

Journal of Evidence-Based Complementary & Alternative Medicine

<http://chp.sagepub.com/>

Growing Evidence for Human Health Benefits of Boron

Forrest H. Nielsen and Susan L. Meacham

Journal of Evidence-Based Complementary & Alternative Medicine published online 9 May 2011

DOI: 10.1177/2156587211407638

The online version of this article can be found at:

<http://chp.sagepub.com/content/early/2011/05/07/2156587211407638>

Published by:



<http://www.sagepublications.com>

Additional services and information for *Journal of Evidence-Based Complementary & Alternative Medicine* can be found at:

Email Alerts: <http://chp.sagepub.com/cgi/alerts>

Subscriptions: <http://chp.sagepub.com/subscriptions>

Reprints: <http://www.sagepub.com/journalsReprints.nav>

Permissions: <http://www.sagepub.com/journalsPermissions.nav>

Growing Evidence for Human Health Benefits of Boron

Journal of Evidence-Based
Complementary & Alternative Medicine
000(00) 1-12
© The Author(s) 2011
Reprints and permission:
sagepub.com/journalsPermissions.nav
DOI: 10.1177/2156587211407638
http://cam.sagepub.com



Forrest H. Nielsen, PhD¹ and Susan L. Meacham, PhD²

Abstract

Growing evidence from a variety of experimental models shows that boron is a bioactive and beneficial (perhaps essential) element for humans. Reported beneficial actions of boron include arthritis alleviation or risk reduction, bone growth and maintenance, central nervous system function, cancer risk reduction, hormone facilitation, and immune response, inflammation, and oxidative stress modulation. The diverse effects of boron indicate that it influences the formation and/or activity of an entity that is involved in many biochemical processes. Formation of borooesters with the ribose moiety of compounds involved in numerous reactions, such as S-adenosylmethionine and oxidized nicotinamide adenine dinucleotide (NAD⁺) might be the reason for boron bioactivity. Both animal and human data suggest that boron intakes should be >1.0 mg/d. Many people consume less than this amount. Thus, a low boron intake should be considered a health concern, which can be prevented by diets rich in fruits, vegetables, nuts, and pulses.

Keywords

boron nutrition, action mechanisms, intakes, S-adenosylmethionine, NAD, borooesters, ribose

Boron is widely distributed in nature and always found bound to oxygen. The ancient Egyptians have been credited with using boron compounds for mummification and in medicinal applications. However, the first conclusive evidence for the use of borax (Na₂B₄O₇·10H₂O) for medicinal purposes dates from the 8th century in Mecca and Medina.¹ The presence of boron in plants has been known since 1857.² In the 1870s, it was discovered that sodium borate and boric acid could be used to preserve foods.³ For about the next 50 years, borate addition was considered one of the best methods for preserving and extending the palatability of foods such as meat and dairy products. Boron had a vital role as a preservative in preventing food crises during both World War I and World War II. However, as early as 1902, German and American scientists began to question whether large amounts of borates in foods were innocuous. In 1904, Wiley⁴ reported that boric acid in doses greater than 500 mg/d (77 mg boron per day) for 50 days resulted in disturbances in appetite, digestion, and health in human volunteers and concluded that boric acid at 4000 mg/d (699 mg boron per day) was the limit beyond which a harm to humans would occur. Subsequent to his report, the opinion that boron posed a risk to health grew. By the 1950s, boron as a food preservative was essentially forbidden throughout the world.

In the 1920s, boron was found to be an essential nutrient for plants.^{5,6} Over the next 20 years, several unsuccessful attempts at showing boron essentiality in higher animals were made. This resulted in generations of students in nutrition being taught that boron was a unique element because it was essential

for plants but not for higher animals and humans. In 1981, 2 reports appeared that suggested that boron could have nutritional benefits. Newnham⁷ suggested that boron could alleviate arthritic symptoms, and Hunt and Nielsen⁸ found that boron deprivation exacerbated gross bone abnormalities in chicks fed marginal amounts of vitamin D. Since then, an increasing number of reports have indicated that boron is a beneficial bioactive (if not essential) trace element for humans.

Boron Essentiality

Boron has been shown to be essential for the completion of the life cycle (ie, deficiency causes impaired growth, development, or maturation such that procreation is prevented) for organisms in all phylogenetic kingdoms. Higher animals that require boron to complete their life cycle are frogs^{9,10} and zebrafish.^{11,12} Boron-deprived male frogs exhibited atrophied testes, decreased sperm counts, and sperm dysmorphology. Female frogs exhibited atrophied ovaries and impaired oocyte

¹ USDA, ARS, Grand Forks Human Nutrition Research Center, Grand Forks, ND, USA

² University of Nevada Las Vegas, Las Vegas, NV, USA

Corresponding Author:

Forrest H. Nielsen, PhD, USDA, ARS, Grand Forks Human Nutrition Research Center, 2420 2 Ave N, Stop 9034, Grand Forks, ND, 58202-9034, USA
Email: forrest.nielsen@ars.usda.gov

maturation. Boron deprivation resulted in necrotic eggs and high mortality in embryos from frogs fed a boron-deficient diet. Boron deficiency also induced high mortality in zebrafish embryos. Although there are data suggesting that boron deprivation impairs early embryonic development in mice,¹³ the critical experiment demonstrating that boron is essential for a mammal to complete the life cycle is lacking. However, boron has been shown to be a bioactive mineral element that has numerous beneficial actions in nutritional amounts for humans and higher animals.

Beneficial Actions

Arthritis

Since 1981,⁷ occasional reports have appeared suggesting that boron can ameliorate or prevent arthritic symptoms. Based on limited observations in several countries, Newnham¹⁴ reported that the occurrence of arthritis is negatively correlated with the amount of boron in the soil and in the food and water supply. In areas where daily boron intakes were typically ≤ 1.0 mg, the estimated incidence of arthritis ranged from 20% to 70%. In areas where daily boron intakes ranged from 3 to >10 mg, the estimated incidence of arthritis ranged from 0% to 10%. Newnham also stated that arthritic dogs, horses, and cattle given 3 mg of boron for every 25 kg of body weight generally showed improvement in 2 to 4 weeks. This observation apparently has not been confirmed by controlled experiments. Although the evidence in these early reports by Newnham¹⁴ is weak, his suggestion that boron can help alleviate arthritic symptoms could have some merit. In a double-blind study conducted in Australia, 20 patients with confirmed osteoarthritis were given a placebo or a supplement providing 6 mg of boron daily for 8 weeks; 15 patients completed the study.¹⁵ Of the 7 patients consuming the boron supplement, 5 reported improved subjective measures for their arthritic condition (eg, less pain on movement), whereas only 1 of 8 patients consuming the placebo reported an improvement in their arthritic condition. Shortly thereafter, it was reported that boron concentrations in bone and synovial fluid were lower in rheumatoid arthritis patients than in healthy controls.¹⁶ A recent study of 20 patients with mild, moderate, or severe osteoarthritis also found that boron supplementation alleviated subjective measures of arthritis.¹⁷ Patients with mild to moderate arthritis supplemented daily with 6 mg of boron as calcium fructoborate (a naturally occurring boron complex commonly found in fruits and vegetables) reported markedly reduced pain. By week 8, 80% of the test participants reduced or eliminated their use of painkillers. Joint rigidity essentially disappeared, and mobility was markedly increased at 8 weeks. Patients with severe arthritis, who were supplemented daily with 12 mg of boron as calcium fructoborate, exhibited a more subdued improvement in mobility and rigidity but still reported a significant reduction in the use of painkillers. These findings, however, are weakened by the non-blinding to treatment and lack of placebo controls. Interestingly, Keshan-Beck disease (characterized by degeneration of

the articular cartilage between joints) has been associated with low boron concentrations in hair¹⁸ and with deficient boron in soils and crops in China.¹⁹

Arthritic conditions are characterized by chronic inflammatory stress. Animal and cell culture studies have shown that boron can inhibit inflammatory stress such as that found in arthritic conditions. The incidence of severe paw swelling in rats following an intradermal injection of *Mycobacterium butyricum* to induce arthritis was less in rats fed 2.1 mg/kg of boron in the diet compared with rats fed 0.1 mg/kg in the diet.²⁰ In another study, a 20-mg/kg boron diet compared with a 0.2-mg/kg boron diet significantly decreased the incidence of arthritis 12 days postinjection with *M tuberculosis* to induce arthritis in rats.²¹ Supplementing 5 mg/kg boron to a diet containing about 2 mg/kg boron decreased the localized swelling response induced by an intradermal injection of phytohemagglutinin in pigs.²² The boron-supplemented pigs also had increased serum concentrations of the inflammatory cytokine, tumor necrosis factor- α .

In vitro studies also indicate that boron can affect the production of inflammatory cytokines by cartilage cells and cells involved in the inflammatory response. Boron as boric acid was found to stimulate the synthesis and release of tumor necrosis factor- α by chick embryo cartilage²³ and fibroblasts.²⁴ Recently, it was found that calcium fructoborate increased tumor necrosis factor- α protein in the culture media of RAW 264.7 macrophages stimulated by lipopolysaccharide.²⁵ The production and release of interleukin-1 β and interleukin-6 were decreased by the stimulated macrophages cultured with calcium fructoborate. In contrast to the tumor necrosis factor- α findings above, Cao et al²⁶ reported that boric acid inhibited the lipopolysaccharide-induced tumor necrosis factor- α formation in cultured THP-1 monocytes.

A carefully controlled study of the effect of boron on objective indicators of arthritic symptoms with a sufficient number of human participants has not been reported. However, animal and cell culture studies suggest that a low boron status could increase the risk and severity of chronic inflammatory stress associated with arthritis. In addition, 2 experiments using subjective measures in a small number of participants and some limited epidemiological findings suggest that a boron intake higher than 1.0 mg/d could decrease the risk and severity of arthritic symptoms in humans. These provocative findings indicate that more studies should be done to determine whether boron supplementation has therapeutic value for some individuals at risk for or who have arthritis.

Bone Growth and Maintenance

Considerable evidence exists to support the contention that boron has a beneficial (if not an essential) function, influencing especially trabecular and alveolar bone growth and maintenance. Early findings indicating that boron deprivation was detrimental to bone growth independent of another stressor affecting bone health included decreased maturation of the bone growth plate in chicks²⁷ and induced limb teratogenesis

in frogs.¹⁰ More recently, microcomputed tomography of the fourth lumbar vertebra found that boron deprivation (0.1 vs 3 mg/kg diet) decreased bone volume fraction and trabecular thickness and increased trabecular separation and structural model index (a lower value or more plate-like structure is preferable) in rats.²⁸ Boron deprivation (0.07 mg/kg diet vs 3 mg/kg diet) in rats has also been shown to decrease alveolar bone (primary support structure for teeth) repair, which is initiated immediately after tooth extraction.²⁹ Histological examination revealed that boron deprivation decreased osteoblast surface and increased quiescent bone-forming surface in the alveolus. In addition, boron deprivation without tooth extraction impaired alveolar bone formation. Boron deprivation (0.07 mg/kg diet vs 3 mg/kg diet) for 9 weeks in mice decreased osteoblast surface and increased bone-forming surface in both the lingual and buccal side of the periodontal alveolar bone.³⁰ Boron supplementation also has been found to stimulate dental bone formation in rabbits³¹ and increase mineralized nodule formation by cultured osteoblasts (MC3T3-E1).³² Bioactive glasses have been modified to contain boron; this modification enhanced bone formation by the glasses,^{33,34} which are used for bone tissue engineering and in situ bone tissue regeneration. In addition to adversely affecting bone, boron deprivation reduced enamel thickness (hypoplasia) in maturing dental enamel of rats.³⁵

The changes in bone structure and formation induced by boron deprivation apparently affect bone strength and could increase the risk of osteoporosis. Boron deprivation decreased bone strength variables determined by a 3-point bending test of femurs from pigs^{36,37} and rats.^{28,38} Boron supplementation was found to increase mean trabecular density and thickness, trabecular bone volume, and cortical bone volume of femurs from rats with retinoic-induced osteoporosis.³⁹ Calcium fructoborate incorporated into margarine was found to improve bone density in 66 of 100 patients with osteoporosis.⁴⁰ As a result, it was concluded that calcium fructoborate could be a good adjuvant in the treatment of osteoporosis.

Brain Function

Findings showing that nutritional intakes of boron have beneficial effects on the central nervous system are more limited than those showing similar effects on the bone. However, the studies are among the most supportive of the suggestion that boron is a beneficial bioactive element for humans. Under well-controlled dietary conditions, boron supplementation after deprivation resulted in electroencephalograms indicative of improved behavior activation (eg, less drowsiness) and mental alertness, improved psychomotor skills of motor speed and dexterity, and improved cognitive processes of attention and short-term memory in older men and women.^{41,42} Animal findings support the suggestion that boron is beneficial to brain function. Early studies found that boron deprivation affected brain electrical activity in rats in a manner similar to nonspecific malnutrition and heavy-metal toxicity.⁴³ More recently, it was found that boron deprivation alters rat behavior differently

when dietary fat was supplied as fish oil instead of safflower oil.⁴⁴ Boron-deprived rats were less active than boron-supplemented rats when fed the diet with safflower oil, based on reduced number, distance, and time of horizontal movements, front entries, margin distance, and vertical breaks and jumps in a spontaneous activity evaluation. Feeding fish oil instead of safflower oil attenuated the less-active responses of boron-deprived rats. Boron-deficient zebrafish developed photophobia, which apparently was caused by photoreceptor dystrophy.¹²

Cancer

After an epidemiological study found an inverse association between dietary boron and prostate cancer,⁴⁵ Barranco and Eckhart initiated studies showing that boric acid completely inhibited the growth of the cultured prostate cancer cells, DU-145.⁴⁶⁻⁴⁸ Subsequently, Carper et al⁴⁹ found that variable, often dose-dependent, amounts of boron gave responses indicating controlled apoptosis as opposed to a toxic or cytotoxic effect on DU-145, PC-3, and LNCaP-cultured prostate cells. Their initial studies found that 1 mmol/L boric acid markedly inhibited growth of DU-145 cells, moderately inhibited growth of cultured LNCaP cells, and had a muted effect on growth of cultured PC-3 cells. Boron analogs such as phenylboronic acid and hydroxymethylphenylboronic acid also had similar inhibitory effects on growth that were consistent with those on growth of cancer cells in vitro reported by others using boric acid.^{46,50} Boric acid at 1 mmol/L also was found to inhibit growth of breast cancer cells in vitro.⁵¹ Cultured SK-BR-3 and ZR-75-1 breast cells were only partially inhibited—an effect that was less than that with cultured DU-145 prostate cells. However, an apoptotic response occurred in cultured ZR-75-1 cells after 7 days of exposure to either boric acid or phenylboronic acid. Caspase 3 activities confirmed apoptotic, or programmed death, rather than a cytotoxic or necrotic death induced by the boron compounds. In contrast to MgCl₂, which stimulated cell attachment, boric acid and phenylboronic acid inhibited cell attachment.⁵¹ Phenylboronic acid induced a dose-dependent block in the S-phase of the cell cycle in the detached ZR-75-A cells.

Changes in focal adhesion kinase were targeted as a possible mechanism of action through which boric acid induced apoptosis in both cultured breast and cultured prostate cancer cells. Focal adhesion kinase is overexpressed in several human cancer cell lines and is essential in the integrin-mediated signal transduction pathway; it participates in cell migration, angiogenesis, and inflammation/wound healing. Through phosphorylation reactions, focal adhesion kinase conformation changes elicit both intracellular and extracellular responses that suppress apoptosis and promote cell migration. A 4-fold reduction in phosphorylated focal adhesion kinase with concurrent increased caspase-3 occurred with boric acid treatment, which indicated apoptotic activity in the cancer cells.⁵²

It should be noted that boron has been associated with other forms of cancer. A study of cervical smears from 472 women

with a high mean boron intake (8.41 mg/d) and 587 with marginal mean boron intake (1.26 mg/d) found 15 cases of cytopathological indications of cervical cancer in boron-low women and none in the boron-high women.⁵³ In a study of 763 women with lung cancer and 838 matched healthy controls, boron intake was inversely associated with the incidence of cancer; odds increased substantially if the women were not on hormone replacement therapy.⁵⁴

Hormone Facilitator

Numerous studies indicate that boron intake affects the presence or function of hormones, including vitamin D, estrogen, thyroid hormone, insulin, and progesterone. The seminal finding indicating that boron is a bioactive beneficial element in nutritional amounts was that boron deprivation exacerbated gross bone abnormalities in chicks fed marginal amounts of vitamin D.⁸ Hunt⁵⁵ found that boron deprivation exacerbated the distortion of marrow sprouts (location of calcified scaffold erosion and new bone formation), increased the number of osteoclasts within the marrow sprouts of the proximal tibial epiphyseal plate, and delayed initiation of cartilage calcification induced by marginal vitamin D in the chick. Subsequently, it was found that boron deprivation exacerbated marginal vitamin D deficiency-induced decreased calcium and phosphorus absorption and balance in rats,⁵⁶ increased plasma glucose and triglycerides in chicks,²⁷ and decreased growth and femur calcium concentrations in chicks.⁵⁷ Boron supplementation also has been found to increase plasma 1, 25-OH₂-vitamin D concentrations in rats.⁵⁸ In older men and women, boron supplementation (3 mg/d) after 63 days of boron deprivation (0.25 mg/d) increased serum 25-OH-vitamin D concentrations.⁵⁹ These findings suggest that boron could be beneficial to people with marginal vitamin D status, especially those living in areas where winter months provide minimal amounts of ultraviolet light for the synthesis of vitamin D in skin.

Boron has been shown to increase the efficacy of estrogen supplementation in both rats and humans. In ovariectomized rats fed an AIN-76 diet (high in sugar and oils that cause oxidative stress) containing 0.4 mg/kg boron, the addition of boron to the diet (5 mg/kg) significantly increased the beneficial effect of 17 β -estradiol supplementation on trabecular bone volume fraction, bone growth plate density, and trabecular separation.⁶⁰ The combination of boron and 17 β -estradiol versus either of these alone markedly improved the absorption of calcium, phosphorus, and magnesium and the retention of calcium and magnesium.⁶¹ In postmenopausal women, the increases in serum 17 β -estradiol and plasma copper induced by estrogen therapy were significantly higher when the women consumed 3.25 mg/d boron instead of 0.25 mg/d.⁶² The higher boron intake enhanced the effect of estrogen therapy on serum triglyceride and immunoreactive ceruloplasmin concentrations. Boron intake also had a positive influence on the association between reduced lung cancer risk and hormone replacement therapy.⁵⁴ Women who consumed a diet low in boron and did

not use hormone replacement therapy had substantially increased odds for lung cancer risk.

Boron also apparently influences thyroid hormone metabolism. Boron deprivation decreased the rate of tail resorption in larvae during their development into frogs. Addition of 100 fg thyroxine/L of medium, a known enhancer of tail absorption, reversed the delayed tail absorption.¹⁰ In pigs, supplementing a low-boron diet (1-2 mg/kg) with 5 mg/kg boron during the nursery and growth stages decreased serum triiodothyronine and thyroxine.⁶³ Boron supplementation (2.5 mg/d) for 90 days decreased serum triiodothyronine in perimenopausal women after consuming a placebo for 90 days.⁶⁴

Limited evidence suggests that boron can facilitate insulin action. Boron supplementation (2 mg/kg diet) of rats fed a boron-deficient diet (0.2 mg/kg diet) reduced plasma insulin but did not change plasma glucose concentrations.⁶⁵ Peak insulin release from the isolated, perfused pancreas of boron-deprived chicks was almost 75% higher than that from the pancreas of boron-supplemented chicks; the difference was especially noticeable when the perfusate was supplemented with glucose. An effect on insulin utilization could be the basis for the observation that boron deprivation induced a modest but significantly increased fasting serum glucose concentration in older men and women fed a low-magnesium, marginal copper diet.⁶⁶

Boron was found to facilitate progesterone action in frog development. Incomplete frog oocyte maturation caused by boron deficiency could not be induced by the administration of exogenous progesterone.⁶⁷ Progesterone successfully induced germinal vesicle breakdown in oocytes from females fed a boron-supplemented diet.

Immune Response, Inflammation, and Oxidative Stress

In addition to the effects on inflammation described in the arthritis section above, other reports indicate that boron can affect the immune response, the populations of blood cells involved in the inflammatory response, and reactive oxygen species metabolism occurring with chronic inflammatory stress or the acute inflammatory response. Animal studies include the finding that supplementing 3 mg/kg boron to boron-deficient (0.2 mg/kg) diet more than doubled the serum total antibody concentrations in response to human typhoid vaccine injection in rats.⁶⁸ In mice, boron deprivation downregulated 30 of 31 cytokines or chemokines associated with the inflammatory response 6 days postprimary infection with the nematode *Heligmosomoides bakeri*.⁶⁹ An opposite pattern was found, especially 21 days postchallenge; mice consuming low and marginally boron-deficient diets had >100% increases in 23 of the 31 cytokines or chemokines.

Animal studies showing that boron can affect blood cell populations include one in which rats were fed diets where the fat source was fish oil (high in anti-inflammatory n-3 fatty acids) or safflower oil (high in n-6 fatty acids).⁷⁰ Compared with safflower oil, fish oil increased white blood cell numbers,

with most of the increase in the lymphocyte fraction, in boron-supplemented (3 mg/g diet) but not in boron-deprived (0.1 mg/g diet) rats. Fish oil instead of safflower oil increased monocyte and basophil numbers in boron-deprived but not in boron-supplemented rats. In another study, boron supplemented (2.0 mg/kg diet) rats had lower circulating concentrations of natural killer cells and CD8a+/CD4- cells than did boron-deficient (0.1 mg/kg diet) rats after injection with an antigen (*M butyricum* in mineral oil).²⁰

One human study found that perimenopausal women excreting an average of 1.1 and 3.0 mg/d boron during placebo and boron supplementation periods, respectively, had increased white blood cell numbers, an increased percentage of neutrophils, and a decreased percentage of lymphocytes during the boron supplementation period.⁶⁴

Activation of neutrophils and phagocytes during the inflammation process results in the production of reactive oxygen species such as superoxide, hydrogen peroxide, and the hydroxyl radical that are used for microbicidal purposes. Excess reactive oxygen species are destroyed in reactions involving glutathione, superoxide dismutase, and catalase. Evidence exists indicating that boron status can affect the destruction of reactive oxygen species. Boron supplementation (3.0 mg/d) significantly increased erythrocyte superoxide dismutase concentration in boron-deprived (0.25 mg/d) men and women.⁵⁹ Low doses (eg, 5 mg/L) of boron were found to support antioxidant enzyme activities, including superoxide dismutase and catalase, in human blood cultures.⁷¹ Calcium fructoborate has been found to decrease the intracellular production or amount of superoxide ions in cultured cells exposed to oxidative stress.⁴⁰ Findings in an experiment with cultured THP-1 monocytes suggested that boron can limit inflammatory injury (lipopolysaccharide-induced tumor necrosis factor- α formation) even in the presence of glutathione deficiency.²⁶

Related to a possible influence of boron on the response to infection is that 3 of the limited number of known natural biomolecules containing boron are antibiotics. These are boromycin from a strain of *Streptomyces antibioticus*,⁷² tartrolon B produced by the myxobacterium *Sorangium cellulosum*,⁷³ and aplasmomycin produced by *Streptomyces griseus*.⁷⁴

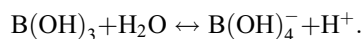
Possible Mechanisms of Actions

Biochemistry of Boron

The diverse responses reported for apparently deficient intakes of boron have made it difficult to identify a primary mechanism responsible for its beneficial bioactivity. The wide range of responses probably is secondary to boron influencing a cell signaling system or the formation and/or activity of an entity that is involved in many biochemical processes. The biochemistry of boron gives some clues about the possible basis for its bioactivity.

Boron biochemistry is essentially that of boric acid. Boric acid acts as a Lewis acid and accepts an electron pair

from a base (H_2O) to form tetravalent compounds such as $B(OH)_4^-$. Thus, the reaction



At the pH of blood (7.4) this reaction results in a dilute aqueous solution composed of $B(OH)_3$ (boric acid) and $B(OH)_4^-$ (borate). Because the pK_a of boric acid is 9.25, the abundance of these 2 species in blood should be 98.4% and 1.6%, respectively. Boric acid forms ester complexes with hydroxyl groups of organic compounds; this preferably occurs when the hydroxyl groups are adjacent and in *cis* orientation. This property results in boron as boric acid forming complexes with several biologically important sugars, including ribose. The fact that borate can stabilize ribose has given support to the speculation that early life on Earth was one in which RNA was the only genetically encoded component of biological catalysts.⁷⁵ Without boron, RNA would have been unlikely to form spontaneously in prebiotic conditions because its ribose component would have decomposed under the harsh conditions of early Earth.

Hypothesized Boron Mechanisms of Action Involving Adenosine

Ribose is a component of adenosine. The diverse actions of boron could occur through its reactions with biomolecules containing adenosine or formed from adenosine precursors, including those shown in Figure 1. S-adenosylmethionine and diadenosine phosphates have higher affinities for boron than any other recognized boron ligands in animal tissues.⁷⁶ Diadenosine phosphates are present in all animal cells and function as signal nucleotides associated with neuronal response. As shown in Figure 2, S-adenosylmethionine is synthesized from adenosine triphosphate and methionine; it is one of the most frequently used enzyme substrates in the body.⁷⁷ About 95% of S-adenosylmethionine is used in methylation reactions, which influence the activity of DNA, RNA, proteins, phospholipids, hormones, and transmitters. The methylation reactions result in the formation of S-adenosylhomocysteine, which can be hydrolyzed into homocysteine.⁷⁸ High circulating homocysteine and depleted S-adenosylmethionine have been implicated in many of the disorders that can be affected by nutritional intakes of boron, including arthritis, osteoporosis, cancer, diabetes, and impaired brain function. Support for the hypothesis that boron bioactivity could be associated with S-adenosylmethionine are the findings that plasma homocysteine increased and liver S-adenosylmethionine and S-adenosylhomocysteine decreased in boron-deprived (0.05 to 0.15 mg/kg diet) compared with boron-supplemented (3 mg/kg diet) rats.⁷⁹ Additional support includes the finding that the bacterial quorum-sensing signal molecule, auto-inducer-2, is a furanosyl borate ester synthesized from S-adenosylmethionine.⁸⁰ Quorum sensing is the cell-to-cell communication between bacteria accomplished through the exchange of extracellular signaling molecules (autoinducers).

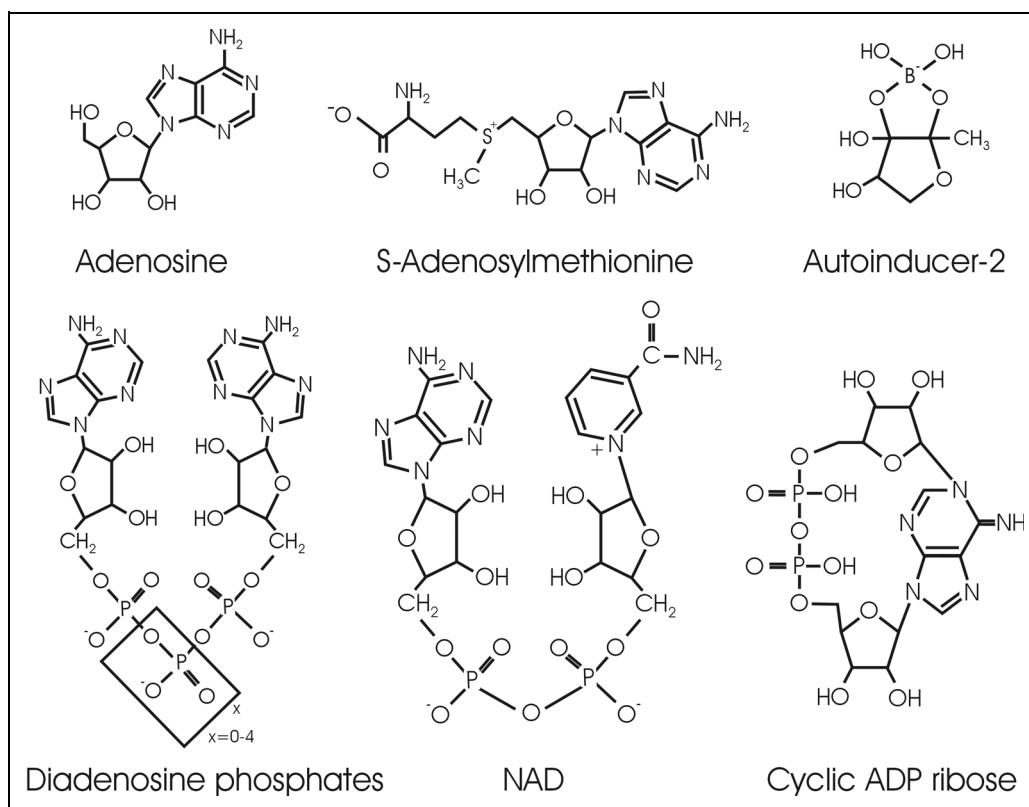


Figure 1. Ribose-containing biomolecules that bind boron

Boron also strongly binds oxidized nicotinamide adenine dinucleotide (NAD⁺) and thus could influence reactions in which it is involved.⁷⁶ One role of extracellular NAD⁺ is binding to the plasma membrane receptor CD38, which is an adenosine diphosphate (ADP)-ribosyl cyclase that converts NAD⁺ to cyclic ADP ribose. Cyclic ADP ribose is released intracellularly and binds to the ryanodine receptor, which results in the release of Ca²⁺ from the endoplasmic reticulum. Cell culture studies show that boric acid binds to and is a reversible inhibitor of cyclic ADP ribose.^{81,82} Boric acid in concentrations that can be found in blood decreased Ca²⁺ release from ryanodine receptor-sensitive stores.⁸² Thus, Eckhart⁸³ has hypothesized that boron could be bioactive through binding NAD⁺ and/or cyclic ADP ribose and inhibiting the release of Ca²⁺. Ca²⁺ is a signal ion for many processes in which boron has been shown to have an effect, including insulin release, bone formation, immune response, and brain function.

Hypothesized Mechanism of Actions Involving Cell Membrane Function, Integrity, and Signaling

Boron can also be bioactive through forming diester borate complexes with phosphoinositides, glycoproteins, and glycolipids, which contain *cis*-hydroxyl groups in membranes. Diester borate polyol complexes could act as calcium chelators and/or redox metabolism modifiers⁸⁴ that affect membrane integrity and function.⁸⁵ A diester borate complex in membranes could

be the molecular species that controls the transmembrane partitioning of boron.⁸⁶ Transporters across the cell membrane have been identified for plants,⁸⁷ yeast,⁸⁸ and animal cells.⁸⁹ The finding that the borate transporter NaBC1, which apparently is essential for boron homeostasis in animal cells, conducts Na⁺ and OH⁻ across cell membranes in the absence of boron,⁸⁹ supports the suggestion that boron affects the transduction of regulatory or signaling ions across cell membranes. It has been hypothesized that a primary essential role for boron in plants, perhaps involving an interplay between boron and calcium, is at the cell membrane level that affects signaling events.⁹⁰ Plant findings also have led to the hypothesis that boron could act as a cellular signal that interacts with transcription factors and thus affects the expression of some genes.⁹¹ These hypotheses suggest that boron could influence cell differentiation, organogenesis, and embryogenesis.^{92,93} Support for this suggestion comes from boron deprivation findings from frog, zebrafish, and bacteria studies. In the boron-deprived *Xenopus* model, abnormal gastrulation is characterized by bleeding yolk and exogastrulation, which suggests disturbed cell membrane structure or function.^{9,10} The most prevalent pathological changes before death of boron-deprived zebrafish during the zygote and cleavage periods were extensive membrane blebbing and extrusion of cytoplasm.^{11,12} The changes occurred when cells were producing prodigious amounts of membranes, and they were consistent with membrane alterations reported for boron-deficient cyanobacteria.⁹⁴

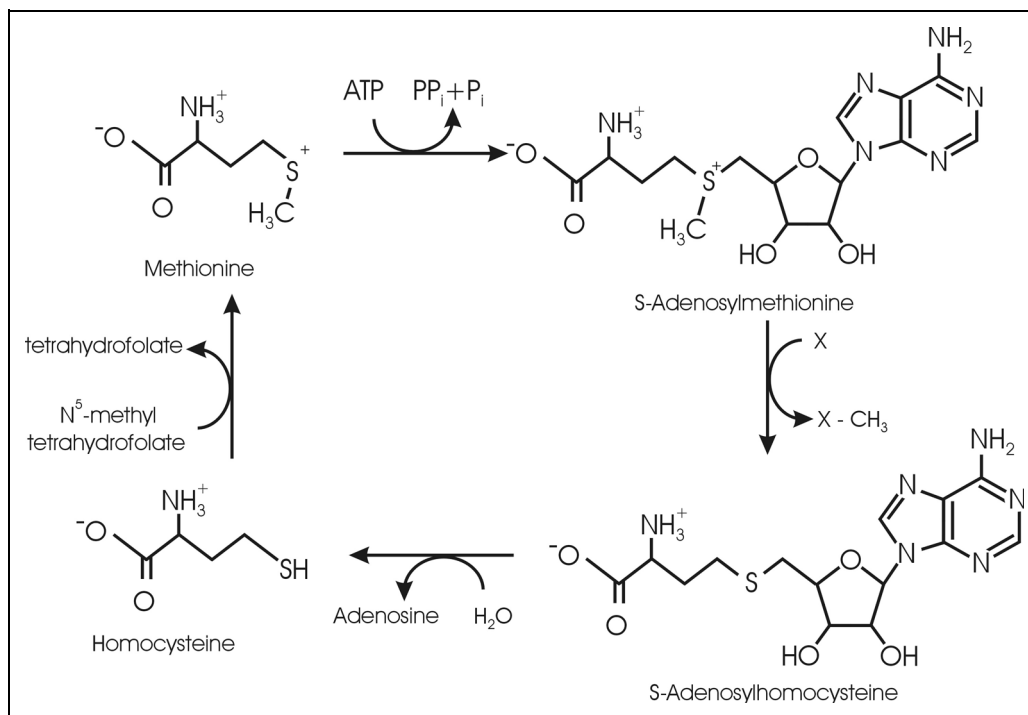


Figure 2. Pathway showing the formation of S-adenosylmethionine, S-adenosylhomocysteine, and homocysteine

Intakes That Affect Health

Nutritional

Both animal and human data were used by the World Health Organization (WHO)⁹⁵ to suggest that an acceptable safe range of population mean intakes of boron for adults could be 1 to 13 mg/d. This suggestion implies that intakes <1.0 mg/d is inadequate for optimal boron beneficial activity. The 1994-1996 Continuing Survey of Food Intakes by Individuals indicated that boron intakes ranged from a low of about 0.35 mg/d to a high of about 3.25 mg/d for adults.⁹⁶ The median intake for various age groups of adults ranged from 0.87 to 1.13 mg/d. These figures suggest that a significant number of people consume less than desirable amounts of boron. Findings from a study involving 43 postmenopausal women in eastern North Dakota⁶⁴ also indicate a similar conclusion. Average urinary excretion of boron (a good indicator of intake) was <0.5 mg/d for 2 women and between 0.5 and 1.0 mg/d for 14 women. In human depletion-repletion experiments, participants responded to a 3 mg/d boron supplement after consuming a diet supplying only 0.2 to 0.4 mg/d boron for 63 days.^{41,42,59,62} These findings indicate that usual intakes of boron above 1.0 mg/d would promote human health.

Safe Upper Level of Intake

Despite the large array of findings showing beneficial effects of boron in animal and human studies, the US Institute of Medicine Food and Nutrition Board⁹⁶ did not set an adequate intake level for boron. However, they set a tolerable upper intake level

of 20 mg/d. The World Health Organization first suggested that 13 mg/d would be a safe upper intake level⁹⁵ but later increased this to 0.4 mg/kg body weight or about 28 mg/d for a 70-kg person.⁹⁷ The European Union established an upper intake level for total boron intake based on body weight that results in about 10 mg/d for adults.⁹⁸ Interestingly, they suggested 1.0 mg/L to be a safe drinking water standard for boron.⁹⁹ This resulted in considerable debate because in some regions in the world, boron concentrations are normally much higher than this. One study found that 10% of 600 drinking water sources had boron concentrations exceeding 1.0 mg/L in the European Union.¹⁰⁰ The World Health Organization first suggested 0.3 mg/L¹⁰¹ then later 0.5 mg/L as a tolerable level for boron.⁹⁷ This increase was appropriate based on the finding that the mortality rate in northern France was significantly lower when drinking water contained >0.3 mg/L than when it contained <0.3 mg/L.¹⁰² Moreover, the low upper intake levels set for drinking water seem inconsistent with the upper intake levels established throughout the world and with the lack of finding of adverse effects in areas of the world where the drinking water is high in boron. For example, Sayli and coworkers¹⁰³⁻¹⁰⁶ found that high concentrations in drinking water, and consequently in food, in Turkey did not negatively affect health. In a population exposed to drinking water containing up to 29 mg/L boron and to boron mining and production, no adverse effects on health or fertility were found over 3 generations. In another study, no adverse effects were found in 66 men (mean age of 39 years) residing in a high boron area for 36 years who had a calculated boron excretion of 6.77 mg/L, which

indicated a high intake of boron. The drinking water in the area where they resided had boron concentrations that ranged from 2.05 to 29.00 mg/L, with a mean of 10.2 ± 4.1 mg/L. This level is 200 to 1000 times the typical values reported for most surface waters (0.01-0.05 mg/L).¹⁰⁷ The findings with drinking water containing high levels of boron and recent findings on the beneficial actions of boron suggest that relaxation of current safe drinking water standards for boron might be appropriate.

Assessment of Boron in the Diet

The preceding discussion indicates that people consuming <1 mg/d will benefit from increased intakes of boron and that it would be best not to exceed intakes above the upper intake level (20 mg/d in the United States and Canada). Assessing human diets to ascertain whether they contain boron in the range of 1 to 20 mg/d requires a thorough analysis of foods, water, and supplements. Several reports have given the boron content of commonly consumed foods and, consequently, an estimated daily dietary intake. The major reports have been by Hunt et al,¹⁰⁸ Anderson and Cunningham,¹⁰⁹ and Hunt and Meacham¹¹⁰ who used different analytical and digestion techniques to determine boron levels. Rainey and Nyquist¹¹¹ used literature boron values for foods to estimate daily dietary boron intakes in several countries. Foods of plant origin, especially fruits, leafy vegetables, nuts, and legumes, are rich in boron, as are wine, cider, and beer. Some of the highest boron concentrations ($\mu\text{g/g}$ fresh weight) are found in avocado (14.3 ± 0.4), peanut butter (5.9 ± 0.2), prune juice (5.6 ± 0.0), chocolate powder (4.3 ± 0.4), wine (3.6 ± 0.0), grape juice (3.4 ± 0.0), and pecans (2.6 ± 0.1).¹¹⁰ These values can vary based on the environment in which the foods are produced.

Improvements in boron assessment methods are needed to more accurately assess boron intakes throughout the world. Because boron analysis is expensive and time-consuming, quality analyses of its presence in food and supplements are limited. In addition, values obtained for foods before 1990 are questionable. At present, urinary excretion is often used as an indication of dietary boron intake. The most widely used method to determine nutrient intake is using values for foods whose intakes have been estimated by food diaries—usually 3-day food records. This determination usually involves software programs that use databases of food composition tables. For boron, this has resulted in some questionable boron intake estimations.

An example of misleading boron intakes comes from a study in which a popular computer software was used to estimate the boron intakes from a single set of diet records. The boron content was available for only 322 foods, or 1.2% of the 26 000 foods in the database. Moreover, boron values reported in the database erroneously used mg/g instead of $\mu\text{g/g}$ of food, which resulted in the limited foods with boron values giving a much higher than expected boron intake and masking the fact that a very limited number of foods in the database had boron values.¹¹² In this study from which diet records were obtained, duplicate plate collections were obtained. This allowed for an

actual boron determination in the diets that was compared with repeat software boron determinations over time as new versions of the software program were updated and released. The versions gave intakes of 4.5, 4.97, and 5.25 mg/d boron in comparison with the analytical determination of 1.2 mg/d. The software values were 3 to 4 times higher than values from other determinations of dietary boron intake.¹¹²⁻¹¹⁵

Dietary supplements are another complicating factor in assessing dietary boron intake. The US National Library of Medicine Dietary Supplements Labels Database reveals an increasing number of supplements with boron as a listed ingredient. In 2009, boron ranged from 0.07 to 3 mg/unit boron in 203 products; only 3 contained 3 mg/unit. One product showed a value of 60 mg/unit, but the issuer confirmed that this was a reporting error; the corrected value was 750 $\mu\text{g}/15$ mL unit for the liquid mineral supplement. In addition to supplements, other unexpected sources of boron can influence its dietary intake; examples include sea salts and chia seeds.

The preceding indicates that although assessment methods for boron are improving, estimates of boron status or dietary intake by using food records must be viewed with caution. Analytical determinations of all items (food, water, and supplements) over a period of time would be the most accurate method for assessing boron intake. Urinary boron excretion could be used to support the accuracy of a boron intake determination.

Summary and Conclusions

The evidence that boron is a bioactive beneficial trace element is substantial. The evidence has come from numerous laboratories that have used a variety of experimental models, including humans. Boron apparently has diverse effects through influencing a cell signaling system or the formation and/or activity of an entity involved in many biochemical processes. Findings have shown that boron is needed to complete the life cycle of some higher animals; in nutritional amounts, it promotes bone health, brain function, and the immune or inflammatory response; alleviates or decreases the risk for arthritis; facilitates the action or utilization of several hormones; and is associated with decreased risk for some cancers. This suggests that boron intakes above 1 mg/d could help people “live longer and better.” Increased intakes of boron through consuming foods such as fruits, vegetables, nuts, and pulses should be recognized as a reasonable dietary recommendation.

Author Contributions

Forrest H. Nielsen was responsible for the overall preparation and final editing of the manuscript. Susan L. Meacham was responsible for writing the sections on cancer, intakes that affect health, and assessment of boron in the diet.

Declaration of Conflicting Interests

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Funding

The author(s) received no financial support for the research, authorship, and/or publication of this article.

References

- Travis NJ, Cocks EJ. *The Tincal Trail: A History of Borax*. London, UK: Harraps; 1984.
- Ploquin J. Le bore dans l'alimentation. *Bull Soc Sci Hyg Aliment*. 1967;55:70-113.
- Gordon V. The case of the toxic life-preserver. *Borax Rev*. 1987; 2:10-12.
- Wiley HW. *Influence of Food Preservation and Artificial Colors on Digestion and Health: I. Boric Acid and Borax*. Washington, DC: Government Printing Office; 1904. US Department of Agriculture Bulletin No 84, Pt 1.
- Warington K. The effect of boric acid and borax on the broad bean and certain other plants. *Ann Bot (Lond)*. 1923;37:629-672.
- Sommer AL, Lipman CB. Evidence of the indispensable nature of zinc and boron for higher green plants. *Plant Physiol*. 1926;1: 231-249.
- Newnham RE. Mineral imbalance and boron deficiency. In: McC Howell J, Gawthorne JM, White CL, eds. *Trace Element Metabolism in Man and Animals (TEMA-4)*. Canberra, Australia: Australian Academy of Science; 1981:400-402.
- Hunt CD, Nielsen FH. Interaction between boron and cholecalciferol in the chick. In: McC Howell J, Gawthorne JM, White CL, eds. *Trace Element Metabolism in Man and Animals (TEMA-4)*. Canberra, Australia: Australian Academy of Science; 1981: 597-600.
- Fort DJ, Stover EL, Strong PL, et al. Chronic feeding of a low boron diet adversely affects reproduction and development in *Xenopus laevis*. *J Nutr*. 1999;129:2055-2060.
- Fort DJ, Rogers RL, McLaughlin DW, et al. Impact of boron deficiency on *Xenopus laevis*: a summary of biological effects and potential biochemical roles. *Biol Trace Elem Res*. 2002;90:117-142.
- Rowe RI, Eckhart CD. Boron is required for zebrafish embryogenesis. *J Exp Biol*. 1999;202:1649-1654.
- Eckhart CE, Rowe RI. Embryonic dysplasia and adult retinal dystrophy in boron-deficient zebrafish. *J Trace Elem Exp Med*. 1999; 12:213-219.
- Lanoué L, Strong PL, Keen CL. Adverse effects of a low boron environment on the preimplantation development of mouse embryos in vitro. *J Trace Elem Exp Med*. 1999;12:235-250.
- Newnham RE. How boron is being used in medical practice. In: Goldbach HE, Rerkasem B, Wimmer MA, Brown PH, Thellier M, Bell RW, eds. *Boron in Plant and Animal Nutrition*. New York, NY: Kluwer Academic/Plenum; 2002:59-62.
- Fracp RLT, Rennie GC, Newnham RE. Boron and arthritis: the results of a double-blind pilot study. *J Nutr Med*. 1990;1:127-132.
- Havercroft JM, Ward NI. Boron and other elements in relation to rheumatoid arthritis. In: Momčilović B, ed. *Trace Elements in Man and Animals 7*. Zagreb, Croatia: IMI; 1991:8.2-8.3.
- Miljkovic D, Scorei RI, Cimpoișu VM, Scorei ID. Calcium fructoborate: plant-based dietary boron for human nutrition. *J Diet Suppl*. 2009;6:211-226.
- Peng X, Lingxia Z, Schrauzer GN, Xiong G. Selenium, boron, and germanium deficiency in the etiology of Keshan-Beck disease. *Biol Trace Elem Res*. 2000;77:193-197.
- Fang W, Wu P, Hu R, Huang Z. Environmental Se-Mo-B deficiency and its possible effects on crops and Keshan-Beck disease (KBD) in the Chousang area, Yao County, Shaanxi Province, China. *Environ Geochem Health*. 2003;25:267-280.
- Hunt CD, Idso JP. Dietary boron as a physiological regulator of the normal inflammatory response: a review and current research progress. *J Trace Elem Exp Med*. 1999;12:221-233.
- Hunt CD. Dietary boron: an overview of the evidence for its role in immune function. *J Trace Elem Exp Med*. 2003;16: 291-306.
- Armstrong TA, Spears JW. Effect of boron supplementation of pig diets on the production of tumor necrosis factor- α and interferon- γ . *J Anim Sci*. 2003;81:2552-2561.
- Benderdour M, Hess K, Dzondo-Gadet M, et al. Effect of boric acid solution on cartilage metabolism. *Biochem Biophys Res Commun*. 1997;234:263-268.
- Benderdour M, Hess K, Dzondo-Gadet M, et al. Boron modulates extracellular matrix and TNF α synthesis in human fibroblasts. *Biochem Biophys Res Commun*. 1998;246:746-751.
- Scorei RI, Ciofrangeanu C, Ion R, et al. In vitro effects of calcium fructoborate upon production of inflammatory mediators by LPS-stimulated RAW 264.7 macrophages. *Biol Trace Elem Res*. 2010;135:334-344.
- Cao J, Jiang L, Zhang X, et al. Boric acid inhibits LPS-induced TNF- α formation through a thiol-dependent mechanism in THP-1 cells. *J Trace Elem Med Biol*. 2008;22:189-195.
- Hunt CD, Herbel JL, Idso JP. Dietary boron modifies the effects of vitamin D3 nutrition on indices of energy substrate utilization and mineral metabolism in the chick. *J Bone Miner Res*. 1994;9: 171-181.
- Nielsen FH, Stoecker BJ. Boron and fish oil have different beneficial effects on strength and trabecular microarchitecture of bone. *J Trace Elem Med Biol*. 2009;23:195-203.
- Gorustovich AA, Steimetz T, Nielsen FH, Guglielmotti MB. Histomorphometric study of alveolar bone healing in rats fed a boron-deficient diet. *Anat Rec (Hoboken)*. 2008;291:441-447.
- Gorustovich AA, Steimetz T, Nielsen FH, Guglielmotti MB. A histomorphometric study of alveolar bone modeling and remodeling in mice fed a boron-deficient diet. *Arch Oral Biol*. 2008;53: 677-682.
- Uysai T, Ustdal A, Sonmez MF, Ozturk F. Stimulation of bone formation by dietary boron in an orthopedically expanded suture in rabbits. *Angle Orthod*. 2009;79:984-990.
- Hakki SS, Bozkurt BS, Hakki E. Boron regulates mineralized tissue-associated proteins in osteoblasts (MC3T3-E1). *J Trace Elem Med Biol*. 2010;24:243-250.
- Gorustovich AA, López JMP, Guglielmotti MB, Cabrini RL. Biological performance of boron-modified bioactive glass particles implanted in rat tibia bone marrow. *Biomed Mater*. 2006;1: 100-105.
- Xie Z, Liu X, Jia W, et al. Treatment of osteomyelitis and repair of bone defect by degradable bioactive borate glass releasing vancomycin. *J Control Release*. 2009;139:118-126.

35. Durand LAH, Mesones RV, Nielsen FH, Gorustovich AA. Histomorphometric and microchemical characterization of maturing dental enamel in rats fed a boron-deficient diet. *Biol Trace Elem Res.* 2010;135:242-252.
36. Armstrong TA, Spears JW, Crenshaw TD, Nielsen FH. Boron supplementation of a semipurified diet for weanling pigs improves feed efficiency and bone strength characteristics and alter plasma lipid metabolites. *J Nutr.* 2000;139:2575-2581.
37. Armstrong TA, Flowers WL, Spears JW, Nielsen FH. Long-term effects of boron supplementation on reproductive characteristics and bone mechanical properties in gilts. *J Anim Sci.* 2002;80:154-161.
38. Naghii MR, Torkaman G, Mofid M. Effects of boron and calcium supplementation on mechanical properties of bone in rats. *Biofactories.* 2006;28:195-201.
39. Xu P, Hu WB, Guo X, et al. Therapeutic effect of dietary boron supplement on retinoic acid-induced osteoporosis in rats [in Chinese]. *Nan Fang Yi Ke Da Xue Xue Bao* 2006;26:1785-1788.
40. Scorei R, Rotaru P. Calcium fructoborate: potential anti-inflammatory agent [published online ahead of print January 28, 2011]. *Biol Trace Elem Res.* doi:10.1007/s12011-011-8972-6.
41. Penland JG. Quantitative analysis of EEG effects following experimental marginal magnesium and boron deprivation. *Magnes Res.* 1995;8:341-358.
42. Penland JG. The importance of boron nutrition for brain and psychological function. *Biol Trace Elem Res.* 1998;66:299-317.
43. Penland JG, Eberhardt MJ. Effects of dietary boron and magnesium on brain function of mature male and female Long-Evans rats. *J Trace Elem Exp Med.* 1993;6:53-64.
44. Nielsen FH, Penland JG. Boron deprivation alters rat behavior and brain mineral composition differently when fish oil instead of safflower oil is the diet fat source. *Nutr Neurosci.* 2006;9:105-112.
45. Cui Y, Winton MI, Zhang ZF, et al. Dietary boron intake and prostate cancer risk. *Oncol Rep.* 2004;11:887-892.
46. Barranco WT, Eckhart CD. Boric acid inhibits human prostate cancer cell proliferation. *Cancer Lett.* 2004;216:21-26.
47. Barranco WT, Eckhart CD. Cellular changes in boric acid-treated DU-145 prostate cancer cells. *Br J Cancer.* 2006;94:884-890.
48. Barranco WT, Hudak PF, Eckhart CD. Evaluation of ecological and in vitro effects of boron on prostate cancer risk (United States). *Cancer Causes Control.* 2007;18:71-77.
49. Carper SW, Hall C, Meacham SL. Boric acid and phenyl boric acid induce apoptosis in prostate cancer cell lines. *Cell Biol Toxicol.* 2007;24:S30.
50. Gallardo-Williams MT, Chapin RE, King PE, et al. Boron supplementation inhibits growth and local expression of IGF-1 in human prostate adenocarcinoma (LNCaP) tumors in nude mice. *Toxicol Pathol.* 2004;32:73-78.
51. Elegbede AF. *Boric Acid Inhibits Cell Growth and Induces Apoptosis in Breast Cancer Cells [master's thesis]*. Las Vegas, NV: University of Las Vegas Nevada; 2007.
52. Wallace A, Meacham S, Abel-Santos E, Fiscus R. Boric acid and focal adhesion kinase: a mechanism to induce apoptosis in breast and prostate cancer cells. *West Reg Mtg Am Chem Soc.* 2008: 61883.
53. Korkmaz M, Uzgören E, Bakirdere S, et al. Effects of dietary boron on cervical cytopathology and on micronucleus frequency in exfoliated buccal cells. *Environ Toxicol.* 2007;22:17-25.
54. Mahabir S, Spitz MR, Barrera SL, et al. Dietary boron and hormone replacement therapy as risk factors for lung cancer in women. *Am J Epidemiol.* 2008;167:1070-1080.
55. Hunt CD. Dietary boron modified the effects of magnesium and molybdenum on mineral metabolism in the cholecalciferol-deficient chick. *Biol Trace Elem Res.* 1989;22:201-220.
56. Hegsted M, Keenan MJ, Siver F, Wozniak P. Effect of boron on vitamin D deficient rats. *Biol Trace Elem Res.* 1991;28:243-255.
57. Bai Y, Hunt CD. Dietary boron enhances efficacy of cholecalciferol in broiler chicks. *J Trace Elem Exp Med.* 1996;9:117-132.
58. Naghii MR, Samman S. The effect of boron on plasma testosterone and plasma lipids in rats. *Nutr Res.* 1997;17:523-531.
59. Nielsen FH. Evidence for the nutritional essentiality of boron. *J Trace Elem Exp Med.* 1996;9:215-229.
60. Sheng MH-C, Taper LJ, Veit H, et al. Dietary boron supplementation enhanced the action of estrogen, but not that of parathyroid hormone, to improve trabecular bone quality in ovariectomized rats. *Biol Trace Elem Res.* 2001;82:109-123.
61. Sheng MH-C, Taper LJ, Veit H, et al. Dietary boron supplementation enhances the effects of estrogen on bone mineral balance in ovariectomized rats. *Biol Trace Elem Res.* 2001;81:29-45.
62. Nielsen FH, Gallagher SK, Johnson LK, Nielsen EJ. Boron enhances and mimics some effects of estrogen therapy in postmenopausal women. *J Trace Elem Exp Med.* 1992;5:237-246.
63. Armstrong TA, Spears JW, Lloyd KE. Inflammatory response, growth, and thyroid hormone concentrations are affected by long-term boron supplementation in gilts. *J Anim Sci.* 2001;79:1549-1556.
64. Nielsen FH, Penland JG. Boron supplementation of perimenopausal women affects boron metabolism and indices associated with macromineral metabolism, hormonal status and immune function. *J Trace Elem Exp Med.* 1999;12:251-261.
65. Bakken NA, Hunt CD. Dietary boron decreases peak pancreatic in situ insulin release in chicks and plasma insulin concentrations in rats regardless of vitamin D or magnesium status. *J Nutr.* 2003;133:3577-3583.
66. Nielsen FH. Biochemical and physiologic consequences of boron deprivation in humans. *Environ Health Perspect.* 1994;102(suppl 7):59-63.
67. Fort DJ. Boron deficiency disables *Xenopus laevis* oocyte maturation events. *Biol Trace Elem Res.* 2002;85:157-169.
68. Bai Y, Hunt CD., Newman SM Jr. Dietary boron increases serum antibody (IgG and IgM) concentrations in rats immunized with human typhoid vaccine. *Proc N D Acad Sci.* 1997;51:81.
69. Bourgeois A-C, Scott ME, Sabally K, Koski KG. Low dietary boron reduces parasite (nematode) survival and alters cytokine profiles but the infection modifies liver minerals in mice. *J Nutr.* 2007;137:2080-2086.
70. Nielsen FH, Poellot R. Boron status affects differences in blood immune cell populations in rats fed diets containing fish oil or safflower oil. In: Anke M, Flachowsky G, Kisters K, et al, eds. *Macro and Trace Elements (Mengen- und Spurenelemente) Workshop 22*. Vol 2. Leipzig, Germany: Schubert-Verlag; 2004:959-964.

71. Tükez H, Geyikoğlu F, Tatar A, et al. Effects of some boron compounds on peripheral human blood. *Z Naturforsch C*. 2007;62: 889-896.
72. Hutter R, Keller-Schierlein W, Knusel F, et al. Stoffwechselprodukte von Mikroorganismen: Boromycin. *Helv Chim Acta*. 1967;50:1533-1539.
73. Schummer D, Irschik H, Reichenbach H, Höfle G. Antibiotics from gliding bacteria. LVII. Tartrolons: new boron-containing macrodiolides from *Sorangium cellulosum*. *Liebigs Ann Chem*. 1994:283-289.
74. Sato K, Okazaki T, Maeda K, et al. New antibiotics, aplasmomycins B and C. *J Antibiot*. 1978;31:632-635.
75. Ricardo A, Carrigan MA, Olcott AN, Benner SA. Borate minerals stabilize ribose. *Science*. 2004;303:196.
76. Ralston NVC, Hunt CD. Diadenosine phosphates and S-adenosylmethionine: novel boron binding biomolecules detected by capillary electrophoresis. *Biochim Biophys Acta*. 2001;1527:20-30.
77. Loenen WAM. S-adenosylmethionine: jack of all trades and master of everything? *Biochem Soc Trans*. 2006;34:330-333.
78. Grillo MA, Colombatto S. S-adenosylmethionine and its products. *Amino Acids*. 2008;34:187-193.
79. Nielsen FH. Boron deprivation decreases liver S-adenosylmethionine and spermidine and increases plasma homocysteine and cysteine in rats. *J Trace Elem Med Biol*. 2009;23: 204-213.
80. Chen X, Schauder S, Potier N, et al. Structural identification of a bacterial quorum-sensing signal containing boron. *Nature*. 2002; 415:545-549.
81. Kim D, Que Hee SQ, Norris A, et al. Boric acid inhibits ADP-ribosyl cyclase non-competitively. *J Chromatogr A*. 2006;1115: 246-252.
82. Henderson K, Stells SL Jr, Kobylewski S, Eckhart CD. Receptor activated Ca²⁺ release is inhibited by boric acid in prostate cancer cells. *PLoS One*. 2009;4:e6009.
83. Eckhart CD. Other trace elements. In: Shils ME, Shike M, Ross AC, Caballero B, Cousins RJ, eds. *Modern Nutrition in Health and Disease*. 10th ed. Philadelphia, PA: Lippincott Williams & Wilkins. 2006:338-350.
84. Goldbach HE, Huang L, Wimmer MA. Boron functions in plants and animals: recent advances in boron research and open questions. In: Xu F, Goldbach HE, Brown PH, et al, eds. *Advances in Plant and Animal Boron Nutrition*. Dordrecht, the Netherlands: Springer; 2007:3-25.
85. Wimmer MA, Lochnit G, Bassil E, et al. Membrane-associated, boron-interacting proteins isolated by boronate affinity chromatography. *Plant Cell Physiol*. 2009;50:1292-1304.
86. Ralston NVC, Hunt CD. Transmembrane partitioning of boron and other elements in RAW 264.7 and HL60 cell cultures. *Biol Trace Elem Res*. 2004;98:181-191.
87. Takano J, Miwa K, Yuan L, et al. Endocytosis and degradation of BOR1, a boron transporter of *Arabidopsis thaliana*, regulated by boron availability. *Proc Natl Acad Sci U S A*. 2005;102: 12276-12281.
88. Kaya A, Karakaya H, Fomenko DE, et al. Identification of a novel system for boron transport: Atr1 is a main boron exporter in yeast. *Mol Cell Biol*. 2009;29:3665-3674.
89. Park M, Li Q, Shcheynikov N, et al. NaBC1 is a ubiquitous electrogenic Na⁺-coupled borate transporter essential for cellular boron homeostasis and cell growth and proliferation. *Mol Cell*. 2004;16:331-341.
90. Bolaños L, Lukaszewski K, Bonilla I, Blevins D. Why boron? *Plant Physiol Biochem*. 2004;42:907-912.
91. González-Fontes A, Rexach J, Navarro-Gochicoa MT, et al. Is boron involved solely in structural roles in vascular plants? *Plant Signal Behav*. 2008;3:24-26.
92. Redondo-Nieto M, Reguera M, Bonilla I, Bolaños L. Boron dependent membrane glycoproteins in symbiosome development and nodule organogenesis: a model for a common role of boron in organogenesis. *Plant Signal Behav*. 2008;3:298-300.
93. Reguera M, Espí A, Bolaños L, et al. Endoreduplication before cell differentiation fails in boron-deficient legume nodules: is boron involved in signaling during cell cycle regulation? *New Phytol*. 2009;183:8-12.
94. Garcia-González M, Mateo P, Bonilla I. Boron protection for O₂ diffusion in heterocysts of *Anabella* sp. PCC 7119. *Plant Physiol*. 1988;87:785-789.
95. World Health Organization. Boron. *Trace Elements in Human Nutrition and Health*. Geneva, Switzerland: World Health Organization; 1996:175-179.
96. Institute of Medicine, Food and Nutrition Board. *Dietary Reference Intakes for Vitamin A, Vitamin K, Arsenic, Boron, Chromium, Copper, Iodine, Iron, Manganese, Molybdenum, Nickel, Silicon, Vanadium, and Zinc*. Washington, DC: National Academy Press; 2001:618-619.
97. World Health Organization, International Programme on Chemical Safety. *Environmental Health Criteria 204 Boron*. Geneva, Switzerland: World Health Organization; 1998.
98. European Food Safety Authority. Opinion of the scientific panel on dietetic products, nutrition and allergies on a request from the commission related to the tolerable upper intake level of boron (sodium borate and boric acid). *EFSA J*. 2004;80:1-22.
99. European Food Safety Authority. EFSA opinion of the scientific panel on contaminants in food chain on a request of the commission related to concentration limits for boron and fluoride in natural mineral waters. *EFSA J*. 2005;237:1-8.
100. Weinthal E, Parag Y, Vengosh A, et al. The EU drinking water directive: the boron standard and scientific uncertainty. *Eur Environ*. 2005;15:1-12.
101. World Health Organization. *Guidelines for Drinking Water Quality*. Geneva, Switzerland: World Health Organization; 1993.
102. Yazbeck C, Kloppmann W, Cottier R, et al. Health impact evaluation of boron in drinking water: a geographical risk assessment in Northern France. *Environ Geochem Health*. 2005;27: 419-427.
103. Sayli BS. An assessment of fertility in boron-exposed Turkish subpopulations 2: evidence that boron has no effect on human reproduction. *Biol Trace Elem Res*. 1988;66:409-422.
104. Sayli BS. The sex ratio of offspring of people exposed to boron. *Reprod Toxicol*. 1998;12:673-674.
105. Sayli BS. Assessment of fertility and infertility in boron-exposed Turkish subpopulations 3: evaluation of fertility among sibs and in "borate families." *Biol Trace Elem Res*. 2001;81:255-267.

106. Sayli BS, C  l M, Elhan AH, Gen Y. Assessment of fertility and infertility in boron-exposed Turkish subpopulation 6: relevant data from all centers. *J Ankara Med Sch.* 2003;25:165-173.
107. Howe PD. A review of boron effects in the environment. *Biol Trace Elem Res.* 1998;66:153-166.
108. Hunt CD, Shuler TR, Mullen LM. Concentration of boron and other elements in human foods and personal-care products. *J Am Diet Assoc.* 1991;91:558-568.
109. Anderson DL, Cunningham WC, Lindstrom TR. Concentrations and intakes of H, B, S, K, Na, Cl and NaCl in foods. *J Food Comp Anal.* 1994;7:59-82.
110. Hunt CD, Meacham SL. Aluminum, boron, calcium, copper, iron, magnesium, manganese, molybdenum, phosphorus, potassium, sodium, and zinc: concentrations in common Western foods and estimated daily intakes by infants; toddlers; and male and female adolescents, adults, and seniors in the United States. *J Am Diet Assoc.* 2001;101:1058-1060.
111. Rainey C, Nyquist L. Multicountry estimation of dietary boron intake. *Biol Trace Elem Res.* 1998;66:79-86.
112. Meacham SL. What do we know about boron in relation to human health? In: Konuk A, Kurama H, Ak H, Iphar M, eds. *Proceedings of the 4th International Boron Symposium.* Ankara, Turkey: Gurup Matbaacilik. 2009:535-545.
113. Meacham SL, Hunt CD. Dietary boron intakes of selected populations in the United States. *Biol Trace Elem Res.* 1998;66:65-78.
114. Meacham SL, Taper LJ, Volpe SL. Effects of boron supplementation on bone mineral density, and dietary, blood, and urinary calcium, phosphorus, magnesium, and boron in female athletes. *Environ Health Perspect.* 1994;102(suppl 7):79-82.
115. Meacham SL, Taper LJ, Volpe SL. Effect of boron supplementation on blood and urinary calcium, magnesium, and phosphorus, and urinary boron in athletic and sedentary women. *Am J Clin Nutr.* 1995;61:341-345.