FIG. 2. Deceased *Coeranoscincus reticulatus*.

reticulatus is currently listed as Vulnerable under Australian federal legislation. Known major threats for *C. reticulatus* include land clearing, inappropriate fire, and habitat disturbance due to cattle grazing and feral pigs.

In Australia, feral cats (*Felis catus*) are opportunistic mesopredators (Doherty et al. 2018. J. Biogeogr. 42:964–975) feeding on a wide range of birds, mammals, and reptiles ranging from those with several grams of body weight up to ca. 4 kg (Bonnaud et al. 2011. Biol. Invasions 13:581–603; Spencer et al. 2014. J. Mammal. 95:1278–1288; Woinarski et al. 2017. Biol. Conserv. 216:1–9). It has been speculated that cats might prey on *C. reticulatus*, however, to date no direct evidence has been found.

At 1000 h on 11 December 2018, a subadult male *F. catus* was found deceased (cause unknown) on Brindle Creek Road, within the world heritage-listed Border Ranges National Park, Australia (28.2251°S, 153.0639°E; WGS 84; 971 m elev.). Upon dissecting and examining the stomach contents (Fig. 1), five reptile species were identified, including *C. reticulatus* (Fig. 2), *Cacophis squamulosus* (Golden-crowned Snake), *Cryptophis nigrescens* (Small-eyed Snake), *Hemiaspis signata* (Marsh Snake), and *Cyclodomorphus gerrardii* (Pink-tongued Skink). Given that all of these species are either nocturnal or mostly active at night during summer months, it is likely they were prey items from the night prior to discovery. Our account represents the first record of a feral cat consuming *C. reticulatus*, and therefore highlights that predation by feral cats might pose a threat to *C. reticulatus* populations.

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***DIPORIPHORA AUSTRALIS* (Tommy Roundhead). ENDOPARASITES.** *Diphoriphora australis* is widespread along the north-eastern coast of Australia, extending north to the Cape York Peninsula, Queensland and south to northern New South Wales (Melville et al. 2019. Mem. Mus. Victoria 78:23–55). To our knowledge, the only helminth report for *D. australis* is the nematode *Strongyluris davisi* described by Harwood (1948. Proc. Linn. Soc. New South Wales 72:311–312). In this note we add to the helminth list of *D. australis*.

A sample of 15 *D. australis* (mean SVL = 61.9 mm ± 4.2 SD, range: 58–70 mm) collected in 1971 from the vicinity of Townsville, Queensland, Australia (19.2590°S, 148.8169°E; WGS 84) was examined from the University of Michigan Museum of Zoology (UMMZ): UMMZ 200869, 200896, 200897, 200900, 200902, 200905, 200907–200909, 200912–200917.

The specimens had been preserved in 10% formalin and stored in 70% ethanol. The body cavity was opened by a longitudinal incision and the digestive tract was removed and opened. The esophagus, stomach, small intestine and large intestine were examined for helminths under a dissecting microscope. Each helminth was placed in a drop of lactophenol on a glass slide, a cover slip was added, and identification was made after study under a compound microscope. We found two species of Nematoda: *Maxvachonia chabaudi* (small, large intestines), prevalence = 27%; mean intensity of infection (mean number of helminths) ± SD = 2.3 ± 0.96, 1–3; *Kreisiella chrysocampa* (small intestine), 1 individual. Identifications were made by study of the original descriptions: Mawson (1972. Trans. R. Soc. S. Aust. 96:101–113) and Jones (1985. J. Nat. Hist. 19:1231–1237). *Maxvachonia chabaudi* and *K. chrysocampa* in *D. australis* are new host records. Voucher helminths were deposited in the Harold W. Manter Parasitology Laboratory (HWML), The University of Nebraska, Lincoln, USA as *Maxvachonia chabaudi* (HWML 111446) and *Kreisiella chrysocampa* (HWML 111445).

Maxvachonia chabaudi was described from *Morethia lineoocellata* by Mawson (1972, *op. cit.*), and other hosts have been previously described from Australia (Mawson 1972, *op. cit.*; Jones 2003. Pap. Proc. R. Soc. Tasmania. 137:7–12; Goldberg and Bursey 2012. Comp. Parasitol. 79:247–268), Malaysia (Goldberg et al. 2018. Sauria 40:92–94), Oceania (Goldberg et al. 2000. Comp. Parasitol. 67:118–121; Goldberg et al. 2005. Pac. Sci. 59:609–614; Bursey and Goldberg 2001. J. Parasit. 87:135–138; Goldberg and Bursey 2002. J. Nat. Hist. 361:2249–2264), and Papua New Guinea (Goldberg et al. 2010. Pac. Sci. 64:131–139).

Kreisiella chrysocampa was described from *Egernia inornata* (from Australia) by Jones (1985, *op. cit.*). Other hosts have been described from Australia (Jones 1995. Aust. J. Zool. 43:141–164, Goldberg and Bursey 2000. Trans. Roy. Soc. S. Aust. 124:127–133) and Papua New Guinea (Jones 1985, *op. cit.*; Goldberg et al. 2008. J. Nat. Hist. 42:1923–1935).

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***ECPLEOPUS GAUDICHAUDII*. REPRODUCTION.** *Ecpleopus gaudichaudii* is a small gymnophthalmid lizard (up to 50 mm SVL) endemic to the Atlantic Forest and found mainly in south-eastern and southern Brazil (Dias and Rocha 2013. Check List 9:607–609). Their reproductive biology is poorly known, but females seemingly produce a single egg per clutch from late winter (September) to early summer (January; Uzzell 1969. Postilla 135:1–23; Perini and Butti 2008. Herpetol. Rev. 39:222). Here, we report on the first record of egg-laying and relative clutch mass (RCM) of the species and provide evidence that female reproductive season is longer than previously shown.

On 22 March 2012, a gravid female *E. gaudichaudii* (IBSP. CRIB 413: 35 mm SVL, 60 mm tail length, 0.577 g) was collected at Parque Estadual Restinga de Bertioga, São Paulo, Brazil (23.77826°S, 46.07745°W; WGS 84; 11 m elev.). The female was

brought to our laboratory, placed in a plastic container with leaf litter, and kept at room temperature (22–26°C). A couple of days later, the female laid a single egg (7.51 × 3.94 mm, 0.078 g). The egg was incubated in a plastic container with moistened vermiculite and kept at room temperature, but it spoiled due to fungal contamination. RCM (total clutch mass/maternal body mass after oviposition + total clutch mass; sensu Vitt and Price 1982. Herpetologica 38:237–255) was 0.119, which is similar to that of many gymnophthalmids (Mesquita et al. 2016. Am. Nat. 187:689–705) and active forager lizards (Vitt and Price 1982, *op. cit.*). Our record of egg-laying in late March indicates that the reproductive season of female *E. gaudichaudii* extends throughout rainy season.

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***EGERNIA KINGII* (King's Skink). TAIL BIFURCATION.** Caudal autotomy and subsequent regeneration is an effective anti-predation strategy used by members of many lizard families (Arnold 1984. J. Nat. Hist. 18:127–169; Bateman and Fleming 2009. J. Zool. 277:1–14). However, in certain cases, either from an incomplete autotomy, or from a significant caudal wound, additional tails can be regenerated, producing an individual with a multifurcated tail (Bellairs and Bryant 1985. *In* Gans and Billett [eds.], Biology of the Reptilia, Volume 15: Development B, pp. 301–410. John Wiley & Sons, New York, New York; Barr et al. 2019. Herpetol. Rev. 50:567). Here, we report on three observations of *Egernia kingii* with regenerative bifurcations, two from the preserved collection of the Western Australian Museum, and one from a field population on Penguin Island, Western Australia (32.30584°S, 115.69134°E; WGS 84).

Egernia kingii is a large (up to 0.5 m) skink endemic to Western Australia and its surrounding Islands (Storr 1978. Rec. West. Aust. Mus. 6:147–187; Cogger 2014. Reptiles and Amphibians of Australia. CSIRO Publishing, Victoria, Australia. 544 pp.). Data was obtained from preserved specimens of the Western Australia Museum from May 2017–August 2018 and for field data from November 2017–February 2018 and November 2018–February 2019. Standard morphometrics including snout–vent length (SVL), tail length (TL) and regeneration length (RL) were measured using a plastic ruler to the nearest mm. Field individuals were sexed using hemipene probing as per Brown (2012. A Guide to Australian Skinks in Captivity. Reptile Pub. Burleigh, Queensland, Australia. 360 pp.).

The entire collection of preserved specimens of *E. kingii* from the Western Australian Museum (N = 300) was assessed as a part of another study (Barr et al. 2018. Biol. J. Linn. Soc. 126:268–275), and 105 *E. kingii* individuals were assessed over the two field seasons from Penguin Island. Tails of 60 individuals from the museum collection were not attached to the specimen, most likely lost posthumously, and could not be assessed. The first individual with caudal bifurcation from the Western Australian Museum (R36041), a large adult (unknown sex) measured 240 mm SVL and 215 mm TL and possessed a large, mature bifurcation 65 mm from its cloacae, with the primary (longer) tail portion measuring 150 mm, and secondary tail measuring 117 mm (Fig. 1). The second individual with caudal bifurcation from the Western Australian Museum (R9348), a juvenile (unknown sex), measured 136 mm SVL and 178 mm TL and possessed a small bifurcation 150 mm from its cloaca, with the bifurcation length measuring 20 mm. The specimen from Penguin Island, an adult female, measured 194 mm SVL and 172 mm TL, had a small distal bifurcation measuring 13 mm. The incidence of bifurcation in the museum specimens overall was 0.83% (2/240) and 0.95% (1/105) for Penguin Island, with incidence of bifurcation for those showing evidence of regeneration as 1.54% (2/130) for museum specimens, and 1.22% (1/82) for the Penguin Island population. Observation of regenerative bifurcations in populations or museum collections, when reported, is generally low (Cordes and Walker 2013. Herpetol. Rev. 44:319; Vrcibradic and Niemeyer 2013. Herpetol. Rev. 44:510–511; Sorlin et al. 2019. Herpetol. Rev. 50:377–378), with some ecological evidence that this may be caused from inefficient predation or wounding (e.g., *Cyclura* spp. attacked by invasive rodents; Hayes et al. 2012. Biodivers. Conserv. 21:1893–1899).

FIG. 1. Two preserved *Egernia kingii* specimens (top: R36041; bottom: R9348) from the Western Australia Museum showing caudal bifurcation.

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***GYMNODACTYLUS GECKOIDES*. BEHAVIOR.** *Gymnodactylus geckoides* (Phyllodactylidae) is endemic to the Caatinga and distributed in the following states in Brazil: Tocantins, Mato Grosso, Goiás, Piauí, Ceará, Rio Grande do Norte, Paraíba, Pernambuco, Alagoas and Bahia (Vanzolini 2004. An. Acad. Bras. Ciênc. 76:663–698; Costa and Bérnils 2018. Herpetol. Bras. 7:11–57). Lizards may exhibit various pre-copulatory reproductive behaviors, ranging from cloacal friction on theioids, nodding on tropidurids, extending barbed on polyrotrids to darkening of the skin on leiosaurids (Carpenter 1962. Am. Midl. Nat. 67:132–151; Carpenter 1977. Herpetologica 33:285–289; Jenssen 1977. Am. Zool. 17:203–215; Lima and Sousa 2006. Rev. Bras. Zooc. 8:193–197). The most recurrent post-copulatory behavior is polygyny (Anderson and Vitt 1990. Oecologia 84:145–157; Olsson 1993. Behav. Ecol. Sociobiol. 32:337–341; Censky 1995. Behav. Ecol. Sociobiol.