Does high-field MR imaging improve cortical lesion detection in multiple sclerosis?

\[ \text{Abstract} \] Objective Cortical lesions in multiple sclerosis (MS) are notoriously difficult to visualize with standard MR imaging (MRI) techniques. However, the use of higher field-strengths with intrinsically higher signal-to-noise, which can partly be used to increase spatial resolution, may improve cortical lesion detection. Therefore, in this post mortem study, the sensitivity of high-field-strength MRI (4.7 T) for cortical lesions was investigated, and compared to that of standard field-strength (1.5 T). Methods At 1.5 T, dual-echo T2-weighted spin-echo, as well as 3D-FLAIR images of seventeen formalin-fixed coronal MS and four control hemispheres were acquired. At 4.7 T, the same specimens were imaged with a mainly proton-density (PD)-weighted sequence. Proteolipid protein (PLP)-stained tissue sections (10 \mu m) of the same brain slices were matched to the corresponding MR images, and cortical lesions were scored on all three MR sequences (blinded to histology) and in tissue sections (blinded to MRI). Sensitivity of the sequences for four cortical lesion types was calculated. Additionally, an unblinded, retrospective MR scoring was performed. Results Sensitivity for purely intracortical lesions (histopathological lesion types II, III, and IV; n = 128) was below 10% for both 1.5 T and 4.7 T MRI, while mixed gray matter-white matter (type I) lesions (n = 5) were detected in four out of five cases. All lesion counts increased upon retrospective (unblinded) scoring. However, up to 80% of the intracortical lesions still remained undetected. Conclusions MRI sensitivity for post mortem detection of cortical lesions is low, even when a higher field-strength was used. It varies, however, for different subtypes of cortical lesions.

\[ \text{Keywords} \] high-field MRI \cdot sensitivity \cdot cortical lesions \cdot histopathology \cdot type I lesions

Introduction

Multiple sclerosis (MS) is classically regarded as a disorder that predominantly affects the white matter (WM). However, involvement of the gray matter (GM) was already acknowledged in early pathology studies [12, 16] and has been reported with increasing frequency over the recent years. Histopathological detection of intracortical MS lesions has been improved by the use of myelin protein immunohistochemistry [8, 9, 21, 31], revealing a high prevalence of cortical MS lesions [8, 21, 24, 25].

Based on Dawson’s initial observations [16], a classification system for cortical lesions has been proposed [8, 31], which was used in the current study. This system
distinguishes between mixed GM-WM lesions (type I lesions) and purely intracortical lesions (types II-III-IV). Type II lesions are small intracortical lesions, usually surrounding a blood vessel; type III lesions are larger, extending from the pia downwards without reaching the subcortical WM; type IV lesions affect the entire width of the cortex from the pial surface down to the subcortical WM. Studies using standard histochemistry to describe cortical demyelination [12, 24] initially reported that type I lesions were most frequently observed, but more modern immunohistochemical methods revealed that type III lesions actually constitute the most common cortical lesion type [8, 21, 31].

In a clinical setting, (juxta)cortical lesions are of diagnostic importance [2, 28], and WM abnormalities alone cannot explain the full range of symptoms observed in MS [1]. It has been hypothesized that pathology involving the GM may substantially contribute to clinical decline, and indeed juxtacortical lesions detected by magnetic resonance imaging (MRI) have been correlated to physical disability, cognitive impairment, and MS-associated epilepsy and depression [19, 26, 29, 32, 34]. Furthermore, changes in cortical volume [3, 14, 18] have been reported, as well as changes in quantitative MR measures of the “normal-appearing” cortical GM [10, 13, 20, 37], even in early phases of the disease [15]. To what extent those diffuse abnormalities can be explained by the presence of cortical lesions is as yet unclear, and in this perspective, it is essential to study the effects of cortical lesions (and their specific intracerebral distribution) on clinical deterioration, including neurocognitive deficits.

Unfortunately, however, studies that have correlated conventional MRI techniques to histopathology have proved that it is difficult to visualize lesions located within the cortical GM [21, 24]. One of these studies [21] showed that up to 95% of the intracortical lesions as defined by histopathology, go undetected on MRI at 1.5 T. Higher field-strengths with intrinsically higher signal-to-noise ratio (SNR) may be expected to improve intracortical lesion detection.

Therefore, in the present study, high field-strength (4.7 T) MR images of 17 formalin-fixed coronal MS brain slices were compared to images obtained at standard field-strength (1.5 T), and the relative sensitivities of the different MR techniques for cortical lesions (types I–IV) were determined.

---

**Patients and methods**

### Patients and autopsy procedure

Seventeen coronally cut 10-mm thick hemispheric brain slices of ten chronic MS cases (seven females; mean age: 66.8 years) were selected after rapid autopsy (mean post mortem delay: 8 h and 25 min), as well as four slices from three non-neurological controls (one female; mean age: 41.7 years). The slices were placed in a 10% formalin solution for several weeks, to allow for appropriate fixation. For details concerning demographic and neuropathologic data of patient and control

<table>
<thead>
<tr>
<th>Case</th>
<th>No</th>
<th>Sex</th>
<th>Age</th>
<th>PMD</th>
<th>DD</th>
<th>Type</th>
<th>COD</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>2</td>
<td>F</td>
<td>74</td>
<td>06:00</td>
<td>15</td>
<td>SP</td>
<td>cardiac arrest</td>
</tr>
<tr>
<td>2</td>
<td>1</td>
<td>F</td>
<td>48</td>
<td>04:50</td>
<td>20</td>
<td>SP</td>
<td>euthanasia</td>
</tr>
<tr>
<td>3</td>
<td>2</td>
<td>F</td>
<td>72</td>
<td>12:00</td>
<td>14</td>
<td>PP</td>
<td>pneumonia</td>
</tr>
<tr>
<td>4</td>
<td>1</td>
<td>M</td>
<td>77</td>
<td>04:15</td>
<td>27</td>
<td>PP</td>
<td>stroke</td>
</tr>
<tr>
<td>5</td>
<td>1</td>
<td>F</td>
<td>48</td>
<td>05:50</td>
<td>18</td>
<td>SP</td>
<td>cong. heart failure</td>
</tr>
<tr>
<td>6</td>
<td>2</td>
<td>F</td>
<td>65</td>
<td>06:00</td>
<td>25</td>
<td>PR</td>
<td>cardiac failure</td>
</tr>
<tr>
<td>7</td>
<td>2</td>
<td>F</td>
<td>84</td>
<td>08:45</td>
<td>12</td>
<td>SP</td>
<td>euthanasia</td>
</tr>
<tr>
<td>8</td>
<td>2</td>
<td>M</td>
<td>73</td>
<td>06:45</td>
<td>16</td>
<td>SP</td>
<td>septicemic shock</td>
</tr>
<tr>
<td>9</td>
<td>2</td>
<td>M</td>
<td>59</td>
<td>22:15</td>
<td>22</td>
<td>SP</td>
<td>myocardial infarct</td>
</tr>
<tr>
<td>10</td>
<td>2</td>
<td>F</td>
<td>68</td>
<td>07:30</td>
<td>23</td>
<td>SP</td>
<td>pneumonia</td>
</tr>
</tbody>
</table>

Mean (SD) 66.8 (12.0) 08:25 (05:20) 19.2

<table>
<thead>
<tr>
<th>Case</th>
<th>No</th>
<th>Sex</th>
<th>Age</th>
<th>PMD</th>
<th>DD</th>
<th>Type</th>
<th>COD</th>
</tr>
</thead>
<tbody>
<tr>
<td>11</td>
<td>1</td>
<td>F</td>
<td>57</td>
<td>unknown</td>
<td>NA</td>
<td>control</td>
<td>cardiac failure</td>
</tr>
<tr>
<td>12</td>
<td>2</td>
<td>M</td>
<td>22</td>
<td>unknown</td>
<td>NA</td>
<td>control</td>
<td>HK + AC</td>
</tr>
<tr>
<td>13</td>
<td>1</td>
<td>M</td>
<td>46</td>
<td>unknown</td>
<td>NA</td>
<td>control</td>
<td>DC + PPH</td>
</tr>
</tbody>
</table>

Mean (SD) 41.7 (17.9)

No number of hemispheric slices included (1 or 2) per case; PMD post mortem delay (hrs:min); DD disease duration (yrs); type of disease (SP secondary progressive; PP primary progressive; PR progressive relapsing; control non-neurological control); COD cause of death; HK + AC hyperglycemic ketoadocisosis (diabetes mellitus type I) in combination with alcoholic cardiomyopathy; DC + PPH decompensatio cordis due to primary pulmonary hypertension; NA not applicable
cases, see Table 1. Permission for performing autopsies, use of tissue and access to medical records for research purposes was granted by the local institutional ethics review board, and all subjects gave informed consent prior to death.

**Standard-field (1.5 T) and high-field (4.7 T) MR imaging**

The formalin-fixed brain slices were separately put into a purpose-built perspex single-slice holder (100 × 70 × 10 mm) and immersed in a non-magnetic oil (Fomblin; perfluorinated polyether, Solvay Solexis, Weesp, The Netherlands), after having removed the meninges. This way, inconvenient partial volume effects, resulting from suboptimal intrasulcal suppression of remaining small quantities of fixative and water, could be reduced. Furthermore, Fomblin has a low dielectric effect, which eliminates unwanted susceptibility artifacts near the boundaries of tissue samples. For measurements at clinical field-strength (1.5 T; Siemens Vision, Erlangen, Germany), the perspex single-slice holder was placed in a larger multi-slice holder, which fits exactly into the head coil of the MR system, as described before [6]. At 1.5 T, the sequences used were optimized for clinical scanning protocols. Dual-echo T2-weighted conventional spin-echo (T2SE) images (TR/TE/NEX: 2755ms/45 and 90ms/2; field of view (FoV): 80 × 128 mm; matrix size: 160 × 256; slice thickness: 3 mm; acquisition time: 7 min 25 s), as well as 3D fluid-attenuated inversion recovery (3D-FLAIR) images (TR/TE/TI/NEX: 6500 ms/120 ms/2200 ms/1; 8 partitions per slab; partition thickness: 1.25 mm; FoV: 125 × 200 mm; matrix size: 160 × 256; acquisition time: 7 min 54 s) were obtained from each brain slice.

For measurements at high-field (experimental animal scanner, 4.7 T horizontal bore, Varian, Palo Alto, CA, USA), the holder was placed and fixed into a birdcage volume coil (0.85 mm, length 120 mm). At 4.7 T, the slices were imaged with a mainly proton density (PD)-weighted sequence (FSE3D; TR/TE/NEX: 4000 ms/9 ms/2; echo train length: 8; partition thickness: 0.39 mm; 64 partitions per slab; FoV: 100 × 100 mm; matrix size: 256 × 256; acquisition time: 4h 55 min), which mimicked the appearance of the short echo of the T2-weighted spin-echo sequence at 1.5 T, with the combined benefit of better spatial resolution and higher SNR.

**MR images of the controls were used to study normal appearance, as well as possible artifacts (Fig. 1A). Moreover, contrast-to-noise ratios (CNR) were determined for the different MR image types, on the basis of signal intensity (SI) measurements in regions-of-interest (i.e., GM, GM lesions, WM, WM lesions, Fomblin [noise]). The CNR between two tissue types was defined as: [SI1 – SI2]/standard deviation (noise), taking into account a correction for underestimation of the noise in magnitude images [23]. In this definition, the absolute difference of signal intensities is used, so that negative values in the numerator are accounted for. CNRs were calculated for WM and GM contrast (WM-GM CNR; both in MS and control samples), GM and GM lesions (GM-GM CNR; in MS samples only), and WM and WM lesions (WM-WM CNR; MS samples). For calculation of T2SE CNRs, the short-echo images were used.**

**Histology and immunohistochemistry**

After the MR images had been obtained, the 10-mm thick formalin-fixed brain slices were cut in half to reveal the imaged plane, and the 5-mm slices were subsequently paraffin-embedded. Serial 10-μm thick sections were cut, mounted onto Superfrost glass slides (Menzel-gläser), and dried overnight at 37 °C. Sections were deparaffinized in a series of xylene, 100% alcohol (ethanol), 96% alcohol, 70% alcohol and water. Standard Luxol Fast-Blue (LFB)-Periodic Acid Schiff (PAS) stainings were performed to enable visual matching of histological sections to the standard- and high-field MR images.

For immunohistochemistry, endogenous peroxidase activity was blocked by incubating the sections in methanol with 0.3% H2O2. After having allowed the sections to cool sufficiently, they were rinsed. All washes were carried out for 30 min with 0.01 M phosphate-buffered saline (PBS; pH 7.4). Antibodies were diluted in PBS containing 0.1% bovine serum albumin (BSA). To prevent non-specific binding, sections were preincubated with PBS containing 5% BSA for 10 min at room temperature.

Primary antibodies (Proteolipid Protein (PLP), Serotec, Oxford, UK) were diluted 1:3000 in PBS-BSA and incubated for 1 h at room temperature. After washing, immunolabelling with primary antibodies was detected with biotinylated rabbit anti-mouse (1:500) for
30 min at room temperature and avidin-biotin-peroxidase complexes (sABC-HRP 1:200, DAKO, Glostrup, Denmark) for 60 min at room temperature. Peroxidase activity was demonstrated with 0.5 mg/ml 3,3diaminobenzidinetetrahydrochloride (DA; Sigma, St Louis, MO) in PBS containing 0.03 % H2O2 for 5 min, which leads to a brown reaction product. Sections were lightly counterstained with haematoxylin and mounted with Depex (BDH, Poole, UK).

MRI to histopathology matching and analysis

Hemispheric tissue sections were carefully matched to the corresponding MR imaging planes of the 1.5 T and 4.7 T images, using cortical anatomy and WM lesions as landmarks [6,21]. An example of an LFB-PAS stained tissue section that was successfully matched to the corresponding 4.7 T PD image is shown in Fig. 2.

Numbers of cortical lesions were scored by the MR reader, first on the short-echo of the 1.5 T T2SE, then on the 1.5 T 3D-FLAIR, and finally on the 4.7 T PD images (allowing for appropriate time intervals between scorings to avoid recall bias). The following criteria were used to define cortical lesions: 1) lesions should appear hyperintense on MRI, intermediate to the signal intensities of WM lesions and adjacent normal cortex; 2) lesional borders should be irregularly shaped (as in histology), and 3) lesions should not be clearly traceable over several subsequent slices (to avoid aberrant scoring of vascular structures). The MR reader’s scoring was blinded to histopathology. Not all cortical areas studied histologically could be analyzed on the MR images, since field inhomogeneities at the upper and lower boundaries of the (larger) brain slices (especially at 4.7 T) occasionally caused the images to be poorly interpretable in these areas (illustrated in Fig. 1B). Regions with such signal loss in any MR image type were excluded from all analyses. The pathology reader also scored cortical lesion numbers (blinded to MR data) and classified them (mixed GM-WM or type I lesions; intracortical lesions, types II, III or IV [9, 21, 31]), using the PLP-stained tissue sections. Finally, total numbers of type I–IV lesions scored with the different MR sequences were expressed as percentages of the total number of lesions scored histologically (i.e. the sensitivity of the three MR sequences for the different cortical lesion types). After the blinded scoring of the MR images, precise localizations of all lesions were revealed to the MR reader and a second (retrospective) assessment of the images was performed.

Results

Normal appearance of the three MR sequences (1.5 T T2SE, 1.5 T 3D-FLAIR and 4.7 T PD) was investigated and artifacts were marked. In Fig. 1A, cortical Virchow-Robin spaces can be recognized as punctate hyperintensities on a 4.7 T PD image of a control brain slice. On 4.7 MR images, these areas could be distinguished morphologically from focal cortical lesions in the MS brain slices. In total, 27 cortical lesions, of which 26 type III lesions and one type IV lesion had to be excluded from further analysis, due to local field inhomogeneities at 4.7 T and subsequent poor visibility of these lesions on scans.

In the MS brain slices, a total of 133 cortical lesions were detected, only five of which were mixed GM-WM (type I) lesions. The total number of lesions that were located entirely within the cortical GM (intracortical types II, III, and IV lesions) was therefore 128, most of which (n = 101) consisted of type III (subpial) lesions, followed in number by the smaller type II lesions (n = 22), and the cortex-spanning type IV lesions (n = 5). Table 2 shows the numbers of lesions found in the tissue sections, as well as the proportions of those lesions detected by standard- and high-field MRI.

On both standard-field and high-field MR sequences, nearly all of the type I lesions were seen (one lesion was not scored prospectively on 3D-FLAIR; see Table 2). However, the intracortical part of mixed lesions was often most clear on the high-field PD sequence (Fig. 3). Intracortical lesions (types II, III, and IV) could be identified in a minority of cases only (Table 2, Fig. 4). Prospectively, the sensitivity of the sequences for these lesions at both field-strengths remained below 10%.

![Fig. 2 Matching of a Luxol Fast-Blue (LFB) – Periodic Acid Schiff (PAS) stained hemispheric tissue section (right) to the 4.7 T PD image of the same brain slice (left). The visual matching procedure relies on the presence of different anatomical landmarks, and a high accuracy could be achieved (for example, see the anatomical linings of claustrum and pontes grisei of the lentiform nucleus). Areas that were damaged (black arrow) or otherwise difficult to match (white arrows) were excluded from analysis.](image-url)
Table 2  Number of cortical lesions found with standard- (1.5 T) and high-field (4.7 T) MRI, and sensitivities of the different MR sequences for the different cortical lesion types

<table>
<thead>
<tr>
<th>Lesion type</th>
<th>1.5T MRI</th>
<th>4.7T MRI</th>
<th>Retrospective scoring</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>T2SE</td>
<td>3D-FLAIR</td>
<td>PD</td>
</tr>
<tr>
<td>Type I</td>
<td>5 (100)</td>
<td>4 (80)</td>
<td>5 (100)</td>
</tr>
<tr>
<td>Type II</td>
<td>22 (4.5)</td>
<td>2 (9.1)</td>
<td>3 (13.6)</td>
</tr>
<tr>
<td>Type III</td>
<td>101 (7.9)</td>
<td>9 (8.9)</td>
<td>4 (4.0)</td>
</tr>
<tr>
<td>Type IV</td>
<td>5 (2)</td>
<td>1 (20)</td>
<td>0 (0)</td>
</tr>
<tr>
<td>Type II-III-IV</td>
<td>128 (8.6)</td>
<td>12 (9.4)</td>
<td>7 (5.5)</td>
</tr>
</tbody>
</table>

Table 3  Contrast-to-noise ratios (SD)

<table>
<thead>
<tr>
<th>Contrast-to-noise ratio (CNR)</th>
<th>1.5T</th>
<th>4.7T</th>
</tr>
</thead>
<tbody>
<tr>
<td>WM-GM</td>
<td>9.4 (1.8)</td>
<td>6.3 (2.1)</td>
</tr>
<tr>
<td>WM-WML</td>
<td>13.6 (4.4)</td>
<td>21.4 (8.4)</td>
</tr>
<tr>
<td>GM-GML</td>
<td>2.8 (2.0)</td>
<td>1.7 (0.9)</td>
</tr>
</tbody>
</table>

WM-GM white matter to gray matter CNR; WM-WML white matter to white matter lesion CNR; GM-GML gray matter to gray matter lesion CNR

Upon retrospective scoring, lesion counts increased as expected, but still up to about 80% of the intracortical lesions could not be detected, even though the exact localization of the lesions had at that time been revealed to the MR reader. The differences between MR sequences and field-strengths, in terms of sensitivity for the separate intracortical lesion types, were minute (Table 2). Therefore, data were presented purely descriptively.

The 4.7 T PD images showed highest WM-GM and WM-WML CNRs. However, GM-GML CNRs of all three sequences (both field-strengths) were comparably low (Table 3), indicating that the contrast between GM and GM lesions is very low, independent of sequence or field-strength (for an illustration, see Fig. 4 and also the intracortical part of the type I lesion shown in Fig. 3).

Discussion

This study shows that the sensitivity of MRI for cortical lesions is low, even when using higher field-strengths to improve spatial resolution and SNR. However, MR sensitivity depends on lesion type: mixed type I lesions can be visualized relatively well at both field-strengths, while intracortical lesions (histopathological types II, III, and IV) are seen in less than 10% of the cases.

The use of specific myelin immunohistochemistry (like PLP and Myelin Basic Protein [MBP]), as opposed to standard histochemistry (Luxol Fast-Blue [LFB]), has greatly contributed to the detection of (intra)cortical lesions in post mortem MS brain tissue. However, increased detection of GM lesions under the microscope has also made clear that the majority of these lesions are missed by (standard) in vivo MR imaging techniques [21, 24]. This hinders further research into the clinical effects of GM abnormalities, as well as more fundamental studies focusing on pathogenetic mechanisms of GM pathology.

Low sensitivity of standard MRI for cortical lesions was already reported in an earlier study [21], and the standard-field results in the present study, obtained from a different material (different patients and formalin-fixed instead of fresh tissue), are largely in agreement with those previous results. In that same study, it was hypothesized that imaging at higher field-strengths may be beneficial when trying to image (intra)cortical lesions. In the current study, 4.7 T MR imaging was used to substantially improve spatial resolution, in combination with good SNR. However, as already mentioned and contrary to our expectations, detection of (intra)cortical lesions was not increased. Since the observed cortical lesion numbers were too small, no statistical analysis was performed, and the seemingly different numbers in Table 2 should be cautiously interpreted.

Several factors may underlie the low sensitivity of MRI for cortical lesions. For example, purely intracortical lesions were reported to be largely non-inflammatory [8, 11] and neuroaxonal elements seem to be relatively preserved [5, 16], although they may still be affected to some degree [31, 35, 38]. In addition, because of low intrinsic myelin, demyelination in the subpial layers will only have a very limited effect on the T2-relaxation times. As a result of these effects, intracortical lesions may generate only little contrast on MRI. This seems to be confirmed by the low GM-GML CNRs at both 1.5 T and 4.7 T MRI. It should be noted, however, that although CNRs have been calculated, the goal of this
study was not to quantitatively compare CNRs between sequences or field strengths. Instead, we merely aimed to improve cortical lesion detection by using high-field MRI to predominantly increase spatial resolution, using standardly available imaging sequences. Clinical field-strength (1.5 T) was used as a reference, and sequence parameters at 4.7 T were chosen such that both spatial resolution and SNR were enhanced compared to 1.5 T. Obviously, voxel dimensions and acquisition times were different, and contrast can be expected to decrease due to shortening of T2-relaxation times at higher field-strengths. T1- and T2-relaxation times are also subject to change following death and fixation [4, 27, 30], and the clinical sequences used in this study were not specifically optimized for the post mortem situation. These methodological limitations are likely to have contributed to some extent to the observed insensitivity of the MR techniques to cortical lesions. This also holds for partial volume effects with fluid surrounding the brain slices, which may hamper adequate cortical lesion detection [21]. The non-magnetic oil Fomblin, in which the slices were completely immersed, was used in an attempt to eliminate the high signal intensity from the watery fluid in the sulci. In the narrower sulci, however, optimal results could not always be obtained (data not shown); this may create problems, especially when trying to visualize the superficially localized (type III) lesions. The fact that type III lesions are particularly difficult to image is unfortunate, since it is the most abundant intracortical lesion type [9, 21, 31], which may occasionally extend over several gyri, resulting in a pattern of general cortical subpial demyelination [7, 9, 25].

Further difficulties in post mortem MRI-histopathology correlative studies may be expected in matching tissue sections (of 10 µm thickness in this study) to post mortem MR images (with slice thickness in the mm-range in this study). However, with the large hemispheric samples used in the current study, highly accurate matching could be obtained, since many different anatomical landmarks could be used (see Fig. 2 and also [6]). Areas that could still not be matched (due to e.g. small deviations in angulation during the sectioning process) were excluded from analysis.

As opposed to the intracortical lesion types II, III and IV, type I lesions were easier to detect, both at standard- and at high-field. Apart from the fact that the more conspicuous WM part of type I lesions generally draws the observer’s eye to the spot, the GM part of type I lesions is also usually more easy to distinguish on MRI than the purely intracortical lesions. As mentioned before, this may be essentially related to the fact that the lower layers of the cortical GM are more densely myelinated than

**Fig. 3** Example of a mixed GM-WM (type I) lesion in a coronally cut MS brain slice; A T2SE image of part of the top area of an MS brain slice showing a type I lesion (arrow). The GM part of the lesion is difficult to appreciate; B same area (arrow) on 3D-FLAIR: again the WM part is better visible than the GM part; C high-field PD MR image of the same lesion (arrow). The intracortical part of the lesion can now be slightly better distinguished (arrowheads); D PLP stain (2.5x magnification) of the same (matched) area, showing in detail the demyelination in both the cortical GM (arrowheads) and subcortical WM (arrow), as was already visible on the 4.7 T PD.
the upper layers, so that demyelination within these areas may create greater T2 contrast. It has also been shown that type I lesions, including the GM part, may exhibit higher numbers of T-lymphocytes and of activated microglia/macrophages than the purely intracortical lesion types [8, 11]. Whether increased inflammation within type I cortical lesions is responsible for the higher MR conspicuity of these lesions could not be determined in the current study. This remains an important issue for future research.

It has already been shown on several occasions that focal MS lesions in the WM are easy to image with standard MR methods in the post mortem setting [17, 21, 24, 33, 36]. Therefore, WM lesion detection was not specifically addressed in this study. However, both focal and diffuse WM abnormalities could be visualized well with all sequences (for an example of WM hyperintensities as visualized by 4.7T PD, see Fig. 4B), and WM-WML CNRs of all three sequences were high.

In the search for imaging modalities that allow for improved intracortical lesion detection, newly developed techniques such as Double Inversion Recovery (DIR), suppressing both WM and cerebrospinal fluid, may prove to be beneficial. In vivo, a multi-slab 3D-DIR sequence was recently reported to increase the detection of intracortical lesions and improve the distinction between the purely intracortical lesions and mixed GM-WM lesions [22]. Adapting this particular sequence for post mortem purposes may be helpful. Imaging brain slices at even higher field-strengths than those used in the current study should also be considered. For the radiological practice, results thus obtained might be interesting in the long run, when stronger magnets (e.g. 7 T) will have become more widely available.

Acknowledgments The authors would like to thank Mrs. Christa van Urk – van den Berg for excellent technical support and the Netherlands Brain Bank for supply of the brain material. J.G. Geurts (grant numbers: 00–427 and 02–358b) and L. Bö (grant number: 02–358b) were sponsored by the Dutch MS Research Foundation, Voorschoten, The Netherlands.
References

19. Feinstein A, Roy F, Lobaugh N, Fein-
myelination and diffuse white matter injury in multiple sclerosis. Brain 28:2705–2712


