

Review Article

Metabolic syndrome, haemostasis and thrombosis

Marie-Christine Alessi, Irène Juhan-Vague

Laboratoire d'Hématologie, Faculté de Médecine, Université de la Méditerranée, Inserm UMR 626, Marseille, France

Summary

The metabolic syndrome (metS), a concurrence of abdominal fat, disturbed glucose and insulin metabolism, dyslipidemia, and hypertension has been strongly associated not only with subsequent development of type 2 diabetes but also with atherothrombosis. The physiopathology of this association is complex. The metS affects the thrombogenicity of circulating blood. Apart from its effect on platelets, a procoagulant and hypofibrinolytic state has been identified; mainly the result of the inflammatory state, dyslipidemia, and liver fat accumulation that accompany the

MetS. Among haemostasis disturbances, the strong rise in the inhibitor of plasminogen activator type I plasma level is the most documented abnormality implicating the participation of the oxidative stress and inflammatory state developed during the metS. Endothelial dysfunction is also a central feature. Moreover, secretion products of fat tissues (adipokines) are now thought to have direct modulating effects on the vascular and the circulating cells. In support of these data, the metS, may predispose not only to atherosclerosis but also to venous thrombosis.

Keywords

Haemostasis, metabolic syndrome, visceral obesity, platelet, coagulation, fibrinolysis, endothelial dysfunction

Thromb Haemost 2008; 99: 995–1000

Introduction

The metabolic syndrome (metS), is a clinical entity of substantial heterogeneous traits represented by the co-occurrence of abdominal fat, impaired glucose tolerance, dyslipidemia (high triglycerides and low high-density lipoprotein [HDL] cholesterol levels) and hypertension. The conference on metS definition of the American Heart Association (1) underlined two additional components that are a proinflammatory and a prothrombotic states and confirmed cardiovascular disease and type 2 diabetes as major clinical outcomes of metS.

Although the prevalence of the components of the metS is increased in obesity (2), it is important to notice that not all obese subjects develop metS, and even non-obese individuals can carry metS.

Pathophysiology of this syndrome mainly involved three ways of thinking: (i) central obesity also known as ectopic fat disease, (ii) insulin resistance, and (iii) a constellation of independent factors (e.g. molecules of hepatic, vascular, and adipose origin) that mediate specific components of the syndrome.

In this review we describe the relationships between the components of the metS and alterations in hemostasis that may predispose to both arterial and venous thrombosis.

Metabolic syndrome and platelet hyperactivity

Increased platelet response is seen in individuals with the metS (3–8) supported by the main features of metS: insulin resistance, dyslipidemia, products of adipose tissue (adipokines) and inflammation.

Insulin resistance refers to resistance to the metabolic and vascular activities of insulin in a variety of cells, including mainly hepatocytes, muscle cells, and adipocytes. Insulin possesses antiplatelet properties (6). Attenuation of platelet response in insulin resistant patients have been connected to a reduced platelet sensitivity to insulin (9). Loss of platelet inhibition by insulin in patients with insulin resistance may be related to loss of insulin-mediated suppression of ADP-induced P2y12 signaling as well as decreased P2y12 inhibition by receptor antagonists (10). This may explain the association between diabetes and resistance to the antiplatelet effects of clopidogrel (11). Other mechanisms have been documented to explain platelet hyperactivity as an elevated cytosolic Ca(2+) and the increase of oxidative stress, which elicits isoprostane production from arachidonic acid (12).

Hypertriglyceridemia and increased concentration of free fatty acids exert a proaggregating effect *in vitro* (13). Hypo-

Correspondence to:
Marie-christine Alessi
Laboratoire d'Hématologie, Faculté de Médecine
Université de la Méditerranée, Inserm UMR 626
27 Bd Jean Moulin, Marseille, F-13385 France
Tel.: +33 4 91 38 60 48, Fax: +33 4 91 94 23 32
E-mail: marie-christine.alessi@univmed.fr

Footnote:
This review article is the last contribution to the March 2008 Theme issue
in connection with the GTH congress in Wiesbaden, Germany.

Received November 16, 2007
Accepted after major revision April 25, 2008

Prepublished online May 7, 2008
doi:10.1160/TH07-11-0682

HDLaemia also influences platelet aggregation, possibly because HDL opposes the activation properties of low-density lipoprotein (LDL) on platelets (14). The exact mechanisms of these effects are not clearly elucidated.

Adipokines may directly influence platelet function. Plasma leptin reflects mainly adipose tissue mass, and a strong association was found between circulating leptin and insulin resistance (15). Leptin receptors have been identified in the cardiovascular system (16) as well as in platelets. Leptin potentiates the normal response of platelets to adenosine diphosphate and thrombin (17–19). However, four-day leptin infusion only slightly increases platelet aggregation in healthy volunteers (20). Moreover carriers of leptin deficiency exhibited increased platelet aggregation relative to controls, and incubation *in vitro* with leptin did not increase up platelet aggregation (21). Thus, one may argue that factors other than leptin modulate platelet function in extreme obesity or that obesity is associated with impaired platelet sensitivity to leptin, i.e. leptin resistance resembling the reduced sensitivity of platelets to insulin in insulin resistance (22, 23). This needs to be confirmed *in vivo*. The adipokine, adiponectin, is abundantly present in plasma. Its plasma concentration decreases in obesity, type 2 diabetes, and patients with coronary artery disease. In recent years, a great amount of literature has suggested a close connection between adiponectin and each inherent trademark of the metS (24). Results obtained in adiponectin knockout mice indicate that adiponectin acts as an endogenous antithrombotic factor (25). *In-vitro* platelet aggregation induced by low concentration of agonists was enhanced in adiponectin knockout mice, and recombinant adiponectin overcame the enhanced platelet aggregation (25). However, addition of adiponectin did not affect either aggregation or adhesion of human platelets; even at supra-physiological concentrations (26). Lastly platelet, may be activated by the low-grade systemic inflammatory state seen in visceral obesity (27).

Pharmacological inhibition of platelet functions is at the center of the treatment of active cardiovascular disease and in the secondary prevention of cardiovascular events. However, there are divergent views, between guidelines, on which patients with type 2 diabetes require aspirin in the primary prevention strategy as well as on the dose of aspirin (28). In addition, there is no data regarding antiplatelet therapy in patients with metS without diabetes despite prognostically important coronary arterial disease in these patients. This high level of uncertainty will deserve to be precised in randomized clinical trials taking into account the lower potency of antiplatelet therapy described in patients with diabetes (28, 29) and insulin-resistant obesity (30).

Metabolic syndrome and hypercoagulability

Plasma from subjects with the metS formed denser clots compared with subjects free from metS. In addition clot density increased progressively with increasing number of metS components (31). Analysis of clot density in prospective studies is warranted to document the pathogenicity of this haemostasis trait in patients with metS, as stiffer and denser clots were associated with premature cardiovascular disease (32).

Tissue factor (TF), the key initiator of coagulation is widely expressed in atherosclerotic plaques and found in macrophages,

smooth muscle cells, extracellular matrix and acellular lipid-rich core. The blood-borne TF encrypted on the circulating microparticles derived from vascular cells is a marker of vascular injury and a source of procoagulant activity. Evidence indicates that elevated levels of blood-borne or circulating TF has been associated with metS (33) and is a candidate biomarker for future cardiovascular events (34). The elevated TF level may result from various stimulants which accompany the metS such as C-reactive protein, oxidized LDL, tumor growth factor (TGF) beta, angiotensin II, hyperglycemia, and adipocytokines (35). Among them, hyperinsulinemia may be of particular relevance. Adipose and circulating TF are potentiated by insulin administration in obese mice (36, 37) and humans (38), respectively. Despite the important role of TF in initiation of coagulation, the relevance of blood-borne TF for thrombosis in metS deserves to be documented.

In non-diabetic elderly men and women, increased levels of vitamin K-dependent coagulation proteins clustered with dyslipidemia and inflammation, whereas they were not related to anthropometric parameters or arterial pressure nor glucidic metabolism (39). These results may be in favour of a potentiation of hepatic synthesis of vitamin K-dependent proteins during the metS. Liver steatosis could play an important role in this process. Liver fat is highly significantly and linearly correlated with all components of the metS, independent of obesity. Overproduction of coagulation factors in addition to glucose and very low-density lipoprotein (VLDL) by the fatty liver could contribute to the excess risk of cardiovascular disease associated with the metS. In agreement a strong relationship has been reported between circulating levels of vitamin K-dependent proteins and that of the hepatic enzyme gamma glutamyl transferase (40).

Fibrinogen levels associates importantly with metS cluster (41) as factor VIII (FVIII) (42, 43). These elevations has to be brought closer to the inflammatory state that accompanies the metS. Indeed, FVIII circulates as an inactive procofactor in complex with the acute-phase protein von Willebrand factor, which slows down FVIII elimination. The increase in IL-6 levels that accompagnies the metS may be responsible for the slight increase in hepatic synthesis of fibrinogen (44, 45).

It has been reported that dyslipidemia may directly affect activation of coagulation factors. VLDL produced in excess during the metS supports activation of factor VII by the Xa/Va (46, 47) and HDL that levels are diminished during the metS, attenuate the expression of TF and downregulates thrombin generation via the enhancement of the anticoagulant protein C pathway (48). Therefore, the hypoHDLaemia which accompanies the metS could be involved in the thrombotic risk by increasing thrombin generation.

Metabolic syndrome and hypofibrinolysis

Subjects with metS had prolonged clot lysis times compared with those without metS (31), partly due to increased circulating levels of plasminogen activator inhibitor 1 (PAI-1) that is the most important and visible change of the haemostatic system in the metS (49).

Increased concentration of PAI-1 leads to impairment of the removal of thrombi from the vascular system (50) and may in-

fluence the development of atherosclerotic lesions as well (51). In large epidemiological studies, elevated plasma levels of PAI-1 proved to be predictors of myocardial infarction (49). Remarkably, the predictive ability of PAI-1 disappears after adjustment for markers of the metS (52). These results suggest that the presence of abdominal obesity and insulin resistance is a prerequisite for the increased PAI-1 levels in patients at risk of atherothrombosis and have led to the proposal that increased PAI-1 level can be considered as a true component of the metS (53).

The increase in plasma PAI-1 levels associated with abdominal obesity may be attributed to PAI-1 production by ectopic adipose tissues (54–58) and fatty liver (59, 60). Overall these findings suggest that circulating PAI-1 levels reflect fat redistribution and may be considered a biomarker of ectopic fat storage disease, a feature of central obesity.

Tissue expression of PAI-1 is not constitutive but mainly inducible. Many inducers of PAI-1 synthesis during visceral obesity have been identified that may exert their effect locally or more remotely (49). Establishment of inflammation or oxidative stress at the macrophage level as fundamental precursors of PAI-1 overexpression in metS is tempting.

Circulating PAI-1 levels predict development of type 2 diabetes (61–64) and more recently metS (65), suggesting that PAI-1 may be causally related to deterioration of metabolic homeostasis. Three groups found that fat accumulation was prevented in mice lacking PAI-1 in both a nutritionally induced (66, 67) and a genetic (68) murine model of obesity. Results obtained by our group (69–71) followed this direction, showing an effect of pharmacological inhibition of PAI-1 on weight gain and on insulin sensitivity.

In addition, PAI-1 deficiency may exert beneficial effects through improved insulin sensitivity in adipocytes (72). This effect may be mediated through the ability of PAI-1 to impair the cooperation between integrin $\alpha_v\beta_3$ and insulin signalling (73, 74) or to block the deleterious effect of tumor necrosis factor (TNF) on insulin sensitivity (72).

Findings suggest that targeted PAI-1 overexpression in macrophages and adipocytes impairs adipose tissue growth in mice (75), which agrees with the recently described inhibitory effect of PAI-1 on murine adipocyte differentiation (72) not reproduced by another study (76). This finding may, at first glance, appear to be at odds with that obtained in PAI-1-deficient mice, but it must be interpreted in connection with the multiple facets of PAI-1, which render it a serpin that acts locally at various sites and perhaps systemically through endocrine effects. Interestingly, old transgenic mice overexpressing PAI-1 and maintained on a standard fat diet exhibit significantly higher insulinaemia and a tendency toward higher triglyceride levels, despite lower body fat (71). These data are not inconsistent with those obtained in PAI-1-deficient mice and indicate that PAI-1 overexpression might worsen the metabolic profile. This requires confirmation because this deleterious effect was not found in younger transgenic mice fed a diet high in fat (75).

Overall, these data support the concept that PAI inhibition (77) has the potential to reduce obesity and improve insulin sensitivity, and may represent a new therapeutic target. This requires confirmation in different experimental models, and the mechanisms involved should be precisely defined.

Metabolic syndrome and endothelial dysfunction

In healthy conditions insulin promotes glucose disposal and stimulates the endothelial production of nitric oxide (NO), which in turn, through NO-dependent increases in blood flow to skeletal muscle, may account for 25% to 40% of the increase in glucose uptake in response to insulin stimulation (78). A physiologic increment in plasma insulin concentration particularly increases microvascular blood volume, consistent with a mechanism of capillary recruitment (79).

Insulin resistance is characterized by pathway-specific impairment in phosphatidylinositol 3-kinase-dependent signalling, which in endothelium may cause imbalance between production of NO and secretion of endothelin-1, leading to decreased blood flow, which worsens insulin resistance (80–82). Experimental inhibition of phosphatidylinositol 3-kinase with wortmannin not only blocked the ability of insulin to stimulate increased expression of endothelial NO synthase but also increased expression of vascular cellular adhesion molecules-1 and E-selectin, and increased rolling interactions of monocytes with endothelial cells, showing that inhibition of the metabolic branch of insulin signalling leads to an enhanced atherogenic action of insulin in endothelial cells (83).

In parallel with inadequate vasodilatation, in obesity endothelial cells take a proinflammatory phenotype with increased expression of VCAM1, ICAM1, E selectin, a release of microparticles (84) and shedding products, and an increased synthesis and release of the adhesive protein von Willebrand factor which levels correlated with parameters of the metS (42, 85) and inflammatory parameters (42, 86–88). These endothelial disorders may arise at a very early age in obese children (89).

Metabolic syndrome and thrombosis

The metS is a well recognized risk factor of acute cardiovascular disease (90–93). Given the evidence of an hypercoagulability, hypofibrinolysis and endothelial dysfunction in the carriers of the metS there is a rationale to hypothesize that the metS may also predispose patients to develop venous thromboembolism (VTE) (94). In addition VTE was found to be associated with atherosclerosis more frequently than expected (95) leading to several hypotheses among which that of the metS as a common antecedent.

Some individual components of the metabolic syndrome have been associated with VTE mainly dyslipoproteinemia involving high triglyceride (TG) levels, low HDL particles and small LDL particles (96–99). A recent metanalysis assessed the association between cardiovascular risk factors and VTE (100). A total of 63,552 subjects met the inclusion criteria. Compared with control subjects the risk of VTE was 2.33 for obesity and 1.42 for diabetes mellitus. HDL cholesterol was inversely and consistently correlated with VTE, and triglycerides were on average 21 mg/dl higher in patients with VTE than in controls.

Few data have been provided on visceral obesity or the metS considered as a whole. The prospective study of men born in 1913 showed that men with a waist circumference of more than 100 cm had a higher cumulative incidence of VTE than men with a waist circumference less than 100 cm leading to a adjusted

relative risk of 3.92 (101). More recently Ray et al. (102) investigated the association between VTE and features of the metS in a prospective cohort of adults with cardiovascular disease or diabetes and additional risk factor. This cohort, derived from the HOPE-2 randomized clinical trial, enrolled 5,522 subjects older than 55 years and followed for a median of five years. Again elevated waist circumference was significantly associated with VTE.

Two recent small size case-control studies have investigated the association between the metS defined according to NCEP-ATPIII criteria (103) and the occurrence of VTE. In the first study (96) the metS was significantly more common in patients with idiopathic VTE than in controls in uni- and multivariate analysis. Ay et al. (104) found that patients with recurrent VTE had significantly higher body mass index, waist-to-hip ratio and triglyceride levels than controls. The metS was diagnosed in 35% of patients and 20% of controls leading to an adjusted odds ratio of 2.1. Interestingly it was not the presence of a single com-

ponent but rather the constellation of multiple components that is crucial in this association.

Conclusion

The metS is accompanied by important changes in the haemostatic system that may favour the development of thrombosis. Hyperactivity of platelets and hypercoagulability favour platelet and fibrin deposits, and hypofibrinolysis due to the PAI-1 excess prevents their elimination. The increased PAI-1 expression that accompanies abdominal obesity is the most documented abnormality associated with the metS. As PAI-1 could also be directly involved in the physiopathology of obesity, it could represent an original target for preventing both the thrombotic and metabolic risks. Whereas strong epidemiological evidences have established the contribution of metS to cardiovascular disease, better illustration is needed to establish whether metS is a relevant risk factor for VTE.

References

1. Grundy SM, Brewer HB Jr, Cleeman JI, et al. Definition of metabolic syndrome: Report of the National Heart, Lung, and Blood Institute/American Heart Association Circulation 2004; 109: 433–438.
2. Abbasi F, Brown BW Jr, Lamendola C, et al. Relationship between obesity, insulin resistance, and coronary heart disease risk. *J Am Coll Cardiol* 2002; 40: 937–943.
3. Trovati M, Anfossi G. Insulin, insulin resistance and platelet function: similarities with insulin effects on cultured vascular smooth muscle cells. *Diabetologia* 1998; 41: 609–622.
4. Anfossi G, Trovati M. Pathophysiology of platelet resistance to anti-aggregating agents in insulin resistance and type 2 diabetes: implications for anti-aggregating therapy. *Cardiovasc Hematol Agents Med Chem* 2006; 4: 111–128.
5. Davi G, Guagnano MT, Ciabattini G, et al. Platelet activation in obese women: role of inflammation and oxidant stress. *J Am Med Assoc* 2002; 288: 2008–2014.
6. Trovati M, Anfossi G. Influence of insulin and of insulin resistance on platelet and vascular smooth muscle cell function. *J Diabetes Complications* 2002; 16: 35–40.
7. Arteaga RB, Chirinos JA, Soriano AO, et al. Endothelial microparticles and platelet and leukocyte activation in patients with the metabolic syndrome. *Am J Cardiol* 2006; 98: 70–74.
8. Basili S, Pacini G, Guagnano MT, et al. Insulin resistance as a determinant of platelet activation in obese women. *J Am Coll Cardiol* 2006; 48: 2531–2538.
9. Westerbacka J, Yki-Järvinen H, Turpeinen A, et al. Inhibition of platelet-collagen interaction: an in vivo action of insulin abolished by insulin resistance in obesity. *Arterioscler Thromb Vasc Biol* 2002; 22: 167–172.
10. Ferreira IA, Mocking AI, Feijge MA, et al. Platelet inhibition by insulin is absent in type 2 diabetes mellitus. *Arterioscler Thromb Vasc Biol* 2006; 26: 417–422.
11. Angiolillo DJ, Fernandez-Ortiz A, Bernardo E, et al. Platelet function profiles in patients with type 2 diabetes and coronary artery disease on combined aspirin and clopidogrel treatment. *Diabetes* 2005; 54: 2430–2435.
12. Anfossi G, Russo I, Trovati M. Platelet resistance to the anti-aggregating agents in the insulin resistant states *Curr Diabetes Rev* 2006; 2: 409–430.
13. Englyst NA, Taube JM, Aitman TJ, et al. A novel role for CD36 in VLDL-enhanced platelet activation. *Diabetes* 2003; 52: 1248–1255.
14. Korporaal SJ, Akkerman JW. Platelet activation by low density lipoprotein and high density lipoprotein. *Pathophysiol Haemost Thromb* 2006; 35: 270–280.
15. Haffner, SM, Miettinen, H, Mykkanen, et al. Leptin concentrations and insulin sensitivity in normoglycemic men *Int J Obes Relat Metab Disord* 1997; 21: 393–399.
16. Beltowski J. Leptin and atherosclerosis. *Atherosclerosis* 2006; 189: 47–60.
17. Konstantinides S, Schafer K, Loskutoff DJ. The prothrombotic effects of leptin possible implications for the risk of cardiovascular disease in obesity. *Ann NY Acad Sci* 2001; 947: 134–141.
18. Konstantinides S, Schafer K, Koschnick S, et al. Leptin-dependent platelet aggregation and arterial thrombosis suggests a mechanism for atherothrombotic disease in obesity. *J Clin Invest* 2001; 108: 1533–1540.
19. Nakata M, Yada T, Soejima N, et al. Leptin promotes aggregation of human platelets via the long form of its receptor. *Diabetes* 1999; 48: 426–429.
20. Ozata M, Avcu F, Durmus O, et al. Leptin does not play a major role in platelet aggregation in obesity and leptin deficiency. *Obes Res* 2001; 9: 627–630.
21. Canavan B, Salem RO, Schurgin S, et al. Effects of physiological leptin administration on markers of inflammation, platelet activation, and platelet aggregation during caloric deprivation. *J Clin Endocrinol Metab* 2005; 90: 5779–5785.
22. Giandomenico G, Dellas C, Czekay RP, et al. The leptin receptor system of human platelets. *J Thromb Haemost* 2005; 3: 1042–1049.
23. Corsonello A, Perticone F, Malara A, et al. Leptin-dependent platelet aggregation in healthy, overweight and obese subjects. *Int J Obes Relat Metab Disord* 2003; 27: 566–573.
24. Gualillo O, González-Juanatey JR, Lago F. The emerging role of adipokines as mediators of cardiovascular function: physiologic and clinical perspectives. *Trends Cardiovasc Med* 2007; 17: 275–283.
25. Kato H, Kashiwagi H, Shiraga M, et al. Adiponectin acts as an endogenous antithrombotic factor. *Arterioscler Thromb Vasc Biol* 2006; 26: 224–230.
26. Elbatarny HS, Netherton SJ, Ovens JD, et al. Adiponectin, ghrelin, and leptin differentially influence human platelet and human vascular endothelial cell functions: implication in obesity-associated cardiovascular diseases. *Eur J Pharmacol* 2007; 558: 7–13.
27. Von Hundelshausen P, Weber C. Platelets as immune cells: bridging inflammation and cardiovascular disease. *Circ Res* 2007; 100: 27–40.
28. Nicolucci A, De Berardis G, Sacco M, et al. ADA/ADA vs. ESC/EASD recommendations on aspirin as a primary prevention strategy in people with diabetes: how the same data generate divergent conclusions. *Eur Heart J* 2007; 28: 1925–1927.
29. Angiolillo DJ, Shoemaker SB, Desai B, et al. Randomized comparison of a high clopidogrel maintenance dose in patients with diabetes mellitus and coronary artery disease: results of the Optimizing Antiplatelet Therapy in Diabetes Mellitus (OPTIMUS) study. *Circulation* 2007; 115: 708–716.
30. Tamminen M, Lassila R, Westerbacka J, et al. Obesity is associated with impaired platelet-inhibitory effect of acetylsalicylic acid in nondiabetic subjects. *Int J Obes Relat Metab Disord* 2003; 27: 907–911.
31. Carter AM, Cymbalista CM, Spector TD, et al. Heritability of clot formation, morphology, and lysis: the EuroCLOT study. *Arterioscler Thromb Vasc Biol* 2007; 27: 2783–2789.
32. Collet JP, Allali Y, Lesty C, et al. Altered fibrin architecture is associated with hypofibrinolysis and premature coronary atherothrombosis. *Arterioscler Thromb Vasc Biol* 2006; 26: 2567–2573.
33. Diamant M, Nieuwland R, Pablo RF, et al. Elevated numbers of tissue-factor exposing microparticles correlate with components of the metabolic syndrome in uncomplicated type 2 diabetes mellitus. *Circulation* 2002; 106: 2442–2447.
34. Meerarani P, Moreno PR, Cimmino G, et al. Atherothrombosis: role of tissue factor; link between diabetes, obesity and inflammation. *Indian J Exp Biol* 2007; 45: 103–110.
35. Napoleone E, Di Santo A, Amore C, et al. Leptin induces tissue factor expression in human peripheral blood mononuclear cells: a possible link between obesity and cardiovascular risk? *J Thromb Haemost* 2007; 5: 1462–1468.

36. Samad F, Pandey M, Loskutoff DJ. Tissue factor gene expression in the adipose tissues of obese mice. *Proc Natl Acad Sci USA* 1998; 95: 7591–7596.
37. Samad F, Pandey M, Loskutoff DJ. Regulation of tissue factor gene expression in obesity. *Blood* 2001; 98: 3353–3358.
38. Vaidyula VR, Rao AK, Mozzoli M, et al. Effects of hyperglycemia and hyperinsulinemia on circulating tissue factor procoagulant activity and platelet CD40 ligand. *Diabetes* 2006; 55: 202–208.
39. Sakkinen PA, Wahl P, Cushman M, et al. Clustering of procoagulation, inflammation, and fibrinolysis variables with metabolic factors in insulin resistance syndrome. *Am J Epidemiol* 2000; 152: 897–907.
40. Godsland IF, Crook D, Proudler AJ, et al. Hemostatic risk factors and insulin sensitivity, regional body fat distribution, and the metabolic syndrome. *J Clin Endocrinol Metab* 2005; 90: 190–197.
41. Kraja AT, Province MA, Arnett D et al. Do inflammation and procoagulation biomarkers contribute to the metabolic syndrome cluster? *Nutr Metab (Lond)* 2007; 4: 28.
42. Folsom AR, Conlan MG, Davis CE, et al. Relations between hemostasis variables and cardiovascular risk factors in middle-aged adults. *Atherosclerosis Risk in Communities (ARIC) Study. Ann Epidemiol* 1992; 2: 481–494.
43. Fibrinogen Studies Collaboration. Plasma fibrinogen level and the risk of major cardiovascular diseases and nonvascular mortality: an individual participant meta-analysis. *J Am Med Assoc* 2005; 294: 1799–1809.
44. Fain JN, Madan AK, Hiler ML, et al. Comparison of the release of adipokines by adipose tissue, adipose tissue matrix, and adipocytes from visceral and subcutaneous abdominal adipose tissues of obese humans. *Endocrinology* 2004; 145: 2273–2282.
45. Yudkin JS, Kumari M, Humphries SE, et al. Inflammation, obesity, stress and coronary heart disease: is interleukin-6 the link? *Atherosclerosis* 2000; 148: 209–214.
46. Carvalho de Sousa J, Bruckert E, Giral P, et al. Coagulation Factor VII and plasma triglycerides. Decreased catabolism as a possible mechanism of factor VII hyperactivity. *Haemostasis* 1989; 19: 125–130.
47. Grant PJ. Diabetes mellitus as a prothrombotic condition. *J Intern Med* 2007; 262: 157–172.
48. Mineo C, Deguchi H, Griffin JH, et al. Endothelial and antithrombotic actions of HDL. *Circ Res* 2006; 98: 1352–1364.
49. Alessi MC, Juhan-Vague I. PAI-1 and the metabolic syndrome: links, causes, and consequences. *Arterioscler Thromb Vasc Biol* 2006; 26: 2200–2207.
50. Eren M, Painter CA, Atkinson JB, et al. Age-dependent spontaneous coronary arterial thrombosis in transgenic mice that express a stable form of human plasminogen activator inhibitor-1. *Circulation* 2002; 106: 491–496.
51. Sobel BE. Increased plasminogen activator inhibitor-1 and vasculopathy. A reconcilable paradox. *Circulation* 1999; 99: 2496–2498.
52. Juhan-Vague I, Pyke SDM, Alessi MC, et al. Fibrinolytic factors and the risk of myocardial infarction or sudden death in patients with angina pectoris. *Circulation* 1996; 94: 2057–2063.
53. Mertens I, Verrijken A, Michiels JJ, et al. Among inflammation and coagulation markers, PAI-1 is a true component of the metabolic syndrome. *Int J Obes* 2006; 30: 1308–1314.
54. Morange PE, Alessi MC, Verdier M, et al. PAI-1 produced *ex vivo* by human adipose tissue is relevant to PAI-1 blood level. *Arterioscler Thromb Vasc Biol* 1999; 9: 1361–1365.
55. Alessi MC, Peiretti F, Morange P, et al. Production of plasminogen activator inhibitor 1 by human adipose tissue: possible link between visceral fat accumulation and vascular disease. *Diabetes* 1997; 46: 860–867.
56. Shimomura I, Funahashi T, Takahashi M, et al. Enhanced expression of PAI-1 in visceral fat: possible contributor to vascular disease in obesity. *Nat Med* 1996; 2: 800–803.
57. Bastelica D, Morange P, Berthet B, et al. Stromal cells are the main plasminogen activator inhibitor-1 producing cells in human fat: evidence of differences between visceral and subcutaneous deposits. *Arterioscler Thromb Vasc Biol* 2002; 22: 173–178.
58. Fain JN, Madan AK, Hiler ML, et al. Comparison of the release of adipokines by adipose tissue, adipose tissue matrix, and adipocytes from visceral and subcutaneous abdominal adipose tissues of obese humans. *Endocrinology* 2004; 145: 2273–2282.
59. Cigolini M, Targher G, Agostino G, et al. Liver steatosis and its relation to plasma haemostatic factors in apparently healthy men—role of the metabolic syndrome. *Thromb Haemost* 1996; 76: 69–73.
60. Alessi MC, Bastelica D, Mavri A, et al. Plasma PAI-1 levels are more strongly related to liver steatosis than to adipose tissue accumulation. *Arterioscler Thromb Vasc Biol* 2003; 23: 1262–1268.
61. Festa A, D'Agostino R Jr, Tracy RP, et al. Insulin Resistance Atherosclerosis Study. Elevated levels of acute-phase proteins and plasminogen activator inhibitor-1 predict the development of type II diabetes: the insulin resistance atherosclerosis study. *Diabetes* 2002; 51: 1131–1137.
62. Festa A, Williams K, Tracy RP et al. Progression of plasminogen activator inhibitor-1 and fibrinogen levels in relation to incident type II diabetes. *Circulation* 2006; 113: 1753–1759.
63. Kanaya AM, Wassel Fyr C, Vittinghoff E, et al. Adipocytokines and incident diabetes mellitus in older adults: the independent effect of plasminogen activator inhibitor 1. *Arch Intern Med* 2006; 166: 350–356.
64. Meigs JB, O'donnell CJ, Tofler GH, et al. Hemostatic markers of endothelial dysfunction and risk of incident type 2 diabetes: the Framingham Offspring Study. *Diabetes* 2006; 55: 530–537.
65. Ingelsson E, Pencina MJ, Tofler GH, et al. Multi-marker approach to evaluate the incidence of the metabolic syndrome and longitudinal changes in metabolic risk factors: the Framingham Offspring Study. *Circulation* 2007; 116: 984–992.
66. Ma LJ, Mao SL, Taylor KL, et al. Prevention of obesity and insulin resistance in mice lacking plasminogen activator inhibitor 1. *Diabetes* 2004 ;53: 336–46.
67. De Taeye BM, Novitskaya T, Gleaves L, et al. Bone marrow plasminogen activator inhibitor-1 influences the development of obesity. *J Biol Chem* 2006; 281: 32796–32805.
68. Schafer K, Fujisawa K, Konstantinides S, et al. Disruption of the plasminogen activator inhibitor I gene reduces the adiposity and improves the metabolic profile of genetically obese and diabetic ob/ob mice. *FASEB J* 2001; 15: 1840–1842.
69. Crandall DL, Quinet EM, El Ayachi S, et al. Modulation of adipose tissue development by pharmacologic inhibition of PAI-1. *Arterioscler Thromb Vasc Biol* 2006; 26: 2209–2215.
70. Lijnen HR, Alessi MC, Frederix L, et al. Tiplaxtinin impairs nutritionally induced obesity in mice. *Thromb Haemost* 2006; 96: 731–737.
71. Lijnen HR, Alessi MC, Van Hoef B, et al. On the role of plasminogen activator inhibitor-1 in adipose tissue development and insulin resistance in mice. *J Thromb Haemost* 2005; 3: 1174–1179.
72. Liang X, Kanjanabuch T, Mao SL, et al. Plasminogen activator inhibitor-1 modulates adipocyte differentiation. *Am J Physiol Endocrinol Metab* 2006; 290: E103–E113.
73. Lebrun P, Baron V, Hauck CR, et al. Cell adhesion and focal adhesion kinase regulate insulin receptor substrate-1 expression. *J Biol Chem* 2000; 275: 38371–38377.
74. Lopez-Alemany R, Redondo JM, Nagamine Y, et al. Plasminogen activator inhibitor type-1 inhibits insulin signaling by competing with alphavbeta3 integrin for vitronectin binding. *Eur J Biochem* 2003; 270: 814–821.
75. Lijnen HR, Maquoi E, Morange P, et al. Nutritionally induced obesity is attenuated in transgenic mice overexpressing plasminogen activator inhibitor-1. *Arterioscler Thromb Vasc Biol* 2003; 23: 78–84.
76. Scroyen I, Christiaens V, Lijnen HR. No functional role of plasminogen activator inhibitor-1 in murine adipogenesis or adipocyte differentiation. *J Thromb Haemost* 2007; 5: 139–145.
77. Schalkwijk CG, Stehouwer CD. PAI-1 inhibition in obesity and the metabolic syndrome: a promising therapeutic strategy. *Thromb Haemost* 2006; 96: 698–699.
78. Baron AD, Steinberg H, Brechtel G, et al. Skeletal muscle blood flow independently modulates insulin-mediated glucose uptake. *Am J Physiol* 1994; 266: E248–253.
79. Coggins M, Lindner J, Rattigan S, et al. Physiologic hyperinsulinemia enhances human skeletal muscle perfusion by capillary recruitment. *Diabetes* 2001; 0: 2682–2690.
80. Baron AD, Tarshoby M, Hook G, et al. Interaction between insulin sensitivity and muscle perfusion on glucose uptake in human skeletal muscle, evidence for capillary recruitment. *Diabetes* 2000; 49: 768–774.
81. Lteif A, Vaishnava P, Baron AD, et al. Endothelin limits insulin action in obese/insulin-resistant humans. *Diabetes* 2007; 56: 728–734.
82. Kim JA, Montagnani M, Koh KK, et al. Reciprocal relationships between insulin resistance and endothelial dysfunction: molecular and pathophysiological mechanisms. *Circulation* 2006; 113: 1888–1904.
83. Montagnani M, Golovchenko I, Kim I, et al. Inhibition of phosphatidylinositol 3-kinase enhances mitogenic actions of insulin in endothelial cells. *J Biol Chem* 2002; 277: 1794–1799.
84. Arteaga RB, Chirinos JA, Soriano AO, et al. Endothelial microparticles and platelet and leukocyte activation in patients with the metabolic syndrome. *Am J Cardiol* 2006; 98: 70–74.
85. Meigs JB, Mittleman MA, Nathan DM, et al. Hyperinsulinemia, hyperglycemia, and impaired hemostasis: the Framingham Offspring Study. *J Am Med Assoc* 2000; 283: 221–228.
86. Juhan-Vague I, Thompson SG, Jespersen J. Involvement of the hemostatic system in the insulin resistance syndrome. A study of 1500 patients with angina pectoris. The ECAT Angina Pectoris Study Group. *Arterioscler Thromb* 1993; 13: 1865–1873.
87. Ziccardi P, Nappo F, Giugliano G, et al. Reduction of inflammatory cytokine concentrations and improvement of endothelial functions in obese women after weight loss over one year. *Circulation* 2002; 105: 804–849.
88. Picchi A, Gao X, Belmadani S, et al. Tumor necrosis factor-alpha induces endothelial dysfunction in the prediabetic metabolic syndrome. *Circ Res* 2006; 99: 69–77.
89. Valle Jimenez M, Estepa RM, Camacho RM, et al. Endothelial dysfunction is related to insulin resistance and inflammatory biomarker levels in obese prepubertal children. *Eur J Endocrinol* 2007; 156: 497–502.

- 90.** Lakka HM, Laaksonen DE, Lakka TA, et al. The metabolic syndrome and total and cardiovascular disease mortality in middle-aged men. *J Am Med Assoc* 2002; 288: 2709–2716.
- 91.** Hu G, Qiao Q, Tuomilehto J, et al, DECODE Study Group. Prevalence of the metabolic syndrome and its relation to all-cause and cardiovascular mortality in nondiabetic European men and women. *Arch Intern Med* 2004; 164: 1066–1076.
- 92.** Meigs JB, Wilson PW, Nathan DM, et al. Prevalence and characteristics of the metabolic syndrome in the San Antonio. Heart and Framingham Offspring Studies. *Diabetes* 2003; 52: 2160–2167.
- 93.** Freeman MS, Mansfield MW, Barrett JH, et al. Insulin resistance: an atherothrombotic syndrome. The Leeds family study. *Thromb Haemost* 2003; 89: 161–168.
- 94.** Malone PC, Agutter PS. The aetiology of deep venous thrombosis. *QJM* 2006; 99: 581–593.
- 95.** Prandoni P. Links between arterial and venous disease. *J Intern Med* 2007; 262: 341–350.
- 96.** Vayá A, Mira Y, Ferrando F, et al. Hyperlipidaemia and venous thromboembolism in patients lacking thrombophilic risk factors. *Br J Haematol* 2002; 118: 255–259.
- 97.** Gonzalez-Ordóñez AJ, Fernandez-Carreira JM, Fernandez-Alvarez CR, et al. The concentrations of soluble vascular cell adhesion molecule-1 and lipids are independently associated with venous thromboembolism. *Haematologica* 2003; 88: 1035–1043.
- 98.** Doggen CJ, Smith NL, Lemaitre RN, et al. Serum lipid levels and the risk of venous thrombosis. *Arterioscler Thromb Vasc Biol* 2004; 24: 1970–1975.
- 99.** Deguchi H, Pecheniuk NM, Elias DJ, et al. High-density lipoprotein deficiency and dyslipoproteinemia associated with venous thrombosis in men. *Circulation* 2005; 112: 893–899.
- 100.** Ageno W, Becattini C, Brighton T, et al. Cardiovascular risk factors and venous thromboembolism: a meta-analysis. *Circulation* 2008; 117: 93–102.
- 101.** Hansson PO, Eriksson H, Welin L, et al. Smoking and abdominal obesity: risk factors for venous thromboembolism among middle-aged men: „the study of men born in 1913“. *Arch Intern Med* 1999; 159: 1886–1890.
- 102.** Ray JG, Lonn E, Yi Q, et al. HOPE-2 investigators. Venous thromboembolism in association with features of the metabolic syndrome. *QJM* 2007; 100: 679–684.
- 103.** Ageno W, Prandoni P, Romualdi E, et al. The metabolic syndrome and the risk of venous thrombosis: a case-control study. *J Thromb Haemost* 2006; 4: 1914–1918.
- 104.** Ay C, Tengler T, Vormittag R, et al. Venous thromboembolism—a manifestation of the metabolic syndrome. *Haematologica* 2007; 92: 374–380.