
- Tsien A, Diaz-Sanchez D, Ma J, Saxon A: **The organic component of diesel exhaust particles and phenanthrene, a major polyaromatic hydrocarbon constituent, enhances IgE production by IgE-secreting EBV-transformed human B cells in vitro.** *Toxicol Appl Pharmacol* 1997, **142**:256–263.
- Yamamoto M, Singh A, Sava F, Pui M, Tebbutt SJ, Carlsten C: **MicroRNA expression in response to controlled exposure to diesel exhaust: attenuation by the antioxidant N-acetylcysteine in a randomized cross-over study.** *Environ Health Perspect* 2013, **121**:670–675.
- Carlsten C, MacNutt MJ, Zhang Z, Sava F, Pui MM: **Anti-Oxidant N-Acetylcysteine Diminishes Diesel Exhaust-Induced Increased Airway Responsiveness in Person with Airway Hyper-Responsiveness.** *Toxicol Sci* 2014, **139**:479–487.
- Holloway JW, Savarimuthu Francis S, Fong KM, Yang IA: **Genomics and the respiratory effects of air pollution exposure.** *Respirology* 2012, **17**:590–600.
- Diaz-Sanchez D, Proietti L, Polosa R: **Diesel fumes and the rising prevalence of atopy: an urban legend?** *Curr Allergy Asthma Rep* 2003, **3**:146–152.
- Feil R, Fraga MF: **Epigenetics and the environment: emerging patterns and implications.** *N Y Group* 2011, **13**:97–109.
- Bjornsson HT, Sigurdsson MI, Fallin MD, Irizarry RA, Aspelund T, Cui H, Yu W, Rongione MA, Ekström TJ, Harris TB, Launer LJ, Eiriksdottir G, Leppert MF, Sapienza C, Gudnason V, Feinberg AP: **Intra-individual change over time in DNA methylation with familial clustering.** *JAMA* 2008, **299**:2877–2883.
- Feinberg AP: **Phenotypic plasticity and the epigenetics of human disease.** *Nature* 2007, **447**:433–440.
- Wong CC, Caspi A, Williams B, Craig IW, Houts R, Ambler A, Moffitt TE, Mill J: **A longitudinal study of epigenetic variation in twins.** *epigenetics* 2010, **5**:516–526.
- Li Y, Zhu J, Tian G, Li N, Li Q, Ye M, Zheng H, Yu J, Wu H, Sun J, Zhang H, Chen Q, Luo R, Chen M, He Y, Jin X, Zhang Q, Yu C, Zhou G, Sun J, Huang Y, Zheng H, Cao H, Zhou X, Guo S, Hu X, Li X, Kristiansen K, Bolund L, Xu J, Wang W, Yang H, Wang J, Li R, Beck S, Wang J, Zhang X: **The DNA methylome of human peripheral blood mononuclear cells.** *PLoS Biol* 2010, **8**:e1000533.
- Hou L, Zhang X, Zheng Y, Wang S, Dou C, Guo L, Byun H-M, Motta V, McCracken J, Díaz A, Kang C-M, Koutrakis P, Bertazzi PA, Li J, Schwartz J, Baccarelli AA: **Altered methylation in tandem repeat element and elemental component levels in inhalable air particles.** *Environ Mol Mutagen* 2014, **55**:256–265.
- Bellavia A, Urch B, Speck M, Brook RD, Scott JA, Albetti B, Behbod B, North M, Valeri L, Bertazzi PA, Silverman F, Gold D, Baccarelli AA: **DNA hypomethylation, ambient particulate matter, and increased blood pressure: findings from controlled human exposure experiments.** *J Am Heart Assoc* 2013, **2**:e000212.
- Tarantini L, Bonzini M, Apostoli P, Pegoraro V, Bollati V, Marinelli B, Cantone L, Rizzo G, Hou L, Schwartz J, Bertazzi PA, Baccarelli A: **Effects of particulate matter on genomic DNA methylation content and iNOS promoter methylation.** *Environ Health Perspect* 2009, **117**:217–222.
- Hou L, Zhang X, Tarantini L, Nordio F, Bonzini M, Angelici L, Marinelli B, Rizzo G, Cantone L, Apostoli P, Bertazzi PA, Baccarelli A: **Ambient PM exposure and DNA**

- methylation in tumor suppressor genes: a cross-sectional study. *Part Fibre Toxicol* 2011, **8**:25.
32. Bredy TW, Sun YE, Kobor MS: How the epigenome contributes to the development of psychiatric disorders. *Dev Psychobiol* 2010, **52**:331–342.
 33. Lam LL, Emberly E, Fraser HB, Neumann SM, Chen E, Miller GE, Kobor MS: Factors underlying variable DNA methylation in a human community cohort. *Proc Natl Acad Sci U S A* 2012, **109**:17253–17260.
 34. Huang DW, Sherman BT, Lempicki RA: Systematic and integrative analysis of large gene lists using DAVID bioinformatics resources. *Nat Protoc* 2009, **4**:444–57.
 35. Carlsten C, Oron AP, Curtiss H, Jarvis S, Daniell W, Kaufman JD: Symptoms in response to controlled diesel exhaust more closely reflect exposure perception than true exposure. *PLoS One* 2013, **8**:e83573.
 36. Valinluck V, Tsai H-H, Rogstad DK, Burdzy A, Bird A, Sowers LC: Oxidative damage to methyl-CpG sequences inhibits the binding of the methyl-CpG binding domain (MBD) of methyl-CpG binding protein 2 (MeCP2). *Nucleic Acids Res* 2004, **32**:4100–4108.
 37. Miousse IR, Chalbot M-CG, Aykin-Burns N, Wang X, Basnakian A, Kavouras IG, Koturbash I: Epigenetic alterations induced by ambient particulate matter in mouse macrophages. *Environ Mol Mutagen* 2014, **55**:428–435.
 38. Mazzoli-Rocha F, Fernandes S, Einicker-Lamas M, Zin WA: Roles of oxidative stress in signaling and inflammation induced by particulate matter. *Cell Biol Toxicol* 2010, **26**:481–498.
 39. Baeuerle PA, Henkel T: Function and activation of NF-kappaB in the immune system. *Annu Rev Immunol* 1994, **12**:141–179.
 40. Donaldson K, Tran L, Jimenez LA, Duffin R, Newby DE, Mills N, MacNee W, Stone V: Combustion-derived nanoparticles: a review of their toxicology following inhalation exposure. *Part Fibre Toxicol* 2005, **2**:10.
 41. Chan K-K, Shen L, Au W-Y, Yuen H-F, Wong K-Y, Guo T, Wong ML, Shimizu N, Tsuchiyama J, Kwong Y-L, Liang RH, Srivastava G: Interleukin-2 induces NF-kappaB activation through BCL10 and affects its subcellular localization in natural killer lymphoma cells. *J Pathol* 2010, **221**:164–174.
 42. Takizawa H, Ohtoshi T, Kawasaki S, Kohyama T, Desaki M, Kasama T, Kobayashi K, Nakahara K, Yamamoto K, Matsushima K: Diesel exhaust particles induce NF-kB activation in human bronchial epithelial cells in vitro: importance in cytokine transcription. *J Immunol* 1999, **162**:4705–4711.
 43. Hashimoto S, Gon Y, Takeshita I, Matsumoto K, Jibiki I, Takizawa H, Kudoh S, Horie T: Diesel exhaust particles activate p38 MAP kinase to produce interleukin 8 and RANTES by human bronchial epithelial cells and N-acetylcytosteine attenuates p38 MAP kinase activation. *Am J Respir Crit Care Med* 2000, **161**:280–285.
 44. Romieu I, Ramirez-Aguilar M, Sienra-Monge JJ, Moreno-Macias H, del Rio-Navarro BE, David G, Marzec J, Hernandez-Avila M, London S: GSTM1 and GSTP1 and respiratory health in asthmatic children exposed to ozone. *Eur Respir J* 2006, **28**:953–959.
 45. Zimniak P, Nanduri B, Pikula S, Bandorowicz-Pikula J, Singhal SS, Srivastava SK, Awasthi S, Awasthi YC: Naturally occurring human glutathione S-transferase GSTP1-1 isoforms with isoleucine and valine in position 104 differ in enzymic properties. *Eur J Biochem* 1994, **224**:893–899.
 46. Miller DP, De Vivo I, Neuberger D, Wain JC, Lynch TJ, Su L, Christiani DC: Association between self-reported environmental tobacco smoke exposure and lung cancer: modification by GSTP1 polymorphism. *Int J Cancer* 2003, **104**:758–763.
 47. Tamer L, Calikoğlu M, Ates NA, Yildirim H, Ercan B, Saritas E, ÜNLÜ A, Atik U: Glutathione-S-transferase gene polymorphisms (GSTT1, GSTM1, GSTP1) as increased risk factors for asthma. *Respirology* 2004, **9**:493–498.
 48. MacIntyre EA, Brauer M, Melén E, Bauer CP, Bauer M, Berdel D, Bergström A, Brunekreef B, Chan-Yeung M, Klümper C, Fuertes E, Gehring U, Gref A, Heinrich J, Herbarth O, Kerkhof M, Koppelman GH, Kozyrskyj AL, Pershagen G, Postma DS, Thiering E, Tiesler CMT, Carlsten C: TAG Study Group: GSTP1 and TNF Gene variants and associations between air pollution and incident childhood asthma: the traffic, asthma and genetics (TAG) study. *Environ Health Perspect* 2014, **122**:418–424.
 49. de Zoeten EF, Wang L, Sai H, Dillmann WH, Hancock WW: Inhibition of HDAC9 increases T regulatory cell function and prevents colitis in mice. *Gastroenterology* 2010, **138**:583–594.
 50. Yamazaki T, Yang XO, Chung Y, Fukunaga A, Nurieva R, Pappu B, Martin-Orozco N, Kang HS, Ma L, Panopoulos AD, Craig S, Watowich SS, Jetten AM, Tian Q, Dong C: CCR6 regulates the migration of inflammatory and regulatory T cells. *J Immunol* 2008, **181**:8391–8401.
 51. Yang AS, Estéicio MRH, Doshi K, Kondo Y, Tajara EH, Issa J-PJ: A simple method for estimating global DNA methylation using bisulfite PCR of repetitive DNA elements. *Nucleic Acids Res* 2004, **32**:e38.
 52. Lucchinetti E, Feng J, Silva RD, Tolstogon GV, Schaub MC, Schumann GG, Zaugg M: Inhibition of LINE-1 expression in the heart decreases ischemic damage by activation of Akt/PKB signaling. *Physiol Genomics* 2006, **25**:314–324.
 53. Bennett EA, Keller H, Mills RE, Schmidt S, Moran JV, Weichenrieder O, Devine SE: Active Alu retrotransposons in the human genome. *Genome Res* 2008, **18**:1875–1883.
 54. Gaudet F, Hodgson JG, Eden A, Jackson-Grusby L, Dausman J, Gray JW, Leonhardt H, Jaenisch R: Induction of tumors in mice by genomic hypomethylation. *Science* 2003, **300**:489–492.
 55. Cho N-Y, Kim B-H, Choi M, Yoo EJ, Moon KC, Cho Y-M, Kim D, Kang GH: Hypermethylation of CpG island loci and hypomethylation of LINE-1 and Alu repeats in prostate adenocarcinoma and their relationship to clinicopathological features. *J Pathol* 2007, **211**:269–277.
 56. Wang X, Fan J, Liu D, Fu S, Ingvarsson S, Chen H: Spreading of Alu methylation to the promoter of the MLH1 gene in gastrointestinal cancer. *PLoS One* 2011, **6**:e25913.
 57. Lunyak W, Prefontaine GG, Núñez E, Cramer T, Ju B-G, Ohgi KA, Hutt K, Roy R, García-Díaz A, Zhu X, Yung Y, Montoliu L, Glass CK, Rosenfeld MG: Developmentally regulated activation of a SINE B2 repeat as a domain boundary in organogenesis. *Science* 2007, **317**:248–251.
 58. Byun H-M, Nordio F, Coull BA, Tarantini L, Hou L, Bonzini M, Apostoli P, Bertazzi PA, Baccarelli A: Temporal stability of epigenetic markers: sequence characteristics and predictors of short-term DNA methylation variations. *PLoS One* 2012, **7**:e39220.
 59. He L, Hannon GJ: MicroRNAs: small RNAs with a big role in gene regulation. *Nat Rev Genet* 2004, **5**:522–531.
 60. Bollati V, Marinelli B, Apostoli P, Bonzini M, Nordio F, Hoxha M, Pegoraro V, Motta V, Tarantini L, Cantone L, Schwartz J, Bertazzi PA, Baccarelli A: Exposure to metal-rich particulate matter modifies the expression of candidate microRNAs in peripheral blood leukocytes. *Environ Health Perspect* 2010, **118**:763–768.
 61. Cheng Y, Liu X, Zhang S, Lin Y, Yang J, Zhang C: MicroRNA-21 protects against the H(2)O(2)-induced injury on cardiac myocytes via its target gene PDCD4. *J Mol Cell Cardiol* 2009, **47**:5–14.
 62. Lu TX, Munitz A, Rothenberg ME: MicroRNA-21 is up-regulated in allergic airway inflammation and regulates IL-12p35 expression. *J Immunol* 2009, **182**:4994–5002.
 63. Juniper EF, O'byrne PM, Guyatt GH, Ferrie PJ, King DR: Development and validation of a questionnaire to measure asthma control. *Eur Respir J* 1999, **14**(4):902–907.
 64. Birger N, Gould T, Stewart J, Miller MR, Larson T, Carlsten C: The Air Pollution Exposure Laboratory (APEL) for controlled human exposure to diesel exhaust and other inhalants: characterization and comparison to existing facilities. *Inhal Toxicol* 2011, **23**:219–225.
 65. Jones MJ, Farré P, McEwen LM, Macisaac JL, Watt K, Neumann SM, Emberly E, Cynader MS, Virji-Babul N, Kobor MS: Distinct DNA methylation patterns of cognitive impairment and trisomy 21 in Down syndrome. *BMC Med Genomics* 2013, **6**:58.
 66. Price ME, Cotton AM, Lam LL, Farré P, Emberly E, Brown CJ, Robinson WP, Kobor MS: Additional annotation enhances potential for biologically-relevant analysis of the Illumina Infinium HumanMethylation450 BeadChip array. *Epigenetics Chromatin* 2013, **6**:4.
 67. Du P, Kibbe WA, Lin SM: Lumi: a pipeline for processing Illumina microarray. *Bioinformatics* 2008, **24**(13):1547–1548.
 68. R Development Core Team: *R: A language and environment for statistical computing*. Vienna, Austria: R Foundation for Statistical Computing; 2008. ISBN 3-900051-07-0, URL <http://www.R-project.org>.
 69. Dedeurwaerder S, Defrance M, Calonne E, Denis H, Sotiriou C, Fuks F: Evaluation of the Infinium Methylation 450 K technology. *Epigenomics* 2011, **3**:771–784.
 70. Dabney A, Storey JD, Warnes GR: qvalue: Q-value estimation for false discovery rate control. *R package version 1.34.0*.
 71. Pinheiro J, Bates D, DebRoy S, Sarkar D, R Core Team: nlme: Linear and nonlinear mixed effects models. 2014, R package version 3.1-115, URL: <http://CRAN.R-project.org/package=nlme>.

doi:10.1186/s12989-014-0071-3

Cite this article as: Jiang et al.: Short-term diesel exhaust inhalation in a controlled human crossover study is associated with changes in DNA methylation of circulating mononuclear cells in asthmatics. *Particle and Fibre Toxicology* 2014 **11**:71.