

Platelets in cancer

From basic research to therapeutic implications

E. Mammadova-Bach; P. Mangin; F. Lanza; C. Gachet

UMR_S949, Inserm, Strasbourg, France; Etablissement Français du Sang-Alsace, Strasbourg, France; Université de Strasbourg, France; Fédération de Médecine Translationnelle de Strasbourg, France

Keywords

Platelet physiology, cancer, tumor metastasis, anti-platelet agents

Summary

Platelets are well-known for their major role in primary hemostasis and thrombosis. Cancer patients frequently manifest thrombotic events and present abnormalities in blood coagulation which appear to be linked to altered platelet function and turnover. Moreover, numerous studies indicate an intimate cross-talk between platelets and tumor growth, angiogenesis and metastatic dissemination. Finally, several experimental data and clinical trials suggest possible benefits of anti-platelet drugs on some cancers. Here, we will review the current state of basic biological research regarding the role of platelets in cancer progression. We also critically review the possible clinical applicability of some anti-platelet therapies to limit tumor growth and prevent metastatic dissemination.

Schlüsselwörter

Plättchenphysiologie, Krebs, Tumormetastasen, Thrombozytenaggregationshemmer

Zusammenfassung

Die wichtige Rolle der Plättchen in der primären Hämostase und Thrombose ist gut bekannt. Krebspatienten weisen häufig thrombotische Ereignisse und Unregelmäßigkeiten der Blutgerinnung auf, die mit Veränderungen der Plättchenfunktion und des Thrombozyten-Turnover verbunden zu sein scheinen. Außerdem wiesen zahlreiche Studien auf eine enge Wechselwirkungen zwischen Plättchen, Tumorwachstum, Angiogenese und Metastasierung hin. Schließlich lassen experimentelle Daten und klinische Studien einen möglichen Nutzen von Thrombozytenaggregationshemmern bei einigen Krebsarten vermuten. Wir geben einen Überblick zum aktuellen Stand der biologischen Grundlagenforschung bzgl. der Rolle der Plättchen bei der Krebsprogression. Auch die Eignung einiger Thrombozytenaggregationshemmer zur Eindämmung des Tumorwachstums und zur Prophylaxe der metastatischen Tumorausbreitung haben wir kritisch geprüft.

Correspondence to:

Christian Gachet
UMR_S949 Inserm, Université de Strasbourg,
Etablissement Français du Sang-Alsace (EFS-Alsace)
10 rue Spielmann, B.P. N° 36, 67065 Strasbourg Cedex,
France
E-mail: christian.gachet@efs.sante.fr

Blutplättchen und Krebs

Von der Grundlagenforschung zur
therapeutischen Relevanz
Hämostaseologie 2015; 35: ●●●●
<http://dx.doi.org/10.5482/hamo-14-11-0065>

received: February 5, 2015
accepted in revised form: March 6, 2015
epub ahead of print: August 20, 2015

to establish the fundamentals of cancer biology (1). Accordingly, therapeutic anti-cancer strategies have primarily focused to target tumor cells. Cumulating experimental and preclinical data indicate that carcinogenesis and tumor progression are not cell-autonomous processes, but rather involve complex multiple interactions with tumor microenvironment (2). Tumor microenvironment is the cellular and molecular environment in which tumors grow, including blood vessels, pericytes, fibroblasts, immune cells, bone-marrow derived cells, growth factors, cytokines and extracellular matrix (ECM) molecules. Tumor microenvironment provides a necessary blood supply and a favorable milieu which stimulates their growth and invasion, prevents from immune recognition and promotes survival in circulation, until they extravasate and seed at distant organs. Currently, it is believed that cellular and molecular components of the tumor microenvironment constitute a barrier protecting against certain drugs and therapies and may favor development of resistance against therapeutic approaches (3). These concepts have become widely recognized and increased the relevance for the development of new therapeutic targets to treat human cancers.

The first association between cancer and blood dates back to the Indian surgeon Sushruta, who lived approximately 3000 years ago and who described that tumor entry into the blood stream leads to blood vessel constriction and compression. In 1865, an association between hemostatic abnormalities and cancer was clinically recognized by the French clinician Armand Trousseau. He reported several cases of thrombophlebitis in patients who were later diagnosed with gastric cancer (4). He emphasized the increased formation of platelet-rich thrombi and hypercoagulability

Cancer is the uncontrolled growth of cells in a given tissue to a cell mass which may spread throughout the body to form metastasis, ultimately leading to death. Cancer comprises several pathologies whose symptoms and disease course differ depending

on the cell type initially affected, the multiple cellular and molecular factors involved, the type of tissue affected, genetic predisposition and environmental factors. Characterization of alterations affecting oncogenes and tumor suppressors helped

in these patients. Recent studies indicated that the chance of diagnosing cancer is significantly elevated after pulmonary embolism or primary deep venous thromboembolism (5). More evidence for platelet involvement in the tumor process, relies on the link between an elevated platelet count (thrombocytosis) and malignant tumors which was reported by Reiss et al., in 1872 (6). Since then, thrombocytosis has been shown to be correlated with advanced, often metastatic steps of cancer and appears to be a negative prognostic factor for many human cancers (7).

Platelets are small anucleated cellular fragments, derived from megakaryocytes in the bone marrow. One third of the platelets are sequestered in the spleen while two thirds of them circulate freely in the bloodstream at a count comprised between 150 and 400 thousands per microliter, providing a potent biomarker for diagnostic and clinical studies. The principal duty of platelets is to prevent hemorrhages and post-traumatic bleedings (8). Following vascular injury, platelets are recruited to the exposed subendothelial ECM proteins, leading to their activation. Platelet activation triggers the release of a variety of biological active substances including adhesive glycoproteins, growth factors, cytokines, coagulation factors, from the so-called α -granules, and soluble agonists such as adenosine diphosphate (ADP), adenosine triphosphate (ATP) and serotonin from the dense (δ) granules along with the production and release of thromboxane A₂ (TxA₂). These mediators contribute to the recruitment of circulating platelets by upregulating the affinity of integrin α IIb β 3 for its ligands, soluble fibrinogen (FGN) and von Willebrand factor (VWF), thereby leading to the formation of a hemostatic plug. A functional coagulation system is additionally required for effective hemostasis completion. The coagulation cascade is initiated by tissue factor (TF) which is exposed upon vascular injury and amplified by negatively charged phospholipids exposed at the platelet surface, supporting the assembly of the tenase and prothrombinase complexes. This cascade leads to the generation of thrombin, which mediates the conversion of FGN into an insoluble fibrin network stabilizing the clot.

Beyond hemostasis and thrombosis, platelets are critically involved in many biological processes including inflammation, embryonic development, innate and adaptive immunology and tissue regeneration to cite a few (9–11). Their role in cancer is known for long but the molecular mechanisms underlying the interactions between cancer cells and platelets only begin to be unraveled. On the one hand, tumor cell induced platelet activation and aggregation may trigger thrombosis in patients with cancer. On the other hand, platelets recruited by tumor cells may favor tumor growth, angiogenesis and metastasis. Recent reviews have been published which all exhaustively describe the most up-to-date data concerning the involvement of blood platelets in the progression of cancer and metastasis (12–15). What we would like to do in the present review is to illustrate and to critically analyze the available data on the role played by platelets in various aspects of the tumor process including thrombosis, angiogenesis and metastatic dissemination and to explore potential therapeutic implications either using existing antiplatelet drugs or new drug candidates to target new receptors and pathways in order to improve cancer treatment.

Cancer-thrombosis connection

Cancer patients often suffer from thromboembolic diseases, such as superficial and deep vein thrombosis, pulmonary emboli, as well as arterial thrombosis and embolism. Thromboembolic disease is the second leading cause of death in cancer patients and thromboprophylaxis with low molecular weight heparin significantly reduces the incidence of symptomatic venous thromboembolism (VTE) in ambulatory cancer patients treated with chemotherapy (16). The “Khorana risk assessment model” which includes several clinical variables such as site of cancer, obesity, leukocytosis, anemia and thrombocytosis has been proposed to predict risk of cancer-associated thrombosis. This risk prediction model, which has been validated in several large cohorts of patients in various clinical settings, provided evidence that thrombocy-

tosis is an important biomarker of cancer-associated thrombosis, indicating a role for platelets in this process (7). Cancer-mediated thrombocytosis may be explained by the ability of several tumor cells to produce and regulate thrombopoietin (TPO), (17, 18), a key cytokine which stimulates megakaryocyte formation and platelet production. In a multicenter study involving 619 patients with epithelial ovary cancer, thrombocytosis was found to be associated with high plasma level of TPO and interleukin-6 (IL-6), and linked to an advanced disease and poor survival (17). It has also been reported that circulating IL-6 is a risk marker of thromboembolic manifestations. Interestingly, experiments performed with orthotopic mouse models of ovary cancer, provided evidence and confirmed that tumor cell-derived IL-6 upregulated the production of hepatic TPO (17).

In addition to thrombocytosis, several platelet activation markers were found to be upregulated and could contribute to the prothrombotic state of cancer patients. For example, ovarian cancer patients had elevated number of CD63 positive platelet microparticles reflecting a procoagulant phenotype (19). Moreover, in several studies, key markers of platelet activation, including CD40 ligand, β -thromboglobulin or P-selectin exposed at the platelet surface or soluble in the plasma (13) are increased in cancer patients compared to non-cancer control subjects. Thus, the combination of increased number of circulating platelets along with upregulation of circulating prothrombogenic factors establishes the hypercoagulable state contributing to Trousseau's syndrome.

The concept of cancer cells as inducers of platelet activation and aggregation has been well established in vitro. Tumor cells can trigger changes in platelet activation through several mechanisms. The activation by direct interaction and subsequent aggregation of platelets, termed as TCIPA (tumor cell-induced platelet aggregation) has been shown to occur in vitro with lung, colon, breast, pancreatic and prostate cancer cells (20). One possible mechanism of TCIPA involves sialoglycoprotein Aggrus/podoplanin found in various tumor cell lines, such as glioblastoma, mesothelioma, lung, esophageal squamous cell and

colon carcinoma (21, 22). The C-type lectin-like receptor (CLEC-2) expressed on platelets was identified as one of the counter receptors of podoplanin. Podoplanin binding to CLEC-2 transmits platelet-activation signals through Src family kinases, Syk and phospholipase C γ 2 in platelets (23). Blockade of the podoplanin-CLEC-2 interaction inhibits TCIPA in vitro, as well as metastasis in vivo (24). TCIPA is also induced by tumor derived cathepsin B and matrix metalloproteinases (MMPs) in several tumor cell lines as well as by released ADP (20).

Cancer cells can also trigger indirect platelet activation. They can initiate the coagulation cascade through their ability to express tissue factor, thereby generating thrombin, the most potent platelet agonist. Their ability to release procoagulant micro-particles, can also initiate thrombin generation (25). Finally, platelets can be activated through cancer-induced formation of neutrophil-extracellular DNA traps (NETs), which may result in platelet aggregation and thrombus formation (26) (► Fig. 1).

To what extent all these events occur in patients is difficult to assess and tools lack at the moment to properly interfere with them. We will discuss later the impact of antiplatelet therapy on platelet/cancer cell interaction.

Platelet-mediated tumor angiogenesis

Beyond a certain size (>1–2 mm³), tumors initiate angiogenesis, the formation of new capillaries from preexisting blood vessels, to provide the oxygen and nutrients essential for their growth (27). There is experimental evidence that the role of platelets in angiogenesis may contribute to tumor growth and survival (► Fig. 2a).

Platelets contain both pro- and anti-angiogenic factors, which can be released upon platelet activation. Examples of positive regulators of angiogenesis in platelets are vascular endothelial growth factor (VEGF), platelet-derived growth factor (PDGF), basic fibroblast growth factor (bFGF), insulin-like growth factor 1 (IGF-1), lysophosphatidic acid (LPA), an-

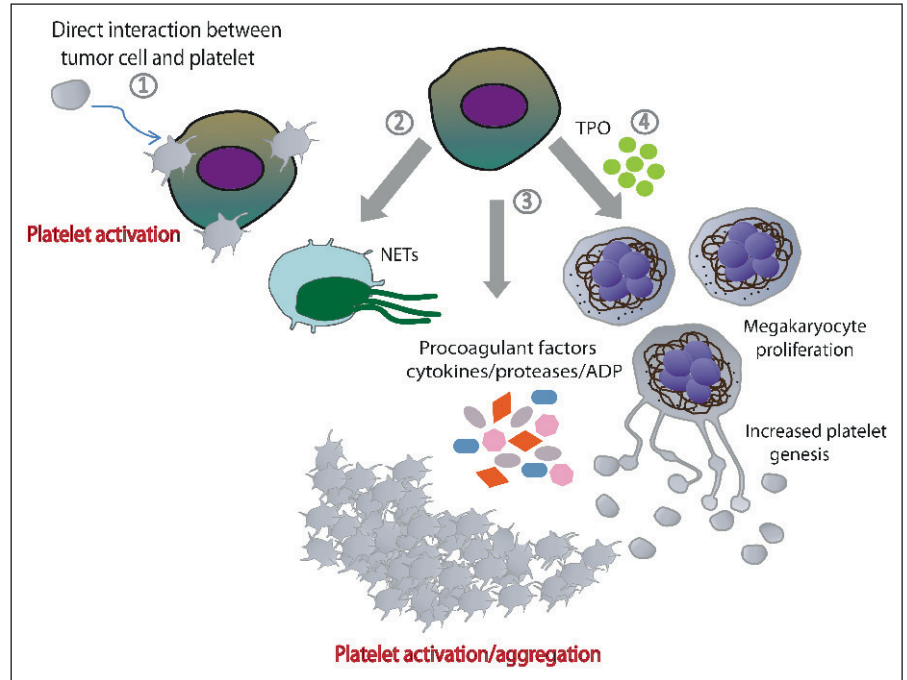
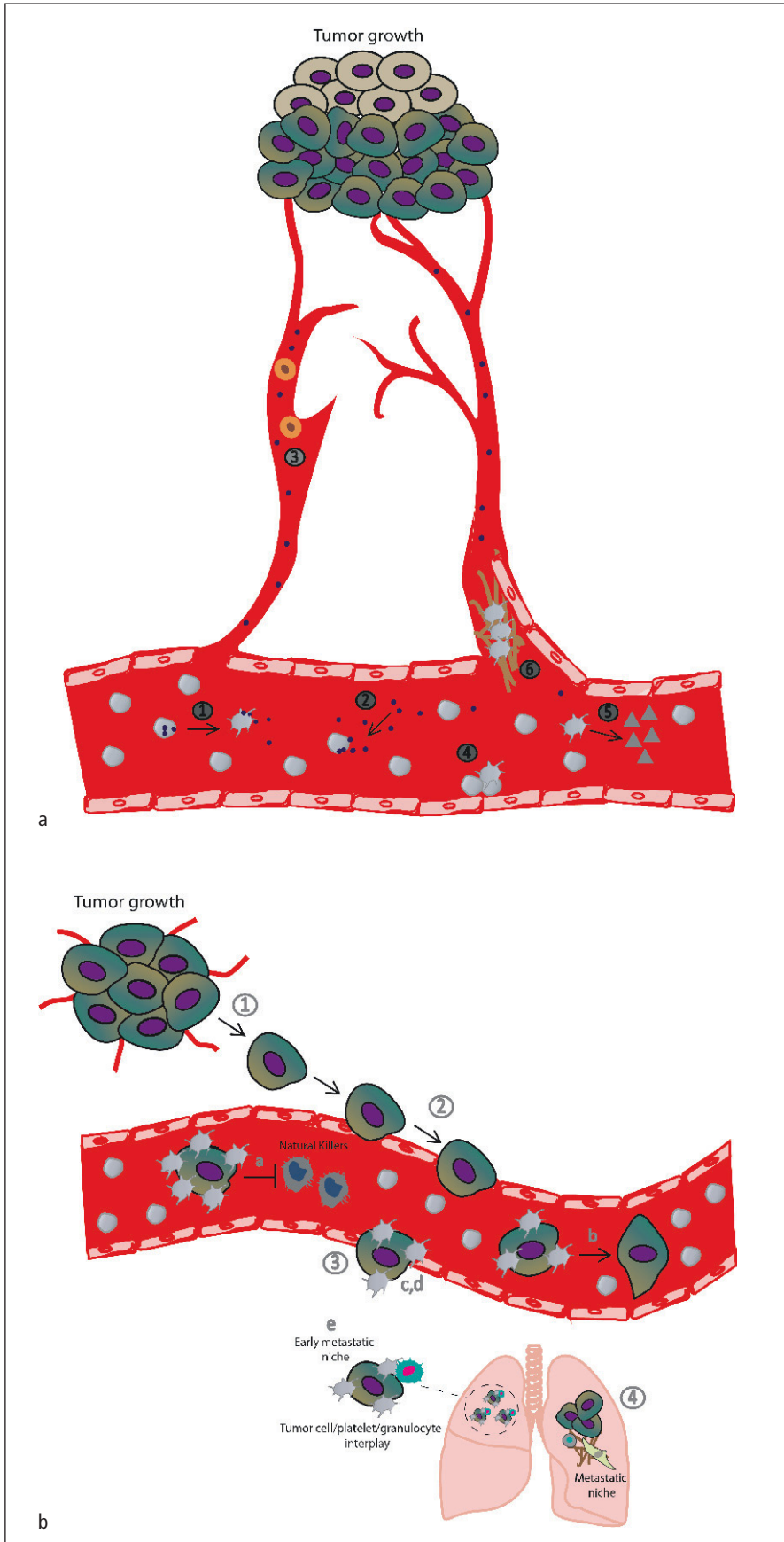


Fig. 1 Mechanisms involved in tumor cell-induced thromboembolic events: Tumor cells through their ability to bind directly platelets (1), to promote NET formation (2), to release pro-activatory and pro-coagulant factors (3) could trigger platelet aggregation and activation. Additionally, tumor cells may regulate TPO levels to stimulate thrombocytosis (4), thereby leading to a prothrombotic phenotype. For the details, see the text.

giopietin (Ang), and MMP-1, -2, and -9, while negative regulators comprise platelet factor-4 (PF-4), plasminogen activator inhibitor type-1 (PAI-1), thrombospondin, tissue inhibitor of MMPs and endostatin. Several studies suggested that release of pro- and anti-angiogenic factors is a tightly regulated process. It has been proposed that pro- and anti-angiogenic factors are stored in distinct α -granules in the same platelet and that their release is regulated by a selective stimulation of the thrombin proteinase-activated receptors (PAR) -1 and PAR-4 (28), while tumor cell derived ADP would promote the release of the pro-angiogenic factor VEGF through the activation of the P2Y₁₂ receptor, without affecting the release of the anti-angiogenic factor endostatin (29, 30). In contrast, TxA₂, another important soluble agonist, has been proposed to promote the release of endostatin but not VEGF (29). Such a tightly regulated mechanism of differential secretion with selective release of pro or anti angiogenic factors is somewhat difficult to reconcile with the common knowledge that platelet activation results from simulta-

neous stimulation of multiple pathways. In addition, the concept of functional co-clustering of proteins in distinct granules was recently challenged in experimental settings using quantitative immunofluorescence microscopy techniques and micro ELISA arrays (31, 32). Thus, it will probably be very challenging to determine under which in vivo conditions the selective stimulation of various platelet receptors, such as PAR-1, PAR-4, P2Y₁₂ and thromboxane receptors may occur.

Platelets have been reported to take up and sequester pro-angiogenic mediators, which could indirectly regulate angiogenesis. It has been shown that a small amount of VEGF secreted from microscopic subcutaneous tumors resulted in elevated levels of platelet VEGF (33). In ischemic hind limb and tumor xenograft hypoxia models, platelets promoted mobilization and recruitment of bone marrow-derived cells to neovascularized hypoxic tissues to favor angiogenesis, and this homing was dependent not only on release of the α -granule content but also on the sequestration of growth factors and cytokines by platelets



(34). In agreement with these results, Kuznetsov et al., showed that human platelets adsorb cytokines released by luminal breast cancer cells in order to deliver them to indolent disseminated tumors ensuring their outgrowth through vessel formation (35).

Beside the release, storage and delivery of soluble mediators, platelets have also been proposed to participate in angiogenesis by directly or indirectly regulating endothelial cell function. Human platelets were able to promote tube formation of human umbilical vein endothelial cells in a matrigel assay (36). This effect was independent of platelet activation or granule release, suggesting that direct platelet/endothelial interactions can enhance angiogenesis in malignancy. Conversely, the coagulation end-product fibrin, which is fre-

Fig. 2 Schematic illustrations

a) multiple ways of platelet-mediated tumor angiogenesis: Platelets may stimulate tumor angiogenesis through the secretion of pro-angiogenic growth factors (1), the take-up and sequestration of pro-angiogenic mediators from tumor and tumor microenvironment (2), and the recruitment of bone-marrow-derived cells to neovascularized tumor sites (3). Direct interactions between platelets and endothelial cells (4) may also stimulate tumor angiogenesis. Finally, angiogenic sprouting may be supported by shedding of microparticles (5) and endothelial cell migration on platelet-enriched fibrin clots (6). For the details, see the text.

b) main steps of metastatic dissemination: Metastatic cascade is a multi-step process which includes: local invasion enabling detachment of tumor cells from primary tumor site (1), tumor cell attachment to the vascular wall and entry into the circulation (intravasation), (2), exit of tumor cells from the bloodstream (extravasation), (3) and finally tumor cell survival and proliferation at secondary organs (4), such as lung to form metastases, through involvement of a local microenvironment (metastatic niche). Within the bloodstream, platelets escort tumor cells through several steps of metastasis and may support this pathological process by: protecting tumor cells from immune system surveillance (a), conferring a mesenchymal phenotype to epithelial tumor cells (b), reinforcing tumor cell/blood vessel interplay (c), promoting tumor cell extravasation (d), and establishing early metastatic niches (e) within the vasculature of distant organs to promote tumor cell survival and proliferation. For the details, see the text.

quently found deposited in tumors, constitutes a provisional matrix that supports endothelial cell adhesion, survival and migration to promote angiogenesis (37).

Platelets have also been shown to stimulate angiogenesis following the shedding of microparticulates. Platelet-derived microparticulates (PMP) promote proliferation, migration and angiogenic sprouting of endothelial cells *in vitro* (38). They increase the mRNA levels of pro-angiogenic factors, such as VEGF, hepatocyte growth factor and MMP-9 in tumor cells, which may subsequently support tumor angiogenesis and growth (39). Brill et al., reported that PMP are capable to induce angiogenic sprouting *in vitro* and *in vivo* to a similar extent as whole platelets (40). An association between PMP and tumor progression has been shown in patients with prostate and gastric cancer (41). In addition, the levels of PMP in patients with gastric cancer were strongly correlated with the levels of angiogenic factors, such as VEGF, IL-6 and RANTES (Regulated on Activation Normal T Cell Expressed), further supporting a possible role of PMP in tumor angiogenesis (42).

Therapies targeting VEGF signaling are often unsuccessful in cancer patients. Approval to use Bevacizumab, a monoclonal antibody against VEGF-A, has been recently revoked by the United States Food and Drug Administration in women with breast cancer (43). Therefore, more appropriate anti-angiogenic strategies aiming to limit cancer progression are needed. Whether targeting the pathways of platelet-mediated angiogenesis could represent a potential anti-cancer strategy is rather speculative but remains to be addressed. Finally, whether angiogenic profiling of platelet α -granules in patients with cancer may constitute a predictive marker of the risk of disease progression and overall prognosis is a question for future stimulating studies.

Mechanisms of platelet-mediated metastasis

Metastasis is the major cause of mortality in patients suffering from cancer, and therapeutic interventions directed against

this pathological process are limited since the underlying mechanisms are not fully understood. To metastasize, tumor cells must undergo successive several steps of cancer progression: detachment from the primary tumor, intravasation into the vascular system directly or via the lymph nodes, survival in the circulation, attachment to the endothelium and extravasation, and finally survival and proliferation in distant organs. Survival and proliferation of disseminated tumor cells at distant sites need a supportive specialized micro-environment. This concept known as seed and soil theory, was formulated by Paget in 1889, suggesting that microenvironment niches (soil) need to be compatible to tumor cells (seed) (44).

The first evidence for a role for platelets in this phenomenon came from studies reported by Gasic et al., in which thrombocytopenia was closely associated with reduced metastasis (45). Moreover, injection of platelets in thrombocytopenic mice restored the capacity to form metastases (46). Interfering with platelet production by disturbing megakaryocyte maturation also resulted in inhibition of metastasis in an experimental mouse model (47). Several mechanisms have been proposed to explain the role played by platelets (► Fig. 2b). They can contribute to metastasis by shielding tumor cells from immune host system, by triggering epithelial-mesenchymal transition, by mediating tumor/vascular wall interaction and by various mechanisms helping extravasation of tumor cells from host vasculature. They are also able to mediate tumor cell survival and growth at distant sites by guiding establishment of metastatic niches.

Platelet-mediated protection from immune system surveillance

Cytotoxic natural killer (NK) lymphocytes, which induce tumor cell lysis, represent the major threat towards tumor cells in the blood circulation. Platelets are likely the first blood cells to interact with tumor cells (48) and have been proposed to serve as physical guards to protect tumor cells from immune system surveillance (49).

Several NK sensitive tumor cell lines exhibited decreased metastatic potential after

platelet depletion. The proposed underlying mechanism relies on platelet adhesion to the tumor surface, thereby providing a shield and protecting them from NK-cell induced cytotoxic effects. Moreover, platelets were shown to transfer MHC (major histocompatibility complex) class I molecules onto tumor cells to provide a self-signal to NK-cells suppressing their killing activities *in vitro* (50). It seems that intact platelet activation is required for efficient metastasis in the presence of NK-cells since mice deficient for G α_q , a G protein crucial for platelet activation, exhibited decreased tumor cell survival and metastasis, an effect that was reversed by immunologic or genetic depletion of NK (51). Others had previously shown that platelets inhibit NK-cell cytotoxic activity through soluble factors (52). This has notably been evidenced by the fact that supernatants of activated platelets decreased NK-cell dependent lysis of human leukemia cells *in vitro* (52). Kopp et al., suggested that platelet-derived TGF- β down-regulates the cytokine NKG2D (Natural Killer Group 2, member D) on NK-cell surface, resulting in decreased NK-cell cytotoxicity *in vitro* (53). Furthermore, neutralization of TGF- β (transforming growth factor- β) in platelet release reversed this effect and restored normal NK-cell function. Interestingly, down-modulation of NKG2D has been associated with elevated TGF- β levels in plasma of patients with colorectal and lung cancer (54). Studies on modulation of NK-cell cytotoxicity by platelet TGF- β shed a new light on future clinical research. Several inhibitors of TGF- β signaling are currently under evaluation in preclinical models and early clinical trials, including soluble protein receptors, TGF- β antibodies, small-molecule kinase inhibitors, oligonucleotides and peptide aptamers. However, recent *in vivo* kinetic studies of lung and liver metastasis casted doubt (55) on the concept of the inhibitory effects of platelets on NK cell anti-metastatic activity (49). The authors reported that platelets exert their pro-metastatic effects within the first 1 hour following intravasation of tumor cells into circulation, whereas anti-metastatic effects of NK occurred between 1 and 6 hours after tumor cell inoculation in mice in a platelet-independent manner. Future studies are

required to determine under which *in vivo* conditions and pathological context platelet TGF- β may modulate NK-cell cytotoxicity.

Role of platelets in epithelial-mesenchymal transition

Epithelial-mesenchymal transition (EMT) represents a major developmental regulatory program, which can be reactivated during the progression of some cancers, conferring mesenchymal cell properties to epithelial cells. Tumor cells undergoing EMT lose their adhesive properties and acquire migratory and proteolytic activities, helping them to support the metastatic process. Within the same tumor, the loss of epithelial and the gain of mesenchymal markers have been described to correlate with tumor progression, metastasis and bad prognosis (56). Platelets have been proposed to promote EMT through several signaling pathways upon release of PDGF and TGF- β (57, 58). Here again, TGF- β released by activated platelets plays a key role, in addition to its effect on NK cells. Indeed, TGF- β activates TGF- β /Smad signaling in tumor cells (58). In addition, a direct interaction between tumor cells and platelets appear sufficient to induce Nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ b) signaling, which in synergy with TGF- β signaling promotes EMT and efficient metastatic seeding (58). Because platelet TGF- β enhances EMT, it may represent an attractive target to interfere with progression of tumor process. Thus, the rationale of targeting TGF- β deserves further studies to unravel the mechanisms and signaling pathways including immune system and anti-tumor effects. An alternative option to inhibit the switch to pro-metastatic EMT phenotype induced by platelet TGF- β may be by targeting NF- κ b activation in tumor cells. However, no drug is currently available to specifically block NF- κ b activation.

Platelets in tumor cell/blood vessel interactions

Platelets were proposed to facilitate tumor cell adhesion to endothelial cells, thereby enhancing their extravasation from the cir-

culatory system into the tissues to establish metastasis. Platelet/tumor cell aggregates travel inside the circulation and roll along activated endothelium. P-selectin and CD44 appear to support this process in colon cancer cells (59) and probably in other cancer cells. In agreement with this, P-selectin knock-out mice exhibited decreased tumor growth and metastasis of melanoma and colon cancer cells (60). Similar results were obtained in mice treated by mutated heparin lacking anticoagulant activity, but with P- and L-selectin inhibiting effects (61).

Platelet integrin α IIb β 3 has been proposed to mediate transition from selectin-dependent rolling of tumor cells along the endothelium to stationary adhesion (62). In experimental models, inhibition of this integrin or genetic deficiency of β 3 integrin in mice decreased tumor cell colonization in lungs and the number of bone metastases (63–65).

Tumor cells also express integrins, such as α V β 3 integrin, which was shown to support tumor cell interaction with platelets allowing adhesion to the vasculature and metastasis (66). This integrin colocalizes with nectin-like molecule 5 (NECL5) at the invasive edge of tumor cells (67). It has been reported that NECL5 on colon cancer cells enhances experimental metastasis. This effect was shown to be mediated by the interaction of NECL5 with its counter-receptor on platelets, probably CD226 enabling tumor cell attachment to the endothelium, ultimately enhancing metastasis (67). Recently, α V β 3 integrin expressed by breast cancer cells has been shown to interact with platelet-derived autotaxin, thereby leading to early bone colonization by these cells and progression of skeletal metastases in mice (68).

Tumor cell/platelet interactions resulting in platelet activation, the release of soluble mediators may activate endothelial cells. Exposure of VWF on activated endothelial cells could support the recruitment of platelet/tumor cell aggregates through its binding to platelet GPIIb β 3. Jain et al, reported that the absence of GPIIb β 3 decreases metastasis of melanoma cells (69), but whether this relies on a loss of tumor cell adhesion to the endothelium remains to be established. In contrast, mice

treated with Fab fragments directed against GPIIb β 3 binding to VWF exhibited enhanced metastasis of the same melanoma cell type (70). Additional studies are required to explain this discrepancy and provide potential mechanisms by which GPIIb β 3 may affect metastasis.

Platelet-mediated tumor cell extravasation

After attachment to endothelial cells, tumor cells spread and actively transigrate through the endothelial barrier. Understanding of the mechanisms of this critical step, known as tumor cell extravasation is essential for development of targeted therapies to prevent metastasis. Various mechanisms by which platelets enhance tumor cell extravasation have been proposed. Overall, a wide range of mediators released by platelets could be involved as shown in mouse models deficient either in granules content or in the secretion machinery (71, 72). More specifically, first the release of adenosine nucleotides stored in the δ -granules has been shown to play a key role. This process appears to be mediated by the endothelial receptor P2Y $_2$, since mice deficient for this receptor, exhibited a reduced migration of tumor cells through the endothelial barrier (71). MMPs stored in the α -granules may also be involved in tumor cell extravasation through their ability to degrade the vascular basement membrane and the subendothelial ECM.

Platelet/tumor cell aggregates arrested in the vasculature may cause endothelial cell retraction and exposure of subendothelial collagen thereby facilitating platelet/tumor cell extravasation. Mice lacking the platelet-restricted collagen receptor GPVI exhibited decreased metastasis of Lewis lung carcinoma and melanoma cells (73). Interestingly, this important platelet receptor in thrombosis, is not critical for normal hemostasis (74), which makes it a potentially interesting target devoid of bleeding complications.

Activated platelets release the endothelial agonist S1P (sphingosine-1-phosphate), a potent inhibitor of vascular leakage. In contrast, LPA stored in α -granules induce permeability of brain endothelial cells (75, 76). These two factors may affect vascular

integrity during tumor cell extravasation. Serotonin is also released from activated platelets and can modulate the vascular tone by inducing vasoconstriction or vasodilation (77). Circulating tumor cells increase serotonin plasma levels and blockade of serotonin receptors or calcium channels have been shown to inhibit experimental liver metastasis (78). However, a direct role of platelet derived serotonin in tumor cell extravasation has not been demonstrated. Another potent vasoactive factor released by platelets is histamine, which enhances vascular permeability and increases leukocyte extravasation (79, 80). Tumor cell incorporation to platelet-leukocyte aggregates arrested at vasculature may generate a favorable milieu stimulating tumor cell extravasation. It remains to be addressed, whether histamine may impact tumor cell extravasation by this mechanism.

NETs have been reported to sequester tumor cells, thereby facilitating tumor cell extravasation in the context of systemic infection (81). Platelets are known to promote formation of NETs in sepsis through platelet-derived Toll-like receptor 4 (TLR4). Recently, platelet derived TLR4 was shown to promote metastasis by interaction with tumor cell-released high-mobility group box1 protein (82). Whether platelet TLR4 may also play a role in tumor cell extravasation or in other steps of cancer progression through mechanisms involving NETs needs to be addressed.

Thus again, a wide range of mediators and various mechanisms have been shown or are suspected to play a role in tumor cell extravasation. Future studies are needed to evaluate at which metastasis step α - and/or δ -granule content could have a functional role. No doubt also that other components of the platelet releasate will appear to play specific roles in these complex processes, including those affecting endothelial permeability, vascular tone and specific cellular mechanisms.

Platelets and metastatic niches

During metastasis, host cells are recruited by disseminated tumor cells to form specialized microenvironment niches. Recently, Labelle et al. demonstrated that

tumor cell-activated platelets release CXC chemokine ligand (CXCL) 5 and CXCL-7, which bind to CXCR2 (CXC chemokine receptor 2) at the surface of granulocytes favoring their recruitment to platelet-tumor cell aggregates. This process establishes an early metastatic niche within 2 hours of tumor cell initial arrest in the lung vasculature. The recruitment of granulocytes to the early metastatic niche was strictly dependent on platelet activation (83).

Early metastatic niche differs from Paget's seed and soil theory, which requires a favorable microenvironment (premetastatic niche) that may evolve allowing tumor cell engraftment (metastatic niche) and proliferation at secondary sites. An interesting question is whether platelets affect metastasis by participating in establishment of metastatic niches, and/or through their interplay with molecular and cellular components of metastatic niches, such as fibronectin, collagen, tenascins, bone marrow derived cells or fibroblasts which provide a permissive milieu for the arrival and growth of tumor cells. Tenascin-C (TNC) has been shown to contribute to the generation of stem like niches supporting cancer and thereby initiating survival and proliferation at newly colonized metastatic sites in breast cancer (84). Moreover, cancer associated fibroblasts producing TNC and VEGF have been reported to provide the permissive "soil" for metastatic colonization (85). TNC is also able to recruit flowing platelets directly through integrin $\alpha 2\beta 1$ and indirectly, through integrin $\alpha 1\text{Ib}\beta 3$ and the GPIb-IX complex which bind the plasma VWF adsorbed onto TNC (86). Moreover, TNC induces accumulation of fibrin through down regulation of tissue plasminogen activator, which is also known to support flowing platelet recruitment (87). Whether the platelet/TNC interplay influences metastasis remains also to be evaluated.

Therapeutic implications

Current therapeutic options targeting platelets to improve cancer treatment are scarce. Available anti-platelet drugs are aspirin, P2Y₁₂ receptor-targeting drugs and

integrin $\alpha 1\text{Ib}\beta 3$ blockers which interfere with platelet activation and aggregation (88). As antithrombotic drugs, they are frequently combined to provide a better clinical outcome, notably aspirin and clopidogrel, which represents the current standard of care in the treatment and secondary prevention of coronary artery disease. Considering the role played by platelets in several steps of the tumor process it may appear obvious to use antiplatelet drugs in order to improve cancer patient outcomes in preventive and therapeutic settings. On the other hand, the mechanisms involved in platelet/tumor cells interactions and in the various steps of tumor progression may not be impaired by existing drugs and may require new pharmacological approaches, more specific of these interactions.

Effects of aspirin on cancer: Do platelets play a role?

Aspirin (acetylsalicylic acid) irreversibly inactivates the cyclooxygenase activity of COX-1 and COX-2. As a result formation of prostanoids is prevented, including PGD₂, PGE₂, PGF₂ α , PGI₂ and TxA₂. COX-1 is the only isoform in platelets, while COX-2 is expressed in a large variety of cells, including epithelial cells, endothelial cells and monocytes. Aspirin is known to inhibit more potently COX-1 (>50–100-fold) in anucleated platelets than COX-2 in other cells, firstly because COX-2 is less sensitive to aspirin and secondly because nucleated cells continuously synthesize this enzyme. An inhibitory role of aspirin in cancer was initially reported by Gasic et al., in 1973, who observed metastatic inhibition of MCA6 ascites sarcoma cells in aspirin treated mice (89). Later, additional studies confirmed this effect of aspirin which prevented tumor growth and metastasis in mice (90). A protumorigenic role was initially attributed to the COX-2 isoform, but more recent studies showed that COX-1 also participates in tumorigenesis. Genetic disruption of either COX-1 or COX-2 was shown to reduce intestinal polyp formation in mice (91). COX-1 and COX-2 pathways operate sequentially in intestinal tumorigenesis. Some studies hypothesized that activated platelets may promote tumorigenesis by

triggering COX-2 expression in stromal cells via release of IL- β , PDGF and TGF- β , which lead to tumor progression (92).

In humans, observational studies indicated that regular use of aspirin was associated with a reduced risk of melanoma, lung, liver, prostate, skin, esophageal and colorectal cancers (90). Moreover, prospective clinical trials have shown that daily aspirin intake as recommended for the prevention of cardiovascular disease reduces the incidence of colorectal cancer after 8–10 years and mortality (93). In more recent analysis of randomized trials, daily treatment with aspirin (75 mg) also decreased the risk of all cancer with metastases (94). Of course, effects of anti-platelet drugs may be dependent on cancer type, stage of tumor progression and cancer risks factors. Aspirin was reported to be more efficient in patients with colorectal cancer than other cancers (94). Reimers et al., reported that aspirin use after diagnosis improves survival of older adults with colon cancer (95). Recently in a meta-analysis of nine observational studies of patients with Barrett's esophagus, aspirin was associated with reduced risk of esophageal adenocarcinoma or high grade dysplasia (96). Conversely, a consensus panel concluded in 2011 that aspirin has only minimal effects on prevention of breast cancer (97).

These data collectively support preventive and therapeutic effects of aspirin on several types of cancer. The key remaining question is whether aspirin exerts its role on cancer through platelet dependent and/or independent mechanisms. The efficacy of the so-called "antiplatelet" lowest dose of 75 mg/day is so far the strongest argument for a role of platelets in cancer-related events.

However, COX-independent mechanisms of aspirin have also been proposed to contribute to its preventive effects in colorectal cancer. Aspirin induces degradation of I κ B α , leading to the nuclear translocation of NF- κ B, thereby resulting in cell apoptosis. This effect has been demonstrated in vitro and in murine models of colorectal cancer (98). In addition, aspirin was shown to dose-dependently inhibit Wnt/ β -catenin, which is a major oncogenic pathway in several cancers (99). Aspirin

could also trigger autophagy in colon cancer cells by inhibiting mammalian Target of Rapamycin (mTOR) signaling effectors S6K1 (S6 kinase 1) and 4E-BP1 (eukaryotic initiation factor 4E binding protein-1) through both AMPK (adenosine monophosphate-activated protein kinase)-dependent and independent mechanisms (100). Of note, these pro-apoptotic and anti-proliferative effects triggered by COX-independent action of aspirin have been evidenced in vitro and usually require very high concentrations of aspirin, in the millimolar range, which likely discounts the occurrence of such effects with anti-platelet doses of 75 to 100 mg/day in humans.

Potential limitations of anti-platelet drugs may include an elevated risk of bleeding, especially in patients who are already thrombocytopenic due to chemotherapy or radiotherapy. However, the use of aspirin showed benefit in patients with advanced stage of colorectal cancer, while no major bleeding complications were observed (101). In contrast, other studies suggested caution in the use of aspirin, because a high risk of gastrointestinal bleedings and hemorrhagic strokes were observed in virtually all studies (94, 102–104). Therefore, identification of relevant biomarkers of response to aspirin is urgently needed.

Large randomized clinical trials in patients with active malignancy are required to establish benefits of aspirin. Thus, so far the popular hypothesis that the chemopreventive and chemotherapeutic effects of aspirin could be due to its anti-platelet effects is still speculative and needs to be clearly established. Ongoing randomized clinical trials in patients with different stages of colorectal, non-small cell lung and breast cancers should answer this question in the near future. In one proposed study the investigators have notably focused on platelet dependent mechanisms of aspirin in patients with colorectal cancer (ClinicalTrials.gov NCT02125409).

Evidence of anti-tumoral effects of P2Y₁₂-targeting drugs

The P2Y₁₂ receptor is a purinergic Gai₂-coupled ADP receptor expressed notably on platelets and known to play a criti-

cal role in thrombus stability in vivo (105). Two different classes of drugs target this receptor. These are the thienopyridine compounds ticlopidine, clopidogrel and prasugrel which are prodrugs. Their active metabolites irreversibly block the binding of ADP to the receptor, resulting in decreased platelet activation and aggregation, due in large part to a reduced inside-out activation of platelet integrin α IIb β 3 (106). Direct reversible P2Y₁₂ receptor antagonists also exist, namely ticagrelor and cangrelor which display similar inhibitory properties. Clopidogrel is widely used clinically to treat coronary artery, cerebrovascular and peripheral vascular diseases (107).

In contrast to aspirin, there is no large scale clinical evidence for any beneficial effect of clopidogrel or any P2Y₁₂ targeting drug in cancer patients. As an example, cancer mortality among the CHARISMA trial patients was not influenced by clopidogrel (108).

However, in vitro data indicate that inhibition of the P2Y₁₂ receptor results in decreased TCIPA, which is not surprising if one remembers the importance of ADP in tumor cells interactions with platelets (109). Recently, in animal models, clopidogrel was shown to reduce cancer progression. Indeed, in syngeneic orthotopic mice models of pancreatic cancer, clopidogrel inhibited tumor development, metastasis and the extent of thrombosis associated with cancer, at a dose of 8 mg/kg which is 4–8 fold the chronic dose in patients and probably induces complete inhibition of ADP-induced platelet aggregation (110). Similarly, ticagrelor has been found to inhibit lung metastasis in mice (111), whereas genetic deficiency of P2Y₁₂, has also been shown to inhibit lung colonization by Lewis lung carcinoma and melanoma cells (112).

Do P2Y₁₂ targeting drugs also affect tumor angiogenesis? Earlier work has shown that ticlopidine displayed anti-angiogenic properties in a rat model of subcutaneous fibrin gel chambers (113). However, the thienopyridine SR 25989 R which is the inactive stereoisomer of clopidogrel and completely devoid of any anti-platelet action also appeared to inhibit angiogenesis in both in vitro and in vivo conditions and exhibits an inhibitory effect in an ex-

perimental model of metastasis (114, 115) clearly indicating an off-target effect of thienopyridine compounds on angiogenesis.

Another aspect to consider is the fact that the P2Y₁₂ receptor is expressed in cells other than platelets (116), such as on osteoclasts (117). Su et al., reported that mice treated with clopidogrel were protected from pathologic osteolysis and bone loss associated with tumor growth (117). The benefits of P2Y₁₂ antagonists in preventing pathological osteolysis and metastasis have been shown in mice, but their effects remain to be studied in humans with malignancy.

Finally, an intriguing observation has been reported concerning prasugrel, the potent third generation thienopyridine. In the TRITON-TIMI 38 trial which compared clopidogrel to prasugrel in the setting of percutaneous coronary intervention, a higher incidence of solid tumors and cancer death was recorded in the prasugrel arm – reviewed by Nanau et al. (118). A possible occurrence in this trial of a chance effect should be taken into consideration. Of note, no tumorigenic effects of prasugrel was reported in mouse xenograft models of prostate, lung and colon cancers and genotoxicity, carcinogenicity assays (119). If the tumor causing effects of prasugrel is confirmed in humans, it would be important to establish whether it is due to an off-target effect or to a more aggressive antiplatelet regimen as compared to clopidogrel.

Effect of integrin $\alpha\text{IIb}\beta 3$ blockade on cancer

Several $\alpha\text{IIb}\beta 3$ antagonists, which are exclusively used in acute phases of cardiovascular diseases (120), notably integrilin, have shown to provide some benefits in experimental metastasis (65). It remains to be confirmed that the effect of integrilin is limited to its action on platelets since it has been reported that some cancer cell lines also express $\alpha\text{IIb}\beta 3$ (121). Zhang et al., proposed an anti-metastatic approach based on the use of a humanized single chain antibody directed against integrin $\beta 3$ (122). One drawback in targeting platelet $\alpha\text{IIb}\beta 3$ is the associated bleeding risk which pre-

cludes chronic use of such drugs in patients (120). A potential approach to limit the risk of bleedings is to specifically target this integrin under its active conformation. This has been proposed by Stoll et al., demonstrating that a single chain antibody directed against activated $\alpha\text{IIb}\beta 3$ provide an antithrombotic effect without increasing the bleeding risk (123). Future studies should be conducted to evaluate the effect of such a tool in animal models of metastasis.

Other pharmacological approaches

Platelet-mimicry of tumor cells – Co-targeting anti-platelet strategies?

Therapeutic strategies disrupting the multiple interactions between platelets and tumor cells at an earlier stage may provide advantages to limit tumor progression and metastasis. However, identification of patients with early disease is rather challenging. Tumor cells may express several megakaryocytic genes, such as integrin $\alpha\text{IIb}\beta 3$, PECAM-1 (Platelet-Endothelial Cell Adhesion Molecule-1)/CD31, thrombin receptors, platelet-type 12-lipoxygenase and acquire a geno-phenotype mimicking platelets (121). This epiphenomenon of platelet mimicry and shared receptors may enable platelet antagonist and inhibitors to simultaneously target tumor cells and platelets and dissipate tumor and platelet cross-talks with tumor microenvironment. Systematic analysis of the markers of platelet mimicry in various cancers and clinical trials investigating the efficacy of shared molecular targets may provide new therapeutic modalities in management of active malignancy.

Platelets as biocompatible drug delivery system?

As mentioned below, targeting of platelet/tumor cell interactions may represent an attractive therapeutic approach. Interestingly, alternative approaches taking advantage of tumor cells preference in interacting with platelets have also been proposed. Indeed, the attributes of platelets, namely,

their tendency to uptake a variety of compounds and release them when activated, have been suggested as an efficient drug delivery system (124). Future studies are needed to address whether this strategy might be more effective in killing cancer cells than normal cells while preserving normal physiology.

Experimental limitations

Existing research has provided compelling biological evidence in support of attempting to disrupt physical and functional platelet/tumor interactions and platelet/tumor microenvironment cross-talks to attenuate or inhibit tumor growth, invasion, angiogenesis and tumor metastasis. In mouse models, anti-platelet drugs were shown to reduce tumor growth, angiogenesis and metastasis. However, some data should be interpreted with caution, since evidence has been accumulated for species specific differences in terms of platelet and tumor cell receptors. For example, human platelets are primarily activated by thrombin through PAR-1 and PAR-4, whereas mouse platelets do not express PAR-1 and are predominantly activated through PAR-3 and PAR-4. In several studies, experimental approaches used to evaluate involvement of platelets to induce thrombocytopenia were essentially based on the use of neuraminidase or anti-platelet antibodies. The major drawback of neuraminidase is that sialic acid removal occurs not only on platelets but also on other vascular cells, potentially affecting several biological functions. Thrombocytopenia inducing antibodies present also disadvantages, since they can trigger platelet activation leading to the release of factors potentially affecting molecular pathways in cancer or other circulating cells. Another important point to revisit is that many mouse models used to evaluate mediators released by platelets or platelet receptors in metastasis dismiss the early stages of the metastatic cascade. For example, the frequently used injection of a large number of tumor cells into the circulatory system does not accurately mimic the disease progression as it occurs in patients. Syngeneic orthotopic and genetic mouse models of human cancers are more relevant since they recapitulate progressive

stages of cancer starting from primary tumor development, to tumor angiogenesis and establishment of metastases. Thus, well-designed pre-clinical models with defined alterations in tumors cells, platelets or in other host cells successfully mimicking human diseases are mandatory to dissect the cellular and molecular mechanisms, and to assess the potential and selectivity of anti-platelet agents.

Studies aiming to understand the role of platelets in different steps of metastasis have been mainly conducted in vitro, notably the interactions of platelets between with tumor cells, endothelial cells, leukocytes. This allowed dissection of many mechanisms involved and modulation of signaling pathways. In vivo rabbit and mouse ear chambers have provided some hints on the dynamics of tumor cell intravasation, migration and arrest along the vasculature and platelet-thrombosis formation. However conditions are different from native environment, which may not recapitulate more relevantly different steps of tumor progression and metastatic cascade. Tumor cell-endothelial interactions have been extensively studied by microscopy techniques in mice, and not so far in real time. There are also difficulties in tracking platelet-tumor cell interactions within the vasculature. Recently, multiphoton microscopy techniques have been developed to track platelet-tumor cell interactions in mouse liver sinusoid vessels (125). Highly resolutive real time imaging techniques are needed, to analyze more profoundly behavior of platelet/tumor/vascular cell interactions in animal models, thereby leading to more fine elucidation of mechanisms.

Conclusion

Literature describing the contribution of platelets to cancer is significantly growing. It has revealed a highly complex role and the involvement of platelets in many bidirectional interactions with tumor cells and tumor microenvironment. Platelets appear to influence many functions of tumor cells and escort them through different stages of tumor progression. In turn, tumor cells may use their ability to hijack important

biological functions of platelets to increase their survival and proliferation capacity promoting the pathogenesis of cancer. In addition, clinical data have led to speculate that anti-platelet medication may provide anti-cancer effects by disturbing platelet-tumor cell cross-talks. However, further fundamental, translational and clinical studies are needed before these drugs can be introduced in clinical cancer care.

Conflict of interest

The authors declare that they have no conflict of interest.

References

- Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell* 2011; 144: 646–674.
- Hanahan D, Coussens LM. Accessories to the crime: functions of cells recruited to the tumor microenvironment. *Cancer Cell* 2012; 21: 309–322.
- Olson OC, Joyce JA. Microenvironment-mediated resistance to anticancer therapies. *Cell Res* 2013; 23: 179–181.
- Varki A. Trousseau's syndrome: multiple definitions and multiple mechanisms. *Blood* 2007; 110: 1723–1729.
- Prandoni P, Falanga A, Piccioli A. Cancer and venous thromboembolism. *Lancet Oncol* 2005; 6: 401–410.
- Tranum BL, Haut A. Thrombocytosis: platelet kinetics in neoplasia. *J Lab Clin Med* 1974; 84: 615–619.
- Khorana AA. Cancer and coagulation. *Am J Hematol* 2012; 87 (Suppl 1): S82–S87.
- Versteeg HH, Heemskerk JW, Levi M, Reitsma PH. New fundamentals in hemostasis. *Physiol Rev* 2013; 93: 327–358.
- Nurden AT. Platelets, inflammation and tissue regeneration. *Thromb Haemost* 2011; 105 (Suppl 1): S13–S33.
- Bertozzi CC, Hess PR, Kahn ML. Platelets: covert regulators of lymphatic development. *Arterioscler Thromb Vasc Biol* 2010; 30: 2368–2371.
- Semple JW, Italiano JE Jr, Freedman J. Platelets and the immune continuum. *Nature reviews. Immunology* 2011; 11: 264–274.
- Stegner D, Dutting S, Nieswandt B. Mechanistic explanation for platelet contribution to cancer metastasis. *Thromb Res* 2014; 133 (Suppl 2): S149–S157.
- Riedl J, Pabinger I, Ay C. Platelets in cancer and thrombosis. *Hämostaseologie* 2014; 34: 54–62.
- Goubran HA, Stakiw J, Radosevic M, Burnouf T. Platelet-cancer interactions. *Semin Thromb Hemost* 2014; 40: 296–305.
- Gay LJ, Felding-Habermann B. Contribution of platelets to tumour metastasis. *Nature reviews. Cancer* 2011; 11: 123–134.

- Di Nisio M, Porreca E, Otten HM, Rutjes AW. Primary prophylaxis for venous thromboembolism in ambulatory cancer patients receiving chemotherapy. *The Cochrane database of systematic reviews* 2014; 8: CD008500.
- Stone RL et al. Paraneoplastic thrombocytosis in ovarian cancer. *N Engl J Med* 2013; 366: 610–618.
- Sasaki Y et al. Production of thrombopoietin by human carcinomas and its novel isoforms. *Blood* 1999; 94: 1952–1960.
- Rank A et al. Circulating microparticles in patients with benign and malignant ovarian tumors. *Anticancer Res* 2012; 32: 2009–2014.
- Jurasz P, Alonso-Escolano D, Radomski MW. Platelet-cancer interactions: mechanisms and pharmacology of tumour cell-induced platelet aggregation. *Br J Pharmacol* 2004; 143: 819–826.
- Raica M, Cimpean AM, Ribatti D. The role of podoplanin in tumor progression and metastasis. *Anticancer Res* 2008; 28(5B): 2997–3006.
- Dang Q, Liu J, Li J, Sun Y. Podoplanin: a novel regulator of tumor invasion and metastasis. *Med Oncol* 2014; 31: 24.
- Lowe KL, Navarro-Nunez L, Watson SP. Platelet CLEC-2 and podoplanin in cancer metastasis. *Thromb Res* 2012; 129 (Suppl 1): S30–S37.
- Takagi S et al. Platelets promote tumor growth and metastasis via direct interaction between Aggrus/podoplanin and CLEC-2. *PloS one* 2013; 8: e73609.
- Han X, Guo B, Li Y, Zhu B. Tissue factor in tumor microenvironment: a systematic review. *J Hematol Oncol* 2014; 7: 54.
- Demers M et al. Cancers predispose neutrophils to release extracellular DNA traps that contribute to cancer-associated thrombosis. *Proc Natl Acad Sci USA* 2012; 109: 13076–13081.
- Carmeliet P, Jain RK. Molecular mechanisms and clinical applications of angiogenesis. *Nature* 2011; 473: 298–307.
- Italiano JE Jr et al. Angiogenesis is regulated by a novel mechanism: pro- and antiangiogenic proteins are organized into separate platelet alpha granules and differentially released. *Blood* 2008; 111: 1227–1233.
- Battinelli EM, Markens BA, Italiano JE Jr. Release of angiogenesis regulatory proteins from platelet alpha granules: modulation of physiologic and pathologic angiogenesis. *Blood* 2011; 118: 1359–1369.
- Bambace NM, Levis JE, Holmes CE. The effect of P2Y-mediated platelet activation on the release of VEGF and endostatin from platelets. *Platelets* 2010; 21: 85–93.
- Kamykowski J, Carlton P, Sehgal S, Storrie B. Quantitative immunofluorescence mapping reveals little functional coclustering of proteins within platelet alpha-granules. *Blood* 2011; 118: 1370–1373.
- Jonnalagadda D, Izu LT, Whiteheart SW. Platelet secretion is kinetically heterogeneous in an agonist-responsive manner. *Blood* 2012; 120: 5209–5216.
- Klement GL et al. Platelets actively sequester angiogenesis regulators. *Blood* 2009; 113: 2835–2842.
- Feng W et al. A novel role for platelet secretion in angiogenesis: mediating bone marrow-derived cell mobilization and homing. *Blood* 2011; 117: 3893–3902.

35. Kuznetsov HS et al. Identification of luminal breast cancers that establish a tumor-supportive macroenvironment defined by proangiogenic platelets and bone marrow-derived cells. *Cancer discovery* 2012; 2: 1150–1165.
36. Pipili-Synetos E, Papadimitriou E, Maragoudakis ME. Evidence that platelets promote tube formation by endothelial cells on matrigel. *Br J Pharmacol* 1998; 125: 1252–1257.
37. Qi J, Goralnick S, Kreutzer DL. Fibrin regulation of interleukin-8 gene expression in human vascular endothelial cells. *Blood* 1997; 90: 3595–3602.
38. Kim HK, Song KS, Chung JH, Lee KR, Lee SN. Platelet microparticles induce angiogenesis in vitro. *Br J Haematol* 2004; 124: 376–384.
39. Janowska-Wieczorek A et al. Microvesicles derived from activated platelets induce metastasis and angiogenesis in lung cancer. *International journal of cancer. J Int Cancer* 2005; 113: 752–760.
40. Brill A, Dashevsky O, Rivo J et al. Platelet-derived microparticles induce angiogenesis and stimulate post-ischemic revascularization. *Cardiovasc Res* 2005; 67: 30–38.
41. Helley D et al. Platelet microparticles: a potential predictive factor of survival in hormone-refractory prostate cancer patients treated with docetaxel-based chemotherapy. *Eur Urol* 2009; 56: 479–484.
42. Kim HK et al. Elevated levels of circulating platelet microparticles, VEGF, IL-6 and RANTES in patients with gastric cancer: possible role of a metastasis predictor. *Eur J Cancer* 2003; 39: 184–191.
43. Twombly R. Avastin's uncertain future in breast cancer treatment. *J Natl Cancer Inst* 2011; 103: 458–460.
44. Fidler IJ. The pathogenesis of cancer metastasis: the 'seed and soil' hypothesis revisited. *Nature reviews. Cancer* 2003; 3: 453–458.
45. Gasic GJ, Gasic TB, Stewart CC. Antimetastatic effects associated with platelet reduction. *Proc Natl Acad Sci USA* 1968; 61: 46–52.
46. Karpatkin S, Pearlstein E, Ambrogio C, Collier BS. Role of adhesive proteins in platelet tumor interaction in vitro and metastasis formation in vivo. *J Clin Invest* 1988; 81: 1012–1019.
47. Camerer E et al. Platelets, protease-activated receptors, and fibrinogen in hematogenous metastasis. *Blood* 2004; 104: 397–401.
48. Labelle M, Hynes RO. The initial hours of metastasis: the importance of cooperative host-tumor cell interactions during hematogenous dissemination. *Cancer Discovery* 2012; 2: 1091–1099.
49. Nieswandt B, Hafner M, Echtenacher B, Mannel DN. Lysis of tumor cells by natural killer cells in mice is impeded by platelets. *Cancer Res* 1999; 59: 1295–1300.
50. Placke T et al. Platelet-derived MHC class I confers a pseudonormal phenotype to cancer cells that subverts the antitumor reactivity of natural killer immune cells. *Cancer Res* 2012; 72: 440–448.
51. Palumbo JS et al. Platelets and fibrin(ogen) increase metastatic potential by impeding natural killer cell-mediated elimination of tumor cells. *Blood* 2005; 105: 178–185.
52. Skov Madsen P, Hokland P, Hokland M. Secretory products from thrombin-stimulated human platelets exert an inhibitory effect on NK-cytotoxic activity. *Acta pathologica, microbiologica, et immunologica Scandinavica. Section C, Immunology* 1986; 94: 193–200.
53. Kopp HG, Placke T, Salih HR. Platelet-derived transforming growth factor-beta down-regulates NKG2D thereby inhibiting natural killer cell anti-tumor reactivity. *Cancer Res* 2009; 69: 7775–7783.
54. Lee JC, Lee KM, Kim DW, Heo DS. Elevated TGF-beta1 secretion and down-modulation of NKG2D underlies impaired NK cytotoxicity in cancer patients. *J Immunol* 2004; 172: 7335–7340.
55. Coupland LA, Chong BH, Parish CR. Platelets and P-selectin control tumor cell metastasis in an organ-specific manner and independently of NK cells. *Cancer Res* 2012; 72: 4662–4671.
56. Steinestel K, Eder S, Schrader AJ, Steinestel J. Clinical significance of epithelial-mesenchymal transition. *Clin Transl Med* 2014; 3: 17.
57. Dovizio M et al. Pharmacological inhibition of platelet-tumor cell cross-talk prevents platelet-induced overexpression of cyclooxygenase-2 in HT29 human colon carcinoma cells. *Mol Pharmacol* 2013; 84: 25–40.
58. Labelle M, Begum S, Hynes RO. Direct signaling between platelets and cancer cells induces an epithelial-mesenchymal-like transition and promotes metastasis. *Cancer cell* 2011; 20: 576–590.
59. Alves CS, Burdick MM, Thomas SN et al. The dual role of CD44 as a functional P-selectin ligand and fibrin receptor in colon carcinoma cell adhesion. *Am J Physiol Cell Physiol* 2008; 294: C907–916.
60. Qi CL et al. P-selectin-mediated platelet adhesion promotes the metastasis of murine melanoma cells. *PLoS One* 2014; 9: e91320.
61. Coupland LA, Parish CR. Platelets, selectins, and the control of tumor metastasis. *Semin Oncol* 2014; 41: 422–434.
62. McCarty OJ, Mousa SA, Bray PF, Konstantopoulos K. Immobilized platelets support human colon carcinoma cell tethering, rolling, and firm adhesion under dynamic flow conditions. *Blood* 2000; 96: 1789–1797.
63. Zhang C et al. Modified heparins inhibit integrin alpha(IIb)beta(3) mediated adhesion of melanoma cells to platelets in vitro and in vivo. *International journal of cancer. J Int Cancer* 2009; 125: 2058–2065.
64. Bakewell SJ et al. Platelet and osteoclast beta3 integrins are critical for bone metastasis. *Proc Natl Acad Sci USA* 2003; 100: 14205–14210.
65. Boucharaba A et al. Platelet-derived lysophosphatidic acid supports the progression of osteolytic bone metastases in breast cancer. *J Clin Invest* 2004; 114: 1714–1725.
66. Pilch J, Habermann R, Felding-Habermann B. Unique ability of integrin alpha(v)beta 3 to support tumor cell arrest under dynamic flow conditions. *J Biol Chem* 2002; 277: 21930–21938.
67. Morimoto K et al. Interaction of cancer cells with platelets mediated by Necl-5/poliiovirus receptor enhances cancer cell metastasis to the lungs. *Oncogene* 2008; 27: 264–273.
68. Leblanc R et al. Interaction of platelet-derived autotaxin with tumor integrin alphaVbeta3 controls metastasis of breast cancer cells to bone. *Blood* 2014; 124: 3141–3150.
69. Jain S et al. Platelet glycoprotein Ib alpha supports experimental lung metastasis. *Proc Natl Acad Sci USA* 2007; 104: 9024–9028.
70. Erpenbeck L, Nieswandt B, Schon M et al. Inhibition of platelet GPIb alpha and promotion of melanoma metastasis. *J Invest Dermatol* 2010; 130: 576–586.
71. Schumacher D, Strilic B, Sivaraj KK et al. Platelet-derived nucleotides promote tumor-cell transendothelial migration and metastasis via P2Y2 receptor. *Cancer Cell* 2013; 24: 130–137.
72. Guerrero JA et al. Gray platelet syndrome: Pro-inflammatory megakaryocytes and alpha-granule loss cause myelofibrosis and confer resistance to cancer metastasis in mice. *Blood* 2014; 124: 3624–3635.
73. Jain S, Russell S, Ware J. Platelet glycoprotein VI facilitates experimental lung metastasis in syngenic mouse models. *J Thromb Haemost* 2009; 7: 1713–1717.
74. Zahid M et al. The future of glycoprotein VI as an antithrombotic target. *J Thromb Haemost* 2012; 10: 2418–2427.
75. Schaphorst KL et al. Role of sphingosine-1 phosphate in the enhancement of endothelial barrier integrity by platelet-released products. *Am J Physiol* 2003; 285: L258–L267.
76. Yin F, Watsky MA. LPA and S1P increase corneal epithelial and endothelial cell transcellular resistance. *Invest Ophthalmol Vis Sci* 2005; 46: 1927–1933.
77. Cote F, Fligny C, Fromes Y et al. Recent advances in understanding serotonin regulation of cardiovascular function. *Trends Mol Med* 2004; 10: 232–238.
78. Skolnik G, Bagge U, Blomqvist G et al. The role of calcium channels and serotonin (5-HT2) receptors for tumour cell lodgement in the liver. *Clin Exp Metastasis* 1989; 7: 169–174.
79. Kuna P et al. RANTES, a monocyte and T lymphocyte chemotactic cytokine releases histamine from human basophils. *J Immunol* 1992; 149: 636–642.
80. Medina VA, Rivera ES. Histamine receptors and cancer pharmacology. *Br J Pharmacol* 2010; 161: 755–767.
81. Cools-Lartigue J et al. Neutrophil extracellular traps sequester circulating tumor cells and promote metastasis. *J Clin Invest* 2013; 123: 3446–3458.
82. Yu LX et al. Platelets promote tumour metastasis via interaction between TLR4 and tumour cell-released high-mobility group box1 protein. *Nature Commun* 2014; 5: 5256.
83. Labelle M, Begum S, Hynes RO. Platelets guide the formation of early metastatic niches. *Proc Natl Acad Sci USA* 2014; 111: E3053–E3061.
84. Oskarsson T et al. Breast cancer cells produce tenascin C as a metastatic niche component to colonize the lungs. *Nature Med* 2011; 17: 867–874.
85. O'Connell JT et al. VEGF-A and Tenascin-C produced by S100A4+ stromal cells are important for metastatic colonization. *Proc Natl Acad Sci USA* 2011; 108: 16002–16007.
86. Schaff M et al. Novel function of tenascin-C, a matrix protein relevant to atherosclerosis, in platelet recruitment and activation under flow. *Arterioscler Thromb Vasc Biol* 2011; 31: 117–124.
87. Brellier F et al. Tenascin-C triggers fibrin accumulation by downregulation of tissue plasminogen activator. *FEBS letters* 2011; 585: 913–920.
88. Patrono C, Baigent C, Hirsh J et al. Antiplatelet drugs: American College of Chest Physicians Evidence-Based Clinical Practice Guidelines. *Chest* 2008; 133 (6 Suppl): 199S–233S.

89. Gasic GJ, Gasic TB, Galanti N et al. Platelet-tumor-cell interactions in mice. The role of platelets in the spread of malignant disease. *Int J Cancer* 1973; 11: 704–718.
90. Alfonso L, Ai G, Spitale RC, Bhat GJ. Molecular targets of aspirin and cancer prevention. *Br J Cancer* 2014; 111: 61–67.
91. Chulada PC, et al. Genetic disruption of Ptg-1, as well as Ptg-2, reduces intestinal tumorigenesis in Min mice. *Cancer Res* 2000; 60: 4705–4708.
92. Sciuilli MG et al. Platelet activation in patients with colorectal cancer. Prostaglandins Leukot Essent Fatty Acids 2005; 72: 79–83.
93. Flossmann E, Rothwell PM. Effect of aspirin on long-term risk of colorectal cancer: consistent evidence from randomised and observational studies. *Lancet* 2007; 369: 1603–1613.
94. Rothwell PM et al. Effect of daily aspirin on risk of cancer metastasis: a study of incident cancers during randomised controlled trials. *Lancet* 2012; 379: 1591–1601.
95. Reimers MS et al. Aspirin use after diagnosis improves survival in older adults with colon cancer: a retrospective cohort study. *J Am Geriatr Soc* 2012; 60: 2232–2236.
96. Zhang S et al. Cyclooxygenase inhibitors use is associated with reduced risk of esophageal adenocarcinoma in patients with Barrett's esophagus: a meta-analysis. *Br J Cancer* 2014; 110: 2378–2388.
97. Cuzick J, et al. Preventive therapy for breast cancer: a consensus statement. *Lancet Oncol* 2011; 12: 496–503.
98. Stark LA et al. Aspirin activates the NF-kappaB signalling pathway and induces apoptosis in intestinal neoplasia in two in vivo models of human colorectal cancer. *Carcinogenesis* 2007; 28: 968–976.
99. Bos CL et al. Effect of aspirin on the Wnt/beta-catenin pathway is mediated via protein phosphatase 2A. *Oncogene* 2006; 25: 6447–6456.
100. Din FV et al. Aspirin inhibits mTOR signaling, activates AMP-activated protein kinase, and induces autophagy in colorectal cancer cells. *Gastroenterology* 2012; 142: 1504–1515.
101. Neugut AI. Aspirin as adjuvant therapy for stage III colon cancer: standard of care? *JAMA Int Med* 2014; 174: 739–741.
102. Seshasai SR et al. Effect of aspirin on vascular and nonvascular outcomes: meta-analysis of randomized controlled trials. *Arch Int Med* 2013; 172: 209–216.
103. Sostres C, Lanas A. Gastrointestinal effects of aspirin. *Nature reviews. Gastroenterol Hepatol* 2011; 8: 385–394.
104. McQuaid KR, Laine L. Systematic review and meta-analysis of adverse events of low-dose aspirin and clopidogrel in randomized controlled trials. *Am J Med* 2006; 119: 624–638.
105. Gachet C. Regulation of platelet functions by P2 receptors. *Annu Rev Pharmacol Toxicol* 2006; 46: 277–300.
106. Gachet C. P2 receptors, platelet function and pharmacological implications. *Thromb Haemost* 2008; 99: 466–472.
107. Cattaneo M. New P2Y(12) inhibitors. *Circulation* 2010; 121: 171–179.
108. Berger JS et al. Smoking, clopidogrel, and mortality in patients with established cardiovascular disease. *Circulation* 2009; 120: 2337–2344.
109. Bastida E, Escolar G, Almirall L, Ordinas A. Platelet activation induced by a human neuroblastoma tumor cell line is reduced by prior administration of ticlopidine. *Thromb Haemost* 1986; 55: 333–337.
110. Mezouar S, Darbousset R, Dignat-George F et al. Inhibition of platelet activation prevents the P-selectin and integrin-dependent accumulation of cancer cell microparticles and reduces tumor growth and metastasis in vivo. *Int J Cancer* 2015; 136: 462–475.
111. Gebremeskel S, LeVatte T, Liwski RS et al. The reversible P2Y12 inhibitor ticagrelor inhibits metastasis and improves survival in mouse models of cancer. *Int J Cancer* 2015; 136: 234–240.
112. Wang Y et al. Platelet P2Y12 is involved in murine pulmonary metastasis. *PLoS One* 2013; 8: e80780.
113. Rohr S et al. Quantitative image analysis of angiogenesis in rats implanted with a fibrin gel chamber. *Nouvelle Revue Francaise d'Hematologie* 1992; 34: 287–294.
114. Klein-Soyer C et al. Angiogenesis inhibitor SR 25989 upregulates thrombospondin-1 expression in human vascular endothelial cells and foreskin fibroblasts. *Biol Cell* 1997; 89: 295–307.
115. Mah-Becherel MC et al. Anti-angiogenic effects of the thienopyridine SR 25989 in vitro and in vivo in a murine pulmonary metastasis model. *Br J Cancer* 2002; 86: 803–810.
116. Gachet C. P2Y(12) receptors in platelets and other hematopoietic and non-hematopoietic cells. *Purinergic Signal* 2012; 8: 609–619.
117. Su X et al. The ADP receptor P2RY12 regulates osteoclast function and pathologic bone remodeling. *J Clin Invest* 2012; 122: 3579–3592.
118. Nanau RM, Delzor F, Neuman MG. Efficacy and safety of prasugrel in acute coronary syndrome patients. *Clin Biochem* 2014; 47: 516–528.
119. Buckley LA et al. Nonclinical assessment of carcinogenic risk and tumor growth enhancement potential of prasugrel, a platelet-inhibiting therapeutic agent. *Int J Toxicol* 2012; 31: 317–325.
120. Bledzka K, Smyth SS, Plow EF. Integrin alphaIIb-beta3: from discovery to efficacious therapeutic target. *Circ Res* 2013; 112: 1189–1200.
121. Timar J et al. Platelet-mimicry of cancer cells: epiphenomenon with clinical significance. *Oncology* 2005; 69: 185–201.
122. Zhang W et al. A humanized single-chain antibody against beta 3 integrin inhibits pulmonary metastasis by preferentially fragmenting activated platelets in the tumor microenvironment. *Blood* 2013; 120: 2889–2898.
123. Stoll P et al. Targeting ligand-induced binding sites on GPIIb/IIIa via single-chain antibody allows effective anticoagulation without bleeding time prolongation. *Arterioscler Thromb Vasc Biol* 2007; 27: 1206–1212.
124. Sarkar S, Alam MA, Shaw J, Dasgupta AK. Drug delivery using platelet cancer cell interaction. *Pharm Res* 2013; 30: 2785–2794.
125. Tanaka K et al. In vivo optical imaging of cancer metastasis using multiphoton microscopy: a short review. *Am J Transl Res* 2014; 6: 179–187.