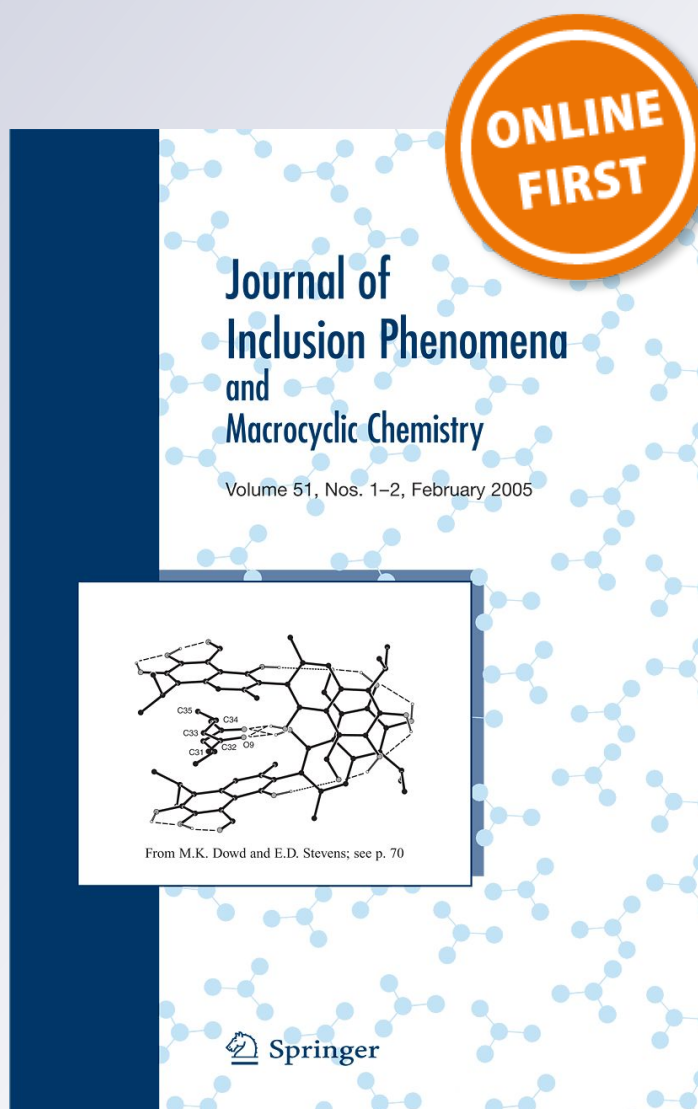# Equilibrium and structural characterization of ofloxacin–cyclodextrin complexation

Gergő Tóth, Réka Mohácsi, Ákos Rácz, Aura Rusu, Péter Horváth, Lajos Szente, Szabolcs Béni & Béla Noszál

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ORIGINAL ARTICLE

Equilibrium and structural characterization of ofloxacin–cyclodextrin complexation

**Gergő Tóth · Réka Mohácsi · Ákos Rácz ·
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Abstract The enantiomer-specific characterization of ofloxacin–cyclodextrin complexes was carried out by a set of complementary analytical techniques. The apparent stability constants of the ofloxacin enantiomers with 20 different cyclodextrins at two different pH values were determined to achieve good resolution capillary electrophoresis enantioseparation either to establish enantioselective drug analysis assay, or to interpret and design improved host–guest interactions at the molecular level. The cyclodextrins studied differed in the nature of substituents, degree of substitution (DS), charge and purity, allowing a systematic test of these properties on the complexation. The seven-membered beta-cyclodextrin and its derivatives were found to be the most suitable hosts. Highest stability and best enantioseparation were observed for the carboxymethylated-beta-cyclodextrin (DS ∼ 3.5). The effect of substitution pattern (SP) was investigated by molecular modeling, verifying that SP greatly affects the complex stability. Induced circular dichroism was observed and found especially significant on carboxymethylated-beta-cyclodextrin. The complex stoichiometry and the

geometry of the inclusion complexes were determined by ^{1}H NMR spectroscopy, including 2D ROESY techniques. Irrespective of the kind of cyclodextrin, the complexation ratio was found to be 1:1. The alfa-cyclodextrin cavity can accommodate the oxazine ring only, whereas the whole tricyclic moiety can enter the beta- and gamma-cyclodextrin cavities. These equilibrium and structural information offer molecular basis for improved drug formulation.

Keywords Ofloxacin · Levofloxacin · Cyclodextrin · Inclusion complex · ROESY · Capillary electrophoresis

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Introduction

Ofloxacin (OFL), chemically known as (±)-9-fluoro-2,3-dihydro-3-methyl-10-(4-methyl-1-piperazinyl)-7-oxo-7H-pyrido[1,2,3-de]-1,4-benzoxazine-6-carboxylic acid (Fig. 1), belongs to a fluoroquinolone class of chemotherapeutic agents with a broad spectrum of activity against gram-positive and gram-negative bacteria in vivo and in vitro [1–3]. OFL is a racemic compound; the antibacterial activities of the isomers are different. The effect of the S-(−) isomer is 128 times exceeding that of the R-(+)-isomer [4, 5]. The eutomer S-(−) isomer, levofloxacin (LEV) is also available on the drug market as a single enantiomer drug. Separation of the two OFL enantiomers is therefore substantial as shown by several studies. The most common separation method is a chiral ligand-exchange chromatography on a reversed phase HPLC column [6–9]. The other widely used technique is the chiral capillary electrophoresis (CE) using various cyclodextrins (CD) as chiral selectors [10–13].
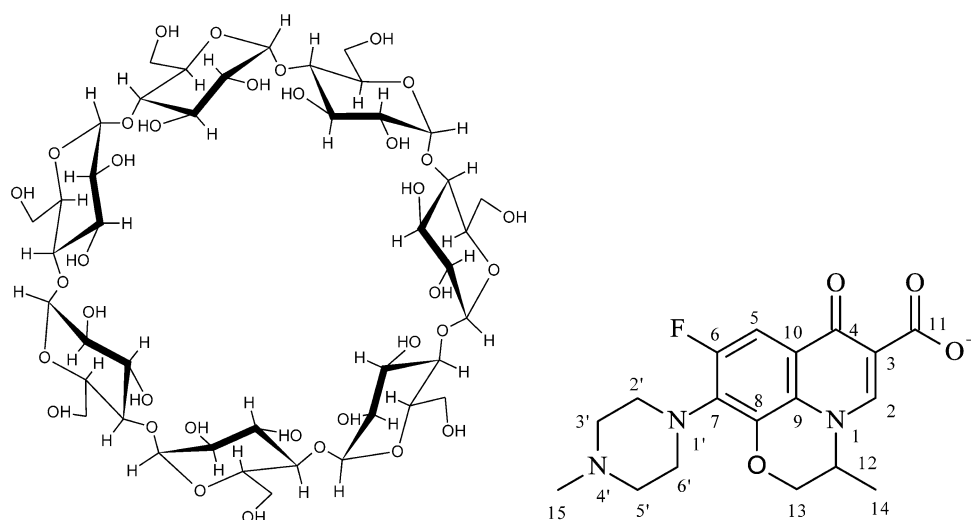
CDs, the cyclic oligosaccharides are composed of 6, 7 or 8, α-(1 → 4)-linked glucopyranose units in the respective α-, β- or γ-CDs, depicted often as a truncated cone (Fig. 1).

Fig. 1 The structure of native β-CD (*left*) and the numbered formula of the anionic OFL (*right*)

The H3, H5 and glycosidic oxygens are located inside the moderately hydrophobic cavity, while H1, H2, H4, and H6 are in the hydrophilic outer surface of the CD. As a consequence of these features CDs can encapsulate molecules inside their cavity through non-covalent interactions (electrostatic, van der Waals, hydrophobic, hydrogen bond) to form inclusion complex of host–guest type [14–16]. Nowadays not only drugs in native CD complex forms are available in the drug market but drugs in modified, ionic CD complexes such as voricinazole in sulfobutyl-ether-β-CD [17, 18]. CD complexation of pharmaceuticals can result in improved properties of the guest, such as solubility, stability, masking of undesirable properties, protection against oxidation, light-induced reactions and loss by evaporation [19, 20]. Hence the optimized OFL-CD complexes are promising candidates to develop new OFL formulations of improved quality. Beside the enantioseparation, solubility and light-induced decomposition of OFL can also be beneficially modified by CDs. Despite the above advantages of CDs, no literature data appeared on the systematic characterization of OFL-CD using a large number of CDs to obtain equilibrium and structural information, important in the drug formulation process. So far, only the β- and hydroxypropyl-β-CD-OFL complexes were characterized in detail. Koester et al. [21] determined that OFL encapsulation with β-CD increased the solubility 2.6 times but the rate of the photodegradation was not reduced. They determined the apparent 1:1 stability constant by a solubility method which was found to be $K = 152$ at pH 8.3. Their ^{1}H NMR and molecular modeling study showed that the *N*-methyl-piperazine ring is located in the inner cavity of the CD. Li et al. [22] prepared and characterized the β- and hydroxy-propyl-β-CD-OFL complexes. They determined the apparent 1:1 stability constant at 3 different pHs by fluorescence method, finding $K = 1640$ and $K = 500$ for β-, and hydroxypropyl-β-CD at pH 7.53, respectively. Their 2D

NOESY NMR studies showed that the tricyclic part of the molecule is in the CD-cavity, while the piperazine moiety is located outside. It can be seen that some data in the literature are contradictory ([21] vs. [22]).

Our aim was to characterize the OFL-CDs complexes by various techniques at two pHs, where the respective average charge of OFL is +1 and −1 [23], and thorough characterization of the complexes by CE, circular dichroism, molecular modeling, 1D and 2D NMR techniques. Here we report the equilibrium and structural parameters of the inclusion complexes formed between 20 CDs and two OFL enantiomers in two charged forms, and the resulting optimized separation of R- and S-OFLs.

Materials and methods

Materials

All native CDs and their derivatives (Table 1) were the products of Cyclolab R&D Ltd. (Budapest, Hungary). Racemic OFL was purchased from Smruthi Organics Ltd. (Solapur, India), LEV (S-OFL) was obtained from Sigma-Aldrich Co. (St. Louis, USA). H_3PO_4, KH_2PO_4 Na_3PO_4, NaOH and HCl were used for buffer preparation using chemicals of analytical grade from commercial suppliers. DMSO, the electro-osmotic flow (EOF) marker in the CE experiment was purchased from Reanal (Budapest, Hungary). D_2O was obtained from Sigma-Aldrich. All solutions were prepared from bidistilled Millipore water (specific conductivity: 1.1 μScm^{-1}).

CE experiments

CE experiments were performed on a Hewlett Packard ^{3D}CE instrument equipped with a photodiode array detector. An untreated fused-silica capillary from Agilent was used.

Table 1 Properties and abbreviations of the 20 investigated CDs

Compound	Abbreviation	Substituents	Degree of substitution	Purity (%)	Molecular mass
α-Cyclodextrin	α-CD	–	0	100	972.9
β-Cyclodextrin	β-CD	–	0	100	1135.0
γ-Cyclodextrin	γ-CD		0	100	1297.2
Hydroxypropyl-α-CD	HPαCD	$-CH_2-CH(OH)-CH_3$	3	>97	1151.0
Hydroxypropyl-β-CD	HPβCD DS3	$-CH_2-CH(OH)-CH_3$	3	>97	1309.0
Hydroxypropyl-β-CD	HPβCD DS4.5	$-CH_2-CH(OH)-CH_3$	4.5	>97	1396.4
Hydroxypropyl-β-CD	HPβCD DS6.3	$-CH_2-CH(OH)-CH_3$	6.3	>98	1497.6
Hydroxypropyl-γ-CD	HPγCD	$-CH_2-CH(OH)-CH_3$	3	>97	1471.4
Methyl-β-CD	RAMEB	$-CH_3$	12	>98	1303.4
Trimethyl-β-CD	TRIMEB	$-CH_3$	21	>98	1429.6
Carboxymethyl-α-CD	CMαCD	CH_2-COO^-	3.5	>95	1212.9
Carboxymethyl-β-CD	CMβCD	CH_2-COO^-	3.5	>95	1375.1
Carboxymethyl-γ-CD	CMγCD	CH_2-COO^-	3.5	>95	1537.1
Carboxyethyl-β-CD	CEβCD	$(CH_2)_2-COO^-$	3	>95	1351.2
Sulfopropyl-α-CD	SPαCD	$(CH_2)_3SO_3^-$	2	>90	1261.2
Sulfopropyl-β-CD	SPβCD	$(CH_2)_3SO_3^-$	2	>90	1423.3
Sulfopropyl-γ-CD	SPγCD	$(CH_2)_3SO_3^-$	2	>90	1585.5
Sulfobutyl-β-CD	SBβCD	$(CH_2)_4SO_3^-$	4	>95	1767.7
Succinyl-β-CD	SuccβCD	$-CO(CH_2)_2COO^-$	3.5	>95	1535.3
6-Monoamino-6-monodeoxy-β-CD	MAβCD	$-NH_2$	1	>97	1170.5

Conditioning of new capillaries was conducted by flushing with 1 M NaOH for 30 min followed by 0.1 M NaOH and buffer for 60 min each. The capillary cassette temperature was set to 25 °C and the voltage to +20 kV. UV detection was performed at 200, 210, 230 and 254 nm and samples were run in triplicate. Between the measurements capillaries were flushed with water (1 min), 0.1 M NaOH (0.5 min), water (1 min) and BGE (3 min). The running buffers were PO_4^{3-}/HPO_4^{2-} at pH 11 and 0.05 M phosphoric acid/$H_2PO_4^-$ at pH 3.5. The ionic strength was constant 0.15 M in both cases. The stock OFL solution (1 mM) contained 0.001 % (v/v) DMSO as EOF marker. Samples were injected hydrodynamically at a pressure of 50 mbar for 3 s.

NMR experiments

All NMR measurements were carried out on a Varian DDR spectrometer (599.9 MHz for ^{1}H) with a 5 mm inverse-detection gradient probehead. Standard pulse sequences and processing routines available in VnmrJ 2.2C/Chempack 4.0 were used. All NMR experiments were carried out in D_2O (15 mM phosphate buffer). The chemical shifts were referenced to internal methanol ($\delta = 3.300$ ppm). The average extent of penetration and the direction of inclusion were investigated by two dimensional phase-sensitive rotating frame nuclear Overhauser effect spectroscopy (ROESY). In ROESY experiments the samples contained 1 mM CDs and 2 mM OFL at pH* 3.5. Different mixing times (300 ms, 500 ms) were applied in these experiments.

Circular dichroism measurements

The spectra were recorded on a Jasco J720 Spectropolarimeter using different cylindrical quartz cells. The spectra were accumulated three times with a bandwidth of 1 nm and a scanning step of 0.2 nm at a scan speed of 50 nm/min. In this experiment the samples contained 1 mM OFL and 2 mM various CDs.

Molecular modeling

Modeling of α-CD-OFL, β-CD-OFL, HPβCD-OFL DS4 with 3 different substitution patterns and HPβCD-OFL DS6 complexes were performed using MMFF94 force field implemented in TINKER program. The initial structures of the CDs were based on structural data found in the literature [24] and on structures 2ZYM and 3CGT in the Brookhaven Protein Database [25, 26]. OFL was placed into the cavity at its wider rim in two orientations: either with the carboxyl group or the N-methyl-piperazine group inside. Each structure was subject to energy minimization with 0.001 RMS gradient criteria, simulating aqueous environment with setting $\varepsilon = 78.3$. Molecular dynamic calculations were

performed using the Velocity-Verlet algorithm in NVT ensemble with 30 Å sized cubic box periodic boundary conditions, $T = 298$ K (10000 steps, with 1.0 fs step increment and 0.1 ps snapshot interval).

The resulting 100 structures/guest orientation/charge state/CD were re-optimized and according to the energy values of the optimized structures, the lowest energy ones were taken into account for the interaction energies. The calculated interaction energy was the difference between the energy of the complex and the sum of the energy of the host and the guest.

Results and discussion

CE experiments: stability constants and enantioseparation study

The binding constants between the analyte and the chiral selector are of fundamental interest to understand the inclusion behavior (information on the analyte–ligand affinity and understanding the molecular interactions) [27, 28]. The crucial requirement of CE in binding analysis is that at least one of the interacting species has to carry a charge. Assuming 1:1 averaged stoichiometry for the complex between the host CD and the guest, the effective mobility depends on the CD concentration according to Eq. (1) [29].

$$\mu_{eff} = \frac{\mu_{free} + \mu_{cplx}K[CD]}{1 + K[CD]} \tag{1}$$

where μ_{eff} is the effective mobility of the guest at the actual CD concentration, μ_{free} and μ_{cplx} are the effective mobilities of the free and complexed ligands, respectively, while K is the apparent averaged complex stability constant.

Apparent complex stability constants were determined according to the x-reciprocal method by plotting the data in the form of $\mu_{eff}^i - \mu_{free}/[CD]$ versus $\mu_{eff}^i - \mu_{free}$, yielding $-K$ as slope. Beside the x-reciprocal method several other linearization functions are published in the literature [30, 31]. However the x-reciprocal one is a simple, fast and robust method which is ideal for fast screening to determine a large number of binding constants. Due to Wren's theory, the optimal CD concentration for the enantioseparation can be calculated as:

$$[CD]_{opt} = \frac{1}{\sqrt{K_R K_S}} \tag{2}$$

where K_R and K_S are the stability constants of the inclusion complexes of the R and S enantiomers, respectively [32].

For the determination of the binding constants, various concentrations of CDs (ranging from 3 to 50 mM) were added to the running buffers. The estimated stability constants and enantioresolution values (calculated at the optimal concentrations according to Wren's theory) are compiled in Table 2. Resolution of the enantioseparation was calculated by the following equation:

$$R_s = 1.18\left(\frac{t_2 - t_1}{w_1 + w_2}\right) \tag{3}$$

where t_1 and t_2 are the migration times of the enantiomers, and w_1 and w_2 are the extrapolated peak widths at the baseline.

Table 2 shows that the most stable complex with the native CDs is formed with β-CD, indicating that it has the most suitable cavity size for OFL. The decreasing trend in stability values follows the β-, α-, γ-CD order for all derivatives. For native CDs higher stability constants were measured at acidic pH. This observation shows that the –COOH form of OFL is more appropriate for the complexation than the anionic carboxylate form. The methylation of native CDs decreased the stability constants in our investigations; however the hydroxypropyl substitution (DS3) slightly increased the stability. This can be explained by assuming more secondary bonds between the host and guest molecules. Higher degree of substitution (DS6.3) decreased the stability as the substituents inhibited the inclusion process. Among the neutral CDs, only the HPα- and HPβCD derivatives were able to resolve the OFL enantiomers. Baseline enantioseparation could be achieved with each HPβCD. The highest enantioresolution, however, could be observed for HPβCD DS6.3. The most stable complexes and the best enantioresolution were detected with negatively charged CDs especially CMβCD at pH 3.5. In this inclusion complex additional ionic interaction can develop between the host and guest molecules. The relatively high stability of the MAβCD OFL complex at pH 8 can also be interpreted in terms of ionic interaction but no enantioseparation could be achieved with the positively charged host. It is noteworthy that R-OFL showed stronger binding in each OFL-CD system.

Based on these observations the enantioseparation was developed with CMβCD to achieve high resolution in 10 min, optimizing the buffer pH, concentration, CMβCD concentration, applied voltage, and the injection parameters. Figure 2 shows the electropherogram of the optimized enantioseparation. This enantioseparation could serve as a starting point in the development of a new enantioselective method for the determination of OFL enantiomers in pharmaceutical formulations.

Molecular modeling study

By molecular modeling the accordance between the computer calculation and the experimental data and the

Table 2 The averaged stability constants of OFL-CD complexes with the enantioresolutions (R_s) of the enantiomers at two pHs

CD	pH 3.5 ($K_{H_2OFL^+-CD}$)			pH 11 (K_{OFL^--CD})
α	$K = 19.5 \pm 0.9$			$K = 16 \pm 0.7$
β	$K = 125.2 \pm 4$			$K = 102.3 \pm 3.5$
γ	$K = 17.2 \pm 0.9$			$K = 12.3 \pm 0.8$
RAMEB	$K = 100.3 \pm 5.1$			$K = 112.2 \pm 3.2$
TRIMEB	$K = 90.6 \pm 7.2$			$K = 108.6 \pm 1.2$
HPα DS3	$K_s = 22.3 \pm 2.2$	$K_R = 26.7 \pm 2.5$	$R_s = 1.32$	$K = 28.1 \pm 1.7$
HPβ DS3	$K_s = 132.5 \pm 1.8$	$K_R = 138.4 \pm 1.6$	$R_s = 1.57$	$K = 147.8 \pm 3.9$
HPγ DS3	$K = 20.3 \pm 0.6$			$K = 24.5 \pm 8.5$
HPβ DS4.5	$K_s = 122.3 \pm 2.1$	$K_R = 129.7 \pm 1.8$	$R_s = 1.61$	$K = 130.5 \pm 5.2$
HPβ DS6.3	$K_s = 108.7 \pm 3.4$	$K_R = 120.6 \pm 3.8$	$R_s = 1.85$	$K = 120.3 \pm 3.7$
CMα DS3	$K_s = 83.3 \pm 1.4$	$K_R = 89.6 \pm 1.5$	$R_s = 1.90$	$K = 32.1 \pm 0.8$
CMβ DS3.5	$K_s = 251.2 \pm 0.7$	$K_R = 513.4 \pm 1.1$	$R_s = 7.85$	$K = 82.5 \pm 1.5$
CMγ DS3	$K_s = 41.6 \pm 1.1$	$K_R = 48.2 \pm 0.9$	$R_s = 1.53$	$K = 24.2 \pm 0.2$
CEβ DS3	$K_s = 105.3 \pm 5.1$	$K_R = 117.2 \pm 4.8$	$R_s = 1.81$	$K = 60.5 \pm 2.4$
SPα DS2	$K_s = 50.3 \pm 1.8$	$K_R = 54.7 \pm 1.5$	$R_s = 1.70$	$K = 28.1 \pm 1.6$
SPβ DS2	$K_s = 131 \pm 0.9$	$K_R = 158.3 \pm 1.3$	$R_s = 3.05$	$K = 68.2 \pm 2.0$
SPγ DS2	$K_s = 26.3 \pm 2.9$	$K_R = 29.4 \pm 3.2$	$R_s = 1.47$	$K = 20.3 \pm 0.4$
SBβ DS4	$K_s = 230.6 \pm 8.2$	$K_R = 470.5 \pm 8.5$	$R_s = 6.50$	$K = 75.5 \pm 3.2$
Succβ DS4	$K_s = 105.3 \pm 2.1$	$K_R = 114.2 \pm 2.4$	$R_s = 1.54$	$K = 50.2 \pm 0.4$
MAβ	$K = 70.1 \pm 7.1$			$K = 145.2 \pm 10.5$

K_S and K_R are the stability constants of S-OFL (LEV) and R-OFL, respectively. S-OFL was usually the first-migrating enantiomer

complexation behavior of CD with different substitution patterns was investigated.

The binding energy between different neutral CDs and two different protonated forms of OFL (H_2OFL^+, OFL^-) was investigated by molecular modeling. The values of the energies are summarized in Table 3.
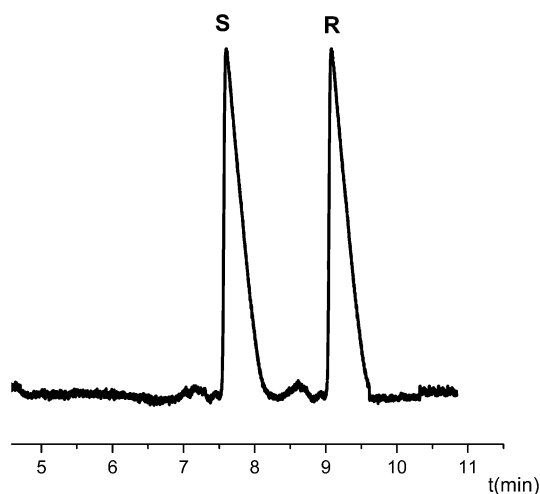

Fig. 2 Optimized enantioseparation of OFL enantiomers with CMβCD (electrolyte 20 mM phosphate buffer with 3 mM CMβCD pH = 4.4, applied voltage 15 kV; injection: 50 mbar for 4 s; total length of the capillary 30 cm (21.5 cm effective length), the OFL concentration was 50 μg/ml.)

For α-CD and β-CD good correlation was found between the experimental observations and the calculated energy values. Two different degrees of substitution were investigated in the case of HPβCD. The energy values were in agreement with the observations in the CE section: an increased degree of substitution decreased the stability of the inclusion complex. This result points out that molecular modeling can predict even subtle binding affinity differences between host and guest molecules [33]. Thus prediction can reduce the costs of encapsulation development. The hydroxyalkylation of native CD results in mixture of positional isomers, therefore molecular interaction with hosts possessing well-defined substitution pattern may be difficult to study experimentally. In the in silico part of our

Table 3 Total energy changes of complex formation for the interaction of OFL and various CDs

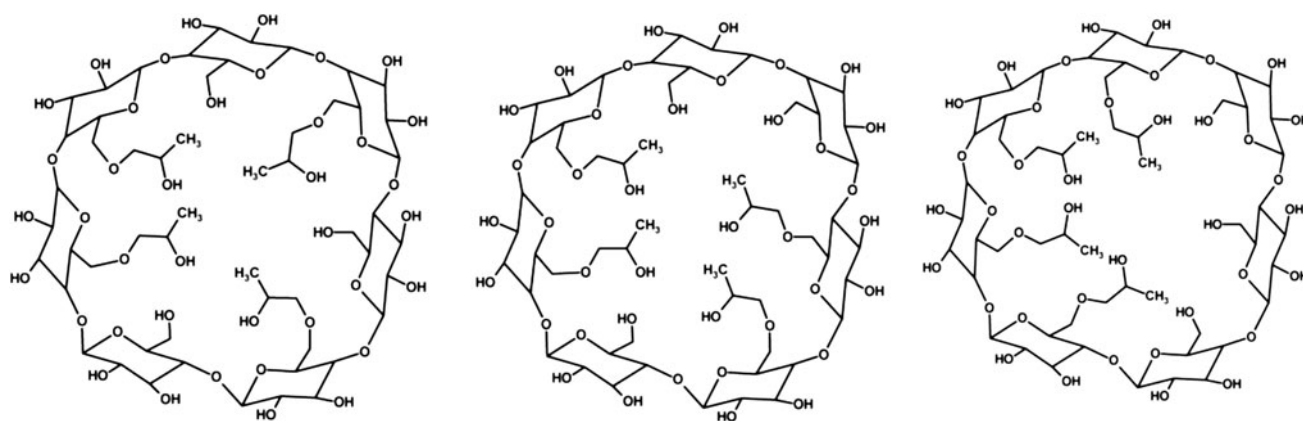|	E (H_2OFL^+) (kcal/mol)	E (OFL^-) (kcal/mol)
α-CD	−13.8	−13.4
β-CD	−21.9	−19.8
HPβ DS4[*]	−17.3	−17.2
HPβ DS4[*]	−18.4	−19.9
HPβ DS4[*]	−21.0	−21.3
HPβ DS6	−16.3	−15.9

* The structures of these CDs are illustrated in Fig. 3

Fig. 3 The structure of the three HPβCDs with different substitution patterns (HPβCD DS4[a], HPβCD DS4[b] and HPβCD DS4[c], respectively)

work three different substitution patterns of HPβCD DS4 were also investigated. Four hydroxypropyl groups were attached to β-CD primary hydroxyl groups in various positions as shown by earlier works [34, 35]. The structure of these three HPβCDs with different substitution patterns can be seen in Fig. 3. These CDs are hydroxyalkylated only at the primary side, which compounds are hardly available by synthesis due to concomitant hydroxyalkylation on the secondary hydroxyl groups. However, these hypothetical guests are suitable for in silico investigation on the role of substitution pattern on guest complexation.

The interaction energy values of these complexes show that the substitution pattern is an important factor influencing complex stability as earlier studies also proved experimentally [36, 37]. The complex formation among the HPβCDs is most favorable energetically in the case of HPβCD DS4[c] (see Table 3). In this CD the substituents are in each other's closest vicinity so the molecule is not overcrowded and OFL can slip into the CD cavity.

Circular dichroism study

It is known that native CDs are inactive in the UV–VIS absorption region. Nevertheless, an achiral chromophore guest may exhibit an induced circular dichroism (ICD) upon complex formation. The magnitude and direction of ICD signs depend on the complex stability, the dynamics within the complex, the extent of the chromophore inclusion in the CD cavity and the orientation of its electronic transition moment relative to the *n*-fold rotation axis of CD [38, 39].

In this investigation α-, β-, γ-, HPβ and CMβCD complexes were examined. In our study significant ICD signal could be observed for CMβCD only as can be seen in Fig. 4. This fact is in accordance with the low complex stability of γ- and α-CDs as well as the magnitude of inclusion of α-CD (see NMR section). The main reason of

the very low ICD signal intensity with β- and HPβCDs relies in the unfavorable dynamics within the complex. Matsuura et al. observed [40] that rigid, tricyclic analogues of benzophenone, like fluorenone and anthrone, which are similar to OFL gave a very weak ICD while benzophenone can attain a twisted (optically active) conformation within the β-CD cavity. A larger positive ICD can be observed between the OFL and CMβCD due to the enhanced complex stability and the ionic interaction. This positive ICD sign indicates that the aromatic ring system is situated in the CD cavity in axial orientation, in agreement with the Harata rules [41].

NMR studies

Job plot titration

The continuous variation method was adopted to verify the stoichiometry of the complexes. The ^{1}H-NMR chemical shifts were measured at different [OFL]/[CD] ratios while the total [OFL] + [CD] was kept constant. The calculated quantities ($\Delta\delta$ [OFL] or [CD]) were plotted as a function of molar ratio. The resulting plots showed a maximum at 0.5 indicating 1:1 binding stoichiometry (invariantly of the actual CD derivative). These results confirm the previously reported findings of Li et al. by fluorescence method [22]. Representative Job plot curves of the two OFL enantiomers and the CMβCD are shown in Fig. 5.

^{1}H NMR study

The inclusion complex formation can be proved from the ^{1}H NMR chemical shift changes. Figure 6 illustrates the aromatic peak changes of OFL upon complexation with CMβCD. It can be seen that at higher molar ratios of CMβCD the aromatic protons of the two enantiomers are separated. This method can be suitable to distinguish the

Table 4 ^{1}H NMR chemical shifts of OFL protons in the absence and presence of various CDs in 1:2 ratio in phosphate buffer at pH* 3.5

Proton	OFL	αOFL	βOFL	γOFL	HP/βOFL	SBβOFL	CEβOFL	CMβOFL
H2	8.670 s	8.668 Δ = −0.002	8.660 Δ = −0.010	8.667 Δ = −0.003	8.661 Δ = −0.011	8.651 s Δ = −0.019 8.635 s Δ = −0.035	8.663 Δ = −0.007	8.649 s Δ = −0.021 8.631 s Δ = −0.039
H5	7.454 d	7.452 Δ = −0.002	7.501 Δ = 0.047	7.460 Δ = 0.006	7.505 Δ = 0.051	7.520 Δ = 0.066 7.500 Δ = 0.046	7.465 Δ = 0.011	7.522 d Δ = 0.068 7.502 d Δ = 0.048
H2′H6′	3.551 m	3.551 Δ = 0.000	3.554 Δ = 0.003	3.552 Δ = 0.001	3.555 Δ = 0.004	3.556 Δ = 0.005	3.555 Δ = 0.004	3.557 Δ = 0.006
H3′H5′	3.631 m	3.632 Δ = 0.001	3.628 Δ = −0.003	3.6330 Δ = −0.001	3.633 Δ = 0.002	3.630 Δ = 0.001	3.629 Δ = −0.002	3.630 Δ = −0.001
H13a	4.417 dd	4.411 Δ = −0.006	4.430 Δ = 0.013	4.419 Δ = 0.002	4.433 Δ = 0.016	4.456 Δ = 0.039 4.436 Δ = 0.019	4.423 Δ = 0.006	4.459 Δ = 0.042 4.439 Δ = 0.022
H13b	4.572 dd	4.567 Δ = −0.005	4.582 Δ = 0.010	4.569 Δ = −0.003	4.584 Δ = 0.012	4.587 Δ = 0.015	4.580 Δ = 0.008	4.589 Δ = 0.017
H14	1.531 d	1.525 Δ = −0.006	1.533 Δ = 0.002	1.527 Δ = −0.004	1.532 Δ = 0.001	1.537 Δ = 0.006	1.534 Δ = 0.003	1.538 Δ = 0.007
H15	2.941 s	2.941 Δ = 0.000	2.942 Δ = 0.001	2.943 Δ = 0.002	2.940 Δ = −0.001	2.943 Δ = 0.002	2.941 Δ = 0.000	2.939 Δ = −0.002

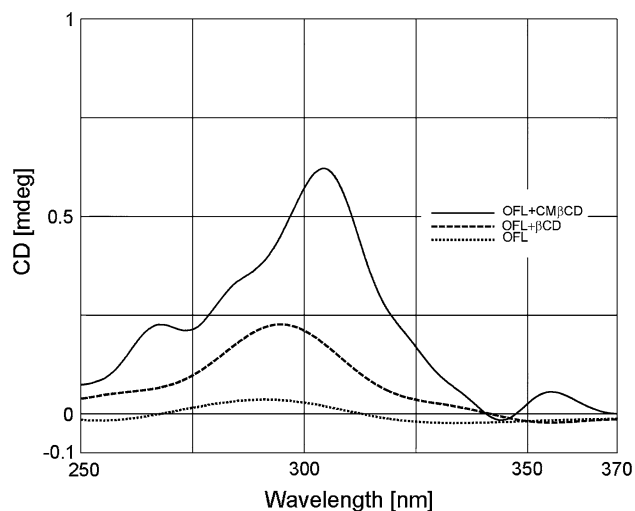pH* is the appropriate notation of the measured pH in D20

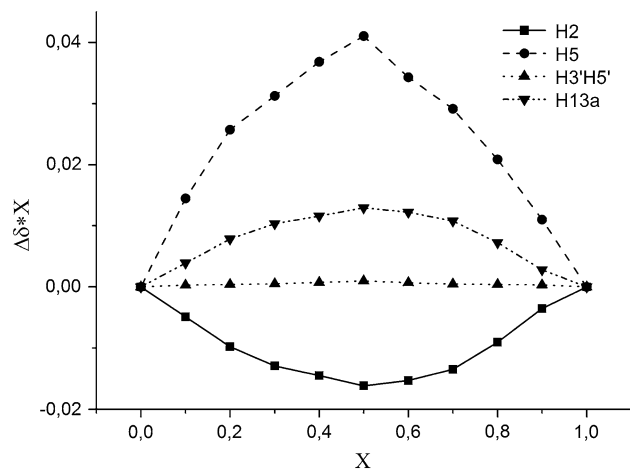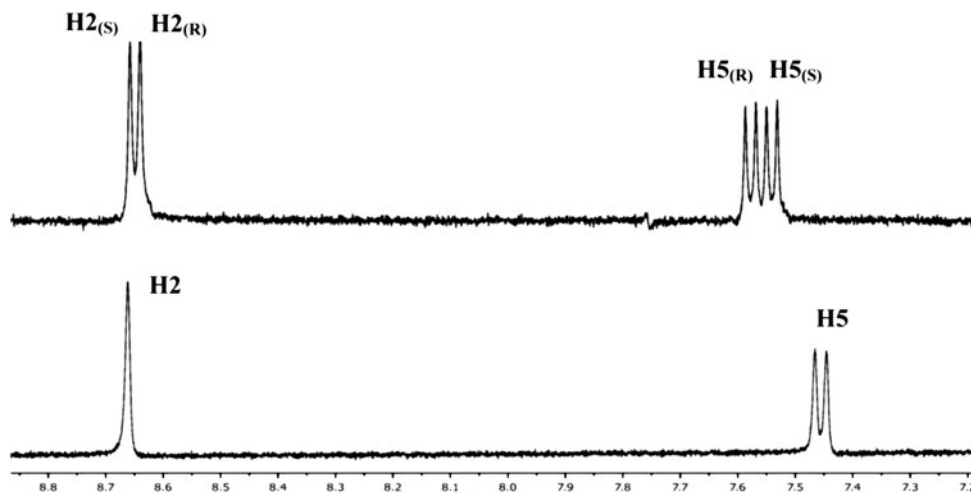**Fig. 4** The induced circular dichroism signals of OFL and its CD complexes

Fig. 5 Representative job plot curves of the two OFL enantiomers with CMβCD

OFL enantiomers. The ^{1}H chemical shifts of OFL in the absence and presence of 8 different CDs are indicated in Table 4.

Table 4 shows that complexation-induced chemical shift changes occur most with β-CDs due to the high complex stability. H5, H2 and H13 protons appear at higher chemical shifts as opposed to H2′, H6′, H3′, H5′ and H15 piperazine protons indicating that the piperazine ring is outside the CD cavity, while the aromatic ring system is inside the cavity. Minor changes occur only with γ-CD in the aromatic region. In contrast, the aromatic proton do not show any change with α-CD but the protons in the oxazine ring show a downfield shift which indicates that only the oxazine ring is in the cavity of α-CD. To verify these hypotheses 2D ROESY NMR experiments were carried out with α, β, γ and CMβCD.

2D ROESY NMR results

The 2D ROESY NMR experiment is suitable to obtain information about the spatial proximity between atoms of the host and guest molecules by observing the intermolecular dipolar cross-correlations. The ROE is a manifestation of cross relaxation between two nonequivalent nuclear spins that are relatively close (<5 Å) in space [42].

Figure 7 shows a ROESY spectrum partial expansion of the OFL-CMβCD inclusion complex. Three cross-peaks can be found between the H2, H13 protons of OFL and H3 of CMβCD as well as between H5 of OFL and H5 of CD. These results indicate that OFL enters the CMβCD cavity from the wider rim and the methyl-piperazine ring remains out of the CD cavity. The analogous γ-CD results were the same, the tricyclic moiety is in the cavity. In the case of α-CD the oxazine protons show correlations with the α-CD verifying that only this part of the molecule enters the CD cavity. The molecular modeling study corroborates these

Fig. 6 Aromatic region of OFL ^{1}H NMR spectrum at pH* 3.5 (*bottom*) and the same region in the presence of 10 mM CMβCD (*top*)

Fig. 7 ROESY spectra of the inclusion complex of OFL with CMβCD using 500 ms mixing time
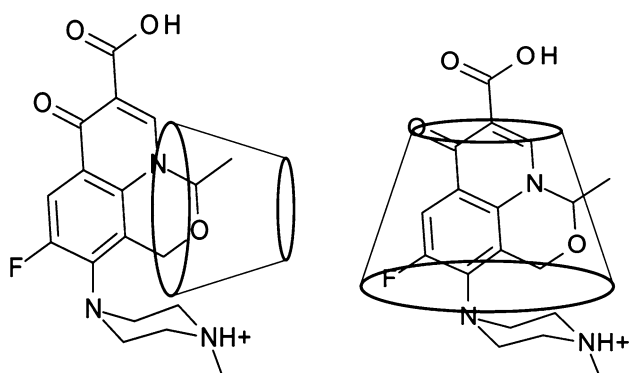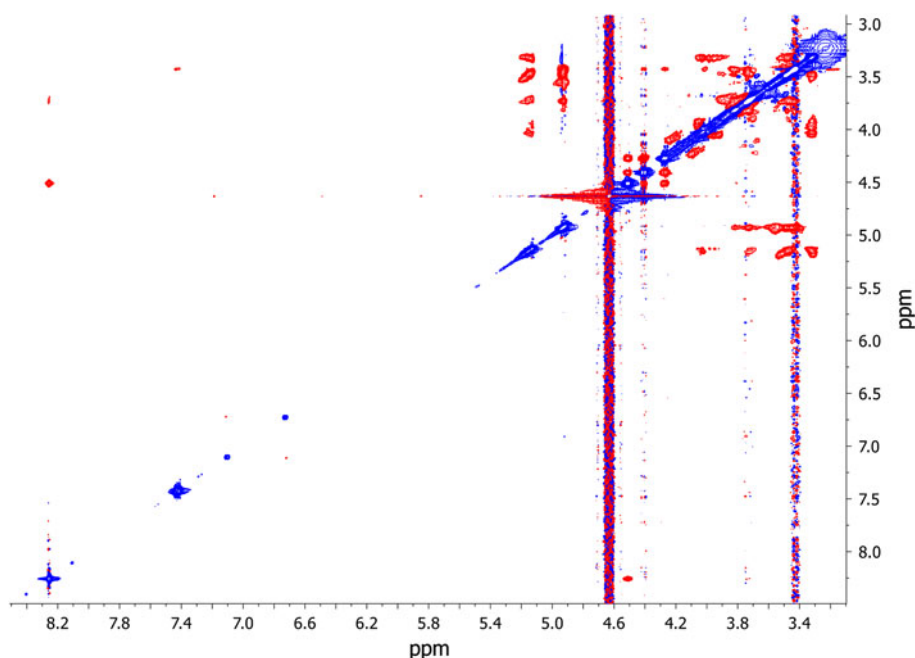

Fig. 8 Molecular structure of OFL—α-CD complex (*left*) and OFL—β-CD complex (*right*)

2D NMR results. Figure 8 shows the simplified molecular model of the inclusion of OFL with different CDs.

basis for improved drug formulation aiming especially at targeted delivery.

Conclusion

The interactions of OFL with various CDs were characterized by CE, circular dichroism, molecular modeling, and various 1D and 2D NMR techniques. Our results showed that the negatively charged CDs formed the most stable complexes at acidic pH. The best enantioseparation was achieved using CMβCD. ^{1}H NMR chemical shift and 2D ROESY data show that β-CDs and γ-CD accommodate the OFL tricycle in the CD cavity, whereas α-CD lets only in the oxazine ring. Molecular modeling studies provided evidence that the substitution pattern plays an important role in the complex stability. These results offer molecular

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