REVIEW

Understanding plant defence responses against herbivore attacks: an essential first step towards the development of sustainable resistance against pests

M. Estrella Santamaria · Manuel Martínez · Inés Cambra · Vojislava Grbic · Isabel Diaz

Received: 22 April 2013/Accepted: 10 June 2013/Published online: 21 June 2013 © Springer Science+Business Media Dordrecht 2013

Abstract Plant-herbivore relationships are complex interactions encompassing elaborate networks of molecules, signals and strategies used to overcome defences developed by each other. Herbivores use multiple feeding strategies to obtain nutrients from host plants. In turn, plants respond by triggering defence mechanisms to inhibit, block or modify the metabolism of the pest. As part of these defences, herbivore-challenged plants emit volatiles to attract natural enemies and warn neighbouring plants of the imminent threat. In response, herbivores develop a variety of strategies to suppress plant-induced protection. Our understanding of the plant-herbivore interphase is limited, although recent molecular approaches have revealed the participation of a battery of genes, proteins and volatile metabolites in attack-defence processes. This review describes the intricate and dynamic defence systems governing plant-herbivore interactions by examining the diverse strategies plants employ to deny phytophagous arthropods the ability to breach newly developed mechanisms of plant

M. E. Santamaria \cdot M. Martínez \cdot I. Cambra \cdot I. Diaz (\boxtimes)

M. E. Santamaria · V. Grbic

Department of Biology, University of Western Ontario, 1151 Richmond St, London, ON N6A 5B7, Canada resistance. A cornerstone of this understanding is the use of transgenic tools to unravel the complex networks that control these interactions.

Keywords Plant defence · Plant-herbivore interaction · Transgenic plants · Signalling pathway

Introduction

Plants and arthropods coexist as friends or foes, sharing 400 million years of evolution. Currently, more than 50 % of a million of known insect species are phytophagous, although they are only represented in nine of the 29 insect orders (Ortego 2012). In addition, among the 5,000 species of acari identified, a significant number are plant-feeders (Helle and Sabelis 1985). The majority of herbivore arthropods are specialists, restricted to feeding on a single or a low number of plant species (mono- or oligo-phagous, respectively), while generalist herbivores (polyphagous) have preserved their association with a wide range of host plants over millions of years (Wheat et al. 2007). Throughout their co-evolution, phytophagous arthropods have developed morphological, physiological and behavioural adaptations, depending on their feeding habits and life styles, to overcome plant defences. They have developed a variety of strategies to avoid, excrete, sequester and degrade plant toxins. Although herbivores eat all plant organs,

Centro de Biotecnología y Genómica de Plantas UPM-INIA, Universidad Politécnica de Madrid, Campus Montegancedo, 28223 Pozuelo de Alarcón, Madrid, Spain e-mail: i.diaz@upm.es

leaf tissues are the main target for potential feeders. Nearly two-thirds of phytophagous arthropods are leaf-feeding species, with sophisticated mouth structures for chewing, snipping or tearing. Alternatively, piercing-sucking herbivores use tube-like structures (thrips and spider mites) or specialised stylets (aphids and whiteflies) to suck nutrients from parenchymatic cells or vascular tissues. Leafminers feed between epidermal cell layers within leaf tissues, where they are protected from plant defences and predators (Howe and Jander 2008). In addition, plant feeders have adapted their digestive physiology (pH, redox potential, enzymes, protease inhibitors and detoxification systems) and have developed digestive strategies (endo- or exo-peritrophic compartments and time-sequenced enzymatic activities) to a particular feeding style, which correlates with their phylogenetic position (Ortego 2012).

Plants have evolved elaborated protection systems against herbivore attacks by combining constitutive basal defences with inducible defences in response to specific arthropods. Moreover, plants have developed an indirect defence mechanism using volatiles and nectars to attract natural enemies of herbivores. Many of the approximately 500,000 secondary metabolites produced by plants are powerful chemicals against pests (Wu and Baldwin 2010). Plant volatiles may also function as signals to prime defence responses of neighbouring plants not still infected by insects or mites (Alba et al. 2012). Even if this battery of protection systems is not fully successful, plants can implement emergency responses and induce localised senescence or cell death processes in the infested tissues to isolate the attacker (Liu et al. 2010; Kerchev et al. 2012).

The co-evolutionary dynamics of plant defences and herbivore adaptations triggers a complex network of molecular events from the plant that ends in a diversification of structures, metabolic pathways and biochemical compounds. Consequently, plants have developed the ability to avoid or minimise being eaten by herbivores (resistance) and to decrease the negative fitness effects of pest damage (tolerance). These two defence mechanisms against pests may complement each other (Carmona and Fornoni 2013). However, damage to crops from field pests is still a limiting factor agriculture, and, in some cases, pests such as aphid or mite species, cause secondary effects by acting as virus vectors. Moreover, stored organs (seeds, tubers) infested with beetles, borers, moths or mites produce only a portion of their nutritive value and germination capacity, and subsequently lead to a decrease in grain sales and/or exportation.

Genetic transformation has emerged as a significant tool in modern agriculture. For years, transgenic plant technologies have primarily been targeted at the enhancement of pest, disease and weed controls to increase crop yields. More recently, plant genetic transformation has focused on the improvement of resistance to abiotic stresses, the enhancement of nutritional quality and the production of modified or new biomolecules and biomaterials for industrial uses. Many recent reviews have reported the main advances in the genetic transformation of plant species (Chandler and Sanchez 2012; Christou 2013; Gambino and Gribaudo 2012; Holme et al. 2013). However, there are still only a few examples of commercial applications of transgenic plant even with the record 170.3 million hectares of biotech crops grown globally in 2012 (http://www.isaaa.org/kc/cropbiotechupdate). As a consequence of the increasing publication of plant and arthropod genomes (Martinez 2012), the functional analysis of genes has become highly relevant in basic research. Studies of functional genomics are becoming more and more important, being the majority of their results based on plant transgenic technologies. The overexpression or silencing of genes involved in the synthesis of specific plant molecules yields new insights and makes it possible to uncover regulatory mechanisms associated with specific metabolic processes. The current and future challenge of plant genetic engineering is to access the functional characterisations of gene networks involved in physiological pathways, with the purpose of exploiting intrinsic biological processes they have for use in breeding and IPM (Integrated Pest Management) programmes.

Plant constitutive defences

Plants have developed an intricate basal defence system to decrease the chances of phytophagous arthropods using them as a source of nutrients. Constitutive defences are most likely essential for plant survival. They include physical barriers such as cuticular and cell wall modifications, surface waxes, trichomes, spines and thorns to hinder or deter herbivore accessibility, and the production of chemical defences, -mainly toxins and deterrents- to reduce palatability and affect growth, development and digestion of herbivores (Fig. 1). Among the plethora of structural defences, the protective role of trichomes has been the most studied. Trichomes are hair-like epidermal structures mainly located on leaves and stems that interfere with herbivore movements or even entrap herbivores. They may also prevent insect/mite egg attachment to plant tissues after oviposition. Glandular trichomes produce, store and secrete different classes of toxic metabolites and repellents associated with resistance to herbivores (Glas et al. 2012). Trichome morphology, size and density influence host plant resistance to pests. Their defensive role has been demonstrated by the negative correlation between trichome density and rates of herbivore damage (Hanley et al. 2007). Moreover, trichome density can also be induced by herbivores and plant hormones (Tian et al. 2012).

Transgenic plant technologies, including manipulative and genetic methods, have been very useful in studying the protective function of trichomes. For example, flea beetle Phyllotreta spp. feeding decreased on transgenic canola (Brassica napus) seedlings with elevated trichome density. Beetles required twice as much time to reach satiation when feeding on leaves with enhanced pubescence than on control leaves (Soroka et al. 2011). This study suggests that dense pubescence developed in canola plants with a strong potential to deter feeding by crucifer flea beetles. Likewise, in a screen of activation-tagged Populus tremula x Populus alba trees, Plett et al. (2010) identified a *fuzzy* mutant line with increased foliar trichome density, which correlated with a significant resistance to feeding by larvae of the white-spotted tussock moth (Orgya lecustigma). The fuzzy trichome phenotype was attributable to activation tagging and

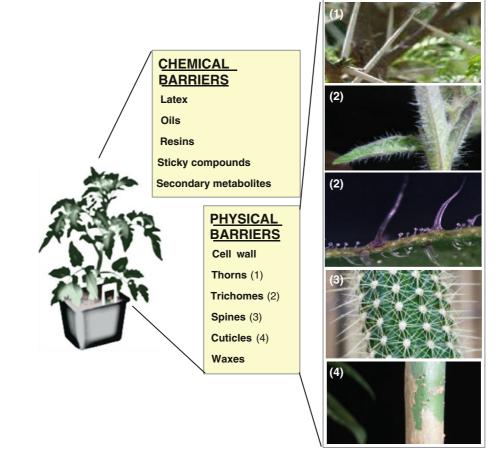


Fig. 1 Plant constitutive defences against phytophagous arthropods include physical and chemical barriers

increased expression of the gene encoding the PtaM-YB186 transcription factor. Moreover, the transgenic overexpression of PtaMYB186 in poplar not only increased trichome density but also improved resistance against insects.

Plant transgenic approaches have also been used to analyse the impact of structural changes of the cell wall on pest behaviour. Sasidharan et al. (2011) recently reviewed how modifying the cell wall composition might result in a wall structure more resistant to physical penetration by invading attackers. In celery, aphid infestation induces a massive transcriptional reprogramming, including a group of genes encoding cell wall-modifying proteins as one of the major functional groups (Divol et al. 2007). Furthermore, silencing of Manduca sexta-induced pectin methyl esterases (PMEs), which catalyse the demethylesterification of pectins and therefore result in cell wall loosening or stiffening, increased herbivore feeding performance in PME knockdown Nicotiana attenuata plants (Korner et al. 2009). PME-silenced plants also had significantly lower levels of jasmonic and salicylic acid. Another experimental approach attempted to produce ferulic acid modifications, based on the potential of ferulic acid to form crosslinks between cell wall polysaccharides. Genetically engineered tall festuca (Festuca arundinacea) expressing a ferulic acid esterase gene showed reduced levels of ferulate cross-linking in cell walls, and an increased susceptibility to Spodoptera frugiperda feeding (Buanafina and Fescemyer 2012).

In addition to the structural adaptations developed by plants, some defensive chemicals are also continuously expressed irrespective of herbivore, although they are dependent on the plant species and their levels vary in plant tissues. These chemical products include peptides, proteins, protease inhibitors and other compounds with anti-nutritional or toxic properties. Approximately 10 % of angiosperms synthesise latex, oils, resins and other sticky substances that trap herbivores (Konno 2011). These substances are exuded from wounded points in plant tissues caused either mechanically or by plant feeders. In particular, latex is a complex mixture enriched in proteins (lectins, poteases and other enzymes) and secondary metabolites (terpenoids, alkaloids, cardenolides, furanocoumarins and phenolics) with important defensive roles (Konno 2011). Enzymatic adaptations to degrade latex ingredients have been described in several herbivores (Hirayama et al. 2007). Secondary metabolites are also essential for reducing plant palatability, and have potential repellent, deterrent or even toxic effects against pests (Ibanez et al. 2012).

The defensive roles of many of these compounds have been revealed by silencing or blocking gene expression in planta through genetic engineering approaches. For example, Nicotiana attenuata transformed with consensus fragments of the putresceine N-methyl transferase gene in either the antisense or inverted-repeat orientation inhibited nicotine production, resulting in the destruction of 3-fold greater leaf area from a variety of herbivores compared to control plants (Steppuhn et al. 2004). Similarly, silencing of geranylgeranyl diphosphate synthase in N. attenuata blocked the synthesis of antifeedant diterpenoid glycosides and impaired resistance to the tobacco hornworm Manduca sexta (Jassabi et al. 2008). Notably, the wide variety of plant chemicals of glandular trichomes includes secondary metabolites, defence proteins and organic acids. The majority, constitutively expressed or induced by herbivores, produces drastic effects on arthropod performance and host-plant selection behaviour (Glas et al. 2012; Tian et al. 2012; War et al. 2012). The molecular engineering of glandular trichome chemistry has allowed a better understanding of plant structural defences to predict how plant morphological responses might influence plant-herbivore interactions. Wang et al. (2001, 2004) reported that the suppression of the cytochrome P450 hydroxylase in transformed tobacco plants, either by antisense inhibition or sense co-suppression, enhanced trichome exudate cembratrieneols and exhibited significant protection from aphid infestation in the field. More recently, Choi et al. (2012) have shown that tobacco plants overexpressing the NtLTP1 gene which plays a role in lipid secretion from trichome heads, resulted in a greater secretion of trichome exudates, including epicuticular wax, than control plants. These transgenic tobacco lines showed improved resistance to aphid infestation. In contrast, RNAi-treated tobacco in which liquid secretion from trichomes was strongly reduced but epicuticular wax secretion was not altered, showed increased aphid infestation levels.

Plant inducible defences

Most plant defences against pests are induced by the feeder, primarily in the tissue where the infestation takes place, in which case it is termed the local response, and then in distal undamaged tissues of the same plant, which is referred to as the systemic response. In both responses, the protection comprises structural and chemical modifications, some of which are similar to those previously described as constitutive defences. The induction of these plant reactions requires appropriated recognition systems (receptors) of the Herbivore-Associated Elicitors (HAEs), Herbivore Associated Molecular Patterns (HAMPs) or herbivore effectors. This recognition triggers signal transduction pathways that connect herbivore-specific elicitors to the expression of suitable defence genes (Fig. 2).

Plant perception

Mechanical wounding of plant tissues is the first consequence of plant-herbivore interactions although damaged tissue is not always an indicator of a pest attack. Plant perception begins with the herbivore contact. The physical pressures, cell disruption,

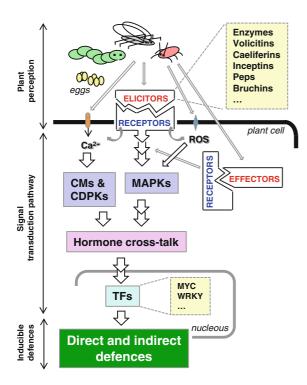


Fig. 2 Plant inducible defences against phytophagous arthropods require appropriated recognition systems to trigger signal transduction pathways that connect herbivore specific elicitors to the expression of suitable defence genes. *CMs* Calmodulins, *CDPKs* Ca²⁺ dependent protein kinases, *MAPKs* mitogen activated protein kinases, *TFs* transcriptional factors

deposition of chemical compounds or arthropod oviposition are differentially recognised by plants as elicitors or effectors of the defence response (Balbyshev and Lorenzen 1997; Peiffer et al. 2009; Bonaventure 2012). Comparisons between host transcriptome profiles induced by mechanical injury or by arthropod feeding present an important overlap (Ralph et al. 2006; Lawrence et al. 2012). Nevertheless, it is clearly demonstrated that plants induce different responses by discriminating between herbivore feeding or application of insect oral secretions (OSs) to wounded sites from mechanical damages (Giri et al. 2006; Howe and Jander 2008; Lawrence et al. 2008, 2012). Moreover, current experimental approaches based on "omic"-technologies have shown that specific damage produced by chewing, phloem-feeding, mesophyll-sucking or leafminer herbivores are coupled to specific plant responses (Carolan et al. 2011; Lawrence et al. 2012; Zhang et al. 2012).

Among the molecules considered as HAEs or HAMPs, salivary enzymes of insect herbivores (ß-glucosidases, glucose oxidases, oxireductases and proteases) and particularly FACs (Fatty acid-Amino acid Conjugates) have been widely studied (Bonaventure et al. 2011; Yoshinaga et al. 2007; Rodriguez and Bos 2013). FACs are fatty acids derived from plant membranes conjugated mainly with amino acids from herbivores, found in OSs. The first FAC characterised was the volicitin, N-(17-hydroxylinolenoyl)-L-glutamine, first identified in Spodoptera exigua (Alborn et al. 1997) and subsequently found in other lepidopteran and dipteran species (Yoshinaga et al. 2007). Other analogues of FACs are the caeliferins, sulphur-containing fatty acids isolated from regurgitants of several insects (Alborn et al. 2007). In addition, cryptic peptides derived from proteins can elicit plant defence. The best studied cryptic peptides are the inceptins, proteolytic products of the plant chloroplastic ATP synthase γ -subunit (cATPC) identified in cowpea plants after Spodoptera frugiperda feeding (Schmelz et al. 2006, 2007). Ingested cATPs are cleaved in insect midguts and subsequently form inceptins. Both volicitins and inceptins bind reversibly to plant cell membranes, most likely through specific receptors and trigger the release of volatile terpenes with a potential function as indirect defences (Truitt et al. 2004; Alborn et al. 2007). The application of volicitins and inceptins on mechanically damaged leaves induces Mitogen Activated Protein kinase (MAPK), jasmonic acid (JA) and ethylene (ET) biosynthesis. Consequently, transcriptomic, proteomic and metabolomic responses are modified, and the amplification of direct and indirect plant defences is induced (Schmelz et al. 2007; Skibbe et al. 2008; Meldau et al. 2009; Wu and Baldwin 2010). In addition, families of elicitor peptides (Peps), which play an important role in plant resistance against pathogens, have also been implicated in defence against herbivores (Huffaker et al. 2013). Peps are plant C-terminal peptides derived from cytoplasmic PROPep precursors, which are very well studied in Arabidopsis where their receptors have also been identified (Yamaguchi et al. 2010; Yamaguchi and Huffaker 2011). In addition, molecules identified in oviposition fluids also elicit specific defence responses in host plants. For example bruchins -long-chain ω, α diols esterified at one or both oxygens with 3-hydroxypropanoic acid-, have been isolated from the oviposition fluid of pea weevils and benzyl cyanides have been detected in cabbage after Pieris brassicae oviposition (Doss et al. 2000; Fatouros et al. 2008).

Plant transgenic technologies have also been applied to study the functionality of genes encoding putative HAEs or HAMPs molecules. Bos et al. (2010) identified 48 effector candidates in the salivary glands of Myzus persicae, using transient plant overexpression assays to analyze elicitation of phenotype and effects on aphid performance. The same group of researchers demonstrated silencing of the aphid C002 effector by dsRNA feeding from plants (Pitino et al. 2011). Recently, Pitino and Hogenhout (2013) have found protein effectors that promote aphid feeding and colonisation in a plant-specific manner. M. persicae reproduction increased in transgenic Arabidopsis producing the C002, PintO1 and PINtO2 effectors, whereas its reproduction was not altered on Arabidopsis producing the Acyrthosiphon pisum orthologues of these proteins. Moreover, a plant-mediated RNAi approach has shown that c002- and Pit02-silenced M. persicae has reduced progeny compared with nonsilenced aphids.

Notably, plant defence responses are induced not only by molecules derived from arthropods or by plants after arthropod feeding but also by endogenous host-plant elicitors as a consequence of wounding or herbivore attacks. Elicitors can be released by cell death, as small molecules or molecule fragments and reactive oxygen species (ROS) are recognised as alarm signals. One example is the wound-signalling peptide systemin, a C-terminal peptide derived from prosystemin which promotes protease inhibitor (PI) and JA synthesis and systemic volatile emissions in tomatoes (McGurl et al. 1992; Degenhardt et al. 2010). There is direct evidence indicating that systemin activates a phospholipase A that releases linolenic acid from membrane lipids in wounded tomato leaves and consequently induces JA synthesis (Narvaez-Vasquez et al. 1999; Sun et al. 2011). When thrips and spider mites fed on prosystemin-overexpressing tomato, leaf damage was decreased and herbivore fecundity was reduced compared to results from control tomato (Li et al. 2002). Remarkably, 35S-prosystemin lines increased volatile terpenes emissions in tomatoes to attract insect enemies compared to wild type plants (McGurl et al. 1994; Li et al. 2002; Degenhardt et al. 2010). In contrast and as expected, transgenic plants expressing an antisense prosystemin cDNA were defective in the systemic expression of PIs, and this resulted in greater susceptibility to pest attacks (McGurl et al. 1992). These results provide strong evidence of an efficient defensive function of the systemin/JA signalling pathway and again highlight the value of transgenic techniques for plant defence research.

Plant receptors

Phytophagous arthropods are recognised when conserved HAE, HAMPs or effectors are detected by plant Pattern Recognition Receptors (PRRs) leading defensive reactions (Bos et al. 2010; Hogenhout and Bos 2011; Erb et al. 2012). This complex mechanism is similar to the PAMP or MAMP (pathogen/microbeassociated molecular patterns) and Effector Triggered Immunity (ETI) systems from microorganisms, widely described and responsible for activating plant immunity (Wu and Baldwin 2010).

A considerable number of trans-membrane receptor-like protein kinases (RLKs) have been associated with plant defence. RLKs often contain an extracellular receptor and a cytoplasmic catalytic kinase domain, identified as a Nucleotide Binding Site (NBS) and a Leucine Rich Repeat (LRR). These receptors transduce an extracellular recognition event into an intracellular signalling cascade to trigger host resistance to herbivores (Bonaventure 2012). In most cases this step requires additional RLKs for full function (Yamaguchi and Huffaker 2011).

Until now, investigations to uncover the function of PRRs in herbivore resistance have not been wellestablished. However, the relation of genes encoding proteins of the NBS-LRR family to pest resistance has been determined by transgenic approaches. The tomato Mi-1 gene was the first cloned NBS-LRR gene that confers resistance to aphids, psyllids and whiteflies when expressed in tomato plants (Rossi et al. 1998; Vos et al. 1998; Nombela et al. 2003; Casteel et al. 2006). Mi-1-mediated resistance to aphids requires the RLK, SISERK1, as confirmed by tomato virus-induced gene silencing experiments (Mantelin et al. 2011). In lettuce, silencing of several members of the RGC2 family encoding NBS-LRR proteins increases their susceptibility to root aphids (Wroblewski et al. 2007). In rice, overexpression of *Bph14*, a CC-NB-LRR encoding gene, activates the salicylic acid (SA) pathway and generates a higher resistance to the brown plant hopper (Du et al. 2009). Similarly, virusinduced gene silencing of the Lectin receptor kinase 1 (LecRK1) in Nicotiana attenuata prevents the full defence response against the lepidopteran Manduca sexta, because LecRK1 participates in the suppression of the insect-mediated inhibition of JA-induced defence responses (Gilardoni et al. 2011). Very recently, it has been described ML3, a putative MD2 lipopolysaccharide-recognition domain protein with a putative function as a regulator for HAMP recognition. ML3 loss-of-function in Arabidopsis plants compromised the upregulation of herbivore-induced genes, and the larvae of Spodoptera littoralisthat fed on ml3 mutant plants gained more weight than those that fed on wild-type plants (Fridborg et al. 2013).

Signal transduction pathway

Direct or indirect recognition of HAEs or HAMPs initiates a signal transduction pathway involving the rapid activation of numerous processes, including Ca^{2+} -signalling, production of ROS, phosphorylation cascades and transcriptional activation events that ultimately culminate in specific defence responses and the transmission of alerts to cells in other parts of the damaged plant (Maffei et al. 2007; Wu and Baldwin 2010; Bonaventure 2012; War et al. 2012).

Early events in plant-herbivore interactions start with ion imbalances, causing variations in membrane potential and increases in the levels of cytosolic free calcium. Calmodulins (CMs) and Ca²⁺-dependent protein-kinases (CDPKs) have been identified as Ca^{2+} sensors modulating the first step of a downstream cascade of defence effects. Calmodulin is activated in transgenic tobacco lines overexpressing a glutamate decarboxylase whose product, Gamma-aminobutyrate (GABA) is an inhibitory neurotransmitter. The activation of calmodulin correlates with GABA hyper accumulation, as those plants are more resistant to budworm larvae than control plants (MacGregor et al. 2003). Similarly, silencing of Nicotiana attenuata Ca²⁺-dependent Protein Kinases CDPK4 and CDPK5 strongly up-regulates JA levels after wounding or Manduca sexta infestation with a parallel delay in the arva development rate (Yang et al. 2012). Ca^{2+} is also associated with NO synthesis and ROS production by modulating NADPH oxidase and polyphenol oxidase (PPO) activities (Wu et al. 2013). Although ROS function in signalling has not been extensively studied during herbivore attacks, it has been reported that insect/acari feeding increases plant ROS levels (Maffei et al. 2007; Santamaria et al. 2012). Transgenic tomato and poplar plants overexpressing PPO genes presented changes in redox levels and resulted in higher mortality of Spodotera litura and Malacosoma disstria, respectively (Wang and Constabel 2004; Mahanil et al. 2008). In contrast, rboh D-silenced plants, which lacked functional NADPH oxidases, were more vulnerable to insects (Miller et al. 2009; Wu et al. 2013).

In addition, strong evidence suggests that ROS signalling is closely related to hormone-signalling pathways in the plant-herbivore interactions. In particular, jasmonates have been proposed as the "master regulators" of plant defences. Herbivore elicits JA, and plant cells transduce JA signals into the activation of transcriptional regulation of defence-genes, with many dependent on the MAP kinase pathway (Kanchiswamy et al. 2010; Bonaventure et al. 2011). In a major branch of the JA pathway, JA conjugates with the amino acid isoleucine (JA-Ile) bind to the F-Box protein receptor COI1 (Yan et al. 2009) and causes ubiquitination and degradation of JAZ protein repressors (Chini et al. 2007; Thines et al. 2007). Consequently, their target transcription factors, such as the MYC class factors, are liberated (Pauwels et al. 2010). One of the MYC proteins, MYC2, is shown to activate several metabolic pathways involved in plant defence

against herbivores (Dombrecht et al. 2007). Moreover, Stotz et al. (2000) reported that *coi1-1* mutants were more susceptible to *Spodoptera littoralis* feeding than the wild type plants, and the triple mutant *myc2 myc3 myc4* was as impaired as *coi1-1* in the activation of defence against this pest (Fernández-Calvo et al. 2011). As part of this defence pathway, Schweighofer et al. (2007) showed that the PP2C-type phosphatase AP2C1 negatively regulated MPK4 and MPK6, modulated JA and ethylene levels in Arabidopsis and influenced the response to phytophagous mites and pathogens.

WRKY transcription factors are also important regulators of the JA response. The application of Manduca sexta FACs on Nicotiana attenuatawounded leaves induces the activation of MAPKs, JA and ET biosynthesis and signalling, and WRKY factors, which ultimately produce the amplification and modification of wounding-induced transcriptomic, proteomic and metabolomic responses. This transduction pathway ends with the induction of direct and indirect plant defences (Wu et al. 2007; Skibbe et al. 2008). In contrast, silencing of WRKY3, WRKY6 or both in N. attenuate rendered plants highly susceptible to Manduca sexta attack (Skibbe et al. 2008). Once more, transgenic approaches have been used to analyse gene functionality in vivo. Plants that overor under-express several JA-regulated genes (LOX, prosystemin, or pin2) enhance or impair resistance to lepidopteran caterpillars (Royo et al. 1999; Ryan 2000; Kandoth et al. 2007).

Many articles and reviews have covered particular aspects of the JA-induced responses against pests, including a complex cross-talk with other hormones (Ballare 2010; Robert-Seilaniantz et al. 2011). Two other hormones with a key role in orchestrating the expression of plant defences against herbivores are SA and ET. Generally, it is accepted that SA antagonises JA-induced resistance, but SA and ET may both have positive and/or negative effects. In this way, Pieris brassicae oviposition induced SA accumulation and reduced JA-responsive genes, leading to reduced Arabidopsis resistance against S. littoralis (Bruessow et al. 2010). Ethylene signalling reduces the resistance of Arabidopsis towards Spodoptera littoralis but not against Plutella xylostella. Moreover, the hookless1 and ein2 mutations, which affect components of ET signalling, conferred resistance to S. littoralis (Stotz et al. 2000). Auxins, abscisic acid, brassinosteroids, citokinins and gibberellins have received less attention as herbivore resistance modulators but the role of these hormones in mediating specificity in herbivoreinduced defence responses has been recently reported (Robert-Seilaniantz et al. 2011; Erb et al. 2012). This hormonal cross-talk is summarised in Fig. 3. Furthermore, recent reports have revealed an integration of ROS and hormone signalling pathways in chloroplast in response to insects (Kerchev et al. 2012; Nabity et al. 2013). It can be concluded that phytohormones work in a very complex fashion, either I a cooperative or antagonistic role, and have become of increasing importance in plant defence research.

Future challenges

As demonstrated throughout this review, there is a complex system of steps, signals and molecules induced in a plant after its interaction with phytophagous arthropods. The application of transgenic approaches has been essential for understanding the function of genes involved in the cascade of plant reactions to specifically develop either resistance or tolerance against each herbivore species.

An effective plant transgenic technology has been crucial for functional analysis of genes to understand

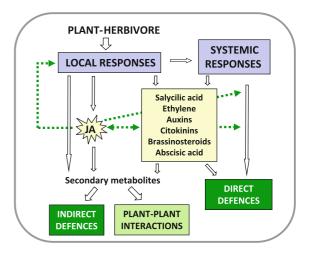


Fig. 3 The expression of plant defences locally or systematically induced by feeders, are mainly regulated by jamonic acid (JA) and a complex cross-talk with other hormones (salicylic acid, ethylene, brassinosteroids, abscisic acid, auxins and citokinins). Consequently, direct and indirect defences against herbivores are produced and herbivore-challenged plants can emit volatiles to warn neighbour plants of the pest presence

their in vivo roles. However, this knowledge is still fragmentary, and increasing the understanding of what happens in the plant-herbivore interphase is critical for having more and new alternatives to enhance plant defences against pests. Hopefully, in the coming years a wider availability of plant genome sequences will broaden their usage and multiply the possible applications of transgenic approaches. In the near future, the induced plant responses to pests can be engineered genetically so that the defensive compounds can be constitutively produced in plants challenged by herbivores. Moreover, induced resistance can be manipulated by using chemical elicitors or secondary metabolites to confer resistance to herbivores. Elicitors of induced responses could be sprayed on crop plants to build up the natural defence system against specific damage caused by a specific herbivore. However, more advances should be made to improve transformation efficiencies, to reduce plant manipulation time-consumption, to overcome the transformation of recalcitrant species and finally, to increase the number of cultivars, variants and plant genome backgrounds to be transformed. It is our hope that continued advancements in basic plant research to translate will be applied to enhance the defence systems of agriculturally important crops.

Acknowledgments Project funding by the Ministerio de Economía y Competitividad of Spain (AGL2011-23650), by the Government of Canada through Genome Canada and the Ontario Genomics Institute (OGI-046) and by Ontario Research Fund- Global Leadership in Genomics and Life Sciences (GL2-01-035).

References

- Alba JM, Bleeke PM, Glas JJ, Schimme BCJ, Wijk M, Sabelis MW, Schuurink RC, Kant MR (2012) The impact of induced plant volatiles on plant-arthropod interactions. In: Smagghe G, Diaz I (eds) Arthropod-plant interactionsnovel insights and approaches for IPM. Springer, Dordrecht, pp 15–73
- Alborn T, Turlings TCJ, Jones TH, Stenhagen G, Loughrin JH, Tumlinson JH (1997) An elicitor of plant volatiles from beet armyworm oral secretions. Science 276:945–949
- Alborn HT, Hansen TV, Jones TH, Bennet DC, Tumlinson JH, Schmelz EA, Teal PEA (2007) Disulfooxy fatty acids from the American birdgrasshopper Schistocerca americana, elicitors of plant volatiles. Proc Natl Acad Sci USA 104:12976–12981
- Balbyshev NF, Lorenzen JH (1997) Hypersensitivity and egg drop: a novel mechanism of host plant resistance to

Colorado potato beetle (Coleoptera: Chrysomelidae). J Econ Entomol 90:652–657

- Ballare CL (2010) Jasmonate-induced defenses: a tale of intelligence, collaborators and rascals. Trends Plant Sci 16: 249–257
- Bonaventure G (2012) Perception of insect feeding by plants. Plant Biol 14:872–880
- Bonaventure G, Van Doorn A, Baldwin IT (2011) Herbivoreassociated elicitors: FAC signaling and metabolism. Trends Plant Sci 16:294–299
- Bos JI, Prince D, Pitino M, Maffei ME, Win J, Hogenhout SA (2010) A functional genomics approach identifies candidate effectors from the aphid species *Myzus persicae* (green peach aphid). PLoS Genet 6:3100216
- Bruessow F, Gouhier-Darimont C, Buchala A, Metraux JP, Reymond P (2010) Insect eggs suppress plant defence against chewing herbivores. Plant J 62:876–885
- Buanafina MMO, Fescemyer HW (2012) Modification of esterifed cell wall phenolics increases vulnerability of tall fescue to herbivory by the fall armyworm. Planta 236:513–523
- Carmona D, Fornoni J (2013) Herbivores can select for mixed defensive strategies in plants. New Phytol 197:576–585
- Carolan JC, Caragea D, Reardon KT, Mutti NS, Dittmer N, Pappan K, Cui F, Castaneto M, Poulain J, Dossat C, Tagu D, Reese JC, Reeck GR, Wilkinson TL, Edwards OR (2011) Predicted effector molecules in the salivary secretome of the pea aphid Acyrthosiphon pisum: a dual transcriptomic/proteomic approach. J Proteome Res 10:1505–1508
- Casteel CL, Walling LL, Paine TD (2006) Behaviour and biology of the tomato psyllid, *Bactericerca cockerelli*, in response to the *Mi-1.2* gene. Entomol Exp Appl 121:67–72
- Chandler SF, Sanchez C (2012) Genetic modification; the development of transgenic ornamental plant varieties. Plant Biotechnol J 10:891–903
- Chini A, Fonsec S, Chico JM, Fernández-Calvo P, Solano R (2007) The JAZ family of repressors is the missing link in jasmonate signalling. Nature 448:666–671
- Choi YE, Lim S, Kim HJ, Han JY, Lee MH, Yang Y, Kim JA, Kim YS (2012) Tobacco NtLTP1, a glandular-specific lipid transfer protein, is required for lipid secretion from glandular trichomes. Plant J 70:480–491
- Christou P (2013) Plant genetic engineering and agricultural biotechnology 1983–2013. Trends Biotechnol 31:125–127
- Degenhardt D, Refi-Hind S, Strarmann JW, Lincoln DE (2010) Systemin and jasmonic acid regulate constitutive and herbivore-induced systemic volatile emission in tomato, *Solanum lycopersicum*. Phytochemistry 71:2024–2037
- Divol F, Vilaine F, Thibivilliers S, Kusiak C, Sauge MH, Dinant S (2007) Involvement of the xyloglucan endotransglycosylase/hydrolases encoded by celery XTH1 and *Arabidopsis* XTH33 in the phloem response to aphids. Plant, Cell Environ 30:187–201
- Dombrecht B, Xue GP, Sprague SJ, Kirkegaard JA, Ross JJ, Reid JB, Fitt GP, Sewelam N, Schenk PM, Manners JM, Kazan K (2007) MYC2 differentially modulates diverse jasmonate-dependent functions in *Arabidopsis*. Plant Cell 19:2225–2245
- Doss RP, Oliver JE, Proebsting WM, Potter SW, Kuy S, Clement SL, Williamson RT, Carney JR, DeVilbiss ED (2000) Bruchins: insect-derived plant regulators that

stimulate neoplasm formation. Proc Natl Acad Sci USA 97:6218-6223

- Du B, Zhang W, Liu B, Hu J, Wei Z, Shi Z, He R, Zhu L, Chen R, Han B, He G (2009) Identification and characterization of *Bph14*, a gene conferring resistance to brown planthopper in rice. Proc Natl Acad Sci USA 106:22163–22168
- Erb M, Meldau S, Howe GA (2012) Role of phytohormones in insect-specific plant reactions. Trends Plant Sci 17:250–259
- Fatouros NE, Broekgaarden C, Bukovinszkine'Kiss G, van Loon JJ, Mumm R, Huigens ME, Dicke M, Hilker M (2008) Male-derived butterfly antiaphrodisiac mediates induced indirect plant defense. Proc Natl Acad Sci USA 105:10033–10038
- Fernández-Calvo P, Chini A, Fernández-Barbero G, Chico JM, Gimenez-Ibanez S, Geerinck J, Eeckhout D, Schweizer F, Godoy M, Franco-Zorrilla JM, Pauwels L, Witters E, Puga MI, Paz-Ares J, Goossens A, Reymond P, De Jaeger G, Solano R (2011) The *Arabidopsis* bHLH transcription factors MYC3 and MYC4 are targets of JAZ repressors and act additively with MYC2 in the activation of jasmonate responses. Plant Cell 23:701–715
- Fridborg I, Johansson A, Lagensjö J, Leelarasamee N, Floková K, Tarkowská D, Meijer J, Bejai S (2013) ML3: a novel regulator of herbivory-induced responses in *Arabidopsis thaliana*. J Exp Bot 64:935–948
- Gambino G, Gribaudo I (2012) Genetic transformation of fruit trees: current status and remaining challenges. Transgenic Res 21:1163–1181
- Gilardoni PA, Hettenhausen C, Baldwin IT, Bonaventure G (2011) Nicotiana attenuata LECTIN RECEPTOR KINASE1 suppresses the insect-mediated inhibition of induced defense responses during Manduca sexta herbivory. Plant Cell 23:3512–3532
- Giri AP, Wunsche H, Mitra S, Zavala JA, Muck A, Svatos A, Baldwin IT (2006) Molecular interactions between the specialist herbivore *Manduca sexta* (Lepidoptera, phingidae) and its natural host *Nicotiana attenuata*. VII. Changes in the plant's proteome. Plant Physiol 142:1621–1641
- Glas JJ, Schimmel BC, Alba JM, Esobar-Bravo R, Chuurink RXC, Kant MR (2012) Plant glandular trichomes as targets for breeding of resistance to herbivores. Int J Mol Sci 13:17077–17103
- Hanley ME, Lamont BB, Fairbanks MM, Rafferty CM (2007) Plant structural traits and their role in anti-herbivore defence. Persp Plant Ecol Evol Syst 8:157–178
- Helle W, Sabelis MW (1985) World crop pests, spider mites: their biology, natural enemies, and control. Elsevier Science Publishers, New York
- Hirayama C, Konno K, Wasano N, Nakamura M (2007) Differential effects of sugar-mimic alkaloids in mulberry latex on sugar metabolism and disaccharidases of Eri and domesticated silkworms: enzymatic adaptation of *Bombyx mori* to ulberry defense. Insect Biochem Mol Biol 37:1348–1358
- Hogenhout SA, Bos JI (2011) Effector proteins that modulate plant-insect interactions. Current Opin Plant Biol 14:422–428
- Holme IB, Wendt T, Holm PB (2013) Intragenesis and cisgenesis as alternatives to transgenic crop development. Plant Biotechnol J. doi:10.1111/pbi.12055
- Howe GA, Jander G (2008) Plant immunity to insect herbivores. Ann Rev Plant Biol 59:41–66

- Huffaker A, Pearce G, Veyrat N, Erb M, Turlings TEC, Sartor R, Shene Z, Briggs SP, Vaughan MM, Alborm HT, Teal PEA, Schmelz EA (2013) Plant elicitor peptides are conserved signals regulating direct and indirect anti-herbivore defense. Proc Natl Acad Sci USA 14:5707–5712
- Ibanez S, Gallet C, Despres L (2012) Plant insecticidal toxins in ecological networks. Toxins 4:228–243
- Jassabi AR, Gase K, Hettenhausen C, Schmidt A, Baldwin IT (2008) Silencing geranylgeranyl diphosphate symthase in *Nicotiana attenuata* dramatically impairs resistance to tobacco hornworm. Plant Physiol 146:974–986
- Kanchiswamy CN, Takahashi H, Quadro S, Maffei M, Bossi S, Bertea C, Zebelo SA, Muroi A, Ishihama N, Yoshioka H, Boland W, Takabayashi J, Endo Y, Sawasaki T, Arimura G (2010) Regulation of *Arabidopsis* defense responses against *Spodoptera littoralis* by CPK-mediated calcium signaling. BMC Plant Biol 10:97
- Kandoth PK, Ranf S, Pancholi SS, Jayanty S, Walla MD, Miller W, Howe GA, Lincoln DE, Stratmann JW (2007) Tomato MAPKs LeMPK1, LeMPK2, and LeMPK3 function in the systemin-mediated defense response against herbivorous insects. Proc Natl Acad Sci USA 104:12205–12210
- Kerchev PL, Fenton B, Foyer CH, Hancock RD (2012) Plant responses to insect herbivory: interactions between photosynthesis, reactive oxygen species and hormonal signalling pathways. Plant, Cell Environ 35:441–453
- Konno K (2011) Plant latex and other exudates as plant defense systems: roles of various defense chemicals and proteins contained therein. Phytochemistry 72:1510–1530
- Korner E, von Dahl CC, Bonaventure G, Baldwin IT (2009) Pectin methylesterase NaPME1 contributes to the emission of methanol during insect herbivory and to the elicitation of defence responses in *Nicotiana attenuata*. J Exp Bot 60:2631–2640
- Lawrence SD, Novak NG, Ju CJ, Cooke JE (2008) Potato, Solanum tuberosum, defense against Colorado potato beetle, Leptinotarsa decemlineata (Say): microarray gene expression profiling of potato by Colorado potato beetle regurgitant treatment of wounded leaves. J Chem Ecol 34:1013–1025
- Lawrence SD, Novak NG, Kayal WEl, Ju CHT, Cooke JEK (2012) Root herbivory: molecular analysis of the maize transcriptome upon infestation by Southern corn root-worm, *Diabrotica undecimpunctata howardi*. Physiol Plant 144:303–319
- Li C, Williams MM, Loh YT, Lee GI, Howe GA (2002) Resistance of cultivated tomato to cell content-feeding herbivores is regulated by the octadecanoid-signalling pathway. Plant Physiol 130:494–503
- Liu X, Williams CE, Nemacheck JA, Wang H, Subramanyam S, Zheng C, Chen MS (2010) Reactive oxygen species are involved in plant defense against a gall midge. Plant Physiol 152:985–999
- MacGregor KB, Shelp BJ, Peiris S, Bown AW (2003) Overexpression of glutamate decarboxylase in transgenic tobacco plants deters feeding by phytophagous insect larvae. J Chem Ecol 29:2177–2182
- Maffei ME, Mithöfer A, Boland W (2007) Before gene expression: early events in plant-insect interaction. Trends Plant Sci 12:310–316

- Mahanil S, Attajarusit J, Stout MJ, Thipyapong P (2008) Overexpression of tomato polyphenol oxidase increases resistance to common cutworm. Plant Sci 174:456–466
- Mantelin S, Peng HC, Li B, Atmian HS, Takken FLW, Kaloshian I (2011) The receptor-like SISERK1 is required for Mi-1-mediated resistance to potato aphids in tomato. Plant J 67:459–471
- Martinez M (2012) Co-evolution of genes for specification in arthropod-plant interaction: a bioinformatics analysis in plant and arthropod genomes. In: Smagghe G, Diaz I (eds) Arthropod-plant interactions- novel insights and approaches for IPM. Springer, Dordrecht, pp 15–73
- McGurl B, Pearce G, Orozco-Cardenas M, Ryan CA (1992) Structure, expression, and antisense inhibition of the system in precursor gene. Science 255:1570–1573
- McGurl B, Orozco-Cardenas M, Pearce G, Ryan CA (1994) Overexpression of the prosystemin gene in transgenic tomato plants generates a systemic signal that constitutively induces proteinase inhibitor synthesis. Proc Natl Acad Sci USA 91:9799–9802
- Meldau S, Wu JQ, Baldwin IT (2009) Silencing two herbivoryactivated MAP kinases, SIPK and WIPK, does not increase *Nicotiana attenuata*'s susceptibility to herbivores in the glasshouse and in nature. New Phytol 181:161–173
- Miller G, Schlauch K, Tam R, Cortes D, Torres MA, Shulaev V, Dangl JL, Mittler R (2009) The plant NADPH oxidase RBOHD mediates rapid systemic signaling in response to diverse stimuli. Sci Signal 2:ra45
- Nabity PD, Zavala JA, DeLucia EH (2013) Herbivore induction of jasmonic acid and chemical defences reduce photosynthesis in *Nicotina attenuate*. J Exp Bot 64:685–694
- Narvaez-Vasquez J, Florin-Christensen J, Ryan CA (1999) Positional specificity of a phospholipase A activity induced by wounding, system in and oligosaccharide elicitors in tomato leaves. Plant Cell 11:2249–2260
- Nombela G, Williamson VM, Muñiz M (2003) The root-knot nematode resistance gene *Mi-1.2* of tomato is responsible for resistance against the whitefly *Bemisia tabaci*. Mol Plant Microbe Interact 16:645–649
- Ortego F (2012) Physiological adaptations of the insect gut to herbivory. In: Smagghe G, Diaz I (eds) Arthropod-plant interactions- novel insights and approaches for IPM. Springer, Dordrecht, pp 15–73
- Pauwels L, Barbero GF, Geerinck J, Grunewalsd W, Perez AC, Chico JM, Bossche RV, Sewell J, Gil E, Garcia-Casado G, Witters E, Inze D, Long JA, Laeqer G, Solano R, Goossens A (2010) NINJA connects the co-repressor TOPLESS to jasmonate signaling. Nature 464:788–791
- Peiffer M, Tooker JF, Luthe DS, Felton GW (2009) Plants on early alert: glandular trichomes as sensors for insect herbivores. New Phytol 184:644–656
- Pitino M, Hogenhout SA (2013) Aphid protein effectors promote aphid colonization in plant species-specific manner. Mol Plant-Microbe Inter 1:130–139
- Pitino M, Coleman AD, Maffei ME, Ridout CJ, Hogenhout SA (2011) Silencing of aphid genes by dsRNA feeding from plants. PLoS ONE 6:e25709
- Plett JM, Wilkins O, Campbell MM, Ralph SG, Regan S (2010) Endogenous overexpression of *Populus* MYB186 increases trichome density, improves insect pest resistance, and impacts plant growth. Plant J 64:419–432

- Ralph SG, Yueh H, Friedmann M, Aeschliman D, Zeznik JA, Nelson CC, Butterfield YS, Kirkpatrick R, Liu J, Jones SJ, Marra MA, Douglas CJ, Ritland K, Bohlmann J (2006) Conifer defense against insect: microarray gene expression profiling of Sitka spruce (*Picea sitchensis*) induced by mechanical wounding or feeding by spruce budworms (*Choristoneura occidentalis*) or white pine weevils (*Pissodes strobe*) reveals large-scale changes of the host transcriptome. Plant, Cell Environ 29:1545–1570
- Robert-Seilaniantz A, Garnt M, Jones JDG (2011) Hormone crosstalk in plant disease and defense: more than just JASMONATES-SALICYLATE antagonism. Ann Rev Phytopathol 49:317–343
- Rodriguez PA, Bos JI (2013) Toward understanding the role of aphid effectors in plant infestation. Mol Plant Microbe Interact 26:25–30
- Rossi M, Goggin FL, Milligan SB, Kaloshian I, Ullman DE, Williamson VM (1998) The nematode resistance gene *Mi* of tomato confers resistance against the potato aphid. Proc Natl Acad Sci USA 95:9750–9754
- Royo J, Leon J, Vancanneyt G, Albar JP, Rosahl S, Ortego F, Castanera P, Sanchez-Serrano JJ (1999) Antisense-mediated depletion of a potato lipoxygenase reduces wound induction of proteinase inhibitors and increases weight gain of insect pests. Proc Natl Acad Sci USA 96:1146–1151
- Ryan CA (2000) The systemin signaling pathway: differential activation of defensive genes. Biochim Biophys Acta 1477:112–122
- Santamaria ME, Cambra I, Martinez M, Pozancos C, González-Melendi P, Grbic V, Castañera P, Ortego F, Diaz I (2012) Gene pyramiding of peptidase inhibitors enhances plant resistance to the spider mite *Tetranychus urticae*. PLoS ONE 7:e43011
- Sasidharan R, Voesenek LACJ, Pierik R (2011) Cell wall modifying proteins mediate plant acclimatization to biotic and abiotic stresses. Crit Rev Plant Sci 30:548–562
- Schmelz EA, Carroll MJ, LeClere S, Phipps SM, Meredith J, Chourey PS, Alborn HT, Teal PEA (2006) Fragments of ATP synthase mediated plant perception of insect attack. Proc Natl Acad Sci USA 103:8894–8899
- Schmelz EA, LeClere S, Carroll MJ, Alborn HT, Teal PEA (2007) Cowpea chloroplasts ATP synthase is the source of multiple plant defense elicitors during insect herbivory. Plant Physiol 144:793–805
- Schweighofer A, Kazanaviciute V, Scheikl E, Teige M, Doczi R, Hirt H, Schwanninger M, Kant M, Mauch F, Buchala A, Cardinale F, Meskiene I (2007) The PP2C-type phosphatase AP2C1, which negatively regulates MPK4 and MPPK6, modulates innate immunity, jasmonic acids, and ethylene levels in Arabidopsis. Plant Cell 19:2213–2224
- Skibbe M, Qu N, Galis I, Baldwin IT (2008) Induced plant defenses in the natural environment: *Nicotiana attenuata* WRKY3 and WRKY6 coordinate responses to herbivory. Plant Cell 20:1984–2000
- Soroka JJ, Holowachuk JM, Gruber MY, Grenkow LF (2011) Feeding by flea beetles (Coleoptera: Chrysomelidae; *Phyllotreta* spp.) is decreased on canola (*Brassica napus*) seedlings with increased trichome density. J Econ Entomol 104:125–136
- Steppuhn A, Gase K, Kroch B, Halitschke R, Baldwin IT (2004) Nicotine's defensive function in nature. PLoS Biol 2:e382

- Stotz HU, Pittendrigh BR, Kroymann J, Weniger K, Fritsche J, Bauke A, Mitchell-Olds T (2000) Induced plant defense responses against chewing insects. Ethylene signaling reduces resistance of *Arabidopsis* against Egyptian cotton worm but not diamondback moth. Plant Physiol 124:1007–1017
- Sun JQ, Jiang HL, Li CY (2011) Systemin/jasmonate-mediated systemic defense signalling in tomato. Mol Plant 4:607–615
- Thines B, Katsir L, Melotto M, Niu Y, Mandaokar A, Liu G, Nomura K, He SY, Howe GA, Browse J (2007) JAZ repressor proteins are targets of the SCF (COI1) complex during jasmonate signalling. Nature 448:661–665
- Tian D, Tooker J, Peiffer M, Chung SH, Felton GW (2012) Role of trichomes in defense against herbivores: comparison of herbivore response to woolly and hairless trichome mutants in tomato (*Solanum lycopersicum*). Planta 236:1053–1066
- Truitt CL, Wei HX, Pare PW (2004) A plasma membrane protein from *Zea mays* binds with the herbivore elicitor volicitin. Plant Cell 16:523–532
- Vos P, Simons G, Jesse T, Wijbrandi J, Heinen L, Hogers R, Frijters A, Groenendijk J, Diergaarde P, Reijans M, Fierens-Onstenk J, de Both M, Peleman J, Liharska T, Hontelez J, Zabeau M (1998) The tomato *Mi-1* gene confers resistance to both root-knot nematodes and potato aphids. Nat Biotechnol 16:1365–1369
- Wang J, Constabel CP (2004) Polyphenol oxidase overexpression in transgenic *Populus* enhances resistance to herbivory by forest tent caterpillar (*Malacosoma disstria*). Planta 220:87–96
- Wang E, Wang R, Deparasis J, Loughrin JH, Gan SS, Wagner GJ (2001) Suppression of a P450 hydroxylase gene in plant trichome glands enhances natural-product-based aphid resistance. Nat Biotechnol 19:371–374
- Wang E, Hall JT, Wagner GJ (2004) Transgenic Nicotiana tabacum L. with enhanced trichome exudates cembratrieneols has reduced aphid infestation in the field. Mol Breeding 13:49–57
- War AR, Paulraj MG, Ahmad TA, Buhroo AA, Hussain B, Ignacimuthu S, Sharma HC (2012) Mechanisms of plant defence against insect herbivores. Plant Signal Behav 7:1–15
- Wheat CW, Vogel H, Wittstock U, Braby MF, Underwood D, Mitchell-Olds T (2007) The genetic basis of a plant-insect

coevolutionary key innovation. Proc Natl Acad Sci USA 104:20427–20431

- Wroblewski T, Piskurewicz U, Tomczak A, Ochoa O, Michelmore RW (2007) Silencing of the major family of NBS-LRR-encoding genes in lettuce results in the loss of multiple resistance specificities. Plant J 51:803–818
- Wu J, Baldwin IT (2010) New insights into plant responses to the attack from insect herbivores. Annu Rev Genet 44:1–24
- Wu J, Hettenhausen C, Meldau S, Baldwin IT (2007) Herbivory rapidly activates MAPK signalling in attacked and unattacked leaf regions but not between leaves of *Nicotiana attenuata*. Plant Cell 19:1096–10122
- Wu J, Wang L, Wünsche H, Baldwin IT (2013) Narboh D, a respiratory burst oxidase homolog in Nicotiana attenuata, is required for late defense responses after herbivore attack. J Integ Plant Biol 55:187–198
- Yamaguchi Y, Huffaker A (2011) Endogenous peptide elicitors in higher plants. Current Opin Plant Biol 14:351–357
- Yamaguchi Y, Huffaker A, Bryan AC, Tax FE, Ryan AC (2010) PEPR2 is a second receptor for the Pep1 and Pep2 peptides and contributes to defenses in Arabidopsis. Plant Cell 22:508–522
- Yan J, Zhang C, Gu M, Bai Z, Zhang W, Qi T, Cheng Z, Peng W, Luo H, Nan F, Wang Z, Xie D (2009) The Arabidopsis coronatine insensitive1 protein is a jasmonate receptor. Plant Cell 21:2220–2236
- Yang DH, Hettenhausen C, Baldwin IT, Wu J (2012) Silencing Nicotiana attenuata calcium-dependent protein kinases, CDPK4 and CDPK5, strongly up-regulate wound- and herbivory-induced jasmonic acid accumulations. Plant Physiol 159:1591–1607
- Yoshinaga N, Aboshi T, Ishikawa C, Fukui M, Shimoda M, Nishida R, Lait CG, Tumlinson JH, Mori N (2007) Fatty acid amides, previously identified in caterpillars, found in the cricket *Teleogryllus taiwanemma* and the fruit fly *Drosophila melanogaster* larvae. J Chem Ecol 33:1376–1381
- Zhang S, Zhang Z, Kang L (2012) Transcriptome response analysis of *Arabidopsis thaliana* to leaf miner (*Liriomyza huidobrensis*). BMC Plant Biol 12:234