## Europe PMC Funders Group Author Manuscript Cancer Epidemiol Biomarkers Prev. Author manuscript; available in PMC 2011 March 01.

Published in final edited form as: *Cancer Epidemiol Biomarkers Prev.* 2010 September ; 19(9): 2143–2151. doi: 10.1158/1055-9965.EPI-10-0374.

# Missense Variants in *ATM* in 26,101 Breast Cancer Cases and 29,842 Controls

Olivia Fletcher<sup>1,\*</sup>, Nichola Johnson<sup>1</sup>, Isabel dos Santos Silva<sup>1</sup>, Nick Orr<sup>1</sup>, Alan Ashworth<sup>1</sup>, Heli Nevanlinna<sup>2</sup>, Tuomas Heikkinen<sup>2</sup>, Kristiina Aittomäki<sup>2</sup>, Carl Blomqvist<sup>2</sup>, Barbara Burwinkel<sup>3</sup>, Claus R. Bartram<sup>3</sup>, Alfons Meindl<sup>3</sup>, Rita K. Schmutzler<sup>3</sup>, Angela Cox<sup>4</sup>, Ian Brock<sup>4</sup>, Graeme Elliott<sup>4</sup>, Malcolm W. R. Reed<sup>4</sup>, Melissa C. Southey<sup>5</sup>, Letitia Smith<sup>5</sup>, Amanda B. Spurdle<sup>5</sup>, John L. Hopper<sup>5</sup>, Fergus J. Couch<sup>6</sup>, Janet E. Olson<sup>6</sup>, Xianshu Wang<sup>6</sup>, Zachary Fredericksen<sup>6</sup>, Peter Schürmann<sup>7</sup>, Regina Waltes<sup>7</sup>, Michael Bremer<sup>7</sup>, Thilo Dörk<sup>7</sup>, Peter Devilee<sup>8</sup>, Christie J. van Asperen<sup>8</sup>, Rob A.E.M. Tollenaar<sup>8</sup>, Caroline Seynaeve<sup>8</sup>, Per Hall<sup>9</sup>, Kamila Czene<sup>9</sup>, Keith Humphreys<sup>9</sup>, Jianjun Liu<sup>9</sup>, Shahana Ahmed<sup>10</sup>, Alison M. Dunning<sup>10</sup>, Melanie Maranian<sup>10</sup>, Paul D.P. Pharoah<sup>10</sup>, Georgia Chenevix-Trench<sup>11</sup>, Jonathan Beesley<sup>11</sup>, kConFab Investigators<sup>11</sup>, AOCS Group<sup>11</sup>, Natalia V. Bogdanova<sup>12</sup>, Natalia N. Antonenkova<sup>12</sup>, losif V. Zalutsky<sup>12</sup>, Hoda Anton-Culver<sup>13</sup>, Argyrios Ziogas<sup>13</sup>, Hiltrud Brauch<sup>14</sup>, Yon-Dschun Ko<sup>14</sup>, Ute Hamann<sup>14</sup>, the GENICA Consortium<sup>14</sup>, Peter A. Fasching<sup>15</sup>, Reiner Strick<sup>15</sup>, Arif B. Ekici<sup>15</sup>, Matthias W. Beckmann<sup>15</sup>, Graham G. Giles<sup>16</sup>, Gianluca Severi<sup>16</sup>, Laura Baglietto<sup>16</sup>, Dallas R. English<sup>16</sup>, Roger L. Milne<sup>17</sup>, Javier Benítez<sup>17</sup>, José Ignacio Arias<sup>17</sup>, Guillermo Pita<sup>17</sup>, Børge G. Nordestgaard<sup>18</sup>, Stig E. Bojesen<sup>18</sup>, Henrik Flyger<sup>18</sup>, Daehee Kang<sup>19</sup>, Keun-Young Yoo<sup>19</sup>, Dong Young Noh<sup>19</sup>, Arto Mannermaa<sup>20</sup>, Vesa Kataja<sup>20</sup>, Veli-Matti Kosma<sup>20</sup>, Montserrat García-Closas<sup>21</sup>, Stephen Chanock<sup>21</sup>, Jolanta Lissowska<sup>21</sup>, Louise A. Brinton<sup>21</sup>, Jenny Chang-Claude<sup>22</sup>, Shan Wang-Gohrke<sup>22</sup>, Annegien Broeks<sup>23</sup>, Marjanka K Schmidt<sup>23</sup>, Flora E van Leeuwen<sup>23</sup>, Laura J Van 't Veer<sup>23</sup>, Sara Margolin<sup>24</sup>, Annika Lindblom<sup>24</sup>, Manjeet K. Humphreys<sup>25</sup>, Jonathan Morrison<sup>25</sup>, Radka Platte<sup>25</sup>, Douglas F. Easton<sup>25</sup>, and Julian Peto<sup>1</sup> on behalf of the Breast **Cancer Association Consortium** 

<sup>1</sup> British Breast Cancer Study (BBCS): Breakthrough Breast Cancer Research Centre, London, UK [OF, NJ, NO, AA]; London School of Hygiene and Tropical Medicine, London, UK [OF, IdSS, JP]; Institute of Cancer Research, Sutton, Surrey, UK [JP]. <sup>2</sup> Helsinki Breast Cancer Study (HEBCS): Departments of Obstetrics and Gynecology [HN, TH], Clinical Genetics [KA] and Oncology [CB], Helsinki University Central Hospital, Helsinki, Finland <sup>3</sup> German Consortium for Hereditary Breast and Ovarian Cancer (GC-HBOC): Department of Obstetrics and Gynecology [BB] and Institute of Human Genetics [CRB], University of Heidelberg, Heidelberg, Germany; Molecular Epidemiology Group, German Cancer Research Center (DKFZ), Heidelberg, Germany [BB]; Department of Gynaecology and Obstetrics, Technical University of Munich, Munich, Germany [AM]; Department of Gynaecology and Obstetrics, Clinical Center University of Cologne, Köln, Germany [RKS] <sup>4</sup> Sheffield Breast Cancer Study (SBCS): Institute for Cancer Studies [AC, IB, GE], Academic Unit of Surgical Oncology [MWRR], Sheffield University Medical School, Sheffield, UK <sup>5</sup> Australian Breast Cancer Family Study (ABCFS): The University of Melbourne, Victoria, Australia [MCS, LS, JLH], Queensland Institute of Medical Research [ABS] <sup>6</sup>

Austrailan Ovarian Cancer Study: http://www.aocstudy.org/

<sup>&</sup>lt;sup>\*</sup> Corresponding author Olivia Fletcher The Breakthrough Breast Cancer Research Centre, Institute of Cancer Research, 237 Fulham Road, London, SW3 6JB, UK Tel: +44 (0) 20 7878 3813 olivia.fletcher@icr.ac.uk Fax: +44 (0) 20 7878 3858.

Web Addresses

BCAC: http://www.srl.cam.ac.uk/consortia/bcac/

Applied Biosystems: http://www.appliedbiosystems.com/

dbSNP: http://www.ncbi.nlm.nih.gov/projects/SNP/

Mayo Clinic Breast Cancer Study (MCBCS): Department of Laboratory Medicine and Pathology [FC.XW] and Department of Health Sciences Research [FC, JEO, ZF]. Mayo Clinic, Rochester. MN, USA. <sup>7</sup> Hannover Breast Cancer Study (HABCS): Department of Obstetrics and Gynaecology [TD, PS], Department of Radiation Oncology [RW, MB], Hannover Medical School, Hannover, Germany.<sup>8</sup> Leiden University Medical Centre Breast Cancer Study (ORIGO): Department of Human Genetics [PD], Department of Pathology [PD], Department of Clinical Genetics [CJvA] and Department of Surgery [RAEMT]. Leiden University Medical Centre, Leiden. The Netherlands; Department of Medical Oncology, Rotterdam Family Cancer Clinic, Erasmus MC-Daniel den Hoed Cancer Center, Rotterdam, The Netherlands [CS]. <sup>9</sup> Singapore and Swedish Breast Cancer Study (SASBAC): Department of Medical Epidemiology and Biostatistics, Karolinska Institute, Stockholm, Sweden [PH, KC, KH] and Human Genetics Laboratory, Genome Institute of Singapore, Singapore [JL]. <sup>10</sup> Studies of Epidemiology and Risk Factors in Cancer Heredity (SEARCH): Department of Oncology [SA, AMD, MM, PDPP] and Department of Public Health and Primary Care [PDPP], University of Cambridge, Cambridge, UK. <sup>11</sup> Kathleen Cuningham Foundation Consortium for Research into Familial Breast Cancer and Australian Ovarian Cancer Study (kConFab/AOCS): Queensland Institute of Medical Research, Brisbane, Australia [GC-T, JB, AOCS] and Peter MacCallum Cancer Center, Melbourne, Austalia [kConFab, AOCS], <sup>12</sup> Hannover-Minsk Breast Cancer Study (HMBCS); Department of Obstetrics and Gynaecology and Department of Radiation Oncology, Hannover Medical School, Hannover, Germany [NVB]; N.N. Alexandrov Research Institute of Oncology and Medical Radiology, Minsk, Belarus [NVB, NNA, IVZ]. <sup>13</sup> University of California Irvine Breast Cancer Study (UCIBCS): Department of Epidemiology, University of California Irvine, Irvine, California, USA <sup>14</sup> Gene Environment Interaction and Breast Cancer in Germany (GENICA): Dr. Margarete Fischer-Bosch-Institute of Clinical Pharmacology, Stuttgart, and University of Tübingen [HB, CJ]; Molecular Genetics of Breast Cancer, Deutsches Krebsforschungszentrum, Heidelberg [UH], Department of Internal Medicine, Evangelische Kliniken Bonn gGmbH, Johanniter Krankenhaus, Bonn [YDK], BGFA-Research Institute of Occupational Medicine of the German Social Accident Insurance, Institute of Ruhr University Bochum, Germany.<sup>15</sup> Bavarian Breast Cancer Cases and Controls (BBCC): University Breast Center [PAF, RS, MWB] and Institute of Human Genetics [ABE]. University Hospital Erlangen, Erlangen, Germany; Department of Gynecology and Obstetrics, David Geffen School of Medicine, Division of Hematology and Oncology, University of California at Los Angeles, CA, USA [PAF]. <sup>16</sup> Melbourne Collaborative Cohort Study (MCCS): Cancer Epidemiology Centre, The Cancer Council Victoria, Melbourne, Australia [GGG, GS, LB] and Centre for Molecular Environmental, Genetic, and Analytic Epidemiology, The University of Melbourne, Australia [DRE]. <sup>17</sup> Spanish National Cancer Centre Breast Cancer Study (CNIO-BCS) Study: Spanish National Cancer Centre, Madrid, Spain [RLM, JB, GP]; CIBERER, Spain [JB]; Monte Naranco Hospital, Oviedo, Spain [JIA] <sup>18</sup> Copenhagen Breast Cancer Study and Copenhagen General Population Study (CGPS): Department of Clinical Biochemistry, and Department of Breast Surgery, Herlev University Hospital, University of Copenhagen, Denmark. <sup>19</sup> Seoul Breast Cancer Study (SEBCS); Seoul National University College of Medicine, Seoul, Korea. <sup>20</sup> Kuopio Breast Cancer Project (KBCP): Institute of Clinical Medicine, Pathology and Forensic Medicine, University of Kuopio; Department of Pathology, University Hospital of Kuopio and Biocenter Kuopio, Kuopio, Finland [AM, V-MK]; Department of Oncology, University Hospital of Kuopio and Biocenter Kuopio, Kuopio, Finland; Department of Oncology, Vaasa Central Hospital, Vaasa, Finland [VK].<sup>21</sup> Polish Breast Cancer Study (PBCS): Division of Cancer Epidemiology and Genetics [MG-C, SJC, LAB], National Cancer Institute, Rockville, MD, USA; Department of Epidemiology and Cancer Prevention, The M. Sklodowska-Curie Cancer Centre and Institute of Oncology, Warsaw, Poland [JL]. 22 Genetic Epidemiology Study of Breast Cancer by Age 50 (GESBC): German Cancer Research Center (DFKZ), Heidelberg, Germany [JC-C]; Department of Gynecology and Obstetrics, Ulm Medical School, Ulm, Germany [SW-G]. 23 Amsterdam Breast Cancer Study (ABCS): Netherlands Cancer Institute, Amsterdam, The

Netherlands [AB, MKS, FevL, LJVtV] <sup>24</sup> Karloinska Breast Cancer Study (KARBAC): Dept. Molecular Medicine & Surgery, Dept. Oncology & Pathology, Karolinska Institutet, S 17176 Stockholm, Sweden [SM, AL] <sup>25</sup> Breast Cancer Association Consortium (BCAC): Cancer Research UK Genetic Epidemiology Unit, Department of Public Health and Primary Care, University of Cambridge, Cambridge, UK.

## Abstract

**Background**—Truncating mutations in *ATM* have been shown to increase the risk of breast cancer but the effect of missense variants remains contentious.

**Methods**—We have genotyped five polymorphic (MAF 0.9% to 2.6%) missense single nucleotide polymorphisms (SNPs) in *ATM* (S49C, S707P, F858L, P1054R, L1420F) in 26,101 breast cancer cases and 29,842 controls from 23 studies in the Breast Cancer Association Consortium (BCAC).

**Results**—Combining data from all five SNPs, the OR was 1.05 for being a heterozygote for any of the SNPs and 1.51 for being a rare homozygote for any of the SNPs with an overall trend OR=1.06 ( $P_{trend}$ =0.04). The trend OR among bilateral and familial cases was 1.12 (95% CI 1.02-1.23;  $P_{trend}$ =0.02).

**Conclusions**—In this large combined analysis, these 5 missense *ATM* SNPs were associated with a small increased risk of breast cancer, explaining an estimated 0.03% of the excess familial risk of breast cancer.

**Impact**—Testing the combined effects of rare missense variants in known breast cancer genes in large collaborative studies should clarify their overall contribution to breast cancer susceptibility.

## INTRODUCTION

Ataxia-telangiectasia (A-T) is an autosomal recessive disorder characterized by cerebellar ataxia, telangiectases, immune defects, radiosensitivity and a predisposition to malignancy (MIM #208900). The gene that is mutated in A-T, *ATM* (MIM #607585), encodes a protein kinase that plays a key role in cellular responses to DNA damage. The large majority of A-T cases are known to harbour mutations in *ATM* leading to a truncated or absent protein. Epidemiological studies of families of A-T patients have shown a two to fivefold increased risk of breast cancer for female relatives who are obligate heterozygous carriers of an A-T mutation (1, 2).

The increased risk of breast cancer in *ATM* mutation carriers has been confirmed by direct analysis of *ATM* mutations in breast cancer cases compared to controls. In a study of British familial breast cancer cases and controls, Renwick and colleagues identified nine mutations that result in premature termination or exon-skipping among 443 strongly familial cases (2.0%) compared to two in 551 controls (0.4%, P = 0.028) (3). They also found three cases and no controls who carried one of two missense variants for which there is strong a priori evidence of a pathogenic phenotype in individuals with A-T (V2424G or SV2855\_2856RI). Bernstein and colleagues identified seven heterozygotes for the V2424G missense variant among 3,743 population-based breast cancer cases (0.2%) unselected for family history and none among 1,268 controls (P = 0.1) (4). Based on the breast cancer history of first- and second-degree relatives of carrier cases, the breast cancer risk to age 70 years for heterozygotes was estimated to be 52% (95% CI: 28 - 80%; P <0.0001).

An association between other *ATM* variants, particularly amino acid substitutions that are not expected to be associated with A-T, and breast cancer has also been hypothesised (5),

but to date there has been little evidence to support this (6, 7). In a previous study we genotyped nine missense variants in *ATM* in 473 bilateral breast cancer cases and 2,463 controls as part of a high-throughput screen of 1,037 non-synonymous single nucleotide polymorphisms (nsSNPs) within candidate "cancer genes" (8). None of these variants was common, with minor allele frequencies (MAFs) in controls ranging from <0.1% (0/4924 chromosomes) to 2.4% (116/4926 chromosomes). Although no single *ATM* missense variant was significantly associated with breast cancer risk there was a significant trend in risk with increasing numbers of variant *ATM* SNPs (odds ratio (OR) = 1.27, 95% CI: 1.04 - 1.56; P<sub>trend</sub>= 0.02). We selected the 4 variants with MAF>1% (S707P (rs4986761), F858L (rs1800056), P1054R (rs1800057) and L1420F (rs1800058)) for further analysis in 26,101 invasive breast cancer cases and 29,842 controls in 23 studies within the Breast Cancer Association Consortium (BCAC). We also included a fifth variant (S49C (rs1800054)) with MAF 1.2% which was not genotyped in our previous analysis (8) but for which there had been some prior evidence of an association with breast cancer risk (OR = 1.13, 95% CI 0.99 - 1.30 P = 0.08) in an earlier BCAC analysis (9) that included a subset of the current studies.

## MATERIALS AND METHODS

#### Study populations and genotyping

Table 1s (supplementary online) summarises study details and genotyping platform for all studies that contributed data. Genotyping was performed by 5' nuclease assay (Taqman®), Sequenom iPLEX or Illumina Golden Gate technology. Taqman genotyping reagents were designed by Applied Biosystems as Assays-by-Design<sup>SM</sup> and distributed by the University of Cambridge group to each of the centres that used this technology. Genotyping was performed using the ABI PRISM 7900HT or 7500 Sequence Detection Systems according to manufacturer's instructions. For five studies, SNPs were genotyped using matrix assisted laser desorption/ionization time of flight mass spectrometry (MALDI-TOF MS) for the determination of allele-specific primer extension products using Sequenom's MassARRAY system and iPLEX technology (Sequenom, San Diego, CA, USA). The design of oligonucleotides was carried out according to the guidelines of Sequenom and performed using MassARRAY Assay Design software (version 1.0). In one study SNPs were genotyped using customised Illumina Sentrix Bead Arrays according to manufacturers instructions.

#### Quality control criteria

We applied BCAC standard quality control (QC) guidelines (http://www.srl.cam.ac.uk/ consortia/bcac/). In addition, we imposed a threshold of 99% for the call rate (compared with the standard threshold of 95%) and we excluded SNPs from studies where cluster plots, scored from 1 (poor) to 4 (good), scored by a single reader blinded to identifiers scored 2 or less. These more stringent thresholds were imposed because the minor alleles of these SNPs are rare, and therefore more susceptible to differential calling between cases and controls. S49C was not genotyped by 3 studies and data were excluded from analyses for QC criteria for 3 studies. S707P was not genotyped by 2 studies and data were excluded from analyses for QC criteria for 8 studies. F858L was genotyped by all studies; data were excluded from analyses for QC criteria for 1 study. P1054R was genotyped by all studies and data were excluded from analyses for QC criteria for 3 studies. L1420F was not genotyped by 2 studies and data were excluded from analyses for QC criteria for 8 studies for QC criteria for 8 studies. Full details of studies that contributed data for each SNP, numbers of cases and controls genotyped by each study and genotypes of cases and controls for each SNP are given in supplementary tables 1s and 2s online.

#### Statistical methods

The OR for each SNP and for being a carrier or rare homozygote for any SNP was tested using logistic regression with "study" as a stratifying covariate. To maximise the amount of data included in the analysis, SNPs that were not genotyped by a study or were excluded for QC criteria were coded as 0 for all subjects for the analysis of being a carrier or rare homozygote for any SNP. The effect of this will be to bias our OR estimate, marginally, towards the null.

LD metrics between SNPs ( $r^2$  and D', supplementary table 3s) were computed separately for each study using the Tagzilla module as implemented in GLU version 1.0a6. rs1800056 (F858L) and rs1800057 (P1054R) are correlated ( $r^2$ =0.38 – 0.71; supplementary table 3s), otherwise these rare SNPs are independent of each other ( $r^2$ <0.001). Maximum likelihood estimates of haplotype frequencies for the four alleles defined by F858L and P1054R (namely F858+P1054, F858+1054R, 858L+1054R, and 858L+P1054) were estimated in cases and controls separately and in each of the studies separately using HaploStats (http:// mayoresearch.mayo.edu/mayo/research/schaid\_lab/software.cfm); supplementary table 3s). ORs for F858+1054R, 858L+1054R and 858L+P1054 versus the common allele F858+ P1054 were estimated using unconditional logistic regression weighted for the phase assignment probability and with study as a stratifying covariate.

Statistical analyses were performed using STATA version 10 (State College, Texas, US). All P values reported are two-sided. Meta analyses (Figure 1) were carried out using the Metan routine within STATA, using inverse variance weighting of the study specific estimates. Cochran's Q statistic and the  $l^2$  statistic (10) to quantify the proportion of the total variation due to heterogeneity between studies were calculated.

### Results

The distribution of genotypes in cases and controls in each study for each *ATM* SNP is shown in table 2s (supplementary online). Subjects reporting ethnicities other than Caucasian were excluded (table 2s, footnote). The MAFs for each of the five SNPs genotyped in this analysis differed significantly (P<0.007, footnote table 1s) between the 22 studies of Caucasian subjects; medians (and ranges) were: S49C 1.2% (0.2%-1.7%), S707P 0.9% (0.6%-1.6%), F858L 1.5% (0.2%-2.4%), P1054R 2.6% (0.6%-3.7%) and L1420F 1.6% (0.2%-2.7%). In the one study in which the majority of subjects were of Asian ethnicity (SEBCS) three SNPs were monomorphic (S49C, S707P and F858L) and for the other two SNPs (P1054R and L1420F) there was only one carrier among 872 control subjects.

In the combined analysis across studies, the point estimates for each of the heterozygote ORs were above 1.0 and the estimates of the homozygote ORs were higher (table 1). The only significantly elevated OR was for L1420F homozygotes (OR=5.31, 95% CI 1.35-20.87). Two SNPs F858L (rs1800056) and P1054R (rs1800057) are correlated ( $r^2$  0.38 to 0.71 across studies, supplementary table 3s). The G allele of rs1800057 (1054R) is more common than the C allele of rs1800056 (858L; table 1s) thus the rare C allele of rs1800056 (858L) is almost completely contained on the rare G allele of rs1800057 (1054R) such that there are 3 main haplotypes for these two allelic variants (F858\_P1054, F858\_1054R and 858L\_1054R) and one extremely rare haplotype (858L\_P1054, supplementary table 3s). The trend OR estimates for each of the two haplotypes that carried the rare (C) allele of rs1800056 (858L\_1054R and 858L\_P1054) compared to the most common haplotype (F858\_P1054) were 1.05 (95% CI; 0.95-1.16, P=0.47) and 1.12 (95% CI; 0.61 – 2.05, P=0.71) respectively. The OR estimate for the haplotype that carried the rare (G) allele of

rs1800057 with the common T allele of rs1800056 (F858\_1054R) was 0.97 (95% CI; 0.86-1.10, P=0.65).

Combining data from all five SNPs, the OR was 1.05 for being a heterozygote for any of the SNPs and 1.51 for being a rare homozygote for any of the SNPs ( $P_{trend}=0.04$ ), with an overall OR<sub>trend</sub> of 1.06 (table 1) and no evidence of heterogeneity between studies (figure 1a, Cochrane Q=21.5 21df, P=0.43,  $\vec{P}=2.4\%$ ). Restricting the analysis to bilateral cases and those with a family history of breast cancer the overall OR<sub>trend</sub> was stronger (OR<sub>trend</sub>=1.12,  $P_{trend}=0.02$ , table 1) with no evidence of heterogeneity between studies (figure 1b, Cochrane Q=15.5 18df, P=0.62,  $\vec{P}=0\%$ )

## Discussion

Based on Swift's demonstration that carrier status for recessively inherited A-T is associated with a three-fold increase in risk of female breast cancer (1) and the more recent molecular validation of this observation (3) it is arguable that there is a high prior likelihood that a subset of polymorphic (MAF>1%) missense ATM variants will be associated with a modest increase in breast cancer risk. In our previous analysis of the combined effects of nine missense ATM variants (MAF <0.1% -2.4%) we demonstrated that on average, each missense ATM SNP was associated with an OR of 1.27 (95% CI: 1.04-1.56) in bilateral breast cancer cases, implying an OR of 1.13 (95% CI: 1.02–1.25) for cases with a single primary breast cancer (11, 12).

We selected five SNPs for further investigation. Despite restricting our follow-up analysis to SNPs with MAFs estimated to be 1% we did not have power to estimate individual effects for these SNPs or the effects of individual haplotypes. The aim of this present analysis was, therefore, to test the composite hypothesis that rare polymorphic ATM variants are, on average, associated with an increased risk of breast cancer. The five SNPs we genotyped in this analysis had a combined carrier frequency of ~12.5%; by genotyping 20,000 cases and 20,000 controls we had 90% power at 1% significance to detect an OR of 1.10.

Our OR estimate of 1.06 (95% CI 1.00-1.12) provides independent evidence that polymorphic missense variants in ATM are associated with a very modest increase in breast cancer risk, albeit at a nominal level of statistical significance (P=0.04). The stronger OR estimate for bilateral cases and cases with a family history of breast cancer (OR=1.12, 95% CI; 1.02-1.23, P=0.02) provides additional support.

We identified four previous studies (13-16) in which at least 100 Caucasian breast cancer cases and 100 Caucasian controls were genotyped and for which individual effect sizes for S49C (rs1800054), S707P (rs4986761), F858L (rs1800056), P1054R (rs1800057) or L1420F (rs1800058) were reported (table 2); we also obtained data for all five variants from the Wellcome Trust Case Control Consortium analysis (Table 2, (17)). For three of these (13, 14, 16), the case control series overlap with the current analysis; the other two (15, 17) do not support an association but are entirely consistent with a per SNP OR of 1.06. A recent analysis of rare (MAF<1%), evolutionarily unlikely missense substitutions in ATM (18) reported a per SNP OR estimate of 1.14 (0.90-1.44, P=0.39) for the combined effects of 121 variants in 1,948 cases and 1,852 controls. We also identified two studies that compared the frequency of ATM variants in bilateral breast cancer cases versus unilateral breast cancer cases. One (19) reported no difference in the frequency of missense variants between bilateral cases and unilateral cases overall but a longer median time to developing a second cancer in carriers of a missense variant who also received radiotherapy. In the other (20), a study of gene-environment interactions (WECARE study) in which bilateral cases were counter-matched to unilateral "controls" on the basis of exposure to radiotherapy, rare

(MAF<1%) A-T associated variants and those that were classified as deleterious according to the prediction algorithm SIFT (21) were associated with a non-significantly increased risk of a second breast cancer while those that were classified as tolerated and several of the more common missense variants were associated with a protective effect. For the linked variants F858L and P1054R, this was statistically significant (OR=0.5, 95% CI 0.3-1.0 and OR=0.5, 95% CI: 0.3-0.9 for F858L and P1054R respectively) raising the possibility of an interaction between radiotherapy and a subset of ATM variants.

It is not yet clear whether polymorphic (MAF>1%) missense variants in ATM and other validated breast cancer genes could make a contribution to explaining the excess familial risk of breast cancer. With a combined carrier frequency of 12.6% in Caucasian controls and an estimated average OR of 1.06, these five *ATM* variants explain 0.03% of excess familial risk of breast cancer, compared to between 0.07% and 1.7% explained by each of the common variants identified in recent GWA studies (7, 22-27). Rare SNPs (MAF 5%), however, account for a relatively large proportion of genetic variation (28); there are 83 rare missense SNPs in *ATM* listed in dbSNP (including the five genotyped in this study) and large numbers in other breast cancer genes. (29-32).

Testing the combined effects of rare missense variants in known breast cancer genes in large collaborative studies should, eventually, clarify their overall contribution to breast cancer susceptibility. Gutierrez-Enriquez et al (33) compared radiosensitivity of lymphoblastoid cell lines (LCLs) from breast cancer cases who were carriers of one or more rare allele(s) of S707P, F858L, P1054R and L1420F to LCLs from healthy controls. They demonstrated increased radiosensitivity in the LCLs from the breast cancer cases compared to controls generally, and specifically for the six LCLs from patients with at least one copy of the 858L + 1054R haplotype. Incorporating information from such functional assays and from next-generation *in silico* prediction algorithms may help to identify a subset that are most likely to be predictive of risk (34-36).

## Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

### Acknowledgments

The authors would like to thank the thousands of women who participated in this research. The HEBCS thanks Dr. Kirsimari Aaltonen and RN Hanna Jäntti for their help with the patient data and gratefully acknowledge the Finnish Cancer registry for the cancer data. The **GC-HBOC** thanks Sandrine Tchatchou for participating in genotyping. The SBCS thanks Sabapathy Balasubramanian, Simon Cross, Helen Cramp, and Dan Connley for their contribution to the study. The ABCFS thanks Maggie Angelakos, Judi Maskiell and Gillian Dite. The HABCS, and HMBCS gratefully acknowledge their German colleague Johann H. Karstens for his support of the breast cancer studies at Hannover Medical School. The ORIGO study thanks P.E.A. Huijts, E. Krol-Warmerdam, and J. Blom for patient accrual, administering questionnaires, and managing clinical information. The SEARCH study thanks the SEARCH and EPIC teams for recruitment of case patients and control subjects. kConFab thanks Heather Thorne, Eveline Niedermayr, all the kConFab research nurses and staff, the heads and staff of the Family Cancer Clinics, the Clinical Follow Up Study for its contributions to the resource, and the many families who contribute to kConFab. The AOCS Management Group (D Bowtell, G Chenevix-Trench, A deFazio, D Gertig, A Green, P Webb) gratefully acknowledges the contribution of all the clinical and scientific collaborators, the AOCS and the ACS Management Group (A Green, P Parsons, N Hayward, P Webb, D Whiteman), as well as all of the project staff, collaborating institutions and study participants. The GENICA study acknowledges Christian Baisch for the collection of clinical and histopathological data, Beate Pesch, Volker Harth and Thomas Brüning for their involvement in the recruitment of study subjects and the collection of epidemiological data as well as Christina Justenhoven for genotyping and data management. The CNIO-BCS thanks Primitiva Menendez from the Hospital Central Universitario de Asturias (HUCA-Oviedo), Pilar Zamora from the La Paz University Hospital in Madrid and Anna González-Neira, Charo Alonso and Tais Moreno from the CNIO. The KBCP is thankful Helena Kemiläinen and Aija Parkkinen for their contribution. The PBCS thanks Drs. Neonila Szeszenia- Dabrowska and Beata Peplonska of the Nofer Institute of Occupational Medicine (Lodz, Poland), Witold Zatonski of the Department of Cancer Epidemiology and Prevention, The M. Sklodowska-Curie Cancer Center and Institute of

Oncology (Warsaw, Poland), Mark Sherman from the Division of Cancer Epidemiology and Genetics of the National Cancer Institute, USA, Jeff P Struewing from the National Human Genetics Research Institute USA, and Pei Chao from Information Management Services (Sliver Spring MD, USA), for their valuable contributions to the study. The **GESBC** thanks Ursula Eilber for competent data coordination and management and Tanja Koehler for excellent technical assistance. **ABCS** acknowledges L. Braaf, R. van Hien, R. Tollenaar and other contributors to the "BOSOM" study and the support of H.B. Bueno-de-Mesquita for organising the release of control DNA.

#### Funding

The BBCS is funded by Cancer Research UK and Breakthrough Breast Cancer and acknowledges NHS funding to the NIHR Biomedical Research Centre, and the National Cancer Research Network (NCRN). The HEBCS study has been financially supported by the Helsinki University Central Hospital Research Fund, Academy of Finland [110663], the Finnish Cancer Society and the Sigrid Juselius Foundation. The GC-HBOC study was supported by Deutsche Krebshilfe [107054], the Center of Molecular Medicine, Cologne, the Helmholtz society and the Dietmar-Hopp Fondation. The SBCS was supported by Yorkshire Cancer Research and the Breast Cancer Campaign. The ABCFS was supported by the National Health and Medical Research Council of Australia (NHMRC) [145604], the United States National Institutes of Health (NIH) [CA102740-01A2], and by the United States National Cancer Institute, National Institutes of Health [CA-95-011] through cooperative agreements with members of the Breast Cancer Family Registry and principal investigators Cancer Care Ontario [CA69467], Columbia University [CA69398], Fox Chase Cancer Center [CA69631], Huntsman Cancer Institute [CA69446], Northern California Cancer Center [CA69417], University of Melbourne [CA69638]. The content of this manuscript does not necessarily reflect the views or policies of the National Cancer Institute or any of collaborating centers in the Breast CFR, nor does mention of trade names, commercial products, or organizations imply endorsement by the US Government or the Breast CFR. The ABCFS was initially supported by the NHMRC, the New South Wales Cancer Council and the Victorian Health Promotion Foundation. J.L.H. is an Australia Fellow of the NHMRC and Victorian Breast Cancer Research Consortium Group Leader, M.C.S and A.B.S are Senior Research Fellows of the NHMRC. Genotyping was in part supported by the Prostate Cancer Foundation of Australia. The MCBCS was supported by the NIH [CA122340, CA128978] an NIH breast cancer SPORE award to the Mayo Clinic [CA116201] and a Susan G. Komen Breast Cancer Foundation award. The HABCS has been supported by an intramural grant from Hannover Medical School and by a grant from the German Research Foundation [DFG, Do761/2-1]. The HMBCS was supported by short-term fellowships from the German Academic Exchange Program [to N.B], and the Friends of Hannover Medical School [to N.B.]. The ORIGO study was supported by the Dutch Cancer Society. The SASBAC study was supported by the Agency for Science, Technology and Research of Singapore (A\*STAR), the NIH and the Susan G. Komen Breast Cancer Foundation. SEARCH is funded by Cancer Research UK (CR-UK) programme grant [C490/A11021]. AMD is supported by CR-UK [C8197/A10865] &. P.D.P.P. is a Senior Clinical Research Fellow of CR-UK. kConFab is supported by grants from the National Breast Cancer Foundation, the NHMRC, the Queensland Cancer Fund, the Cancer Councils of New South Wales, Victoria, Tasmania and South Australia and the Cancer Foundation of Western Australia. The kConFab Clinical Follow Up Study was funded by the NHMRC [145684, 288704, 454508]. Financial support for the AOCS was provided by the United States Army Medical Research and Materiel Command [DAMD17-01-1-0729], the Cancer Council of Tasmania and Cancer Foundation of Western Australia and the NHMRC [199600]. G.C.T. and P.W. are supported by the NHMRC. The UCIBCS is supported by the National Institutes of Health, National Cancer Institute USA grant CA-58860 and the Lon V Smith Foundation grant LVS-18840. The GENICA study was supported by the German Human Genome Project and the German Federal Ministry of Education and Research (BMBF) [01KW9975/5, 01KW9976/8, 01KW9977/0 01KW0114]. Genotyping analysis was supported by the Robert Bosch Foundation of Medical Research, Stuttgart, Germany and the Deutsches Krebsforschungszentrum, Heidelberg, Germany. The work of the BBCC was partly funded by ELAN-Fond of the University Hospital of Erlangen. Infrastructure support for the MCCS recruitment and follow-up is provided by The Cancer Council Victoria, while cohort recruitment was partly funded by VicHealth. This work using the MCCS was supported by NHMRC [209057, 251533, 396414] and genotyping was in part supported by the Prostate Cancer Foundation of Australia. The CNIO-BCS was supported by the Genome Spain Foundation, the Red Temática de Investigación Cooperativa en Cáncer and grants from the Asociación Española Contra Cáncer and the Fondo de Investigación Sanitario [PI081120 to J.B., PI081583 to R.L.M.]. The CGPS was supported by the Chief Physician Johan Boserup and Lise Boserup Fund, the Danish Medical Research Council and Copenhagen University Hospital, Herlev Hospital. The SEBCS was supported the National Research and Development (R&D) Program for Cancer Control [0620410-1] and the Korea Health 21 R&D Project [AO30001], Ministry of Health and Welfare, Republic of Korea. KBCP is supported by grants from EVO funds of Kuopio University Hospital and the Finnish Cancer Foundation. The PBCS was funded by Intramural Research Funds of the National Cancer Institute, Department of Health and Human Services, USA. The GESBC was supported by the Deutsche Krebshilfe e. V. [70492]. Funding for the ABCS was provided by the Dutch Cancer Society [grants NKI 2001-2423; 2007-3839] and the Dutch National Genomics Initiative. KARBAC acknowledges funding from the Swedish Cancer Society and the Gustav V Julilee Foundation. The BCAC is funded by CR-UK [C1287/A10118, C1287/A7497]. Meetings of the BCAC have been funded by the European Union COST programme [BM0606]. D.F.E. is a Principal Research Fellow of CR-UK.

## References

- Swift M, Reitnauer PJ, Morrell D, Chase CL. Breast and other cancers in families with ataxiatelangiectasia. N Engl J Med. 1987; 316:1289–94. [PubMed: 3574400]
- Thompson D, Duedal S, Kirner J, et al. Cancer risks and mortality in heterozygous ATM mutation carriers. J Natl Cancer Inst. 2005; 97:813–22. [PubMed: 15928302]
- 3. Renwick A, Thompson D, Seal S, et al. ATM mutations that cause ataxia-telangiectasia are breast cancer susceptibility alleles. Nat Genet. 2006; 38:873–5. [PubMed: 16832357]
- 4. Bernstein JL, Teraoka S, Southey MC, et al. Population-based estimates of breast cancer risks associated with ATM gene variants c.7271T>G and c.1066-6T>G (IVS10-6T>G) from the Breast Cancer Family Registry. Hum Mutat. 2006; 27:1122–8. [PubMed: 16958054]
- Gatti RA, Tward A, Concannon P. Cancer risk in ATM heterozygotes: a model of phenotypic and mechanistic differences between missense and truncating mutations. Mol Genet Metab. 1999; 68:419–23. [PubMed: 10607471]
- Khanna KK, Chenevix-Trench G. ATM and genome maintenance: defining its role in breast cancer susceptibility. J Mammary Gland Biol Neoplasia. 2004; 9:247–62. [PubMed: 15557798]
- 7. Ahmed M, Rahman N. ATM and breast cancer susceptibility. Oncogene. 2006; 25:5906–11. [PubMed: 16998505]
- Johnson N, Fletcher O, Palles C, et al. Counting potentially functional variants in BRCA1, BRCA2 and ATM predicts breast cancer susceptibility. Hum Mol Genet. 2007; 16:1051–7. [PubMed: 17341484]
- Cox A, Dunning AM, Garcia-Closas M, et al. A common coding variant in CASP8 is associated with breast cancer risk. Nat Genet. 2007; 39:352–8. [PubMed: 17293864]
- Higgins JP, Thompson SG, Deeks JJ, Altman DG. Measuring inconsistency in meta-analyses. BMJ. 2003; 327:557–60. [PubMed: 12958120]
- Antoniou AC, Easton DF. Polygenic inheritance of breast cancer: Implications for design of association studies. Genet Epidemiol. 2003; 25:190–202. [PubMed: 14557987]
- Fletcher O, Johnson N, Palles C, et al. Inconsistent association between the STK15 F31I genetic polymorphism and breast cancer risk. J Natl Cancer Inst. 2006; 98:1014–8. [PubMed: 16849685]
- 13. Dork T, Bendix R, Bremer M, et al. Spectrum of ATM gene mutations in a hospital-based series of unselected breast cancer patients. Cancer Res. 2001; 61:7608–15. [PubMed: 11606401]
- Spurdle AB, Hopper JL, Chen X, et al. No evidence for association of ataxia-telangiectasia mutated gene T2119C and C3161G amino acid substitution variants with risk of breast cancer. Breast Cancer Res. 2002; 4:R15. [PubMed: 12473176]
- Bretsky P, Haiman CA, Gilad S, et al. The relationship between twenty missense ATM variants and breast cancer risk: the Multiethnic Cohort. Cancer Epidemiol Biomarkers Prev. 2003; 12:733– 8. [PubMed: 12917204]
- Stredrick DL, Garcia-Closas M, Pineda MA, et al. The ATM missense mutation p.Ser49Cys (c. 146C>G) and the risk of breast cancer. Hum Mutat. 2006; 27:538–44. [PubMed: 16652348]
- Burton PR, Clayton DG, Cardon LR, et al. Association scan of 14,500 nonsynonymous SNPs in four diseases identifies autoimmunity variants. Nat Genet. 2007; 39:1329–37. [PubMed: 17952073]
- Tavtigian SV, Oefner PJ, Babikyan D, et al. Rare, evolutionarily unlikely missense substitutions in ATM confer increased risk of breast cancer. Am J Hum Genet. 2009; 85:427–46. [PubMed: 19781682]
- Broeks A, Braaf LM, Huseinovic A, et al. The spectrum of ATM missense variants and their contribution to contralateral breast cancer. Breast Cancer Res Treat. 2008; 107:243–8. [PubMed: 17393301]
- 20. Concannon P, Haile RW, Borresen-Dale AL, et al. Variants in the ATM gene associated with a reduced risk of contralateral breast cancer. Cancer Res. 2008; 68:6486–91. [PubMed: 18701470]
- 21. Ng PC, Henikoff S. SIFT: Predicting amino acid changes that affect protein function. Nucleic Acids Res. 2003; 31:3812–4. [PubMed: 12824425]
- Easton DF, Pooley KA, Dunning AM, et al. Genome-wide association study identifies novel breast cancer susceptibility loci. Nature. 2007; 447:1087–93. [PubMed: 17529967]

- Hunter DJ, Kraft P, Jacobs KB, et al. A genome-wide association study identifies alleles in FGFR2 associated with risk of sporadic postmenopausal breast cancer. Nat Genet. 2007; 39:870–4. [PubMed: 17529973]
- 24. Stacey SN, Manolescu A, Sulem P, et al. Common variants on chromosomes 2q35 and 16q12 confer susceptibility to estrogen receptor-positive breast cancer. Nat Genet. 2007; 39:865–9. [PubMed: 17529974]
- Stacey SN, Manolescu A, Sulem P, et al. Common variants on chromosome 5p12 confer susceptibility to estrogen receptor-positive breast cancer. Nat Genet. 2008; 40:703–6. [PubMed: 18438407]
- 26. Zheng W, Long J, Gao YT, et al. Genome-wide association study identifies a new breast cancer susceptibility locus at 6q25.1. Nat Genet. 2009; 41:324–8. [PubMed: 19219042]
- Thomas G, Jacobs KB, Kraft P, et al. A multistage genome-wide association study in breast cancer identifies two new risk alleles at 1p11.2 and 14q24.1 (RAD51L1). Nat Genet. 2009; 41:579–84. [PubMed: 19330030]
- Frazer KA, Murray SS, Schork NJ, Topol EJ. Human genetic variation and its contribution to complex traits. Nat Rev Genet. 2009; 10:241–51. [PubMed: 19293820]
- Shattuck-Eidens D, Oliphant A, McClure M, et al. BRCA1 sequence analysis in women at high risk for susceptibility mutations. Risk factor analysis and implications for genetic testing. JAMA. 1997; 278:1242–50. [PubMed: 9333265]
- 30. Seal S, Thompson D, Renwick A, et al. Truncating mutations in the Fanconi anemia J gene BRIP1 are low-penetrance breast cancer susceptibility alleles. Nat Genet. 2006; 38:1239–41. [PubMed: 17033622]
- 31. Rahman N, Seal S, Thompson D, et al. PALB2, which encodes a BRCA2-interacting protein, is a breast cancer susceptibility gene. Nat Genet. 2007; 39:165–7. [PubMed: 17200668]
- 32. Whibley C, Pharoah PD, Hollstein M. p53 polymorphisms: cancer implications. Nat Rev Cancer. 2009; 9:95–107. [PubMed: 19165225]
- Gutierrez-Enriquez S, Fernet M, Dork T, et al. Functional consequences of ATM sequence variants for chromosomal radiosensitivity. Genes Chromosomes Cancer. 2004; 40:109–19. [PubMed: 15101044]
- Tavtigian SV, Greenblatt MS, Goldgar DE, Boffetta P. Assessing pathogenicity: overview of results from the IARC Unclassified Genetic Variants Working Group. Hum Mutat. 2008; 29:1261–4. [PubMed: 18951436]
- 35. Tavtigian SV, Greenblatt MS, Lesueur F, Byrnes GB. In silico analysis of missense substitutions using sequence-alignment based methods. Hum Mutat. 2008; 29:1327–36. [PubMed: 18951440]
- Goldgar DE, Easton DF, Byrnes GB, Spurdle AB, Iversen ES, Greenblatt MS. Genetic evidence and integration of various data sources for classifying uncertain variants into a single model. Hum Mutat. 2008; 29:1265–72. [PubMed: 18951437]

Fletcher et al.



Figure 1. Trend OR estimates for S49C, S707P, F858L, P1054R and L1420F combined by study in (a) all cases and all controls and (b) bilateral cases and cases with a family history of breast cancer and all controls

ORs and  $P_{trends}$  were calculated coding individuals who were common homozygotes for all genotyped SNPs as 0, individuals who were heterozygous for any rare variant as 1 and individuals who were rare homozygotes as 2 (statistical methods). Horizontal lines represent 95% CIs. The diamond represents the combined, fixed-effects estimate of the OR and 95% CI. The vertical line indicates the null effect (OR = 1.0).

Europe PMC Funders Author Manuscripts

Ц
50
14
Ę
pu
∕a
41
05
P1
្រា
[8]
8
ц.
7P
20
S
Ŭ
49
S
jõ
os
ati
S L
qq
ŏ
pu
Ite
q-
an
te
80
Zy
no
IOL
- <u>-</u>
ote
δõ
, ZC
er(
let
yŁ
lar.
ШС
un
Ś

SNP	MAF <sup>I</sup> (range)	N cases	Heterozygote OR (95% CI)	Homozygote OR (95% CI)	Trend OR (95% CI)
S49C	1.2 (0.2, 1.7)	22,011 25,865	$   \begin{array}{c}     1.08 \\     (0.95 - 1.22)   \end{array} $	1.44 (0.39 – 5.32)	1.08 (0.96 - 1.22)
S707P	0.9 (0.6, 1.6)	17,068 22,330	1.1 (0.96 – 1.26)	5.56 (0.58 - 53.02)	1.12 (0.97 - 1.28)
F858L	1.5 (0.2, 2.4)	26,455 29,785	1.03 (0.93 – 1.14)	1.58 (0.62 - 4.05)	1.04 (0.94 - 1.15)
P1054R	2.6 (0.6, 3.7)	24,191 27,048	1.01 (0.93 – 1.10)	1.04 (0.57 – 1.89)	1.01 (0.94 - 1.10)
L1420F	1.6 (0.2, 2.7)	18,607 22,565	1.05 (0.95 – 1.17)	5.31 (1.35 – 20.87)	1.07 (0.97 - 1.20)
		F858L P105	4R haplotype <sup>2</sup>		
858L+1054R	1.5 (0.2, 2.4)	24,191 27,048	1.04 (0.94 - 1.16)	1.67 (0.59 - 4.73)	1.05 (0.95 - 1.16)
F858+1054R	1.1 (0.4, 1.9)	24,191 27,048	0.98 (0.87 - 1.10)	0.72 (0.21 - 2.46)	0.97 (0.86 - 1.10)
858L+P1054	0.1 (0.04, 0.2)	24,191 27,048	1.06 (0.53 - 2.12)	1.93 (0.22 - 16.67)	1.12 (0.61 - 2.05)
		An	y SNP		
All cases	6.3 <i>3</i>	26,101 29,842	$1.05 \frac{4}{0.99 - 1.11}$	$\begin{array}{c} 1.51 \ \mathcal{5} \\ (0.95 - 2.41) \end{array}$	1.06 (1.00 – 1.12)
					$P_{trend} = 0.04$
Bilateral & familial		5,750 29,842	1.12 (1.02 - 1.23)	1.22 (0.55 – 2.72)	1.12 (1.02 - 1.23)
cases					$P_{trend} = 0.02$
CI: confidence interval	MAF: minor allele f	requency in	controls expressed as	a nerrentage OR- od	lds ratio N/A · No

Cancer Epidemiol Biomarkers Prev. Author manuscript; available in PMC 2011 March 01.

ot available b, Ĺ 5 5 5

*I* Median and range

<sup>2</sup>the OR for being a compound heterozygote was 1.04 (0.94 - 1.15). Due to the correlation between F858L and P1054R, however, 1587/1690 (93.9%) of compound heterozygotes were carriers of the 858L 1054R haplotype.

<sup>3</sup>To calculate the combined MAF we assumed all carriers of the rare allele of F858L also carried the rare allele of P1054R and independence between the other SNPs

 $^{4}_{\rm Heterozygote for any of the five SNPs}$ 

Fletcher et al.

Europe PMC Funders Author Manuscripts

ſт	
Ö	
-Ñ	
4	
5	
Γ,	
К	
4	
3	
Ξ	
Ъ	
î	
SI	
$\widetilde{\mathbf{v}}$	
<sub>2</sub>	
щ	
Ľ.	
F	
0	
5	
<b>.</b>	
υĴ	
õ	
4	
01	
Ä	
2	
ta	
Ja	
el-	
-9	
S	
ő	
- 2	
ğ	
>	
- <del>5</del> .	
Ξ.	
p,	
2	
- <u>C</u>	
aı	
Ч	
ě	
42	
Ë	
ę	
- S	
- <del>.</del> .	
- TS	
ರ	
.2	
e,	
- Id	
ų.	
0	
7	
ar	
В	
Ē	
n	
S	

Nos cases/controls1000/5001453/793110/110856/10421978/2361045/1476S49C $$ $   1.60(0.88-2.90)$ $1.87(1.14-3.11)$ $1.26(0.81-1.96)$ S49C $$ $     0.60(0.55-1.46)$ $1.06(0.55-1.46)$ S707P $2.4(1.0-5.6)$ $1.08(0.59-1.97)$ $0.66(0.05-5.90)$ $0.47(0.23-0.93)$ $1.25(0.80-1.94)$ $0.90(0.55-1.46)$ F85RL $1.4(0.7-2.7)$ $ 2.02(0.10-120.15)$ $2.03(1.05-3.90)$ $1.12(0.67-1.86)$ $0.66(0.40-1.10)$ P1054R $1.4(0.8-2.2)$ $1.35(0.85-1.98)$ $0.83(0.19-3.36)$ $  0.94(0.58-1.20)$ P1054R $1.5(0.9-2.7)$ $1.35(0.85-1.98)$ $0.83(0.19-3.36)$ $  -$ D1054R $1.5(0.9-2.7)$ $1.25(0.89-1.77)$ $0.75(0.25-2.25)$ $1.22(0.84-1.77)$ $0.93(0.63-1.35)$ Combined $1.56(1.11-2.20)$ $1.25(0.89-1.77)$ $0.75(0.25-2.25)$ $1.22(0.84-1.77)$ $0.93(0.78-1.18)$	Study (reference)	Dork (13)	Spurdle (14)	Bretsky (15)	Stredrick (USRT) (16)	Stredrick (Poland) (16)	WTCC (17)
S49C $    1.60(0.88-2.90)$ $1.87(1.14-3.11)$ $1.26(0.81-1.96)$ S707P $2.4(1.0-5.6)$ $1.08(0.59-1.97)$ $0.66(0.05-5.90)$ $0.47(0.23-0.93)$ $1.25(0.80-1.94)$ $0.90(0.55-1.46)$ F858L $1.4(0.7-2.7)$ $ 2.02(0.10-120.15)$ $2.03(1.05-3.90)$ $1.12(0.67-1.86)$ $0.66(0.40-1.10)$ P1054R $1.4(0.8-2.2)$ $1.35(0.85-1.98)$ $0.83(0.19-3.36)$ $0.33(1.05-3.90)$ $1.12(0.67-1.86)$ $0.66(0.40-1.10)$ P1054R $1.4(0.8-2.2)$ $1.35(0.85-1.98)$ $0.83(0.19-3.36)$ $  0.34(0.58-1.22)$ P1054R $1.5(0.9-2.7)$ $1.35(0.85-1.98)$ $0.66(0.05-5.90)$ $  0.33(1.05-3.90)$ $0.112(0.67-1.86)$ D1050R $1.56(1.11-2.20)$ $1.25(0.89-1.77)$ $0.75(0.25-2.25)$ $1.22(0.84-1.77)$ $0.93(1.04-1.81)$ $0.96(0.78-1.18)$	Nos cases/controls	1000/500	1453/793	110/110	856/1042	1978/2286	1045/1476
S707P $2.4 (1.0 - 5.6)$ $1.08 (0.59 - 1.97)$ $0.66 (0.05 - 5.90)$ $0.47 (0.23 - 0.93)$ $1.25 (0.80 - 1.94)$ $0.90 (0.55 - 1.46)$ F858L $1.4 (0.7 - 2.7)$ $ 2.02 (0.10 - 120.15)$ $2.03 (1.05 - 3.90)$ $1.12 (0.67 - 1.86)$ $0.66 (0.40 - 1.10)$ P1054R $1.4 (0.8 - 2.2)$ $1.35 (0.85 - 1.98)$ $0.83 (0.19 - 3.36)$ $0.33 (1.05 - 3.90)$ $1.12 (0.67 - 1.86)$ $0.64 (0.58 - 1.22)$ P1054R $1.4 (0.8 - 2.2)$ $1.35 (0.85 - 1.98)$ $0.83 (0.19 - 3.36)$ $  0.94 (0.58 - 1.22)$ P1054R $1.5 (0.9 - 2.7)$ $1.25 (0.85 - 1.98)$ $0.76 (0.05 - 5.90)$ $  0.94 (0.58 - 1.22)$ Combined $1.5 (0.1 - 2.20)$ $1.25 (0.89 - 1.77)$ $0.75 (0.25 - 2.25)$ $1.22 (0.84 - 1.77)$ $1.37 (1.04 - 1.81)$ $0.96 (0.78 - 1.18)$	S49C				1.60 (0.88 – 2.90)	1.87 (1.14 – 3.11)	$1.26\ (0.81 - 1.96)$
F858L         1.4 (0.7 - 2.7)          2.02 (0.10 - 120.15)         2.03 (1.05 - 3.90)         1.12 (0.67 - 1.86)         0.66 (0.40 - 1.10)           P1054R         1.4 (0.8 - 2.2)         1.35 (0.85 - 1.98)         0.83 (0.19 - 3.36)           0.84 (0.58 - 1.22)           P1054R         1.5 (0.9 - 2.7)         1.35 (0.85 - 1.98)         0.83 (0.16 - 5.90)           0.93 (0.53 - 1.32)           L1420F         1.5 (0.9 - 2.7)         1.25 (0.89 - 1.77)         0.75 (0.25 - 5.90)         1.22 (0.84 - 1.77)         1.37 (1.04 - 1.81)         0.96 (0.78 - 1.18)	S707P	2.4 (1.0 – 5.6)	1.08 (0.59 – 1.97)	$0.66\ (0.05 - 5.90)$	0.47~(0.23-0.93)	$1.25\ (0.80-1.94)$	$0.90\;(0.55-1.46)$
P1054R         1.4 (0.8 - 2.2)         1.35 (0.85 - 1.98)         0.83 (0.19 - 3.36)         —         —         0.84 (0.58 - 1.22)           L1420F         1.5 (0.9 - 2.7)         —         0.66 (0.05 - 5.90)         —         0.93 (0.63 - 1.35)           Combined         1.56 (1.11 - 2.20)         1.25 (0.89 - 1.77)         0.75 (0.25 - 2.25)         1.22 (0.84 - 1.77)         1.37 (1.04 - 1.81)         0.96 (0.78 - 1.18)	F858L	1.4 (0.7 – 2.7)	-	$2.02 \ (0.10 - 120.15)$	2.03 (1.05 – 3.90)	1.12 (0.67 – 1.86)	$0.66\ (0.40-1.10)$
L1420F         1.5 (0.9 - 2.7)         -         0.66 (0.05 - 5.90)         -         0.93 (0.63 - 1.35)           Combined         1.56 (1.11 - 2.20)         1.25 (0.89 - 1.77)         0.75 (0.25 - 2.25)         1.22 (0.84 - 1.77)         1.37 (1.04 - 1.81)         0.96 (0.78 - 1.18)	P1054R	1.4 (0.8 – 2.2)	1.35 (0.85 – 1.98)	0.83 (0.19 – 3.36)	—	-	$0.84\ (0.58-1.22)$
Combined         1.56 (1.11 - 2.20)         1.25 (0.89 - 1.77)         0.75 (0.25 - 2.25)         1.22 (0.84 - 1.77)         1.37 (1.04 - 1.81)         0.96 (0.78 - 1.18)	L1420F	1.5 (0.9 – 2.7)	_	$0.66\ (0.05 - 5.90)$	—	_	$0.93\ (0.63 - 1.35)$
	Combined	1.56 (1.11 – 2.20)	1.25 (0.89 – 1.77)	0.75 (0.25 – 2.25)	1.22 (0.84 – 1.77)	1.37 (1.04 – 1.81)	0.96 (0.78 – 1.18)