



Coordination of physiological and structural traits in Amazon forest trees

S. Patiño^{1,2,†}, N. M. Fyllas², T. R. Baker², R. Paiva³, C. A. Quesada^{2,4}, A. J. B. Santos^{3,4,†}, M. Schwarz¹, H. ter Steege⁵, O. L. Phillips², and J. Lloyd^{2,6}

¹Max-Planck-Institut für Biogeochemie, Postfach 100164, 07701, Jena, Germany

²School of Geography, University of Leeds, LS2 9JT UK

³Instituto Nacional de Pesquisas da Amazônia, Manaus, AM, Brazil

⁴Departamento de Ecologia, Universidade de Brasília, DF, Brazil

⁵Dept. of Plant Ecology and Biodiversity, Utrecht University, The Netherlands

⁶School of Earth and Environmental Sciences, James Cook University, Cairns, Qld 4871, Australia

[†]deceased

Correspondence to: J. Lloyd (j.lloyd@leeds.ac.uk)

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Abstract. Many plant traits covary in a non-random manner reflecting interdependencies associated with “ecological strategy” dimensions. To understand how plants integrate their structural and physiological investments, data on leaf and leaflet size and the ratio of leaf area to sapwood area (Φ_{LS}) obtained for 1020 individual trees (encompassing 661 species) located in 52 tropical forest plots across the Amazon Basin were incorporated into an analysis utilising existing data on species maximum height (H_{max}), seed size, leaf mass per unit area (M_A), foliar nutrients and $\delta^{13}C$, and branch xylem density (ρ_x).

Utilising a common principal components approach allowing eigenvalues to vary between two soil fertility dependent species groups, five taxonomically controlled trait dimensions were identified. The first involves primarily cations, foliar carbon and M_A and is associated with differences in foliar construction costs. The second relates to some components of the classic “leaf economic spectrum”, but with increased individual leaf areas and a higher Φ_{LS} newly identified components for tropical tree species. The third relates primarily to increasing H_{max} and hence variations in light acquisition strategy involving greater M_A , reductions in Φ_{LS} and less negative $\delta^{13}C$. Although these first three dimensions were more important for species from high fertility sites the final two dimensions were more important for low fertility species and were associated with variations linked to reproductive and shade tolerance strategies.

Environmental conditions influenced structural traits with ρ_x of individual species decreasing with increased soil fertility and higher temperatures. This soil fertility response

appears to be synchronised with increases in foliar nutrient concentrations and reductions in foliar [C]. Leaf and leaflet area and Φ_{LS} were less responsive to the environment than ρ_x .

Thus, although genetically determined foliar traits such as those associated with leaf construction costs coordinate independently of structural characteristics such as maximum height, others such as the classical “leaf economic spectrum” covary with structural traits such as leaf size and Φ_{LS} . Coordinated structural and physiological adaptations are also associated with light acquisition/shade tolerance strategies with several traits such as M_A and [C] being significant components of more than one ecological strategy dimension. This is argued to be a consequence of a range of different potential underlying causes for any observed variation in such “ambiguous” traits. Environmental effects on structural and physiological characteristics are also coordinated but in a different way to the gamut of linkages associated with genotypic differences.

1 Introduction

Plant traits are widely used in ecology and biogeochemistry. In particular, sets of functional characters can serve as the basis for identifying important adaptations that improve the success of different taxa at different environments. Over the last decade significant advances have been made in terms of our understanding of plant trait inter-relationships and associated trade-offs (Reich et al., 1997; Westoby et al., 2002),

especially in terms of the so called “leaf economic spectrum” (Wright et al., 2004) with well documented systematic and co-ordinated changes in leaf nitrogen and phosphorus concentrations, leaf mass per unit area, M_A and leaf lifetimes.

Attention has also been paid to the relationships between physiological and structural characteristics of leaves and other plant traits. For example, it has been reported that leaf size declines with wood density, ρ_w (Pickup et al., 2005; Wright et al., 2006, 2007; Malhado et al., 2009) and it has been suggested that this is because the ratio of leaf area to sapwood area (Φ_{LS}) should also decline with increasing wood density due to hydraulic constraints (Wright et al., 2007). Nevertheless, although Φ_{LS} may decline with ρ_w for trees in some ecosystems that are clearly water-limited (Ackerly, 2004; Cavender-Bares et al., 2004), Φ_{LS} sometimes actually increases with ρ_w (Wright et al., 2006; Meinzer et al., 2008). The latter study also found that associated with these higher Φ_{LS} and high wood density stems were lower stem hydraulic conductances, more negative mid-day leaf water potentials, and more negative bulk leaf osmotic potentials at zero turgor. Thus, leaves of some high wood density species may be characterised by physiological and structural adaptations allowing them to function at more severe water deficits than is the case for low wood density species.

The Panama study of Meinzer et al. (2008) also found that higher ρ_w species tended to have higher M_A . Although similar positive correlations between M_A and ρ_w have also been reported for other ecosystems (e.g. for sclerophyllous forest: Ishida et al., 2008) when examining the bivariate relationship between ρ_w and M_A across a range of tropical forest sites, Wright et al. (2007) observed no significant relationship. Likewise, when examining variation in leaf and stem traits for 17 dipterocarp species growing in a common garden in southern China, Zhang and Cao (2009) also found no significant correlation between ρ_w and M_A .

Variations in M_A may also be related to a suite of additional plant physiological characteristics (Poorter et al., 2009), varying negatively with dry-weight foliar nitrogen and phosphorus concentrations (Wright et al., 2004; Fyllas et al., 2009) as well as tending to increase with increasing tree height (Thomas and Bazzaz, 1999; Kenzo et al., 2006; Lloyd et al., 2010). Potential tree height, H_{max} , has also been related to a number of wood traits (Chave et al., 2009) with taller plants tending to have bigger conduits in their trunks, but fewer conduits overall (Coomes et al., 2007).

Within a given stand, taller and generally more light-demanding rain forest species also tend to have larger leaves, this being associated with shallower crown and a more efficient light capture (Poorter et al., 2006; Poorter and Rozendaal, 2008). Leaf-size may also be influenced by other factors. For example, Australian rain forests growing on oligotrophic soils typically have a greater abundance of smaller leaved species than for nearby forests found on more mesotrophic soil types (Webb, 1968).

Seed size may also relate to the above plant functional traits. For example, one of “Corner’s rules” describes a tendency for species with thick twigs to have large appendages (leaves and fruit). The range of viable seed size also tends to increase with plant height (Moles et al., 2005; Grubb et al., 2005). Forests on the more fertile soils of western Amazonia tend to have smaller average seed masses than their less fertile counterparts on the Guyana Shield and elsewhere (ter Steege et al., 2006), this perhaps being related to several advantages attributable to large seeded species under nutrient-poor conditions, *viz.* greater initial nutrient stores, greater initial root zone expansion, and increased mycorrhizal infection, all of which would be expected to increase the probability of seedling survival (Foster, 1986).

This paper presents new data on leaf and leaflet size and Φ_{LS} for 661 species located in 52 plots across the Amazon Basin. The trees sampled form a subset of those also examined for variations in branch xylem density (Patiño et al., 2009), and for foliar nutrients, M_A and $\delta^{13}C$ (Fyllas et al., 2009), which had previously been analysed separately. We here investigate the inter-relationships between these structural and physiological parameters also considering taxonomic variations in H_{max} (Baker et al., 2009) and seed mass (ter Steege and Hammond, 2001; ter Steege et al., 2006). Specifically, we were interested to assess the degree to which the observed variations in the studied structural and physiological traits were coordinated with each other into identifiable integrated trait dimensions: for example, those associated with leaf construction costs, light acquisition, and/or shade tolerance.

2 Materials and methods

2.1 Study sites

In the analysis here, RAINFOR sample plots have been aggregated as discussed in Fyllas et al. (2009), with further plot details available in Patiño et al. (2009) and Quesada et al. (2010). Ten plots in Fyllas et al. (2009) have not been included due to insufficient structural trait data having been collected, but the range of soils encountered here is still substantial with the sum of exchangeable bases (0–0.3 m), for example ranging from less than 1 mmol_c kg⁻¹ to nearly 100 mmol_c kg⁻¹. Total soil phosphorus ranged from 26 mg kg⁻¹ for an ortseinc podzol to 727 mg kg⁻¹ for a eutric cambisol (Quesada et al., 2010). Mean annual precipitation varies from less than 1.5 m a⁻¹ on sites at the north and southern periphery of the basin to more than 3.0 m a⁻¹ for sub-montane sites close to the Andes.

2.2 Structural traits

For most trees sampled in Patiño et al. (2009) and Fyllas et al. (2009), and from the same terminal branches for which data has already been presented in those studies, all leaves from the branch had also been counted. From that branch, a sub-sample of 10–20 leaves was randomly chosen to estimate individual leaf area, L_A , and leaflet area, ℓ_A (when a species had compound leaves), and to estimate the total leaf area of the branch. All age and size leaves or leaflets were selected for this analysis except for very young leaves or those which were obviously senescent. The chosen leaves were usually scanned fresh on the same day of collection. When this was not possible the same day, they were stored for a maximum of two days in sealed plastic bags to avoid desiccation and any consequent reduction of the leaf area. Scans were analysed using “Win Folia Basic 2001a” (Regent Instruments Inc., 4040 rue Blain Quebec, QC., G2B 5C3 Canada) to obtain L_A and ℓ_A .

The distal (sapwood+pith) and pith diameters for each branch were also measured with a digital caliper (Mitutoyo Corporation, Japan) with sapwood area, A_S , then estimated by subtracting pith area from the total branch area with $\Phi_{LS} = n\bar{L}_A/A_S$ where n is the number of leaves distal to the piece of branch sampled and \bar{L}_A is the average area of the individual leaves sub-sampled for the estimation of L_A and/or ℓ_A .

Branch xylem density data for the same samples were obtained as described in Patiño et al. (2009). In brief, this consisted of the estimation of the volume of a branch segment, approximately 1 cm in diameter and 5–10 cm long using calipers, with the pith removed as necessary and dry weight subsequently determined. Species maximum height taken from the database developed by Baker et al. (2009) with estimates made to the species level for 80% of the trees identified, and the bulk of the remainder being genus level averages. Seed mass (S) was taken as a genus level dependent variable and was already on a \log_{10} ordinal scale (ter Steege et al., 2006).

2.3 Physiological foliar traits

Foliar traits used here are as described/measured in Fyllas et al. (2009) and Lloyd et al. (2010) and include leaf mass per unit area (M_A) and foliar [N], [C], [P], [Ca], [K] and [Mg] expressed on dry-weight basis. Foliar $^{13}\text{C}/^{12}\text{C}$ discrimination, Δ , was estimated from measurements of foliar $\delta^{13}\text{C}$ (Fyllas et al., 2009) using an assumed value for the isotopic composition of source air equal to -8.0‰ (Farquhar et al., 1989) and subsequently transformed to a diffusional limitation index, ϕ , according to (Fyllas et al., 2012)

$$\phi = 1 - \sqrt{\frac{(\Delta - 4.4)/25.6 - 0.2}{0.8}} \quad (1)$$

which utilises the well known relationship between Δ and the ratio of internal to ambient CO_2 concentrations, c_i/c_a (Farquhar et al., 1989). Equation (1) assumes that at current day c_a , photosynthesis can be considered a roughly linear function of c_i and with a maximum practical c_i/c_a (indicating minimal diffusional limitation) of 0.8. Here we have taken a value of 4.4‰ for the fractionation against $^{13}\text{CO}_2$ during diffusion into the leaf and 30.0‰ for the fractionation against $^{13}\text{CO}_2$ during photosynthetic fixation (Farquhar et al., 1989). Increasing ϕ values are associated with lower c_i/c_a , and thus, other things being equal, a higher water use efficiency, W , this being the ratio of carbon gained to water lost during photosynthetic CO_2 assimilation. Equation (1) relies on a simplified expression for Δ which ignores difference between gas- and liquid-phase fractionations within the leaf (Farquhar et al., 1989), but this should not seriously compromise its utility in the current context.

2.4 Climate and soils

The soil and climate predictors table used was the same as in Fyllas et al. (2009), using a set of measured soil properties (Quesada et al., 2010) with precipitation variables and temperature from the “WorldClim” dataset (<http://www.worldclim.org>). Estimates of mean annual solar radiation are from New et al. (2002). As in Fyllas et al. (2009) we separate soils into two fertility classes based on their total phosphorus concentration and the total sum of reserve bases, (Quesada et al., 2010). In brief this categorisation gives rise to arenosols, podzols, ferralsols, and most acrisols being classified as low fertility soils. High fertility soils include plinthosols, cambisols, fluvisols, gleysols and most alisols.

2.5 Statistical analysis

This paper implements a similar set of statistical analyses to that described in detail in Fyllas et al. (2009). Preliminary tests included analysis of normality (Shapiro-Wilk) and homogeneity of variance (Fligner-Killeen) for each of the structural traits of interest. The foliar related structural traits (L_A , ℓ_A and Φ_{LS}) presented a right skewed distribution and thus were all \log_{10} transformed. As ρ_x , H_{\max} and S (the latter already provided as size classes on a \log_{10} scale) were more or less symmetrically distributed around their mean we did not apply this transformation for these variables, even though the Shapiro test failed to identify strict normality. The non-parametric Kruskal-Wallis test (Hollander and Wolfe, 1999) was used to explore for differences between fertility groups as well as for differences between families, genera within a family and species within a genus. All analyses were performed with the R statistical platform (R Development Core Team, 2010).

2.5.1 Partitioning of variance and estimation of taxonomic and environmental effects

A multilevel model was initially fitted for all traits (including those previously analysed separately in Fyllas et al. (2009) and Patiño et al. (2009) because this was a slightly different dataset), except H_{\max} and S according to

$$\Theta = \mu + p + f/g/s + \epsilon, \quad (2)$$

where μ is the overall mean value of each trait, Θ ; p is the plot effect, i.e. the effect of the location that each individual is found, and $f/g/s$ represents the genetic structure of the data, i.e. that each individual belongs to a species (s), nested in a genus (g), nested in a family (f), and ϵ is the error term. All parameters were estimated by the Residual Maximum Likelihood (REML) method with the `lme4` library available within R (Bates and Sarkor, 2007). Fyllas et al. (2009) have already discussed further details of the above formulations and the advantage in being able to partition the variance from the family to the species level, also taking into account the location (thus the environmental contribution to trait variation) where the trait was measured. The Supplementary Information (II) of that paper also provides an empirical validation of the approach used. Note, that whilst theoretically possible, we do not include interaction terms in Eq. (2), this is because there is insufficient species replication across different sites. Nevertheless, investigations into the likely magnitude of such effects have been undertaken as part of the analyses in both Patiño et al. (2009) and Fyllas et al. (2009) and have not been found to be significant. Again we were interested in exploring the taxonomic (estimated as the sum of family \pm genus \pm species random effects) and environmental terms, using bivariate relationships as well as multiple non-parametric regressions of plot effect contributions on a set of environmental predictors. For the latter we used Kendall's τ as our measure of association calculating the significance of partial correlations using our own specifically written code, using the R statistical platform.

For H_{\max} and S no multilevel model was fitted or environmental effect assumed, the available data being considered to express directly the genetic potential of each species. We also note that our estimates of S are resolved at the genus level only (ter Steege and Hammond, 2001) and are only on a \log_{10} categorical scale. This introduces potential errors into the analyses where S is involved because all other traits have been resolved at the species level. Thus, even though a small portion of the observed variation in S generally occurs at the species level (Casper et al., 1992), bivariate and multivariate analyses involving this trait as presented here may carry somewhat more “noise” than would otherwise be the case.

2.5.2 Bivariate relationships

Relationships were initially assessed with the Pearson's correlation coefficient (r) with subsequent Standardized Ma-

yor Axis (SMA) line fits where significant correlations were identified. In this study, SMA line fits are applied to the raw dataset (including all measured traits and thus intraspecific variation), to the taxonomic component of trait variation (i.e. each species is represented by a single data point) as well as to the plot level effects (i.e. each plot is contributing a single data point). In each case we initially fitted separate lines for each fertility group, and when a common SMA slope was identified we tested for differences in elevation and/or slope between fertility groups, using the `smatr` library available within R (Warton et al., 2006).

We explored the plot level effect of each structural trait, through non-parametric correlation analysis on selected soil and environmental predictors, with the soil variables reduced to three principal axes to avoid multicollinearity (Fyllas et al., 2009). The climatic variables of mean annual temperature, total annual precipitation, dry season precipitation and mean annual radiation were also examined. As extensively discussed in Fyllas et al. (2009) we dealt with spatial autocorrelation issues by fitting appropriate simultaneous autoregressive models (SAR) which include a spatial error term (Lichstein et al., 2002) to help interpret the significance of full and partial Kendall's τ coefficients as a measure of association between plot-level trait effects and environmental predictors.

2.5.3 Multivariate analyses

Inferred taxonomic effects were analysed jointly for species found on fertile versus infertile soils (excluding those found on both soil types) by calculating separate variance-covariance matrices for the two species groups and then using the common principal components (CPC) model of Flury (1988) as implemented by Phillips and Arnold (1999). Within this model, it is assumed that the two populations of species have the same eigenvectors (principal components; denoted here as U) but that the relative loading of the various U as expressed through their eigenvalues (λ) may potentially vary between the two populations. Flury's model provides a hierarchy of tests corresponding to a range of possible relationships between matrices including equality, proportionality, common principal components, partial common principal components or unrelated (Flury, 1988; Phillips and Arnold, 1999). CPC can thus be seen as a method for summarizing the variation in two or more matrices. Nevertheless, caution needs to be applied when using CPC to address the more complex goal of diagnosing and understanding the nature of the changes that underlie the difference between the matrices. This is because CPC tends to spread any differences over many of the vectors it extracts and often over all of them (Houle et al., 2002).

As the CPC model does not strictly apply to correlation matrices (Flury, 1988), we standardised each variable before calculating the input variance-covariance matrix by dividing each variable by its observed range (across both high and

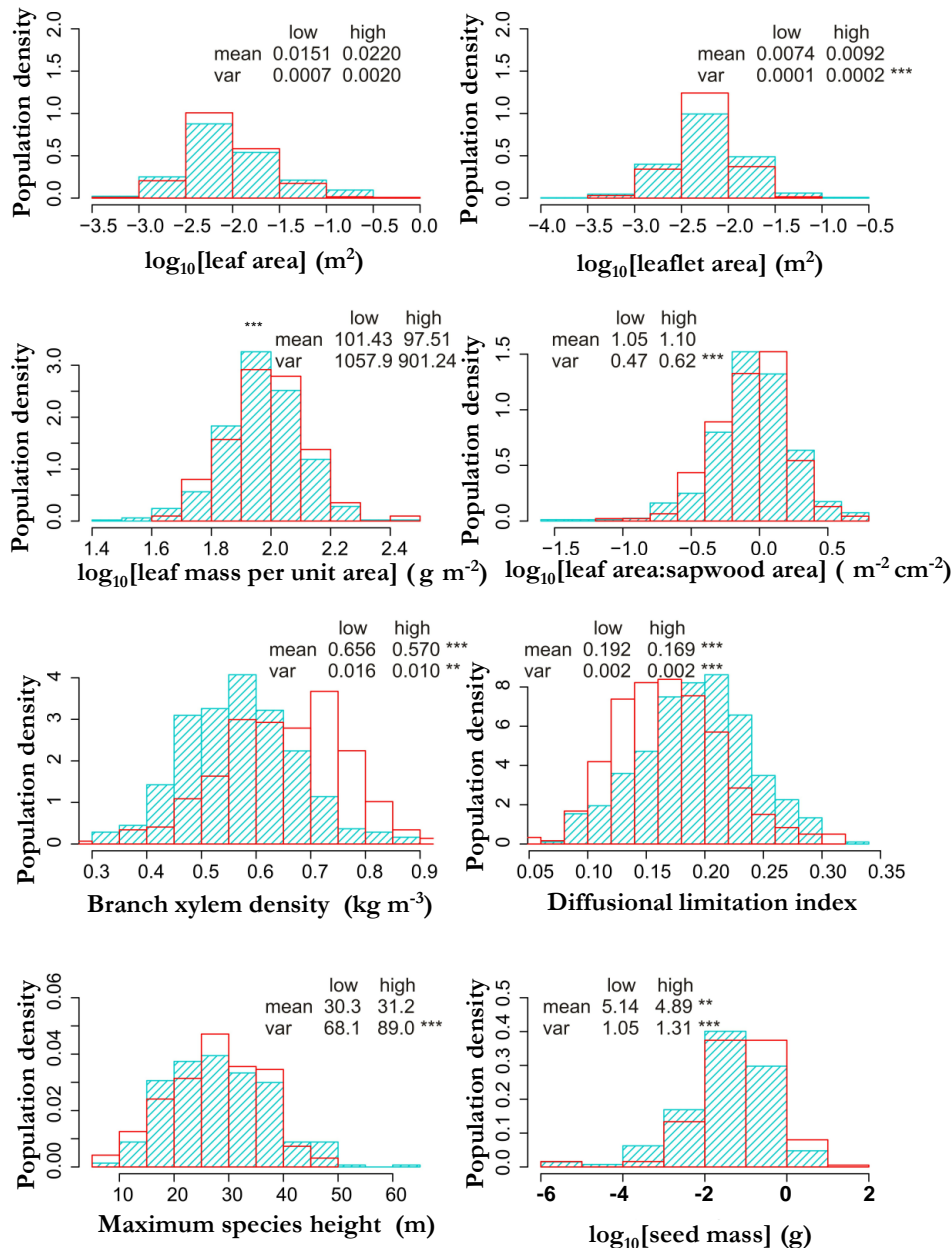


Fig. 1. Probability density histograms of raw data per fertility group for leaf area (L_A ; m²), leaflet area (ℓ_A ; m²), leaf mass per unit area (M_A ; g m⁻²), (m²), leaf area:sapwood area ratio (Φ_{LS} ; cm² m⁻²), branch xylem density (ρ_x ; kg m⁻³), ϕ = stomatal limitation index (dimensionless; see Eq. 1), species maximum height (H_{max} ; m) and seed mass (S ; g). Open red bars represent low and blue dashed bars high soil fertility plots, as defined by the quantitative determinations of the level of total reserve bases from 0.0–0.3 m depth (Fyllas et al., 2009; Quesada et al., 2010). Also given for each histogram are the mean and the variance for each trait. Significant differences in mean values and/or variances between the two fertility groups were identified with the Fligner-Killeen test respectively. Significance codes: *** < 0.001, ** < 0.01, * < 0.05.

low fertility soils) as first proposed by Gower (1966) but, due to the presence of the occasional outlier, taking the effective range as the 0.1 to 0.9 quantiles. Standard errors of the U and λ for the CPC models were estimated assuming asymptotic normality as described in Flury (1988).

All other multivariate analyses (e.g. PCA of the derived environmental effects) were implemented with the `ade4` package (Thioulouse et al., 1997) available within the R statistical platform with the environmental effect PCA undertaken on the correlation matrix.

3 Results

3.1 Trait distribution in relation to soil type

The structural traits distributions along with those for M_A and \odot for the complete dataset divided to low and high fertility groups are shown in Fig. 1 with overall mean values, range and variances for each plot for all traits also provided in the Supplementary Information (Table S1). The three leaf related traits introduced here (L_A , ℓ_A and Φ_{LS}) did not differ significantly between low and high fertility sites (Fig. 1). On the other hand, ρ_x and S showed significant differences between the two fertility groups, with their distributions shifted to the left for fertile sites, i.e. higher ρ_x and S were found for species found on infertile soils. This is similar to the shifted distributions identified for most leaf mineral concentrations across fertility gradients (Fyllas et al., 2009) but in the opposite direction, i.e. with higher structural carbon and lower mineral investment in less fertile environments. As expected from our prior analysis of the statistical distribution of foliar $\delta^{13}C$ (Fyllas et al., 2009), the diffusional limitation index of Eq. 1 \odot tended to be lower for trees growing on low fertility soils. Despite a difference in variance between low and high fertility sites, there was, however, no overall effect of soil fertility classification on the average H_{max} .

3.2 Partitioning of the variance

The variation apportioned to different taxonomic levels varies for each of the traits examined (Fig. 2). When leaf size was expressed per leaflet, most of the variation was attributed at the species level (0.31) with the overall taxonomic component (i.e. family \pm genus \pm species) adding up to a very high (0.62) proportion. When leaf size was expressed at the leaf level, most of the variation was attributed at the family level (0.29) with a very high overall taxonomic component (0.71). In contrast to L_A and ℓ_A , plot level contributions to the total variance were substantial for the other structural traits: being around 0.30 for ρ_x and 0.27 for Φ_{LS} . These are not necessarily higher than their respective taxonomic components, but underline the importance of the site growing conditions in influencing structural traits such as ρ_x and Φ_{LS} . As for the foliar traits reported in Fyllas et al. (2009) this must have direct implications for different physiological processes. In that study, leaf mass per unit area and [C], [N] and [Mg] emerged as highly constrained by the taxonomic affiliation, but with others, such as [P], [K] and [Ca] also strongly influenced by site growing conditions. That study also found foliar $\delta^{13}C$ to be strongly influenced by site growing conditions, consistent with its analogue here (\odot) having its environmental component as the dominant source for its variation. Overall, there was a tendency for the residual component (related to intra-species variations not accountable for by different plot locations and experimental error) to increase as the proportion of variation accountable for by taxonomic affiliation declined

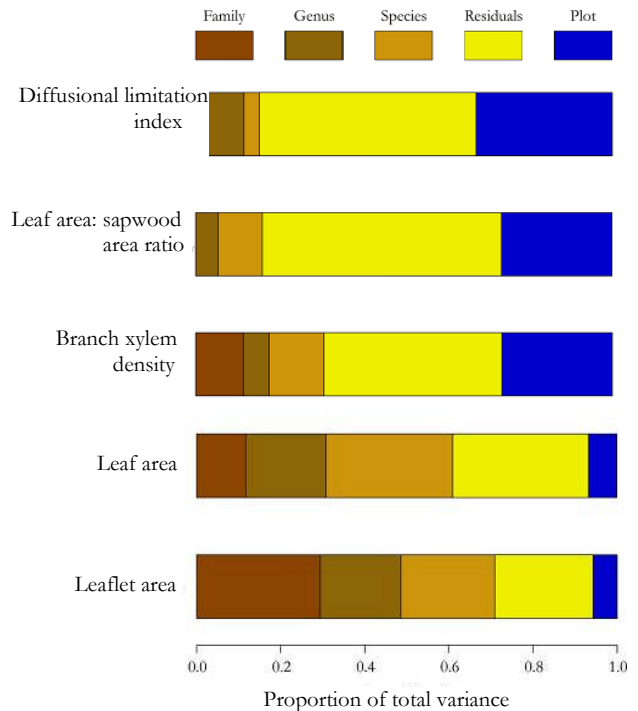


Fig. 2. Partitioning of the total variance for each studied property into taxonomic (family/genus/species), environmental (plot) and error (residual) components. Traits are sorted from less to more taxonomically constrained. Significance of each variance component was tested with a likelihood ratio test (Galwey, 2006). Significance codes: *** < 0.001, ** < 0.01, * < 0.05.

and with the proportion attributable to plot location tending to increase as the residual component became larger.

3.3 Bivariate relationships: raw data

These are not considered in any detail here, but for the interested reader data are summarised in the Supplementary Information, Table S2A.

3.4 Bivariate relationships: taxonomic components

Considering data from both low and high fertility sites together, Table 1 lists correlations and SMA slopes for the derived taxonomic components with this same information shown in more detail (including confidence intervals) in the Supplementary Information (Table S2A) and with low and high fertility species separated for OLS and SMA regression analyses in Table S2B. Within Table 1, the SMA slopes reflect the relationship $y \leftrightarrow x$, with the x as the column headers and the y being the row labels. Figures 3 through 6 illustrate the more important relationships involving the sampled structural traits. Due to considerations associated with multiple testing, we focus only on relationships significant

Table 1. Relationships between the derived genetic components of the observed plant traits: M_A = leaf mass per unit area (g m^{-2}); elemental concentrations are on a dry weight basis (mg g^{-1}), L_A = leaf area (m^2), ℓ_A = leaflet area (m^2), Φ_{LS} = leaf area:sapwood area ratio ($\text{cm}^2 \text{g}^{-1}$), ρ_x = branch xylem density (kg m^{-3}), \varnothing = stomatal limitation index (see Eq. 1), S = seed mass (g), H_{\max} = species maximum height (m). Values above the diagonal represent the slope of the relationship (y axis as columns labels, x axis as row labels). Values below the diagonal represent the correlation coefficient. Values significant at $P < 0.05$ are given in bold. NS = no slope estimated as the relationship was not significant.

Variable	M_A	[C]	log[N]	log[P]	log[Ca]	log[K]	log[Mg]	log(L_A)	log(ℓ_A)	log(Φ_{LS})	ρ_x	\varnothing	log(S)	H_{\max}
log(M_A)	–	0.37	–1.01	–1.21	NS	–1.65	–2.18	NS	4.32	–1.27	0.88	NS	19.2	167
[C]	0.15	–	NS	NS	–6.28	–4.43	–5.85	17.30	NS	NS	NS	NS	51.4	NS
log[N]	–0.43	0.07	–	1.20	NS	1.63	NS	6.36	–4.18	1.22	NS	0.22	–18.6	NS
log[P]	–0.41	–0.02	0.66	–	1.93	1.36	1.79	5.37	NS	1.03	–0.72	0.19	NS	NS
log[Ca]	–0.07	–0.51	0.02	0.14	–	0.7	0.93	NS	NS	NS	–0.37	0.10	–8.3	NS
log[K]	–0.28	–0.45	0.18	0.46	0.46	–	1.32	NS	2.60	NS	–0.52	NS	–11.4	NS
log[Mg]	–0.14	–0.45	0.05	0.18	0.65	0.59	–	–2.98	1.97	NS	–0.40	0.10	–8.8	NS
log(L_A)	–0.09	0.14	0.27	0.37	–0.03	–0.01	–0.14	–	0.65	0.19	–0.13	0.04	NS	NS
log(ℓ_A)	0.17	–0.08	–0.11	0.04	0.07	0.18	0.15	0.41	–	NS	–0.2	NS	NS	NS
log(Φ_{LS})	–0.24	0.06	0.20	0.14	0.00	0.04	–0.09	0.26	0.01	–	NS	NS	NS	NS
ρ_x	0.13	0.07	–0.08	–0.20	–0.21	–0.24	–0.12	–0.10	–0.22	0.07	–	NS	21.6	NS
\varnothing	0.09	0.03	0.23	0.27	0.12	0.06	0.09	0.12	–0.08	–0.08	–0.09	–	–81.8	731
log(S)	0.12	0.18	–0.16	–0.08	–0.34	–0.23	–0.25	0.02	0.00	–0.10	0.25	–0.20	–	8.8
H_{\max}	0.17	0.04	–0.03	0.00	–0.06	–0.02	–0.08	–0.02	0.00	–0.07	–0.03	0.11	0.14	–

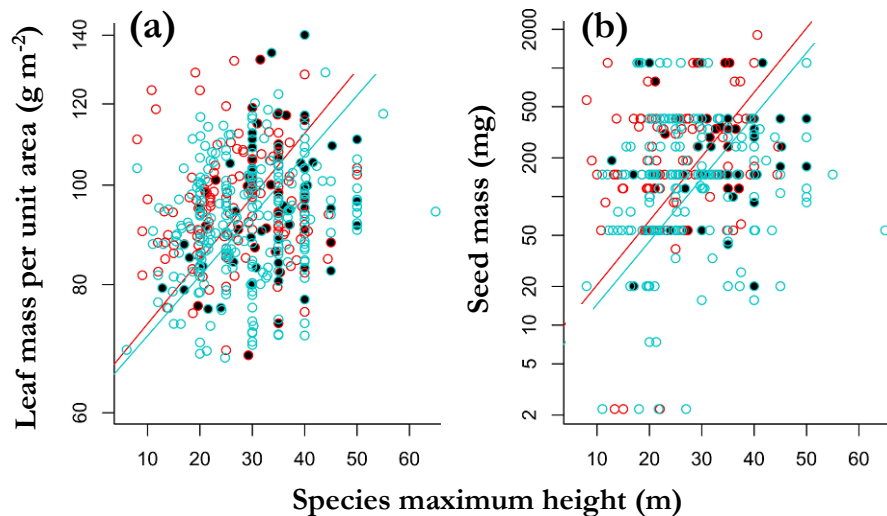


Fig. 3. Standard Major Axis (SMA) regression lines between species maximum height (H_{\max}) and the derived taxonomic components of leaf mass per unit area (M_A) for the same species and the associated average seed mass (S) for the associated genus. Red open circles indicate species found on low fertility sites and the blue open circles indicate species found on high fertility sites. Species found on both soil fertility groups are indicated with closed circles (see text for details). Red solid lines show the SMA model fit for low fertility species which is significantly different to the blue solid lines for high fertility soil species.

at $p \leq 0.001$ though, where interesting and/or informative, statistically less significant relationships are also considered.

3.4.1 Maximum tree height

Generally only poor correlations were observed for H_{\max} , these being significant only for $\log_{10}(M_A)$ ($p \leq 0.001$) and $\log_{10}(S)$ ($p \leq 0.01$). The $M_A \leftrightarrow H_{\max}$ and $S \leftrightarrow H_{\max}$ relationships are shown in Fig. 3. Here, due to differences in the

SMA slope and/or intercept between the species associated with the two soil fertility classes (see Supplementary Information, Table S2B) we have fitted separate lines for species found on low and high fertility soils. This shows that for species associated with low fertility soils, both M_A and S tend to be slightly higher at a given H_{\max} than their higher fertility counterparts. Especially for $S \leftrightarrow H_{\max}$ the variation is considerable, particularly at low H_{\max} , with S varying three orders of magnitude for H_{\max} between 10 and 30 m.

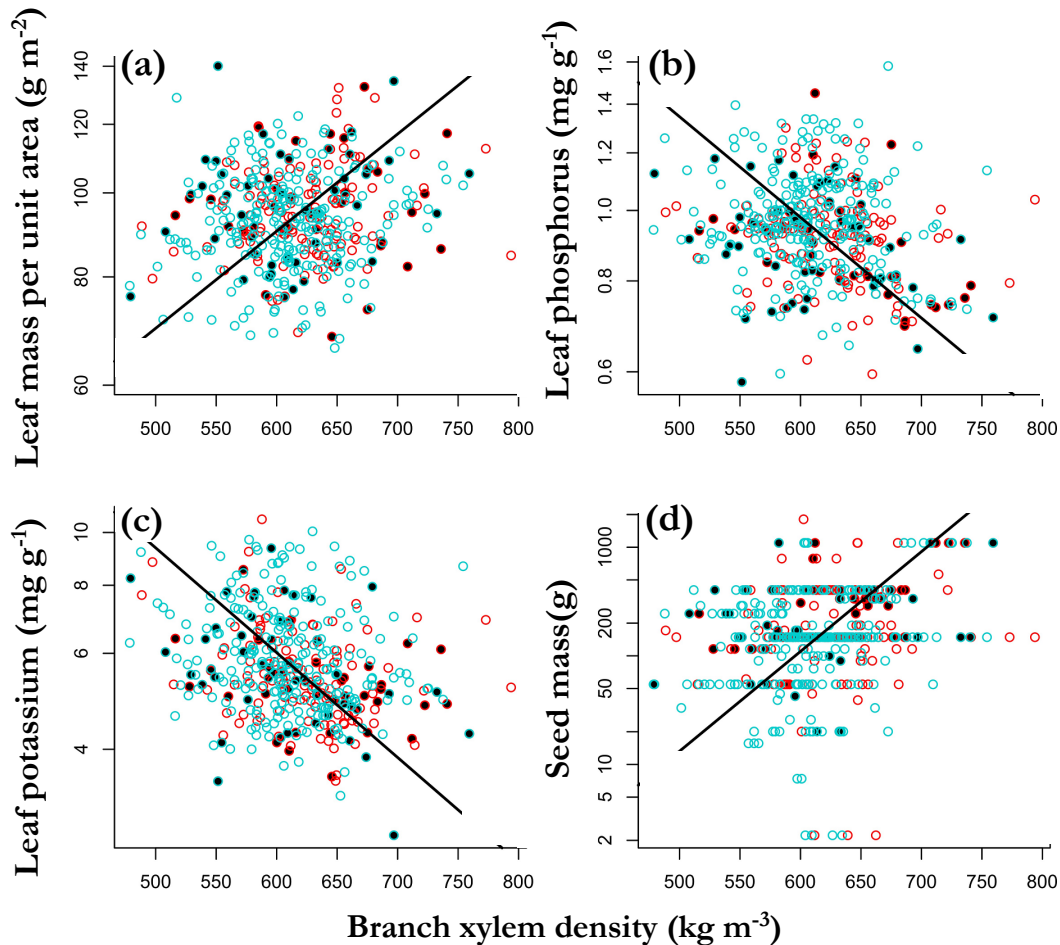


Fig. 4. Standard Major Axis (SMA) regression lines between the derived species components of branch xylem density (ρ_x) and those for mass per unit area (M_A), foliar [P] and foliar [K] for the same species and the average seed mass (S) for the associated genus. Red open circles indicate species found on low fertility sites and the blue open circles indicate species found on high fertility sites. Species found on both soil fertility groups are indicated with closed circles (see text for details). The black solid lines show the SMA model fit which did not depend on soil fertility.

3.4.2 Branch xylem density

As detailed in Table 1, the derived taxonomic component of ρ_x was negatively correlated with $\log_{10}[\text{P}]$, $\log_{10}[\text{Ca}]$, $\log_{10}[\text{K}]$, $\log_{10}(\ell_A)$ and positively associated with $\log_{10}(S)$ ($p \leq 0.001$). A weaker but significant positive correlation was also observed with $\log_{10}(M_A)$ and a negative correlation with $\log_{10}[\text{Mg}]$ ($p \leq 0.01$). Of minor significance was a negative association with $\log_{10}(L_A)$ ($p \leq 0.05$). Some of these relationships are illustrated in Fig. 4 which shows the relationships between ρ_x and both [P] and [K] to be particularly compelling and, as is also the case for M_A and S , with no difference for species associated with low versus high fertility soils.

3.4.3 Leaf area: sapwood area ratio

Reasonably strong correlations were found for $\log_{10}(\Phi_{LS})$ with $\log_{10}(M_A)$, $\log_{10}[\text{N}]$ and $\log_{10}(L_A)$ ($p \leq 0.001$) with the relationship between $\log_{10}(\Phi_{LS})$ and $\log_{10}[\text{P}]$ also significant ($p \leq 0.01$). The relevant biplots are shown in Fig. 5. The slope for the taxonomic component $M_A \leftrightarrow \Phi_{LS}$ relationship is $1/-1.27 = -0.79$. Thus, as Φ_{LS} increases across species, then M_A declines proportionally less. That is to say, species with a higher Φ_{LS} also tend to carry a greater weight of (generally larger) leaves per unit stem area with those leaves also tending to have higher foliar [N] and [P].

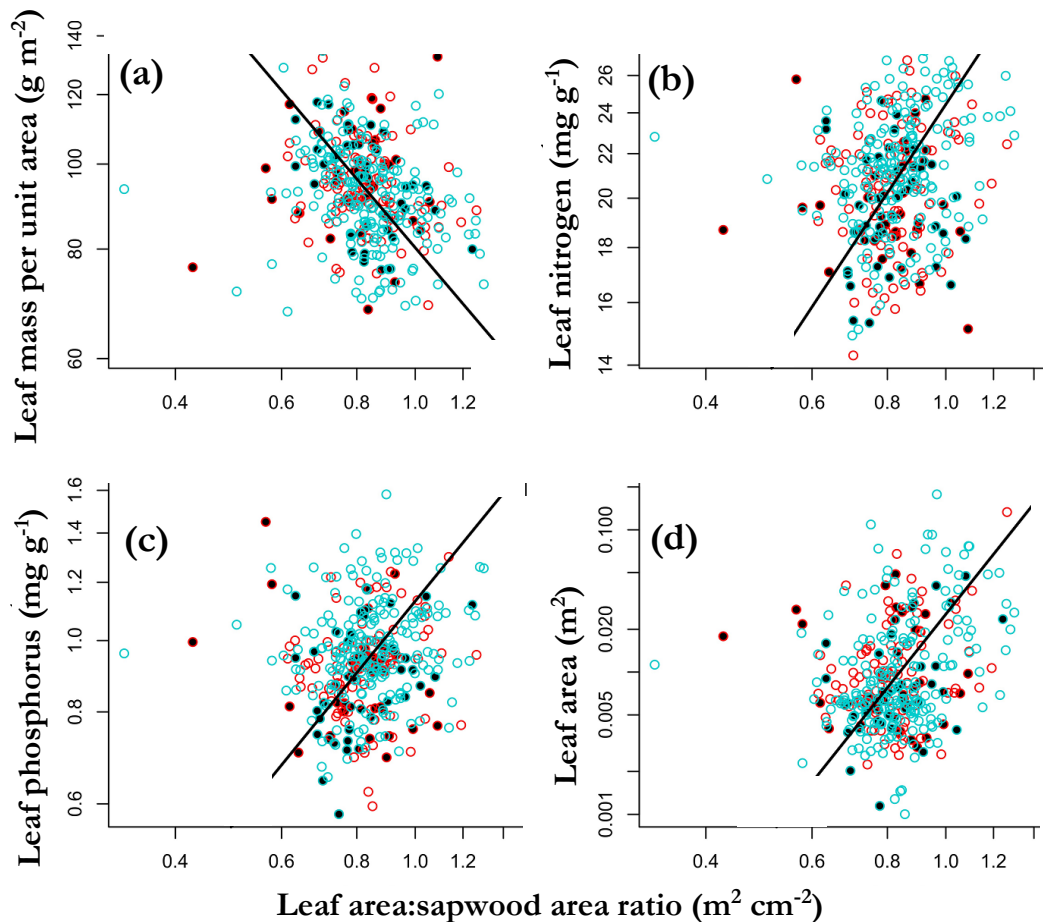


Fig. 5. Standard Major Axis (SMA) regression lines between the derived species components of leaf area/sapwood area ratio (Φ_{LS}) and those for mass per unit area M_A , foliar [N], foliar [P] and average leaf size for the same species. Red open circles indicate species found on low fertility sites and the blue open circles indicate species found on high fertility sites. Species found on both soil fertility groups are indicated with closed circles (see text for details). Solid lines show the SMA model fit which did not depend on soil fertility.

3.4.4 Leaf nutrients and other structural traits

Strong positive correlations ($p \leq 0.001$) were also observed for $\log_{10}(L_A)$ with $\log_{10}[N]$ and $\log_{10}[P]$ as well as between $\log_{10}[Ca]$ and S . Interestingly, both the slope and intercept of these relationships are dependent on the soil fertility with which a species is associated (Supplementary Information Table S2B). Species found on low fertility soils tend to have a higher L_A at any given foliar [N] and/or [P].

For the $[Ca] \leftrightarrow S$ pairing the negative slope is also large (-8.3), though in this case with no soil fertility effect detected. Though not shown in Fig. 6, also of note is the positive $[C] \leftrightarrow S$ relationship ($p \leq 0.001$) with species with a low seed mass also tending to have a low foliar carbon content.

3.5 Common Principal Component modelling (taxonomic components)

Results from the CPC modelling are shown in Table 2, with the full model output, details of the rationale for eigenvector inclusion and assessments of the overall model fit given in the Supplementary Information Tables S3, S4 and S5 and their accompanying captions. The five eigenvectors selected are listed in Table 2 in order of their importance, as derived from the characteristic roots (eigenvectors, λ). These results can be interpreted as in the case of an ordinary principal components analysis, the difference here being that the relative weightings (λ) have been allowed to differ for species on high vs. low fertility soils.

The first eigenvector, U_1 , had somewhat higher λ for high vs. low fertility associated species (accounting for 0.24 and 0.27 of the dataset variance respectively) and with high positive coefficients for all three foliar cations and to a lesser

Table 2. Common principal component analysis of derived genetic effects for species associated with low and high fertility soils. Values in brackets represent standard errors for each component. Coefficients given in bold are either those whose absolute values are 0.50 or more, or 0.30 or more with a standard error of less than 0.1. M_A = leaf mass per unit area; elemental concentrations are on a dry weight basis, L_A = leaf area; Φ_{LS} = leaf area:sapwood area ratio, ρ_x = branch xylem density, \emptyset = diffusion limitation index (see Eq. 1), S = seed mass, H_{max} = species maximum height.

Variable	Component				
	U_1	U_2	U_3	U_4	U_5
$\log(M_A)$	-0.22(0.05)	-0.23 (0.06)	0.44 (0.07)	-0.22 (0.11)	0.35 (0.09)
[C]	-0.35 (0.05)	0.24 (0.07)	0.01 (0.07)	0.06 (0.12)	0.34 (0.09)
$\log[N]$	0.15 (0.10)	0.53 (0.04)	-0.02 (0.09)	0.22 (0.09)	-0.03 (0.08)
$\log[P]$	0.25 (0.08)	0.45 (0.05)	0.12 (0.09)	0.31 (0.05)	0.08 (0.06)
$\log[Ca]$	0.42 (0.03)	-0.13 (0.08)	0.15 (0.09)	-0.31 (0.06)	0.00 (0.08)
$\log[K]$	0.48 (0.02)	-0.01 (0.09)	0.00 (0.08)	0.16 (0.09)	0.05 (0.11)
$\log[Mg]$	0.49 (0.04)	-0.21 (0.09)	0.07 (0.06)	0.06 (0.07)	0.19 (0.07)
$\log(L_A)$	-0.01 (0.09)	0.48 (0.05)	0.25 (0.13)	-0.35 (0.16)	-0.16 (0.10)
$\log(\Phi_{LS})$	-0.01 (0.07)	0.29 (0.06)	-0.44 (0.11)	-0.53 (0.16)	0.18 (0.11)
ρ_x	-0.14 (0.03)	-0.03 (0.05)	-0.22 (0.10)	0.12 (0.21)	0.26 (0.11)
\emptyset	0.10 (0.04)	0.14 (0.05)	0.39 (0.09)	-0.10 (0.13)	0.60 (0.08)
$\log(S)$	-0.23 (0.03)	0.01 (0.06)	0.19 (0.10)	0.48 (0.10)	0.59 (0.08)
H_{max}	-0.10 (0.04)	0.07 (0.06)	0.53 (0.10)	-0.13 (0.22)	-0.47 (0.09)
Characteristic roots					
$\lambda_{low,j}$	1876 (259)	1472 (203)	641(89)	717 (99)	698 (96)
$\lambda_{high,j}$	2341 (237)	1641 (166)	898 (91)	564 (57)	318 (32)

Table 3. Bivariate relationships for the derived environmental component of the observed plant traits. Values above the diagonal represent the slope of the relationship (y axis as columns labels, x axis as row labels). Values below the diagonal represent the correlation coefficient. Values significant at $P < 0.05$ are given in bold. NS = no slope estimated as the relationship was not significant. For units and symbols, see Table 1.

Variable	$\log(M_A)$	[C]	$\log[N]$	$\log[P]$	$\log[Ca]$	$\log[K]$	$\log[Mg]$	$\log(L_A)$	$\log(\ell_A)$	$\log(\Phi_{LS})$	ρ_x	\emptyset
$\log(M_A)$	–	0.31	-1.06	NS	-4.97	NS	-1.32	NS	NS	-3.49	NS	0.57
[C]	0.63	–	-3.38	NS	-15.86	NS	-4.22	NS	NS	NS	4.10	1.82
$\log[N]$	-0.52	-0.30	–	2.69	4.68	NS	1.25	NS	NS	3.06	NS	NS
$\log[P]$	-0.04	-0.09	0.48	–	1.74	1.53	NS	NS	NS	NS	-0.45	0.20
$\log[Ca]$	-0.28	-0.54	0.28	0.50	–	0.88	0.27	NS	NS	NS	-0.26	NS
$\log[K]$	-0.01	-0.13	0.23	0.74	0.49	–	NS	NS	NS	NS	-0.30	0.13
$\log[Mg]$	-0.54	-0.72	0.28	0.04	0.50	0.05	–	NS	NS	NS	NS	NS
$\log(L_A)$	0.08	-0.06	-0.09	-0.03	0.11	0.21	0.07	–	0.85	NS	NS	NS
$\log(\ell_A)$	0.07	-0.20	-0.16	-0.11	0.09	0.20	0.15	0.90	–	NS	-0.79	-0.35
$\log(\Phi_{LS})$	-0.29	-0.25	0.36	-0.07	-0.02	-0.21	0.10	0.07	0.08	–	NS	NS
ρ_x	0.08	0.27	-0.22	-0.64	-0.46	-0.82	-0.06	-0.25	-0.31	0.17	–	NS
\emptyset	0.32	0.27	0.24	0.49	0.25	0.31	-0.22	-0.14	-0.28	-0.09	-0.18	–

extent foliar [P], and negative coefficients for foliar [C] with smaller but still significant coefficients for M_A and S . In terms of cations, carbon and M_A , this first component seems similar to that first described by Poorter and de Jong (1999) and thus we dub it the Poorter-De Jong (PDJ) dimension, \mathcal{A}_{PDJ} .

The second component, U_2 , accounts for an additional 0.18 and 0.19 of the dataset variances for low and high fertility species respectively, and is characterised by high positive

coefficients for foliar [N] and [P] as well as L_A and, to a lesser extent, Φ_{LS} . Also notable are modestly negative coefficients for M_A and foliar [Mg]. In terms of [N], [P] and M_A , U_2 , seems to reflect some components of what is considered the classic leaf economic spectrum (Reich et al., 1997; Wright et al., 2004). We thus label this the Reich-Wright dimension, \mathcal{A}_{RW} , of tropical tree functional trait coordination.

Although H_{max} would seem to have little influence on either \mathcal{A}_{PDJ} or \mathcal{A}_{RW} it emerges as the dominant term for U_3

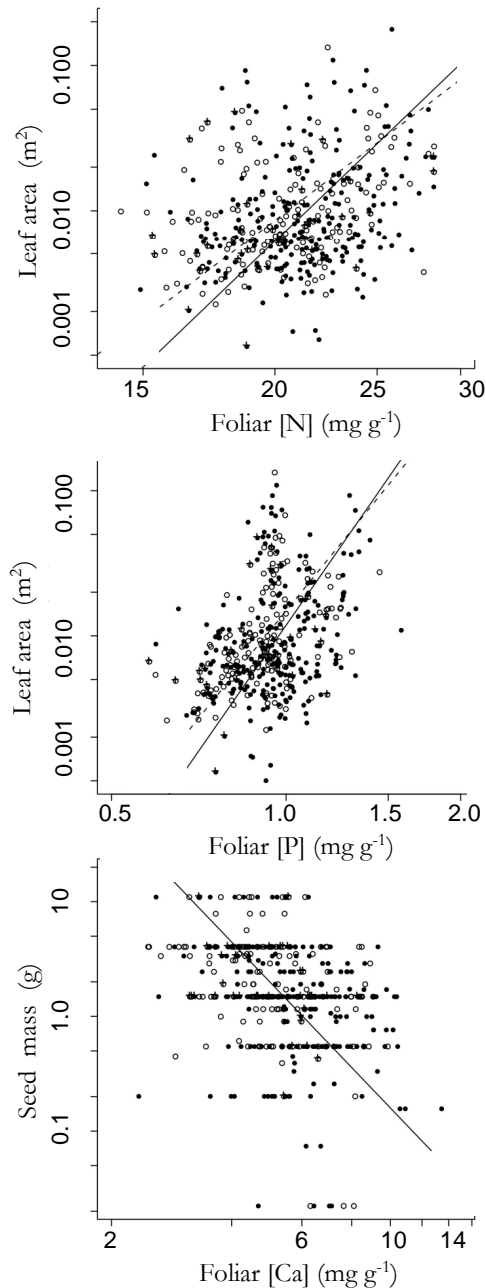


Fig. 6. Standard Major Axis (SMA) regression lines between derived taxonomic components of foliar [N] and foliar [P] and leaf mass per unit area (L_A) for the top two panels and between species estimated foliar [Ca] associated average seed mass (S) for the associated genus (bottom panel). Open circles indicate species found on low fertility sites and the closed circles indicate species found on high fertility sites. Species found on both soil fertility groups are designated by a “+” (see text for details). For the top two panels, solid lines show the SMA fit for low fertility soil species which are significantly different to the dashed lines for high fertility soil species. For the bottom panel the solid line shows the SMA model fit which did not depend on soil fertility.

along with M_A and, of opposite sign, Φ_{LS} . Also of note here is the relatively high value for the coefficient of the diffusion limitation index, ϕ which is positively associated with both H_{max} and M_A . Interestingly, for this component L_A varies in the opposite direction to Φ_{LS} (albeit with a large standard error) suggesting that there is a tendency towards considerably fewer but also significantly larger leaves in taller stunted species. There also being a modest but significant negative contribution of ρ_x to this dimension. We consider U_3 , which on its own accounts for 0.08 and 0.10 of the variation in the dataset respectively, to contain several features similar to those described by Falster and Westoby (2005) for climax tropical forest in Australia, and it is thus denoted as \bar{A}_{FW} .

The fourth component axis is dominated by S and Φ_{LS} with these coefficients of different sign. Associated with the higher S are also lower [Ca] but higher foliar [P] and L_A . With lower values for their coefficients and higher standard errors, also being of different sign, are the M_A and [N] terms. As mentioned in the Discussion, U_4 (accounting for 0.09 and 0.07 of the population variance for low and high fertility species respectively) seems to be dominated by the presence of large seeded members of the Leguminaceae whose importance in the phytogeography of Amazon forest has already been recognised by ter Steege et al. (2006). We therefore denote this dimension as \bar{A}_{TS} .

The last eigenvector included in our analysis, U_5 , differs from the others in having a substantially greater importance for low fertility versus high fertility species (accounting for 0.09 and 0.04 of the population variances respectively). This component is characterised by H_{max} and M_A having opposite signs (in contrast to \bar{A}_{FW}) and with higher S and ϕ also being associated with a lower H_{max} ; this also being along with a less substantial but significant coefficient for ρ_x . Also of influence in characterising U_5 are greater foliar [C] associated with the higher M_A and ϕ . Although, U_5 presents some trait combinations as reported previously in the literature, this component, mostly related with species found at low fertility soils, does not seem to have been recognised before. It is thus here denoted as \bar{A}_{PFL} .

Overall the five eigenvectors selected, all of which we believe to be physiologically relevant (see Supplementary Information), accounted for 0.68 of the total variance for both low and high fertility soil species.

The first three axes species scores (normalised to ± 100) are plotted against each other in Supplementary Information Fig. S1. This shows the required lack of any systematic correlations between the species scores as expected for the output from any good fit of a principle components model. Clearly a wide range of combinations of these three trait dimensions can occur. Figure 8a also shows that it is (generally speaking) only species typically associated with high fertility soils that have high scores for both \bar{A}_{PDJ} and \bar{A}_{RW} .

Figure 7 shows the major components of the three major CPCs and their overlap of traits in diagrammatic form. This

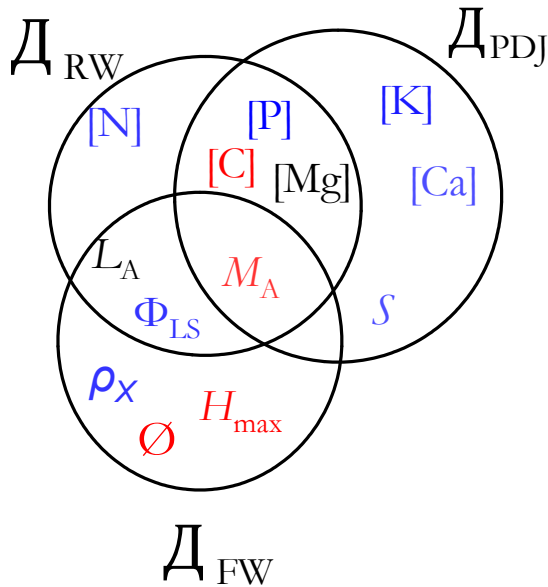


Fig. 7. Euler diagram showing overlaps between the first three dimensions for the individual measured traits (where significant): blue; positive relationship with dimension, red; negative relationship with dimension, black; of different sign depending on the dimension. Abbreviations are as in Table 5, with the three trait dimensions as defined in Sect. 3.5

illustrates that many traits seem to be “shared”, especially M_A which is an important factor for all three of \bar{D}_{PDJ} , \bar{D}_{RW} and \bar{D}_{FW} . Also occurring in $(\bar{D}_{PDJ} \cap \bar{D}_{RW})$ and of the same sign is [C], but with [P] and [Mg] varying in opposite directions with respect to M_A for these two trait dimensions. Intersecting \bar{D}_{RW} and \bar{D}_{FW} and in the same direction relative to M_A is Φ_{LS} . Although with a high estimated standard error as part of \bar{D}_{FW} , we have also included L_A in $(\bar{D}_{RW} \cap \bar{D}_{FW})$, this also showing that it varies in the opposite direction relative to M_A and Φ_{LS} for \bar{D}_{RW} cf. \bar{D}_{FW} .

3.6 Bivariate relationships: environmental components

Considering data from both low and high fertility sites together, Table 3 lists correlations and SMA slopes for the environmental effects with this information provided in more detail (including confidence intervals) in the Supplementary Information (Table S2A). As for Table 1, the SMA slopes reflect the relationship $y \leftrightarrow x$, with the x as the column headers and the y being the row labels. For the structural traits, the most significant relationships are all negative and appear between ρ_x and $\log_{10}[\text{P}]$, $\log_{10}[\text{Ca}]$, $\log_{10}[\text{K}]$ and, to a lesser extent $\log_{10}(\ell_A)$. The slopes observed (-0.26 to -0.41) are, however, much less than for the associated slopes for the taxonomic components as listed in Table 1 (-0.37 to -0.72).

Table 4. Summary of the Principal Components Analysis of the correlation matrix for the derived environmental/soil effects on observed structural and physiological traits. Coefficients given in bold are those whose values are 0.3 or more. M_A = leaf mass per unit area; elemental concentrations are on a dry weight basis, L_A = leaf area; Φ_{LS} = leaf area: sapwood area ratio, ρ_x = branch xylem density, \emptyset = diffusion limitation index (see Eq. 1).

Variable	Component	
	ψ_1	ψ_2
$\log(M_A)$	-0.196	-0.443
[C]	-0.300	-0.412
$\log[\text{N}]$	0.320	0.111
$\log[\text{P}]$	0.406	-0.276
$\log[\text{C}_A]$	0.453	0.099
$\log[\text{K}]$	0.392	-0.300
$\log[\text{Mg}]$	0.245	0.416
$\log(L_A)$	0.087	-0.009
$\log(\Phi_{LS})$	0.025	0.271
ρ_x	-0.383	0.287
\emptyset	0.174	-0.340
Eigenvalue	6.23	2.54
Proportion of variance explained	0.33	0.25

3.7 Principal component analysis of environmental effects

Given the correlations between the environmental effects for ρ_x and several foliar nutrients (Table 3; Fig. 8), it was of additional interest to see if coordinated structural/leaf biochemical responses to the environment exist for Amazon forest. We therefore undertook a PCA analysis of the full plot effects correlation matrix (excluding H_{max} and S both of which were considered to be environmentally invariant for any given species) with the results shown in Table 4. This shows that 0.33 of the total variation in the 11 traits examined could be explained by the first PCA axis (ψ_1) with ρ_x an important contributor and this also relating positively to foliar [C] and M_A , but negatively with all foliar nutrients examined and also with \emptyset . The second axis of the PCA on the plot effects correlation matrix (ψ_2) is also significant, accounting for 0.25 of the variance, with substantial negative weightings for M_A , foliar [C] and \emptyset (and to a lesser extent foliar [P]) being balanced by positive weightings for foliar [Mg] in particular, but also with contributions from Φ_{LS} and ρ_x .

3.8 Relationship between plot effect PCAs and soil/climate

The most significant relationships between the PCA site axis scores of Table 4, and previously calculated soil and climate characteristics of the same sites are shown in Fig. 9. First, the top panel of Fig. 9 shows ψ_1 as a function of the first soil PCA axis of Fyllas et al. (2009), the latter considered a strong

Table 5. Kendall’s partial correlation coefficient, τ_p , for the environmental contribution (plot effect estimate) of each foliar property (controlling for the effects of the other environmental predictors) with their significance estimated as detailed in Maghsoodloo and Laszlo Pallos (1981). Bold values indicate a very strong correlation ($p < 0.001$) and italics indicate significant correlations at $p < 0.01$; see text for details. M_A = leaf mass per unit area; elemental concentrations are on a dry weight basis, L_A = leaf area; ℓ_A = leaflet area, Φ_{LS} = leaf area/sapwood area ratio, ρ_x = branch xylem density, \oslash = diffusion limitation index (see Eq. 1) and η_1 and η_2 are the first two principal components of the PCA analysis on the environmental effects correlation matrix (See Table 4).

	M_A	[C]	[N]	[P]	[Ca]	[K]	[Mg]	L_A	ℓ_a	$\log(\Phi_{LS})$	ρ_x	\oslash	η_1	η_2
Soil fertility PCA axis, ϕ_F	-0.20	-0.23	0.20	0.48	0.48	0.33	0.22	-0.09	-0.07	-0.04	-0.32	0.20	0.56	0.00
Soil texture PCA axis, ϕ_T	0.05	0.10	0.12	0.04	-0.27	-0.17	-0.18	-0.03	0.02	0.05	0.19	0.02	-0.22	-0.07
Mean annual temperature, T_a	0.11	0.051	-0.38	-0.26	-0.08	-0.41	0.03	-0.08	-0.04	0.07	0.35	-0.13	-0.23	0.21
Mean annual precipitation, P_a	0.33	0.30	-0.18	0.17	-0.01	0.11	-0.31	-0.01	-0.01	0.17	-0.12	0.24	-0.07	-0.44
Mean annual radiation, Q_a	-0.06	0.15	0.02	0.12	-0.14	0.08	0.00	-0.11	-0.10	0.08	0.02	0.12	-0.04	-0.11

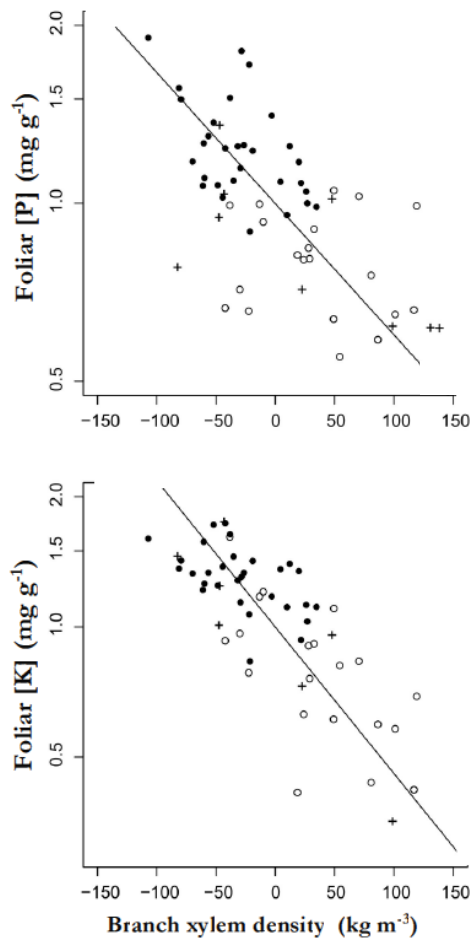


Fig. 8. Standard Major Axis (SMA) regression lines between the derived environmental components of branch xylem density (ρ_x) and foliar [P] and foliar [K]. Open circles indicate species found on low fertility sites and the closed circles indicate species found on high fertility sites. Species found on both soil fertility groups are designated by a “+” (see text for details). Solid lines show the SMA model fits.

integrated measure of soil fertility and denoted ϕ_F . The strong relationship observed suggests an integrated response of Amazon tropical forest trees to soil fertility, with most nutrients increasing, and with foliar [C] and ρ_x decreasing as ϕ_F increases. Interestingly, the Kendall’s τ for this plot of η_1 versus ϕ_F of 0.63 is greater than for any of the original variables examined by Fyllas et al. (2009), the highest of which was 0.56 for foliar [P]. Comparison with Fyllas et al. (2009) also shows that the η_2 contains significant weightings of leaf-level variables that, individually, were all strongly correlated with mean annual precipitation (P_A) viz. positive correlations with foliar [C] and M_A and a negative correlation with foliar [Mg]. It is therefore not surprising, as is shown in the second panel of Fig. 9, that η_2 and P_A also show strong association, but with examination of Table 4 also suggesting that for any given species, both Φ_{LS} and ρ_w also decline with increasing precipitation and, somewhat counter intuitively, with \oslash increasing.

Finally, as in Fyllas et al. (2009) we show values for Kendall’s partial τ (denoted τ_p) for all traits of interest as well as η_1 and η_2 as functions of ϕ_F , ϕ_T , T_a , P_a and Q_a in Table 5. Here the calculated value of τ_p and associated probability giving an indication of the effect of each soil/environmental parameter after accounting for the effect of the other four. Taking into account the potential confounding effects of spatial autocorrelation (Fyllas et al., 2009) we only consider relationships with $p \leq 0.01$ or better. As for the (full) Kendall’s τ shown in Fig. 9, Table 5 suggests the ϕ to be superior predictors than the individual variables, the only exception being T_a . In that case, [N], [K] and ρ_w all show relationships not present when regressing the plot effect PCs as dependent variables.

4 Discussion

Some of the data used here have been presented previously (Fyllas et al., 2009; Patiño et al., 2009), with the current analysis integrating those datasets with structural traits introduced as part of this study (viz. L_A , ℓ_A , Φ_{LS} , S and H_{max})

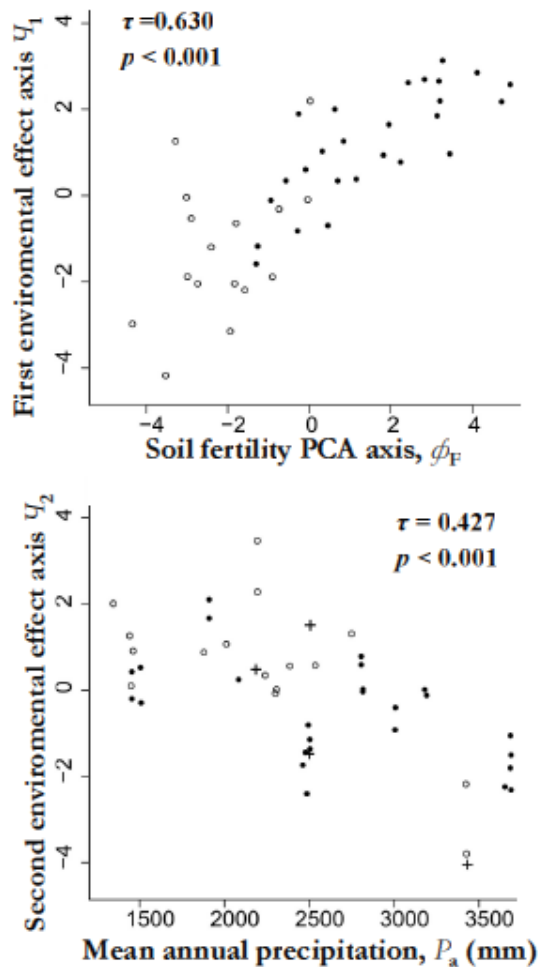


Fig. 9. Relationship between derived environmental effect principal components (Table 5) and soil/environmental parameters for various plots across the Amazon. Top panel, first principal component of the environmental effects versus the first principal component of the PCA of soil chemical and physical characteristics as derived by Fyllas et al. (2009) on the basis of data provided by Quesada et al. (2010). Second panel, second principal component of the environmental effects PCA versus mean annual precipitation. Open circles indicate low fertility sites and the closed circles indicate high fertility sites as defined by Fyllas et al. (2009).

as well as with foliar $^{13}\text{C}/^{12}\text{C}$ ratios as reinterpreted through the diffusional limitation index, ϕ , as defined by Eq. (1). We first consider the bivariate relationships between the structural components introduced as part of this study as well as relationships between these structural traits and the others already presented (Fyllas et al., 2009; Patiño et al., 2009), this then being extended to a consideration of how variations in these traits coordinate in response to differences in species and/or environment. Here we emphasise that, as estimated within the study, our “environmental effects” reflect modulation of taxon specific trait values by soils and/or cli-

mate. To this extent they reflect a systematic component of intra-species variability, i.e. that predictable from where a particular species is growing, as opposed to a more random within population component, such as might be expected when comparing the same species growing nearby under the same edaphic and climatic conditions. This portion, along with experimental error is theoretically included in the residual component of the analysis (i.e., that not accounted for by the fitted model itself) which, as shown in Figure 2, can sometimes be substantial. The extent to which this component of trait variation relates to within plot variability in microclimate or soil characteristics (rather than intrinsic within-species differences or sampling/measurement error) remains to be established.

4.1 Bivariate relationships for the taxonomic component of trait variation

4.1.1 Maximum tree height, branch xylem density and leaf mass per unit area

These three structural traits have often been associated with each other with significant positive $\rho_x \leftrightarrow M_A$ correlations such as for our taxonomic component in Fig. 4 also reported by Bucci et al. (2004), Ishida et al. (2008) and Meinzer et al. (2008). Those studies interpreted this relationship in terms of higher density wood species having lower hydraulic conductances leading to a requirement for more robust leaves capable of sustaining more severe soil water deficits. This notion is supported by more negative osmotic potentials being reported for the leaves of higher M_A and ρ_x species (Bucci et al., 2004; Ishida et al., 2008; Meinzer et al., 2008). On the other hand, it is also the case that M_A tends to increase with actual or potential (maximum) tree height (Falster and Westoby, 2005; Kenzo et al., 2008; Lloyd et al., 2010) and that ρ_w and H_{\max} are sometimes negatively correlated (Falster and Westoby, 2005; van Gelder et al., 2006). This then implying that ρ_w and M_A should be negatively (as opposed to positively) correlated as well.

One reason for this apparent contradiction may be that wood density and xylem vessel traits do not necessarily represent the same axis of ecophysiological variation (Preston et al., 2006; Martínéz-Cabrera et al., 2009; Poorter et al., 2010; Baraloto et al., 2010). For example, decreasing wood density associated with increasing foliar P concentration and lower LMA is also likely associated with decreasing investment in wood physical and chemical defences (Augsburger, 1984; Putz et al., 1983; King, 1986; Chao et al., 2008), including resistance against breakage (Romero and Bolker, 2008). One interpretation of Fig. 4a–c is then simply that tropical tree species with traits associated with a higher photosynthetic potential such as a high foliar [P] (Domingues et al., 2010), also tend to invest less towards wood defensive strategies (but see Larjavaara and Muller-Landau, 2010).

Our observation of significant within-species variation in ρ_x as illustrated in more detail by Patiño et al. (2009) and also observed for ρ_w by Omolodun et al. (1991), Hernández and Restrepo (1995), Gonzalez and Fisher (1998), Weber and Montes (2008) and Sungpalee et al. (2009), shows important intraspecific variation in xylem and/or wood density even within one plot (as also evidenced by the “residual” term for ρ_x in Fig. 2) as well as being systematically affected by soil fertility (Table 5). Thus, although we do not dispute that xylem traits and ρ_w/ρ_x may not necessarily be closely or mechanistically linked (as discussed above), studies which simply compare wood density “species values” as measured in one study or studies with values of ρ_w/ρ_x for the same species but gathered from a completely independent source (Russo et al., 2010; Zanne et al., 2010) are effectively comparing bananas with wombats. Thus, also not employing robust regression techniques more applicable to such analyses (McKean et al., 2009) they must under-estimate the actual significance of any relationship, be it functional or not.

So, does the observation of large diameter xylem vessels with a high K_S also being associated with a greater H_{\max} (e.g., Poorter et al., 2010; Zach et al., 2010) mean that the tendency of mature forests species of a greater H_{\max} to also have a lower ρ_w (Falster and Westoby, 2005; van Gelder et al., 2006; Baker et al., 2009; Poorter et al., 2009) is indicative of some sort of functional linkage? Or does it more simply reflect that the fast-growing and light-demanding species characteristic of “dynamic” tropical forests also tend to have a lower ρ_w – this presumably allowing a faster height and diameter growth rate? On the basis of the discussion above, we suggest the latter, also noting that ρ_w is actually generally better correlated with juvenile light-exposure than H_{\max} (van Gelder et al., 2006; Poorter et al., 2009).

The positive relationship between M_A and H_{\max} of Fig. 3a and as also evident in the data of Falster and Westoby (2005) can also be inferred from the positive M_A vs. tree height relationships as reported by Thomas and Bazzaz (1999), Kenzo et al. (2006) and Lloyd et al. (2010). This is also seen within \bar{A}_{FW} in the CPC analysis of Table 3, with the leaves of (potentially) taller trees being thicker (Kenzo et al., 2006; Rozendaal et al., 2006) with a greater mesophyll thickness associated with a higher photosynthetic capacity per unit area (Kenzo et al., 2006). This increase in M_A with tree height being mostly associated with a greater mesophyll thickness should allow for a more efficient use of the higher rates of insolation towards the canopy top through higher photosynthetic capacities per unit leaf area (Rijkers et al., 2000). Along with more negative osmotic potentials, the greater tissue densities associated with a higher M_A and H_{\max} should also help sustain leaves of such taller trees in the face of the more severe water deficits expected for sun exposed leaves higher up in the canopy (Cavaleri et al., 2010; Lloyd et al., 2010).

4.1.2 Leaf size, nutrients and Φ_{LS}

Species with intrinsically higher foliar nutrient concentrations also tend to be found on more fertile soils (Fyllas et al., 2009), and so the positive correlation between the taxonomic components of leaf size variation, foliar [N] and foliar [P] observed here (Fig. 6) is consistent with the observation that Australian tropical forest tree species associated with poorer soils tend to have smaller leaves than those associated with more eutric conditions (Webb, 1968). This was also found to be the case for south-eastern Australian woodland species once precipitation effects were also taken into account (McDonald et al., 2003). Such a relationship has also been observed for pre-montane subtropical forest species in Argentina (Easdale and Healey, 2009) and has been suggested to be a widespread phenomenon (Givnish, 1987) perhaps being explainable by low N and/or P leaves typically having lower gas exchange rates than those of a higher fertility status (Domingues et al., 2010); with associated lower latent heat loss rates due to lower stomatal conductances. This would give rise to a greater rate of sensible heat loss being required to avoid over-heating during times of high insolation being achieved through the higher boundary layer conductance of smaller leaf sizes (Yates et al., 2010). Alternatively, and consistent with the general notion of plants growing on less fertile soils having more conservative growth strategies (Westoby et al., 2002), smaller leaves may be favoured on low nutrient soils despite their relatively higher construction costs. This is because they also have shorter expansion times with an associated reduction in herbivory losses during this susceptible phase of foliar development (Moles and Westoby, 2000). If the “heat budget” explanation were to be correct, then an even better correlation with ℓ_A would be expected for both foliar [N] and [P]. But this was not the case (Table 1) with both foliar [N] and [P] much more closely correlated with L_A . On the other hand, the relationship between leaf size and expansion time does not appear to differ strongly between simple vs. compound leaves (Moles and Westoby, 2000). This suggests that the herbivory hypothesis may be the more correct.

Although not significant across the dataset as a whole, there was a significant negative correlation between L_A and ρ_x for species characteristic of low fertility sites ($r^2 = -0.17$, $p \leq 0.05$: Supplementary Information, Table S2B) as has also been reported for Australian tree/shrub species by Pickup et al. (2005) and Wright et al. (2007) and for neotropical forest tree species by Swenson and Enquist (2008), Malhado et al. (2009), Baraloto et al. (2010) and, with a much lower correlation ($r^2 = -0.02$) by Wright et al. (2006). Exactly as to why this should be the case is currently unclear. Earlier arguments have revolved around not only ρ_x and K_S being closely linked, but also with the assumption that variations in L_A should to a large extent reflect variations in Φ_{LS} (Wright et al., 2006). But, as discussed in Sect. 4.1.1, wood density and plant hydraulics may not be as closely linked as

once thought and, although Φ_{LS} is indeed correlated with L_A (Fig. 6), our data do not actually show any appreciable correlations between Φ_{LS} and ρ_x (Table 1; Supplementary Information, Table S2B). This suggests that for tropical trees at least, this correlation may be more “casual” than mechanistic. Indeed, both for the dataset as a whole and for the individual fertility groupings, ρ_x was better (negatively) correlated with ℓ_A than L_A (Table 1, Supplementary Information, Table S2B). Given that compound leaves are generally associated with faster diameter increment species (Givnish, 1978; Malhado et al., 2010) as is a generally lower ρ_x (Keeling et al., 2008) this then suggests that the negative correlation between laminar size and wood density may just reflect both traits being associated with faster growth rates. As well as tending to have lower ρ_w (Sect. 4.1.2) such species also tend to exhibit less branching than more shade tolerant species (Poorter et al., 2006; Poorter and Rozendaal, 2006; Takahashi and Mikami, 2008). Presumably (along with wider spacings) this allows for larger leafed upper-canopy species to have greater rates of direct light interception (Falster and Westoby, 2003).

Increasing M_A with decreasing Φ_{LS} as shown in Fig. 5 does not seem to have been detected in other studies with tropical tree species (Meinzer et al., 2008; Zhang and Cao, 2009). Although it is notable that working with a range of emergent or upper-canopy dipterocarp species, Zhang and Chao (2009) did find a significant negative relationship between Φ_{LS} and leaf thickness, the latter being associated with variations in M_A with tree height for dipterocarp species (Kenzo et al., 2006). Sampling across a range of sites in south-eastern Australia, Pickup et al. (2005) also found a negative relationship between M_A with Φ_{LS} but this relationship was, overall, not significant for species sampled within individual sites. Our own data suggest a stronger linkage of M_A with Φ_{LS} than either L_A or (indeed even of different sign) ℓ_A . This suggests (as is discussed further in Sect. 4.2) that this linkage may be mostly related to plant hydraulics considerations. The positive relationship between ℓ_A and M_A may reflect constraints on the range of possible combinations of leaf(let) size and M_A , with larger laminar areas necessarily requiring a greater (minimum) M_A due to structural constraints (Grubb, 1998).

Not surprisingly, L_A and Φ_{LS} were related, but with a scaling coefficient of only 0.17, meaning that a greater leaf size was to a substantial degree compensated for by reduced numbers of leaves per unit sapwood area A_S . This points to Φ_{LS} being a relatively invariable trait as has also been reported by others (e.g., Westoby and Wright, 2003). Of note, Φ_{LS} was also correlated with foliar [P] and [N] (Fig. 5), although this correlation was weaker for L_A , especially in the case of foliar phosphorus. But for both nitrogen and phosphorus, the slope was still positive and close to 1.0. Thus tropical tree species with larger leaves tend to have not only higher [P] and [N] (and by implication higher gas exchange rates) but also a higher Φ_{LS} . As there is little evidence of greater diffusional

limitations on gas exchange for such leaves (as shown by the lack of any significant relationship between Φ_{LS} , [N], [P] or L_A with Φ), this implies that accompanying a higher Φ_{LS} are also increased K_S as also observed by Vander Willigen et al. (2000) for subtropical trees and also by Cavender-Bares and Holbrook (2001) for a range of *Quercus* species.

4.1.3 Seed mass

We first note that unlike the other parameters investigated in this study, seed mass has been resolved only at the genus level. This is potentially an issue as there are large genera present in this dataset (e.g. *Pouteria*, *Ocotea* and *Eschweilera*) within which there may be a rather broad variation in seed mass that has the potential to mask causative patterns reported here (C. Baraloto, personal communication, 2011). Nevertheless, as is evident from Fig. 1, seed mass varies by nearly five orders of magnitude which is much greater than the relative variability even in leaf area. Thus, although it must be accepted that any causative relationships may well have been stronger if seed mass had been more accurately determined, where relationships have been found in this data there is little reason to suspect that they are an artifact of our less than ideal species level measurements of S .

Bearing this in mind, we note that, as has been reported by others, seed mass showed significant positive correlations with both H_{max} (Fig. 3; Foster and Janson, 1985; Hammond and Brown, 1995; Kelly, 1995; Metcalfe and Grubb, 1995; Grubb and Coomes, 1997), and ρ_w (Fig. 4; ter Steege and Hammond, 2001), although the latter relationship was not detected by Wright et al. (2006), perhaps because of methodological issues (Williamson and Weimann, 2010). Generally speaking, a greater seed size should confer a greater ability for survival and thus tend to be favoured under less favorable environmental conditions such as deep shade or nutrient poor soils (Westoby et al., 2002; ter Steege et al., 2006). This readily provides a basis for indirect correlations between S and wood/stem density to exist as high values of ρ_x or ρ_w are similarly associated with shade and/or dystrophic soil conditions (Sect. 4.1; Kitajima, 1994). More controversial is the basis of the relationship between S and H_{max} . For example, the suggestion of Moles et al. (2005) that, by analogy with Charnov’s life history theory for mammals, larger statured species may have larger seeds because they require a longer juvenile period has been contested by Grubb et al. (2005) who maintain that it is simply the range of feasible seed sizes that a species can have that increases with H_{max} . Moreover, for tropical trees at least, there is probably little correlation between juvenile period and H_{max} , with faster-growing low-wood density pioneer type trees attaining greater heights than their smaller statured shade counterparts and in a shorter time (Baker et al., 2009). Indeed, by applying a general scaling model Falster et al. (2008) showed that longer juvenile periods alone are not sufficient to generate a correlation between height and seed size. They suggested that size-asymmetric

competition among recruits (i.e. competition for light) may be the main factor having caused evolution towards larger offspring size. In this scheme of things, correlations with adult height come about because larger adults have a greater total reproductive output, thus generating more intense competition among recruits. That model tested dynamics only with a single species at a time, but it is likely to still apply in more complex species systems such as tropical forests, even though relative size at the onset of maturity is much more variable for tropical trees species than for animal systems (Thomas, 1996; Wright et al., 2005). We also consider it unlikely that simple physical constraints can account for much of the relationships (also seen in Fig. 3) as even small statured species can have reasonably large seeds and/or fruits (for example *Theobroma*, or many members of the genus *Licania*: Prance, 1972). Likewise, wind dispersed species have both small seeds and a tendency to occur in the upper canopy strata where higher wind velocities aiding dispersal are greater (Hughes et al., 1994), one obvious example from the Amazon Basin being the widespread neotropical species *Jacaranda copaia* (Jones et al., 2005).

As was also found by Wright et al. (2006), the study gives little support for one of “Corner’s rules”, viz. that due to their mutual dependence on the available supporting twig mass that leaf size and seed size should be positively correlated (Corner, 1949). There may be two reasons for this. First, as pointed out by Grubb et al. (2005) such biomechanical explanations would only be expected to apply where there is little flexibility in the number of fruits per inflorescence. Second, as for Φ_{LS} (Fig. 5) the ratio of total leaf area to the supporting stem mass is to a large degree independent of L_A (Wright et al., 2007). Indeed, if anything, what our data suggest is that reproductive structures compete with leaves for available space as there is a nearly significant correlation between Φ_{LS} and S ($r^2 = -0.09$, $p = 0.07$) with this negative relationship significant for the low fertility species (Supplementary Information, Table 2). Thus, in contrast to vegetation types from more xeric habitats where leaf areas may be substantially constrained by hydraulic considerations, leaf area per unit available stem area or mass may actually be constrained by the requirements for simultaneous allocation of available carbohydrate to reproductive structures for most tropical forest trees. That being consistent with their tropical forest productivity being carbon limited as argued by Lloyd and Farquhar (2008).

Competition between foliage and developing fruit may also be the reason for the negative relationship between seed size and foliar [Ca] shown in Fig. 6, an observation also made for sub-tropical montane tree species by Easdale and Healey (2009). It has long been known that calcium is relatively immobile in plants (e.g., Kirby and Pilbeam, 1984) with high rates of calcium supply to developing fruit essential for cell wall development and for longer term maintenance of membrane integrity. Sufficient levels of calcium are also required to maintain the integrity of the fruit flesh includ-

ing resistance to fungal attack even after abscised from the plant (Bangerth, 1979). Due to its immobility, this calcium accumulation in fruit tissues must occur at the expense of the leaves, and thus Fig. 6 does not necessarily imply that Ca itself may be limiting for either reproductive tissue development or leaf physiological function. Indeed, the SMA slope fit of -8.3 suggests that for each doubling of S foliar [Ca] declines by only about 10%, a value roughly consistent with the similar [Ca] in both seed and leaf tissue (as evidenced from the seed data of Grubb and Coomes (1997)) and with about 0.1 of total South American tropical forest “soft” litterfall occurring as reproductive organs (Chave et al., 2010). Even though such a result does not, therefore, necessarily imply direct effect of Ca availability on tree function, it is interesting to note that species growing on extremely cation poor spodosols are characterised by relatively small seed masses as compared to more fertile nearby forests (Grubb and Coomes, 1997) as well as with leaf photosynthetic rates showing an apparent dependence of leaf calcium concentrations (Reich et al., 1995). Moreover, for forests on such nutrient poor soils, carbon allocation to photosynthetic organs is apparently prioritised over that to reproduction (Chave et al., 2010). This is consistent with neotropical forest reproductive structure frequency being highly sensitive to soil fertility as inferred (apparently) from soil nitrogen status (Gentry and Emmons, 1987), and markedly lower for forests growing on less fertile soils. Overall, these observations suggest, as also discussed in Sect. 4.1.2, that foliar and reproductive tissue development may be in direct competition for either carbon or available nutrients where soil fertility is low.

4.2 Integration of structural and physiological traits

Although an examination of the various bivariate relationships, as discussed in Sect. 4.1 has hopefully proved informative, it is also of additional interest to quantify the extent to which all the various traits examined coordinate in their variability as a whole. In this respect, PCA was considered the most appropriate approach, as the first dimension of a PCA analysis can also be considered (with data normalisations prior to analysis as undertaken here) as the multivariate equivalent of an SMA model fit (Warton et al., 2006). We therefore interpret Table 2 as indicating five discrete integrated trait dimensions of tropical tree function and with the relative importance of these effects varying between high and low fertility species. This interpretation is made even though some of the measured properties such as M_A and ρ_x are modelled as having significant contributions to several dimensions. This is argued as reasonable on two counts. First, variations in some of the traits measured may have different underlying causes. For example, changes in M_A may be a consequence of variations in leaf thickness, tissue density or both (Witkowski and Lamont, 1991; Niinemets, 1999; Poorter et al., 2009) and likewise, variations in ρ_x could reflect differences in the proportions of gas, air and

dry matter content (for hydrated tissue) in a wide range of combinations (Poorter, 2008). Second, as selective pressures are multiple, it is quite likely that contrasting combinations of individual traits have evolved for different reasons.

4.2.1 \bar{A}_{PDJ} : Leaf structural costs and lifespan

Although it is often considered that the primary dimension of the leaf economic spectrum is that proposed by Wright et al. (2004) *viz.* systematic variations in rates of photosynthetic carbon acquisitions (dry weight basis) being linked with foliar dry-weight concentrations of nitrogen, phosphorus, M_A and leaf longevity, our analysis found that U_1 (accounting for the greatest component of the total variation in the dataset) did not involve nitrogen at all, and was actually dominated by leaf cation concentrations and (of opposite sign) a low carbon content. We suggest that this dimension, \bar{A}_{PDJ} , reflects different plant strategies in terms of leaf construction costs, with the tendency for low M_A in these leaves of high mineral content presumably attributable to a low tissue density associated with thinner, less lignified cell walls and with the higher cations content presumably also balanced by higher levels of organic acids (Poorter and de Jong, 1999). Such leaves also being with lower overall construction costs and less investment of phenols and other carbon rich compounds in defense (Poorter and Villar, 1997). Presumably associated with \bar{A}_{PDJ} are also variations in leaf water relations. For example, it seems reasonable to expect that, associated with lower levels of lignification and reduced tissue densities, would be relatively more flexible cell walls and a low bulk modulus of elasticity (Niinemets, 2001), also with the high cation concentrations, especially potassium making a substantial contribution (in association with organic acids) to leaf tissue osmotic potentials (Olivares and Medina, 1992). These attributes, combined with the likely relatively low allocation of carbon resources to defense associated compounds such as lignin and phenols suggests that in many ways leaves of species with high \bar{A}_{PDJ} scores may be able to expand quite rapidly but also be shorter lived and with more “deciduous like” characteristics than their lower \bar{A}_{PDJ} counterparts (see also Sobrado, 1986).

4.2.2 \bar{A}_{RW} : an extension of the classic “leaf economic spectrum”

Our second identified CPC, \bar{A}_{RW} is that usually considered to be the principal dimension of the leaf economic spectrum (Wright et al., 2004), some aspects of which have also been presented for tropical forest tree species (Sandquist and Cordell, 2007; Santiago and Wright, 2007; Fyllas et al., 2009; Baltzer and Thomas, 2010; Baraloto et al., 2010; Domingues et al., 2010). Although we did not measure the photosynthetic or respiratory components, our analysis does suggest that for tropical forest species, L_A should also be included as part of \bar{A}_{RW} , effectively incorporating the relationships between L_A and foliar [N] and/or [P] status

(Fig. 6) within this dimension. Such an involvement of L_A in the classic resource acquisition/utilisation spectrum has also been suggested from a data analysis involving 29 subtropical montane tree species across 12 ha of permanent sample plots in Tucumán, Argentina (Easdale and Healey, 2009). Although not considered significant on the basis of penalty corrected p -values, correlations between leaf size and [N] and [P] of a similar strength to that reported here (and hence included as part of \bar{A}_{RW}) were also reported for tropical forest leaves sampled across a range of soil substrates in French Guiana (Baraloto et al., 2010). They concluded, however, that L_A was not closely linked with either [N] or [P]. This could be for several reasons. First, their sampling strategy covered a range of (undefined) soil types and as discussed in Sect. 4.3, these are likely to have modulated foliar nutrient levels but not L_A . Second, our sampling has covered a much wider range of environments and soils, presumably bringing wider species-level variation into the dataset as whole. Thirdly, our analysis shows the L_A is also an important component of \bar{A}_{FW} . This means that considered in simple bivariate relationships such as with leaf size, relationships may be less clear than when examined in conjunction with additional covariates as done here.

Also identified as part of \bar{A}_{RW} was Φ_{LS} , this being consistent with the general trend of Φ_{LS} to increase with increasing L_A (Fig. 5d). Especially as there was little contribution of \emptyset to this dimension (Table 2), this suggests, other things being equal, that trees with a higher \bar{A}_{RW} should also have increased rates of water transport per unit A_S .

Also of note (though of lesser significance than the above) was the increase in both M_A and [Mg] with decreasing \bar{A}_{RW} . The former is, of course, well documented and, for woody plants at least, seems to be associated with an increased foliar tissue density rather than changes in leaf thickness (Niinemets, 1999; Poorter et al., 2009) and with a concurrent reduction in photosynthetic nutrient efficiency when expressed on a dry weight basis (Niinemets, 1999; Domingues et al., 2010). One possibility to account for this is low internal conductances to CO_2 transfer for higher M_A species (Lloyd et al., 1992; Syvertsen et al., 1995; Warren and Adams, 2006), as perhaps evidenced by a small but significant positive contribution in \emptyset to this dimension (0.014 ± 0.05 ; Table 2). Alternatively, relatively more nitrogen being allocated to cell walls of low \bar{A}_{RW} species (Onoda et al., 2004; Takashima et al., 2004), much of which would be expected to be in the form of defense related proteins (Feng et al., 2009). The decrease in [Mg] with higher values of \bar{A}_{RW} does not seem to have been reported before and may be related to its role as a coordination compound within the chlorophyll molecule. This is because, in the absence of variation in the within-canopy light regime, leaf chlorophyll contents should be relatively conserved on an area (as opposed to mass) basis (Rijkers et al., 2000; Lloyd et al., 2010), this giving rise to attendant reductions in mass based magnesium concentrations as M_A decreases.

4.2.3 \bar{A}_{FW} : tree height and light acquisition

Unlike the previous two dimensions considered, \bar{A}_{FW} does not involve foliar nutrient concentrations, but incorporates into one dimension variations in H_{max} , Φ_{LS} , \varnothing , M_A and to a lesser extent ρ_x . This linkage is most likely through the hydraulics/plant height considerations already discussed as part of Sect. 4.1.1 and 4.1.2. That is to say, as H_{max} increases, a suite of trait adjustments occur; these including a reduction in Φ_{LS} with estimates of \varnothing also suggesting that leaves with a high H_{max} also tend to operate at a lower c_i/c_a . As it seems likely that the higher M_A with increasing H_{max} is mostly attributable to increased leaf/mesophyll thickness and hence increases in photosynthetic capacity per unit leaf area, A_{max} (Sect. 4.1.1), this reduction in c_i/c_a may be attributable to stomatal capacity increasing less with H_{max} than should A_{max} . Such a tendency to operate at a lower c_i/c_a would also help to conserve water for species more likely to be higher-up in the canopy and hence exposed to higher levels of insolation and an associated greater evaporative demand (Lloyd et al., 2010).

Although H_{max} was not determined in their study, many of the above measured and/or inferred traits, *viz.* Φ_{LS} and A_{max} , were found to co-vary in a similar manner as for \bar{A}_{FW} across a range of tropical forest trees in Panama by Meinzer et al. (2008). Though in that case, variations in ρ_x were considered of key importance in terms of trait coordination, especially through linkages to plant hydraulic parameters such as K_S . Our observed contribution of ρ_x is likewise significant (-0.22 ± 0.10), though as discussed in Sect. 4.1.1 taken across a wide range of species and sites the strong relationship between ρ_x and/or ρ_w and K_S as observed by Meinzer et al. (2008) and also in some other studies (e.g., Santiago et al., 2004a) may not necessarily always apply.

Interestingly, in contrast to \bar{A}_{RW} , variations in Φ_{LS} were not accompanied by commensurate changes in L_A . Indeed, if anything, L_A tend to increase with decreasing Φ_{LS} as H_{max} increases within \bar{A}_{FW} . Thus, we find integrated together within \bar{A}_{FW} the tendency of potentially taller trees to have fewer but larger leaves than their more vertically challenged counterparts. But with a lower Φ_{LS} overall. This lower Φ_{LS} presumably serves to help maintain favourable water relations by counteracting greater resistances in the hydraulic pathway for potentially taller trees. Nevertheless, along with a higher \varnothing , this lower Φ_{LS} must also serve to reduce overall rates of whole tree carbon gain such as otherwise might be expected on the basis of higher A_{max} and a greater probability of high levels of incoming radiation. This trade-off associated with a greater H_{max} may be one reason for the observation that light demanding species with a low ρ_w do not necessarily show higher above-ground growth rates than their more shade tolerant counterparts (Keeling et al., 2008).

4.2.4 \bar{A}_{TS} : large seeds at the expense of leaf area

As mentioned in Sect. 4.1.3, a major factor in accounting for this trait dimension is the presence of many large seeded Fabaceae, especially on nutrient poor soils, for whom it turns out do not have as large a Φ_{LS} as they would otherwise be expected to have on the basis of their other trait values. Thus species with a high \bar{A}_{TS} should best be regarded as those having a larger than average seed size with that being associated with a lower than average Φ_{LS} as compared to trees of an equivalent \bar{A}_{FW} and/or \bar{A}_{RW} . This lower Φ_{LS} is also accompanied by reduction in L_A suggesting that it is not so much competition for lateral meristems (Kleiman and Aarssen, 2007) that gives rise to the negative association between Φ_{LS} and S within this dimension. But rather some sort of mechanical constraint such as the total mass capable of being borne per unit stem weight (Westoby and Wright, 2003) or a simple competition for carbon as discussed in Sect. 4.1.2.

The small but significant contributions of M_A , [N] and [P] to \bar{A}_{TS} may be mostly genetic associations as members of the typically large seeded Fabaceae typically have a lower M_A and higher [N] and [P] than members of other plant families (Fyllas et al., 2009). On the other hand, as is discussed in Sect. 4.1.3, the lower foliar [Ca] levels associated with larger seed size is probably functionally linked though high calcium requirements of developing fruits and seeds.

4.2.5 \bar{A}_{PFL} : shade tolerance and long-term viability

The fifth dimension identified, *viz.* \bar{A}_{PFL} , includes a significant positive contribution of increased M_A , presumably associated with a high tissue density (as opposed to leaf thickness) and associated increased leaf toughness (Kitajima and Poorter, 2010) and with high [C] linked through higher than average levels of more reduced structural compounds such as lignin as well as the typically high C-content defense related compounds such as tannins and phenols (Fine et al., 2006; Read and Stokes, 2006; Read et al., 2009). Also associated with this is a higher \varnothing , which may be suggestive of a greater internal resistance to CO_2 diffusion within the leaves of high tissue density woody species (Lloyd et al., 1992; Syvertsen et al., 1995; Warren and Adams, 2006). Interestingly, as well as these correlated leaf traits in this dimension there is the coordinated involvement of a lower H_{max} . Species with strong weightings along this trait dimension are also characterised by larger seeds as would be expected for shade adapted trees (Sect. 4.1.3). Along with a small but significant contribution of ρ_x , \bar{A}_{PFL} is thus strongly suggestive of a coordinated trait dimension associated with shade tolerance and longevity. Not surprisingly then, it seems to play a much greater role in accounting for the trait variations of species associated with low fertility as opposed to high fertility soils as indicated by the different values for the characteristic roots ($\lambda_{low} = 698, \lambda_{high} = 318$, Table 2).

4.2.6 Significance of integrated trait dimensions and their components

Although it is axiomatic that, to be included in any of the above integrated dimensions, a trait would have had to have been measured, what is perhaps more subtle, is that the mix of suites of traits coming together on any one PCA (or CPC) axis is also dependent on what is *not* measured. For example, our differentiation of the first two components of the CPC analysis of Table 2, *viz.* \bar{A}_{PDJ} and \bar{A}_{RW} occurred mainly as consequence of [C] varying positively with M_A for \bar{A}_{PDJ} (this being interpreted as less allocation of photosynthate to more reduced structural compounds such as lignin in low M_A /high cation leaves), but increasing with decreasing M_A within \bar{A}_{RW} (this likely being an effect of higher lipid contents in higher photosynthetic capacity leaves). If leaf [C] is omitted from the analysis, then these two dimensions actually collapse into the one due to the strong correlations between all of the cations, nitrogen and phosphorus and (negatively) M_A (results not shown). This seems likely to have been the case for the results of Easdale and Healey (2009) and Baraloto et al. (2010) where, along with M_A , cations, nitrogen and phosphorus were all considered part of the one dimension.

It is thus clear, that in the presence of additional parameter measurements (for example direct determination of K_S) our derived dimensions may well have been different. Nevertheless, as discussed above, all five identified \bar{A} relate in some way to previously identified trait groupings; though in some cases (as for example with S and M_A in \bar{A}_{PFL}) not previously specifically linked through the species dependent variance-covariance matrix. It would be of great interest to see how the identified trait combinations vary with phylogeny and if they trace back through evolutionary time as discrete combinations. Nevertheless, these ambitions may be confounded by traits such as M_A being significant in almost all dimensions. As discussed at the start of Sect. 4.2.6 this may be because variations in M_A can be surrogates for variations in tissue density, leaf thickness or both, and similarly from the discussion in Sect. 4.1.1 to 4.1.5 above, variations in [C], Φ_{LS} , L_A and \odot are all potentially attributable to a range of different underlying causes. It is also probably for this reason that considerable ambiguity exists between different studies in terms of the significance (or even the sign) of some bivariate relationships. For example, if the primary source of variation in L_A and M_A were to be in association with \bar{A}_{FW} (this being similar in many ways to the light acquisition axis identified by Zhang and Cao (2009) for dipterocarps growing in a Chinese common garden) then a positive association between M_A and L_A would be expected, with leaves of upper canopy trees being both larger and thicker than those for trees lower down in the canopy (as was found to be the case for temperate deciduous trees, for example, by Niinemets, 1998). On the other hand, where foliar N and/or P dry weight concentrations are the main source of variation via \bar{A}_{RW} , then a

negative relationship between M_A and L_A would be expected to be observed as, for example, was found to be the case for a range of herbaceous angiosperms by Shipley (1995). Or, as was found in some cases by Pickup et al. (2005) we can predict that in some circumstances there should be no relationship between L_A and M_A , for example where \bar{A}_{PDJ} is the primary source of variation in the latter (as L_A is effectively absent from this dimension). Indeed, although much touted as a fundamental plant trait (e.g., Poorter et al., 2009; Asner et al., 2011; Kattge et al., 2011) M_A seems to us to be too confounded a measurement to be practically useful in differentiating different plant growth strategies as evidenced by its contribution to the five \bar{A} above. Future work would be better directed towards separate measurements of foliar tissue density and thickness as well as leaf dry matter content (Witkowski and Lamont, 1991; Wilson et al., 1999). It is probably because of its ambiguous nature that M_A does not seem to be as good a predictor of demographic rates as first thought, especially when comparisons are done across different sites (Poorter et al., 2008).

Our results give no support for the supposed “second dimension” of the leaf economics spectrum proposed by Baltzer and Thomas (2010). That study, primarily based on data from Bornean forest trees did, however, fail to differentiate between taxonomic versus soil effects on foliar properties as has been done here. And with their “second dimension” (hardly likely to be orthogonal to the first dimension in any case) most likely simply reflecting soil fertility effects on foliar [P] as already well documented by Fyllas et al. (2009) and considered further below.

4.3 Coordinated trait responses to environmental variability

As evidenced by the 0.3–0.4 portion of the total variance associated with the Φ_{LS} , ρ_x and \odot “plot effect” terms (Fig. 2), values of all these traits are not independent of where a species is growing and with there being strong environmental correlations between ρ_x and all of $\log[\text{N}]$, $\log[\text{P}]$, $\log[\text{Ca}]$ and $\log[\text{K}]$. This results in this structural parameter aligning itself along with elemental concentrations (including [C] with a negative weighting) in the first environmental PCA axis, η_1 (Table 4), which was itself closely correlated with a PCA of soil chemical and physical properties (Fyllas et al., 2009; Quesada et al., 2010). This dimension, relating to what seems to be a soil fertility mediated effect, bears some resemblance to \bar{A}_{RW} but with a more easily discernible effect on ρ_x . As mentioned in Sect. 4.1.1, such a fertility effect on ρ_x has been seen before as mediated by soil phosphorus availability for eucalypt and mangrove (Thomas et al., 2005; Lovelock et al., 2006). Although working with Brazilian savanna trees, Bucci et al. (2006) found it was nitrogen (as opposed to phosphorus) fertilisation that induced changes in ρ_x and K_S and in their case with N-fertilisation causing attendant increases in Φ_{LS} not detected here (Table 4).

It seems likely that higher foliar [P], especially in combination with the lower M_A also associated with η_1 would give rise to higher photosynthetic rates on an area basis (Domingues et al., 2010; Mercado et al., 2011). Thus, with tropical forest tree hydraulics and photosynthetic capacity being closely linked (Brodribb and Field, 2000; Brodribb et al., 2002; Santiago et al., 2004a) the likely increase in K_S accompanying a decrease in ρ_x with improved nutrient status may serve to help maintain some homeostasis in leaf water relations, this offsetting the higher rates of water-use per leaf area that would be expected to accompany any increase in η_1 . This suggestion supported by the only modest contribution of ϕ to this dimension (Table 4). As to how such a coordination could occur is currently not clear, although the greater rates of cambial activity in the wood of higher P status trees giving rise to a lower ρ_x might be attributable through sugar signalling mechanisms (Rolland et al., 2006; Hölttä et al., 2010), this resulting in less secondary thickening of vessels walls and a higher conduit area (Thomas et al., 2005). Other elements may also be involved though, for example effects of calcium and/or potassium on sapwood cambial activity (Fromm, 2010).

The second integrated environmental response dimension identified, η_2 , essentially represents an integration of previous observed foliar trait responses to precipitation, *viz.* increased M_A , [C] and ϕ and decreased [Mg] as mean annual precipitation increases as detailed in Fyllas et al. (2009). Although this response to P_A seems at odds with the general observation from inter-species analyses that leaves of more arid environments should have a higher M_A and often with a higher ϕ (Miller et al., 2001; Santiago et al., 2004b) as discussed by Fyllas et al. (2009) this tendency towards more structurally rigid leaves at higher P_A may reflect different populations of the same species having different characteristics according to their prevailing environment. An aligned interpretation is that as severe dry season water deficits become increasingly less of a driving force in determining leaf lifetimes, leaves of any given species become more “evergreen” in their structural characteristics. And indeed it is worth noting that the distinction between evergreen and deciduous phenologies for tropical forest trees is a somewhat arbitrary one (Brodribb and Holbrook, 2005; Williams et al., 2008). In such an interpretation, an increase in ϕ with P_A could be interpreted as either a tendency towards more conservative stomatal behavior in evergreen species where the precipitation regime is not strongly seasonal (Lloyd and Farquhar, 1994) or, alternatively to an increased resistance to CO₂ diffusion within higher M_A leaves due to a higher cell wall resistance (Syvertsen et al., 1995).

Although not emerging as any sort of integrated response through the PCA analysis of the derived environmental effects, the temperature responses of [N], [K] and ρ_x are all also of note; these have already been considered separately by Fyllas et al. (2009) and Patiño et al. (2009).

5 Conclusions

Extending beyond a simple bivariate analysis approach, this study has separated environmental from taxonomic effects for a range of structural and physiological traits for Amazon forest trees then using Common Principal Component Analysis to reveal as many as five discrete integrated axes of taxonomic variation. The relative weightings of the axes varies between low and high fertility soil associated species. The first component (accounting for the highest proportion of the total variance in the dataset) was not the classic “leaf economic spectrum”, but rather relates mostly to variations in leaf construction costs per unit dry weight. The leaf economic spectrum was the second most important dimension identified in terms of variance accounted for, with our results suggesting that it also involves differences in leaf size as well as in leaf area: sapwood area ratios. Our third dimension brings together several structural traits, including species specific maximum height, individual leaf areas, leaf mass per unit area and xylem density and leaf magnesium concentrations. The fourth and fifth dimensions were interpreted as relating to a seed size/leaf area trade-off and shade tolerance characteristics respectively.

Several traits, in particular leaf mass per unit area, foliar carbon content and xylem density had significant weighting on many axes of variation, this being attributed to their somewhat ambiguous “proxy” nature for a range of underlying and more fundamental plant physiological properties. In particular, variations in twig xylem density may arise as a consequence of differences in a range of different underlying phenomena and with its generally poor correlation with other plant traits suggesting that it may not be as good a proxy for plant hydraulic conductivity as once thought.

Significant effects of environment on many plant traits were also identified. Some of these integrated into discrete dimensions of variation and with discrete but different changes being associated with variations in soil fertility versus differences in mean annual precipitation. Whether these differences relate to strict “environmental effects” or reflect systematic patterns in intra-specific trait variation with soils and/or climate remains to be established.

Supplementary material related to this article is available online at:

<http://www.biogeosciences.net/9/775/2012/bg-9-775-2012-supplement.pdf>.

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SUPPLEMENTARY INFORMATION TO:

Coordination of physiological and structural traits in Amazon forest trees

by Sandra Patiño, Nikolaos M. Fyllas, Timothy R. Baker, Romilda Paiva, Carlos Alberto Quesada, Alexandre J. B. Santos, Michael Schwarz, Hans ter Steege, Oliver Phillips and Jon Lloyd*

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Table S5: Full results from the CPC analysis (all eigenvectors)

Table S6: Covariance (\mathbf{F}) and correlation (\mathbf{R}) matrices of CPCs.

Figure S1. Summary of CPC analysis results (First three axes)

* To whom correspondence should be addressed: j.lloyd@leeds.ac.uk

Table S1: Mean value of each foliar property per plot. Seed class input data was in \log_{10} steps [log classes: 1 = 10^{-5} - 10^{-4} , 2 = 10^{-4} - 10^{-3} ,.....8 = >100] as per ter Steege and Hammond (2001)

Plot	lat	long	L_A ($\times 10^3$ m 2)	ℓ_a ($\times 10^3$ m 2)	Φ_{LS} (m 2 cm $^{-2}$)	ρ_x (kg m $^{-3}$)	\dot{O}	Seed Class	H_{max} (m)	M_A (g m $^{-2}$)	C (mg g $^{-1}$)	N (mg g $^{-1}$)	P (mg g $^{-1}$)	Ca (mg g $^{-1}$)	K (mg g $^{-1}$)	Mg (mg g $^{-1}$)
AGP-01	-3.74	-70.31	26.8 \pm 45.4	13.1 \pm 10.1	1.23 \pm 0.79	569 \pm 69	0.13 \pm 0.03	5.5 \pm 1.1	27.9 \pm 7.0	103 \pm 31	455 \pm 41	20.9 \pm 4.2	1.06 \pm 0.38	9.8 \pm 6.0	9.1 \pm 6.0	2.8 \pm 1.7
AGP-02	-3.74	-70.30	26.7 \pm 45.5	11.7 \pm 8.1	0.91 \pm 0.46	560 \pm 80	0.13 \pm 0.03	5.6 \pm 0.8	30.9 \pm 7.8	96 \pm 19	457 \pm 29	19.2 \pm 3.4	0.96 \pm 0.24	9.9 \pm 4.3	6.8 \pm 2.6	2.8 \pm 1.8
ALP-11	-3.95	-73.43	2.4 \pm 0.0	4.9 \pm 3.2	ND	630 \pm 64	0.22 \pm 0.05	4.6 \pm 0.9	32.0 \pm 10.9	140 \pm 51	525 \pm 31	17.6 \pm 4.8	0.84 \pm 0.32	7.3 \pm 2.3	6.1 \pm 3.7	2.1 \pm 0.6
ALP-12	-3.95	-73.44	6.2 \pm 3.4	6.2 \pm 3.4	ND	644 \pm 64	0.22 \pm 0.05	5.2 \pm 0.8	32.7 \pm 3.0	124 \pm 19	513 \pm 16	20.3 \pm 4.7	0.78 \pm 0.19	6.9 \pm 2.5	4.9 \pm 1.6	1.6 \pm 0.7
ALP-21	-3.95	-73.44	3.4 \pm 1.4	3.3 \pm 1.5	ND	720 \pm 135	0.21 \pm 0.05	5.0 \pm 0.6	28.4 \pm 11.6	134 \pm 27	524 \pm 19	20.4 \pm 6.2	0.98 \pm 0.31	4.6 \pm 2.9	4.1 \pm 0.9	1.7 \pm 0.6
ALP-22	-3.95	-73.44	9.6 \pm 7.5	4.8 \pm 4.6	ND	670 \pm 74	0.20 \pm 0.03	5.3 \pm 0.8	31.9 \pm 3.8	95 \pm 23	497 \pm 24	21.2 \pm 5.4	1.02 \pm 0.19	7.6 \pm 4.1	6.7 \pm 2.5	1.8 \pm 0.8
ALP-30	-3.95	-73.43	19.9 \pm 23.8	3.2 \pm 2.3	ND	779 \pm 50	0.19 \pm 0.05	5.2 \pm 0.6	29.7 \pm 7.5	115 \pm 26	527 \pm 18	20.3 \pm 4.6	1.02 \pm 0.45	6.6 \pm 3.3	3.5 \pm 1.0	1.9 \pm 0.8
BNT-04	-2.63	-60.15	10.6 \pm 11.1	6.5 \pm 3.4	0.94 \pm 0.43	730 \pm 47	0.16 \pm 0.03	5.7 \pm 0.8	28.0 \pm 7.7	103 \pm 25	491 \pm 16	19.9 \pm 4.3	0.54 \pm 0.15	2.2 \pm 1.2	3.2 \pm 0.9	1.3 \pm 0.4
BOG-01	-0.70	-76.48	29.3 \pm 51.7	7.8 \pm 5.6	1.37 \pm 1.12	526 \pm 96	0.19 \pm 0.04	4.8 \pm 0.9	27.4 \pm 10.4	91 \pm 26	455 \pm 43	25.3 \pm 5.8	1.61 \pm 0.50	13.2 \pm 8.0	9.2 \pm 4.0	2.6 \pm 1.2
BOG-02	-0.70	-76.47	41.9 \pm 126.3	15.4 \pm 25.8	1.12 \pm 0.64	546 \pm 121	0.20 \pm 0.04	4.8 \pm 0.9	29.4 \pm 6.7	95 \pm 24	439 \pm 50	23.0 \pm 5.3	1.44 \pm 0.46	16.2 \pm 11.4	13.0 \pm 6.9	3.6 \pm 3.0
BRA-01	-0.83	-46.64	5.2 \pm 1.8	5.2 \pm 1.8	1.54 \pm 1.25	761 \pm 175	0.12 \pm 0.03	5.4 \pm 1.0	33.3 \pm 7.0	129 \pm 16	409 \pm 15	19.6 \pm 4.7	1.23 \pm 0.47	8.3 \pm 2.9	11.3 \pm 3.6	7.4 \pm 2.9
CAX-01	-1.74	-51.46	17.9 \pm 21.1	6.0 \pm 2.5	1.05 \pm 0.58	641 \pm 162	0.15 \pm 0.05	5.5 \pm 0.8	31.6 \pm 6.8	82 \pm 19	464 \pm 32	23.8 \pm 6.3	0.60 \pm 0.17	3.1 \pm 1.9	2.9 \pm 2.2	3.2 \pm 1.3
CAX-02	-1.74	-51.46	13.0 \pm 16.3	6.0 \pm 2.8	1.35 \pm 1.03	784 \pm 112	0.14 \pm 0.04	5.8 \pm 0.9	29.6 \pm 9.4	85 \pm 19	465 \pm 24	22.1 \pm 3.8	0.74 \pm 0.13	5.8 \pm 4.3	2.3 \pm 1.0	3.1 \pm 1.7
CUZ-03	-12.50	-68.96	17.5 \pm 8.7	9.1 \pm 5.3	1.16 \pm 0.51	585 \pm 151	0.12 \pm 0.04	5.9 \pm 1.2	26.3 \pm 8.4	88 \pm 26	438 \pm 31	21.8 \pm 4.8	1.68 \pm 0.68	14.1 \pm 4.9	11.7 \pm 4.5	2.5 \pm 1.0
ELD-12	6.10	-61.40	29.9 \pm 32.0	10.2 \pm 5.4	1.23 \pm 0.42	523 \pm 124	0.14 \pm 0.05	5.3 \pm 0.9	28.2 \pm 5.0	80 \pm 16	491 \pm 18	20.5 \pm 4.3	0.66 \pm 0.18	4.5 \pm 3.0	5.8 \pm 3.4	2.4 \pm 1.1
HCC-21	-14.56	-60.75	16.7 \pm 13.6	12.5 \pm 15.3	0.66 \pm 0.29	557 \pm 121	0.21 \pm 0.04	4.4 \pm 1.0	27.8 \pm 7.6	76 \pm 22	484 \pm 24	30.7 \pm 3.8	1.03 \pm 0.16	12.3 \pm 3.9	8.9 \pm 2.6	2.7 \pm 1.0
HCC-22	-14.57	-60.75	25.3 \pm 19.8	9.1 \pm 3.9	0.62 \pm 0.35	696 \pm 64	0.16 \pm 0.03	5.2 \pm 0.8	30.8 \pm 8.2	90 \pm 22	439 \pm 45	28.8 \pm 6.1	1.32 \pm 0.45	15.7 \pm 8.6	13.0 \pm 9.3	4.8 \pm 2.9
JAC-04	-2.61	-60.22	18.2 \pm 14.6	9.8 \pm 6.8	0.88 \pm 0.65	671 \pm 84	0.17 \pm 0.04	5.4 \pm 0.6	32.0 \pm 9.9	127 \pm 27	480 \pm 38	12.6 \pm 4.0	0.48 \pm 0.10	3.1 \pm 1.9	5.2 \pm 3.5	2.5 \pm 1.5
JAC-12	-2.61	-60.21	15.0 \pm 25.8	7.9 \pm 10.0	0.56 \pm 0.28	537 \pm 96	0.22 \pm 0.03	4.8 \pm 1.0	31.4 \pm 9.5	103 \pm 25	491 \pm 28	22.3 \pm 5.8	0.60 \pm 0.18	2.6 \pm 1.5	3.7 \pm 1.8	1.9 \pm 1.0
JAS-02	-1.07	-77.62	18.0 \pm 36.1	6.0 \pm 3.3	1.14 \pm 1.00	531 \pm 75	0.21 \pm 0.03	4.7 \pm 1.0	37.5 \pm 8.4	113 \pm 35	482 \pm 35	23.3 \pm 5.1	1.03 \pm 0.28	7.9 \pm 3.9	9.5 \pm 4.9	2.0 \pm 0.8
JAS-03	-1.08	-77.61	9.9 \pm 5.1	9.9 \pm 5.1	0.89 \pm 0.14	540 \pm 82	0.22 \pm 0.04	5.1 \pm 0.8	31.8 \pm 7.5	103 \pm 36	470 \pm 39	24.5 \pm 6.7	1.24 \pm 0.45	11.8 \pm 5.9	9.9 \pm 4.8	2.3 \pm 1.2

Plot	lat	long	L_A	ℓ_a	Φ_{LS}	ρ_x	\hat{O}	Seed	H_{max}	M_A	C	N	P	Ca	K	Mg
			($\times 10^3$ m 2)	($\times 10^3$ m 2)	(m 2 cm $^{-2}$)	(kg m $^{-3}$)		Class	(m)	(g m $^{-2}$)	(mg g $^{-1}$)	(mg g $^{-1}$)	(mg g $^{-1}$)	(mg g $^{-1}$)	(mg g $^{-1}$)	(mg g $^{-1}$)
JAS-04	-1.07	-77.61	23.3 \pm 25.3	7.8 \pm 5.2	1.33 \pm 1.49	465 \pm 70	0.22 \pm 0.03	4.6 \pm 0.8	32.1 \pm 6.4	120 \pm 34	500 \pm 33	18.9 \pm 3.3	0.83 \pm 0.27	7.3 \pm 2.8	6.2 \pm 4.6	1.6 \pm 0.8
JAS-05	-1.06	-77.62	5.2 \pm 3.6	4.5 \pm 3.0	0.93 \pm 0.40	776 \pm 115	0.22 \pm 0.05	5.0 \pm 0.7	25.0 \pm 7.1	94 \pm 21	482 \pm 33	27.0 \pm 4.9	2.10 \pm 0.71	12.6 \pm 6.8	11.5 \pm 5.0	2.7 \pm 1.4
JRI-01	-0.89	-52.19	0.0 \pm 0.0	0.0 \pm 0.0	ND	ND	0.18 \pm 0.05	5.3 \pm 0.4	30.8 \pm 8.0	120 \pm 29	483 \pm 19	17.9 \pm 4.6	0.57 \pm 0.14	5.0 \pm 2.9	3.4 \pm 2.2	2.2 \pm 0.7
JUR-01	-8.88	-72.79	8.8 \pm 7.2	8.8 \pm 7.2	1.06 \pm 0.94	555 \pm 102	0.14 \pm 0.04	4.3 \pm 1.1	32.7 \pm 12.1	97 \pm 32	396 \pm 28	25.3 \pm 6	1.56 \pm 0.27	11.4 \pm 2.9	8.9 \pm 3.8	3.3 \pm 0.1
LFB-01	-14.56	-60.93	8.1 \pm 6.3	6.0 \pm 2.6	1.07 \pm 0.32	601 \pm 71	0.16 \pm 0.03	4.5 \pm 1.9	29.9 \pm 10.0	79 \pm 24	449 \pm 28	22.5 \pm 4.4	0.90 \pm 0.37	5.5 \pm 1.6	10.6 \pm 4.6	2.7 \pm 1.0
LFB-02	-14.58	-60.83	43.4 \pm 83.6	10.5 \pm 4.1	1.18 \pm 0.74	621 \pm 35	0.14 \pm 0.05	6.0 \pm 0.5	23.3 \pm 10.5	96 \pm 19	468 \pm 34	20.7 \pm 4.6	0.78 \pm 0.23	4.2 \pm 1.7	6.3 \pm 2.3	2.0 \pm 0.8
LOR-01	-3.06	-69.99	28.4 \pm 53.9	10.0 \pm 5.2	1.20 \pm 0.74	562 \pm 125	0.13 \pm 0.03	5.5 \pm 0.8	30.0 \pm 6.3	101 \pm 21	488 \pm 18	19.3 \pm 3.4	1.07 \pm 0.41	4.8 \pm 2.6	6.9 \pm 3.0	2.2 \pm 1.0
LOR-02	-3.06	-69.99	11.0 \pm 8.6	3.9 \pm 1.8	0.70 \pm 0.40	638 \pm 97	0.18 \pm 0.05	3.5 \pm 2.2	26.9 \pm 11.6	93 \pm 20	472 \pm 29	21.3 \pm 6.0	1.14 \pm 0.47	5.5 \pm 2.6	8.8 \pm 6.1	2.3 \pm 1.0
LSL-01	-14.41	-61.14	3.1 \pm 0.9	3.1 \pm 0.9	0.89 \pm 0.42	547 \pm 63	0.17 \pm 0.03	3.3 \pm 2.7	30.7 \pm 7.3	77 \pm 20	483 \pm 15	23.9 \pm 4.6	1.18 \pm 0.41	6.5 \pm 4.1	7.8 \pm 2.9	2.2 \pm 1.0
LSL-02	-14.41	-61.14	11.9 \pm 12.9	7.4 \pm 6.1	1.01 \pm 0.42	646 \pm 106	0.18 \pm 0.02	5.3 \pm 0.7	33.9 \pm 7.5	73 \pm 20	455 \pm 31	20.1 \pm 2.8	1.06 \pm 0.22	6.6 \pm 2.6	8.1 \pm 4.8	2.3 \pm 1.0
MBO-01	-1.45	-48.45	15.2 \pm 22.0	5.9 \pm 2.6	0.81 \pm 0.44	586 \pm 58	ND	4.8 \pm 0.9	33.4 \pm 9.7	90 \pm 20	474 \pm 28	21.4 \pm 5.2	0.74 \pm 0.16	2.9 \pm 1.7	3.9 \pm 1.1	2.3 \pm 1.1
RES-04	-10.80	-68.77	82.8 \pm 25.9	11.0 \pm 6.3	1.63 \pm 0.68	555 \pm 98	ND	5.3 \pm 0.7	31.5 \pm 10.0	93 \pm 30		19.5 \pm 4.1				
RES-05	-10.57	-68.31	37.1 \pm 48.9	7.7 \pm 6.3	1.36 \pm 0.80	611 \pm 80	0.18 \pm 0.05	4.8 \pm 0.6	31.4 \pm 12.4	85 \pm 20	424 \pm 45	22.6 \pm 4.7	0.95 \pm 0.21	13.2 \pm 10.2	7.6 \pm 5.0	3.4 \pm 1.5
RES-06	-10.56	-68.30	6.6 \pm 5.3	6.3 \pm 5.2	1.07 \pm 0.82	544 \pm 94	0.17 \pm 0.05	4.8 \pm 1.1	26.4 \pm 9.0	85 \pm 23	436 \pm 46	23.2 \pm 5.3	1.20 \pm 0.38	14.9 \pm 10.5	9.0 \pm 4.2	4.5 \pm 3.3
RIO-12	8.11	-61.69	17.3 \pm 22.7	9.2 \pm 5.2	1.08 \pm 0.59	603 \pm 90	0.14 \pm 0.03	5.8 \pm 0.8	33.6 \pm 6.9	96 \pm 27	455 \pm 42	16.6 \pm 5.4	0.59 \pm 0.16	7.4 \pm 7.7	4.4 \pm 3.1	3.4 \pm 1.9
SCR-01	1.93	-67.02	18.9 \pm 12.5	4.4 \pm 0.3	0.55 \pm 0.33	641 \pm 139	0.17 \pm 0.05	7.0 \pm 0.0	25.4 \pm 3.2	124 \pm 25	504 \pm 11	15.3 \pm 2.9	1.40 \pm 1.04	1.1 \pm 1.0	9.5 \pm 7.8	1.0 \pm 0.3
SCR-04	1.93	-67.04	16.3 \pm 15.6	6.2 \pm 2.3	0.15 \pm 0.33	562 \pm 84	0.17 \pm 0.04	5.5 \pm 1.1	28.7 \pm 7.6	141 \pm 39	512 \pm 15	12.5 \pm 5.2	1.03 \pm 0.53	5.2 \pm 3.2	5.7 \pm 1.9	1.9 \pm 0.8
SCR-05	1.93	-67.04	9.3 \pm 4.8	5.8 \pm 2.5	0.99 \pm 0.48	598 \pm 115	0.17 \pm 0.04	5.7 \pm 1.0	31.8 \pm 10.2	155 \pm 55	540 \pm 18	16.1 \pm 4.1	0.64 \pm 0.16	1.4 \pm 0.8	5.1 \pm 1.5	1.0 \pm 0.4
SUC-01	-3.25	-72.91	18.0 \pm 28.5	16.2 \pm 27.1	ND	636 \pm 122	0.16 \pm 0.05	5.2 \pm 1.3	33.1 \pm 11.1	127 \pm 56	510 \pm 38	19.9 \pm 4.3	0.90 \pm 0.28	5.8 \pm 2.5	6.3 \pm 3.1	2.5 \pm 1.6
SUC-02	-3.25	-72.90	28.8 \pm 36.9	18.8 \pm 29.4	ND	659 \pm 90	0.19 \pm 0.05	5.4 \pm 1.1	29.5 \pm 9.7	113 \pm 29	493 \pm 30	18.2 \pm 5.5	0.90 \pm 0.21	5.4 \pm 3.3	6.5 \pm 2.5	2.0 \pm 0.8
SUC-03	-3.25	-72.92	10.7 \pm 14.0	5.9 \pm 6.0	ND	698 \pm 128	0.17 \pm 0.05	5.5 \pm 0.9	30.8 \pm 10.8	109 \pm 28	495 \pm 24	18.5 \pm 2.8	0.91 \pm 0.20	8.7 \pm 4.4	5.2 \pm 1.9	2.8 \pm 1.6

Plot	lat	long	L_A ($\times 10^3 \text{ m}^2$)	ℓ_a ($\times 10^3 \text{ m}^2$)	Φ_{LS} ($\text{m}^2 \text{ cm}^{-2}$)	ρ_x (kg m^{-3})	\dot{O}	Seed Class	H_{\max} (m)	M_A (g m^{-2})	C (mg g^{-1})	N (mg g^{-1})	P (mg g^{-1})	Ca (mg g^{-1})	K (mg g^{-1})	Mg (mg g^{-1})
SUM-01	-1.75	-77.63	49.7±68.8	11.2±6.2	1.28±0.79	501±77	0.25±0.03	4.8±0.9	35.9±8.9	104±33	476±21	25.3±7.4	1.61±0.50	7.6±2.6	8.2±2.7	2.0±0.9
TAM-01	-12.84	-69.29	15.0±15.8	6.9±5.7	1.35±0.69	579±96	0.21±0.03	5.2±0.4	34.4±7.7	100±21	471±32	22.1±5.1	1.20±0.37	6.6±2.6	8.4±2.9	1.9±0.8
TAM-02	-12.83	-69.29	18.4±21.4	8.1±7.3	1.03±0.39	625±113	0.21±0.04	5.0±0.7	32.8±8.9	104±27	475±37	22.9±5.5	1.16±0.42	5.0±3.1	9.0±4.6	3.1±2.2
TAM-03	-12.84	-69.28	7.0±4.1	7.0±4.1	0.81±0.35	468±98	0.18±0.03	3.9±1.4	33.1±6.4	110±15	485±35	18.8±4.3	1.31±0.33	4.2±1.4	8.3±4.8	1.9±1.4
TAM-04	-12.84	-69.28	10.8±7.8	10.8±7.8	0.97±0.52	595±94	0.20±0.03	4.7±1.3	32.0±8.6	115±25	496±28	21.7±4.7	1.31±0.34	2.2±0.9	7.5±3.0	2.1±0.9
TAM-05	-12.83	-69.27	11.1±11.3	5.8±5.3	1.10±0.41	637±86	0.21±0.05	5.2±0.8	32.4±7.5	101±24	507±34	24.0±5.5	1.05±0.20	2.8±2.5	6.1±3.2	2.2±1.4
TAM-06	-12.84	-69.30	13.6±20.1	6.5±7.9	0.95±0.27	585±82	0.21±0.03	5.1±1.0	34.1±7.6	96±21	485±35	24.8±6.7	1.88±0.84	8.4±4.3	8.2±3.5	2.3±1.0
TAM-07	-12.83	-69.26	19.9±32.0	6.6±5.1	1.01±0.76	629±99	0.19±0.03	5.0±1.4	30.3±9.1	114±22	511±33	21.6±5.0	0.98±0.23	1.9±0.8	6.4±2.3	2.2±0.9
TAP-04	-2.85	-54.95	32.2±66.9	9.3±5.7	1.01±0.63	631±144	0.17±0.04	4.8±1.0	29.9±6.3	99±31	463±43	22.6±7.7	0.75±0.24	7.5±4.2	3.7±2.2	2.7±1.4
TAP-123	-3.31	-54.94	12.2±29.3	5.6±3.6	1.42±0.82	692±97	0.19±0.04	4.8±0.8	27.4±15.7	90±19	467±35	21.7±5.6	0.70±0.18	5.2±3.0	3.0±1.2	2.5±1.1
TIP-03	-0.64	-76.15	9.5±9.2	4.3±3.2	0.71±0.29	565±34	0.18±0.04	4.8±0.5	31.5±8.8	92±48	472±26	28.4±3.3	2.04±0.65	7.3±3.6	7.0±3.1	2.2±1.1
TIP-05	-0.64	-76.14	44.2±67.7	10.5±8.8	1.02±0.43	578±116	0.19±0.05	4.9±0.7	37.7±9.1	112±40	470±32	21.8±3.8	1.23±0.39	11.5±4.9	8.2±4.3	3.2±0.8
YAN-01	-3.44	-72.85	20.5±33.6	6.2±3.5	ND	570±97	0.22±0.04	4.5±1.1	33.7±10.5	86±29	474±36	19.7±5.3	1.24±0.33	19.1±10.7	9.1±4.3	3.2±1.5
YAN-02	-3.43	-72.84	13.6±8.0	13.6±8.0	ND	508±58	0.18±0.03	4.5±1.3	43.5±8.4	105±14	463±43	20.0±3.7	1.2±0.4	17.5±9.5	8.9±2.8	4.2±1.0

Table S2A: Pair-wise relationships of key foliar properties using the raw data (left panel), pooled genetic (middle panel) and pooled plot-environmental effects (right panel). For the raw data the intercept and slope are given, but for both the genetic and environmental effects only the slope is presented as all relationships were forced through (0,0). Also given are the 0.95 quantile confidence intervals, Pearson's r correlation coefficient, the significance of the correlation (sig), and n the number of cases used. The (NS) sign indicates that the respective estimates could not be made. The genetic and plot effects lines are forced through (0,0).

		<u>raw data</u>									<u>genetic effects</u>					<u>plot-environmental effects</u>						
y	x	intercept	intercept		slope	slope		r	sig	n	slope	slope 0.95 ci		r	sig	n	slope	slope 95%ci		r	sig	n
			low	high		low	high					low	high					low	high			
$\log(L_A)$	$\log(\ell_d)$	0.92	0.76	1.07	1.32	1.25	1.39	0.62	0.000	870	1.53	1.40	1.66	0.41	0.000	455	1.18	1.05	1.32	0.90	0.000	61
$\log(L_A)$	$\log(\Phi_{LS})$	-1.92	-1.96	-1.87	1.44	1.34	1.55	0.15	0.000	683	5.26	4.77	5.79	0.26	0.000	388	NS	NS	NS	0.07	0.621	50
$\log(L_A)$	ρ_x	0.23	0.08	0.38	-3.69	-3.46	-3.94	-0.19	0.000	870	-7.62	-6.95	-8.35	-0.10	0.033	456	-1.49	-1.15	-1.91	-0.25	0.059	60
$\log(L_A)$	\hat{O}	NS	NS	NS	NS	NS	NS	-0.02	0.548	813	28.58	26.04	31.36	0.11	0.019	443	NS	NS	NS	-0.14	0.272	60
$\log(L_A)$	$\log(S)$	NS	NS	NS	NS	NS	NS	NS	0.171	815	NS	NS	NS	NS	0.672	420	NS	NS	NS	NA	NA	NA
$\log(L_A)$	H_{max}	NS	NS	NS	NS	NS	NS	NS	0.128	863	NS	NS	NS	NS	0.662	448	NS	NS	NS	NA	NA	NA
$\log(L_A)$	$\log(M_A)$	NS	NS	NS	NS	NS	NS	NS	0.442	855	NS	NS	NS	NS	0.068	453	NS	NS	NS	0.08	0.546	61
$\log(L_A)$	[C]	NS	NS	NS	NS	NS	NS	NS	0.709	826	0.02	0.02	0.02	0.13	0.006	444	NS	NS	NS	-0.08	0.563	60
$\log(L_A)$	$\log[N]$	-7.03	-7.37	-6.69	3.80	3.55	4.06	0.12	0.000	837	6.38	5.83	6.97	0.27	0.000	449	NS	NS	NS	-0.09	0.467	61
$\log(L_A)$	$\log[P]$	-1.94	-1.98	-1.90	2.29	2.14	2.44	0.18	0.000	833	5.39	4.95	5.88	0.37	0.000	445	NS	NS	NS	-0.03	0.815	60
$\log(L_A)$	$\log[Ca]$	-2.94	-3.01	-2.86	1.23	1.15	1.31	0.06	0.109	834	NS	NS	NS	NS	0.619	446	NS	NS	NS	0.11	0.421	60
$\log(L_A)$	$\log[K]$	-3.07	-3.15	-2.99	1.42	1.33	1.52	0.11	0.001	834	NS	NS	NS	NS	0.819	446	NS	NS	NS	0.21	0.111	60
$\log(L_A)$	$\log[Mg]$	NS	NS	NS	NS	NS	NS	0.00	0.979	834	-2.99	-2.72	-3.27	-0.15	0.002	446	NS	NS	NS	0.07	0.586	60
$\log(\ell_d)$	$\log(\Phi_{LS})$	-2.17	-2.20	-2.13	1.01	0.94	1.09	0.12	0.002	693	NS	NS	NS	0.01	0.905	392	NS	NS	NS	0.08	0.582	50
$\log(\ell_d)$	ρ_x	-0.51	-0.62	-0.39	-2.83	-2.65	-3.02	-0.22	0.000	888	-5.03	-4.60	-5.50	-0.22	0.000	464	-1.26	-0.99	-1.62	-0.31	0.015	60
$\log(\ell_d)$	\hat{O}	-0.93	-1.03	-0.84	-7.34	-6.86	-7.86	-0.12	0.001	825	NS	NS	NS	-0.08	0.110	449	-2.84	-2.21	-3.64	-0.28	0.027	60
$\log(\ell_d)$	$\log(S)$	NS	NS	NS	NS	NS	NS	0.05	0.166	831	NS	NS	NS	-0.01	0.842	426	NS	NS	NS	NA	NA	NA
$\log(\ell_d)$	H_{max}	NS	NS	NS	NS	NS	NS	0.00	0.999	881	NS	NS	NS	0.00	0.928	456	NS	NS	NS	NA	NA	NA
$\log(\ell_d)$	$\log(M_A)$	-7.66	-8.02	-7.30	2.74	2.56	2.93	0.09	0.007	869	4.34	3.97	4.75	0.17	0.000	459	NS	NS	NS	0.07	0.604	61
$\log(\ell_d)$	[C]	1.97	1.69	2.26	-0.01	-0.01	-0.01	-0.12	0.001	838	NS	NS	NS	-0.08	0.086	450	NS	NS	NS	-0.19	0.145	60
$\log(\ell_d)$	$\log[N]$	NS	NS	NS	NS	NS	NS	-0.03	0.445	851	-4.20	-3.84	-4.61	-0.11	0.021	455	NS	NS	NS	-0.16	0.214	61
$\log(\ell_d)$	$\log[P]$	NS	NS	NS	NS	NS	NS	0.05	0.162	845	NS	NS	NS	0.04	0.456	451	NS	NS	NS	-0.11	0.394	60
$\log(\ell_d)$	$\log[Ca]$	NS	NS	NS	NS	NS	NS	0.06	0.096	846	NS	NS	NS	0.06	0.215	452	NS	NS	NS	0.09	0.496	60
$\log(\ell_d)$	$\log[K]$	-3.04	-3.11	-2.98	1.10	1.03	1.18	0.16	0.000	846	2.62	2.39	2.87	0.18	0.000	452	NS	NS	NS	0.20	0.132	60
$\log(\ell_d)$	$\log[Mg]$	-2.78	-2.83	-2.73	1.56	1.45	1.66	0.13	0.000	846	1.98	1.80	2.17	0.14	0.003	452	NS	NS	NS	0.15	0.256	60

Table S2A (continued)

		raw data									genetic effects					plot-environmental effects						
y	x	intercept	intercept		slope	slope		r	sig	n	slope	slope		r	sig	n	slope	slope 0.95 c.i.		r	sig	n
			0.95 ci	low		high	0.95 ci					low	high					low	high			
$\log(\Phi_{LS})$	ρ_x	-1.66	-1.78	-1.54	2.61	2.42	2.81	0.09	0.012	712	NS	NS	NS	0.07	0.153	402	NS	NS	NS	0.17	0.236	50
$\log(\Phi_{LS})$	\emptyset	1.18	1.08	1.28	-7.08	-6.57	-7.63	-0.09	0.017	686	NS	NS	NS	-0.08	0.113	396	NS	NS	NS	-0.09	0.531	49
$\log(\Phi_{LS})$	$\log(S)$	NS	NS	NS	NS	NS	NS	-0.03	0.373	696	NS	NS	NS	-0.09	0.069	374	NA	NA	NA	NA	NA	NA
$\log(\Phi_{LS})$	H_{\max}	NS	NS	NS	NS	NS	NS	0.01	0.785	741	NS	NS	NS	-0.06	0.206	403	NA	NA	NA	NA	NA	NA
$\log(\Phi_{LS})$	$\log(M_A)$	5.03	4.67	5.39	-2.59	-2.41	-2.78	-0.26	0.000	727	-1.26	-1.14	-1.38	-0.24	0.000	405	-3.49	-2.66	-4.60	-0.29	0.041	50
$\log(\Phi_{LS})$	[C]	3.91	3.62	4.21	-0.01	-0.01	-0.01	-0.13	0.000	699	NS	NS	NS	0.05	0.284	396	-0.01	-0.01	-0.01	-0.25	0.081	49
$\log(\Phi_{LS})$	$\log[N]$	-3.65	-3.91	-3.39	2.72	2.53	2.92	0.19	0.000	713	1.22	1.11	1.34	0.20	0.000	401	3.06	2.34	4.00	0.36	0.011	50
$\log(\Phi_{LS})$	$\log[P]$	NS	NS	NS	NS	NS	NS	0.01	0.782	705	1.03	0.93	1.13	0.13	0.007	397	NS	NS	NS	-0.07	0.638	49
$\log(\Phi_{LS})$	$\log[Ca]$	NS	NS	NS	NS	NS	NS	0.02	0.613	706	NS	NS	NS	0.00	0.954	398	NS	NS	NS	-0.02	0.914	49
$\log(\Phi_{LS})$	$\log[K]$	NS	NS	NS	NS	NS	NS	-0.03	0.400	706	NS	NS	NS	0.04	0.406	398	NS	NS	NS	-0.21	0.149	49
$\log(\Phi_{LS})$	$\log[Mg]$	NS	NS	NS	NS	NS	NS	0.03	0.446	706	NS	NS	NS	-0.09	0.067	398	NS	NS	NS	0.10	0.501	49
ρ_x	\emptyset	1.09	1.05	1.12	-2.63	-2.46	-2.80	-0.13	0.000	868	NS	NS	NS	-0.09	0.064	469	NS	NS	NS	-0.18	0.181	59
ρ_x	$\log(S)$	0.02	-0.02	0.06	0.12	0.11	0.13	0.23	0.000	876	0.05	0.04	0.05	0.25	0.000	444	NA	NA	NA	NA	NA	NA
ρ_x	H_{\max}	NS	NS	NS	NS	NS	NS	0.01	0.673	930	NS	NS	NS	-0.03	0.492	477	NA	NA	NA	NA	NA	NA
ρ_x	$\log(M_A)$	-1.32	-1.45	-1.19	0.98	0.92	1.05	0.10	0.002	897	0.88	0.80	0.96	0.12	0.007	477	NS	NS	NS	0.08	0.532	60
ρ_x	[C]	-0.91	-1.01	-0.81	0.00	0.00	0.00	0.10	0.004	882	NS	NS	NS	0.07	0.143	470	0.00	0.00	0.00	0.27	0.039	59
ρ_x	$\log[N]$	2.01	1.92	2.10	-1.06	-0.99	-1.13	-0.18	0.000	895	NS	NS	NS	-0.07	0.111	475	NS	NS	NS	-0.22	0.089	60
ρ_x	$\log[P]$	0.59	0.58	0.60	-0.64	-0.60	-0.68	-0.45	0.000	889	-0.72	-0.66	-0.79	-0.20	0.000	471	-0.45	-0.37	-0.55	-0.64	0.000	59
ρ_x	$\log[Ca]$	0.87	0.85	0.89	-0.34	-0.32	-0.37	-0.32	0.000	890	-0.37	-0.34	-0.41	-0.21	0.000	472	-0.26	-0.20	-0.33	-0.46	0.000	59
ρ_x	$\log[K]$	0.90	0.88	0.92	-0.39	-0.37	-0.42	-0.52	0.000	890	-0.53	-0.48	-0.57	-0.24	0.000	472	-0.30	-0.25	-0.34	-0.82	0.000	59
ρ_x	$\log[Mg]$	0.81	0.79	0.83	-0.56	-0.53	-0.60	-0.15	0.000	890	-0.40	-0.36	-0.43	-0.12	0.011	472	NS	NS	NS	-0.06	0.657	59
\emptyset	$\log(S)$	0.40	0.38	0.41	-0.04	-0.04	-0.05	-0.18	0.000	877	-0.38	-0.35	-0.41	-0.20	0.000	449	NA	NA	NA	NA	NA	NA
\emptyset	H_{\max}	0.02	0.00	0.03	0.01	0.00	0.01	0.13	0.000	934	0.00	0.00	0.00	0.13	0.005	484	NA	NA	NA	NA	NA	NA
\emptyset	$\log(M_A)$	-0.52	-0.57	-0.48	0.35	0.33	0.38	0.20	0.000	923	0.23	0.21	0.25	0.10	0.031	491	0.57	0.45	0.73	0.32	0.011	61
\emptyset	[C]	-0.39	-0.43	-0.36	0.001	0.001	0.001	0.08	0.015	926	NS	NS	NS	0.04	0.335	488	0.002	0.001	0.002	0.28	0.032	61
\emptyset	$\log[N]$	-0.34	-0.38	-0.31	0.40	0.37	0.42	0.16	0.000	936	0.22	0.20	0.24	0.23	0.000	492	NS	NS	NS	0.24	0.065	61
\emptyset	$\log[P]$	0.19	0.18	0.19	0.24	0.23	0.25	0.36	0.000	931	0.19	0.17	0.21	0.27	0.000	488	0.20	0.16	0.25	0.49	0.000	61
\emptyset	$\log[Ca]$	0.09	0.08	0.09	0.13	0.12	0.14	0.17	0.000	932	0.09	0.09	0.10	0.11	0.016	489	NS	NS	NS	0.25	0.055	61
\emptyset	$\log[K]$	0.07	0.06	0.08	0.15	0.14	0.16	0.22	0.000	932	NS	NS	NS	0.06	0.210	489	0.13	0.10	0.17	0.31	0.015	61
\emptyset	$\log[Mg]$	NS	NS	NS	NS	NS	NS	0.00	0.930	932	0.10	0.09	0.11	0.09	0.050	489	NS	NS	NS	-0.22	0.088	61

Table S2A (continued)

		raw data									genetic effects					plot-environmental effects							
y	x	intercept	intercept 0.95 ci		slope	slope 0.95 ci		r	sig	n	slope	slope 0.95 ci		r	sig	n	slope	slope		r	sig	n	
			low	high		low	high					low	high					low	high				
$\log(S)$	H_{\max}	1.18	0.92	1.45	0.12	0.12	0.13	0.18	0.000	941	0.17	0.16	0.17	0.14	0.002	457	NA	NA	NA	NA	NA	NA	NA
$\log(S)$	$\log(M_A)$	-11.64	-12.73	-10.55	8.46	7.92	9.02	0.11	0.001	905	90.28	82.45	98.86	0.12	0.014	456	NA	NA	NA	NA	NA	NA	NA
$\log(S)$	[C]	-8.13	-9.00	-7.27	0.03	0.03	0.03	0.12	0.001	891	0.23	0.21	0.26	0.18	0.000	448	NA	NA	NA	NA	NA	NA	NA
$\log(S)$	$\log[N]$	17.06	16.27	17.84	-9.13	-8.56	-9.74	-0.14	0.000	905	86.20	78.70	94.42	-0.16	0.001	453	NA	NA	NA	NA	NA	NA	NA
$\log(S)$	$\log[P]$	4.80	4.71	4.89	-5.66	-5.31	-6.04	-0.18	0.000	898	0.19	0.17	0.21	0.27	0.000	488	NA	NA	NA	NA	NA	NA	NA
$\log(S)$	$\log[Ca]$	7.25	7.08	7.41	-3.01	-2.82	-3.21	-0.23	0.000	899	-38.13	-34.92	-41.62	-0.34	0.000	450	NA	NA	NA	NA	NA	NA	NA
$\log(S)$	$\log[K]$	7.52	7.34	7.70	-3.42	-3.21	-3.64	-0.24	0.000	899	-53.10	-48.52	-58.11	-0.23	0.000	450	NA	NA	NA	NA	NA	NA	NA
$\log(S)$	$\log[Mg]$	6.74	6.60	6.88	-4.94	-4.63	-5.27	-0.15	0.000	899	-39.38	-36.14	-42.91	-0.25	0.000	450	NA	NA	NA	NA	NA	NA	NA
H_{\max}	$\log(M_A)$	-103.16	-111.59	-94.73	67.84	63.72	72.22	0.15	0.000	960	526.33	482.23	574.46	0.18	0.000	490	NA	NA	NA	NA	NA	NA	NA
H_{\max}	[C]	-75.39	-82.17	-68.60	0.22	0.21	0.24	0.08	0.011	949	NS	NS	NS	0.05	0.253	483	NA	NA	NA	NA	NA	NA	NA
H_{\max}	$\log[N]$	NS	NS	NS	NS	NS	NS	-0.04	0.167	963	NS	NS	NS	-0.03	0.534	488	NA	NA	NA	NA	NA	NA	NA
H_{\max}	$\log[P]$	NS	NS	NS	NS	NS	NS	0.00	0.979	956	NS	NS	NS	0.00	0.918	484	NA	NA	NA	NA	NA	NA	NA
H_{\max}	$\log[Ca]$	48.32	46.96	49.68	-24.10	-22.62	-25.68	-0.07	0.030	957	NS	NS	NS	-0.06	0.167	485	NA	NA	NA	NA	NA	NA	NA
H_{\max}	$\log[K]$	NS	NS	NS	NS	NS	NS	-0.03	0.302	957	NS	NS	NS	-0.03	0.506	485	NA	NA	NA	NA	NA	NA	NA
H_{\max}	$\log[Mg]$	44.33	43.18	45.48	-39.81	-37.38	-42.41	-0.11	0.001	957	NS	NS	NS	-0.09	0.054	485	NA	NA	NA	NA	NA	NA	NA

Table S2B. Pair-wise relationships between the genetic components of key foliar properties of species found in low and high fertility plots. The genetic component is computed by summing the *Family + Genus + Species* effect as estimated from the multilevel model. *Slope* of the SMA, Pearson's *r* correlation coefficient, *sig* the significance of the correlation, and *n* the number of cases used. Boldface indicates significant difference ($p < 0.05$) in slope or elevation and/or shift across the SMA axis.

<i>y</i>	<i>x</i>	<u>low fertility</u>										<u>high fertility</u>						<u>significance</u>				
		int. low	intercept 0.95 ci		slope	slope 0.95 ci		<i>r</i>	<i>sig</i>	<i>n</i>	intercept high	intercept 0.95 ci		slope	Slope 0.95 ci		<i>r</i>	<i>sig</i>	<i>n</i>	slope	elev	shift
			low	high		low	high					low	high		low	high						
$\log(L_A)$	$\log(\ell_d)$	0.09	0.03	0.15	1.55	1.34	1.79	0.30	0.000	168	0.06	0.01	0.10	1.47	1.33	1.63	0.47	0.000	279	0.575	0.423	0.768
$\log(L_A)$	$\log(\Phi_{LS})$	0.08	0.01	0.15	5.19	4.43	6.09	0.22	0.007	147	0.00	-0.05	0.06	5.42	4.81	6.12	0.32	0.000	240	0.664	0.122	0.451
$\log(L_A)$	ρ_x	0.10	0.03	0.17	-6.72	-5.79	-7.81	-0.17	0.028	169	NS	NS	NS	NS	NS	NS	-0.06	0.317	279	-	-	-
$\log(L_A)$	\emptyset	0.09	0.02	0.16	24.65	21.16	28.71	0.16	0.040	164	NS	NS	NS	NS	NS	NS	0.05	0.393	273	-	-	-
$\log(L_A)$	$\log(S)$	NS	NS	NS	NS	NS	NS	0.00	0.956	163	NS	NS	NS	NS	NS	NS	0.04	0.563	253	-	-	-
$\log(L_A)$	H_{\max}	NS	NS	NS	NS	NS	NS	-0.11	0.165	168	NS	NS	NS	NS	NS	NS	0.00	0.979	272	-	-	-
$\log(L_A)$	$\log(M_A)$	0.16	0.09	0.23	-6.17	-5.32	-7.16	-0.20	0.011	169	NS	NS	NS	NS	NS	NS	-0.06	0.360	276	-	-	-
$\log(L_A)$	[C]	NS	NS	NS	NS	NS	NS	0.08	0.302	167	NS	NS	NS	NS	NS	NS	0.11	0.081	269	-	-	-
$\log(L_A)$	$\log[N]$	0.06	0.00	0.12	5.77	5.00	6.67	0.33	0.000	168	-0.05	-0.11	0.01	6.61	5.89	7.42	0.25	0.000	273	0.151	0.016	0.097
$\log(L_A)$	$\log[P]$	0.10	0.04	0.15	5.10	4.46	5.84	0.46	0.000	167	-0.01	-0.07	0.04	5.50	4.91	6.16	0.33	0.000	270	0.405	0.006	0.099
$\log(L_A)$	$\log[Ca]$	NS	NS	NS	NS	NS	NS	-0.03	0.655	167	NS	NS	NS	NS	NS	NS	-0.02	0.806	271	-	-	-
$\log(L_A)$	$\log[K]$	NS	NS	NS	NS	NS	NS	0.05	0.503	167	NS	NS	NS	NS	NS	NS	0.00	0.954	271	-	-	-
$\log(L_A)$	$\log[Mg]$	-0.11	-0.19	-0.04	-2.99	-2.58	-3.48	-0.20	0.009	167	NS	NS	NS	NS	NS	NS	-0.10	0.099	271	-	-	-
$\log(\ell_d)$	$\log(\Phi_{LS})$	NS	NS	NS	NS	NS	NS	0.05	0.528	146	NS	NS	NS	NS	NS	NS	0.02	0.815	244	-	-	-
$\log(\ell_d)$	ρ_x	0.00	-0.04	0.05	-4.42	-3.82	-5.13	-0.23	0.003	171	-0.04	-0.08	0.00	-5.45	-4.86	-6.11	-0.21	0.000	285	0.028	0.156	0.043
$\log(\ell_d)$	\emptyset	NS	NS	NS	NS	NS	NS	-0.08	0.336	166	NS	NS	NS	NS	NS	NS	0.09	0.132	277	-	-	-
$\log(\ell_d)$	$\log(S)$	NS	NS	NS	NS	NS	NS	-0.06	0.440	165	NS	NS	NS	NS	NS	NS	0.06	0.329	258	-	-	-
$\log(\ell_d)$	H_{\max}	NS	NS	NS	NS	NS	NS	-0.02	0.805	170	NS	NS	NS	NS	NS	NS	-0.05	0.453	278	-	-	-
$\log(\ell_d)$	$\log(M_A)$	-0.08	-0.13	-0.04	4.07	3.50	4.72	0.19	0.011	171	-0.01	-0.05	0.03	4.64	4.13	5.22	0.15	0.015	280	0.168	0.018	0.273
$\log(\ell_d)$	[C]	NS	NS	NS	NS	NS	NS	-0.10	0.200	169	NS	NS	NS	NS	NS	NS	-0.10	0.111	273	-	-	-
$\log(\ell_d)$	$\log[N]$	NS	NS	NS	NS	NS	NS	-0.09	0.236	170	0.08	0.04	0.12	-4.51	-4.01	-5.08	-0.12	0.049	277	-	-	-
$\log(\ell_d)$	$\log[P]$	NS	NS	NS	NS	NS	NS	-0.01	0.948	169	NS	NS	NS	NS	NS	NS	0.05	0.413	274	-	-	-
$\log(\ell_d)$	$\log[Ca]$	NS	NS	NS	NS	NS	NS	0.10	0.192	169	NS	NS	NS	NS	NS	NS	0.03	0.591	275	-	-	-
$\log(\ell_d)$	$\log[K]$	0.07	0.02	0.11	2.49	2.15	2.89	0.21	0.006	169	0.01	-0.03	0.05	2.68	2.38	3.01	0.17	0.004	275	0.455	0.060	0.011
$\log(\ell_d)$	$\log[Mg]$	NS	NS	NS	NS	NS	NS	0.14	0.072	169	0.05	0.01	0.09	2.06	1.83	2.32	0.17	0.006	275	-	-	-

Table S2B (continued)

Y	x	<u>low fertility</u>										<u>high fertility</u>						<u>significance</u>				
		int. low	intercept 0.95 ci		slope	slope 0.95 ci		r	sig	n	intercept high	intercept 0.95 ci		slope	Slope 0.95 ci		r	sig	n	slope	elev	shift
			low	high		low	high					low	high		low	high						
log(Φ_{LS})	ρ_x	NS	NS	NS	NS	NS	NS	0.05	0.576	151	NS	NS	NS	NS	NS	NS	0.09	0.148	248	-	-	-
log(Φ_{LS})	\emptyset	NS	NS	NS	NS	NS	NS	-0.06	0.471	151	NS	NS	NS	NS	NS	NS	-0.11	0.082	243	-	-	-
log(Φ_{LS})	log(S)	NS	NS	NS	NS	NS	NS	-0.15	0.058	151	NS	NS	NS	NS	NS	NS	-0.06	0.408	226	-	-	-
log(Φ_{LS})	H_{max}	NS	NS	NS	NS	NS	NS	-0.06	0.487	155	NS	NS	NS	NS	NS	NS	-0.11	0.096	245	-	-	-
log(Φ_{LS})	log(M_A)	0.02	0.00	0.03	-1.17	-1.00	-1.37	-0.19	0.016	156	0.01	0.00	0.02	-1.28	-1.14	-1.45	-0.28	0.000	246	0.372	0.615	0.035
log(Φ_{LS})	[C]	NS	NS	NS	NS	NS	NS	0.04	0.582	154	NS	NS	NS	NS	NS	NS	0.05	0.469	239	-	-	-
log(Φ_{LS})	log[N]	NS	NS	NS	NS	NS	□S	0.08	0.307	155	-0.01	-0.03	0.00	1.27	1.12	1.44	0.26	0.000	243	-	-	-
log(Φ_{LS})	log[P]	NS	NS	NS	NS	NS	NS	0.04	0.596	154	-0.01	-0.02	0.01	1.04	0.92	1.18	0.17	0.007	240	-	-	-
log(Φ_{LS})	log[Ca]	NS	NS	NS	NS	NS	NS	0.09	0.245	154	NS	NS	NS	NS	NS	NS	-0.07	0.256	241	-	-	-
log(Φ_{LS})	log[K]	NS	NS	NS	NS	NS	NS	0.03	0.727	154	NS	NS	NS	NS	NS	NS	0.04	0.531	241	-	-	-
log(Φ_{LS})	log[Mg]	NS	NS	NS	NS	NS	NS	-0.02	0.823	154	0.00	-0.01	0.01	-0.56	-0.50	-0.64	-0.14	0.033	241	-	-	-
ρ_x	\emptyset	NS	NS	NS	NS	NS	NS	0.00	0.983	176	-0.01	-0.02	-0.01	-3.98	-3.55	-4.47	-0.14	0.022	286	-	-	-
ρ_x	log(S)	-0.26	-0.30	-0.22	0.05	0.04	0.06	0.20	0.007	177	-0.23	-0.25	-0.20	0.05	0.04	0.05	0.20	0.001	265	0.193	0.779	0.001
ρ_x	H_{max}	NS	NS	NS	NS	NS	NS	-0.02	0.781	182	NS	NS	NS	NS	NS	NS	-0.01	0.834	286	-	-	-
ρ_x	log(M_A)	-0.01	-0.02	0.00	0.94	0.81	1.09	0.16	0.032	181	NS	NS	NS	NS	NS	NS	0.11	0.056	288	-	-	-
ρ_x	[C]	-0.01	-0.02	0.00	0.00	0.00	0.00	0.15	0.049	179	NS	NS	NS	NS	NS	NS	-0.02	0.682	282	-	-	-
ρ_x	log[N]	NS	NS	NS	NS	NS	NS	-0.09	0.249	180	NS	NS	NS	NS	NS	NS	-0.06	0.280	286	-	-	-
ρ_x	log[P]	0.00	-0.01	0.01	-0.78	-0.68	-0.90	-0.23	0.002	179	0.00	-0.01	0.01	-0.69	-0.62	-0.78	-0.15	0.012	283	0.197	0.606	0.000
ρ_x	log[Ca]	-0.02	-0.03	-0.01	-0.42	-0.36	-0.48	-0.27	0.000	179	-0.01	-0.02	0.00	-0.38	-0.34	-0.42	-0.15	0.012	284	0.261	0.248	0.000
ρ_x	log[K]	-0.02	-0.03	-0.01	-0.59	-0.51	-0.67	-0.31	0.000	179	-0.01	-0.02	0.00	-0.49	-0.43	-0.55	-0.21	0.000	284	0.046	0.423	0.000
ρ_x	log[Mg]	-0.02	-0.04	-0.01	-0.45	-0.39	-0.52	-0.17	0.024	179	NS	NS	NS	NS	NS	NS	-0.10	0.101	284	-	-	-

Table S2B (continued)

		<u>low fertility</u>										<u>high fertility</u>						<u>significance</u>				
<i>y</i>	<i>x</i>	int. low	intercept		slope	slope		<i>r</i>	<i>sig</i>	<i>n</i>	intercept high	intercept		slope	Slope		<i>r</i>	<i>sig</i>	<i>n</i>	slope	elev	shift
			0.95 ci	low		high	0.95 ci					low	high		low	high						
\emptyset	log(<i>S</i>)	NS	NS	NS	NS	NS	NS	-0.13	0.080	178	0.05	0.05	0.06	-0.01	-0.01	-0.01	-0.25	0.000	264	-	-	-
\emptyset	H_{\max}	NS	NS	NS	NS	NS	NS	0.01	0.856	184	-0.04	-0.04	-0.03	0.00	0.00	0.00	0.14	0.020	286	-	-	-
\emptyset	log(M_A)	NS	NS	NS	NS	NS	NS	0.12	0.099	185	NS	NS	NS	NS	NS	NS	0.02	0.687	293	-	-	-
\emptyset	[C]	NS	NS	NS	NS	NS	NS	0.14	0.053	184	NS	NS	NS	NS	NS	NS	-0.05	0.441	290	-	-	-
\emptyset	log[N]	0.00	0.00	0.00	0.24	0.20	0.27	0.28	0.000	185	-0.01	-0.01	0.00	0.21	0.19	0.24	0.24	0.000	293	0.265	0.007	0.245
\emptyset	log[P]	0.00	0.00	0.00	0.21	0.18	0.24	0.35	0.000	184	0.00	-0.01	0.00	0.17	0.15	0.19	0.24	0.000	290	0.033	0.002	0.164
\emptyset	log[Ca]	NS	NS	NS	NS	NS	NS	0.10	0.190	184	0.00	0.00	0.00	0.09	0.08	0.10	0.22	0.000	291	-	-	-
\emptyset	log[K]	NS	NS	NS	NS	NS	NS	0.12	0.117	184	NS	NS	NS	NS	NS	NS	0.08	0.150	291	-	-	-
\emptyset	log[Mg]	NS	NS	NS	NS	NS	NS	0.07	0.379	184	0.00	0.00	0.00	0.09	0.08	0.11	0.14	0.013	291	-	-	-
log(<i>S</i>)	H_{\max}	1.87	1.36	2.38	0.12	0.10	0.13	0.18	0.012	185	1.55	1.11	1.99	0.11	0.10	0.13	0.16	0.009	265	0.826	0.002	0.184
log(<i>S</i>)	log(M_A)	NS	NS	NS	NS	NS	NS	0.08	0.275	183	4.83	4.66	5.00	19.17	17.02	21.59	0.17	0.005	266	-	-	-
log(<i>S</i>)	[C]	4.82	4.61	5.02	0.05	0.04	0.06	0.16	0.029	181	4.80	4.63	4.97	0.05	0.04	0.05	0.17	0.005	260	0.331	0.742	0.000
log(<i>S</i>)	log[N]	NS	NS	NS	NS	NS	NS	-0.06	0.413	182	5.22	5.05	5.38	-18.90	-16.79	-21.26	-0.24	0.000	264	-	-	-
log(<i>S</i>)	log[P]	NS	NS	NS	NS	NS	NS	0.08	0.296	181	5.09	4.92	5.27	-15.77	-13.98	-17.80	-0.14	0.019	261	-	-	-
log(<i>S</i>)	log[Ca]	4.68	4.49	4.86	-8.06	-7.02	-9.26	-0.35	0.000	181	4.73	4.57	4.88	-8.67	-7.72	-9.73	-0.32	0.000	262	0.429	0.523	0.000
log(<i>S</i>)	log[K]	4.77	4.56	4.98	-11.49	-9.93	-13.28	-0.15	0.043	181	4.85	4.68	5.01	-10.81	-9.60	-12.16	-0.24	0.000	262	0.522	0.646	0.000
log(<i>S</i>)	log[Mg]	4.59	4.38	4.79	-9.08	-7.86	-10.48	-0.21	0.004	181	4.65	4.48	4.82	-8.50	-7.55	-9.57	-0.23	0.000	262	0.491	0.790	0.000
H_{\max}	log(M_A)	26.2	24.6	27.9	161.6	140.2	186.1	0.19	0.010	189	29.17	27.78	30.55	170.85	152.64	191.23	0.24	0.000	288	0.546	0.006	0.347
H_{\max}	[C]	NS	NS	NS	NS	NS	NS	0.03	0.686	187	NS	NS	NS	NS	NS	NS	0.10	0.111	282	-	-	-
H_{\max}	log[N]	NS	NS	NS	NS	NS	NS	-0.10	0.183	188	NS	NS	NS	NS	NS	NS	-0.05	0.403	286	-	-	-
H_{\max}	log[P]	NS	NS	NS	NS	NS	NS	-0.11	0.143	187	NS	NS	NS	NS	NS	NS	-0.04	0.522	283	-	-	-
H_{\max}	log[Ca]	NS	NS	NS	NS	NS	NS	-0.13	0.084	187	NS	NS	NS	NS	NS	NS	-0.08	0.205	284	-	-	-
H_{\max}	log[K]	NS	NS	NS	NS	NS	NS	-0.09	0.239	187	NS	NS	NS	NS	NS	NS	-0.10	0.106	284	-	-	-
H_{\max}	log[Mg]	23.5	21.7	25.4	-77.6	-67.3	-89.5	-0.16	0.031	187	NS	NS	NS	NS	NS	NS	-0.10	0.109	284	-	-	-

Table S3: Decomposition of the χ^2_{total} according to the Flury hierarchy. In this procedure a range of models are examined ranging from the two covariance matrices being unrelated through partial common principal components, CPC(1) through CPC(11), then extending to the full CPC model, followed by the proportional or equality models. According to the “step-up” procedure outlined by Flury (1988), the CPC model clearly emerges as the best model fit. See also Phillips and Arnold (1999).

Model		χ^2	df	P	$\frac{\chi^2}{\text{df}}$	AIC for Higher Model
Higher	Lower					
Equality	Proportional	3.318	3	0.3451	1.106	564.800
Proportional	CPC	162.709	36	0.0000	4.520	567.482
CPC	CPC(11)	3.836	3	0.2797	1.279	476.773
CPC(11)	CPC(10)	5.362	6	0.4983	0.894	478.937
CPC(10)	CPC(9)	10.801	9	0.2896	1.200	485.575
CPC(9)	CPC(8)	19.704	12	0.0729	1.642	492.774
CPC(8)	CPC(7)	15.762	15	0.3980	1.051	497.071
CPC(7)	CPC(6)	41.291	18	0.0014	2.294	511.308
CPC(6)	CPC(5)	33.360	21	0.0424	1.589	506.018
CPC(5)	CPC(4)	39.074	24	0.0268	1.628	514.657
CPC(4)	CPC(3)	58.826	27	0.0004	2.179	523.583
CPC(3)	CPC(2)	39.902	30	0.1068	1.330	518.757
CPC(2)	CPC(1)	78.028	33	0.0000	2.364	538.855
CPC(1)	Unrelated	52.828	36	0.0348	1.467	526.828
Unrelated						546.00

Table S4. Test results for sphericity of adjacent eigenvectors (λ) according to Eq. 1.40 of Chapter 3 of Flury (1988). According to the χ^2 test the sixth eigenvector was not unique to the seventh and thus only the first five λ were retained (See Table 2).

λ	(1, 2)	(2, 3)	(3, 4)	(4, 5)	(5, 6)	(6, 7)	(7, 8)	(8, 9)	(9,10)	(10,11)	(11,12)	(12,13)
χ^2	7.7	35.34	10.84	15.95	35.38	3.46	5.96	6.54	6.87	3.64	38.85	2.06
P	0.05	<0.01	0.01	<0.01	<0.01	0.33	0.11	0.09	0.07	0.03	<0.01	0.56

Table S5. Common Principal Component Analysis for species associated with low and high fertility soils. Values in brackets represent standard errors and, for each component. M_A = leaf mass per unit area; elemental concentrations are on a dry weight basis, L_A = leaf area; Φ_{LS} = leaf area:sapwood area ratio, ρ_X = branch xylem density, \hat{O} = stomatal limitation index (see Eq. 1), S = seed mass, H_{max} = species maximum height.

Variable	Component												
	U_1	U_2	U_3	U_4	U_5	U_6	U_7	U_8	U_9	U_{10}	U_{11}	U_{12}	U_{13}
$\log(M_A)$	-0.220 (0.053)	-0.230 (0.061)	0.440 (0.071)	0.220 (0.107)	0.350 (0.088)	0.160 (0.121)	-0.060 (0.141)	0.280 (0.124)	0.280 (0.192)	-0.300 (0.147)	-0.460 (0.142)	0.190 (0.062)	0.080 (0.080)
[C]	-0.350 (0.049)	0.240 (0.070)	0.010 (0.071)	-0.060 (0.115)	0.340 (0.088)	-0.110 (0.098)	-0.230 (0.106)	0.330 (0.123)	0.240 (0.227)	-0.150 (0.173)	0.580 (0.115)	-0.280 (0.081)	-0.170 (0.108)
$\log[N]$	0.150 (0.097)	0.530 (0.039)	-0.020 (0.087)	-0.220 (0.085)	-0.030 (0.080)	-0.280 (0.073)	0.120 (0.122)	-0.100 (0.101)	0.080 (0.130)	-0.510 (0.072)	-0.190 (0.145)	0.330 (0.135)	-0.360 (0.121)
$\log[P]$	0.250 (0.083)	0.450 (0.052)	0.120 (0.067)	-0.310 (0.052)	0.080 (0.063)	0.030 (0.076)	-0.040 (0.120)	0.150 (0.071)	0.080 (0.097)	0.020 (0.085)	-0.210 (0.067)	-0.350 (0.224)	0.640 (0.123)
$\log[Ca]$	0.420 (0.033)	-0.130 (0.081)	0.150 (0.068)	0.310 (0.062)	0.000 (0.081)	0.120 (0.080)	-0.020 (0.129)	-0.180 (0.093)	-0.120 (0.107)	-0.460 (0.070)	0.030 (0.142)	-0.610 (0.085)	-0.210 (0.215)
$\log[K]$	0.480 (0.023)	-0.010 (0.091)	0.000 (0.075)	-0.160 (0.094)	0.050 (0.105)	0.210 (0.083)	0.050 (0.137)	0.500 (0.084)	0.210 (0.133)	0.390 (0.095)	-0.110 (0.149)	-0.030 (0.173)	-0.480 (0.040)
$\log[Mg]$	0.490 (0.042)	-0.210 (0.091)	0.070 (0.059)	-0.060 (0.069)	0.190 (0.065)	0.140 (0.063)	0.080 (0.078)	-0.040 (0.100)	0.160 (0.201)	-0.210 (0.151)	0.530 (0.093)	0.450 (0.118)	0.310 (0.161)
$\log(L_A)$	-0.010 (0.092)	0.480 (0.047)	0.250 (0.133)	0.350 (0.155)	-0.160 (0.095)	0.500 (0.127)	-0.310 (0.198)	-0.300 (0.121)	0.180 (0.107)	0.210 (0.084)	0.100 (0.112)	0.170 (0.047)	-0.070 (0.068)
$\log(\Phi_{LS})$	-0.010 (0.065)	0.290 (0.055)	-0.440 (0.113)	0.530 (0.164)	0.180 (0.108)	0.190 (0.175)	0.390 (0.214)	0.340 (0.135)	-0.260 (0.100)	-0.080 (0.092)	0.000 (0.123)	0.090 (0.060)	0.130 (0.046)
ρ_X	-0.140 (0.034)	-0.030 (0.051)	-0.220 (0.104)	-0.120 (0.206)	0.260 (0.113)	0.210 (0.170)	0.530 (0.134)	-0.420 (0.162)	0.540 (0.110)	0.090 (0.133)	-0.050 (0.213)	-0.200 (0.052)	-0.050 (0.081)
\hat{O}	0.100 (0.043)	0.140 (0.051)	0.390 (0.088)	0.100 (0.132)	0.600 (0.080)	-0.310 (0.116)	0.150 (0.146)	-0.240 (0.147)	-0.350 (0.112)	0.370 (0.107)	0.030 (0.175)	0.030 (0.063)	-0.100 (0.045)
$\log(S)$	-0.100 (0.043)	0.070 (0.061)	0.530 (0.100)	0.130 (0.223)	-0.470 (0.090)	-0.180 (0.195)	0.570 (0.125)	0.210 (0.155)	0.110 (0.125)	0.050 (0.104)	0.220 (0.088)	-0.060 (0.045)	0.040 (0.042)
H_{max}	-0.230 (0.033)	0.010 (0.060)	0.190 (0.098)	-0.480 (0.098)	0.590 (0.081)	0.200 (0.116)	0.050 (0.196)	0.030 (0.160)	-0.490 (0.085)	-0.160 (0.123)	0.080 (0.194)	0.000 (0.062)	-0.110 (0.042)
Characteristic roots													
$\lambda_{low,j}$	1876. (258.9)	1472. (203.1)	641. (88.5)	717. (99.0)	698. (96.3)	454. (62.7)	540. (74.5)	442. (61.0)	271. (37.4)	315. (43.5)	240. (33.1)	166. (22.9)	138. (19.0)
$\lambda_{high,j}$	2341. (237.1)	1641. (166.2)	898. (91.0)	564. (57.1)	318. (32.2)	707. (71.6)	560. (56.7)	408. (41.3)	379. (38.4)	265. (26.8)	318. (32.2)	134. (13.6)	116. (11.7)

Table S6: Covariance (\mathbf{F}) and correlation (\mathbf{R}) matrices of CPCs. The generally low correlations between the various vectors are of little practical consequence and indicate a reasonable model fit (Flury 1988).

$\mathbf{R}_{\text{low}} \setminus \mathbf{F}_{\text{low}} =$

1869.87	-50.57	55.93	177.24	-293.55	168.46	-59.64	13.26	-92.68	-57.31	112.82	-17.27	-63.85
-0.03	1465.59	83.40	-157.88	123.83	-85.35	-166.14	-48.23	-83.57	9.77	-42.26	24.18	-11.81
0.05	0.09	647.98	-40.42	170.45	-65.95	-75.39	33.56	-59.68	58.17	-20.97	-27.34	-21.33
0.15	-0.15	-0.06	715.87	-105.51	162.36	50.95	64.20	-11.96	-11.25	43.78	-22.24	-3.79
-0.23	0.11	0.23	-0.13	855.89	4.24	72.03	-56.99	-47.80	12.06	-19.87	-23.74	4.09
0.21	-0.12	-0.14	0.33	0.01	333.31	-50.07	34.32	37.40	14.77	-22.48	-56.48	-7.33
-0.06	-0.19	-0.13	0.08	0.11	-0.12	506.83	27.49	22.09	-20.47	49.18	39.80	-34.93
0.01	-0.06	0.06	0.11	-0.09	0.09	0.06	437.01	2.03	-7.12	-29.58	-18.40	23.46
-0.13	-0.13	-0.14	-0.03	-0.10	0.12	0.06	0.01	269.98	-16.70	-1.68	14.44	-33.31
-0.07	0.01	0.13	-0.02	0.02	0.05	-0.05	-0.02	-0.06	317.07	-18.66	-29.31	-3.81
0.17	-0.07	-0.05	0.11	-0.04	-0.08	0.14	-0.09	-0.01	-0.07	237.93	-15.66	7.16
-0.03	0.05	-0.08	-0.06	-0.06	-0.24	0.14	-0.07	0.07	-0.13	-0.08	166.54	0.20
-0.13	-0.03	-0.07	-0.01	0.01	-0.03	-0.13	0.10	-0.17	-0.02	0.04	0.00	136.43

$\mathbf{R}_{\text{high}} \setminus \mathbf{F}_{\text{high}} =$

2328.48	13.29	-48	-57.22	-302.31	230.41	145.18	-5.86	92.51	25.13	-69.53	10.98	34.65
0.01	1631.92	-71.03	45.95	-14.14	42.24	86.74	27.27	69.42	-7.35	42.78	0.79	6.25
-0.03	-0.06	906.44	-2.92	95.54	-58.9	-13.88	-4.51	39.76	-14.66	17.46	10.58	14.85
-0.05	0.05	0.56	1.82	-138.36	122.81	63.24	-46.59	18.72	3.21	-46.77	12.66	4.00
-0.28	-0.02	0.14	-0.26	512.91	75.94	-0.01	-16.31	-84.25	-49.87	29.91	10.12	-7.64
0.22	0.05	-0.09	0.23	0.15	490.35	-50.05	-15.66	85.61	19.57	-3.75	22.2	12.62
0.13	0.09	-0.02	0.11	0.00	-0.15	547.19	-9.88	24.84	18.19	-47.27	-14.18	18.12
-0.01	0.03	-0.01	-0.1	-0.04	-0.04	-0.02	403.65	-1.55	0.79	43.46	5.51	-9.59
0.1	0.09	0.07	0.04	-0.19	0.2	0.05	0.37	9.58	-2.38	1.91	-1.21	9.52
0.03	-0.01	-0.03	0.01	-0.14	0.05	0.05	0	-0.01	265.53	-15.53	13.66	-0.42
-0.08	0.06	0.03	-0.11	0.07	-0.01	-0.11	0.12	0.01	-0.05	314.62	3.08	-1.18
0.02	0	0.03	0.05	0.04	0.09	-0.05	0.02	-0.01	0.07	0.01	134.96	1.09
0.07	0.01	0.05	0.02	-0.03	0.05	0.07	-0.04	0.05	0	-0.01	0.01	114.97

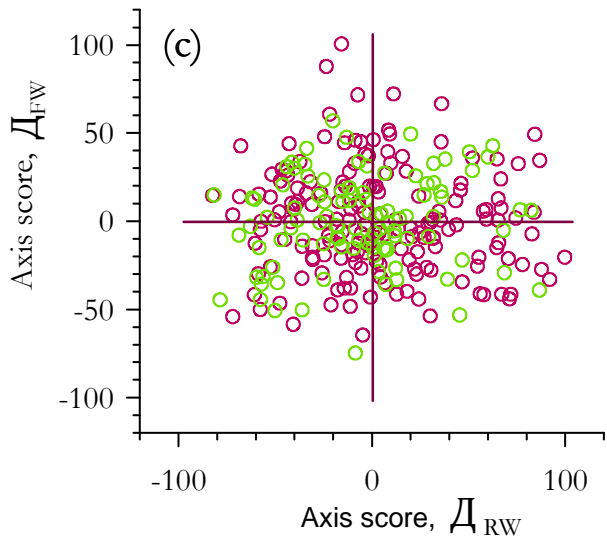
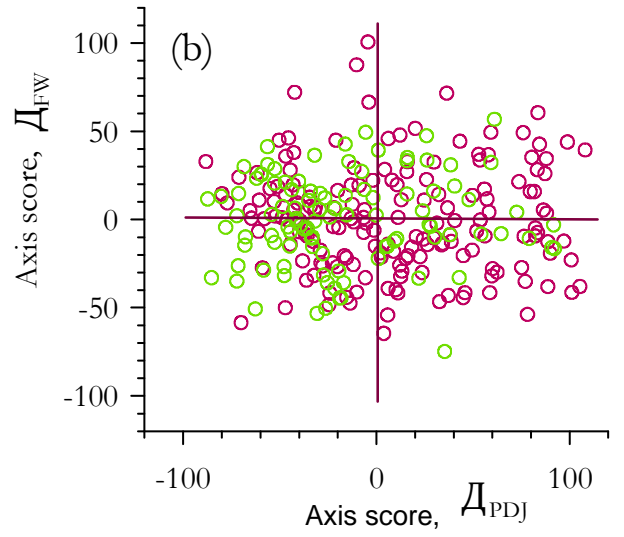
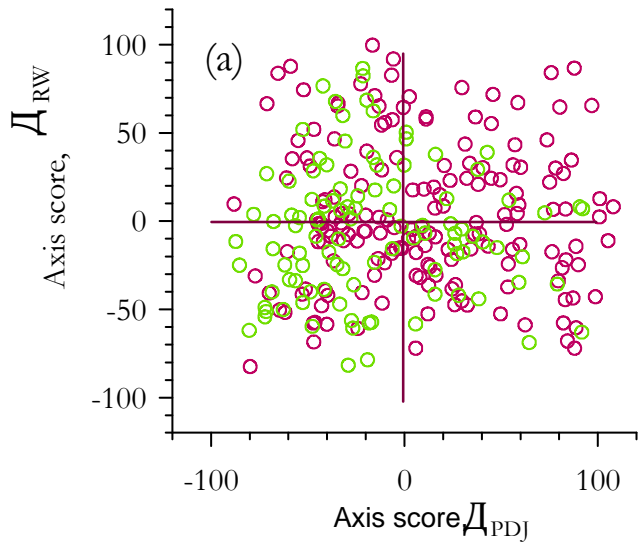


Figure S1. Summary of CPC analysis results (a-c), first three axis scores plotted against each other showing a lack of correlation required for a good fit; green: species associated with low fertility soil, purple: species associated with high fertility soils.