

## GEP- NETS UPDATE

## Genetics of neuroendocrine tumors

Joakim Crona and Britt Skogseid

Department of Medical Sciences, Uppsala University, Rudbecklaboratoriet, Dag Hammarskjölds väg 20, 75185 Uppsala, Sweden

Correspondence should be addressed to J Crona  
**Email**  
joakim.crona@medsci.uu.se

## Abstract

Neuroendocrine tumors (NETs) are a heterogeneous group of neoplasms, arising from neuroendocrine cells that are dispersed throughout the body. Around 20% of NETs occur in the context of a genetic syndrome. Today there are at least ten recognized NET syndromes. This includes the classical syndromes: multiple endocrine neoplasias types 1 and 2, and von Hippel–Lindau and neurofibromatosis type 1. Additional susceptibility genes associated with a smaller fraction of NETs have also been identified. Recognizing genetic susceptibility has proved essential both to provide genetic counseling and to give the best preventive care. In this review we will also discuss the knowledge of somatic genetic alterations in NETs. At least 24 genes have been implicated as drivers of neuroendocrine tumorigenesis, and the overall rates of genomic instability are relatively low. Genetic intra-tumoral, as well as inter-tumoral heterogeneity in the same patient, have also been identified. Together these data point towards the common pathways in NET evolution, separating early from late disease drivers. Although knowledge of specific mutations in NETs has limited impact on actual patient management, we predict that in the near future genomic profiling of tumors will be included in the clinical arsenal for diagnostics, prognostics and therapeutic decisions.

*European Journal of  
Endocrinology*  
(2016) 174, R275–R290

## Neuroendocrine tumors

Neuroendocrine tumors (NETs) are rare neoplasms originating from neuroendocrine cells most commonly located in the endocrine glands but also in the gastrointestinal and bronchopulmonary systems (Fig. 1) (1). NETs usually display slow proliferation but have a high rate of advanced stage at presentation (2, 3). Neuropeptide hypersecretion is a hallmark of these diseases and often results in distinct hormonal syndromes. These syndromes are associated with both increased morbidity and mortality (2, 4). The median survival of patients with well differentiated NETs is about 10 years, contributing to the relatively high prevalence of these diseases (5, 6).

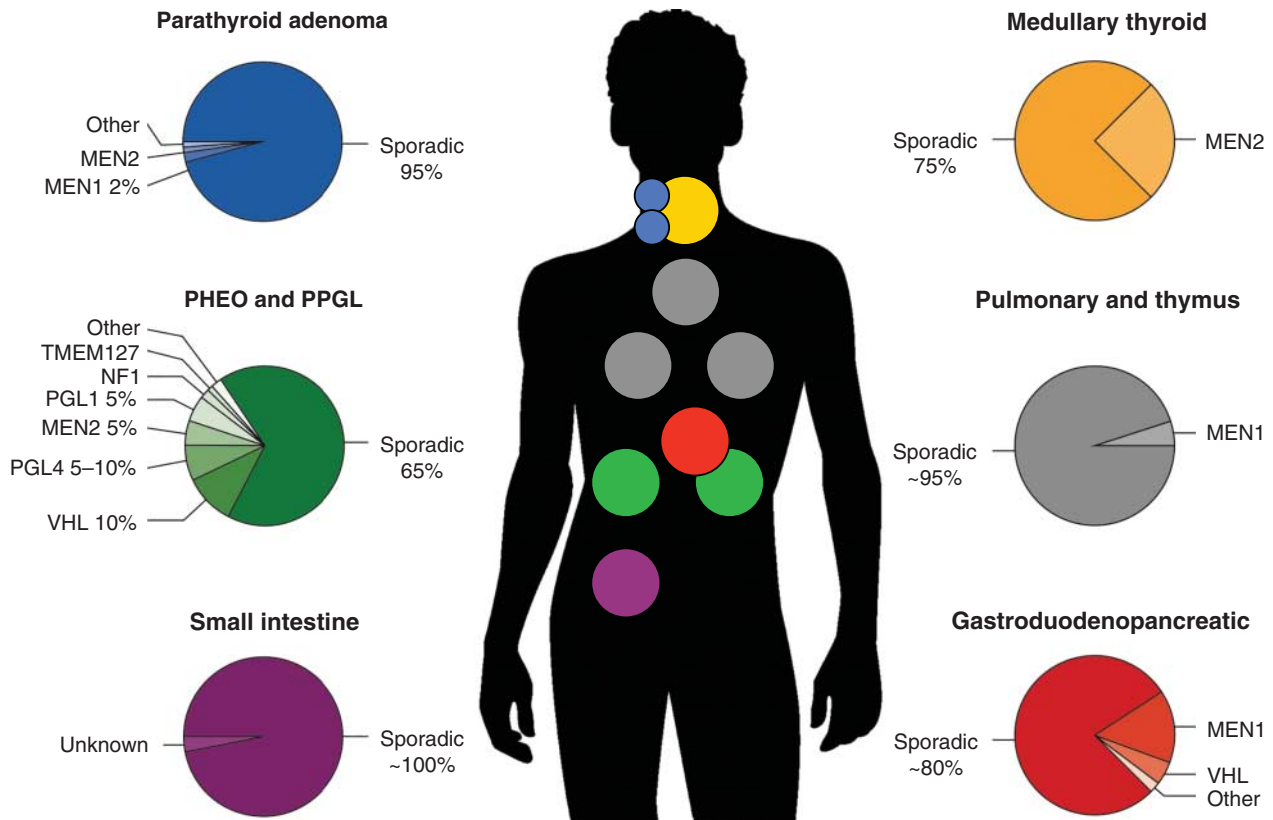
NET is one of the most heritable groups of neoplasms that feature in at least ten genetic syndromes. Additional susceptibility genes associated with a smaller fraction of NETs have also been identified. In this review we will present relevant genetic information regarding these

diagnostic entities. We will also summarize data from tumor sequencing studies that provided comprehensive maps of the genomic landscape of NETs. Finally we comment on the current and future potential of these data as tools for the development of biomarkers as well as novel druggable targets. Below we present syndromes and additional susceptibility genes with increased risk of NET development (Table 1). The present paper reviews genetic profiles, by means of early and late mutations, of relevance for tumorigenesis in NETs.

## Genetic syndromes

## Multiple endocrine neoplasia type 1

Multiple endocrine neoplasia type 1 (MEN1; OMIM 131100) is one of the first described autosomal dominantly inherited complex endocrine syndromes (7). MEN1

**Figure 1**

Overview of genes with recurrent mutations in NETs and their distribution accordingly to anatomical location.

has a high penetrance, over 90% by age 40, and is present in about three per 100 000 individuals. Gene carriers most frequently develop tumors in the parathyroid glands (95%), the anterior pituitary (20–40%), and in the endocrine cells of the pancreas/duodenum (40–80%) (8). Any of these can be the presenting lesion, and typically clinically detectable in the young adult, although rare cases of childhood tumors are reported. By definition MEN1 is present if two of these classical target tissues are affected by tumors, and familial MEN1 includes at least one relative with a corresponding tumor. Apart from the above-mentioned organs, MEN1 carriers frequently develop NETs of the foregut, e.g. bronchial, thymic, and gastric ECLomas. Adrenocortical lesions, mostly non-secreting, hyperplasias or adenomas but also rare cases of adrenocortical carcinomas are also present in MEN1 patients. The most lethal of the MEN1 lesions are the pancreatic and the thymic NETs which frequently develop into metastatic disease. Several non-endocrine tumors are also overrepresented in MEN1 patients, such as facial angiofibromas, collagenomas, lipomas and meningiomas. The typical

clinical picture of the disease is highly variable between family members and dependent of the affected organs and the pattern of hypersecreted hormones in each case.

In 1988 the *MEN1* gene was linked to chromosome 11q13 and suggested to be a suppressor (9). The gene was finally identified by positional cloning, and heterozygous germline inactivating *MEN1* mutations was revealed in 70% of typical index cases (10). The proportion of MEN1 patients with a causative *MEN1* mutation was later estimated to 75–95% (11). By now more than 1000 mutations have been recognized. The vast majority causes truncation of the protein. The most likely cases to reveal a germline *MEN1* mutation are probands developing lesions at young age and show multiple tumors per organ. There is no convincing genotype–phenotype correlation identified so far. Furthermore if a mutation cannot be found it does not exclude MEN1 and recently alternative genes giving rise to syndromes resembling MEN1 have been suggested, e.g. germline *CDKN1B* (p27kip) mutations resulting in MEN4 (12). It has also been postulated that common genetic variants within *CDKN1B* could act as disease

**Table 1** Genetic syndromes with NET manifestation. Overview of genetic syndromes that involve NETs; penetrance of NETs at the specified location is presented in parenthesis; figures on penetrance are presented when available.

Name	Gene	NET manifestation	Other manifestation
MEN1	<i>MEN1</i>	Parathyroid (>90%), gastroenteropancreatic (50%), anterior pituitary (30%), lung and thymus (10%)	Adrenocortical tumors
MEN2A (FMTC)	<i>RET</i>	Medullary thyroid (90–100%), adrenal medulla (20–80%), parathyroid (20%) <sup>a</sup>	
MEN2B	<i>RET</i>	Parathyroid, medullary thyroid (100%), adrenal medulla (50%)	Marfanoid habitus
MEN4 Neurofibromatosis type 1	<i>CDKN1B</i> <i>NF1</i>	Parathyroid, pancreas, pituitary Adrenal medulla (1–5%), duodenum	Neurofibroma, cafe-au-lait spots renal carcinoma
von Hippel–Lindau	<i>VHL</i>	Adrenal medulla and sympathetic ganglia (15%), pancreas (10%)	Hemangioblastoma, renal carcinoma
Familial PGL 1-5	<i>SDHA-D</i> , <i>SDHAF2</i>	Sympathetic and parasympathetic paraganglia, Adrenal medulla	GIST, renal carcinoma
Familial PCC and PGL syndromes	<i>TMEM127</i> , <i>MAX</i> , <i>FH</i> , <i>MDH2</i>	Adrenal medulla, sympathetic ganglia ( <i>TMEM127</i> 30%)	
Polycytemia paraganglioma syndrome	<i>EPAS1</i>	Sympathetic ganglia, adrenal medulla, duodenum	Polycytemia
Tuberous sclerosis complexes HPT-JT syndrome	<i>TSC1</i> , <i>TSC2</i> <i>HRPT2</i>	Pancreas Parathyroid adenoma (80%) and carcinoma (15%)	Hamartoma

<sup>a</sup>Penetrance in MEN2 is specific to the particular mutation.

modifiers in MEN1, possibly contributing to the observed clinical heterogeneity (13).

Classical MEN1 tumorigenesis depends on a second *MEN1* hit, by means of a somatic mutation eliminating also the WT allele. But some NETs in MEN1, such as thymic and duodenal NETs, do not to necessary require a complete inactivation of the gene (14). Somatic homozygous inactivation of *MEN1* is also frequently seen in sporadic tumors of the MEN1 target organs.

The MEN1 protein, named menin, is ubiquitously expressed and preferentially located in the nucleus (10). It has been suggested to be a scaffold protein with more than 40 interacting proteins and thus are involved in a large number of biological functions, such as chromatin modification, DNA repair, transcription, cell division, protein degradation, motility and adhesion (15). Several *Men1* knock-out mouse models mimicking the human syndrome are available.

### Multiple endocrine neoplasias type 2 and familial medullary thyroid cancer

Multiple endocrine neoplasias type 2 (MEN2) and familial medullary thyroid carcinoma (FMTC) are autosomal dominantly inherited disorders characterized by development of multiple endocrine MTC, pheochromocytoma

(PCC) and parathyroid adenomas (reviewed in Wells *et al.* (16)). MEN2 subtype A (OMIM 171400) accounts for about 80% of cases with an almost complete penetrance for MTC, while PCC and parathyroid adenoma are seen in about 50 and 30% of patients (17). MEN2B (Omim 162300) includes more aggressive MTC, dysmorphic marfanoid features but no parathyroid adenoma (18).

MEN2 and FMTC are caused by gain of function mutations in the rearranged during transfection (*RET*) protooncogene, localized on chromosome 10, which encodes a receptor tyrosine kinase (19, 20, 21). Gain of function mutations in *RET* result in autonomous activation that transduces activating signals through the RAS/MAPK and PI3K/AKT pathways (22). A majority of MEN2 and FMTC cases reveal constitutional mutations in the cysteine-rich extracellular domains (exons 10–11) of the *RET* gene (23), while disease causing variants within *RET* non-cysteine regions (exons 13–16) are less common and the related phenotype is characterized by pronounced heterogeneity (24). Patients with constitutional *RET* mutations show strong genotype–phenotype correlations, e.g. codon 634 transitions linked to MEN2A and codon 918 mutations linked to MEN2B. It is critical to recognize the specific *RET* mutation in order to optimize patient and family management, particularly for the timing of prophylactic thyroidectomy and to provide information

on the risk of developing PCC and parathyroid adenoma (25, 26).

FMTc (OMIM 155240) is characterized exclusively by MTC. It has been suggested that FMTc show less aggressive disease characteristics than MTC occurring in the context of MEN2A and B (16, 27).

#### Multiple endocrine neoplasia type 4

Multiple endocrine neoplasia type 4 (MEN4; OMIM 610755) is a rare syndrome that is thought to predispose development of NETs, mainly parathyroid and pituitary adenomas. The trait is inherited through germline mutations in *CDKN1B* encoding p27kip (12). MEN4 seems to occur in an autosomal dominant fashion and is linked to loss of function mutations in the cell cycle regulator *CDKN1B* (12). Future studies have to be performed to confirm the phenotype and penetrance of *CDKN1B* mutations.

#### Neurofibromatosis type 1

Neurofibromatosis type 1 (NF1; OMIM 162200) is an autosomal dominant syndrome that is characterized by multiple endocrinopathies and nervous system manifestations (28). The most common features are fibromatous skin tumors, lichen eye nodules, optic gliomas and café-au-lait spots (29). Endocrinopathies are less common and include PCCs and duodenal NETs (30). Neurofibromatosis type 1 is caused by loss of function mutations in *NF1* that has been linked to deregulation of both rat sarcoma viral oncogene homolog (RAS) proteins and the ERK/MAPK signaling pathway (31, 32).

#### von Hippel–Lindau syndrome

The von Hippel–Lindau syndrome (VHL; OMIM 193300) syndrome has an incidence of ~1/36 000 individuals (33) and is an autosomal dominant disease. VHL is characterized by increased risk of tumors and cysts; retinal and CNS hemangioblastomas, PCC, paragangliomas (PGLs), renal clear cell carcinomas, renal cysts, pancreatic NETs, pancreatic cysts and endolymphatic sac tumors.

The syndrome is caused by inactivating mutations in the *VHL* tumor suppressor gene, located in 3p25 that is involved in the oxygen-sensing pathway through regulation of hypoxia-inducible factors (34). Truncation of *VHL* results in decreased ubiquitination of HIF transcription factors resulting in increased expression of target genes such as VEGFA etc. *VHL* is subclassified into

type 1 (truncating mutations) and type 2 (missense mutations) that differ in clinical presentation (35). Subtype 1 presents without PCC whereas type 2 have a high penetrance of PCC. Subtype 2 may be further subclassified into type 2A without renal cell carcinoma or pancreatic cysts that may be present in subtype 2B.

#### PCC and PGL syndromes

There are at least 12 genes that have proved to confer susceptibility to PCCs and/or PGLs: *SDHA* (36), *SDHB* (37), *SDHC* (38), *SDHD* (39), *SDHAF2* (40), *FH* (41), *VHL* (42), *EPAS1* (43), *NF1* (32), *RET* (20), *TMEM127* (44) and *MAX* (45). The classic genetic syndromes MEN2, NF1 and VHL have been presented above. In the following section we will give an overview to more recently described diagnoses that show pronounced heterogeneity both in term of the affected organs, disease penetrance and mode of inheritance.

#### Familial PGL

Familial PGL syndromes types 1–5 are transmitted in an autosomal dominant manner. These syndromes are caused by the loss of function mutations in *SDHD* (PGL 1), *SDHAF2* (PGL 2), *SDHC* (PGL 3), *SDHB* (PGL 4) and *SDHA* (PGL 5). *SDHx* genes (*SDHA*, *SDHB*, *SDHC* and *SDHD*) encode succinate dehydrogenase subunits that are catalyzing reactions in tricarboxylic acid cycle and in the respiratory electron transfer chain. Due to the disruption of the tricarboxylic acid cycle the associated PGL harbor unique metabolic profiles with accumulation of oncometabolites (46). Although familial PGL syndromes share pathogenic mechanisms, with disruption of the succinate dehydrogenase complex, the phenotype varies between the different subtypes (47). The molecular rationale for this heterogeneity remains to be identified. Paternal transmission has been shown to occur in PGL 1 and 2.

Familial PGL type 1 (OMIM 168000) have an almost complete penetrance for parasympathetic tumors in the head–neck region (48, 49). Furthermore, unilateral PCC and/or sympathetic PGLs are seen in about 25% of patients respectively (50, 51).

Familial PGL type 2 (OMIM 601650) has so far only been detected in a few European families (40, 52, 53, 54), and all reported patients have presented with parasympathetic lesions.

Familial PGL type 3 (605373) is also a rare condition that is mainly manifested by parasympathetic tumors (55, 56, 57).

Familial PGL type 4 (OMIM 115310) is associated with significant morbidity and increased mortality due to the

substantial risk of development of malignant sympathetic PGLs (51, 58, 59). PCC and parasympathetic PGL also occur. In addition, patients with PGL 4 have an increased risk of developing gastrointestinal stromal tumors (GIST) as well as renal cell carcinoma (50).

Familial PGL type 5 (OMIM 614165) is associated with PCC, PGL as well as GIST. Concomitant presentation of two or more of these three tumor types seems to be exceedingly rare (36, 60, 61).

### Familial PCC and PGL syndromes

Patients with familial PCC and PGL of the TMEM127 subtype (OMIM 613403) have an estimated penetrance of PCC of 30% whereas abdominal PGL are less frequent (44, 62, 63, 64). No other phenotypes have been described. This disorder presents in patients with loss of function mutations in the *Transmembrane protein 127* gene that is linked to dysinhibition of the mammalian target of rapamycin (mTOR) pathway.

*Myc-associated factor X* (MAX) associated familial PCC and PGL (OMIM 154950) is an autosomal dominant disorder with a suggested paternal mode of transmission (45, 65). These patients show susceptibility to PCCs and PGLs with unknown penetrance. No other phenotypes have been described. This syndrome is caused by loss of function mutations in MAX that results in deregulation the MYC-MAX-MXD1 pathway (45).

*Fumarate hydratase* (FH) associated PCC and PGL (OMIM 136850) is an autosomal dominant syndrome previously known to result in susceptibility to leiomyomatosis and renal cell cancer (OMIM 150800 (66)). PCC and PGL was recently recognized as a feature of this syndrome and the penetrance is currently unknown (41, 67, 68). The syndrome is caused by function mutations in FH that results in reduced enzymatic activity with fumarate accumulation.

A recent study described germline mutations in *MDH2* as a cause of PGL (69). Although this finding needs to be confirmed, it is indeed remarkable that *MDH2* is the third Krebs cycle gene suggested to be involved in PGL tumorigenesis (69).

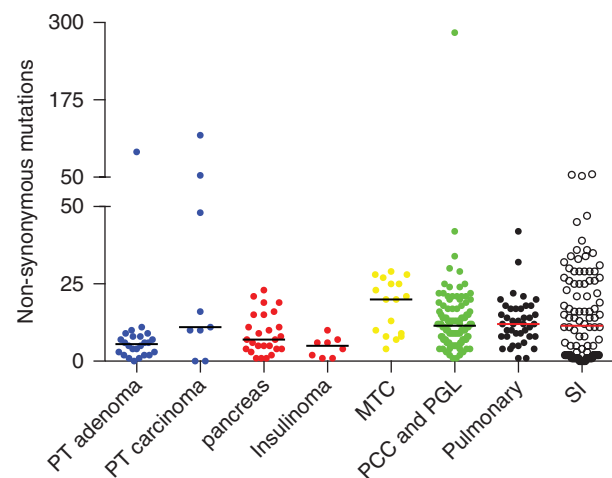
### Polycytemia and PGL syndrome

Disruption of oxygen sensing is also a recurrent disease mechanism in NETs. The recently recognized polycytemia and PGL syndrome was shown to occur as a result gain of function mutations in *EPAS1* that resulted in a pseudohypoxic state through reduction in HIF2 $\alpha$

degradation mediated by disturbed VHL binding (43, 70, 71, 72). The mode of inheritance for the polycytemia and PGL syndrome (OMIM 603349) is currently unknown due to near exclusive presentation in mosaicism. These patients are prone to develop PGL, PCC, polycytemia and somatostatinoma (72). A recent study also suggested ocular manifestations in patients with polycytemia and PGL syndrome (73). A majority of patients with mosaic mutations have had female gender and the underlying mechanism for this phenomenon is unknown (72).

### Tuberous sclerosis complex

Tuberous sclerosis complex subtypes 1 and 2 (OMIM 191100 and 613254) are autosomal dominant syndromes that occur in 1:6000–10 000 individuals (74, 75, 76). These patients show multiorgan manifestations; mainly hamartomas but also angiomyolipomas, renal cell carcinoma and pulmonary lymphangiomyomatosis (reviewed by Curatolo *et al.* (77)). Pancreatic NET is a less frequent manifestation (78). These syndromes are due to the loss of function mutations in *TSC1* and *TSC2* that encode proteins forming the tuberin-hamartin complex that is essential for mTOR signaling (79). Somatic mutations in *TSC2* occur frequently in pancreatic NETs (80).



**Figure 2**

Overview of mutational burden in NETs merged from raw data in (80, 85, 86, 87, 98, 99, 100, 104, 117, 120, 121, 122, 128, 136, 137). Each dot represents a unique tumor, and the line shows the median number of mutations of each category. PT, parathyroid.

## Hyperparathyroidism-jaw tumor syndrome

Hyperparathyroidism-jaw tumor syndrome (OMIM 145001) is an autosomal dominant syndrome caused by truncating mutations in the *HRPT2* located on 1q31.2 (81). It is characterized by adenomatous or malignant lesions of the parathyroids, jaw tumors, as well as renal and uterine tumors (82, 83).

## Genetic landscape of NETs

Compared with other tumor types the genetic landscape of NETs is characterized by relatively few mutations and chromosomal aberrations per tumor (Fig. 2). So far, around 24 genes have been associated with neuroendocrine tumorigenesis (Table 2). These genes seem to interconnect to important pathways involved in cell metabolism, chromatin modification and growth control (Fig. 3).

### Parathyroid adenoma and carcinoma

Constitutional mutations in *CDC73* (81), *MEN1* (84), *RET* (20) and possibly *CDKN1B* (12) and *CASR* (83) cause susceptibility to parathyroid adenoma. Two exome sequencing studies investigated the genomic landscape

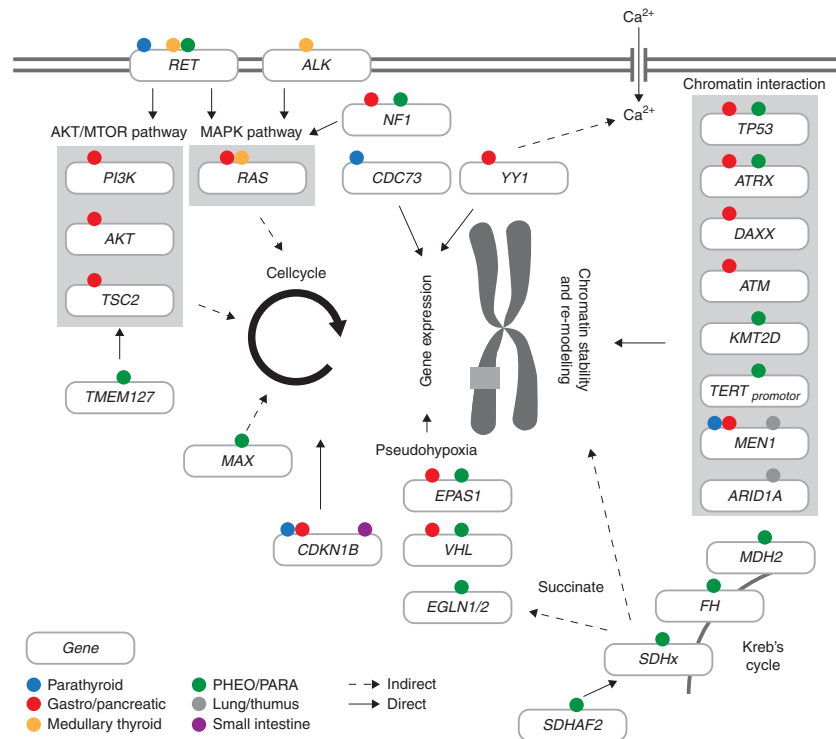
of parathyroid adenomas and discovered somatic *MEN1* mutations in 35% of cases (85, 86). A third study investigated the exomes of parathyroid carcinomas identified recurrent mutations in the *PRUNE2* gene (87). The median number of somatic amino acid substituting mutations per adenoma was eight (range 2–100) (86). Parathyroid carcinomas harbored more mutations (synonymous average 51, range 3–176) that showed an apolipoprotein B mRNA editing enzyme, catalytic polypeptide-like (APOBEC) like mutational signature (87). APOBEC associated genomic instability is a result of cytosine-to-uracil deamination that is catalyzed by APOBEC3A-H enzymes and have been shown to occur in multiple different types of carcinomas (88).

### Gastric, duodenal and pancreatic NETs

Constitutional mutations in *MEN1* (84), *VHL* (42), *NF1* (32), *EPAS1* (somatostatinoma) (43), as well as *TSC1* and 2 (74, 75) cause susceptibility to pancreatic NETs. *CDKN1B* have also been suggested as a susceptibility locus, but limited numbers of cases have been presented (12). In sporadic pancreatic NETs *ATRX*, *DAXX*, *MEN1*, *TP53*, *ATM* and mTOR pathway related genes are commonly mutated in a somatic fashion (80, 89, 90, 91). Pancreatic NETs harboring somatic mutations in *ATRX* and *DAXX* have

**Table 2** List of genes involved in neuroendocrine tumorigenesis. Overview of genes involved in NET tumorigenesis. ALT, alternative lengthening of telomeres.

Gene	Gene function	Chronological classification	Organ specificity
<i>ATM</i>	Chromatin integrity	Unknown	Pancreas
<i>ATRX</i>	ALT	Late	Pancreas, adrenal medulla, paraganglia
<i>CDKN1B</i>	Cell cycle	Unknown	Pancreas, small intestine, parathyroid, anterior pituitary
<i>DAXX</i>	ALT	Late	Pancreas
<i>EPAS1</i>	Cell signaling	Early	Paraganglia, adrenal medulla, duodenum
<i>H-, K-RAS</i>	Cell signaling	Early	Thyroid C-cell, adrenal medulla
<i>FH</i>	Metabolism	Early	Adrenal medulla, paraganglia
<i>KTMD2</i>	Chromatin modification	Unknown	Adrenal medulla
<i>MAX</i>	Cell signaling	Early	Paraganglia, adrenal medulla
<i>MDH2</i>	Metabolism	Unknown	Paraganglia
<i>MEN1</i>	Unknown	Early	Parathyroid, anterior pituitary, endocrine cells in pancreas, duodenum, gastrum, lung, thymus
<i>NF1</i>	Cell signaling	Early	Adrenal medulla, duodenum
<i>RET</i>	Cell signaling	Early	thyroid C-cell, adrenal medulla, parathyroid
<i>SDHx</i>	Metabolism	Early	Paraganglia, adrenal medulla
<i>TERT</i> promoter	Telomere maintenance	Late	Paraganglia
<i>TMEM127</i>	Cell signaling	Early	Paraganglia, adrenal medulla
<i>TP53</i>	Chromatin integrity, Cell signaling	Unknown	Endocrine pancreas, adrenal medulla
<i>TSC1-2</i>	Cell signaling	Unknown	Pancreas
<i>YY1</i>	Transcriptional regulation	Early	Pancreas

**Figure 3**

Simplified overview of genes and pathways involved in NET tumorigenesis.

been associated with chromosomal instability and reduced survival (92). It has been suggested that *ATRX* and *DAXX* mutations are not involved in the initial phase of tumorigenesis. Instead these mutations are thought to be late events that result in malignant transformation of PNETs (92, 93). The proposed pathogenic mechanism of *ATRX* and *DAXX* deficiency is the activation of the alternative lengthening of telomeres phenotype (94, 95). This phenotype has been associated with genome rearrangements, defects in the G2/M checkpoint and altered double-strand break repair (95). It has also been suggested that *ATRX/DAXX* mutations lead to differential epigenetic deregulation (96). The mean number of amino acid substituting somatic mutations in non-functioning pancreatic NETs were 16 (80). The degree of chromosomal aberrations is more variable and ranges from no somatic copy number alterations to genome-wide aneuploidy (92, 97).

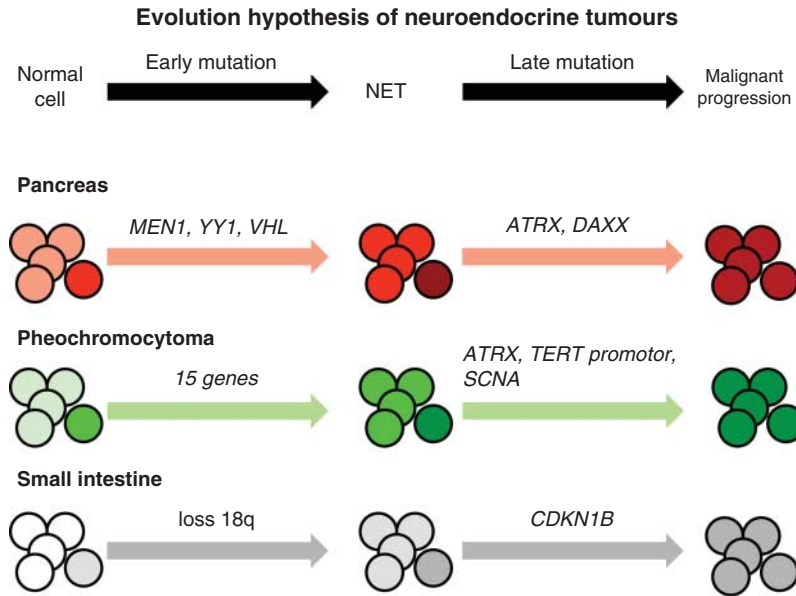
Insulin producing pancreatic NETs with manifest hypoglycemic symptoms, i.e. insulinomas, have been identified with recurrent somatic mutations in *YY1*. This gene encodes a nuclear transcription factor and the described recurrent mutation Thr372Arg has been shown to result in neomorphic effects and altered transcription

(98, 99, 100). Insulinomas exhibit a low mutational burden and the average amino acid substituting mutations were four per tumor (98, 99, 100). High chromosomal instability has been recognized in a subset of insulinomas especially in those with malignant features (101).

Although limited studies have been presented, evidence indicates that the genetics of duodenal and gastric NETs overlap with the profile observed in pancreatic NETs (90, 102).

### Medullary thyroid carcinoma

Germline mutations in *RET* cause MEN 2 and FMTC; both syndromes showing high penetrance of MTC (20). MTC from MEN2 patients rarely show any other somatic driver mutations (103). Investigating the genomic landscape of sporadic MTCs revealed mutually exclusive oncogenic mutations in *RET* and *RAS* subtypes *K* and *H* in 75 and 15% respectively (104, 105, 106). Furthermore there were an average of 18 amino acid substituting somatic mutations per MTC (104). *ALK* gene fusions were recently described in two out of 98 investigated cases suggesting that *ALK* inhibition might further be evaluated for treatment of these rare tumors (107, 108).

**Figure 4**

Theory of evolution, with relevant driver genes, in NET tumorigenesis. The 15 genes involved in pheochromocytoma

and PGL tumorigenesis are: *SDHx, SDHAF2, MDH2, VHL, EPAS1, NF1, RET, HRAS, TMEM127, MAX, KIF1B* and *KMT2D*.

### PCC and PGLs

Mutations in up to 20 genes have been suggested with PCC and PGL tumorigenesis. A total of 12 genes is believed cause of genetic susceptibility to PCC and PGL and a further six loci have been suggested but remains to be validated. These include *KIF1Bβ* (109, 110), *IDH1* (111), *MDH2* (69), *BAP1* (112) as well as *EGLN* subtypes 1 and 2 (113, 114). Somatic mutations in *HRAS* have been shown to cause these diseases (115, 116). Several genes have also been suggested to work as the disease modifiers of PCC and PGL. *ATRX* have been identified in a subset of malignant PCC and PGL having the alternative lengthening of telomeres phenotype (117). *TERT* promoter mutations have been identified to occur exclusively in succinate dehydrogenase deficient PGL (118, 119). A recent study also described the recurrent somatic mutations in *KMT2D*. Whether this gene acts as a disease driver or modifier remains to be identified (120). The genomic landscape of PCC and PGL is associated with relatively low degree of genomic instability having an average of 19 non-synonymous mutations per tumor (117, 121, 122). Clinical criteria have classified these tumors accordingly to location into PCCs (adrenal medulla) and PGLs (extra adrenal). Accumulating evidence show that it is possible to sub-classify PCC and PGL into at least three distinct molecular clusters (based on methylome, transcriptome

and siRNAome) having unique mutational profiles as well as different clinical characteristics (41, 121, 123). Cluster 1a is enriched with noradrenaline producing PGLs that show mutations in genes involved in cell metabolism through the Krebs cycle (*SDHX, FH* and *MDH2*) (121). Cluster 1b has noradrenaline producing PCC and PGL with a pseudohypoxic phenotype (*VHL* and *EPAS1*) (43, 123). Cluster 2 has PCC and PGLs with a mixed adrenaline/noradrenaline phenotype having aberrantly activated signaling in the MAPK and PI3K/AKT signaling pathways (*RET, NF1, TMEM127, MAX* and *HRAS*) (45, 115, 124, 125) (Fig. 4).

### NETs of the lung

This section describes the genomic findings of well differentiated NETs derived from the bronchopulmonary system. These have classically been denoted typical and atypical pulmonary carcinoids based on morphological criteria (126). Pulmonary carcinoids occur in about 5% of MEN1 patients (127). One study examining genome-wide mutation status in 44 sporadic pulmonary carcinoids identified three genes with recurrent mutations: *MEN1, PSIP1* and *ARID1A* (128). Network analysis of genes with somatic mutations extended the number of cases with relevant mutations, 40% had covalent histone modifiers



and 22% subunits of the SWI/SNF complex (22%) (128). The occurrence of somatic *MEN1* mutations in sporadic pulmonary carcinoids have previously been highlighted (129). There were the averages of 13 protein-altering mutations per sample (128). Differential expression of miRNAs in typical vs atypical lung carcinoids has also been reported (130).

### Thymic carcinoid

NET of the thymus is a part of MEN1 syndrome with a penetrance of <10% (8). CGH analysis found recurrent copy number alterations with the most commonly disturbance identified being gain of 8q24 (*MYC* locus) (131). Mutational screening of tumor tissue has been performed on a per case-basis with the finding of a few somatic *MEN1* mutations (132).

### Small intestinal NETs

Small intestinal NETs (SI-NETs) have also been described to occur with familial aggregation in a small fraction of patients (133, 134). Despite substantial efforts no definitive genetic basis for this phenomenon has been described. A recent study used linkage analysis and exome sequencing that revealed *IPMK* germline mutations in a single family (135). Although experimental investigations seem to indicate a pathogenic effect of the *IPMK* mutation a validation effort that included 32 additional families did not reveal the presence of any additional mutations (135). Thus *IPMK* inactivation is not likely to be a significant cause of familial SI-NET. The genomic landscape of SI-NETs has been investigated with exome coverage in close to 100 tumor samples (136, 137). Recurrent mutations were only identified in the *CDKN1B* gene with a mutation prevalence of 9% of SI-NET patients (136, 137, 138). Instead, the most frequent genomic alteration is hemizygous deletions affecting chromosome 18q (133, 136, 137, 139). Recent data have shown that SI-NETs can be sub-classified based on tumor methylome into three distinct clusters (140, 141). Sub classification of SI-NETs based on gene expression may also provide relevant information (142).

### Genetic heterogeneity and tumor evolution

As clinicians seek to select treatments based on genetic biomarkers, knowledge of spatial and temporal heterogeneity could be important and should be researched further. Indeed genetic heterogeneity within and between paired tumors has been described in most NET types. Consistent with the theoretical assumption that MEN1 patients experience parallel development of independent

NET clones, studies using loss of heterozygosity (LOH) LOH markers were able to show that paired tumors from MEN1 patients have unique genetic compositions (143, 144). In contrast, X-chromosome inactivation studies of sporadic parathyroid adenomas, gastrinomas, gastric-NETs and MTCs indicate that these tumors develop in a monoclonal fashion (145, 146, 147, 148). Multiple intestinal tumor lesions have been observed in about 20% of SI-NET patients (149). Whether these are the consequences of independent tumor formation or metastatic spread remain to be settled (144, 145). Recent studies highlighted that a subset of SI-NETs show pronounced genetic heterogeneity within and between tumor lesions (136, 138). Korpershoek *et al.* (150) determined, by using LOH markers, that there is genetic heterogeneity within different areas of selected PCCs and PGLs. Furthermore they recognized that adrenal medullary hyperplasia in MEN2 patients is in fact a precursor lesion for PPGL (151). Widespread genetic heterogeneity within these tumors as well as and between paired lesions has been recently shown (152). Of particular notice was the observed differences between paired primary and metastatic tumor lesions as well as indication of parallel evolution of different metastatic clones (152).

### Authors perspectives and future implications

Precision medicine, where the unique properties of patients and tumors are used to tailor diagnostic and therapeutic procedures is now the standard of care of selected cancer types. For instance in carcinomas of the lung and gastrointestinal system knowledge of tumor mutation status is used to predict the response of treatment and detect the emergence of resistance during treatment with kinase signaling inhibitors (153, 154). The concept of precision medicine has also been useful in the management of patients with NETs, mainly for identification of those with genetic syndromes, enabling genetic counseling followed by appropriate diagnostic measures. However, knowledge of tumor mutation status is not yet used on a routine basis in NETs. Ongoing research is currently investigating whether specific *RET* mutations could predict the success of treatment with receptor kinase inhibitors (NCT01945762) in MTC. Another hypothesis that needs to be tested is that mutations in genes involved in mTOR signaling could be used as biomarkers of rapalogue response (155, 156).

Recent times also saw the publication of experimental findings in NETs that could be important for the

development of future therapies: both combination therapy with *FAK* and mTOR inhibitors (157) as well as inhibition of B-catenin in MEN1 deficient pancreatic NETs (158). Immunotherapy is also an emerging branch in cancer treatment where the knowledge of tumor genetics and biology has proved to be important for prediction. Through determination of the degree of genomic instability and the neo-antigen immunoreactivity the success of PD-1 and CTLA-4 blockade may be estimated (159, 160). On the experimental level mapping the unique somatic mutations present in a particular tumor can also be used to design personalized tumor vaccination protocols (161). Furthermore, genetic instability could be targeted in *ATRX* or *DAXX* mutated tumors as they harbor the unique alternative lengthening of telomeres phenotype. Recent experimental data suggest that such tumors are sensitive to ATR inhibition (162).

However hypotheses based on extrapolation of data from basic NET research needs to be thoroughly tested in a clinical setting. There is an on-going debate whether currently available NET models are representative for human NET disease, supported by the discrepancies of the genomic landscape between NETs and commonly used cell lines (163, 164). Interpretation of basic science data must therefore be made with caution.

In order to screen for genetic disturbances in the established NET genes in a cost effective manner, reliable protocols for novel deep sequencing techniques should be implemented (165, 166). These techniques could also facilitate improvement of diagnostic yield of patients with suspicion of heritable disease. It has also been proposed that deep sequencing can improve the sensitivity in the detection of somatic mutations, especially in scenarios with a low tumor cell fraction (167). Current strategies to analyze tumor genomes are limited by the need of sufficient amount of good quality tumor tissue. In the future peripheral blood samples (liquid biopsies) containing either circulating tumor cells or circulating cell free DNA might instead be used for recognition of the genomic landscape of the individual tumors (168, 169).

#### Declaration of interest

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the review.

#### Funding

Our research is supported by Lions Cancerforskningsfond Uppsala and Swedish Cancer Foundation. Joakim Crona holds a research position funded by Akademiska Sjukhuset.

#### Acknowledgements

We thank Professor Peter Stålberg for his valuable suggestions and comments.

#### References

- 1 Modlin IM, Oberg K, Chung DC, Jensen RT, de Herder WW, Thakker RV, Caplin M, Delle Fave G, Kaltsas GA, Krenning EP *et al*. Gastroenteropancreatic neuroendocrine tumours. *Lancet. Oncology* 2008 **9** 61–72. (doi:10.1016/S1470-2045(07)70410-2)
- 2 Ekeblad S, Skogseid B, Dunder K, Oberg K & Eriksson B. Prognostic factors and survival in 324 patients with pancreatic endocrine tumor treated at a single institution. *Clinical Cancer Research* 2008 **14** 7798–7803. (doi:10.1158/1078-0432.CCR-08-0734)
- 3 Norlen O, Stalberg P, Oberg K, Eriksson J, Hedberg J, Hessman O, Janson ET, Hellman P & Akerstrom G. Long-term results of surgery for small intestinal neuroendocrine tumors at a tertiary referral center. *World Journal of Surgery* 2012 **36** 1419–1431. (doi:10.1007/s00268-011-1296-z)
- 4 Grande E, Capdevila J, Barriuso J, Anton-Aparicio L & Castellano D. Gastroenteropancreatic neuroendocrine tumor cancer stem cells: do they exist? *Cancer Metastasis Reviews* 2012 **31** 47–53. (doi:10.1007/s10555-011-9328-6)
- 5 Hallet J, Law CH, Cukier M, Saskin R, Liu N & Singh S. Exploring the rising incidence of neuroendocrine tumors: a population-based analysis of epidemiology, metastatic presentation, and outcomes. *Cancer* 2015 **121** 589–597. (doi:10.1002/cncr.29099)
- 6 Yao JC, Hassan M, Phan A, Dagohoy C, Leary C, Mares JE, Abdalla EK, Fleming JB, Vauthey JN, Rashid A *et al*. One hundred years after "carcinoid": epidemiology of and prognostic factors for neuroendocrine tumors in 35,825 cases in the United States. *Journal of Clinical Oncology* 2008 **26** 3063–3072. (doi:10.1200/JCO.2007.15.4377)
- 7 Wermer P. Genetic aspects of adenomatosis of endocrine glands. *American Journal of Medicine* 1954 **16** 363–371. (doi:10.1016/0002-9343(54)90353-8)
- 8 Brandi ML, Gagel RF, Angeli A, Bilezikian JP, Beck-Peccoz P, Bordi C, Conte-Devolx B, Falchetti A, Gheri RG, Libroia A *et al*. Guidelines for diagnosis and therapy of MEN type 1 and type 2. *Journal of Clinical Endocrinology and Metabolism* 2001 **86** 5658–5671. (doi:10.1210/jcem.86.12.8070)
- 9 Larsson C, Skogseid B, Oberg K, Nakamura Y & Nordenskjold M. Multiple endocrine neoplasia type 1 gene maps to chromosome 11 and is lost in insulinoma. *Nature* 1988 **332** 85–87. (doi:10.1038/332085a0)
- 10 Chandrasekharappa SC, Guru SC, Manickam P, Olufemi SE, Collins FS, Emmert-Buck MR, Debelenko LV, Zhuang Z, Lubensky IA, Liotta LA *et al*. Positional cloning of the gene for multiple endocrine neoplasia-type 1. *Science* 1997 **276** 404–407. (doi:10.1126/science.276.5311.404)
- 11 Thakker RV. Multiple endocrine neoplasia type 1 (MEN1) and type 4 (MEN4). *Molecular and Cellular Endocrinology* 2014 **386** 2–15. (doi:10.1016/j.mce.2013.08.002)
- 12 Pellegata NS, Quintanilla-Martinez L, Siggelkow H, Samson E, Bink K, Hoffer H, Fend F, Graw J & Atkinson MJ. Germ-line mutations in p27Kip1 cause a multiple endocrine neoplasia syndrome in rats and humans. *PNAS* 2006 **103** 15558–15563. (doi:10.1073/pnas.0603877103)
- 13 Longuini VC, Lourenco DM Jr, Sekiya T, Meirelles O, Goncalves TD, Coutinho FL, Francisco G, Osaki LH, Chammas R, Alves VA *et al*. Association between the p27 rs2066827 variant and tumor multiplicity in patients harboring MEN1 germline mutations. *European Journal of Endocrinology/European Federation of Endocrine Societies* 2014 **171** 335–342. (doi:10.1530/EJE-14-0130)

- 14 Anlauf M, Perren A, Henopp T, Rudolf T, Garbrecht N, Schmitt A, Raffel A, Gimm O, Weihe E, Knoefel WT *et al.* Allelic deletion of the MEN1 gene in duodenal gastrin and somatostatin cell neoplasms and their precursor lesions. *Gut* 2007 **56** 637–644. (doi:10.1136/gut.2006.108910)
- 15 Agarwal SK. Multiple endocrine neoplasia type 1. *Frontiers of Hormone Research* 2013 **41** 1–15. (doi:10.1159/000345666)
- 16 Wells SA Jr, Pacini F, Robinson BG & Santoro M. Multiple endocrine neoplasia type 2 and familial medullary thyroid carcinoma: an update. *Journal of Clinical Endocrinology and Metabolism* 2013 **98** 3149–3164. (doi:10.1210/jc.2013-1204)
- 17 Steiner AL, Goodman AD & Powers SR. Study of a kindred with pheochromocytoma, medullary thyroid carcinoma, hyperparathyroidism and Cushing's disease: multiple endocrine neoplasia, type 2. *Medicine* 1968 **47** 371–409. (doi:10.1097/00005792-196809000-00001)
- 18 Morrison PJ & Nevin NC. Multiple endocrine neoplasia type 2B (mucosal neuroma syndrome, Wagenmann–Froboese syndrome). *Journal of Medical Genetics* 1996 **33** 779–782. (doi:10.1136/jmg.33.9.779)
- 19 Donis-Keller H, Dou S, Chi D, Carlson KM, Tushima K, Lainmore TC, Howe JR, Moley JF, Goodfellow P & Wells SA Jr. Mutations in the RET proto-oncogene are associated with MEN 2A and FMTC. *Human Molecular Genetics* 1993 **2** 851–856. (doi:10.1093/hmg/2.7.851)
- 20 Mulligan LM, Kwok JB, Healey CS, Elsdon MJ, Eng C, Gardner E, Love DR, Mole SE, Moore JK, Papi L *et al.* Germ-line mutations of the RET proto-oncogene in multiple endocrine neoplasia type 2A. *Nature* 1993 **363** 458–460. (doi:10.1038/363458a0)
- 21 Sippl J. The association of pheochromocytoma with carcinoma of the thyroid gland. *American Journal of Medicine* 1961 **31** 163–166. (doi:10.1016/0002-9343(61)90234-0)
- 22 Wells SA Jr & Santoro M. Targeting the RET pathway in thyroid cancer. *Clinical Cancer Research* 2009 **15** 7119–7123. (doi:10.1158/1078-0432.CCR-08-2742)
- 23 Ponder BA. The phenotypes associated with ret mutations in the multiple endocrine neoplasia type 2 syndrome. *Cancer Research* 1999 **59** 1736s–1741s; discussion 1742s.
- 24 Mukherjee S & Zakalik D. RET codon 804 mutations in multiple endocrine neoplasia 2: genotype–phenotype correlations and implications in clinical management. *Clinical Genetics* 2011 **79** 1–16. (doi:10.1111/j.1399-0004.2010.01453.x)
- 25 Kloos RT, Eng C, Evans DB, Francis GL, Gagel RF, Gharib H, Moley JF, Pacini F, Ringel MD, Schlumberger M *et al.* Medullary thyroid cancer: management guidelines of the American Thyroid Association. *Thyroid* 2009 **19** 565–612. (doi:10.1089/thy.2008.0403)
- 26 Chen H, Sippel RS, O'Dorisio MS, Vinik AI, Lloyd RV & Pacak K. The North American Neuroendocrine Tumor Society consensus guideline for the diagnosis and management of neuroendocrine tumors: pheochromocytoma, paraganglioma, and medullary thyroid cancer. *Pancreas* 2010 **39** 775–783. (doi:10.1097/MPA.0b013e3181ebb4f0)
- 27 Farndon JR, Leight GS, Dilley WG, Baylin SB, Smallridge RC, Harrison TS & Wells SA Jr. Familial medullary thyroid carcinoma without associated endocrinopathies: a distinct clinical entity. *British Journal of Surgery* 1986 **73** 278–281. (doi:10.1002/bjs.1800730411)
- 28 Crowe F, Schull W, Neel J. *A Clinical, Pathological and Genetic Study of Multiple Neurofibromatosis*. Springfield, IL: Charles C Thomas, 1956.
- 29 McGaughran JM, Harris DI, Donnai D, Teare D, MacLeod R, Westerbeeck R, Kingston H, Super M, Harris R & Evans DG. A clinical study of type 1 neurofibromatosis in north west England. *Journal of Medical Genetics* 1999 **36** 197–203.
- 30 Griffiths DF, Williams GT & Williams ED. Duodenal carcinoid tumours, phaeochromocytoma and neurofibromatosis: islet cell tumour, phaeochromocytoma and the von Hippel–Lindau complex: two distinctive neuroendocrine syndromes. *Quarterly Journal of Medicine* 1987 **64** 769–782.
- 31 Martin GA, Viskochil D, Bollag G, McCabe PC, Crosier WJ, Haubruck H, Conroy L, Clark R, O'Connell P, Cawthon RM *et al.* The GAP-related domain of the neurofibromatosis type 1 gene product interacts with ras p21. *Cell* 1990 **63** 843–849. (doi:10.1016/0092-8674(90)90150-D)
- 32 Wallace MR, Marchuk DA, Andersen LB, Letcher R, Odeh HM, Saulino AM, Fountain JW, Brereton A, Nicholson J, Mitchell AL *et al.* Type 1 neurofibromatosis gene: identification of a large transcript disrupted in three NF1 patients. *Science* 1990 **249** 181–186. (doi:10.1126/science.2134734)
- 33 Maher ER, Iselius L, Yates JR, Littler M, Benjamin C, Harris R, Sampson J, Williams A, Ferguson-Smith MA & Morton N. Von Hippel–Lindau disease: a genetic study. *Journal of Medical Genetics* 1991 **28** 443–447. (doi:10.1136/jmg.28.7.443)
- 34 Nordstrom-O'Brien M, van der Luijt RB, van Rooijen E, van den Ouweland AM, Majoor-Krakauer DF, Lolkema MP, van Brussel A, Voest EE & Giles RH. Genetic analysis of von Hippel–Lindau disease. *Human Mutation* 2010 **31** 521–537. (doi:10.1002/humu.21219)
- 35 Chen F, Kishida T, Yao M, Hustad T, Glavac D, Dean M, Gnarr JR, Orcutt ML, Duh FM, Glenn G *et al.* Germline mutations in the von Hippel–Lindau disease tumor suppressor gene: correlations with phenotype. *Human Mutation* 1995 **5** 66–75. (doi:10.1002/humu.1380050109)
- 36 Burnichon N, Briere JJ, Libe R, Vescovo L, Riviere J, Tissier F, Jouanno E, Jeunemaitre X, Benit P, Tzagoloff A *et al.* SDHA is a tumor suppressor gene causing paraganglioma. *Human Molecular Genetics* 2010 **19** 3011–3020. (doi:10.1093/hmg/ddq206)
- 37 Astuti D, Latif F, Dallol A, Dahia PL, Douglas F, George E, Skoldberg F, Husebye ES, Eng C & Maher ER. Gene mutations in the succinate dehydrogenase subunit SDHB cause susceptibility to familial pheochromocytoma and to familial paraganglioma. *American Journal of Human Genetics* 2001 **69** 49–54. (doi:10.1086/321282)
- 38 Niemann S & Muller U. Mutations in SDHC cause autosomal dominant paraganglioma, type 3. *Nature Genetics* 2000 **26** 268–270. (doi:10.1038/81551)
- 39 Baysal BE, Ferrell RE, Willett-Brozick JE, Lawrence EC, Myssiorek D, Bosch A, van der Mey A, Taschner PE, Rubinstein WS, Myers EN *et al.* Mutations in SDHD, a mitochondrial complex II gene, in hereditary paraganglioma. *Science* 2000 **287** 848–851. (doi:10.1126/science.287.5454.848)
- 40 Hao HX, Khalimonchuk O, Schraders M, Dephore N, Bayley JP, Kunst H, Devilee P, Cremers CW, Schiffman JD, Bentz BG *et al.* SDH5, a gene required for flavination of succinate dehydrogenase, is mutated in paraganglioma. *Science* 2009 **325** 1139–1142. (doi:10.1126/science.1175689)
- 41 Letouze E, Martinelli C, Lorient C, Burnichon N, Abermil N, Ottolenghi C, Janin M, Menara M, Nguyen AT, Benit P *et al.* SDH mutations establish a hypermethylator phenotype in paraganglioma. *Cancer Cell* 2013 **21** 00183–00189. (doi:10.1016/j.ccr.2013.04.018)
- 42 Latif F, Tory K, Gnarr J, Yao M, Duh FM, Orcutt ML, Stackhouse T, Kuzmin I, Modi W, Geil L *et al.* Identification of the von Hippel–Lindau disease tumor suppressor gene. *Science* 1993 **260** 1317–1320. (doi:10.1126/science.8493574)
- 43 Zhuang Z, Yang C, Lorenzo F, Merino M, Fojo T, Kebebew E, Popovic V, Stratakis CA, Prchal JT & Pacak K. Somatic HIF2A gain-of-function mutations in paraganglioma with polycythemia. *New England Journal of Medicine* 2012 **367** 922–930. (doi:10.1056/NEJMoa1205119)
- 44 Qin Y, Yao L, King EE, Buddavarapu K, Lenci RE, Chocron ES, Lechleiter JD, Sass M, Aronin N, Schiavi F *et al.* Germline mutations in TMEM127 confer susceptibility to pheochromocytoma. *Nature Genetics* 2010 **42** 229–233. (doi:10.1038/ng.533)
- 45 Comino-Mendez I, Gracia-Aznarez FJ, Schiavi F, Landa I, Leandro-Garcia LJ, Leton R, Honrado E, Ramos-Medina R, Caronia D, Pita G *et al.* Exome sequencing identifies MAX mutations as a cause of

- hereditary pheochromocytoma. *Nature Genetics* 2011 **43** 663–667. (doi:10.1038/ng.861)
- 46 Richter S, Peitzsch M, Rapizzi E, Lenders JW, Qin N, de Cubas AA, Schiavi F, Rao JU, Beuschlein F, Quinkler M *et al.* Krebs cycle metabolite profiling for identification and stratification of pheochromocytomas/paragangliomas due to succinate dehydrogenase deficiency. *Journal of Clinical Endocrinology and Metabolism* 2014 **99** 3903–3911. (doi:10.1210/jc.2014-2151)
- 47 Welander J, Soderkvist P & Gimm O. Genetics and clinical characteristics of hereditary pheochromocytomas and paragangliomas. *Endocrine-Related Cancer* 2011 **18** R253–R276. (doi:10.1530/ERC-11-0170)
- 48 van der Mey AG, Maaswinkel-Mooy PD, Cornelisse CJ, Schmidt PH & van de Kamp JJ. Genomic imprinting in hereditary glomus tumours: evidence for new genetic theory. *Lancet* 1989 **2** 1291–1294. (doi:10.1016/S0140-6736(89)91908-9)
- 49 Burnichon N, Rohmer V, Amar L, Herman P, Leboulleux S, Darrouzet V, Niccoli P, Gaillard D, Chabrier G, Chabolle F *et al.* The succinate dehydrogenase genetic testing in a large prospective series of patients with paragangliomas. *Journal of Clinical Endocrinology and Metabolism* 2009 **94** 2817–2827. (doi:10.1210/jc.2008-2504)
- 50 Neumann HP, Pawlu C, Peczkowska M, Bausch B, McWhinney SR, Muresan M, Buchta M, Franke G, Klisch J, Bley TA *et al.* Distinct clinical features of paraganglioma syndromes associated with SDHB and SDHD gene mutations. *Journal of the American Medical Association* 2004 **292** 943–951. (doi:10.1001/jama.292.8.943)
- 51 van Hulsteijn LT, Dekkers OM, Hes FJ, Smit JW & Corssmit EP. Risk of malignant paraganglioma in SDHB-mutation and SDHD-mutation carriers: a systematic review and meta-analysis. *Journal of Medical Genetics* 2012 **25** 25. (doi:10.1136/jmedgenet-2012-101192)
- 52 van Baars F, Cremers C, van den Broek P, Geerts S & Veldman J. Genetic aspects of nonchromaffin paraganglioma. *Human Genetics* 1982 **60** 305–309. (doi:10.1007/BF00569208)
- 53 Bayley JP, Kunst HP, Cascon A, Sampietro ML, Gaal J, Korpershoek E, Hinojar-Gutierrez A, Timmers HJ, Hoefsloot LH, Hermsen MA *et al.* SDHAF2 mutations in familial and sporadic paraganglioma and pheochromocytoma. *Lancet. Oncology* 2010 **11** 366–372. (doi:10.1016/S1470-2045(10)70007-3)
- 54 Casey R, Garrahy A, Tuthill A, O'Halloran D, Joyce C, Casey MB, O'Shea P & Bell M. Universal genetic screening uncovers a novel presentation of an SDHAF2 mutation. *Journal of Clinical Endocrinology and Metabolism* 2014 **99** E1392–E1396. (doi:10.1210/jc.2013-4536)
- 55 Niemann S, Steinberger D & Muller U. PGL3, a third, not maternally imprinted locus in autosomal dominant paraganglioma. *Neurogenetics* 1999 **2** 167–170. (doi:10.1007/s100480050078)
- 56 Hensen EF, van Duinen N, Jansen JC, Corssmit EP, Tops CM, Romijn JA, Vriends AH, van der Mey AG, Cornelisse CJ, Devilee P *et al.* High prevalence of founder mutations of the succinate dehydrogenase genes in the Netherlands. *Clinical Genetics* 2012 **81** 284–288. (doi:10.1111/j.1399-0004.2011.01653.x)
- 57 Mannelli M, Ercolino T, Giache V, Simi L, Cirami C & Parenti G. Genetic screening for pheochromocytoma: should SDHC gene analysis be included? *Journal of Medical Genetics* 2007 **44** 586–587. (doi:10.1136/jmg.2007.051045)
- 58 Gimenez-Roqueplo AP, Favier J, Rustin P, Rieubland C, Crespin M, Nau V, Khau Van Kien P, Corvol P, Plouin PF & Jeunemaitre X. Mutations in the SDHB gene are associated with extra-adrenal and/or malignant pheochromocytomas. *Cancer Research* 2003 **63** 5615–5621.
- 59 Bogdasarian R & Lotz P. Multiple simultaneous paragangliomas of the head and neck in association with multiple retroperitoneal pheochromocytomas. *Otolaryngology – Head and Neck Surgery* 1979 **87** 648–652.
- 60 Korpershoek E, Favier J, Gaal J, Burnichon N, van Gessel B, Oudijk L, Badoual C, Gadessaud N, Venisse A, Bayley JP *et al.* SDHA immunohistochemistry detects germline SDHA gene mutations in apparently sporadic paragangliomas and pheochromocytomas. *Journal of Clinical Endocrinology and Metabolism* 2011 **96** E1472–E1476. (doi:10.1210/jc.2011-1043)
- 61 Pantaleo MA, Astolfi A, Indio V, Moore R, Thiessen N, Heinrich MC, Gnocchi C, Santini D, Catena F, Formica S *et al.* SDHA loss-of-function mutations in KIT-PDGFRα wild-type gastrointestinal stromal tumors identified by massively parallel sequencing. *Journal of the National Cancer Institute* 2011 **103** 983–987. (doi:10.1093/jnci/djr130)
- 62 Yao L, Schiavi F, Cascon A, Qin Y, Inglada-Perez L, King EE, Toledo RA, Ercolino T, Rapizzi E, Ricketts CJ *et al.* Spectrum and prevalence of FP/TMEM127 gene mutations in pheochromocytomas and paragangliomas. *Journal of the American Medical Association* 2010 **304** 2611–2619. (doi:10.1001/jama.2010.1830)
- 63 Abermil N, Guillaud-Bataille M, Burnichon N, Venisse A, Manivet P, Guignat L, Drui D, Chupin M, Josseume C, Affres H *et al.* TMEM127 screening in a large cohort of patients with pheochromocytoma and/or paraganglioma. *Journal of Clinical Endocrinology and Metabolism* 2012 **97** E805–E809. (doi:10.1210/jc.2011-3360)
- 64 Toledo SP, Lourenco DM Jr, Sekiya T, Lucon AM, Baena ME, Castro CC, Bortolotto LA, Zerbini MC, Siqueira SA, Toledo RA *et al.* Penetrance and clinical features of pheochromocytoma in a six-generation family carrying a germline TMEM127 mutation. *Journal of Clinical Endocrinology and Metabolism* 2015 **100** E308–E318. (doi:10.1210/jc.2014-2473)
- 65 Burnichon N, Cascon A, Schiavi F, Paes Morales N, Comino-Mendez I, Abermil N, Inglada-Perez L, de Cubas AA, Amar L, Barontini MB *et al.* MAX mutations cause hereditary and sporadic pheochromocytoma and paraganglioma. *Clinical Cancer Research* 2012 **18** 2828–2837. (doi:10.1158/1078-0432.CCR-12-0160)
- 66 Smit DL, Mensenkamp AR, Badeloe S, Breuning MH, Simon ME, van Spaendonck KY, Aalfs CM, Post JG, Shanley S, Krapels IP *et al.* Hereditary leiomyomatosis and renal cell cancer in families referred for fumarate hydratase germline mutation analysis. *Clinical Genetics* 2011 **79** 49–59. (doi:10.1111/j.1399-0004.2010.01486.x)
- 67 Clark GR, Sciacovelli M, Gaude E, Walsh DM, Kirby G, Simpson MA, Trembath RC, Berg JN, Woodward ER, Kinning E *et al.* Germline FH mutations presenting with pheochromocytoma. *Journal of Clinical Endocrinology and Metabolism* 2014 **99** E2046–E2050. (doi:10.1210/jc.2014-1659)
- 68 Castro-Vega LJ, Buffet A, De Cubas AA, Cascon A, Menara M, Khalifa E, Amar L, Azriel S, Bourdeau I, Chabre O *et al.* Germline mutations in FH confer predisposition to malignant pheochromocytomas and paragangliomas. *Human Molecular Genetics* 2014 **23** 2440–2446. (doi:10.1093/hmg/ddt639)
- 69 Cascon A, Comino-Mendez I, Curras-Freixes M, de Cubas AA, Contreras L, Richter S, Peitzsch M, Mancikova V, Inglada-Perez L, Perez-Barrios A *et al.* Whole-exome sequencing identifies MDH2 as a new familial paraganglioma gene. *Journal of the National Cancer Institute* 2015 **107** Article ID: djv053. (doi:10.1093/jnci/djv053)
- 70 Lorenzo FR, Yang C, Ng Tang Fui M, Vankayalapati H, Zhuang Z, Huynh T, Grossmann M, Pacak K & Prchal JT. A novel EPAS1/HIF2A germline mutation in a congenital polycythemia with paraganglioma. *Journal of Molecular Medicine* 2012 **91** 507–512. (doi:10.1007/s00109-012-0967-z)
- 71 Comino-Mendez I, de Cubas AA, Bernal C, Alvarez-Escola C, Sanchez-Malo C, Ramirez-Tortosa CL, Pedrinaci S, Rapizzi E, Ercolino T, Bernini G *et al.* Tumoral EPAS1 (HIF2A) mutations explain sporadic pheochromocytoma and paraganglioma in the absence of erythrocytosis. *Human Molecular Genetics* 2013 **22** 2169–2176. (doi:10.1093/hmg/ddt069)
- 72 Pacak K, Jochmanova I, Prodanov T, Yang C, Merino MJ, Fojo T, Prchal JT, Tischler AS, Lechan RM & Zhuang Z. New syndrome of paraganglioma and somatostatinoma associated with polycythemia. *Journal of Clinical Oncology* 2013 **31** 1690–1698. (doi:10.1200/JCO.2012.47.1912)

- 73 Pacak K, Chew EY, Pappo AS, Yang C, Lorenzo FR, Wilson MW, Aronow MB, Young JA, Popovic V & Zhuang Z. Ocular manifestations of hypoxia-inducible factor-2 $\alpha$  paraganglioma-somatostatinoma-polycythemia syndrome. *Ophthalmology* 2014 **121** 2291–2293. (doi:10.1016/j.ophtha.2014.06.019)
- 74 van Slechtenhorst M, de Hoogt R, Hermans C, Nellist M, Janssen B, Verhoef S, Lindhout D, van den Ouweland A, Halley D, Young J *et al.* Identification of the tuberous sclerosis gene TSC1 on chromosome 9q34. *Science* 1997 **277** 805–808. (doi:10.1126/science.277.5327.805)
- 75 Identification and characterization of the tuberous sclerosis gene on chromosome 16. *Cell* 1993 **75** 1305–1315. (doi:10.1016/0092-8674(93)90618-Z)
- 76 Osborne JP, Fryer A & Webb D. Epidemiology of tuberous sclerosis. *Annals of the New York Academy of Sciences* 1991 **615** 125–127. (doi:10.1111/j.1749-6632.1991.tb37754.x)
- 77 Curatolo P, Bombardieri R & Jozwiak S. Tuberous sclerosis. *Lancet* 2008 **372** 657–668. (doi:10.1016/S0140-6736(08)61279-9)
- 78 Dworakowska D & Grossman AB. Are neuroendocrine tumours a feature of tuberous sclerosis? A systematic review *Endocrine-Related Cancer* 2009 **16** 45–58. (doi:10.1677/ERC-08-0142)
- 79 Inoki K, Li Y, Zhu T, Wu J & Guan KL. TSC2 is phosphorylated and inhibited by Akt and suppresses mTOR signalling. *Nature Cell Biology* 2002 **4** 648–657. (doi:10.1038/ncb839)
- 80 Jiao Y, Shi C, Edil BH, de Wilde RF, Klimstra DS, Maitra A, Schulick RD, Tang LH, Wolfgang CL, Choti MA *et al.* DAXX/ATRX, MEN1, and mTOR pathway genes are frequently altered in pancreatic neuroendocrine tumors. *Science* 2011 **331** 1199–1203. (doi:10.1126/science.1200609)
- 81 Carpten JD, Robbins CM, Villablanca A, Forsberg L, Presciutti S, Bailey-Wilson J, Simonds WF, Gillanders EM, Kennedy AM, Chen JD *et al.* HRPT2, encoding parafibromin, is mutated in hyperparathyroidism-jaw tumor syndrome. *Nature Genetics* 2002 **32** 676–680. (doi:10.1038/ng1048)
- 82 Jackson CE, Norum RA, Boyd SB, Talpos GB, Wilson SD, Taggart RT & Mallette LE. Hereditary hyperparathyroidism and multiple ossifying jaw fibromas: a clinically and genetically distinct syndrome. *Surgery* 1990 **108** 1006–1012; discussion 1012–1003.
- 83 Pollak MR, Brown EM, Chou YH, Hebert SC, Marx SJ, Steinmann B, Levi T, Seidman CE & Seidman JG. Mutations in the human Ca(2+)-sensing receptor gene cause familial hypocalciuric hypercalcaemia and neonatal severe hyperparathyroidism. *Cell* 1993 **75** 1297–1303. (doi:10.1016/0092-8674(93)90617-Y)
- 84 Lemmens I, Van de Ven WJ, Kas K, Zhang CX, Giraud S, Wautot V, Buisson N, De Witte K, Salandre J, Lenoir G *et al.* Identification of the multiple endocrine neoplasia type 1 (MEN1) gene. The European Consortium on MEN1. *Human Molecular Genetics* 1997 **6** 1177–1183. (doi:10.1093/hmg/6.7.1177)
- 85 Cromer MK, Starker LF, Choi M, Udelsman R, Nelson-Williams C, Lifton RP & Carling T. Identification of somatic mutations in parathyroid tumors using whole-exome sequencing. *Journal of Clinical Endocrinology and Metabolism* 2012 **97** E1774–E1781. (doi:10.1210/jc.2012-1743)
- 86 Newey PJ, Nesbit MA, Rimmer AJ, Attar M, Head RT, Christie PT, Gorvin CM, Stechman M, Gregory L, Mihai R *et al.* Whole-exome sequencing studies of nonhereditary (sporadic) parathyroid adenomas. *Journal of Clinical Endocrinology and Metabolism* 2012 **97** E1995–E2005. (doi:10.1210/jc.2012-2303)
- 87 Yu W, McPherson JR, Stevenson M, vanEijk R, Heng HL, Newey P, Gan A, Ruano D, Huang D, Poon SL *et al.* Whole-exome sequencing studies of parathyroid carcinomas reveal novel PRUNE2 mutations, distinctive mutational spectra related to APOBEC-catalyzed DNA mutagenesis and mutational enrichment in kinases associated with cell migration and invasion. *Journal of Clinical Endocrinology and Metabolism* 2015 **100** E360–E364. (doi:10.1210/jc.2014-3238)
- 88 Burns MB, Temiz NA & Harris RS. Evidence for APOBEC3B mutagenesis in multiple human cancers. *Nature Genetics* 2013 **45** 977–983. (doi:10.1038/ng.2701)
- 89 Perren A, Komminoth P, Saremaslani P, Matter C, Feurer S, Lees JA, Heitz PU & Eng C. Mutation and expression analyses reveal differential subcellular compartmentalization of PTEN in endocrine pancreatic tumors compared to normal islet cells. *American Journal of Pathology* 2000 **157** 1097–1103. (doi:10.1016/S0002-9440(10)64624-X)
- 90 Zhuang Z, Vortmeyer AO, Pack S, Huang S, Pham TA, Wang C, Park WS, Agarwal SK, Debelenko LV, Kester M *et al.* Somatic mutations of the MEN1 tumor suppressor gene in sporadic gastrinomas and insulinomas. *Cancer Research* 1997 **57** 4682–4686.
- 91 Sadanandam A, Wullschlegel S, Lyssiottis CA, Grotzinger C, Barbi S, Bersani S, Korner J, Wafy I, Mafficini A, Lawlor RT *et al.* A cross-species analysis in pancreatic neuroendocrine tumors reveals molecular subtypes with distinctive clinical, metastatic, developmental, and metabolic characteristics. *Cancer Discovery* 2015 **5** 1296–1313. (doi:10.1158/2159-8290.CD-15-0068)
- 92 Marinoni I, Kurrer AS, Vassella E, Dettmer M, Rudolph T, Banz V, Hunger F, Pasquinelli S, Speel EJ & Perren A. Loss of DAXX and ATRX are associated with chromosome instability and reduced survival of patients with pancreatic neuroendocrine tumors. *Gastroenterology* 2014 **146** 453–460.e455. (doi:10.1053/j.gastro.2013.10.020)
- 93 de Wilde RF, Heaphy CM, Maitra A, Meeker AK, Edil BH, Wolfgang CL, Ellison TA, Schulick RD, Molenaar IQ, Valk GD *et al.* Loss of ATRX or DAXX expression and concomitant acquisition of the alternative lengthening of telomeres phenotype are late events in a small subset of MEN-1 syndrome pancreatic neuroendocrine tumors. *Modern Pathology* 2012 **25** 1033–1039. (doi:10.1038/modpathol.2012.53)
- 94 Heaphy CM, de Wilde RF, Jiao Y, Klein AP, Edil BH, Shi C, Bettgowda C, Rodriguez FJ, Eberhart CG, Hebbar S *et al.* Altered telomeres in tumors with ATRX and DAXX mutations. *Science* 2011 **333** 425. (doi:10.1126/science.1207313)
- 95 Lovejoy CA, Li W, Reisenweber S, Thongthip S, Bruno J, de Lange T, De S, Petrini JH, Sung PA, Jasin M *et al.* Loss of ATRX, genome instability, and an altered DNA damage response are hallmarks of the alternative lengthening of telomeres pathway. *PLoS Genetics* 2012 **8** e1002772. (doi:10.1371/journal.pgen.1002772)
- 96 Pipinikas C, Dibra H, Karpathakis A, Feber A, Novelli M, Oukrif D, Fusai G, Valente R, Caplin M, Meyer T *et al.* Epigenetic dysregulation and poorer outcome in DAXX deficient PNETs. *Endocrine-Related Cancer* 2015 **22** L13–L18. (doi:10.1530/ERC-15-0108)
- 97 Hu W, Feng Z, Modica I, Klimstra DS, Song L, Allen PJ, Brennan MF, Levine AJ & Tang LH. Gene amplifications in well-differentiated pancreatic neuroendocrine tumors inactivate the p53 pathway. *Genes & Cancer* 2010 **1** 360–368. (doi:10.1177/1947601910371979)
- 98 Cao Y, Gao Z, Li L, Jiang X, Shan A, Cai J, Peng Y, Li Y, Jiang X, Huang X *et al.* Whole exome sequencing of insulinoma reveals recurrent T372R mutations in YY1. *Nature Communications* 2013 **4** 2810. (doi:10.1038/ncomms3810)
- 99 Lichtenauer UD, Di Dalmazi G, Slater EP, Wieland T, Kuebart A, Schmittfull A, Schwarzmayr T, Diener S, Wiese D, Thasler WE *et al.* Frequency and clinical correlates of somatic Ying Yang 1 mutations in sporadic insulinomas. *Journal of Clinical Endocrinology and Metabolism* 2015 **100** E776–E782. (doi:10.1210/jc.2015-1100)
- 100 Cromer MK, Choi M, Nelson-Williams C, Fonseca AL, Kunstman JW, Korah RM, Overton JD, Mane S, Kenney B, Malchoff CD *et al.* Neomorphic effects of recurrent somatic mutations in Yin Yang 1 in insulin-producing adenomas. *PNAS* 2015 **112** 4062–4067. (doi:10.1073/pnas.1503696112)
- 101 Jonkers YM, Claessen SM, Perren A, Schmitt AM, Hofland LJ, de Herder W, de Krijger RR, Verhofstad AA, Hermus AR, Kummer JA *et al.* DNA copy number status is a powerful predictor of poor survival in endocrine pancreatic tumor patients. *Endocrine-Related Cancer* 2007 **14** 769–779. (doi:10.1677/ERC-07-0111)

- 102 Debelenko LV, Emmert-Buck MR, Zhuang Z, Epshteyn E, Moskaluk CA, Jensen RT, Liotta LA & Lubensky IA. The multiple endocrine neoplasia type I gene locus is involved in the pathogenesis of type II gastric carcinoids. *Gastroenterology* 1997 **113** 773–781. (doi:10.1016/S0016-5085(97)70171-9)
- 103 Cai J, Li L, Ye L, Jiang X, Shen L, Gao Z, Fang W, Huang F, Su T, Zhou Y *et al.* Exome sequencing reveals mutant genes with low penetrance involved in MEN2A-associated tumorigenesis. *Endocrine-Related Cancer* 2015 **22** 23–33. (doi:10.1530/ERC-14-0225)
- 104 Agrawal N, Jiao Y, Sausen M, Leary R, Bettegowda C, Roberts NJ, Bhan S, Ho AS, Khan Z, Bishop J *et al.* Exomic sequencing of medullary thyroid cancer reveals dominant and mutually exclusive oncogenic mutations in RET and RAS. *Journal of Clinical Endocrinology and Metabolism* 2013 **98** E364–E369. (doi:10.1210/jc.2012-2703)
- 105 Moura MM, Cavaco BM, Pinto AE & Leite V. High prevalence of RAS mutations in RET-negative sporadic medullary thyroid carcinomas. *Journal of Clinical Endocrinology and Metabolism* 2011 **96** E863–E868. (doi:10.1210/jc.2010-1921)
- 106 Ciampi R, Mian C, Fugazzola L, Cosci B, Romei C, Barollo S, Cirello V, Bottici V, Marconcini G, Rosa PM *et al.* Evidence of a low prevalence of RAS mutations in a large medullary thyroid cancer series. *Thyroid* 2013 **23** 50–57. (doi:10.1089/thy.2012.0207)
- 107 Ji JH, Oh YL, Hong M, Yun JW, Lee HW, Kim D, Ji Y, Kim DH, Park WY, Shin HT *et al.* Identification of driving ALK fusion genes and genomic landscape of medullary thyroid cancer. *PLoS Genetics* 2015 **11** e1005467. (doi:10.1371/journal.pgen.1005467)
- 108 Shaw AT, Kim DW, Mehra R, Tan DS, Felip E, Chow LQ, Camidge DR, Vansteenkiste J, Sharma S, De Pas T *et al.* Ceritinib in ALK-rearranged non-small-cell lung cancer. *New England Journal of Medicine* 2014 **370** 1189–1197. (doi:10.1056/NEJMoa1311107)
- 109 Schlisio S, Kenchappa RS, Vredeveld LC, George RE, Stewart R, Greulich H, Shahriari K, Nguyen NV, Pigny P, Dahia PL *et al.* The kinesin KIF1B $\beta$  acts downstream from EglN3 to induce apoptosis and is a potential 1p36 tumor suppressor. *Genes and Development* 2008 **22** 884–893. (doi:10.1101/gad.1648608)
- 110 Welander J, Andreasson A, Juhlin CC, Wiseman RW, Backdahl M, Hoog A, Larsson C, Gimm O & Soderkvist P. Rare germline mutations identified by targeted next-generation sequencing of susceptibility genes in pheochromocytoma and paraganglioma. *Journal of Clinical Endocrinology and Metabolism* 2014 **99** E1352–E1360. (doi:10.1210/jc.2013-4375)
- 111 Gaal J, Burnichon N, Korpershoek E, Roncelin I, Bertherat J, Plouin PF, de Krijger RR, Gimenez-Roqueplo AP & Dinjens WN. Isocitrate dehydrogenase mutations are rare in pheochromocytomas and paragangliomas. *Journal of Clinical Endocrinology and Metabolism* 2010 **95** 1274–1278. (doi:10.1210/jc.2009-2170)
- 112 Wadt K, Choi J, Chung JY, Kiilgaard J, Heegaard S, Drzewiecki KT, Trent JM, Hewitt SM, Hayward NK, Gerdes AM *et al.* A cryptic BAP1 splice mutation in a family with uveal and cutaneous melanoma, and paraganglioma. *Pigment Cell & Melanoma Research* 2012 **25** 815–818. (doi:10.1111/pcmr.12006)
- 113 Ladroue C, Carcenac R, Leporrier M, Gad S, Le Hello C, Galateau-Salle F, Feunteun J, Poussegur J, Richard S & Gardie B. PHD2 mutation and congenital erythrocytosis with paraganglioma. *New England Journal of Medicine* 2008 **359** 2685–2692. (doi:10.1056/NEJMoa0806277)
- 114 Yang C, Zhuang Z, Fliedner SM, Shankavaram U, Sun MG, Bullova P, Zhu R, Elkahloun AG, Kourlas PJ, Merino M *et al.* Germ-line PHD1 and PHD2 mutations detected in patients with pheochromocytoma/paraganglioma-polycythemia. *Journal of Molecular Medicine* 2015 **93** 93–104. (doi:10.1007/s00109-014-1205-7)
- 115 Crona J, Delgado Verdugo A, Maharjan R, Stalberg P, Granberg D, Hellman P & Bjorklund P. Somatic mutations in H-RAS in sporadic pheochromocytoma and paraganglioma identified by exome sequencing. *Journal of Clinical Endocrinology and Metabolism* 2013 **98** E1266–E1271. (doi:10.1210/jc.2012-4257)
- 116 Oudijk L, de Krijger RR, Rapa I, Beuschlein F, de Cubas AA, Dei Tos AP, Dinjens WN, Korpershoek E, Mancikova V, Mannelli M *et al.* H-RAS mutations are restricted to sporadic pheochromocytomas lacking specific clinical or pathological features: data from a multi-institutional series. *Journal of Clinical Endocrinology and Metabolism* 2014 **99** E1376–E1380. (doi:10.1210/jc.2013-3879)
- 117 Fishbein L, Khare S, Wubbenhorst B, DeSloover D, D'Andrea K, Merrill S, Cho NW, Greenberg RA, Else T, Montone K *et al.* Whole-exome sequencing identifies somatic ATRX mutations in pheochromocytomas and paragangliomas. *Nature Communications* 2015 **6** 6140. (doi:10.1038/ncomms7140)
- 118 Papatthomas T, Oudijk L, Zwarthoff EC, Post E, Duijkers FA, van Noesel M, Hofland L, Pollard PJ, Maher ER, Restuccia DF *et al.* TERT promoter mutations in tumors originating from the adrenal gland and extra-adrenal paraganglia. *Endocrine-Related Cancer* 2014 **21** 653–661. (doi:10.1530/ERC-13-0429)
- 119 Liu T, Brown TC, Juhlin CC, Andreasson A, Wang N, Backdahl M, Healy JM, Prasad ML, Korah R, Carling T *et al.* The activating TERT promoter mutation C228T is recurrent in subsets of adrenal tumors. *Endocrine-Related Cancer* 2014 **21** 427–434. (doi:10.1530/ERC-14-0016)
- 120 Juhlin CC, Stenman A, Haglund F, Clark VE, Brown TC, Baranoski J, Bilguvar K, Goh G, Welander J, Svahn F *et al.* Whole-exome sequencing defines the mutational landscape of pheochromocytoma and identifies KMT2D as a recurrently mutated gene. *Genes, Chromosomes & Cancer* 2015 **54** 542–554. (doi:10.1002/gcc.22267)
- 121 Castro-Vega LJ, Letouze E, Burnichon N, Buffet A, Disderot PH, Khalifa E, Lorient C, Elarouci N, Morin A, Menara M *et al.* Multi-omics analysis defines core genomic alterations in pheochromocytomas and paragangliomas. *Nature Communications* 2015 **6** 6044. (doi:10.1038/ncomms7044)
- 122 Flynn A, Benn D, Clifton-Bligh R, Robinson B, Trainer AH, James P, Hogg A, Waldeck K, George J, Li J *et al.* The genomic landscape of pheochromocytoma. *Journal of Pathology* 2015 **236** 78–89. (doi:10.1002/path.4503)
- 123 Dahia PL, Ross KN, Wright ME, Hayashida CY, Santagata S, Barontini M, Kung AL, Sanso G, Powers JF, Tischler AS *et al.* A HIF1 $\alpha$  regulatory loop links hypoxia and mitochondrial signals in pheochromocytomas. *PLoS Genetics* 2005 **1** 72–80. (doi:10.1371/journal.pgen.0010008)
- 124 Burnichon N, Vescovo L, Amar L, Libe R, de Reynies A, Venisse A, Jouanno E, Laurendeau I, Parfait B, Bertherat J *et al.* Integrative genomic analysis reveals somatic mutations in pheochromocytoma and paraganglioma. *Human Molecular Genetics* 2011 **20** 3974–3985. (doi:10.1093/hmg/ddr324)
- 125 Qin Y, Deng Y, Ricketts CJ, Srikantan S, Wang E, Maher ER & Dahia PL. The tumor susceptibility gene TMEM127 is mutated in renal cell carcinomas and modulates endolysosomal function. *Human Molecular Genetics* 2014 **23** 2428–2439. (doi:10.1093/hmg/ddt638)
- 126 Travis WD, Müller-Hermelink HK, Harris CC, Brambilla E. Pathology and Genetics. In *WHO Classification of Tumours*. Lyon: IARC Press, 2004.
- 127 Sachithanandan N, Harle RA & Burgess JR. Bronchopulmonary carcinoid in multiple endocrine neoplasia type 1. *Cancer* 2005 **103** 509–515. (doi:10.1002/cncr.20825)
- 128 Fernandez-Cuesta L, Peifer M, Lu X, Sun R, Ozretic L, Seidel D, Zander T, Leenders F, George J, Muller C *et al.* Frequent mutations in chromatin-remodelling genes in pulmonary carcinoids. *Nature Communications* 2014 **5** 3518. (doi:10.1038/ncomms4518)
- 129 Debelenko LV, Brambilla E, Agarwal SK, Swallow JJ, Kester MB, Lubensky IA, Zhuang Z, Guru SC, Manickam P, Olufemi SE *et al.* Identification of MEN1 gene mutations in sporadic carcinoid tumors of the lung. *Human Molecular Genetics* 1997 **6** 2285–2290. (doi:10.1093/hmg/6.13.2285)
- 130 Rapa I, Votta A, Felice B, Righi L, Giorcelli J, Scarpa A, Speel EJ, Scagliotti GV, Papotti M & Volante M. Identification of microRNAs

- differentially expressed in lung carcinoid subtypes and progression. *Neuroendocrinology* 2015 **101** 246–255. (doi:10.1159/000381454)
- 131 Strobel P, Zettl A, Shilo K, Chuang WY, Nicholson AG, Matsuno Y, Gal A, Laeng RH, Engel P, Capella C *et al.* Tumor genetics and survival of thymic neuroendocrine neoplasms: a multi-institutional clinico-pathologic study. *Genes, Chromosomes & Cancer* 2014 **53** 738–749. (doi:10.1002/gcc.22183)
- 132 Fujii T, Kawai T, Saito K, Hishima T, Hayashi Y, Imura J, Hironaka M, Hosoya Y, Koike M & Fukayama M. MEN1 gene mutations in sporadic neuroendocrine tumors of foregut derivation. *Pathology International* 1999 **49** 968–973. (doi:10.1046/j.1440-1827.1999.00971.x)
- 133 Cunningham JL, Diaz de Stahl T, Sjoblom T, Westin G, Dumanski JP & Janson ET. Common pathogenetic mechanism involving human chromosome 18 in familial and sporadic ileal carcinoid tumors. *Genes, Chromosomes & Cancer* 2011 **50** 82–94. (doi:10.1002/gcc.20834)
- 134 Neklason D, VanDerslice J, Curtin K & Cannon-Albright LA. Evidence for a heritable contribution to neuroendocrine tumors of the small intestine. *Endocrine-Related Cancer* 2016 **23** 93–100. (doi:10.1530/ERC-15-0442)
- 135 Sei Y, Zhao X, Forbes J, Szymczak S, Li Q, Trivedi A, Voellinger M, Joy G, Feng J, Whatley M *et al.* A hereditary form of small intestinal carcinoid associated with a germline mutation in inositol polyphosphate multikinase. *Gastroenterology* 2015 **149** 67–78. (doi:10.1053/j.gastro.2015.04.008)
- 136 Francis JM, Kiezun A, Ramos AH, Serra S, Pedamallu CS, Qian ZR, Banck MS, Kanwar R, Kulkarni AA, Karpathakis A *et al.* Somatic mutation of CDKN1B in small intestine neuroendocrine tumors. *Nature Genetics* 2013 **45** 1483–1486. (doi:10.1038/ng.2821)
- 137 Banck MS, Kanwar R, Kulkarni AA, Boora GK, Metge F, Kipp BR, Zhang L, Thorland EC, Minn KT, Tentu R *et al.* The genomic landscape of small intestine neuroendocrine tumors. *Journal of Clinical Investigation* 2013 **123** 2502–2508. (doi:10.1172/JCI67963)
- 138 Crona J, Gustavsson T, Norlen O, Edfeldt K, Akerstrom T, Westin G, Hellman P, Bjorklund P & Stalberg P. Somatic mutations and genetic heterogeneity at the CDKN1B locus in small intestinal neuroendocrine tumors. *Annals of Surgical Oncology* 2015 **22** (Suppl 3) 1428–1435. (doi:10.1245/s10434-014-4351-9)
- 139 Kulke MH, Freed E, Chiang DY, Philips J, Zahrieh D, Glickman JN & Shivdasani RA. High-resolution analysis of genetic alterations in small bowel carcinoid tumors reveals areas of recurrent amplification and loss. *Genes, Chromosomes & Cancer* 2008 **47** 591–603. (doi:10.1002/gcc.20561)
- 140 Karpathakis A, Dibra H, Pipinikas C, Feber A, Morris T, Francis JM, Oukrif D, Mandair D, Pericleous M, Mohmaduvish M *et al.* Prognostic impact of novel molecular subtypes of small intestinal neuroendocrine tumour. *Clinical Cancer Research* 2016 **22** 250–258. (doi:10.1158/1078-0432.CCR-15-0373)
- 141 Verdugo AD, Crona J, Starker L, Stalberg P, Akerstrom G, Westin G, Hellman P & Bjorklund P. Global DNA methylation patterns through an array-based approach in small intestinal neuroendocrine tumors. *Endocrine-Related Cancer* 2014 **21** L5–L7. (doi:10.1530/ERC-13-0481)
- 142 Edfeldt K, Bjorklund P, Akerstrom G, Westin G, Hellman P & Stalberg P. Different gene expression profiles in metastasizing midgut carcinoid tumors. *Endocrine-Related Cancer* 2011 **18** 479–489. (doi:10.1530/ERC-10-0256)
- 143 Hessman O, Skogseid B, Westin G & Akerstrom G. Multiple allelic deletions and intratumoral genetic heterogeneity in men1 pancreatic tumors. *Journal of Clinical Endocrinology and Metabolism* 2001 **86** 1355–1361. (doi:10.1210/jcem.86.3.7332)
- 144 Katona TM, Jones TD, Wang M, Abdul-Karim FW, Cummings OW & Cheng L. Molecular evidence for independent origin of multifocal neuroendocrine tumors of the enteropancreatic axis. *Cancer Research* 2006 **66** 4936–4942. (doi:10.1158/0008-5472.CAN-05-4184)
- 145 Guo Z, Li Q, Wilander E & Ponten J. Clonality analysis of multifocal carcinoid tumours of the small intestine by X-chromosome inactivation analysis. *Journal of Pathology* 2000 **190** 76–79. (doi:10.1002/(SICI)1096-9896(200001)190:1 <76::AID-PATH499>3.0.CO;2-1)
- 146 Goebel SU, Vortmeyer AO, Zhuang Z, Serrano J, Jensen RT & Lubensky IA. Identical clonality of sporadic gastrinomas at multiple sites. *Cancer Research* 2000 **60** 60–63.
- 147 Baylin SB, Gann DS & Hsu SH. Clonal origin of inherited medullary thyroid carcinoma and pheochromocytoma. *Science* 1976 **193** 321–323. (doi:10.1126/science.935869)
- 148 Friedman E, Sakaguchi K, Bale AE, Falchetti A, Streeten E, Zimering MB, Weinstein LS, McBride WO, Nakamura Y, Brandi ML *et al.* Clonality of parathyroid tumors in familial multiple endocrine neoplasia type 1. *New England Journal of Medicine* 1989 **321** 213–218. (doi:10.1056/NEJM198907273210402)
- 149 Strosberg JR, Weber JM, Feldman M, Coppola D, Meredith K & Kvols LK. Prognostic validity of the American Joint Committee on Cancer staging classification for midgut neuroendocrine tumors. *Journal of Clinical Oncology* 2013 **31** 420–425. (doi:10.1200/JCO.2012.44.5924)
- 150 Korpershoek E, Stobbe CK, van Nederveen FH, de Krijger RR & Dinjens WN. Intra-tumoral molecular heterogeneity in benign and malignant pheochromocytomas and extra-adrenal sympathetic paragangliomas. *Endocrine-Related Cancer* 2010 **17** 653–662. (doi:10.1677/ERC-10-0072)
- 151 Korpershoek E, Petri BJ, Post E, van Eijck CH, Oldenburg RA, Belt EJ, de Herder WW, de Krijger RR & Dinjens WN. Adrenal medullary hyperplasia is a precursor lesion for pheochromocytoma in MEN2 syndrome. *Neoplasia* 2014 **16** 868–873. (doi:10.1016/j.neo.2014.09.002)
- 152 Crona J, Backman S, Maharjan R, Mayrhofer M, Stalberg P, Isakson A, Hellman P & Bjorklund P. Spatio-temporal heterogeneity characterizes the genetic landscape of pheochromocytoma and defines early events in tumorigenesis. *Clinical Cancer Research* 2015 **21** 4451–4460. (doi:10.1158/1078-0432.CCR-14-2854)
- 153 Paez JG, Janne PA, Lee JC, Tracy S, Greulich H, Gabriel S, Herman P, Kaye FJ, Lindeman N, Boggon TJ *et al.* EGFR mutations in lung cancer: correlation with clinical response to gefitinib therapy. *Science* 2004 **304** 1497–1500. (doi:10.1126/science.1099314)
- 154 Karapetis CS, Khambata-Ford S, Jonker DJ, O'Callaghan CJ, Tu D, Tebbutt NC, Simes RJ, Chalhah H, Shapiro JD, Robitaille S *et al.* K-ras mutations and benefit from cetuximab in advanced colorectal cancer. *New England Journal of Medicine* 2008 **359** 1757–1765. (doi:10.1056/NEJMoa0804385)
- 155 Wagle N, Grabiner BC, Van Allen EM, Hodis E, Jacobus S, Supko JG, Stewart M, Choueiri TK, Gandhi L, Cleary JM *et al.* Activating mTOR mutations in a patient with an extraordinary response on a phase I trial of everolimus and pazopanib. *Cancer Discovery* 2014 **4** 546–553. (doi:10.1158/2159-8290.CD-13-0353)
- 156 Wagle N, Grabiner BC, Van Allen EM, Amin-Mansour A, Taylor-Weiner A, Rosenberg M, Gray N, Barletta JA, Guo Y, Swanson SJ *et al.* Response and acquired resistance to everolimus in anaplastic thyroid cancer. *New England Journal of Medicine* 2014 **371** 1426–1433. (doi:10.1056/NEJMoa1403352)
- 157 Francois RA, Maeng K, Nawab A, Kaye FJ, Hochwald SN & Zajac-Kaye M. Targeting focal adhesion kinase and resistance to mTOR inhibition in pancreatic neuroendocrine tumors. *Journal of the National Cancer Institute* 2015 **107**. (doi:10.1093/jnci/djv123)
- 158 Jiang X, Cao Y, Li F, Su Y, Li Y, Peng Y, Cheng Y, Zhang C, Wang W & Ning G. Targeting  $\beta$ -catenin signaling for therapeutic intervention in MEN1-deficient pancreatic neuroendocrine tumours. *Nature Communications* 2014 **5** 5809. (doi:10.1038/ncomms6809)
- 159 Snyder A, Makarov V, Merghoub T, Yuan J, Zaretsky JM, Desrichard A, Walsh LA, Postow MA, Wong P, Ho TS *et al.* Genetic basis for clinical response to CTLA-4 blockade in melanoma. *New England Journal of Medicine* 2014 **371** 2189–2199. (doi:10.1056/NEJMoa1406498)

- 160 Brown SD, Warren RL, Gibb EA, Martin SD, Spinelli JJ, Nelson BH & Holt RA. Neo-antigens predicted by tumor genome meta-analysis correlate with increased patient survival. *Genome Research* 2014 **24** 743–750. (doi:10.1101/gr.165985.113)
- 161 Kreiter S, Vormehr M, van de Roemer N, Diken M, Lower M, Diekmann J, Boegel S, Schrors B, Vascotto F, Castle JC *et al.* Mutant MHC class II epitopes drive therapeutic immune responses to cancer. *Nature* 2015 **520** 692–696. (doi:10.1038/nature14426)
- 162 Flynn RL, Cox KE, Jeitany M, Wakimoto H, Bryll AR, Ganem NJ, Bersani F, Pineda JR, Suva ML, Benes CH *et al.* Alternative lengthening of telomeres renders cancer cells hypersensitive to ATR inhibitors. *Science* 2015 **347** 273–277. (doi:10.1126/science.1257216)
- 163 Vandamme T, Peeters M, Dogan F, Pauwels P, Van Assche E, Beyens M, Mortier G, Vandeweyer G, de Herder W, Van Camp G *et al.* Whole-exome characterization of pancreatic neuroendocrine tumor cell lines BON-1 and QGP-1. *Journal of Molecular Endocrinology* 2015 **54** 137–147. (doi:10.1530/JME-14-0304)
- 164 Vandamme T, Beyens M, Peeters M, Van Camp G & de Beeck KO. Next generation exome sequencing of pancreatic neuroendocrine tumor cell lines BON-1 and QGP-1 reveals different lineages. *Cancer Genetics* 2015 **208** 523. (doi:10.1016/j.cancergen.2015.07.003)
- 165 Crona J, Delgado Verdugo A, Granberg D, Welin S, Stålberg P, Hellman P & Björklund P. Next generation sequencing in genetic screening of pheochromocytoma and paraganglioma. *Endocrine Connections* 2013 **2** 104–111. (doi:10.1530/EC-13-0009)
- 166 Crona J, Ljungstrom V, Welin S, Walz MK, Hellman P & Björklund P. Bioinformatic challenges in clinical diagnostic application of targeted next generation sequencing: experience from pheochromocytoma. *PLoS ONE* 2015 **10** e0133210. (doi:10.1371/journal.pone.0133210)
- 167 Simbolo M, Mian C, Barollo S, Fassan M, Mafficini A, Neves D, Scardoni M, Pennelli G, Rugge M, Pelizzo MR *et al.* High-throughput mutation profiling improves diagnostic stratification of sporadic medullary thyroid carcinomas. *Virchows Archiv* 2014 **465** 73–78. (doi:10.1007/s00428-014-1589-3)
- 168 Khan MS, Kirkwood AA, Tsigani T, Lowe H, Goldstein R, Hartley JA, Caplin ME & Meyer T. Early changes in circulating tumor cells are associated with response and survival following treatment of metastatic neuroendocrine neoplasms. *Clinical Cancer Research* 2016 **22** 79–85. (doi:10.1158/1078-0432.CCR-15-1008)
- 169 Khan MS, Kirkwood A, Tsigani T, Garcia-Hernandez J, Hartley JA, Caplin ME & Meyer T. Circulating tumor cells as prognostic markers in neuroendocrine tumors. *Journal of Clinical Oncology* 2013 **31** 365–372. (doi:10.1200/JCO.2012.44.2905)

---

Received 30 September 2015

Revised version received 17 December 2015

Accepted 21 December 2015