Are microRNAs the Molecular Link Between Metabolic Syndrome and Alzheimer's Disease?

Juan F. Codocedo¹ · Juvenal A. Ríos¹ · Juan A. Godoy¹ · Nibaldo C. Inestrosa^{1,2,3,4,5}

Received: 8 December 2014 / Accepted: 29 April 2015 / Published online: 15 May 2015 © Springer Science+Business Media New York 2015

Abstract Alzheimer's disease (AD) is the most common cause of dementia in people over 65 years of age. At present, treatment options for AD address only its symptoms, and there are no available treatments for the prevention or delay of the disease process. Several preclinical and epidemiological studies have linked metabolic risk factors such as hypertension, obesity, dyslipidemia, and diabetes to the pathogenesis of AD. However, the molecular mechanisms that underlie this relationship are not fully understood. Considering that less than 1 % of cases of AD are attributable to genetic factors, the identification of new molecular targets linking metabolic risk factors to neuropathological processes is necessary for improving the diagnosis and treatment of AD. The dysregulation of microRNAs (miRNAs), small non-coding RNAs that regulate several biological processes, has been implicated in the development of different pathologies. In this review, we summarize some of the relevant evidence that points to the role of miRNAs in metabolic syndrome (MetS) and AD and propose

Nibaldo C. Inestrosa ninestrosa@bio.puc.cl

- ¹ Centro de Envejecimiento y Regeneración (CARE), Departamento de Biología Celular, Molecular, Facultad de Ciencias Biológicas, Pontificia Universidad Católica de Chile, Santiago, Chile
- ² Centre for Healthy Brain Ageing, School of Psychiatry, Faculty of Medicine, University of New South Wales, Sydney, Australia
- ³ Centro UC Síndrome de Down, Pontificia Universidad Católica de Chile, Santiago, Chile
- ⁴ Centro de excelencia en Biomedicina de Magallanes (CEBIMA), Universidad de Magallanes, Punta Arenas, Chile
- ⁵ CARE Biomedical Research Center, Pontificia Universidad Católica de Chile, Av. Alameda 340, Santiago, Chile

that miRNAs may be a molecular link in the complex relationship between both diseases.

Keywords Alzheimer's disease · Metabolic syndrome · microRNA · Lifestyle

Introduction

Alzheimer's disease (AD) is the most common type of dementia, in which the death or malfunction of neurons causes changes in memory, behavior, and cognition. In AD, this dysfunction results in an impaired ability to carry out such basic functions as walking and swallowing and is ultimately lethal [1]. At present, AD has no cure or preventive treatment [2, 3]. Late-onset AD (LOAD) occurs in individuals over the age of 65 and is reported in greater than 90 % of Alzheimer's cases [4]. Several of the risk factors associated with LOAD have been classified as genetic or non-genetic factors. The strongest genetic risk factor for AD is related to the apolipoprotein E (APOE) gene, which can confer an increased probability of developing LOAD. The presence of the $\varepsilon 4$ allele of the APOE gene not only dose-dependently increases the risk for AD but also lowers the age of onset [5]. However, many factors other than genetics contribute to the development of AD.

Preclinical evidence with experimental animal models suggests that the relationship between cardiometabolic risk factor and AD is robust and clear, including spontaneous-induced experimental type 2 diabetes (T2DM), leading to AD-like pathology [6–8], induction of hypertension by angiotensin II infusion, and promoting AD lesions [9], hypercholesterolemia [10], and obesity [11]. On the other hand, clinical and epidemiological evidence indicates that the clusters of metabolic syndrome (MetS) such as hypertension [12], dyslipidemia [13], obesity [14], and T2DM [15] in a separate manner promote development of mild cognitive impairment (MCI), dementia, and AD, even considering some contradictions between studies [16], given the difficulties of the homogenization of the sample populations.

Early work indicates that T2DM is associated with a twoto threefold increase in the relative risk for AD, independent of the risk for vascular dementia [17]. In a global manner, patients with MCI and MetS had a higher risk of progression to dementia compared with patients with MCI alone [18–20]. A growing body of epidemiological evidence has allowed the development of a pathophysiological model termed "metabolic-cognitive syndrome" that aims to understand the complex relationship between metabolic disorders and cognitive disturbances and thereby generate therapeutic strategies against MetS that could help prevent or ameliorate the cognitive disturbances [18, 21–23]. Despite a wealth of evidence supporting this relationship, the molecular mechanisms underlying metabolic-cognitive syndrome are not fully understood.

Considering that less than 1 % of cases of AD are attributable to genetic factors [1], the identification of new molecular targets linking metabolic risk factors to neuropathological processes is necessary for improving the diagnosis and treatment of the disease. As AD and MetS are multifactorial pathologies, multiple mechanisms have been proposed in base to the etiological overlap observed. In this context, it has been suggested a role for inflammation in the etiology of MCI and AD [24]. Oxidative stress could also precede the onset of clinical and pathological AD symptoms and MetS [25]. Amyloidosis is another interesting hypothesis which is common to diabetes mellitus and AD [26]. The amylin aggregates observed in the pancreas are morphologically similar to the amyloid fibrils of AD, and both present neurotoxicity [27] and contains APOE [28].

We previously discussed how deregulation in several signaling pathway, including leptin, angiotensin, Wnt, and others, could explain the interesting crossroad between MetS and AD [29]; Now, we want to expand this work view including the role of microRNAs (miRNAs) as a molecular link between MetS and AD.

miRNAs are small endogenous RNAs, approximately 22 nucleotides (nt) in length, that have emerged as important post-transcriptional regulators of different protein-coding genes [30, 31]. The study of miRNAs is relatively new, but these small molecules have an important impact in most biological processes. The development of techniques such as gene amplification, gene sequencing, and microarray analysis has allowed the expression profiles of a large number of miRNAs in human diseases, including metabolic disorders [32], obesity [33], T2DM [34], hypertension [35], and cardiovascular disease [36], to be established. Because several of those "metabolic miRNAs" have been shown to be altered in AD patients (Table 1), it is possible that miRNAs might constitute a molecular link that explains the functional

relationship between metabolic risk factors and the development of AD. Additionally, a series of preclinical studies show that the metabolic signals produced in peripheral tissues like insulin or leptin regulate the expression of miRNAs in several brain regions, and the metabolic distress observed in MetS alters the expression of miRNAs related to human AD.

Importantly, the understanding that the risk factors associated to MetS and AD are lifestyle dependent, such as poor nutrition and a lack of exercise, and that miRNAs respond to changes in diet and physical activity could open new therapeutic avenues to the prevention and diagnosis of AD.

MicroRNA Biogenesis and Function

Precursor forms of miRNAs, which are hundreds of nt in length, are processed to reach their mature forms, which in turn are subsequently loaded in the miRNA-induced silencing complex (miRISC). This finely orchestrated process may be subject to regulation at different levels to control the function of miRNAs and gene expression [38–40]. We review some basics of the miRNA biogenetic pathway and discuss the unique functional modalities of miRNAs that can help to understand their possible role as a molecular link between MetS and AD.

miRNA Transcription

The transcription of miRNA generates a primary miRNA transcript (pri-miRNA), from either individual genes containing their own promoters (intergenic) or intragenically from spliced portions of protein-coding genes (intronic or exonic). The transcription of intragenic miRNAs is dependent on the regulatory elements of the host gene promoter. For that reason, intronic miRNAs are normally co-transcribed with their host gene, and pri-miRNA results from the splicing of the host gene transcript precursor. In many cases, this co-transcription is related to the cooperative function of the gene host products and the miRNA [41, 42]. In other cases, the co-transcription results in the silencing of the host in a negative feedback loop [43].

miRNAs are almost exclusively transcribed by RNA polymerase II (RNA Pol II) and their expression is thus subject to the same types of transcriptional control as other cellular genes including RNA Pol II-associated transcription factors and epigenetic regulators. One of the best described examples of transcription factors that regulate miRNA expression corresponds to the p53 tumor suppressor protein that regulates the expression of stress response genes including miR-34 family as the most prevalent p53-induced miRNAs [44]. It is very interesting to note that p53/miR-34 axes are downregulated in cancer [45] but are upregulated in neuropathologies including AD [46–48] which is consistent with the inverse occurrence of both diseases [49–53]. In the same logic, the axis p53/miR-34 has been described to be upregulated in clusters of MetS such

Alzheimer	T2dm	Hypertension	Obesity	Fatty Liver
hsa-mir-21	hsa-mir-21	hsa-mir-21	hsa-mir-21	hsa-mir-21
hsa-mir-103a-1	hsa-mir-103a-1	hsa-mir-103a-1		
hsa-mir-103a-2	hsa-mir-103a-2	hsa-mir-103a-2		
hsa-mir-17		hsa-mir-17	hsa-mir-17	
hsa-mir-107	hsa-mir-107		hsa-mir-107	
hsa-mir-20a		hsa-mir-20a		
hsa-mir-146a	hsa-mir-146a			
hsa-mir-144	hsa-mir-144			
	hsa-mir-143		hsa-mir-143	
		hsa-mir-122		hsa-mir-122
hsa-mir-106b	hsa-mir-486	hsa-mir-1-1	hsa-mir-132	hsa-mir-451
hsa-mir-101-1	hsa-mir-20b	hsa-mir-1-2	hsa-mir-18a	hsa-mir-10b
hsa-mir-590	hsa-mir-24-1	hsa-mir-133a-1	hsa-mir-30e	hsa-mir-34a
hsa-mir-128-1	hsa-mir-24-2	hsa-mir-133a-2	hsa-mir-146b	
hsa-mir-128-2	hsa-mir-15a	hsa-mir-155	hsa-mir-221	
hsa-mir-137	hsa-mir-126	Hsa-mir-208b		
hsa-mir-181c	hsa-mir-191	hsa-mir-204		
hsa-mir-9-1	hsa-mir-197	hsa-mir-637		
hsa-mir-9-2	hsa-mir-223	hsa-mir-133a-1		
hsa-mir-9-3	hsa-mir-320a	hsa-mir-637		
hsa-mir-137	hsa-mir-483	hsa-mir-296		
hsa-mir-181c	hsa-mir-99a	Hsa-mir-133b		
hsa-mir-29a		hsa-let-7e		
hsa-mir-29b-1		hsa-mir-328		
hsa-mir-29b-2		hsa-mir-150		
hsa-mir-124-1		hsa-mir-424		
hsa-mir-124-2		hsa-mir-503		
hsa-mir-124-3		hsa-mir-145		
hsa-mir-125b-1				
hsa-mir-125b-2				
hsa-mir-195				
hsa-mir-153-1				
hsa-mir-153-2				
hsa-mir-34c				

Table 1 Human miRNAs associated to AD and component of MetS

miRNAs associated to each disease were obtained from HMDD v2.0 which is a database that curated experiment-supported evidence for human microRNA (miRNA) and disease associations [37]. In red, miR-21, which is associated to AD and all the diseases of MetS. In blue, miRNAs associated to AD and other two components of MetS. In green, miRNAs associated to AD and one component of MetS. In yellow, miRNAs associate between components of MetS but not with AD. In white, miRNAs without association between AD and MetS

as non-alcoholic fatty liver disease (NAFLD) [54] which support the idea that regulation of miRNAs is part of the

pathological mechanism that associates the occurrence of MetS and AD (Fig. 1).

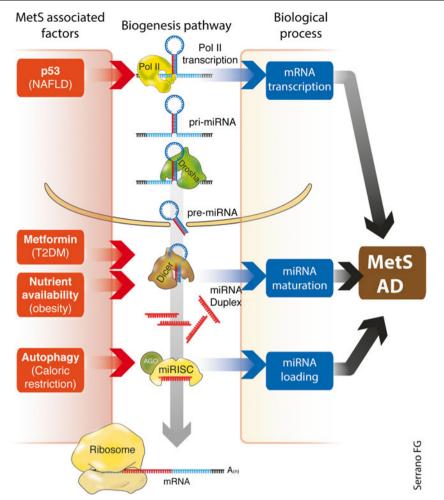


Fig. 1 MiRNA biogenesis and their regulation with MetS-associated factors. The miRNA biogenesis pathway produces pri-miRNA transcripts by RNA polymerase II (*Pol II*) from miRNA genes. Next, the Drosha microprocessor complex processes pri-miRNA transcripts into pre-miRNAs. Pre-miRNAs are exported from the nucleus via exportin 5 and subsequently cleaved by Dicer, and the miRNA/miRNA* duplex is unwound via Argonaute (*AGO*) and loaded into the miRNA-induced silencing complex (*miRISC*). The binding of target mRNAs to miRNAs in RISC is followed by the inhibition of translation and/or mRNA degradation. The miRNA biogenesis pathway may be subject to regulation at different levels to control the function of miRNAs and thus gene expression. In the figure, we show some examples of environmental (external)

miRNA Processing

For most miRNAs, the processing of pri-miRNAs to the mature form involves the sequential cropping of nuclear and cytosolic complexes in a process known as the canonical pathway. Once transcribed, the pri-miRNA are recognized by the nuclear microprocessor complex, which generates a shorter stem loop precursor-miRNA (pre-miRNA) that is ~60–70-nt long [55, 56]. The microprocessor complex is composed of two core proteins, DiGeorge Syndrome Critical Region 8 (DGCR8), which contains a RNA-binding domain that recognizes the pri-miRNA and binds to Drosha, a class II RNase III enzyme that cleaves double-stranded RNA in a staggered

factors related to MetS that control the expression of miRNA levels through different mechanisms. At the transcription level, the transcription factor p53 controls the expression of miR-34 cluster in response to a diet that generates non-alcoholic fatty liver disease (*NAFLD*). At the processing level, metabolic interventions like fasting or obesity regulate Dicer expression, thereby increasing the processing and generation of mature miRNAs. Finally, at the miRNA loading level, metabolic alterations that regulate the autophagy process affect the stability of AGO and the transcriptional repression of several miRNA targets. The biological consequence or output of this regulation is related to MetS and possibly to AD progression

manner and creates a 2-nt overhang on the 3' end of its products [57]. This overhang is recognized by exportin 5, which transports the pre-miRNA to the cytoplasm in a GTPdependent fashion [58–60]. In the cytoplasm, the premiRNA is processed by Dicer, a class III RNase III, into a ~22-nt-long miRNA/*miRNA duplex [61].

Any modulation of the miRNA processing proteins has a significant effect on the levels of miRNA produced by the canonical pathway and, consequently, on the biological processes in which they are involved. For example, mice deficient in Dicer do not survive beyond E8.5 [62], and tissue-specific Dicer deletion results in developmental defects in several organs, including the brain [63–66]. Conditional knockout (KO)

of Dicer in the brain of adult mice results in a progressive neurodegeneration, reduced brain size, neuroinflammation, apoptosis, hyperphosphorylation of endogenous tau protein, and impaired dendritic spine morphology similar to that observed in AD brains [63, 67, 68]. A recent study shows that the expression of Dicer in the hypothalamus is regulated by nutrient availability (Fig. 1). Fasting caused specific upregulation of Dicer mRNA levels, and obesity generates a decrease in Dicer expression [69]. Importantly, conditional deletion of Dicer in hypothalamic neurons resulted in obesity, hyperleptinemia, defective glucose metabolism, and alterations in the pituitary–adrenal axis which was paralleled by a neuron degenerative process [69].

These evidences suggest that any external factor that has the ability to regulate Dicer-dependent miRNA expression has the potential to affect neuronal pathological process associated to MetS and AD (Fig. 1).

miRISC Assembly

To control the expression of their targets, the 22-nt-long miRNA/*miRNA duplex needs to be associated with several proteins that conform the miRISC, where it is separated into its mature strand and its complementary strand, the latter of which is degraded in most cases [70]. The core proteins of the miRISC are Dicer, Argonaute (AGO), and TAR RNA binding protein (TRBP), a double-stranded RNA binding protein that is required for the recruitment of AGO to the miRNA bound by Dicer [71]. AGO proteins function as effectors by recruiting factors that induce translational repression, mRNA deadenylation, and mRNA decay [72].

Altering the expression or the activity of different components of miRISC alters miRNA-mediated silencing without affecting miRNA expression or processing. For example, in primary rat hippocampal neurons, AGO2 silencing generates an increase in the protein levels of the amyloid precursor

Fig. 2 Functional modalities of miRNAs. a miRNAs can control the transcription of tissue-specific or tissue-enriched targets, affecting biological process associated to brain or periphery, respectively. b miRNAs can control the transcription of targets ubiquitously expressed and probably have a more general role in several tissues and associated diseases. c Convergence of two or more miRNAs in the regulation of the same targets induces an increase in the transcriptional repression

🖄 Springer

protein (APP). However, no significant alteration of APP mRNA was observed, suggesting that the APP translation is regulated by an AGO2/miRNA pathway [73].

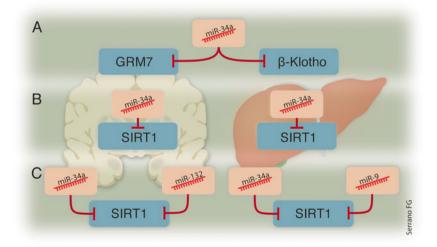
The stability and activity of AGO are regulated by several modifications including prolyl 4-hydroxylation, phosphorylation, poly(ADP-ribosyl)ation, proteasome-mediated degradation, and autophagy [74]. In particular, it has been reported that the homeostatic turnover of unloaded AGO proteins (miRNA-free) is mediated by autophagy [75, 76] which corresponds to a catabolic mechanism that involves degradation of unnecessary or dysfunctional cellular components through the actions of lysosomes [77]. Inhibition of autophagy at the level of autophagosome biogenesis generates an increase in the unloaded AGO levels with a decrease in the transcriptional repression of several miRNA targets [76].

These evidences have implications for human diseases linked to misregulated autophagy and miRNA levels, like MetS [78, 79] and AD [80, 81] (Fig. 1).

miRNA Target Recognition and Functional Modalities

In the miRISC, miRNA functions as a guide by base pairing with its target mRNAs. The initial bases of the miRNA sequence play an important role in defining the mRNA targets. Bases 2–7, known as the miRNA "seed," are responsible for the definition of target specificity [82]. With few exceptions, most of the miRNA recognition elements (MRE) are located within the 3'UTR of the targeted mRNA [31].

Considering that the miRNA–mRNA binding site is short, each miRNA has the potential to regulate many genes as its targets, while one gene may be targeted by many miRNAs [31]. In this sense, the same miRNA could regulate different biological process in function of their target context (Fig. 2a). For example, miR-34a regulate the postprandial metabolic responses by targeting the hepatic co-receptor β -Klotho [83] but in hippocampus contribute to the mood regulation through



the regulation of metabotropic glutamate receptor 7 [84]. Some of the targets are ubiquitously expressed and probably have a more general role in several tissues and associated diseases (Fig. 2b). For example, miR-34a has been described to be upregulated in both the liver of obese models [85] and the brain of aged mice [86]. Parts of the phenotypes, observed in both tissues, are related to the repression of their target. Silent information regulator 1 (SIRT1) which is a NADdependent deacetylase is critically involved in diverse cellular processes including MetS, cancer, aging, and neuroprotection [87, 88].

Another layer of complexity is related to the convergent action of different miRNAs on a common target (Fig. 2c). Continuing with the example of SIRT1, the repression mediated by miR-34a could be reinforced by additional miRNAs [89] including the brain-enriched miR-132 [90] or the action of miR-9 in brain [91] and peripheral tissues [92]. The convergence of two or more co-expressed miRNAs on a given target normally results in an increased degree of translational repression [93, 94].

All these characteristics are important to understand that the regulation of miRNAs is part of the mechanism underlying the development of human diseases. The identification of the noxious stimulus, risk factors, and signaling pathways that affect their biogenesis and function in both, brain, and peripheral tissues could contribute to understand the close association between AD and conditions like MetS.

miRNAs and Metabolic Syndrome

MetS is defined by a combination of metabolic disorders that are used to identify patients at increased risk for cardiovascular disease (CVD), T2DM, and all-cause mortality. The epidemiological concept of MetS originated from the observation that several metabolic risk factors often coincide in patients at high risk for CVD, namely visceral obesity, dyslipidemia, hypertension, NAFLD, and insulin resistance [95, 96]. The worldwide prevalence of MetS is increasing and is currently estimated to be approximately 25 %, largely as a result of the increasing rates of obesity and sedentary lifestyles [97].

Metabolic homeostasis is driven by gene expression programs that respond to cues such as cholesterol, lipids, leptin, glucose, and insulin. This response is mediated by a series of transcription factors including LXRs, sterol regulatory element-binding proteins (SREBPs), proliferator-activated receptors (PPARs), carbohydrate response element-binding protein (CHREBP), CCAAT-enhancer-binding protein (C/EBP), and forkhead box protein O1 (FOXO1). Recently, a series of studies has described the role of miRNAs in the control of metabolic homeostasis and metabolic disorders [32]. In this section, we highlight some examples of miRNAs deregulated in peripheral tissues and the consequential phenotype associated to MetS.

miRNA in Blood Lipid Disorder

Altered lipid levels and hypercholesterolemia represent a critical risk factor for cardiometabolic diseases prevalent in the developed world, such as MetS [32]. Additionally, alterations in cholesterol and lipid metabolism result in major neurodegenerative disorders, including AD [98]. Several miRNAs have been shown to post-transcriptionally regulate the expression of key genes involved in lipid homeostasis. One of the best characterized is miR-33, which acts in coordination with their host gene, SREBP, to increase the intracellular levels of cholesterol, primarily through the transcriptional repression of the ATP-binding cassette transporters 1 (ABCA1) [99], which mediates an essential step in the formation of high-density lipoprotein (HDL or "good cholesterol") [100]. Low HDL levels are strongly associated with risk for CVD [101]. Interestingly, a high-carbohydrate diet increases the levels of SREBP and miR-33 and decreases the levels of the ABCA1 transporter in liver of non-human primates. This could explain the low HDL levels observed in individuals suffering from MetS [102]. In addition to the regulation of genes involved in cholesterol efflux, the list of predicted miR-33a/b targets includes genes that promote fatty acid β -oxidation and negative regulators of SREBPs. This reflects an integrated network of functional interactions between the SREBP transcription factors and their intronic miRNAs to regulate cholesterol and lipid homeostasis [32]. It is important to note that in the brain, ABCA1 acts to lipidate APOE, which is essential for its interaction with A β and subsequent clearance, and a growing body of evidence suggests that ABCA1 plays a critical role in A β metabolism and accumulation [103]. Considering that ABCA1 transporter is regulated by several miRNAs that respond to metabolic signals is important to study the possible role of brain miRNAs in the regulation of ABCA1 transporter and other regulators of lipid homeostasis in the progression of AD.

miRNA in Obesity

Adipose tissue expansion generates insulin resistance and hyperlipidemia, thereby causing detrimental steatosis in other tissues. Several profiling studies reveal that miRNAs are differentially expressed in the adipose tissue of obese individuals and in mouse models of obesity [104]. For example, miR-103 and miR-107, which are upregulated in adipogenesis, are significantly downregulated in adipocytes from a mouse model of diet-induced obesity [105]. These miRNAs have been proposed to regulate the insulin response through the repression of caveolin 1 (CAV1), a critical regulator of the insulin receptor [106] (Fig. 3d). Another validated target of miR-103/107 is

Dicer (Fig. 3d), a critical enzyme in the biogenetic pathway of miRNAs (Fig. 1). Therefore, the dietary regulation of these miRNAs can generate additional modifications to the full collection of miRNAs in the genome (miRNome), thereby affecting other biological functions that contribute to the development of MetS. The same concept is suggested for AD, because as we will see later, miR-103/107 are downregulated in early stages of AD patients and in the brain of animal models supplemented with a high-fat diet (HFD).

miRNA in NAFLD

Being overweight or obese triggers pathologies such as NAFL D, which is defined as the excess accumulation of hepatic triglycerides and fatty acids and is identified as the hepatic manifestation of MetS [107]. The increased hepatic expression of miR-34a and the subsequently reduced expression of SIRT1 expression have been observed in the NAFLD model [85]. Because the use of natural activators of SIRT1, such as resveratrol, has proven to reduce hepatosteatosis in mice with diet-induced obesity [108], it has been proposed that miR-34a, through the regulation of SIRT1, could be a new therapeutic target for the treatment of NAFLD and other obesity-related diseases.

Another NAFLD-related miRNA is miR-122, which is the most abundant miRNA in the liver and regulates metabolic pathways such as cholesterol biosynthesis, fatty acid synthesis, and oxidation [109]. Studies in patients with NAFLD showed that hepatic and serum miR-122 levels were associated with hepatic steatosis and fibrosis [110–112]. Part of the mechanism whereby miR-122 participates in the development of NAFLD involves the repression of prolyl 4-hydroxylase subunit α -1 (P4H41), a key enzyme in collagen synthesis and, thus, the progression of fibrosis [113] (Fig. 3e).

miRNA and Insulin Signaling

Insulin resistance is a condition in which cells from different tissues do not respond adequately to the normal actions of the hormone insulin and is the most prevalent metabolic dysfunction of all of the MetS risk factors [114]. Many studies have established that miRNAs play significant roles in multiple aspects of insulin signaling [32]. For example, the co-expression of miR-124a and miR-375 regulates insulin exocytosis partly through the regulation of their target Myotrophin, a protein that participates in vesicular fusion [115].

Other miRNAs regulate insulin response by targeting proteins that promote signaling, such as miR-29 a/b, which targets caveolin 2 [116], or miR-126, which represses insulin receptor substrate 1 (IRS1), an adapter protein that plays a key role in signal transduction between the insulin receptor and the phosphoinositide 3-kinase pathway [117] (Fig. 3c).

miRNA and Hypertension

Arterial hypertension (HTA) is a common systemic condition when the individuals have consistent blood pressure measurements exceeding 140/90 mmHg [118]. To date, the main events and system involucrate in pathogenesis of HTA include salt-sensitive theory, abnormal activation of renin-angiotensin system (RAS), and renal fibrosis [119]. The role of miRNAs has been recently described in hypertension and also in other cardiovascular diseases as a putative biomarker of disease risk and progression [120-122]. For example, a recent study shows that hypertensive patients showed a significantly lower miR-9 and miR-126 expression levels compared with healthy controls [123]. Genetics studies show that some polymorphisms influence the expression of miRNAs and the disease pathogenesis. For example, the rs4705342 single nucleotide polymorphism in miR-143 promoter is associated with a decrease in their expression and genetic susceptibility to essential hypertension [124]. In the same line, single nucleotide polymorphisms (SNPs) located in the MRE in RAS genes can influence blood pressure and risk of myocardial infarction [125].

miRNAs can contribute to the development of HTA through the regulation of several component of the reninangiotensin system in vascular smooth muscle cells. The treatment with angiotensin II generates a robust change in the expression of several miRNAs including miR-483-3p which, in turn, is able to regulate the expression of four different RAS genes including angiotensinogen and angiotensin-converting enzyme 1 (ACE-1) proteins in vascular smooth muscle cells [126] (Fig. 3f).

MicroRNAs Studies in AD

AD is the most frequent form of dementia in the elderly. It is a neurodegenerative disorder that it is characterized by neuron destruction and synaptic loss, which result in a progressive decline in memory and other cognitive functions [2]. The two core histopathological hallmarks of AD are amyloid plaques (senile plaques) and neurofibrillary tangles. Amyloid plaques are formed by the deposition of the $A\beta$ peptides generated by proteolytic cleavage from the fulllength APP through the amyloidogenic pathway. In this process, APP is sequentially cleaved by β -secretase (BACE1) and the γ -secretase complex. The cleavage of APP by α secretase occurs within the AB peptide containing region, precluding the formation of this peptide (non-amyloidogenic pathway). The toxicity of the $A\beta$ peptide is dependent on its conformational state and length. Small oligomers of $A\beta$ can be more toxic than mature fibrils, and $A\beta_{1-42}$ peptide aggregates more readily than does $A\beta_{1-40}$ [127]. The ratio of these two isoforms is influenced by the pattern of cleavage from APP by the α , β , and γ secretases [128].

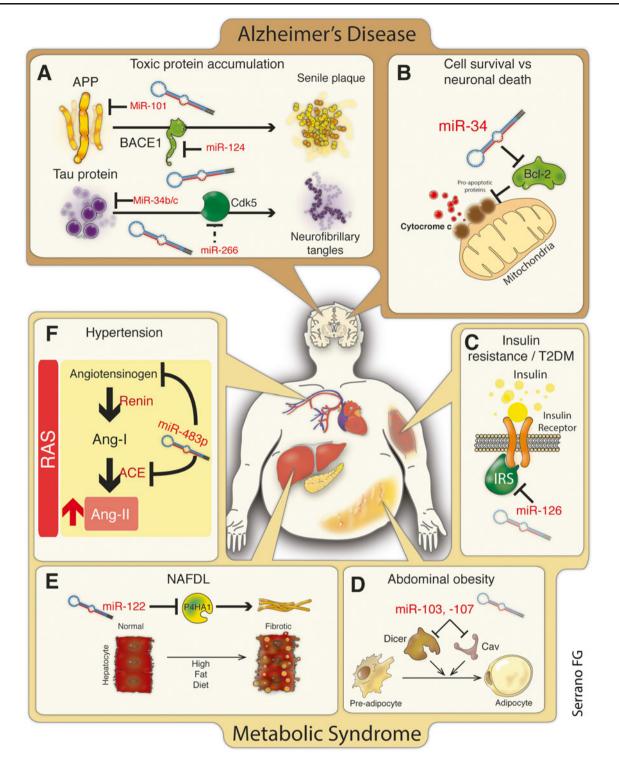
Another hallmark of AD is the neurofibrillary tangles formed by *tau*, a microtubule-associated protein. During neurodegeneration, tau becomes highly phosphorylated, causing conformational changes that result in disassociation from the microtubule aggregation [129]. Therefore, the balance between tau kinases and phosphatases is vital for fine-tuning the phosphorylation state of tau to regulate its biological activity.

Considering the complex molecular network that is involved in the development of AD and the poor diagnostic tools that are currently available, the identification of molecular clues with a clear relationship to the onset and development of the disease is fundamental. Thus, several studies have attempted to generate a miRNA profile in different models of AD. Studies carried out in tissue of human patients, AD animal models, or cell cultures have shown significant changes in subpopulations of miRNAs with respect to their matched controls; however, the changes observed in individual miRNAs have been inconsistent. One remarkable example is miR-9, a brain-enriched miRNA consistently altered in several profiling studies. However, post-mortem samples of human hippocampus have shown both increased [130] and decreased [131] miR-9 levels. Similar studies in the cortex have shown the same inconsistencies [132, 133]. These results may be explained by the use of different experimental approaches or the degree of disease progression. Interestingly, in primary mouse hippocampal neurons, treatment with $A\beta_{1-42}$ has been reported to decrease the levels of miR-9 [134, 135], suggesting that miR-9 deregulation in response to an insult such as A β may be an important factor that contributes to the cascade of events leading to AD.

Another important approach for understanding the role of the misregulation of miRNAs in AD is the identification of the miRNA targets implicated in the accumulation of toxic proteins (Fig. 3a) or proteins that promote or inhibit cell survival (for example anti-apoptotic proteins such as Bcl2) (Fig. 3b). miRNAs can regulate these proteins either directly, by interacting with the 3'UTR region of their mRNA, or indirectly, by regulating the mRNA of proteins that affect their expression, processing, or function. For example, miR-101, which has been reported to be reduced in AD cortex [135, 136], possesses two MRE in the APP 3'UTR. In vitro validation of this direct regulation was performed by luciferase assay and gain of function of miR-101 in primary rat hippocampal neurons and human cell cultures [137], suggesting that the downregulation of miR-101 generates an increase in APP levels and contributes to the development of AD (Fig. 3a). Conversely, several miRNAs have been described to regulate proteins that participate in APP processing, including BACE1 [133, 138, 139] (Fig. 3a). Thus, the loss or gain of function of these miRNA controls the generation of A β independently of Fig. 3 miRNA overview in MetS and AD. Research into the specific activity of miRNA in several tissues suggests an important role in the generation of metabolic disorders associated with MetS and the histopathological hallmarks of AD. a Toxic protein accumulation. miRNAs can regulate the generation of toxic protein directly by repressing APP and tau or indirectly by repressing the proteins associated with the pathogenic process, such as BACE1 or Cdk5. b Cell survival. Several miRNAs regulate the expression of anti-apoptotic proteins such as Bcl2 and influence cell survival processes. In this example, miR-34, a miRNA related to MetS and AD, represses the expression of Bcl2 and induces mitochondrial pathways of apoptosis. c Insulin resistance. miRNAs regulate multiple aspects of insulin signaling such as insulin secretion, insulin sensitivity, and regulation of glucose uptake in target tissues. Here, miR-126 regulates the expression of IRS1, which has a key role in signal transduction between insulin receptors and intracellular pathways. d Abdominal obesity. miR-103 and miR-107 are upregulated in adipogenesis and significantly downregulated in adipocytes in mouse models of diet-induced obesity. These miRNAs regulate insulin sensitivity by repressing caveolin 1 (CAVI), a critical regulator of the insulin receptor. Another important target is Dicer, which can potentially induce the maturation of several additional miRNAs. e Non-alcoholic fatty liver disease (NAFLD). NAFLD is a hepatic manifestation of MetS and is characterized by the excess accumulation of hepatic triglycerides and fatty acids, hepatic steatosis, and fibrosis. MiR-122 participates in the development of NAFLD by repressing Prolyl 4-hydroxylase subunit alpha-1 (P4H41), a key enzyme in collagen synthesis. f Hypertension. The increase in angiotensin II generates a robust change in the expression of several miRNAs including miR-483-3p which in turn is able to regulate the expression of different renin angiotensin system genes including angiotensinogen and angiotensin-converting enzyme 1 (ACE-1) proteins in vascular smooth muscle cells

the changes in APP expression. Similar studies have examined the miRNAs regulating tau expression (miR-34 family) [140] and the proteins involved in the hyperphosphorylation of tau observed in AD (for example, miR-26b indirectly affects cytoplasmic export of cyclin-dependent kinase 5 (Cdk5), a major tau kinase [141]) (Fig. 3a).

The miRNome has been shown to be affected during the progression of AD; further, miRNAs have demonstrated their potential as non-invasive biomarkers from blood and serum for a wide variety of human pathologies [142]. Recent studies have sought to establish whether the changes in miRNA expression described in neurons of AD models or patients correlate with the expression of peripheral miRNAs that were obtained from peripheral cells such as platelets or mononuclear cells [143-145] or with that of circulating miRNAs [146, 147] and whether they could potentially act as biomarkers for AD. Some miRNAs were consistently identified in AD, including some let-7 family members, miR-9, miR-181, and miR-29 (Fig. 3f). These miRNAs seem to be involved in processes that have previously been associated with AD, such as inflammation and immunological response [148]. Although the concept is very attractive, extensive validation and follow-up studies in larger cohorts of patients are needed before an analysis of miRNAs can be applied as a diagnostic tool in the clinic.



microRNAs as a Molecular Link Between MetS and AD

In a practical exercise, you simple can start to enumerate the coincidental characteristics observed in two pathological conditions to try to establish a relationship. In that sense, we are able to highlight several links between MetS and AD, for example inflammation [149], amyloidosis [150], and Wnt signaling imbalance [29], to name just a few. We can do the same with miRNAs, and we can start to compare tables in miRNAdisease database and try to find some coincidences. The last release of the Human miRNA Disease Database V2.0

(HMDD, June 20, 2013) includes experimental-supported evidence for 572 human miRNA associated to 378 diseases, reported in 3,511 papers [37]. For AD, the database includes 33 miRNAs, for hypertension 24, obesity 9, T2DM 20, and NAFLD 5. In Table 1, a cluster of coincidental miRNAs that are deregulated in AD and one or more component of the MetS is described. The obvious conclusion is that all these miRNAs are ideal examples of how miRNAs are in fact a molecular link between AD and MetS, but a coincidental observation does not always implicate a direct relationship. In fact, miR-21, miR-17, miR-20a, and 146a are between the top ten miRNAs with the biggest disease spectrum width (DSW) [37] which is a parameter that evaluates the effect of a miRNA in human diseases [151]. That means that those miRNAs with high DSW are, in fact, altered in several human diseases. In the case of miR-21, which has the biggest DSW, their deregulation was reported in 123 human diseases of a total of 378, including all types of cancer, several infections like HIV, and of course AD. In conclusion, those miRNAs are important markers of a general state of disease but coincidentally do not give evidence of causality.

The other problem of use coincident in the regulation of miRNA, to establish a relation of MetS and AD, is related to the up- or downregulation observed for a particular miRNA in a given condition. For example, if we use the data in the literature, we can assume that a miRNA that is upregulated in AD and MetS is a better candidate than a miRNA that shows opposite regulation (up and down). This is not necessarily true if we consider the phenomenon of resistance observed in AD, in which the levels of metabolic signals like insulin and leptin are different in plasma in relation to cerebrospinal fluid (CSF) or brain, and therefore, the effect on the expression of miRNAs could be different to that of peripheral tissues.

To understand whether miRNAs are part of the epidemiologic associations between MetS and AD, we discuss a series of preclinical studies, showing that metabolic manipulations induce changes in the expression of miRNAs related to AD in different brain areas. The deregulation of metabolic signals observed in MetS as well as its ability to cross the blood–brain barrier (BBB) are fundamental to understand the impact in the brain function, miRNA regulation, and the possible link with AD.

Metabolic Manipulations and miRNA Deregulations in the Brain

Leptin is an adipocyte-derived cytokine involved in long-term regulation of energy intake and expenditure, body weight, and neuroendocrine functions in mammals [152]. In the nonobese condition, energy intake increases leptin secretion and, in the brain, leptin induces a negative feedback on energy intake via stimulating the expression of anorexigenic neuropeptides.

However, leptin has different effects depending upon the developmental stage. Peripheral leptin is able to enter CSF and the CNS (crossing the BBB and choroid plexus), and subsequently, in the CNS, leptin interacts with specific areas of the brain such as the hypothalamus and hippocampus [153, 154]. Besides leptin transport into the CNS and CSF, several studies indicated that leptin can also be produced in human and rodent brains, for example in the hypothalamus, cortex, and cerebellum [155-157). Besides a role in energy intake, the presence of leptin receptors in specific regions of the brain illustrates its potential for being involved in multiple mechanisms related to brain function and structure in many rodent models [158-161]. The effects of leptin on brain structure are determined by its influence on neurogenesis, axon growth, synaptogenesis, and dendritic morphology, which occur during both pre-and postnatal life and are important for the establishment of hypothalamic, hippocampal, and cortical pathways [162, 163].

The role of leptin in the regulation of miRNA expression has been described previously in peripheral tissues. One of the most commonly studied miRNAs is miR-26b, which is downregulated in the adipose tissue of obese models (high leptin peripheral levels) and adipocytes treated with leptin [164]. Obesity induces lower brain leptin levels or an attenuated leptin response (leptin resistance), due to impaired transport of peripheral leptin into the brain [165]. Similarly, in the brain of AD patients, leptin levels are reduced which is associated to weight loss and cognitive decline observed in the intermediated and advanced stages of AD [166]. In accordance with this observation, the levels of miR-26b were reported to be significantly elevated in human postmortem brains, starting from early stages of AD (Braak III), and it has been suggested that contribute to the progression of the disease by regulating the hyperphosphorylation of tau through the regulation of this target retinoblastoma protein (Rb1) [141]. Recently, the group of Ghribi describes the upregulation of miR-26b [167] and the concomitant reduction of leptin levels [168] in the brain cortex of rabbits fed with a diet supplemented with 2 % cholesterol, which induce the development of full-blown AD pathology, including cortical AB deposits and tangles, and other pathological markers also seen in human AD brains [169-174]. Additionally, they describe the deregulation of other miRNAs previously reported to be altered in human AD samples, including miR-125b, miR-98, miR-107, and miR-30, along with three members of the let-7 family [167]. This suggests that the reduction of central levels of leptin by a metabolic distress, like high cholesterol intake, modulates the expression of miRNAs related to AD in brain and could contribute to the progression of the disease.

Another interesting miRNA regulated by leptin is the brain-specific miR-132, which has been related to synaptic plasticity, neuronal outgrowth, integration of newborn neurons in the dentate gyrus, and it shapes synaptic structure. In the hippocampus of the obesity model db/db mice, which is deficient in leptin receptor and the leptin signaling is impaired, the levels of miR-132 are 60 % lower than the wild-type (wt) mice, and treatment of dissociated hippocampal cultures with leptin increased their expression [175]. These animals display cognitive impairments as well as abnormal synaptic structures, which is similar to the phenotype observed in several neurodegenerative diseases. In AD, consistent with the decrease in leptin levels, miR-132 is described to be downregulated in several brain areas of LOAD patients [90, 176-178]. The role of miR-132 in AD could be attributable to their wellknown targets related to synaptic plasticity like p250GAP [179], but other relevant targets include tau, EP300, SIRT1, and members of the FOX family of transcription factors [90].

These findings show that leptin not only regulates the expression of miRNAs with a well-known role in metabolic homeostasis in peripheral tissues and brain (miR-26b) but also shows that leptin regulates the expression of brain-specific miRNA whose role is more associated with synaptic regulation as well as pathological processes associated to AD (miR-132).

Leptin regulates the expression of miRNAs in other brain areas like hypothalamus. In the obese ob/ob mice model, lacking the leptin gene expression, the hypothalamic neural organization and circuitry are impaired, and interestingly, the neuronal development and metabolic phenotype can be rescue by supplementation with leptin. In male adult obese ob/ob mice, the expression of several miRNA was affected compare to the wt mice [180]. The more affected miRNA correspond to miR-200a, which is also upregulated in the hypothalamus of the db/db mice. The intraperitoneal administration by 11 days of leptin reduces miR-200a to levels observed in the wt mice [180]. miR-200a has been observed to increase in the hippocampus and prefrontal cortex of LOAD patients [90] and also in the brain of 6-month-old APP swe/PS Δ E9 transgenic mice [181]. Considering the leptin resistance observed in LOAD patients is possible to speculate that this pathological process can influence the expression of miR-200a and other miRNAs in the hippocampus of LOAD patients.

Other dietary manipulations, known for generate metabolic alterations and cognitive deficit, have been studied for their role in miRNA regulation in the rat hypothalamus. These studies show that a HFD after weaning increases the expression of several miRNAs, compared to rat reared with a normal diet [182, 183]. One of the interesting findings of this work is that some miRNAs, like miR-200a, show a decrease in their levels during the first week of the HFD but displayed an increase in the chronic period, when obesity is clearly established and hypothalamic insulin, leptin, and inflammation signaling are primarily disturbed. Although the emphasis in AD is often placed on the hippocampus and other brain structures directly involved in cognition and memory, the hypothalamus is clearly involved in AD. Significant atrophy has been identified in the hypothalamus of AD subjects by magnetic resonance volumetric analysis [184]. Also, pathological alterations consistent with AD, including neurofibrillary tangles and amyloid plaques, have been found in the hypothalamus of postmortem AD brains [185, 186]. Therefore, understanding how the hypothalamic miRNAs are affected in response to metabolic distress, like leptin resistance, may offer new insights into the metabolic and cognitive dysfunction associated with AD.

Geekiyanage and Chan showed that a decrease in the levels of miR-137, miR-181c, and miR-9 occurred in brain cortices of wt male mice fed with a HFD for a period of 5 months [187]. The levels of miR-137, miR-181c, and miR-9 are also decreased in AD autopsy brain samples. Interestingly, these miRNAs target a serine palmitoyltransferase protein (SPT) that has been shown to be upregulated in patients with sporadic AD; however, the mRNA level of SPT has not been shown to be altered, suggesting that it is subject to a mechanism of post-transcriptional regulation. SPT is a critical enzyme in the de novo synthesis of ceramides, which promotes BACE1 and γ -secretase mislocation to lipid rafts and therefore promotes the formation of A β [188, 189]. The connection of this misregulated miRNA to AD and MetS is strengthened by a posterior study in which the expression levels of miR-137, miR-181c, and miR-9 were also downregulated in the blood serum of probable AD patients and in male wt mice fed a HFD [146].

In contrast to a HFD, the consumption of a low-fat diet, calorie restriction (CR), and maintenance of metabolic function result in preserved cognition, decreased oxidative stress, preserved brain structure, and reduced inflammatory signaling [190], which are essential for reducing the risk of AD [191]. The regulation of miRNAs may be a potential mechanism underlying these beneficial effects of a low-fat diet. Khanna et al. found that the levels of miR-34a, miR-30e, and miR-181-a-1* are significantly lower in brain tissue samples from old mice fed a calorie-restricted diet, i.e., 60 % of their normal ad libitum caloric intake, compared with mice on the usual ad libitum diet [192]. Interestingly, miR-34a are increased in the brain of AD mice models [193] and also in the liver of dietary and genetic obese mice [85, 106].

Physical activity is a necessary complement to diet control, losing weight, and treating MetS. Exercise benefits not only the musculoskeletal and cardiovascular systems but also the brain. It has been clearly established in controlled studies that regular vigorous exercise improves mood and cognition [194]. In fact, voluntary exercise ameliorates HFD-induced memory deficits and A β deposition in APP transgenic mice [195]. It has been established that part of the beneficial effects of exercise in skeletal muscle is attributed to miRNA regulation [196]; however, the evidence of miRNA responsive to exercise in brain is scarce. In the work of Mojtahedi, the brain levels of miR-124 were shown to increase in an exercise intensity-dependent manner [197]. This is interesting because the expression of miR-124 is downregulated in the brains of patients with AD [130, 198]. Furthermore, it has been suggested that miR-124 may target BACE1 and thus reduce ADrelated cell death [138] (Fig. 2a). In a more recent study, the effect of physical activity on the miRNA expression was evaluated in the hippocampus of the spontaneous senescenceaccelerated P8 mouse model (SAMP8) which is currently considered a model of AD [199]. In 8-month-old SAMP8 sedentary mice, the authors observe 18 altered miRNA (of the 84 miRNA tested) compared to the control strain. Among them, miR-30e-5p, miR-125b-5p, and miR-128-3p have also been reported to be altered in post-mortem human AD hippocampus [130, 131]. Interestingly, these miRNA are responsive to physical activity and change their expression in the AD mice model after voluntary exercise [199]. Bioinformatic analysis indicate that these exerciseresponsive miRNAs are involved in the regulation of several biological process including PI-3-kinase, Akt, insulin, and mTOR signaling pathways; all of which are modulated in the brain by exercise [200-202].

Additional studies in animal models of AD are needed to explore the effect of physical activity on miRNA regulation in the brain and further determine the molecular mechanisms behind the beneficial effects of exercise on cognitive processes.

Drug Therapy of MetS and Their Potential Role in AD

The first-line treatment for MetS is change of lifestyle. However, physicians frequently begin a treatment with medication if the MetS biomarkers do not improve within in 3 to 6 months after initiating lifestyle changes. Generally, the individual disorders that comprise the MetS are treated separately. Considering the evidence that demonstrates that improving the metabolic conditions may prevent AD, it was suggested that the use of these drugs medications could also be useful in preventing neurodegenerative conditions such as AD.

For example, statins, a class of hypolipidemic drugs, have been proposed as potential agents for the treatment or prevention of AD [203, 204]. Data from animal models studies suggest that atorvastatin may reduce the A β [205] and BACE1 protein levels [206] and would be of possible benefit in preventing AD. Several studies have attempted to determine the mechanism of action through which statins appear to regulate the development of AD. One avenue explored was the ability of statins to lower cholesterol levels; however, statins possess a number of pleiotropic effects (cholesterolindependent) that have been extensively discussed [207]. Recently, a new mechanism of action of statins involving the regulation of miRNAs has been identified. Simvastatin and atorvastatin induce miR-33 expression, resulting in the reduced expression of sterol transporters and decreased cholesterol efflux [208]. Interestingly, lovastatin is also able to induce the expression of miR-33 and reduces the miR-9 levels in xenograft models of medulloblastoma, a highly malignant primary brain tumor that originates in the cerebellum [209]. Although miR-33 has not been associated with AD, the fact that a drug such as lovastatin, which is capable of crossing the BBB [210], can generate such changes in brain miRNome could explain why the drug treatment of MetS has been suggested as a tool for preventing AD (Fig. 4).

Along the same lines, the oral anti-diabetic drug metformin alters the miRNA levels in several tissues [211–213]. Because metformin can increase the levels of Dicer [214], the enzyme

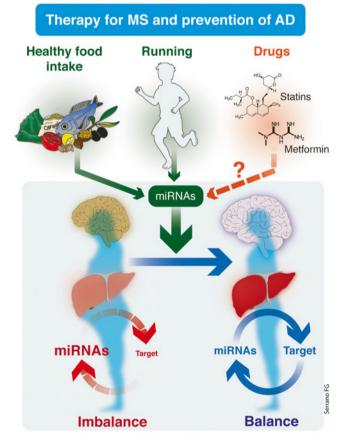


Fig. 4 MetS therapies and their possible impact on AD prevention, through miRNA regulation. Physical activity like running and diet control (e.g., calorie restriction, Mediterranean diet) have been shown to be fundamental in the treatment of MetS through the regulation of several miRNAs in peripheral tissues (liver, adipocytes, vasculature, etc.) and may be important in the prevention of AD by their effects in brain miRNAs that targets protein related to APP processing and survival proteins. Medications for the treatment of individual disorders that compose MetS (e.g., metformin, statins, ARBs) modify miRNAs in peripheral tissues and have the ability to cross the BBB, which suggest a possible role in the regulation of brain miRNAs related to AD progression

responsible for the generation of most of the mature miRNAs (Fig. 1), and is capable of crossing the BBB [215], it is tempting to speculate that some of the controversial roles attributed to metformin in AD [216–218] may be partly dependent on the modulation of miRNAs in the brain (Fig. 4).

Other drugs, such as angiotensin II type 1 receptor antagonists (ARBs), that are used to treat hypertension have been linked to AD [219], but their role in the regulation of miRNAs has not been explored. These drugs can also cross the BBB [220], so more research is necessary to explore the possible regulation of miRNAs that are related to AD.

Concluding Remarks

The concept of "metabolic-cognitive syndrome" emerges from epidemiological and basic research in patients with MetS and cognitive impairment of degenerative or vascular origin [23]. We propose that the miRNAs may act as a molecular link between MetS and AD, may play a role in the complex relationship between metabolic disorders and cognitive disturbances, and may be useful in the generation of therapeutic strategies against MetS that could help prevent or ameliorate the cognitive decline observed in AD and other pathologies. In both animal models and patients with MetS or AD, altered expression levels of different miRNAs have been observed as a consequence of changes at different stages of biogenetic processes, including transcription, processing, and miRISC function (Fig. 1). Importantly, metabolic manipulations generate signals that alter the expression of miRNAs in both peripheral tissues and the brain, inducing changes conducting to MetS and AD through the regulation of ubiquitously expressed target reflecting a common mechanism (ABC1 transporter related to impaired cholesterol homeostasis observed in MetS and AD) or tissue-specific targets that control particular characteristic of each disease (for example, p250GAP related to synaptic loss observed in AD or β-Klotho related to impaired postprandial responses in patients with steatosis) (Fig. 2).

More directed experiments that confirm our hypothesis are required, for example, the use of the obese Zucker rats (ZDF) which possess a (*fa/fa*) mutation and leptin receptor deficiency and develop all the hallmarks of MetS [221] is ideally suited for this purpose. These animals show impairment of synaptic plasticity [222, 223], spatial memory, and learning [224] and even show an increase in brain mRNA levels of APOE [225], protein aggregation, and p-tau levels [226, 227] which correspond to classic markers of AD. The study of miRNAs that change their expression in different brain areas in response to metabolic deterioration could identify key biomarkers that helps to develop diagnostic tools and to prevent the cognitive decline in early stages of the metabolic-cognitive syndrome. Additionally, the use of specific viral delivered miRNA oligomers in the brain that prevent the metabolic-induced change of their levels and restore the cognitive impairment without changes in the metabolic status of the animal might confirm the critical role of miRNAs as a molecular link between MetS and AD.

Lifestyle changes, such as a calorie-restricted diet or increased physical activity, have a positive impact on the expression of the miRNAs that control the proteins involved in APP processing and plaque formation, such as BACE1, suggesting that this is a potential therapeutic approach to prevent deleterious processes that lead towards the development of MetS and AD (Fig. 4). Conversely, the use of medications or drugs to combat the various components of metabolic syndrome may potentially affect the expression of miRNAs in the brain because they can cross the BBB. These unexpected side effects have not been explored in depth, and further research is needed to establish whether this modulation of brain miRNAs can be beneficial or detrimental to the development of AD and other pathologies such as brain cancer. Finally, altered circulating miRNA profiles have already been linked to disease states, including metabolic disorders associated with MetS and AD, and may provide important biomarkers for disease. The use of miRNAs as a molecular link between MetS and AD may provide insight into the initiation and development of these diseases and may offer new therapeutic avenues for diagnostic tests and prevention.

Acknowledgments This work was supported by grants PFB 12/2007 from the Basal Centre for Excellence in Science and Technology, FONDECYT 1120156, MIFAB Foundation, and Fundación Ciencia y Vida to NCI and pre-doctoral fellowships from CONICYT to JAR and JFC. Graphic work was carried out by illustrative science (www. illustrative-science.com).

Statement of Author Contributions NCI and JFC conceived the review concept. JFC and JAR carried out the literature search. JFC designed the figures and table. All authors were involved in writing the paper and had final approval of the submitted and published versions.

References

- Thies W, Bleiler L (2013) 2013 Alzheimer's disease facts and figures. Alzheimers Dement 9:208–245. doi:10.1016/j.jalz.2013. 02.003
- 2. Ballard C, Gauthier S, Corbett A et al (2011) Alzheimer's disease. Lancet 377:1019–1031. doi:10.1016/S0140-6736(10)61349-9
- Inestrosa NC, Varela-Nallar L (2014) Wnt signaling in the nervous system and in Alzheimer's disease. J Mol Cell Biol 6:64–74. doi: 10.1093/jmcb/mjt051
- Hardy J, Selkoe DJ (2002) The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics. Science 297:353–356. doi:10.1126/science.1072994
- Kanekiyo T, Xu H, Bu G (2014) ApoE and Aβ in Alzheimer's disease: accidental encounters or partners? Neuron 81:740–754. doi:10.1016/j.neuron.2014.01.045

- De la Monte SM (2014) Type 3 diabetes is sporadic Alzheimer's disease: mini-review. Eur Neuropsychopharmacol. doi:10.1016/j. euroneuro.2014.06.008
- Li Z, Zhang W, Sima AAF (2007) Alzheimer-like changes in rat models of spontaneous diabetes. Diabetes 56:1817–1824. doi:10. 2337/db07-0171
- Mehla J, Chauhan BC, Chauhan NB (2014) Experimental induction of type 2 diabetes in aging-accelerated mice triggered Alzheimer-like pathology and memory deficits. J Alzheimers Dis 39:145–162. doi:10.3233/JAD-131238
- Csiszar A, Tucsek Z, Toth P et al (2013) Synergistic effects of hypertension and aging on cognitive function and hippocampal expression of genes involved in β-amyloid generation and Alzheimer's disease. Am J Physiol Heart Circ Physiol 305: H1120–H1130. doi:10.1152/ajpheart.00288.2013
- Thirumangalakudi L, Prakasam A, Zhang R et al (2008) High cholesterol-induced neuroinflammation and amyloid precursor protein processing correlate with loss of working memory in mice. J Neurochem 106:475–485. doi:10.1111/j.1471-4159.2008. 05415.x
- Zhang L, Dasuri K, Fernandez-Kim S-O et al (2013) Prolonged diet induced obesity has minimal effects towards brain pathology in mouse model of cerebral amyloid angiopathy: implications for studying obesity-brain interactions in mice. Biochim Biophys Acta 1832:1456–1462. doi:10.1016/j.bbadis.2013.01.002
- Nelson L, Gard P, Tabet N (2014) Hypertension and inflammation in Alzheimer's disease: close partners in disease development and progression. J Alzheimers Dis. doi:10.3233/JAD-140024
- Solomon A, Kivipelto M, Wolozin B et al (2009) Midlife serum cholesterol and increased risk of Alzheimer's and vascular dementia three decades later. Dement Geriatr Cogn Disord 28:75–80. doi:10.1159/000231980
- Whitmer RA, Gustafson DR, Barrett-Connor E et al (2008) Central obesity and increased risk of dementia more than three decades later. Neurology 71:1057–1064. doi:10.1212/01.wnl. 0000306313.89165.ef
- Ahtiluoto S, Polvikoski T, Peltonen M et al (2010) Diabetes, Alzheimer disease, and vascular dementia: a population-based neuropathologic study. Neurology 75:1195–1202. doi:10.1212/ WNL.0b013e3181f4d7f8
- Thambisetty M, Jeffrey Metter E, Yang A et al (2013) Glucose intolerance, insulin resistance, and pathological features of Alzheimer disease in the Baltimore Longitudinal Study of Aging. JAMA Neurol 70:1167–1172. doi:10.1001/jamaneurol. 2013.284
- Ott A, Stolk RP, van Harskamp F et al (1999) Diabetes mellitus and the risk of dementia: The Rotterdam Study. Neurology 53: 1937–1942
- Frisardi V, Solfrizzi V, Seripa D et al (2010) Metabolic-cognitive syndrome: a cross-talk between metabolic syndrome and Alzheimer's disease. Ageing Res Rev 9:399–417. doi:10.1016/j. arr.2010.04.007
- Solfrizzi V, Panza F, Colacicco AM et al (2004) Vascular risk factors, incidence of MCI, and rates of progression to dementia. Neurology 63:1882–1891. doi:10.1212/01.WNL.0000144281. 38555.E3
- Profenno LA, Porsteinsson AP, Faraone SV (2010) Meta-analysis of Alzheimer's disease risk with obesity, diabetes, and related disorders. Biol Psychiatry 67:505–512. doi:10.1016/j.biopsych. 2009.02.013
- De la Monte SM, Tong M (2013) Brain metabolic dysfunction at the core of Alzheimer's disease. Biochem Pharmacol. doi:10. 1016/j.bcp.2013.12.012
- Frisardi V, Solfrizzi V, Capurso C et al (2010) Is insulin resistant brain state a central feature of the metabolic-cognitive syndrome? J Alzheimers Dis 21:57–63. doi:10.3233/JAD-2010-100015

- Panza F, Solfrizzi V, Logroscino G et al (2012) Current epidemiological approaches to the metabolic-cognitive syndrome. J Alzheimers Dis 30(Suppl 2):S31–S75. doi:10.3233/JAD-2012-111496
- Roberts RO, Geda YE, Knopman DS, et al (2010) Metabolic syndrome, inflammation, and nonamnestic mild cognitive impairment in older persons: a population-based study. Alzheimer Dis Assoc Disord 24:11–18. doi: 10.1097/WAD.0b013e3181a4485c
- Luque-Contreras D, Carvajal K, Toral-Rios D et al (2014) Oxidative stress and metabolic syndrome: cause or consequence of Alzheimer's disease? Oxidative Med Cell Longev. doi:10. 1155/2014/497802
- Oskarsson ME, Paulsson JF, Schultz SW et al (2015) In vivo seeding and cross-seeding of localized amyloidosis. Am J Pathol 185:834–846. doi:10.1016/j.ajpath.2014.11.016
- Knowles TPJ, Vendruscolo M, Dobson CM (2014) The amyloid state and its association with protein misfolding diseases. Nat Rev Mol Cell Biol 15:384–396. doi:10.1038/nrm3810
- Guan J, Zhao HL, Sui Y et al (2009) Histopathological correlations of islet amyloidosis and hyaline arteriosclerosis with amylin gene mutations and apolipoprotein E polymorphisms in Chinese patients with type 2 diabetes. Diabetes 58:A368–A368. doi:10. 1097/MPA.0b013e3182965e6e
- Ríos JA, Cisternas P, Arrese M et al (2014) Is Alzheimer's disease related to metabolic syndrome? A Wnt signaling conundrum. Prog Neurobiol. doi:10.1016/j.pneurobio.2014.07.004
- Bartel DP (2004) MicroRNAs: genomics, biogenesis, mechanism, and function. Cell 116:281–297. doi:10.1016/S0092-8674(04) 00045-5
- Bartel DP (2009) MicroRNAs: target recognition and regulatory functions. Cell 136:215–233. doi:10.1016/j.cell.2009.01.002
- Rottiers V, Näär AM (2012) MicroRNAs in metabolism and metabolic disorders. Nat Rev Mol Cell Biol 13:239–250. doi:10.1038/ nrm3313
- McGregor RA, Choi MS (2011) microRNAs in the regulation of adipogenesis and obesity. Curr Mol Med 11:304–316. doi:10. 2174/156652411795677990
- Mao Y, Mohan R, Zhang S, Tang X (2013) MicroRNAs as pharmacological targets in diabetes. Pharmacol Res 75:37–47. doi:10. 1016/j.phrs.2013.06.005
- Ling S, Nanhwan M, Qian J et al (2013) Modulation of microRNAs in hypertension-induced arterial remodeling through the β1 and β3-adrenoreceptor pathways. J Mol Cell Cardiol 65: 127–136. doi:10.1016/j.yjmcc.2013.10.003
- Dimmeler S, Nicotera P (2013) MicroRNAs in age-related diseases. EMBO Mol Med 5:180–190. doi:10.1002/emmm. 201201986
- Li Y, Qiu C, Tu J et al (2014) HMDD v2.0: a database for experimentally supported human microRNA and disease associations. Nucleic Acids Res 42:D1070–D1074. doi:10.1093/nar/gkt1023
- Kawamata T, Tomari Y (2010) Making RISC. Trends Biochem Sci 35:368–376. doi:10.1016/j.tibs.2010.03.009
- Kawamata T, Yoda M, Tomari Y (2011) Multilayer checkpoints for microRNA authenticity during RISC assembly. EMBO Rep 12:944–949. doi:10.1038/embor.2011.128
- Kwak PB, Tomari Y (2012) The N domain of Argonaute drives duplex unwinding during RISC assembly. Nat Struct Mol Biol 19: 145–151. doi:10.1038/nsmb.2232
- Dávalos A, Goedeke L, Smibert P et al (2011) miR-33a/b contribute to the regulation of fatty acid metabolism and insulin signaling. Proc Natl Acad Sci U S A 108:9232–9237. doi:10.1073/pnas. 1102281108
- 42. Najafi-Shoushtari SH, Kristo F, Li Y et al (2010) MicroRNA-33 and the SREBP host genes cooperate to control cholesterol homeostasis. Science 328:1566–1569. doi:10.1126/science.1189123

- 43. Dill H, Linder B, Fehr A, Fischer U (2012) Intronic miR-26b controls neuronal differentiation by repressing its host transcript, ctdsp2. Genes Dev 26:25–30. doi:10.1101/gad.177774.111
- Rokavec M, Li H, Jiang L, Hermeking H (2014) The p53/miR-34 axis in development and disease. J Mol Cell Biol 6:214–230. doi: 10.1093/jmcb/mju003
- Boominathan L (2010) The guardians of the genome (p53, TAp73, and TA-p63) are regulators of tumor suppressor miRNAs network. Cancer Metastasis Rev 29:613–639. doi:10.1007/ s10555-010-9257-9
- Zovoilis A, Agbemenyah HY, Agis-Balboa RC et al (2011) microRNA-34c is a novel target to treat dementias. EMBO J 30: 4299–4308. doi:10.1038/emboj.2011.327
- Hooper C, Meimaridou E, Tavassoli M et al (2007) p53 is upregulated in Alzheimer's disease and induces tau phosphorylation in HEK293a cells. Neurosci Lett 418:34–37. doi:10.1016/j.neulet. 2007.03.026
- 48. Bialopiotrowicz E, Szybinska A, Kuzniewska B et al (2012) Highly pathogenic Alzheimer's disease presenilin 1 P117R mutation causes a specific increase in p53 and p21 protein levels and cell cycle dysregulation in human lymphocytes. J Alzheimers Dis 32:397–415. doi:10.3233/JAD-2012-121129
- Roe CM, Behrens MI (2013) AD and cancer: epidemiology makes for strange bedfellows. Neurology 81:310–311. doi:10.1212/ WNL.0b013e31829c5f16
- 50. Behrens MI, Silva M, Salech F et al (2012) Inverse susceptibility to oxidative death of lymphocytes obtained from Alzheimer's patients and skin cancer survivors: increased apoptosis in Alzheimer's and reduced necrosis in cancer. J Gerontol A Biol Sci Med Sci 67:1036–1040. doi:10.1093/gerona/glr258
- Driver JA, Beiser A, Au R et al (2012) Inverse association between cancer and Alzheimer's disease: results from the Framingham Heart Study. BMJ 344, e1442
- Demetrius LA, Simon DK (2013) The inverse association of cancer and Alzheimer's: a bioenergetic mechanism. J R Soc Interface 10:20130006. doi:10.1098/rsif.2013.0006
- Musicco M, Adorni F, Di Santo S et al (2013) Inverse occurrence of cancer and Alzheimer disease: a population-based incidence study. Neurology 81:322–328. doi:10.1212/WNL. 0b013e31829c5ec1
- Castro RE, Ferreira DMS, Afonso MB et al (2013) miR-34a/ SIRT1/p53 is suppressed by ursodeoxycholic acid in the rat liver and activated by disease severity in human non-alcoholic fatty liver disease. J Hepatol 58:119–125. doi:10.1016/j.jhep.2012.08. 008
- Gregory RI, Yan K-P, Amuthan G et al (2004) The microprocessor complex mediates the genesis of microRNAs. Nature 432:235– 240. doi:10.1038/nature03120
- Denli AM, Tops BBJ, Plasterk RH et al (2004) Processing of primary microRNAs by the microprocessor complex. Nature 432:231–235. doi:10.1038/nature03049
- Han J, Lee Y, Yeom K-H et al (2006) Molecular basis for the recognition of primary microRNAs by the Drosha-DGCR8 complex. Cell 125:887–901. doi:10.1016/j.cell.2006.03.043
- Lund E, Güttinger S, Calado A et al (2004) Nuclear export of microRNA precursors. Science 303:95–98. doi:10.1126/science. 1090599
- Yi R, Qin Y, Macara IG, Cullen BR (2003) Exportin-5 mediates the nuclear export of pre-microRNAs and short hairpin RNAs. Genes Dev 17:3011–3016. doi:10.1101/gad.1158803
- Okada C, Yamashita E, Lee SJ et al (2009) A high-resolution structure of the pre-microRNA nuclear export machinery. Science 326:1275–1279. doi:10.1126/science.1178705
- Macrae IJ, Zhou K, Li F et al (2006) Structural basis for doublestranded RNA processing by Dicer. Science 311:195–198. doi:10. 1126/science.1121638

- Bernstein E, Kim SY, Carmell MA et al (2003) Dicer is essential for mouse development. Nat Genet 35:215–217. doi:10.1038/ ng1253
- Davis TH, Cuellar TL, Koch SM et al (2008) Conditional loss of Dicer disrupts cellular and tissue morphogenesis in the cortex and hippocampus. J Neurosci 28:4322–4330. doi:10.1523/ JNEUROSCI.4815-07.2008
- Cuellar TL, Davis TH, Nelson PT et al (2008) Dicer loss in striatal neurons produces behavioral and neuroanatomical phenotypes in the absence of neurodegeneration. Proc Natl Acad Sci U S A 105: 5614–5619. doi:10.1073/pnas.0801689105
- Kim J, Inoue K, Ishii J et al (2007) A microRNA feedback circuit in midbrain dopamine neurons. Science 317:1220–1224. doi:10. 1126/science.1140481
- Schaefer A, O'Carroll D, Tan CL et al (2007) Cerebellar neurodegeneration in the absence of microRNAs. J Exp Med 204:1553– 1558. doi:10.1084/jem.20070823
- Kawase-Koga Y, Low R, Otaegi G et al (2010) RNAase-III enzyme Dicer maintains signaling pathways for differentiation and survival in mouse cortical neural stem cells. J Cell Sci 123:586– 594. doi:10.1242/jcs.059659
- Hébert SS, Papadopoulou AS, Smith P et al (2010) Genetic ablation of Dicer in adult forebrain neurons results in abnormal tau hyperphosphorylation and neurodegeneration. Hum Mol Genet 19:3959–3969. doi:10.1093/hmg/ddq311
- Schneeberger M, Altirriba J, García A et al (2012) Deletion of miRNA processing enzyme Dicer in POMC-expressing cells leads to pituitary dysfunction, neurodegeneration and development of obesity. Mol Metab 2:74–85. doi:10.1016/j.molmet. 2012.10.001
- Khvorova A, Reynolds A, Jayasena SD (2003) Functional siRNAs and miRNAs exhibit strand bias. Cell 115:209–216. doi:10.1016/S0092-8674(03)00801-8
- Chendrimada TP, Gregory RI, Kumaraswamy E et al (2005) TRBP recruits the Dicer complex to Ago2 for microRNA processing and gene silencing. Nature 436:740–744. doi:10.1038/ nature03868
- Huntzinger E, Izaurralde E (2011) Gene silencing by microRNAs: contributions of translational repression and mRNA decay. Nat Rev Genet 12:99–110. doi:10.1038/nrg2936
- Vilardo E, Barbato C, Ciotti M et al (2010) MicroRNA-101 regulates amyloid precursor protein expression in hippocampal neurons. J Biol Chem 285:18344–18351. doi:10.1074/jbc.M110. 112664
- Ha M, Kim VN (2014) Regulation of microRNA biogenesis. Nat Rev Mol Cell Biol 15:509–524. doi:10.1038/nrm3838
- Martinez NJ, Gregory RI (2013) Argonaute2 expression is posttranscriptionally coupled to microRNA abundance. RNA 19:605– 612. doi:10.1261/ma.036434.112
- Gibbings D, Mostowy S, Jay F et al (2012) Selective autophagy degrades DICER and AGO2 and regulates miRNA activity. Nat Cell Biol 14:1314–1321. doi:10.1038/ncb2611
- Ryter SW, Cloonan SM, Choi AMK (2013) Autophagy: a critical regulator of cellular metabolism and homeostasis. Mol Cell 36:7– 16. doi:10.1007/s10059-013-0140-8
- Ren SY, Xu X (2014) Role of autophagy in metabolic syndromeassociated heart disease. Biochim Biophys Acta. doi:10.1016/j. bbadis.2014.04.029
- Ryter SW, Koo JK, Choi AMK (2014) Molecular regulation of autophagy and its implications for metabolic diseases. Curr Opin Clin Nutr Metab Care 17:329–337. doi:10.1097/MCO. 000000000000068
- Wolfe DM, Lee J-H, Kumar A et al (2013) Autophagy failure in Alzheimer's disease and the role of defective lysosomal acidification. Eur J Neurosci 37:1949–1961. doi:10.1111/ejn.12169

- Salminen A, Kaarniranta K, Kauppinen A et al (2013) Impaired autophagy and APP processing in Alzheimer's disease: the potential role of Beclin 1 interactome. Prog Neurobiol 106–107:33–54. doi:10.1016/j.pneurobio.2013.06.002
- Krol J, Loedige I, Filipowicz W (2010) The widespread regulation of microRNA biogenesis, function and decay. Nat Rev Genet 11: 597–610. doi:10.1038/nrg2843
- Fu T, Choi S-E, Kim D-H et al (2012) Aberrantly elevated microRNA-34a in obesity attenuates hepatic responses to FGF19 by targeting a membrane coreceptor β-Klotho. Proc Natl Acad Sci U S A 109:16137–16142. doi:10.1073/pnas. 1205951109
- Zhou R, Yuan P, Wang Y et al (2009) Evidence for selective microRNAs and their effectors as common long-term targets for the actions of mood stabilizers. Neuropsychopharmacology 34: 1395–1405. doi:10.1038/npp.2008.131
- Lee J, Padhye A, Sharma A et al (2010) A pathway involving famesoid X receptor and small heterodimer partner positively regulates hepatic sirtuin 1 levels via microRNA-34a inhibition. J Biol Chem 285:12604–12611. doi:10.1074/jbc.M109.094524
- Li X, Khanna A, Li N, Wang E (2011) Circulatory miR34a as an RNA-based, noninvasive biomarker for brain aging. Aging (Albany NY) 3:985–1002
- Godoy JA, Zolezzi JM, Braidy N, Inestrosa NC (2014) Role of Sirt1 during the ageing process: relevance to protection of synapses in the brain. Mol Neurobiol. doi:10.1007/s12035-014-8645-5
- Codocedo JF, Allard C, Godoy JA et al (2012) SIRT1 regulates dendritic development in hippocampal neurons. PLoS One 7, e47073. doi:10.1371/journal.pone.0047073
- Choi S-E, Kemper JK (2013) Regulation of SIRT1 by microRNAs. Mol Cell 36:385–392. doi:10.1007/s10059-013-0297-1
- Lau P, Bossers K, Janky R et al (2013) Alteration of the microRNA network during the progression of Alzheimer's disease. EMBO Mol Med 5:1613–1634. doi:10.1002/emmm. 201201974
- Schonrock N, Humphreys DT, Preiss T, Götz J (2012) Target gene repression mediated by miRNAs miR-181c and miR-9 both of which are down-regulated by amyloid-β. J Mol Neurosci 46: 324–335. doi:10.1007/s12031-011-9587-2
- 92. Ramachandran D, Roy U, Garg S et al (2011) Sirt1 and mir-9 expression is regulated during glucose-stimulated insulin secretion in pancreatic β-islets. FEBS J 278:1167–1174. doi:10.1111/j. 1742-4658.2011.08042.x
- Barca-Mayo O, De Pietri TD (2014) Convergent microRNA actions coordinate neocortical development. Cell Mol Life Sci 71: 2975–2995. doi:10.1007/s00018-014-1576-5
- Schouten M, Aschrafi A, Bielefeld P et al (2013) microRNAs and the regulation of neuronal plasticity under stress conditions. Neuroscience 241:188–205. doi:10.1016/j.neuroscience.2013.02. 065
- Alberti KGMM, Zimmet P, Shaw J (2005) The metabolic syndrome—a new worldwide definition. Lancet 366:1059–1062. doi:10.1016/S0140-6736(05)67402-8
- Cerezo C, Segura J, Praga M, Ruilope LM (2013) Guidelines updates in the treatment of obesity or metabolic syndrome and hypertension. Curr Hypertens Rep 15:196–203. doi:10.1007/ s11906-013-0337-4
- 97. Alberti KGMM, Eckel RH, Grundy SM et al (2009) Harmonizing the metabolic syndrome: a joint interim statement of the International Diabetes Federation Task Force on Epidemiology and Prevention; National Heart, Lung, and Blood Institute; American Heart Association; World Heart Federation; International. Circulation 120:1640–1645. doi:10.1161/ CIRCULATIONAHA.109.192644

- Goedeke L, Fernández-Hernando C (2014) microRNAs: a connection between cholesterol metabolism and neurodegeneration. Neurobiol Dis 72:3–8. doi: 10.1016/j.nbd.2014.05.034
- Gerin I, Clerbaux L-A, Haumont O et al (2010) Expression of miR-33 from an SREBP2 intron inhibits cholesterol export and fatty acid oxidation. J Biol Chem 285:33652–33661. doi:10.1074/ jbc.M110.152090
- Yvan-Charvet L, Ranalletta M, Wang N et al (2007) Combined deficiency of ABCA1 and ABCG1 promotes foam cell accumulation and accelerates atherosclerosis in mice. J Clin Invest 117: 3900–3908. doi:10.1172/JCI33372
- Tang C, Oram JF (2009) The cell cholesterol exporter ABCA1 as a protector from cardiovascular disease and diabetes. Biochim Biophys Acta 1791:563–572. doi:10.1016/j.bbalip.2009.03.011
- Rayner KJ, Esau CC, Hussain FN et al (2011) Inhibition of miR-33a/b in non-human primates raises plasma HDL and lowers VLDL triglycerides. Nature 478:404–407. doi:10.1038/ nature10486
- Koldamova R, Fitz NF, Lefterov I (2010) The role of ATP-binding cassette transporter A1 in Alzheimer's disease and neurodegeneration. Biochim Biophys Acta 1801:824–830. doi:10.1016/j. bbalip.2010.02.010
- Alexander R, Lodish H, Sun L (2011) MicroRNAs in adipogenesis and as therapeutic targets for obesity. Expert Opin Ther Targets 15:623–636. doi:10.1517/14728222.2011.561317
- Xie H, Lim B, Lodish HF (2009) MicroRNAs induced during adipogenesis that accelerate fat cell development are downregulated in obesity. Diabetes 58:1050–1057. doi:10.2337/db08-1299
- 106. Trajkovski M, Hausser J, Soutschek J et al (2011) MicroRNAs 103 and 107 regulate insulin sensitivity. Nature 474:649–653. doi: 10.1038/nature10112
- Angulo P (2007) Obesity and nonalcoholic fatty liver disease. Nutr Rev 65:S57–S63. doi:10.1111/j.1753-4887.2007.tb00329.x
- Baur JA, Pearson KJ, Price NL et al (2006) Resveratrol improves health and survival of mice on a high-calorie diet. Nature 444: 337–342. doi:10.1038/nature05354
- Szabo G, Bala S (2013) MicroRNAs in liver disease. Nat Rev Gastroenterol Hepatol 10:542–552. doi:10.1038/nrgastro.2013.87
- 110. Miyaaki H, Ichikawa T, Kamo Y et al (2013) Significance of serum and hepatic microRNA-122 levels in patients with nonalcoholic fatty liver disease. Liver Int. doi:10.1111/liv.12429
- Yamada H, Suzuki K, Ichino N et al (2013) Associations between circulating microRNAs (miR-21, miR-34a, miR-122 and miR-451) and non-alcoholic fatty liver. Clin Chim Acta 424:99–103. doi:10.1016/j.cca.2013.05.021
- 112. Pirola CJ, Gianotti TF, Castaño GO, Sookoian S (2013) Circulating microRNA-122 signature in nonalcoholic fatty liver disease and cardiovascular disease: a new endocrine system in metabolic syndrome. Hepatology 57:2545–2547. doi:10.1002/ hep.26116
- Li J, Ghazwani M, Zhang Y et al (2013) miR-122 regulates collagen production via targeting hepatic stellate cells and suppressing P4HA1 expression. J Hepatol 58:522–528. doi:10.1016/j.jhep. 2012.11.011
- Leavens KF, Birnbaum MJ (2011) Insulin signaling to hepatic lipid metabolism in health and disease. Crit Rev Biochem Mol Biol 46:200–215. doi:10.3109/10409238.2011.562481
- Poy MN, Eliasson L, Krutzfeldt J et al (2004) A pancreatic isletspecific microRNA regulates insulin secretion. Nature 432:226– 230. doi:10.1038/nature03076
- Kim S, Pak Y (2005) Caveolin-2 regulation of the cell cycle in response to insulin in Hirc-B fibroblast cells. Biochem Biophys Res Commun 330:88–96. doi:10.1016/j.bbrc.2005.02.130
- Ryu HS, Park S-Y, Ma D et al (2011) The induction of microRNA targeting IRS-1 is involved in the development of insulin

resistance under conditions of mitochondrial dysfunction in hepatocytes. PLoS One 6, e17343. doi:10.1371/journal.pone.0017343

- Pimenta E, Oparil S (2012) Management of hypertension in the elderly. Nat Rev Cardiol 9:286–296. doi:10.1038/nrcardio.2012.
 27
- Coffman TM (2011) Under pressure: the search for the essential mechanisms of hypertension. Nat Med 17:1402–1409. doi:10. 1038/nm.2541
- Friso S, Carvajal CA, Fardella CE, Olivieri O (2014) Epigenetics and arterial hypertension: the challenge of emerging evidence. Transl Res. doi:10.1016/j.trsl.2014.06.007
- Feinberg AP (2008) Epigenetics at the epicenter of modern medicine. JAMA 299:1345–1350. doi:10.1001/jama.299.11.1345
- Maegdefessel L (2014) The emerging role of microRNAs in cardiovascular disease. J Intern Med. doi:10.1111/joim.12298
- 123. Kontaraki JE, Marketou ME, Zacharis EA et al (2014) MicroRNA-9 and microRNA-126 expression levels in patients with essential hypertension: potential markers of target-organ damage. J Am Soc Hypertens 8:368–375. doi:10.1016/j.jash. 2014.03.324
- 124. Fu X, Guo L, Jiang Z-M et al (2014) An miR-143 promoter variant associated with essential hypertension. Int J Clin Exp Med 7: 1813–1817
- 125. Nossent AY, Hansen JL, Doggen C et al (2011) SNPs in microRNA binding sites in 3'-UTRs of RAAS genes influence arterial blood pressure and risk of myocardial infarction. Am J Hypertens 24:999–1006. doi:10.1038/ajh.2011.92
- Kemp JR, Unal H, Desnoyer R et al (2014) Angiotensin IIregulated microRNA 483-3p directly targets multiple components of the renin-angiotensin system. J Mol Cell Cardiol 75:25–39. doi: 10.1016/j.yjmcc.2014.06.008
- Morris R, Mucke L (2006) Alzheimer's disease: a needle from the haystack. Nature 440:284–285. doi:10.1038/440284a
- Hardy J (2006) Has the amyloid cascade hypothesis for Alzheimer's disease been proved? Curr Alzheimer Res 3:71–73. doi:10.2174/156720506775697098
- Binder LI, Guillozet-Bongaarts AL, Garcia-Sierra F, Berry RW (2005) Tau, tangles, and Alzheimer's disease. Biochim Biophys Acta 1739:216–223. doi:10.1016/j.bbadis.2004.08.014
- Lukiw WJ (2007) Micro-RNA speciation in fetal, adult and Alzheimer's disease hippocampus. Neuroreport 18:297–300. doi:10.1097/WNR.0b013e3280148e8b
- 131. Cogswell JP, Ward J, Taylor IA et al (2008) Identification of miRNA changes in Alzheimer's disease brain and CSF yields putative biomarkers and insights into disease pathways. J Alzheimers Dis 14:27–41. doi:10.1016/j.jalz.2008.05.420
- Sethi P, Lukiw WJ (2009) Micro-RNA abundance and stability in human brain: specific alterations in Alzheimer's disease temporal lobe neocortex. Neurosci Lett 459:100–104. doi:10.1016/j.neulet. 2009.04.052
- 133. Hébert SS, Horré K, Nicolaï L et al (2008) Loss of microRNA cluster miR-29a/b-1 in sporadic Alzheimer's disease correlates with increased BACE1/beta-secretase expression. Proc Natl Acad Sci U S A 105:6415–6420. doi:10.1073/pnas.0710263105
- Schonrock N, Ke YD, Humphreys D et al (2010) Neuronal microRNA deregulation in response to Alzheimer's disease amyloid-beta. PLoS One 5, e11070. doi:10.1371/journal.pone. 0011070
- Nunez-Iglesias J, Liu C-C, Morgan TE et al (2010) Joint genomewide profiling of miRNA and mRNA expression in Alzheimer's disease cortex reveals altered miRNA regulation. PLoS One 5, e8898. doi:10.1371/journal.pone.0008898
- 136. Wang W-X, Huang Q, Hu Y et al (2011) Patterns of microRNA expression in normal and early Alzheimer's disease human temporal cortex: white matter versus gray matter. Acta Neuropathol 121:193–205. doi:10.1007/s00401-010-0756-0

- 137. Long JM, Lahiri DK (2011) MicroRNA-101 downregulates Alzheimer's amyloid-β precursor protein levels in human cell cultures and is differentially expressed. Biochem Biophys Res Commun 404:889–895. doi:10.1016/j.bbrc.2010.12.053
- Fang M, Wang J, Zhang X et al (2012) The miR-124 regulates the expression of BACE1/β-secretase correlated with cell death in Alzheimer's disease. Toxicol Lett 209:94–105. doi:10.1016/j. toxlet.2011.11.032
- 139. Wang W-X, Rajeev BW, Stromberg AJ et al (2008) The expression of microRNA miR-107 decreases early in Alzheimer's disease and may accelerate disease progression through regulation of beta-site amyloid precursor protein-cleaving enzyme 1. J Neurosci 28:1213–1223. doi:10.1523/JNEUROSCI.5065-07.2008
- Dickson JR, Kruse C, Montagna DR et al (2013) Alternative polyadenylation and miR-34 family members regulate tau expression. J Neurochem 127:739–749. doi:10.1111/jnc.12437
- 141. Absalon S, Kochanek DM, Raghavan V, Krichevsky AM (2013) MiR-26b, upregulated in Alzheimer's disease, activates cell cycle entry, tau-phosphorylation, and apoptosis in postmitotic neurons. J Neurosci 33:14645–14659. doi:10.1523/JNEUROSCI.1327-13. 2013
- 142. Keller A, Leidinger P, Bauer A et al (2011) Toward the bloodborne miRNome of human diseases. Nat Methods 8:841–843. doi: 10.1038/nmeth.1682
- Schipper HM (2007) Biomarker potential of heme oxygenase-1 in Alzheimer's disease and mild cognitive impairment. Biomark Med 1:375–385. doi:10.2217/17520363.1.3.375
- 144. Bekris LM, Lutz F, Montine TJ et al (2013) MicroRNA in Alzheimer's disease: an exploratory study in brain, cerebrospinal fluid and plasma. Biomarkers 18:455–466. doi:10.3109/ 1354750X.2013.814073
- 145. Villa C, Ridolfi E, Fenoglio C et al (2013) Expression of the transcription factor Sp1 and its regulatory hsa-miR-29b in peripheral blood mononuclear cells from patients with Alzheimer's disease. J Alzheimers Dis 35:487–494. doi:10.3233/JAD-122263
- Geekiyanage H, Jicha GA, Nelson PT, Chan C (2012) Blood serum miRNA: non-invasive biomarkers for Alzheimer's disease. Exp Neurol 235:491–496. doi:10.1016/j.expneurol.2011.11.026
- 147. Leidinger P, Backes C, Deutscher S et al (2013) A blood based 12miRNA signature of Alzheimer disease patients. Genome Biol 14: R78. doi:10.1186/gb-2013-14-7-r78
- Dorval V, Nelson PT, Hébert SS (2013) Circulating microRNAs in Alzheimer's disease: the search for novel biomarkers. Front Mol Neurosci 6:24. doi:10.3389/fnmol.2013.00024
- Craft S (2005) Insulin resistance syndrome and Alzheimer's disease: age- and obesity-related effects on memory, amyloid, and inflammation. Neurobiol Aging 26(Suppl 1):65–69. doi:10.1016/j.neurobiolaging.2005.08.021
- Westermark GT, Westermark P (2013) Islet amyloid polypeptide and diabetes. Curr Protein Pept Sci 14:330–337
- 151. Qiu C, Chen G, Cui Q (2012) Towards the understanding of microRNA and environmental factor interactions and their relationships to human diseases. Sci Rep 2:318. doi:10.1038/srep00318
- Jéquier E (2002) Leptin signaling, adiposity, and energy balance. Ann N Y Acad Sci 967:379–388
- Peiser C, McGregor GP, Lang RE (2000) Leptin receptor expression and suppressor of cytokine signaling transcript levels in highfat-fed rats. Life Sci 67:2971–81
- 154. Zlokovic BV, Jovanovic S, Miao W et al (2000) Differential regulation of leptin transport by the choroid plexus and blood-brain barrier and high affinity transport systems for entry into hypothalamus and across the blood-cerebrospinal fluid barrier. Endocrinology 141:1434–41. doi:10.1210/endo.141.4.7435
- 155. Morash B, Li A, Murphy PR et al (1999) Leptin gene expression in the brain and pituitary gland. Endocrinology 140:5995–8. doi: 10.1210/endo.140.12.7288

- Wiesner G, Vaz M, Collier G et al (1999) Leptin is released from the human brain: influence of adiposity and gender. J Clin Endocrinol Metab 84:2270–4. doi:10.1210/jcem.84.7.5854
- Ur E, Wilkinson DA, Morash BA, Wilkinson M (2002) Leptin immunoreactivity is localized to neurons in rat brain. Neuroendocrinology 75:264–72
- Grill HJ, Schwartz MW, Kaplan JM et al (2002) Evidence that the caudal brainstem is a target for the inhibitory effect of leptin on food intake. Endocrinology 143:239–46. doi:10.1210/endo.143.1. 8589
- 159. Guan XM, Hess JF, Yu H et al (1997) Differential expression of mRNA for leptin receptor isoforms in the rat brain. Mol Cell Endocrinol 133:1–7
- Huang XF, Koutcherov I, Lin S et al (1996) Localization of leptin receptor mRNA expression in mouse brain. Neuroreport 7:2635–8
- Shioda S, Funahashi H, Nakajo S et al (1998) Immunohistochemical localization of leptin receptor in the rat brain. Neurosci Lett 243:41–4
- Bouret SG (2010) Neurodevelopmental actions of leptin. Brain Res 1350:2–9. doi:10.1016/j.brainres.2010.04.011
- Paz-Filho G, Wong M-L, Licinio J (2010) The procognitive effects of leptin in the brain and their clinical implications. Int J Clin Pract 64:1808–12. doi:10.1111/j.1742-1241.2010.02536.x
- Xu G, Ji C, Shi C et al (2013) Modulation of hsa-miR-26b levels following adipokine stimulation. Mol Biol Rep 40:3577–3582. doi:10.1007/s11033-012-2431-0
- 165. Amoldussen IAC, Kiliaan AJ, Gustafson DR (2014) Obesity and dementia: adipokines interact with the brain. Eur Neuropsychopharmacol 1–18. doi: 10.1016/j.euroneuro.2014.03. 002
- 166. Sergi G, De Rui M, Coin A et al (2013) Weight loss and Alzheimer's disease: temporal and aetiologic connections. Proc Nutr Soc 72:160–165. doi:10.1017/S0029665112002753
- 167. Liu QY, Chang MNV, Lei JX et al (2014) Identification of microRNAs involved in Alzheimer's progression using a rabbit model of the disease. Am J Neurodegener Dis 3:33–44
- Marwarha G, Dasari B, Prasanthi JRP et al (2010) Leptin reduces the accumulation of Abeta and phosphorylated tau induced by 27hydroxycholesterol in rabbit organotypic slices. J Alzheimers Dis 19:1007–1019. doi:10.3233/JAD-2010-1298
- Schreurs BG (2013) Cholesterol and copper affect learning and memory in the rabbit. Int J Alzheimers Dis 2013:518780. doi:10. 1155/2013/518780
- Woodruff-Pak DS, Agelan A, Del Valle L (2007) A rabbit model of Alzheimer's disease: valid at neuropathological, cognitive, and therapeutic levels. J Alzheimers Dis 11:371–383
- 171. Sparks DL (2008) The early and ongoing experience with the cholesterol-fed rabbit as a model of Alzheimer's disease: the old, the new and the pilot. J Alzheimers Dis 15:641–656
- Ghribi O (2008) Potential mechanisms linking cholesterol to Alzheimer's disease-like pathology in rabbit brain, hippocampal organotypic slices, and skeletal muscle. J Alzheimers Dis 15:673– 684
- 173. Ghribi O, Larsen B, Schrag M, Herman MM (2006) High cholesterol content in neurons increases BACE, beta-amyloid, and phosphorylated tau levels in rabbit hippocampus. Exp Neurol 200:460– 467. doi:10.1016/j.expneurol.2006.03.019
- 174. Ghribi O, Golovko MY, Larsen B et al (2006) Deposition of iron and beta-amyloid plaques is associated with cortical cellular damage in rabbits fed with long-term cholesterol-enriched diets. J Neurochem 99:438–449. doi:10.1111/j.1471-4159.2006.04079.x
- Dhar M, Zhu M, Impey S et al (2014) Leptin induces hippocampal synaptogenesis via CREB-regulated microRNA-132 suppression of p250GAP. Mol Endocrinol 28:1073–1087. doi:10.1210/me. 2013-1332
- 176. Cogswell JP, Ward J, Taylor IA et al (2008) Identification of miRNA changes in Alzheimer's disease brain and CSF yields

putative biomarkers and insights into disease pathways. J Alzheimers Dis 14:27–41

- 177. Wyman SK, Knouf EC, Parkin RK et al (2011) Posttranscriptional generation of miRNA variants by multiple nucleotidyl transferases contributes to miRNA transcriptome complexity. Genome Res 21:1450–1461. doi:10.1101/gr. 118059.110
- Wong H-KA, Veremeyko T, Patel N et al (2013) De-repression of FOXO3a death axis by microRNA-132 and -212 causes neuronal apoptosis in Alzheimer's disease. Hum Mol Genet 22:3077–3092. doi:10.1093/hmg/ddt164
- 179. Wayman GA, Davare M, Ando H et al (2008) An activityregulated microRNA controls dendritic plasticity by downregulating p250GAP. Proc Natl Acad Sci U S A 105:9093– 9098. doi:10.1073/pnas.0803072105
- Crépin D, Benomar Y, Riffault L et al (2014) The over-expression of miR-200a in the hypothalamus of ob/ob mice is linked to leptin and insulin signaling impairment. Mol Cell Endocrinol 384:1–11. doi:10.1016/j.mce.2013.12.016
- 181. Ding Y, Tian M, Liu J et al (2012) Expression profile of miRNAs in APP swe/PS∆E9 transgenic mice. Nan Fang Yi Ke Da Xue Xue Bao 32:1280–1283
- Benoit C, Ould-Hamouda H, Crepin D et al (2013) Early leptin blockade predisposes fat-fed rats to overweight and modifies hypothalamic microRNAs. J Endocrinol 218:35–47. doi:10.1530/ JOE-12-0561
- 183. Sangiao-Alvarellos S, Pena-Bello L, Manfredi-Lozano M et al (2014) Perturbation of hypothalamic microRNA expression patterns in male rats after metabolic distress: impact of obesity and conditions of negative energy balance. Endocrinology 155:1838– 1850. doi:10.1210/en.2013-1770
- Callen DJ, Black SE, Gao F et al (2001) Beyond the hippocampus: MRI volumetry confirms widespread limbic atrophy in AD. Neurology 57:1669–1674
- McDuff T, Sumi SM (1985) Subcortical degeneration in Alzheimer's disease. Neurology 35:123–126
- Saper CB, German DC (1987) Hypothalamic pathology in Alzheimer's disease. Neurosci Lett 74:364–370
- 187. Geekiyanage H, Chan C (2011) MicroRNA-137/181c regulates serine palmitoyltransferase and in turn amyloid β, novel targets in sporadic Alzheimer's disease. J Neurosci 31:14820–14830. doi: 10.1523/JNEUROSCI.3883-11.2011
- Lee SJ, Liyanage U, Bickel PE et al (1998) A detergent-insoluble membrane compartment contains A beta in vivo. Nat Med 4:730– 734. doi:10.1038/nm0698-730
- Vetrivel KS, Cheng H, Kim S-H et al (2005) Spatial segregation of gamma-secretase and substrates in distinct membrane domains. J Biol Chem 280:25892–25900. doi:10.1074/jbc.M503570200
- 190. Uranga RM, Bruce-Keller AJ, Morrison CD et al (2010) Intersection between metabolic dysfunction, high fat diet consumption, and brain aging. J Neurochem 114:344–361. doi:10. 1111/j.1471-4159.2010.06803.x
- 191. Singh B, Parsaik AK, Mielke MM et al (2013) Association of Mediterranean diet with mild cognitive impairment and Alzheimer's disease: a systematic review and meta-analysis. J Alzheimers Dis. doi:10.3233/JAD-130830
- 192. Khanna A, Muthusamy S, Liang R et al (2011) Gain of survival signaling by down-regulation of three key miRNAs in brain of calorie-restricted mice. Aging (Albany NY) 3:223–236
- 193. Wang X, Liu P, Zhu H et al (2009) miR-34a, a microRNA upregulated in a double transgenic mouse model of Alzheimer's disease, inhibits bcl2 translation. Brain Res Bull 80:268–273. doi:10.1016/j.brainresbull.2009.08.006
- 194. Bowes A, Dawson A, Jepson R, McCabe L (2013) Physical activity for people with dementia: a scoping study. BMC Geriatr 13: 129. doi:10.1186/1471-2318-13-129

- 195. Maesako M, Uemura K, Kubota M et al (2012) Exercise is more effective than diet control in preventing high fat diet-induced βamyloid deposition and memory deficit in amyloid precursor protein transgenic mice. J Biol Chem 287:23024–23033. doi:10. 1074/jbc.M112.367011
- Zacharewicz E, Lamon S, Russell AP (2013) MicroRNAs in skeletal muscle and their regulation with exercise, ageing, and disease. Front Physiol 4:266. doi:10.3389/fphys.2013.00266
- 197. Mojtahedi S, Kordi MR, Hosseini SE et al (2012) Effect of treadmill running on the expression of genes that are involved in neuronal differentiation in the hippocampus of adult male rats. Cell Biol Int. doi:10.1002/cbin.10022
- Smith P, Al Hashimi A, Girard J et al (2011) In vivo regulation of amyloid precursor protein neuronal splicing by microRNAs. J Neurochem 116:240–247. doi:10.1111/j.1471-4159.2010.07097.x
- 199. Cosín-Tomás M, Alvarez-López MJ, Sanchez-Roige S et al (2014) Epigenetic alterations in hippocampus of SAMP8 senescent mice and modulation by voluntary physical exercise. Front Aging Neurosci 6:51. doi:10.3389/fnagi.2014.00051
- Elfving B, Christensen T, Ratner C et al (2013) Transient activation of mTOR following forced treadmill exercise in rats. Synapse 67:620–625. doi:10.1002/syn.21668
- 201. Bruel-Jungerman E, Veyrac A, Dufour F et al (2009) Inhibition of PI3K-Akt signaling blocks exercise-mediated enhancement of adult neurogenesis and synaptic plasticity in the dentate gyrus. PLoS One 4, e7901. doi:10.1371/journal.pone.0007901
- 202. Muller AP, Gnoatto J, Moreira JD et al (2011) Exercise increases insulin signaling in the hippocampus: physiological effects and pharmacological impact of intracerebroventricular insulin administration in mice. Hippocampus 21:1082–1092. doi:10.1002/hipo. 20822
- Jick H, Zornberg GL, Jick SS et al (2000) Statins and the risk of dementia. Lancet 356:1627–1631. doi:10.1097/00006254-200104000-00019
- Kandiah N, Feldman HH (2009) Therapeutic potential of statins in Alzheimer's disease. J Neurol Sci 283:230–234. doi:10.1016/j.jns. 2009.02.352
- 205. Kurata T, Miyazaki K, Kozuki M et al (2011) Atorvastatin and pitavastatin improve cognitive function and reduce senile plaque and phosphorylated tau in aged APP mice. Brain Res 1371:161– 170. doi:10.1016/j.brainres.2010.11.067
- 206. Murphy MP, Morales J, Beckett TL et al (2010) Changes in cognition and amyloid-β processing with long term cholesterol reduction using atorvastatin in aged dogs. J Alzheimers Dis 22:135– 150. doi:10.3233/JAD-2010-100639
- 207. Barone E, Di Domenico F, Butterfield DA (2013) Statins more than cholesterol lowering agents in Alzheimer disease: their pleiotropic functions as potential therapeutic targets. Biochem Pharmacol. doi:10.1016/j.bcp.2013.10.030
- Allen RM, Marquart TJ, Albert CJ et al (2012) miR-33 controls the expression of biliary transporters, and mediates statin- and diet-induced hepatotoxicity. EMBO Mol Med 4:882–895. doi: 10.1002/emmm.201201228
- 209. Takwi AAL, Li Y, Becker Buscaglia LE et al (2012) A statinregulated microRNA represses human c-Myc expression and function. EMBO Mol Med 4:896–909. doi:10.1002/emmm. 201101045
- 210. Guillot F, Misslin P, Lemaire M (1993) Comparison of fluvastatin and lovastatin blood–brain barrier transfer using in vitro and in vivo methods. J Cardiovasc Pharmacol 21:339–346. doi:10. 1097/00005344-199302000-00022
- Cufi S, Vazquez-Martin A, Oliveras-Ferraros C et al (2012) Metformin lowers the threshold for stress-induced senescence: a

role for the microRNA-200 family and miR-205. Cell Cycle 11: 1235–1246. doi:10.4161/cc.11.6.19665

- Wang Y, Dai W, Chu X et al (2013) Metformin inhibits lung cancer cells proliferation through repressing microRNA-222. Biotechnol Lett 35:2013–2019. doi:10.1007/s10529-013-1309-0
- 213. Li W, Yuan Y, Huang L et al (2012) Metformin alters the expression profiles of microRNAs in human pancreatic cancer cells. Diabetes Res Clin Pract 96:187–195. doi:10.1016/j.diabres.2011. 12.028
- Blandino G, Valerio M, Cioce M et al (2012) Metformin elicits anticancer effects through the sequential modulation of DICER and c-MYC. Nat Commun 3:865. doi:10.1038/ncomms1859
- 215. Łabuzek K, Suchy D, Gabryel B et al (2010) Quantification of metformin by the HPLC method in brain regions, cerebrospinal fluid and plasma of rats treated with lipopolysaccharide. Pharmacol Rep 62:956–965. doi:10.1016/S1734-1140(10)70357-1
- Moore EM, Mander AG, Ames D et al (2013) Increased risk of cognitive impairment in patients with diabetes is associated with metformin. Diabetes Care 36:2981–2987. doi:10.2337/dc13-0229
- 217. Li J, Deng J, Sheng W, Zuo Z (2012) Metformin attenuates Alzheimer's disease-like neuropathology in obese, leptinresistant mice. Pharmacol Biochem Behav 101:564–574. doi:10. 1016/j.pbb.2012.03.002
- 218. Imfeld P, Bodmer M, Jick SS, Meier CR (2012) Metformin, other antidiabetic drugs, and risk of Alzheimer's disease: a populationbased case–control study. J Am Geriatr Soc 60:916–921. doi:10. 1111/j.1532-5415.2012.03916.x
- 219. Kurinami H, Shimamura M, Sato N et al (2013) Do angiotensin receptor blockers protect against Alzheimer's disease? Drugs Aging 30:367–372. doi:10.1007/s40266-013-0071-2
- 220. Michel MC, Foster C, Brunner HR, Liu L (2013) A systematic comparison of the properties of clinically used angiotensin II type 1 receptor antagonists. Pharmacol Rev 65:809–848. doi:10.1124/ pr.112.007278
- 221. Fellmann L, Nascimento AR, Tibiriça E, Bousquet P (2013) Murine models for pharmacological studies of the metabolic syndrome. Pharmacol Ther 137:331–340. doi:10.1016/j.pharmthera. 2012.11.004
- 222. Alzoubi KH, Aleisa AM, Alkadhi KA (2005) Impairment of longterm potentiation in the CA1, but not dentate gyrus, of the hippocampus in obese Zucker rats: role of calcineurin and phosphorylated CaMKII. J Mol Neurosci 27:337–346. doi:10.1385/ JMN:27:3:337
- 223. Gerges NZ, Aleisa AM, Alkadhi KA (2003) Impaired long-term potentiation in obese Zucker rats: possible involvement of presynaptic mechanism. Neuroscience 120:535–539. doi:10.1016/ S0306-4522(03)00297-5
- 224. Kamal A, Ramakers GMJ, Gispen WH et al (2013) Hyperinsulinemia in rats causes impairment of spatial memory and learning with defects in hippocampal synaptic plasticity by involvement of postsynaptic mechanisms. Exp Brain Res 226: 45–51. doi:10.1007/s00221-013-3409-4
- 225. Knight DS, Mahajan DK, Qiao X (2001) Dietary fat up-regulates the apolipoprotein E mRNA level in the Zucker lean rat brain. Neuroreport 12:3111–3115. doi:10.1097/00001756-200110080-00026
- Doherty GH, Beccano-Kelly D, Du Yan S et al (2013) Leptin prevents hippocampal synaptic disruption and neuronal cell death induced by amyloid. Neurobiol Aging 34:226–237. doi:10.1016/j. neurobiolaging.2012.08.003
- 227. Talaei F, Van Praag VM, Shishavan MH et al (2014) Increased protein aggregation in Zucker diabetic fatty rat brain: identification of key mechanistic targets and the therapeutic application of hydrogen sulfide. BMC Cell Biol 15:1. doi:10.1186/1471-2121-15-1