Comments

Dermatology

Dermatology 2003;206:96-105 DOI: 10.1159/000068476

The Role of Innate Immunity in the Pathogenesis of Acne

A. Koreck A. Pivarcsi A. Dobozy L. Kemény

Department of Dermatology and Allergology, University of Szeged, Hungary

Key Words

Acne · Toll-like receptors · CD1d molecule

Abstract

Acne is a multifactorial disease of the pilosebaceous follicle. The most significant pathogenetic factors of acne are: abnormal ductal keratinization, increased sebum secretion, abnormalities of the microbial flora and inflammation. The pilosebaceous unit is an immunocompetent organ. Keratinocytes and sebocytes may act as immune cells capable of pathogen recognition and abnormal lipid presentation, and they might have an important role in initiating and perpetuating the activation of both innate and adaptive immune responses. The elements of the skin immune system are involved in the development of both noninflammatory and inflammatory acne lesions.

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Acne is a chronic inflammatory disease of the pilosebaceous unit. The lesions are localized in the so-called acneprone areas (the cheeks, the nose, the forehead, the midline chest and the back), where sebaceous follicles are most common.

Acne lesions can be divided into noninflammatory and inflammatory lesions. The noninflammatory lesions are represented by microcomedones and comedones. The mi-

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Fax + 41 61 306 12 34 E-Mail karger@karger.ch www.karger.com © 2003 S. Karger AG, Basel 1018–8665/03/2062–0096\$19.50/0 Accessible online at: www.karger.com/drm crocomedo is the first pathogenic feature of acne that can be found in normal-looking skin and can only be identified microscopically [1].

Inflammatory acne lesions consist of papules, pustules and nodules. In these lesions the follicular epithelium is damaged, and a dermal inflammation occurs. Duct rupture can occur but is certainly not an early phenomenon [1, 2]. If rupture occurs the comedo content elicits an intense inflammatory response in the dermis. Long-lasting dermal inflammation may give rise to indurated nodules. Most patients have a mixture of noninflammatory and inflammatory lesions. An interesting feature of acne is the spontaneous resolution of the disease in the majority of young adults and of particular lesions during the disease. The background of this process is still unknown. A possible explanation for the resolution of lesions, especially of microcomedones and comedones, may be represented by the follicular cycling process [3].

There is a general agreement that acne is a multifactorial disease. The most significant pathogenetic factors of acne have been identified as: abnormal ductal keratinization resulting in comedogenesis, increased sebum production resulting in seborrhea, abnormalities of the microbial flora and inflammation [2, 4].

Andrea Koreck, MD, PhD Department of Dermatology and Allergology, University of Szeged Korányi fasor 6 H–6720 Szeged (Hungary) Tel. +36 62 54 52 78, Fax +36 62 54 59 54, E-Mail koreck@derma.szote.u-szeged.hu

Abnormal Ductal Keratinization

Microcomedones and comedones show ductal hyperkeratinization and subsequent obstruction of sebaceous follicles. The hyperproliferation of ductal keratinocytes has been confirmed by immunohistochemical staining with monoclonal antibody Ki-67, by an increase in ³Hthymidine labeling of comedones and the presence of keratins 6 and 16 [5–7]. The mechanism by which this process occurs is not elucidated. One important factor that induces this hyperproliferative state of ductal keratinocytes may be the modified lipid composition of the sebum [2, 8]. Lipids, which have been incriminated in comedogenesis are linoleic acid, free fatty acids and squalene [9]. Several studies have shown that keratinocyte proliferation and differentiation could be influenced by lipids. Nacylated forms of sphingolipids, such as ceramides, have been shown to promote keratinocyte differentiation, and sphingosine and sphingosylphosphorylcholine have been demonstrated to promote keratinocyte proliferation [10].

Beside hyperproliferation of ductal keratinocytes comedogenesis may be related to the failure of ductal keratinocytes to separate: however, conflicting results are available from studies concerning terminal differentiation of keratinocytes and desmosome expression [11, 12]. In comedogenesis hormones could also have a role [2].

Cytokines are another important factor that may induce proliferation of keratinocytes. In experiments performed by Guy and Kealey [13], comedogenesis was induced experimentally by interleukin (IL) 1 α and was blocked by IL-1 receptor antagonist. IL-1 α has been found to be present in high amounts in noninflammatory acne lesions. In contrast, epidermal growth factor markedly reduced comedo formation in vitro [13].

Increased Sebum Production

Almost all acne patients have increased sebum production, but excessive sebum production is not sufficient for the development of acne. It has been demonstrated that in acne patients a considerable heterogeneity in individual follicular sebum secretion exists, which suggests the existence of 'acne-prone' follicles [14]. The induction of sebum hypersecretion in acne is not fully elucidated. Acne generally begins at puberty, when androgen levels increase significantly and stimulate sebum secretion. In young adults acne usually wanes, whereas androgen levels remain high. Several studies have found increased 5α reductase levels and increased expression of androgen

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receptors in sebaceous glands of acne patients [15, 16]. These findings suggest that an end-organ hyperresponse of the sebaceous glands to androgens is probably the most likely explanation for the seborrhea.

In the development of acne, an important feature may be represented beside the increased quantity of sebum by its abnormal composition in certain lipid components.

Abnormalities of the Microbial Flora

A pathogenic factor of acne is the proliferation of normal flora and especially of *Propionibacterium acnes*. Acne is not an infectious disease, but the role of P. acnes is outlined by several data [17]. Sebum production shows a high correlation with P. acnes levels, so the increased sebum secretion of affected follicles provides a very good environment for the growth of P. acnes [18]. In some studies the number of P. acnes was found increased in acne lesions, but there was no correlation between the number of *P*. acnes and the severity and outcome of the disease [17]. Other studies found no difference between the numbers of *P. acnes* in affected follicles and control ones [19]. However, the importance of *P. acnes* is further suggested by the successful antibiotic therapy of acne and by the observation that the presence of resistant strains of P. acnes may be associated with therapeutic failure.

Several studies have demonstrated that in acne patients and especially in subgroups with severe forms, the immune response against P. acnes is abnormal [20]. Ashbee et al. [21] reported that patients with severe acne have significantly higher levels of complement fixing antibodies to P. acnes. Puhvel et al. [22] showed that lymphocyte stimulation by *P. acnes* is significantly increased in patients with nodulocystic acne. Kersey et al. [23] demonstrated that delayed skin test reactivity to P. acnes correlates with the severity of the disease. Recent studies performed by Jappe et al. [24] revealed that P. acnes could activate both peripheral blood mononuclear cells of adults and cord blood mononuclear cells and that the reactions produced are both antigenic and T cell mitogenic. These results are in concordance with other immunological studies, which revealed that the initial cells that infiltrate acne lesions are T helper lymphocytes [1]. P. acnes secretes various biologically active molecules like enzymes and chemotactic factors, which may have a role in the initiation and perpetuation of the local inflammatory response and even in the induction of keratinocyte hyperproliferation [25].

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Inflammation

The fourth main factor in the pathogenesis of acne is inflammation. It is widely accepted that inflammation in acne vulgaris may be mainly induced by an immunological reaction to *P. acnes*. Chemotactive substances released by bacteria attract cells of the immune system such as neutrophils, monocytes and lymphocytes [20, 25]. *P. acnes* may also activate the complement system with the formation of the potent chemoattractant C5a, which further amplifies the recruitment of immune cells [26]. *P. acnes* produces soluble factors that are able to activate immune cells with the consecutive secretion of various proinflammatory cytokines (IL-8, tumor necrosis factor α , IL-1 α) [25, 27].

Recent results indicate a key role for leukotriene B4 in the development of tissue inflammation in acne. Leukotriene B₄ is a proinflammatory mediator synthesized from arachidonic acid. Synthesis of leukotriene B4 is catalyzed by 5-lipoxygenase and leukotriene A₄ hydrolase and is increased by inflammatory mediators including endotoxin, complement fragments tumor necrosis factor α and interleukins [28, 29]. Zouboulis [30] showed that administration of a specific lipoxygenase inhibitor significantly decreased the inflammatory lesions in a small cohort of patients with inflammatory acne. In these patients, a significant reduction in total lipids and especially in free fatty acids in sebum as well as a substantial decrease in lipoperoxides was found. Bivariate analysis indicated that the decrease in total serum lipids, and especially in proinflammatory lipids, was directly correlated with the improvement in inflammatory lesions [30]. In cultured human sebocytes, 5-lipoxygenase was induced at the mRNA and protein levels under arachidonic acid and calcium ionophore treatment. Furthermore, leukotriene A4 hydrolase and peroxisome proliferator-activated receptor α were expressed in sebocytes at the mRNA and protein levels. The presence of the enzymes 5-lipoxygenase, and leukotriene A₄ hydrolase and the detection of peroxisome proliferator-activated receptor α in human sebocytes provide strong evidence that eicosanoids may play an important role in inflammatory sebaceous gland disorders, including acne [31].

These results also support the view that sebum lipids induce inflammation in acne, independent of the presence of bacteria or increased systemic levels of proinflammatory molecules [32]. Ceramides could induce degranulation of neutrophils and increased integrin expression on leukocytes [33]. Ceramides and sphingosylphosphorylcholine have been shown to modulate T lymphocyte proliferation [34]. Modification in the lipid composition also leads to a diminished epidermal barrier function and favors the diffusion of proinflammatory mediators through the follicular epithelium into the dermis [35].

Neuropeptides may also have an important role in the development of inflammatory acne lesions. Seiffert et al. [36] have demonstrated that human sebocytes express neuropeptide receptors and that their agonists possess stimulatory effects on cytokine production. Böhm et al. [37] have shown that the paracrine and/or endocrine actions of β -endorphin are mediated by the μ -opioid receptors expressed by human sebocytes. The presence of melanocortin 1 receptor in human sebocytes has also been demonstrated [38]. Recently, the role of corticotropin-releasing hormone as an autocrine hormone that stimulates lipogenesis in human sebocytes has also been established [39].

Beside the professional cells of the immune system, ductal keratinocytes and sebocytes are also able to secrete amounts of cytokines following various stimuli [40]. Released cytokines stimulate the expression of adhesion molecules on endothelial cells, have chemotactic properties, activate antigen-presenting cells and leukocytes and stimulate the formation of new inflammatory mediators such as leukotrienes and prostaglandins. Even before the rupture of the duct all the immunologically active substances diffuse through the follicular epithelium, which is already thinned. After the disruption of the duct, keratin, hair and lipids from the extruded sebum directly initiate inflammation and the formation of a macrophage giantcell foreign-body reaction.

Both in follicular hyperproliferation of keratinocytes, which is the primary cause of comedo formation, and in the development of inflammatory lesions, growth factors, cytokines and hormones produced locally in the pilosebaceous unit have been supposed to play an important role. The immune system has a high capacity of producing these growth-promoting factors and inflammatory cytokines. In acne lesions, these mediators were found to be upregulated, and activated immune cells could be detected in inflammatory acne lesions. These data suggest that elements of the skin immune system (SIS) might be responsible for producing these mediators and be involved in the development of both noninflammatory (hyperproliferative) and inflammatory acne lesions (table 1).

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 Table 1. Characteristic features of noninflammatory and inflammatory lesions in acne

	Noninflammatory acne lesions	Inflammatory acne lesions
Type of lesions	Microcomedo Comedo (open and closed)	Papules Pustules Nodules
Responsible cells	Keratinocytes Sebocytes?	Keratinocytes Sebocytes Monocytes, neutrophils T lymphocytes, NKT cells?
Responsible mediators	Abnormal lipids? Growth factors Cytokines (IL-1α) Hormones	Proinflammatory cytokines (IL-1α, IL-8, TNF-α) Metabolites of arachidonic acid (leukotrienes, prostaglandins) Enzymes Complement Growth factors Hormones Neuropeptides
Pathogens	P. acnes?	P. acnes

Skin Immune System

The immune system of vertebrates has two components: innate immunity and adaptive immunity. These two systems utilize two very different mechanisms for host defense. The innate immune system relies on a set of germline-encoded receptors that are expressed on a wide variety of cells, like macrophages and neutrophils as well as on epithelial cells situated at host-environment boundaries. The skin represents the largest organ of the human body. In addition to its structural functions, a specific immunological environment has developed in the skin. The theory of the SIS was introduced in 1987, by Bos et al. [41]. The SIS consists of professional immune cells, such as macrophages, neutrophils, dendritic cells and lymphocytes, and nonprofessional immune cells, such as keratinocytes and sebocytes.

The most important function of the innate immune system is to recognize foreign invaders and self-molecules with modified structures. This is a crucial event necessary for the activation of both innate and adaptive defense mechanisms. Abnormal reactions to such stimuli could lead to the development of pathological features. Considering the peculiar aspects of the pilosebaceous unit, which is constantly hosting *P. acnes* and is especially rich in lipids, two mechanisms could have an outstanding role in the development of acne lesions: recognition of pathogens by Toll-like receptors (TLRs) and presentation of abnormal lipids by CD1d molecules, which results in the activa-

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tion of natural killer T (NKT) cells. NKT cells could further activate both the cells of the innate immune system and the cells of the adaptive immune system; they represent a link between innate and adaptive immunity.

TLRs in the Skin: Recognition of Pathogens

Effective host defense against invading microorganisms requires the detection of foreign pathogens. The recognition of invading pathogens is mediated by germlineencoded receptors that are specific for common constituents of pathogenic microorganisms that are called pattern recognition receptors (PRRs). PRRs recognize invariant molecular structures called pathogen-associated molecular patterns (PAMPs) that are relatively invariant within a given class of microorganisms (table 2). Such a PAMP is the lipopolysaccharide (LPS) of gram-negative bacteria, mannan in the yeast cell wall, the mycobacterial cell wall component lipoarabinomannan and the peptidoglycan of gram-positive bacteria. In the past 10 years, a number of PRRs have been described that mediate complement activation, opsonization and phagocytosis. However, until recently only little has been known about the receptors responsible for the induction of immune and inflammatory genes. Recent studies on the recognition of PAMPs have highlighted the critical role of one group of PRRs, the TLRs in innate host defense. TLRs play crucial roles in the induction of antimicrobial responses in different

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Table 2. Common PAMPs and their PRRs[97, 98]

PAMPs	PRRs
LPS	LBP, CD14, TLR ₄ , TLR ₂
Lipoproteins	TLR_2
Peptidoglycan	$CD14, TLR_2$
CpG	TLR ₉
Lipoarabidomannan	TLR_2 , CD1
N-formyl-Met	f-Met receptors 1 and 2
Mannans, mannoproteins, zymosan	mannose-R, MBP, mannose-, glucan-R, TLR ₂
Heat shock proteins	undefined (TLR ₄ ?)
Flagellin	TLR_5

CpG = Deoxycytidylate-phosphate-deoxyguanylate; LBP = lipopolysaccharide-binding protein; R = receptor; MBP = mannose-binding protein.

cells [42]. In the past few years, at least 9 different human TLRs have been identified. Biochemical studies on an in vitro model and investigations on mice deficient in TLR₄ indicated that TLR₄ primarily mediates cellular signaling induced by gram-negative bacteria [43]. Targeted disruption of the TLR₄ gene resulted in abrogation of the response to LPS [44]. In humans, common mutations in the TLR₄ gene are associated with differences in LPS responsiveness [45]. While TLR₄ is highly specific for LPS and is associated with CD14, a coreceptor for LPS, TLR_2 , is implicated in the recognition of multiple products of gram-positive bacteria, mycobacteria and yeast. TLR₂ is required for proinflammatory signaling to lipoteichoic acid, peptidoglycan, lipoproteins, lipoarabinomannan and zymosan [42, 46-48]. The common downstream signaling pathway of TLR₂ and TLR₄ leads to the activation of NF-kB through the myeloid differentiation protein (MyD88) and IL-1-receptor-associated kinase in various cell types [49, 50].

Keratinocytes have been demonstrated to play important regulatory roles in cutaneous inflammatory and immune responses by producing various kinds of cytokines. Keratinocyte-derived cytokines are pivotal in mobilizing leukocytes from the blood and in signaling other cutaneous cells. Keratinocytes are able to produce proinflammatory cytokines such as IL-8 in response to microbial compounds providing initial chemotactic stimuli for neutrophils.

In addition to regulating immunological and inflammatory responses, the epidermal keratinocytes contribute to the protective barrier of the epithelia and participate in the host defense by killing invading microorganisms. It was earlier demonstrated that epidermal keratinocytes have direct candidacidal activity, which can be further increased by UV light [51, 52], IL-1 [53], IL-8 and α -melanocyte-stimulating hormone [54]. IL-1, prostaglandin E_2 and platelet-activating factor have also been demonstrated to be involved in *Candida* killing by human epidermal cells [55], but the mechanism of killing remained unknown. Recently it has been shown that keratinocytes express a new type of mannose binding receptor on their surface, which has a role in the binding and subsequent killing of *Candida albicans* [56]. Keratinocytes have been shown to produce inducible antimicrobial products such as nitric oxide [57], LL-37 [58], antileukoprotease [59] and β -defensins [60]. Keratinocyte-derived nitric oxide and antimicrobial peptides in the epidermis might be responsible for the killing of invading pathogens in the epidermis and for the prevention of systemic invasion by microbes [61, 62].

It was demonstrated that keratinocytes express TLR₂ and TLR₄ and that TLRs are involved in the microbial compound-mediated induction of proinflammatory cytokines in keratinocytes [63, 64]. Microbial compounds induce the nuclear translocation of NF- κ B transcription factor, and the activation of NF- κ B is required for the killing of *Candida* [63].

Recent studies have revealed that the pilosebaceous duct is also characterized by many features of immunological activity such as classical and nonclassical MHC class I expression, presence of intraepithelial Langerhans cells and expression of CD14, TLR₂ and TLR₄ [65, 66]. TLR₂ expressed by sebocytes might have a role in *P*.*acnes*-triggered induction of inflammatory response [67]. Antimicrobial peptides such as human β -defensins are also expressed in pilosebaceous follicles [68]. Investigation of the recently described immortalized sebaceous cell line might give better insight into the role of sebocytes in the innate immunity [69, 70].

CD1d Molecule in the Skin: Link between Innate and Adaptive Immunity

The CD1 molecules represent a family of conserved nonpolymorphic cell surface glycoproteins with a role in the presentation of nonpeptide antigens to T cells. The human CD1 gene family is composed of 5 members (CD1a, b, c, d and e); 4 of these genes are expressed in vivo (CD1a–d). The overall structure of the CD1 molecules resembles that of classical MHC class I molecules; however, the amino acid sequence homology to MHC class I is low which suggests a marked difference in function. On the basis of nucleotide and amino acid sequence homology, CD1 molecules are divided into two groups: group I, consisting of CD1a, b and c, and group II, comprising CD1d [71].

CD1d molecules are capable of presenting lipid and glycolipid antigens to a special type of T cells, the NKT cells [71]. The natural ligands of CD1d molecules are still unknown. In vitro and in vivo experiments have demonstrated that CD1d molecules are capable of binding synthetic glycolipids with a α -anomeric structure, α -galactosylceramide and a-glucosylceramide, and of presenting and stimulating NKT cells with these antigens. Interestingly α -mannosylceramide has been shown to be incapable of stimulating NKT cells, and the same proved to be true for all these ceramides with β -anomeric structure [72]. Additionally, the search for the natural antigen presented by CD1d to NKT cells identified that CD1d is capable of binding cellular phospholipids as phosphatidylinositol and glycosylphosphatidylinositol, and that this association is accomplished in the endoplasmic reticulum, but this complex is inert to the immune system [73].

Joyce [73] has suggested that CD1d molecules could function as 'sensor' molecules, sensing alterations in cellular lipid content. The presentation of a neo-self glycolipid, presumably induced by infectious assaults, such as viral and bacterial infections, activates NKT cells [73].

The CD1d molecule is expressed by a variety of cells: the thymocytes, activated T cells, B lymphocytes, professional antigen-presenting cells and also by intestinal epithelial cells [74–76]. Recently, the expression of CD1d in the skin was also elucidated. Keratinocytes, the major cell type of the epidermis have been shown to express CD1d, as well as sebocytes, dermal dendritic cells and endothelial cells [77]. Moreover, at least two forms of CD1d, one of 30 kD and the other of 47 kD, were found in keratinocytes by Western blot analysis [78].

The role of the CD1d molecule is to present glycolipid antigens to NKT cells [79]. These cells have the ability to

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rapidly secrete large amounts of cytokines following activation and without prior sensitization. NKT cell clones secrete type 1, type 2 or both types of cytokines, which could influence the differentiation of Th0 cells towards Th1 or Th2 cells, depending on the stimulus [80]. Recent studies have suggested that NKT cells also play an important role in cross-talk between components of the innate immune system as well as the adaptive immune system [81]. The rapid activation of these cells is characteristic of innate immunity and probably serves two purposes: to provide a first line of defense against pathogens and to orient the adaptive immune response in an adequate fashion with the nature of the pathogen. The role of CD1d in host defense has been suggested by studies performed in CD1ddeficient mice. Borrelia burgdorferi infection of CD1ddeficient mouse strains normally resistant to this pathogen resulted in arthritis, suggesting that CD1d is important in host defense against spirochetes [82]. CD1d-deficient mice also showed a significantly reduced clearance of Pseudomonas aeruginosa from the lung compared to wild-type mice [83]. CD1d has also been suggested to play a role in the host defense against Mycobacterium tuberculosis, Toxoplasma gondii, Listeria monocytogenes, Plasmodium fal*ciparum* and respiratory syncytial virus [84, 85].

CD1d expression is rapidly induced on keratinocytes in normal skin by physical trauma that disrupts barrier function [86]. This could represent an additional protective mechanism of the skin immune system.

CD1d-activated NKT cells have been suggested to play a role in the pathogenesis of several autoimmune diseases such as diabetes mellitus, rheumatoid arthritis, systemic sclerosis and psoriasis [87, 88].

Lipid antigen presentation in the skin by CD1dexpressing cells could be of major importance both in host defense against pathogens and in the pathogenesis of autoimmune disorders. The skin and especially the epidermis and the pilosebaceous unit are structures rich in lipids. The presence of glycolipids with modified structures could represent an important factor in the initiation and maintenance of immune responses. Bonish et al. [86] have recently shown that coculture of CD1d-positive keratinocytes with an NKT cell clone isolated from patients with psoriasis leads to activation and cytokine release, especially γ -interferon. This reveals that keratinocytes are able to function as nonconventional antigen-presenting cells and activate NKT cells by presenting lipid antigens with the help of CD1d molecules. Many pathogens also possess lipid antigens, which may represent potential antigens for presentation by CD1d-expressing cells to NKT cells.

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Fig. 1. Factors affecting cytokine release in the pilosebaceous unit of acne. TNF- α = Tumor necrosis factor α ; IFN- γ = γ -interferon; LTB₄ = leukotriene B₄.

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The two major cell types involved in the initiation of acne are the keratinocytes and the sebocytes. Since both cell types can participate in innate immunity, if *P. acnes* colonization occurs in the pilosebaceous unit, there is an increased possibility that PAMPs from *P. acnes* can stimulate the immune activity of these cells.

Expression of the LPS receptor CD14 was demonstrated in keratinocytes and sebocytes [66]. Hornemann et al. [89] showed that LPS can stimulate IL-8 secretion by human sebocytes but do not influence IL-1a expression at the mRNA and protein level. Interestingly, linoleic acid reduced the IL-8 secretion of sebocytes [90]. These results implicated the expression of CD14 molecules in human sebocytes and a competition of the proinflammatory activity of LPS by linoleic acid. CD14 expression was reduced in sebocytes by LPS and phorbol myristate acetate (PMA). Interestingly, both LPS and PMA enhanced IL-8 expression, while linoleic acid was unable to inhibit the stimulatory LPS effect. IL-1a expression in SZ95 sebocytes was barely affected by LPS, PMA or linoleic acid. PMA, but not LPS and linoleic acid, stimulated total sebaceous lipids. Therefore, LPS and PMA are likely to serve as proinflammatory signals in human sebocytes inducing CD14-dependent and independent IL-8 expression, respectively. Linoleic acid was unable to antagonize the proinflammatory activity of LPS [66, 91].

It was demonstrated that *Staphylococcus aureus* induces the production and release of IL-8 by keratinocytes in a TLR₂-dependent manner. It was also shown that LPS induced IL-8 expression and release by keratinocytes in a TLR₄-dependent manner [63]. The expressions of TLR₂ and TLR₄ in cultured human sebocytes were demonstrated by Oeff et al. [65]. Upon ligand binding, TLRs activate NF- κ B, which promotes the transcription of proinflammatory cytokines, inducible enzymes, adhesion molecules and antimicrobial peptides. The effect of LPS on TLR expression was also examined in sebocytes; LPS increased TLR₂ expression and was without significant effect on TLR₄ [92].

Recent investigations on the recognition of *P. acnes* have shown that this pathogen induces expression of IL-8 in a TLR₂- and NF- κ B-dependent manner [27, 67]. CD14 is also involved in the *P.-acnes*-induced upregulation of IL-8 expression. Confirmation of the presence of receptors and signaling pathways involved for the recognition of *P. acnes* in cells of the pilosebaceous unit supports the widely accepted view that inflammation in acne vulgaris may be induced or at least sustained by an immunological reaction to extracellular products of *P. acnes*.

The mechanisms by which *P. acnes* induces immune responses in keratinocytes and sebocytes are possibly similar to those of other gram-positive bacteria. Two major components of the gram-positive cell wall that had been described to induce immune responses are peptidoglycan and lipoteichoic acid. The former induces the production of IL-8 in a TLR₂-dependent manner in keratinocytes, while the effect of the latter on keratinocytes is mediated by TLR₄ [63, 64]. In sebocytes lipoteichoic acid suppressed the expression of TLR₂ and TLR₄ [92].

Keratinocytes are biochemically very active cells, which are able to produce various kinds of cytokines, growth factors and antimicrobial compounds. They contain a significant amount of intracellular IL-1 α , which is released after physical stress. IL-1 α is known to amplify the immune response. Guy and Kealey [93] have investigated whether IL-1 α could be responsible for the induction of hypercornification of the infundibulum in a cultivated human pilosebaceous model system. They found that addition of 1 ng/ml IL-1 α to the pilosebaceous infundibulum maintained in keratinocytes serum-free media resulted in hypercornification of the infundibulum similar to that seen in comedones [93].

Walters et al. [94] have investigated the ability of skin commensal microorganisms to modulate IL-1 α release in cultured human keratinocytes. According to their results, low levels of IL-1 α are released constitutively by cultured keratinocytes, but the release of a biologically active form of this cytokine was not increased by microorganisms. They also found that there were individual differences in sensitivity to microbes between individuals [94]. Similarly to keratinocytes, cultured human sebocytes were also shown to produce IL-1 α , but its expression was barely influenced by LPS [89].

Increased IL-8 release could initiate increased antigen presentation since IL-8 is known to upregulate the expression of HLA-DR in cultured human keratinocytes [95]. IL-1 α has no effect on the expression of intercellular adhesion molecules 1 and 3 or HLA-DR in infundibular keratinocytes [13].

The finding of Vega et al. [96] that adapalene, a synthetic retinoid used for the topical treatment of acne, exerts its beneficial effects at least partially through downregulation of the expression of TLR₂, one of the main receptors involved in the recognition of *P. acnes*, may suggest that microorganisms present in acne lesions could make a significant contribution to the promotion and sustainment of inflammation in acne. Adapalene has no effect on the expression of other myeloid markers such as CD14, which is also involved in the recognition of *P. acnes* and on the expression of HLA-DR.

Beside *P. acnes*, abnormal lipids may also affect the activity of keratinocytes and sebocytes. These lipids could

directly influence the proliferation and differentiation status of these cells and the release of various cytokines [10]. Keratinocytes and sebocytes may function as nonprofessional antigen-presenting cells. CD1d, which is expressed by both types of cells [77, 86] may present abnormal lipids from the pilosebaceous unit and stimulate NKT cells. After activation these cells are capable of secreting cytokines, which further activate conventional T cells and cells of the innate immune system [79]. These cytokines could also stimulate keratinocytes and sebocytes contributing to their abnormal proliferation and differentiation (fig. 1).

Conclusion

The SIS plays an important role in the pathogenesis of acne. Beside the implication of immune mechanisms in the induction and maintenance of inflammatory lesions, the SIS may have a role in the development of early, noninflammatory lesions of acne. New aspects of the immune functions of keratinocytes and sebocytes, such as pathogen recognition performed by PRRs and lipid antigen presentation by CD1d molecules, should be considered as possible pathogenetic factors in the development of acne lesions.

References

- Norris JF, Cunliffe WJ: A histological and immunocytochemical study of early acne lesions. Br J Dermatol 1988;118:651–659.
- 2 Cunliffe WJ: The sebaceous gland and acne 40 years on. Dermatology 1998;196:9–15.
- 3 Aldana OL, Holland DB, Cunliffe WJ: Variation in pilosebaceous duct keratinocyte proliferation in acne patients. Dermatology 1998; 196:98–99.
- 4 Gollnick HP, Zouboulis CC, Akamatsu H, Kurokawa I, Schulte A: Pathogenesis and pathogenesis related treatment of acne. J Dermatol 1991;18:489–499.
- 5 Knaggs HE, Holland DB, Morris C, Wood EJ, Cunliffe WJ: Quantification of cellular proliferation in acne using the monoclonal antibody Ki-67. J Invest Dermatol 1994;102:89–92.
- 6 Cunliffe WJ, Holland DB, Clark SM, Stables GI: Comedogenesis: Some new aetiological, clinical and therapeutic strategies. Br J Dermatol 2000;142:1084–1091.
- 7 Hughes BR, Morris C, Cunliffe WJ, Leigh IM: Keratin expression in pilosebaceous epithelia in truncal skin of acne patients. Br J Dermatol 1996;134:247–256.

- 8 Downing DT, Stewart ME, Wertz PW, Strauss JS: Essential fatty acids and acne. J Am Acad Dermatol 1986;14:221–225.
- 9 Kanaar P: Follicular-keratogenic properties of fatty acids in the external ear canal of the rabbit. Dermatologica 1971;142:14–22.
- 10 Wakita H, Matsushita K, Nishimura K, Tokura Y, Furukawa F, Takigawa M: Sphingosylphosphorylcholine stimulates proliferation and upregulates cell surface-associated plasminogen activator activity in cultured human keratinocytes. J Invest Dermatol 1998;110:253–258.
- 11 Toyoda M, Morohashi M: Pathogenesis of acne. Med Electron Microsc 2001;34:29–40.
- 12 Knaggs HE, Hughes BR, Morris C, Wood EJ, Holland DB, Cunliffe WJ: Immunohistochemical study of desmosomes in acne vulgaris. Br J Dermatol 1994;130:731–737.
- 13 Guy R, Kealey T: Modelling the infundibulum in acne. Dermatology 1998;196:32–37.
- 14 Piérard GE: Follicule to follicule heterogeneity of sebum excretion. Dermatologica 1986;173: 61–65.
- 15 Thiboutot D, Gilliland K, Light J, Lookingbill D: Androgen metabolism in sebaceous glands from subjects with and without acne. Arch Dermatol 1999;135:1041–1045.

- 16 Thiboutot D, Harris G, Iles V, Cimis G, Gilliland K, Hagari S: Activity of the type 1 5alpha-reductase exhibits regional differences in isolated sebaceous glands and whole skin. J Invest Dermatol 1995;105:209–214.
- 17 Leyden JJ, McGinley KJ, Vowels B: Propionibacterium acnes colonization in acne and nonacne. Dermatology 1998;196:55–58.
- 18 McGinley KJ, Webster GF, Ruggieri MR, Leyden JJ: Regional variations in density of cutaneous propionibacteria: Correlation of *Propionibacterium acnes* populations with sebaceous secretion. J Clin Microbiol 1980;12: 672–675.
- 19 Leeming JP, Holland KT, Cunliffe WJ: The microbial colonization of inflamed acne vulgaris lesions. Br J Dermatol 1988;118:203– 208.
- 20 Burkhart CG, Burkhart CN, Lehmann PF: Acne: A review of immunologic and microbiologic factors. Postgrad Med J 1999;75:328– 331.

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- 21 Ashbee HR, Muir SR, Cunliffe WJ, Ingham E: IgG subclasses specific to *Staphylococcus epidermidis* and *Propionibacterium acnes* in patients with acne vulgaris. Br J Dermatol 1997; 136:730–733.
- 22 Puhvel SM, Amirian D, Weintraub J, Reisner RM: Lymphocyte transformation in subjects with nodulocystic acne. Br J Dermatol 1977; 97:205–211.
- 23 Kersey P, Sussman M, Dahl M: Delayed skin test reactivity to *Propionibacterium acnes* correlates with severity of inflammation in acne vulgaris. Br J Dermatol 1980;103:651–655.
- 24 Jappe U, Ingham E, Henwood J, Holland KT: Propionibacterium acnes and inflammation in acne: P. acnes has T-cell mitogenic activity. Br J Dermatol 2002;146:202–209.
- 25 Vowels BR, Yang S, Leyden JJ: Induction of proinflammatory cytokines by a soluble factor of *Propionibacterium acnes*: Implications for chronic inflammatory acne. Infect Immun 1995;63:3158–3165.
- 26 Scott DG, Cunliffe WJ, Gowland G: Activation of complement – A mechanism for the inflammation in acne. Br J Dermatol 1979; 101:315–320.
- 27 Chen Q, Koga T, Uchi H, Hara H, Terao H, Moroi Y, Urabe K, Furue M: *Propionibacterium acnes*-induced IL-8 production may be mediated by NF-kappaB activation in human monocytes. J Dermatol Sci 2002;29:97–103.
- 28 Kemeny L, Ruzicka T: Lipid mediators. Hautarzt 1994;45:582–591.
- 29 Crooks SW, Stockley RA: Leukotriene B₄. Int J Biochem Cell Biol 1998;30:173–178.
- 30 Zouboulis CC: Exploration of retinoid activity and the role of inflammation in acne: Issues affecting future directions for acne therapy. J Eur Acad Dermatol Venereol 2001; 15(suppl 3):63–67.
- 31 Alestas T, Fimmel S, Beutler C, Hakij N, Chen W, Muller-Decker K, Zouboulis CC: Presence of the arachidonic acid proinflammatory pathway in human sebocytes in vitro (abstract). J Invest Dermatol 2002;119:737.
- 32 Zouboulis CC: Is acne vulgaris a genuine inflammatory disease? Dermatology 2001;203: 277–279.
- 33 Feldhaus MJ, Weyrich AS, Zimmerman GA, McIntyre TM: Ceramide generation in situ alters leukocyte cytoskeletal organization and beta 2-integrin function and causes complete degranulation. J Biol Chem 2002;277:4285– 4293.
- 34 Tokura Y, Wakita H, Seo N, Furukawa F, Nishimura K, Takigawa M: Modulation of Tlymphocyte proliferation by exogenous natural ceramides and sphingosylphosphorylcholine. J Invest Dermatol Symp Proc 1999;4:184–189.
- 35 Yamamoto A, Takenouchi K, Ito M: Impaired water barrier function in acne vulgaris. Arch Dermatol Res 1995;287:214–218.
- 36 Seiffert K, Zouboulis CC, Seltmann H, Granstein RD: Expression of neuropeptide receptors by human sebocytes and stimulatory effect of their agonists on cytokine production (abstract). Horm Res 2002;53:102.

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- 37 Böhm M, Zouboulis CC, Luger TA: Evidence for expression of mu-opoid receptors on human sebocytes which mediate paracrine and/or endocrine action of beta-endorphin (abstract). J Invest Dermatol 2002;119:278.
- 38 Bohm M, Schiller M, Stander S, Seltmann H, Li Z, Brzoska T, Metze D, Schioth HB, Skottner A, Seiffert K, Zouboulis CC, Luger TA: Evidence for expression of melanocortin-1 receptor in human sebocytes in vitro and in situ. J Invest Dermatol 2002;118:533–539.
- 39 Zouboulis CC, Seltmann H, Hiroi N, Chen W, Young M, Oeff M, Scherbaum WA, Orfanos CE, McCann SM, Bornstein SR: Corticotropinreleasing hormone: An autocrine hormone that promotes lipogenesis in human sebocytes. Proc Natl Acad Sci USA 2002;99:7148–7153.
- 40 Bohm M, Luger TA: The pilosebaceous unit is part of the skin immune system. Dermatology 1998;196:75–79.
- 41 Bos JD, Zonneveld I, Das PK, Krieg SR, van der Loos CM, Kapsenberg ML: The skin immune system (SIS): Distribution and immunophenotype of lymphocytes subpopulations in normal human skin. J Invest Dermatol 1987; 88:569–573.
- 42 Takeuchi O, Hoshino K, Kawai T, Sanjo H, Takada H, Ogawa T, Takeda K, Akira S: Differential roles of TLR2 and TLR4 in recognition of gram-negative and gram-positive bacterial cell wall components. Immunity 1999;11: 443–451.
- 43 Chow JC, Young DW, Golenbock DT, Christ WJ, Gusovsky F: Toll-like receptor-4 mediates lipopolysaccharide-induced signal transduction. J Biol Chem 1999;274:10689–10692.
- 44 Hoshino K, Takeuchi O, Kawai T, Sanjo H, Ogawa T, Takeda Y, Takeda K, Akira S: Cutting edge: Toll-like receptor 4 (TLR4)-deficient mice are hyporesponsive to lipopolysaccharide: Evidence for TLR4 as the Lps gene product. J Immunol 1999;162:3749–3752.
- 45 Arbour NC, Lorenz E, Schutte BC, Zabner J, Kline JN, Jones M, Frees K, Watt JL, Schwartz DA: TLR4 mutations are associated with endotoxin hyporesponsiveness in humans. Nat Genet 2000;25:187–191.
- 46 Yoshimura A, Lien E, Ingalls RR, Tuomanen E, Dziarski R, Golenbock D: Cutting edge: Recognition of gram-positive bacterial cell wall components by the innate immune system occurs via Toll-like receptor 2. J Immunol 1999; 163:1–5.
- 47 Schwandner R, Dziarski R, Wesche H, Rothe M, Kirschning CJ: Peptidoglycan- and lipoteichoic acid-induced cell activation is mediated by toll-like receptor 2. J Biol Chem 1999;274: 17406–17409.
- 48 Brightbill HD, Libraty DH, Krutzik SR, Yang RB, Belisle JT, Bleharski JR, Maitland M, Norgard MV, Plevy SE, Smale ST, Brennan JP, Bloom BR, Godowski PJ, Modlin RL: Host defense mechanisms triggered by microbial lipoproteins through toll-like receptors. Science 1999;285:732–736.
- 49 Medzhitov R, Preston-Hurlburt P, Kopp E, Stadlen A, Chen C, Ghosh S, Janeway CA Jr: MyD88 is an adaptor protein in the hToll/IL-1 receptor family signaling pathways. Mol Cell 1998;2:253–258.

- 50 Medzhitov R, Janeway C: The Toll receptor family and microbial recognition. Trends Microbiol 2000;8:452–456.
- 51 Csato M, Bozoky B, Hunyadi J, Dobozy A: *Candida albicans* phagocytosis by separated human epidermal cells. Arch Dermatol Res 1986;279:136–139.
- 52 Csato M, Kenderessy AS, Dobozy A: Enhancement of *Candida albicans* killing activity of separated human epidermal cells by ultraviolet radiation. Br J Dermatol 1987;116:469–475.
- 53 Kirkpatrick CH, Rich RR, Bennett JE: Chronic mucotaneous candidiasis: Model-building in cellular immunity. Ann Intern Med 1971;74: 955–978.
- 54 Csato M, Kenderessy AS, Dobozy A: Enhancement of *Candida albicans* killing activity of separated human epidermal cells by alphamelanocyte stimulating hormone (letter). Br J Dermatol 1989;121:145–147.
- 55 Csato M, Kenderessy AS, Judak R, Dobozy A: Inflammatory mediators are involved in the *Candida albicans* killing activity of human epidermal cells. Arch Dermatol Res 1990;282: 348–350.
- 56 Szolnoky G, Bata-Csorgo Z, Kenderessy AS, Kiss M, Pivarcsi A, Novak Z, Nagy NK, Michel G, Ruzicka T, Marodi L, Dobozy A, Kemeny L: A mannose-binding receptor is expressed on human keratinocytes and mediates killing of *Candida albicans*. J Invest Dermatol 2001;117: 205–213.
- 57 Bruch-Gerharz D, Ruzicka T, Kolb-Bachofen V: Nitric oxide in human skin: Current status and future prospects. J Invest Dermatol 1998; 110:1–7.
- 58 Frohm M, Agerberth B, Ahangari G, Stahle-Backdahl M, Liden S, Wigzell H, Gudmundsson GH: The expression of the gene coding for the antibacterial peptide LL-37 is induced in human keratinocytes during inflammatory disorders. J Biol Chem 1997;272:15258–15263.
- 59 Wiedow O, Harder J, Bartels J, Streit V, Christophers E: Antileukoprotease in human skin: An antibiotic peptide constitutively produced by keratinocytes. Biochem Biophys Res Commun 1998;248:904–909.
- 60 Harder J, Bartels J, Christophers E, Schroder JM: Isolation and characterization of human (beta)-defensin-3, a novel human inducible peptide antibiotic. J Biol Chem 2001;276: 5707–5713.
- 61 Kenderessy AS, Kemeny L, Dobozy A: Nitric oxide is involved in the *Candida albicans* killing activity of keratinocytes (abstract). J Invest Dermatol 1996;107:452.
- 62 Harder J, Bartels J, Christophers E, Schroder JM: A peptide antibiotic from human skin (letter). Nature 1997;387:861.
- 63 Pivarcsi A, Réthi B, Szell M, Kenderessy AS, Beer ZS, Bata-Csorgo Z, Magocsi M, Rajnavölgyi É, Dobozy A, Kemeny L: Toll-like receptors 2 and 4 are expressed on human keratinocytes and mediate the killing of pathogens (abstract). J Invest Dermatol 2001;117:803.

- 64 Song PI, Park YM, Abraham T, Harten B, Zivony A, Neparidze N, Armstrong CA, Ansel JC: Human keratinocytes express functional CD14 and toll-like receptor 4. J Invest Dermatol 2002;119:424–432.
- 65 Oeff M, Seltmann H, Hakiy N, Bognanoff B, Nastos A, Walters R, Bornstein SR, Zouboulis CC: Toll-like receptor 2 and 4-dependent regulation of inflammatory signaling in human sebocytes (abstract). J Invest Dermatol 2002;119: 736.
- 66 Seltmann H, Zouboulis CC: Human sebocytes express CD14 molecules and their IL8 production induced by both CD14-dependent and independent pathway (abstract). J Invest Dermatol 2001;117:804.
- 67 Kim J, Ochoa MT, Krutzik SR, Takeuchi O, Uematsu S, Legaspi AJ, Brightbill HD, Holland D, Cunliffe WJ, Akira S, Sieling PA, Godowski PJ, Modlin RL: Activation of toll-like receptor 2 in acne triggers inflammatory cytokine responses. J Immunol 2002;169:1535– 1541.
- 68 Chronnell CM, Ghali LR, Ali RS, Quinn AG, Holland DB, Bull JJ, Cunliffe WJ, McKay IA, Philpott MP, Muller-Rover S: Human beta defensin-1 and -2 expression in human pilosebaceous units: Upregulation in acne vulgaris lesions. J Invest Dermatol 2001;117:1120–1125.
- 69 Zouboulis CC, Xia L, Akamatsu H, Seltmann H, Fritsch M, Hornemann S, Ruhl R, Chen W, Nau H, Orfanos CE: The human sebocyte culture model provides new insights into development and management of seborrhoea and acne. Dermatology 1998;196:21–31.
- 70 Zouboulis CC, Seltmann H, Neitzel H, Orfanos CE: Establishment and characterization of an immortalized human sebaceous gland cell line (SZ95). J Invest Dermatol 1999;113:1011– 1020.
- 71 Porcelli SA, Modlin RL: The CD1 system: Antigen-presenting molecules for T cell recognition of lipids and glycolipids. Annu Rev Immunol 1999;17:297–329.
- 72 Nieda M, Nicol A, Koezuka Y, Kikuchi A, Takahashi T, Nakamura H, Furukawa H, Yabe T, Ishikawa Y, Tadokoro K, Juji T: Activation of human Valpha24NKT cells by alpha-glycosylceramide in a CD1d-restricted and Valpha24TCR-mediated manner. Hum Immunol 1999;60:10–19.
- 73 Joyce S: CD1d and natural T cells: How their properties jump-start the immune system. Cell Mol Life Sci 2001;58:442–469.
- 74 Exley M, Garcia J, Wilson SB, Spada F, Gerdes D, Tahir SM, Patton KT, Blumberg RS, Porcelli S, Chott A, Balk SP: CD1d structure and regulation of human thymocytes, peripheral blood T cells, B cells and monocytes. Immunology 2000;100:37–47.

- 75 Canchis PW, Bhan AK, Landau SB, Yang L, Balk SP, Blumberg RS: Tissue distribution of the non-polymorphic major histocompatibility complex class I-like molecule, CD1d. Immunology 1993;80:561–565.
- 76 Somnay-Wadgaonkar K, Nusrat A, Kim HS, Canchis WP, Balk SP, Colgan SP, Blumberg RS: Immunolocalization of CD1d in human intestinal epithelial cells and identification of a beta2-microglobulin-associated form. Int Immunol 1999;11:383–392.
- 77 Koreck A, Szony BJ, Farkas A, Bata Zs, Kemeny L, Drugarin D, Dobozy A: Fokozott CD1d kifejez dés psoriasisos b rben (increased CD1d expression in psoriatic skin). B rgyógy Vener Szle 2001;77:197–200.
- 78 Koreck A, Szell M, Pivarcsi A, Bata-Csorgo Z, Dobozy A, Kemeny L: Distinct isoforms of CD1d are expressed in keratinocytes in a proliferation-differentiation dependent manner (abstract). J Invest Dermatol 2002;119:748.
- 79 Godfrey DI, Hammond KJ, Poulton LD, Smyth MJ, Baxter AG: NKT cells: Facts, functions and fallacies. Immunol Today 2000;21: 573–583.
- 80 Chen H, Paul WE: Cultured NK1.1+ CD4+ T cells produce large amounts of IL-4 and IFNgamma upon activation by anti-CD3 or CD1. J Immunol 1997;159:2240–2249.
- 81 Carnaud C, Lee D, Donnars O, Park SH, Beavis A, Koezuka Y, Bendelac A: Cutting edge: Cross-talk between cells of the innate immune system: NKT cells rapidly activate NK cells. J Immunol 1999;163:4647–4650.
- 82 Kumar H, Belperron A, Barthold SW, Bockenstedt LK: Cutting edge: CD1d deficiency impairs murine host defense against the spirochete, *Borrelia burgdorferi*. J Immunol 2000; 165:4797–4801.
- 83 Nieuwenhuis EE, Matsumoto T, Exley M, Schleipman RA, Glickman J, Bailey DT, Corazza N, Colgan SP, Onderdonk AB, Blumberg RS: CD1d-dependent macrophage-mediated clearance of *Pseudomonas aeruginosa* from lung. Nat Med 2002;8:588–593.
- 84 Actor JK, Olsen M, Hunter RLJ, Geng YJ: Dysregulated response to mycobacterial cord factor trehalose-6,6'-dimycolate in CD1D-/mice. J Interferon Cytokine Res 2001;21: 1089-1096.
- 85 Schofield L, McConville MJ, Hansen D, Campbell AS, Fraser-Reid B, Grusby MJ, Tachado SD: CD1d-restricted immunoglobulin G formation to GPI-anchored antigens mediated by NKT cells. Science 1999;283:225–229.
- 86 Bonish B, Jullien D, Dutronc Y, Huang BB, Modlin R, Spada FM, Porcelli SA, Nickoloff BJ: Overexpression of CD1d by keratinocytes in psoriasis and CD1d-dependent IFN-gamma production by NK-T cells. J Immunol 2000; 165:4076–4085.

- 87 Yanagihara Y, Shiozawa K, Takai M, Kyogoku M, Shiozawa S: Natural killer (NK) T cells are significantly decreased in the peripheral blood of patients with rheumatoid arthritis (RA). Clin Exp Immunol 1999;118:131–136.
- 88 Wilson SB, Kent SC, Patton KT, Orban T, Jackson RA, Exley M, Porcelli S, Schatz DA, Atkinson MA, Balk SP, Strominger JL, Hafler DA: Extreme Th1 bias of invariant Valpha24JalphaQ T cells in type 1 diabetes. Nature 1998;391:177–181.
- 89 Hornemann S, Seltmann H, Kodelja V, Orfanos CE, Zouboulis CC: Interleukin 1α mRNA and protein are expressed in cultured human sebocytes at steady-state and their levels are barely influenced by lipopolysaccharides (abstract). J Invest Dermatol 1997;108:382.
- 90 Seltmann H, Hornemann S, Orfanos CE, Zouboulis CC: Linoleic acid induces accumulation of neutral lipids in undifferentiated human sebocytes and reduce spontaneous IL-8 secretion (abstract). Arch Dermatol Res 1999;291:181.
- 91 Seltmann H, Oeff M, Zouboulis CC: CD14 expression in human sebocytes and IL8 regulation by lipopolysaccharides, phorbol myristate acetate and linoleic acid (abstract). Arch Dermatol Res 2002;294:33.
- 92 Oeff M, Seltmann H, Hakiy N, Bogdanoff B, Nastos A, Walters R, Fimmel S, Bornstein SR, Zouboulis CC: Differential modulation of Tolllike receptor 2 and 4 expression in human sebocytes (abstract). J Invest Dermatol 2002;119: 736.
- 93 Guy R, Kealey T: The effects of inflammatory cytokines on the isolated human sebaceous infundibulum. J Invest Dermatol 1998;110:410– 415.
- 94 Walters CE, Ingham E, Eady EA, Cove JH, Kearney JN, Cunliffe WJ: In vitro modulation of keratinocyte-derived interleukin-1 alpha (IL-1 alpha) and peripheral blood mononuclear cell-derived IL-1 beta release in response to cutaneous commensal microorganisms. Infect Immun 1995;63:1223–1228.
- 95 Kemeny L, Kenderessy AS, Ocsovszky I, Michel G, Ruzicka T, Dobozy A: Interleukin-8 induces HLA-DR expression on cultured human keratinocytes via specific receptors. Int Arch Allergy Immunol 1995;106:351–356.
- 96 Vega B, Jomard A, Michel S: Regulation of human monocyte Toll-like receptor 2 (TLR2) expression by adapalene (abstract). J Eur Acad Dermatol Venereol 2002;16:123–124
- 97 Kopp EB, Medzhitov R: The Toll-receptor family and control of innate immunity. Curr Opin Immunol 1999;11:13–18.
- 98 Zhang G, Ghosh S: Toll-like receptor-mediated NF-kappaB activation: A phylogenetically conserved paradigm in innate immunity. J Clin Invest 2001;107:13–19.