# Understanding transmissibility patterns of Chagas disease through complex vector-host networks

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#### SUMMARY

Chagas disease is one of the most important vector-borne zoonotic diseases in Latin America. Control strategies could be improved if transmissibility patterns of its aetiologic agent, *Trypanosoma cruzi*, were better understood. To understand transmissibility patterns of Chagas disease in Mexico, we inferred potential vectors and hosts of *T. cruzi* from geographic distributions of nine species of Triatominae and 396 wild mammal species, respectively. The most probable vectors and hosts of *T. cruzi* were represented in a Complex Inference Network, from which we formulated a predictive model and several associated hypotheses about the ecological epidemiology of Chagas disease. We compiled a list of confirmed mammal hosts to test our hypotheses. Our tests allowed us to predict the most important potential hosts of *T. cruzi* and to validate the model showing that the confirmed hosts were those predicted to be the most important hosts. We were also able to predict differences in the transmissibility of *T. cruzi* among triatomine species from spatial data. We hope our findings help drive efforts for future experimental studies.

Key words: Trypanosoma cruzi, potential hosts, spatial data mining, ecological epidemiology.

#### INTRODUCTION

Chagas disease is one of the most important vectorborne zoonotic diseases in Latin America, with six to seven million people infected and 70 million being at risk of acquiring the disease (WHO, 2015). Infection prevention programs are still the most effective tool for controlling Chagas disease transmission (Rodrigues-Coura, 2013); however, controlling Chagas disease in endemic countries is difficult (Abad-Franch *et al.* 2013; Rodrigues-Coura, 2013). Control strategies could be improved, if the transmissibility patterns of the aetiologic agent, *Trypanosoma cruzi*, were better understood.

To better understand the transmissibility patterns of *T. cruzi*, it is essential to consider the potential interactions among its vectors and hosts at a more integrative level. The parasite has hundreds of potential vectors (Triatominae Reduviidae Hemiptera) and potential hosts (Mammalia) (Jansen and Roque, 2010). Considering this diversity of potential vectors and hosts, it is logistically impossible to understand the transmissibility patterns of *T. cruzi* by an exhaustive experimental examination of all potential triatomine–mammal interactions. In addition, biotic interactions among vectors and hosts are usually studied at the level of a particular mammal host, rather than considering ecosystemic patterns of the whole host-vector system. Complex Inference Networks (Stephens *et al.* 2009; González-Salazar and Stephens, 2012), applied to zoonoses, provide a useful alternative for understanding the transmissibility patterns of  $T.\ cruzi$ . The network represents the most probable ecosystem of  $T.\ cruzi$  by showing their potential vectors, hosts and their interactions.

In this paper, we derive a vector-host network for the potential ecological factors - Triatominae and wild mammal species - involved in the transmission cycle of T. cruzi in Mexico. Complex Inference Networks in the context of Chagas disease use the statistical significance of co-occurrences of triatomine and wild mammal species as proxies for their potential interactions, where statistically significant co-occurrences are estimated based on the level of overlap of the species' distribution ranges (Stephens et al. 2009; González-Salazar and Stephens, 2012). The patterns recovered from the network allow us to formulate and test several hypotheses about the ecological epidemiology of Chagas disease. Primarily, we test the idea that the lack of randomness in the co-occurrence patterns can be interpreted as a measure of the relative level of importance of the biotic interactions between a given mammal and triatomine species. If we identify the most relevant biotic interactions between mammal and triatomine

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species, then, taking the presence of such an interaction as a necessary (but not necessarily sufficient) condition for the transmission of the pathogen, the potential hosts and vectors of Chagas disease can be inferred and transmission patterns of T. *cruzi* can be drawn. In this sense, the vector-host network helps us to detect the transmissibility patterns of T. *cruzi* in an ecosystemic way. We discuss how our findings can be complemented by future experimental studies.

#### MATERIALS AND METHODS

#### Study area

To define the study area, records of Triatominae species were projected onto a map of ecoregions (Olson *et al.* 2001). We then considered all ecoregions with at least one species record, which were basically the non-desert ecoregions of the country. Therefore, the study area was all of Mexico, except the Chihuahuan, Sonoran and Baja Californian deserts and the Tamaulipan mezquital (Olson *et al.* 2001).

#### Species data

We focused on wild mammal species from nondesert ecoregions of Mexico and nine Triatominae species that are known as the main Chagas disease vectors in Mexico (Ramsey et al. 2015). We compiled georeferenced localities for Triatominae species according to Lent and Wygodzinsky (1979) and Bargues et al. (2008). Triatoma dimidiata sensu Lent and Wygodzinsky (1979) is considered a species complex (Bargues et al. 2008; Dorn et al. 2009; Monteiro et al. 2013), so we analysed the two main lineages of the study area separately. The final dataset of 3425 unique point records was obtained from national entomological collections (Instituto de Diagnóstico y Referencia Epidemiológica, InDRE, Mexico City; Colección Nacional de Insectos, CNIN, UNAM, Mexico City) and published records (Ramsey et al. 2015). The dataset of mammal species consisted of georeferenced localities for 396 species (Ceballos and Arroyo, 2012). This dataset includes 47 942 unique point records compiled from electronic databases (www.gbif.org, www.conabio. gob.mx).

There is, as has been extensively discussed in the literature, an important question of potential sample bias in such point records; for instance, via the *ad hoc* nature of the collections and the large collecting gaps in space and time (Ponder *et al.* 2001; Graham *et al.* 2004) Such sampling bias constitutes a significant challenge for the success and veracity of analyses of data point records (Yañez-Arenas *et al.* 2014). However, in spite of their potential biases, such databases provide large and important

information resources accumulated over long periods (Ponder *et al.* 2001; Graham *et al.* 2004) for trying to determine the distribution of species as a function of space and time. Thus, it is important to leverage these data, while bearing in mind the impact of such biases. This is even more important for urgent problems of great social impact such as that of emerging diseases.

## Inferred interaction network of triatomines and mammals

We adopted a nonparametric spatial data mining approach to infer potential biotic interactions between mammals and triatomine species using the available point collection data. The general modelling methodology of Stephens et al. (2009) is based on the idea that biotic interactions can be inferred from the locations of taxa as a function of space and time. Clearly, biotic and ecological interactions in general are very complex, giving rise to spatiotemporal distributions that depend on an enormous number of variables, both biotic and abiotic. However, it is reasonable to suppose that the spatio-temporal distributions of taxa, or other ecological variables, reflect all of the factors and their causal interactions that determine them. The question is: To what extent can the existence of ecological interactions be deduced by an analysis of the positions of taxa? To give a simple example, one would expect competitive interactions to lead to different spatial distributions than mutualistic interactions.

In Stephens *et al.* (2009), the degree of co-occurrence between taxa was taken as an observable measure with which potential interactions could be inferred. Although co-occurrence is not equal to biological interaction, a significantly non-random cooccurrence distribution is a *necessary* condition for a biotic interaction between taxa, and as such it can be used to formulate hypotheses that can be checked experimentally. However, it is clearly not a *sufficient* condition.

Applying the methodology to the present case we observed those co-occurrences between mammals and triatominae in geographical space that are more common than would be expected by chance (Fig. 1). We focused our attention on the spatial dependence of the distributions and ignored the temporal aspect, as the data used are not capable of describing reliably temporal changes. As a measure of statistical association, we consider the probability to find a triatomine given the occurrence of a mammal,  $P(T_i|M_j)$ , where  $T_i$  and  $M_j$  represent the presence of the *i*th triatomine and *j*th mammal, respectively.

To determine this probability we divide the geographic region of interest into a uniform grid and then count grid cells according to presence of a given triatomine, presence of a given mammal and/

	Mj	Mj Ti	Mj		Mj Ti	Mj		Mj Ti	
		Ti			Mj Ti			Mj Ti	
		Ti			Ti			Mj Ti	
3	2			3.5				N	
R		N			2			1	

Fig. 1. Co-occurrence pattern of a triatomine (*Ti*) species and a mammal species (*Mj*). Epsilon ( $\varepsilon$ ) values increase when the overlap between mammal and triatomine distributions increases. Relative rank (*R*) values are high for species with high  $\varepsilon$  values.

or co-occurrences of both. The choice of grid-cell size has no biological motivation. It is statistically motivated, being associated with maximizing the effective sample size for counting co-occurrences, a co-occurrence essentially being our fundamental statistical unit. The choice of cell size is known in geography as the 'modifiable areal unit problem'. In terms of forming a spatial grid, there are at least two important considerations: the sizes of the statistical samples of the variables and their degree of correlation. Too fine a grid and there will be no co-occurrences, too rough and there will be little to no discrimination. It was checked explicitly in Stephens et al. (2009) that the relative ranking of mammals by the model was quite insensitive to the cell size over the range 5-100 km (see also Sierra and Stephens, 2012). However, even though this previous research has shown that predictions of potential feeding resources are robust to large changes in the grid-cell size, we have independently assayed three different grid-cell sizes to check how robust our predictions were (Table S1, Supporting information). Based on these results, for our analysis we used 3535 square grid cells of linear size 20 km, as this resolution has been found to give good overall results when considering a large number of distributions simultaneously (Stephens et al. 2009; Sierra and Stephens, 2012).

To evaluate the non-random nature of the cooccurrence distribution we considered the following exact binomial statistical test:

$$\varepsilon(T_i|M_j) = \frac{N_{M_j}(P(T_i|M_j) - P(T_i))}{(N_{M_j}P(T_i)(1 - P(T_i)))^{1/2}}$$
(1)

where  $P(T_i | M_j) = N_{Ti}$  and  $M_{j}/N_{Mj}$  with  $N_{Ti}$  and  $M_j$ being the number of cells where there is a co-occurrence of  $T_i$  and  $M_j$ ,  $N_{Mj}$  is the number of cells with presence of  $M_j$  and  $P(T_i) = N_{Ti}/N$ , where  $N_{Ti}$  is the number of grid cells with point collections of species  $T_i$  and N is the total number of grid cells. This binomial test measures the degree of confidence of the statistical association between  $T_i$  and  $M_j$ , relative to the null hypothesis that the spatial distribution of  $T_i$  is independent of  $M_j$  and distributed randomly over the grid, i.e.  $P(T_i)$ . The sampling distribution of the null hypothesis is a binomial distribution, where every cell is given a probability  $P(T_i)$  of having a point collection of  $T_i$ . The numerator of equation (1) is then the difference between the actual number of co-occurrences of  $T_i$  and  $M_j$  relative to the expected number if the spatial distribution of point collections was obtained from a binomial with sampling probability,  $P(T_i)$ . The denominator of equation (1) is the standard deviation of the binomial distribution (Stephens *et al.* 2009; González-Salazar *et al.* 2013).

The quantitative values of  $\mathcal{E}(T_i | M_i)$  can be interpreted in the standard sense of hypothesis testing. We consider the *P*-value as the probability that  $\mathcal{E}(T_i|M_i)$  is at least as large as the observed one and we compare this P-value with the required significance level. In the case where  $N_{Ti} \ge 5-10$ , and  $P(T_i)$  and  $P(T_i|M_i)$  are not close to zero or one, then a normal approximation for the binomial distribution should be adequate, in which case  $\mathcal{E}(T_i | M_i) =$ 1.96 would represent the standard 95% confidence interval. Note that such a statistical association does not necessarily prove that there is a direct 'causal' interaction between mammals and vectors. Rather, it allows for a statistical inference to be made or a hypothesis to be formulated that may be validated subsequently (González-Salazar and Stephens, 2012).

We are interested in hypotheses about the transmissibility of the parasite by a specific vector-host interaction. We estimated epsilon,  $\varepsilon$ , values for a particular mammal species according to its degree of co-occurrence with a given triatomine species. With the values of  $\varepsilon(T_i|M_j)$  in hand for every possible triatomine-mammal pair we can compute and visualize a network by considering the nodes of the network to be the mammal and triatomine species and a link between a mammal,  $M_j$ , and a triatomine,  $T_i$ , to be associated with  $\varepsilon(T_i|M_j)$ . If all values of  $\varepsilon(T_i|M_j)$  are considered, then the network is fully connected. However, if we only draw those links that have a certain degree of statistical significance, then the network has a different topology, that now represents the principal inferred biotic interactions between mammals and triatominae.

# Testing hypotheses of ecological epidemiology of Chagas disease

All else being equal we posit that the higher the value of  $\varepsilon$  for a host the more epidemiologically important it is in ecological terms. The rational for this is that the greater the degree of spatial overlaps between the distributions, the greater the proportion of potentially infected host individuals due to the higher proportion of individuals that can have a biotic interaction with the vector. Here we do not consider the relative epidemiological importance in terms of human infection. Of course, epidemiological importance, both at the ecological and public health levels is highly complex and multi-factorial involving a host of factors, such as host competence, host/vector abundance, host/vector domiciliation, etc. However, spatial coincidence of vector and host is an absolutely necessary condition on which multiple other factors can and should be included. In the absence of comprehensive, systematic data on these multiple other factors however it is useful to build first-order models based only on occurrence data and use empirical data to test associated hypotheses and models.

To test hypotheses about the ecological epidemiology of Chagas disease, we compiled from the literature a list of confirmed mammal hosts. We searched for mammals species with at least one individual reported as being infected with *T. cruzi* (i.e. confirmed) in non-desert ecoregions of Mexico. Records were found in the Web of Knowledge of the Institute for Scientific Information (ISI – Thomson Scientific, Philadelphia, PA, USA), BibTri (bibtri. com.ar) databases and bibliographic collections of the Laboratorio de Biología de Parásitos, Facultad de Medicina, Universidad Nacional Autónoma de México (Appendix 1, Supporting Information).

We considered all records of confirmed mammal species in our analyses. In our results, we also mention the diagnostic methods by which infection with T. cruzi was determined for each mammal species. As the method used here assumes that the larger the geographical overlap the more likely it is that there is a biotic interaction between the species, it may be argued that the mammal species with the largest overlap are simply those with the greatest geographic range, i.e. that a ranking by  $\varepsilon$  is equivalent to a ranking by the distribution range. To check this we searched for statistical differences, using a t-test, between the relative ranking of both  $\varepsilon$  values and the distribution sizes of mammals, i.e. the number of grid cells with records of a given mammal species. Two sample groups were tested: T. dimidiata 2, a relatively widespread species (245 grid cells), and T. picturata, a species with a narrow-distribution (20 grid cells).

#### RESULTS

## Inferred interaction network of triatomines and mammals

Our data mining approach allowed us to identify the statistically significant ( $\varepsilon > 1.96$ ) potential vectorhost pair associations. From these 643 pairs, for each triatomine species we chose the 25% [top quartile (Q4)] of highest  $\varepsilon$  values. Thus, our network represents the most significant (Q4) positive cooccurrence associations between Triatominae and mammal species (Fig. 2). These potential relationships are the most likely (but not surely) to yield an important biotic interaction between a triatomine and a mammal, yielding the most statistically significant geographic overlaps between triatomines and mammals and therefore the highest potential for encounters given our 'all else being equal' assumption. For a given threshold on  $\varepsilon$  it is 'maximal' in that it encapsulates the idea of capturing those triatominemammal pairs that most satisfy the necessary condition of spatial overlap, but without any assumption of other potential conditions that would cause the interaction to either not be present at all-e.g., the triatomine does not feed on that mammal, or that the interaction does not lead to infection, such as if the mammal has very low competence. It is, of course, of great theoretical and practical interest to know what fraction of that 'maximal' network corresponds to real biotic interactions that also correspond to components of the transmission cycle.

Reviewing the network we note that 116 out of the 396 (29%) total wild mammal species considered could interact with at least one species of Triatominae (Table 1); while 86 of them are potentially associated to only one triatomine species. Ten mammal species probably interact with three or more vectors, among which Baiomys musculus and Liomys pictus seem the most important as they are potentially associated with five triatomine species (Fig. 2, Table 1). Once again, this network shows species association patterns that are based on the overlap of their geographic ranges, co-occurrence being a necessary condition for a biotic interaction. It does not, however, necessarily prove that there is a direct 'causal' interaction. It can, though, provide testable hypotheses for vector-host interactions.

To check the sensitivity of the network, and therefore our conclusions, to the model parameters and assumptions, such as the grid size and our threshold on  $\varepsilon$ , we considered how as a base measure the figure of 29% of true positives in Q4, seen in Fig. 3A, changed for three different grid sizes and three different  $\varepsilon$  thresholds. Checks were made with three grid sizes: the true positive percentages in Q4 were 28% (5 km), 25% (10 km) and 25% (50 km), respectively. We noted that the 29% of true positives in the top quartile does not change significantly ( $\chi^2 = 1.3$ , P = 0.52) as a function of grid size. On the other

available

at

Downloaded from https://www.cambri



Fig. 2. Inferred vector-host network of *Trypanosoma cruzi* from Triatominae (squares) and wild mammals (points) species in non-desert areas of Mexico. Circled mammals potentially interact with three or more triatominae species.

Table 1. Ranked list of potential mammal hosts for Trypanosoma cruzi in non-desert areas of Mexico

	R	Mammals	Е	Tri	R	Mammals	Е	Tri
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	1	Peromyscus yucatanicus	17.96	1	59	Rhogeessa tumida	9.41	1
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	2	Orthogeomys hispidus	16.87	1	60	Orthogeomys grandis	$9.36^{\mathrm{a}}$	2
4       Peppogeomys bulleri       16-63"       2       6       Lontra longiculis       9-24         6       Baiomys taylori       15-75"       2       64       Peromyscus pectoralis       9-04         7       Carollia perspicillat       15-75"       2       64       Peromyscus pectoralis       9-04         8       Carollia perspicillat       15-01       1       65       Heteromys desmarstans       9-01         9       Didelphis marsupialis       14-59       1       66       Didelphis virgimiana       8-91         9       Didelphis marsupialis       14-59       1       67       Cryptotis mexicana       8-74         10       Orgoodomys banderanus       13-96       70       Sigmodon alleni       8-65         11       Heteromys gauneri       13-27       1       72       Ictidonys mexicanas       8-53         15       Microtus guasiater       13-27       1       72       Molossus rufus       8-53         16       Reithrodontomys gracilis       13-18       74       71       Margama americana       8-46         10       Otaylomys phylotis       12-84	3	Peromyscus mexicanus	$16.65^{a}$	3	61	Macrotus waterhousii	9.3	1
$      5 \qquad 0 ligoryzomys fulcescens                                 $	4	Pappogeomys bulleri	$16.63^{\mathrm{a}}$	2	62	Lontra longicaudis	9.24	1
	5	Oligoryzomys fulvescens	16.33	1	63	Otospermophilus variegatus	$9.04^{\mathrm{a}}$	2
$ \begin{array}{c} 7 & Carollia perspicillata \\ 8 & Carollia soxelli \\ 14.79 & 166 \\ 14.79 & 166 \\ 14.79 & 166 \\ 14.79 & 166 \\ 14.79 & 166 \\ 14.79 & 166 \\ 14.71 & 168 \\ 14.71 \\ 11 \\ 12 \\ 12 \\ 12 \\ 12 \\ 12 \\ 12 \\ $	6	Baiomys taylori	$15.75^{\mathrm{a}}$	2	64	Peromyscus pectoralis	9.04	1
8       Carollia sovelli       14-79       1       66       Didelphis marsupialis       8-91         9       Didelphis marsupialis       14-79       1       68       Artibus lituratus       8-74         11       Philander opsum       14-08       1       69       Reithrodontomys fulvescens       8-71         12       Orgoodomys handeranus       13-96       1       70       Sigmodon alleni       8-65         13       Peromyscus melanurus       13-92       2       71       Myotis ingricans       8-50         14       Heteromys gameeria       13-12       7       7       Icitalonys mexicanus       8-53         15       Microtus quasiater       13-22       2       73       Molosus rufus       8-53         16       Reithrodontomys gracitis       13-14       1       75       Sigmodon mascotensis       8-50         18       Lepus callotis       13-07       3       76       Masama americana       8-45         20       Peromyscus furcus       12-84       1       77       Vampyrodes major       8-45         21       Dastyprota mexicana       12-15	7	Carollia perspicillata	15.01	1	65	Heteromys desmarestianus	9.01	1
9       Didelphis marsupialis       14-27       167       Cryptotis mexicana       8+24         11       Philander oposum       14-08       1       69       Reithrodontomys fulvescens       8+74         11       Philander oposum       14-08       1       69       Reithrodontomys fulvescens       8+74         12       Osgoodomys banderemus       13-95"       2       71       Myotis migricans       8+65         13       Peromyscus melanurus       13-95"       2       71       Myotis migricans       8+56         14       Heteromys gaumeri       13-22"       73       Molossus rufus       8+53         16       Reithrodontomys graciiis       13-14       1       75       Sigmodon mascotensis       8+50°         17       Orysomys cousei       13-14       1       75       Sigmodon mascotensis       8+50°         18       Lepus calloits       13-07"       3       76       Mascona mercicana       8+45         20       Peromyscus furcus       12-84       1       77       Vampyrodes major       8+45         21       Dasyprocta mexicana       12-47       18	8	Carollia sowelli	14.79	1	66	Didelphis virginiana	8.91	1
10     Peromyscus leucopus $14-27$ 1     68     Artibus lituratus $8-74^{\circ}$ 11     Polinader opsum $14+08$ 16     9     Reithrodontomys fulvescens $8+71$ 12     Orgoodomys handeranus $13-95^{\circ}$ 2     71     Myotis nigricans $8+6^{\circ}$ 13     Peromyscus melanurus $13-95^{\circ}$ 2     71     Myotis nigricans $8+5^{\circ}$ 14     Heteromys gammeri $13-27$ 172     Ictidomys mexicanus $8+5^{\circ}$ 15     Microtus quasiater $13-27^{\circ}$ 72     Ictidomys mexicanus $8+53$ 16     Reithrodontomys graciilis $13+14$ 1     75     Sigmodon mascenenicana $8+46$ 19     Otatylomys phyllotis $12+84$ 1     77     Vampyrodes major $8+45$ 20     Peromyscus furcus $12+84$ 1     77     Vampyrodes major $8+36$ 21     Dasyprocta mexicana $12-28$ 18     Oryconys melanotis $8+19^{\circ}$ 24     Cryptotis mayensis $12+19$ 1     82     Glassomy satutis $7+44$ <	9	Didelphis marsupialis	14.59	1	67	Cryptotis mexicana	8.82	1
11Philander oposum14-08169Reithrodontomys fulcescens8-7112Osgoodomys banderanus13-95"271Myotis nigricans8-65"13Peromysus melanurus13-27172Ictidomys mexicanus8-59"14Heteromys gaumeri13-27172Ictidomys mexicanus8-59"15Microtus quasitater13-1227Molossus rufus8-5316Reithrodontomys gracilis13-18174Platyrrhinus fielleri8-5317Oryzomys cousci13-14175Sigmodon mascotensis8-50"18Lepus callotis13-07"376Mazama americana8-4619Ototylomys phyllotis12-84177Vampyrodes major8-4520Peromyscus furvus12-1518Molossus sinaloae8-3421Dasybrota mexicana12-15183Leptomyteris macrotis8-19"22Sigmodon hispidus12-15183Leptonyteris nicalis7-8423Sciurus deppei12-15184Liomys pitus7-6524Cryptotis mayensis12184Liomys pitus7-6425Otonytomys hatti1-193186Leptonyteris nicalis7-74928Sciurus colliaei11-93186Leptomyteris mealophrys7-1830Reithrodontomys hisratus11-63"589<	10	Peromyscus leucopus	14.27	1	68	Artibeus lituratus	8·74ª	2
12Osgoodomys banderanus $13^{\circ}9^{\circ}$ 70Sigmodon allem $8^{\circ}5^{\circ}$ 13Peromyscus melanurus $13^{\circ}9^{\circ}$ 271Myoits nigricans $8^{\circ}6^{\circ}$ 14Heteromys gaumeri $13^{\circ}27$ 172Ictidomys mexicanus $8^{\circ}9^{\circ}$ 15Microtus guasiater $13^{\circ}27$ 172Ictidomys mexicanus $8^{\circ}53$ 16Reithrodontomys gracilis $13^{\circ}14^{\circ}$ 175Sigmodon mascotensis $8^{\circ}53$ 17Oryzomys couesi $13^{\circ}14^{\circ}$ 175Sigmodon mascotensis $8^{\circ}53^{\circ}$ 19Ototylomys phyllotis $12^{\circ}84$ 177Vampyrodes major $8^{\circ}45^{\circ}$ 20Peromyscus furvus $12^{\circ}83$ 178Hodomys alleni $8^{\circ}43^{\circ}$ 21Dasyprocta mexicana $12^{\circ}28$ 181Oryzomys malaoe $8^{\circ}34$ 22Sigmodon hispidus $12^{\circ}47$ 180Molosus analoae $8^{\circ}34$ 23Sciurus deppei $12^{\circ}28$ 181Oryzomys melanotis $8^{\circ}19^{\circ}4$ 24Cryptotis magensis $12^{\circ}19$ 182Glosophaga commissarisi $7^{\circ}8$ 25Otonyctomys hatti $12^{\circ}15$ 183Leptonycteris verbaluenae $7^{\circ}41$ 28Sciurus colliaei $11^{\circ}93$ 186Leptonycteris verbaluenae $7^{\circ}41$ 29Molosus aztecus $11^{\circ}66^{\circ}$ 90Peromyscus melanophrys $7^{\circ}18$ 31Balant	11	Philander oposum	14.08	1	69	Reithrodontomys fulvescens	8.71	1
13Peromyscus melanurus $13^{+}95^{\circ}$ 271Myots mexicanus $8^{+}59^{\circ}$ 14Heteromys gauasiater $13\cdot27$ 172Ictidomys mexicanus $8^{+}59^{\circ}$ 15Microtus guasiater $13\cdot22^{\circ}$ 273Molosus rufus $8^{+}53$ 16Reithrodontomys gracilis $13\cdot18$ 174Platyrrhinus helleri $8^{+}53$ 17Oryzomys couesi $13\cdot14$ 175Sigmodon mascotensis $8^{+}50^{\circ}$ 18Lepus callotis $13\cdot07^{\circ}$ 76Mazama americana $8^{+}45$ 20Peromysus furvus $12\cdot83$ 78Hodomys alleri $8^{+}45$ 21Dasyprocta mexicana $12\cdot5$ 79Peropteryx macrotis $8^{+}36$ 22Sigmodon hispidus $12\cdot47$ 180Molosus sinaloae $8^{+}34$ 23Sciurus deppei $12\cdot28$ 81Oryzomys melanotis $8^{+}9^{\circ}$ 24Cryptotis magensis $12\cdot19$ 182Glossophaga commissarisi $7^{+}98$ 25Otonyctomys hatti $12\cdot15$ 183Leptonycteris incalis $7^{+}49$ 26Reithrodontomys mexicanus $11\cdot63^{\circ}$ 58Hetonys prictus $7^{-}23$ 30Reithrodontomys hirsutus $11\cdot76^{\circ}$ 88Basariscus sumichrasti $7^{-}23$ 31Balantiopteryx plicata $11\cdot60^{\circ}$ 91Uroderma maginostrum $7^{-}16$ 32Oryzomys alfaroi $11\cdot6^{\circ}$ 92Sciurus sumelanophrys $7^{-}18$ <td>12</td> <td>Osgoodomys banderanus</td> <td>13.96</td> <td>1</td> <td>70</td> <td>Sigmodon alleni</td> <td>8·65ª</td> <td>2</td>	12	Osgoodomys banderanus	13.96	1	70	Sigmodon alleni	8·65ª	2
14       Heteromys gaumeri $15^{27}$ 1       72       Ictidonys mexicanus $8:59^{\circ}$ 15       Microtus quasiater $13:22^{\circ}$ 2       73       Molossus nefus $8:53$ 16       Reithrodontomys gracilis $13:18$ 1       74       Platyrrhinus helleri $8:53$ 17       Oryzomys couesi $13:14$ 1       75       Sigmodon mascotensis $8:50^{\circ}$ 18       Lepus callotis $13:07^{\circ}$ 3       76       Mascama americana $8:44^{\circ}$ 20       Peromyscus furrus $12:83$ 1       78       Hodomys alleni $8:43^{\circ}$ 21       Dasyprocta mexicana $12:5$ 1       79       Peropteryx macrotis $8:36^{\circ}$ 22       Sigmodon hispidus $12:47$ 1       80       Molosus sinaloae $8:34^{\circ}$ 23       Sciurus delpiei $12:15$ 18       10 oryzomys melanotis $7:45^{\circ}$ 24       Cryptotis mayensis $12:15$ 18       12 iomys pictus $7:45^{\circ}$ 25       Otonyctomys hatiti </td <td>13</td> <td>Peromyscus melanurus</td> <td>13.95*</td> <td>2</td> <td>71</td> <td>Myotis nigricans</td> <td>8.6</td> <td>1</td>	13	Peromyscus melanurus	13.95*	2	71	Myotis nigricans	8.6	1
15Microtus quasiater $15^{+}22^{+}$ 273Molossus rifus $8^{+}53$ 16Reithrodontomys gracilis $13^{+}14$ 174Platyrrhimus helleri $8^{+}53$ 17Oryzomys couesi $13^{+}14$ 175Sigmodon mascotensis $8^{+}50^{+}$ 18Lepus callolis $13^{+}07^{a}$ 376Mazama americana $8^{+}46$ 19Ototylomys phyllotis $12^{+}83$ 178Hodomys alleni $8^{+}45^{+}$ 21Dasyprocta mexicana $12^{+}5$ 179Peropteryx macrotis $8^{+}36^{+}$ 22Sigmodon hispidus $12^{+}7$ 180Molossus sinaloae $8^{+}34^{+}$ 23Sciurus depei $12^{-}28$ 181Oryzomys nelanotis $8^{+}19^{+}$ 24Cryptotis mayensis $12^{-}17$ 183Leptonycteris nivalis $7^{+}65^{-}$ 25Otonyctomys hatti $12^{+}17$ 183Leptonycteris nivalis $7^{+}65^{-}$ 25Otonyctomys hexicanus $12^{-}18^{-}18^{-}3^{-}18^{-}8^{-}8^{-}16^{-}18^{-}11^{-}11^{-}18^{-}18^{-}18^{-}18^{-}11^{-}118^{-}11^{-}18^{-}18^{-}18^{-}18^{-}118^{-}118^{-}118^{-}11^{-}11^{-}18^{-}18^{-}18^{-}118^{-}111^{-}11^{-}11^{-}11^{-}111^{-}111^{-}111^{-}111^{-}111^{-}111^{-}111^{-}111^{-}111^{-}111^{-}111^{-}111^{-}111^{-}111^{-}111^{-}1111^{-}1111^{-}111^{-}1111^{-}1111111$	14	Heteromys gaumeri	13.27	1	72	Ictidomys mexicanus	8·59"	2
16     Reithrodontomys gracitis     13-18     1     74     Platyrimus helleri     8-53       17     Oryzomys cousei     13-14     1     75     Sigmodon mascolensis     8-50       18     Lepus callotis     13-07 <sup>a</sup> 3     76     Mazama americana     8-45       19     Ototylomys phyllotis     12-84     1     77     Vampyrodes major     8+45       20     Peromyscus furcus     12-83     1     78     Hodomys alleni     8-43       21     Dasybrocta mexicana     12-5     1     79     Peropteryy macrotis     8-36       22     Sigmodon hispidus     12-47     1     80     Molossus sinaloae     8-34       23     Sciturus deppei     12-28     1     81     Oryzomys nelanotis     8-19 <sup>a</sup> 24     Cryptotis mayensis     12-19     1     82     Glossophaga commissarisi     7-98       25     Otoryctomys hatti     12-17     1     85     Cryptotis goldmani     7-49       28     Sciurus colliaei     11-97     1     85     Cryptotis goldmani     7-49       29     Molosu	15	Microtus quasiater	13.22*	2	73	Molossus rufus	8.53	1
17     Oryzonys couest     15-14     1     75     Sigmodon mascolensis     8-50"       18     Lepus callotis     13-07"     3     76     Mazama americana     8-46       19     Ototylomys phyllotis     12-83     1     78     Hodomys alleni     8-43"       20     Peromyscus furvus     12-83     1     79     Peropteryx macrotis     8-36       21     Dasyprocta mexicana     12-47     1     80     Molossus sinaloae     8-34       22     Sigmodon hispidus     12-47     1     80     Molossus sinaloae     8-36       23     Sciurus deppei     12-28     1     81     Oryzomys melanotis     7-98       24     Cryptotis mayensis     12-19     1     82     Clonsycteris viculis     7-84       25     Otonyctomys hatti     12-15     1     83     Leptomysteris viculis     7-65       27     Sylvilagus gabbi     11-97     1     85     Cryptotis goldmani     7-49       28     Sciurus colliaei     11-93     86     Leptomytetris viculis     7-33       30     Reithrodontomys hirsutus	16	Reithrodontomys gracilis	13.18	1	74	Platyrrhinus helleri	8.53	1
18     Lepus cattotis     13-07     3     76     Mazama americana     8-46       90     Otzylomys phyllotis     12-84     1     77     Vampyrodes major     8-45       20     Peromyscus furvus     12-83     1     78     Hodomys alleni     8-43°       21     Dasyprocta mexicana     12-5     1     79     Peropteryx macrotis     8-36       22     Sigmodon hispidus     12-47     1     80     Molossus sinaloae     8-34       23     Sciurus deppei     12-28     1     81     Oryzomys melanotis     8-19°       24     Cryptotis mayensis     12-19     1     82     Glossophaga commissarisi     7-98       25     Otonyctomys hatti     12-15     1     83     Leptonycteris invalis     7-65       27     Sylvilagus gabbi     11-97     1     85     Cryptotis goldmani     7-49       28     Sciurus collicei     11-93     1     86     Leptonycteris yerbabuene     7-41       29     Molossus aztecus     11-63°     5     89     Leptus flavigularis     7-22       30     Reithrodon	1/	Oryzomys couesi	13.14		75	Sigmodon mascotensis	8.50-	2
19Otolytomys phytotis12:84177Varburgstock amound for the state of the sta	18	Lepus callotis	13.07-	3	/6	Mazama americana	8.46	1
20Peromyscus jurvus12*83178Hodomys allem8*43'1Dasyprocta mexicana12*5179Peropteryx macrotis8:3622Sigmodon hispidus12*47180Molossus sinaloae8:3423Sciurus deppei12:28181Oryzomys melanotis8:19"24Cryptotis mayensis12:19182Glossophaga commissarisi7:9825Otonyctomys hatti12:15183Leptonycteris nivalis7:8426Reithrodontomys mexicanus12184Liomys pictus7:6527Sylvilagus gabi11:97185Cryptotis goldmani7:4928Sciurus colliaei11:93186Leptonycteris yerbabuenae7:4129Molossus aztecus11:86187Nyetomys sumichrasti7:3330Reithrodontomys hirsutus11:66390Peronyscus melanophrys7:1831Baiomys musculus11:66391Uroderma agenirostrum7:1634Dasyprocta punctata11:56192Sylvilagus cunicularius7:09"35Carollia subrufa11:48"293Chrioderma salvini7:04"36Artibeus hirsutus11:31"294Idionyctris phyllotis6:8937Sciurus aureogaster11:04195Artibeus jamaicensis6:6638Liomys irroratus10:72"2 </td <td>19</td> <td>Ototylomys phyllotis</td> <td>12.84</td> <td>1</td> <td>//</td> <td>V ampyrodes major</td> <td>8.45</td> <td>1</td>	19	Ototylomys phyllotis	12.84	1	//	V ampyrodes major	8.45	1
21Dasyprota mexicana12:5179Peroper y macroits8:3022Sigmodon hispidus12:47180Molosus sinaloae8:3423Sciurus deppei12:28181Oryzomys melanotis8:19 <sup>a</sup> 24Cryptotis mayensis12:15182Glossophaga commissarisi7:9825Otonyctomys hatti12:15184Liomys pictus7:6527Sylvilagus gabbi11:97185Cryptotis goldmani7:4928Sciurus colliaei11:93186Leptonycteris yerbabuenae7:4129Molossus aztecus11:86187Nyetomys sumichrasti7:3330Reithrodontomys hirsutus11:76188Basariscus sumichrasti7:2331Baiomys musculus11:6190Peromyscus melanophrys7:1833Balantiopteryx plicata11:60 <sup>a</sup> 391Uroderma magnirostrum7:1634Dasyprota punctata11:31 <sup>a</sup> 294Idionycteris phyllotis6:8937Sciurus aureogaster11:04195Artibeus jamaicensis6:6740Peromyscus spicilegus10:75197Dermanura tolteca6:7741Cunculus paca10:581100Cratagenys fumosus6:5443Myotis carteri10:411101Mephitis macroura6:4345Sciurus gaas oricina10:581 <td< td=""><td>20</td><td>Peromyscus furvus</td><td>12.83</td><td>1</td><td>/8</td><td>Hodomys alleni</td><td>8.43</td><td>2</td></td<>	20	Peromyscus furvus	12.83	1	/8	Hodomys alleni	8.43	2
22Sigmodon inspiratis $12 \cdot 47$ 180Motossis sinatoae8 \cdot 3 \cdot 423Sciurus deppei $12 \cdot 28$ 181Oryzonys melanotis $8 \cdot 19^{a}$ 24Cryptotis mayensis $12 \cdot 19$ 182Glossophaga commissarisi $7 \cdot 98$ 25Otonyctomys hatti $12 \cdot 15$ 183Leptonycteris nivalis $7 \cdot 48$ 26Reithrodontomys mexicanus $12$ 184Liomys pictus $7 \cdot 65$ 27Sylvilagus gabbi $11 \cdot 97$ 185Cryptotis goldmani $7 \cdot 49$ 28Sciurus colliaei $11 \cdot 97$ 186Leptonycteris yerbabuenae $7 \cdot 41$ 29Molossus aztecus $11 \cdot 86$ 187Nyctomys sumichrasti $7 \cdot 33$ 30Reithrodontomys hirsutus $11 \cdot 67$ 188Bassariscus sumichrasti $7 \cdot 23$ 31Baiomys musculus $11 \cdot 63^{a}$ 589Lepus flavigularis $7 \cdot 23$ 32Oryzomys alfaroi $11 \cdot 60^{a}$ 391Uroderma magnirostrum $7 \cdot 16$ 33Balantiopteryx plicata $11 \cdot 60^{a}$ 391Uroderma salvini $7 \cdot 04^{a}$ 34Dasybrocta punctata $11 \cdot 64^{a}$ 293Chiroderma salvini $7 \cdot 04^{a}$ 35Carollia subrufa $11 \cdot 48^{a}$ 293Chiroderma salvini $7 \cdot 04^{a}$ 36Artibeus hirsutus $11 \cdot 31^{a}$ 94Idionycteris philipsii $6 \cdot 84$ 37Sciurus auceagaster	21	Dasyprocta mexicana	12.5	1	/9	Peropteryx macrotis	8.36	1
23Sciurus depet12-28181Oryzomys metanotis81-924Cryptotis mysensis12-19182Glossophaga commissarisi7-9825Otonyctomys hatti12-15183Leptonycteris nivalis7-8426Reithrodontomys mexicanus12184Liomys pictus7-6527Sylvilagus gabbi11-97185Cryptotis goldmani7-4928Sciurus colliaei11-93186Leptonycteris yerbabuenae7-4129Molossus aztecus11-63589Lepus flavigularis7-2330Reithrodontomys hirsutus11-63589Lepus flavigularis7-2331Balantiopteryx plicata11-6690Peromyscus melanophrys7-1833Balantiopteryx plicata11-669190Peromyscus melanophrys7-1834Dasyprocta punctata11-6692Sylvilagus cunicularius7-0935Carollia subrufa11-48 <sup>a</sup> 293Chiroderma magnirostrum7-1634Dasyprocta punctata11-67195Artibeus jamaicensis6-8637Sciurus aureogaster11-04195Artibeus jamaicensis6-8638Liomys irroratus10-75197Dermanura tolteca6-7740Peromyscus spicilegus10-72 <sup>a</sup> 298Sciurus ouclatus6-6141Quiculus baca10-731 <td>22</td> <td>Sigmodon hispidus</td> <td>12.47</td> <td>1</td> <td>80</td> <td>Molossus sinaloae</td> <td>8.34</td> <td>1</td>	22	Sigmodon hispidus	12.47	1	80	Molossus sinaloae	8.34	1
24Cryptoits mayensis12-19182Glossophaga commissarisi7-9825Otonyctomys hatti12-15183Leptonycteris nivalis7-8426Reithrodontomys mexicanus12184Liomys pictus7-6527Sylvilagus gabbi11-97185Cryptotis goldmani7-4928Sciurus colliaei11-93186Leptonycteris yerbabuenae7-4129Molossus aztecus11-86187Nyctomys sumichrasti7-3330Reithrodontomys hirsutus11-66190Peromyscus melanophrys7-1831Baiomys musculus11-66190Peromyscus melanophrys7-1832Oryzomys alfaroi11-66190Peromyscus melanophrys7-1634Dasyprocta punctata11-67191Uroderma anguirostrum7-1634Dasyprocta punctata11-48*293Chiroderma aslvini7-04*36Artibus hirsutus11-31*294Idionycteris phyllotis6-8937Sciurus aureogaster11-04195Artibus jamaicensis6-8638Liomys irroratus10-72*298Sciurus ouclatus6-6741Cuniculus paca10-72*298Sciurus ouclatus6-6442Glossophaga soricina10-281102Myotis velifer6-37*45Sciurus yucatanensis10-2	23	Sciurus deppei	12.28	1	81	Oryzomys melanotis	8.19-	3
25Otonyctomys haiti12*15183Leptonycteris invalis7*8426Reithrodontomys mexicanus12184Liomys pictus7*6527Sylvilagus gabbi11*97185Cryptotis goldmani7*4928Sciurus colliaei11*97186Leptonycteris yerbabuenae7*4129Molossus aztecus11*66187Nyctomys sumichrasti7*3330Reithrodontomys hirsutus11*66188Bassariscus sumichrasti7*2331Baiomys musculus11*66*/a589Lepus flavigularis7*232Oryzomys alfaroi11*6190Peromyscus melanophrys7*1634Dasyprocta punctata11*66*/a91Uroderma magnirostrum7*0935Carollia subrufa11*48*/a293Chiroderma salvini7*04*36Artibeus hirsutus11*31*/a294Idionycteris phyllotis6*8937Sciurus aureogaster11*0495Artibeus jamaicensis6*8638Liomys irroratus10*75*/a97Dermanura tolteca6*7740Peromyscus spicilegus10*72*/a298Sciurus culatus6*6141Cuniculus paca10*59199Glossophaga leachii6*6142Glossophaga soricina10*581100Cratogeomys fumosus6*5444Myotis carteri10*11101Mephitis	24	Cryptotis mayensis	12.19	1	82	Glossophaga commissarisi	7.98	1
20Retitivationity mexicantits12184Lomys pictus7-0527Sylvilagus gabbi11-97185Cryptotis goldmani7-4928Sciurus colliaei11-93186Leptonycteris yerbabuenae7-4129Molossus aztecus11-66187Nyctomys sunichrasti7-3330Reithrodontomys hirsutus11-66189Leptus flavigularis7-231Baiomys musculus11-66190Peromyscus melanophrys7-1833Balantiopteryx plicata11-66190Peromyscus melanophrys7-1834Dasyprocta punctata11-56192Sylvilagus cunicularius7-0935Carollia subrufa11-48 <sup>a</sup> 293Chiroderma salvini7-04 <sup>a</sup> 36Artibeus hirsutus11-31 <sup>a</sup> 294Idionycteris phyllotis6-8937Sciurus aureogaster11-04195Artibeus jamaicensis6-8638Liomys irroratus10-75197Dermanura tolteca6-7740Peromyscus spicilegus10-72 <sup>a</sup> 298Sciurus oculatus6-6141Cuniculus paca10-59199Glossophaga leachii6-6142Glossophaga soricina10-581100Crategeomys funosus6-5443Myotis carteri10-121103Peromyscus spiculus6-3644Megasorex gigas10-2	25	Otonyctomys natti	12.15	1	83	Leptonycteris nivalis	7.84	1
27Sylvilagus gaon $11.97$ $1$ 85Cryptoits golaman $7.49$ 28Sciurus colliaei $11.93$ $1$ 86Leptonycteris yerbabuenae $7.41$ 29Molossus astecus $11.86$ $1$ 87Nyctomys sumichrasti $7.33$ 30Reithrodontomys hirsutus $11.66$ $1$ 88Bassariscus sumichrasti $7.23$ 31Baiomys musculus $11.66$ $1$ 90Peromyscus melanophrys $7.18$ 32Oryzomys alfaroi $11.66$ $1$ 90Peromyscus melanophrys $7.18$ 33Balantiopteryx plicata $11.66$ $1$ $92$ Sylvilagus cunicularius $7.09$ 35Carollia subrufa $11.48^a$ $2$ $93$ Chiroderma salvini $7.04^a$ 36Artibeus hirsutus $11.31^a$ $2$ $94$ Idionycteris phyllotis $6.86$ 38Liomys irroratus $10.88^a$ $3$ $96$ Cryptotis phillipsii $6.84$ 39Eptesicus furinalis $10.75$ $1$ $97$ Dermanura tolteca $6.77$ 40Peromyscus spicilegus $10.72^a$ $2$ $98$ Sciurus oculatus $6.61$ 41Cuniculus paca $10.58$ $1$ $100$ Cratageomys fumosus $6.54$ 43Myotis carteri $10.41$ $1$ $101$ Mephitis macroura $6.43$ 44Megasorex gigas $10.2^a$ $103$ Peromyscus perfulvus $6.36$ 45Sciurus yucatanensis $10.2^a$ $103$	20	Reithroaontomys mexicanus	12	1	84	Liomys pictus	7.05	5
28Scurrus contact11+85180Leptonycleris yeroabilenae7+4129Molossus aztecus11+86187Nyctomys sunichrasti7-3330Reithrodontomys hirsutus11+66188Bassariscus sumichrasti7-2331Baiomys musculus11+66190Peromyscus melanophrys7-1833Balantiopteryx plicata11+66190Peromyscus melanophrys7-1634Dasyprocta punctata11+56192Sylvilagus cunicularius7-0935Carollia subrufa11+48 <sup>a</sup> 293Chiroderma salvini7-04 <sup>a</sup> 36Artibeus hirsutus11-31 <sup>a</sup> 294Idionycteris phyllotis6-8637Sciurus aureogaster11+0495Artibeus jamaicensis6-8638Liomys irroratus10-75197Dermanura tolteca6-7740Peromyscus spicilegus10-72 <sup>a</sup> 298Sciurus oculatus6-6741Cuniculus paca10-59199Glossophaga leachii6-6142Glossophaga soricina10-581100Cratogeomys fumosus6-5443Myotis carteri10-411101Mephitis macroura6-4344Megasorex gigas10-21103Peromyscus perfulvus6-3645Sciurus yucatanensis10-111105Lasiurus blossevillii6-1748Dermanura azteca10-07 <t< td=""><td>27</td><td>Sylvilagus gabbi</td><td>11.02</td><td>1</td><td>85</td><td>Cryptotis golamani</td><td>7.49</td><td>1</td></t<>	27	Sylvilagus gabbi	11.02	1	85	Cryptotis golamani	7.49	1
29Motoskik dziecius11*80187Nyciomys simichrasti7:3330Reithrodontomys hirsutus11*66188Bassariscus sumichrasti7:2331Baiomys musculus11*66190Peromyscus melanophrys7:1832Oryzomys alfaroi11*6190Peromyscus melanophrys7:1833Balantiopteryx plicata11*66190Peromyscus melanophrys7:1834Dasyprocta punctata11*56192Sylvilagus cunicularius7:0935Carollia subrufa11*31*a294Idionycteris phyllotis6:8937Sciurus aureogaster11:04195Artibeus jamaicensis6:8638Liomys irroratus10*88*a396Cryptotis phillipsii6:8439Eptesicus furinalis10*75197Dermanura tolteca6:7740Peromyscus spicilegus10*72*a298Sciurus oculatus6:6142Glosophaga soricina10*581100Cratogeomys fumosus6:5443Myotis carteri10*411101Mephitis macroura6:4344Megasorex gigas10:221103Peromyscus perilubus6:37*a45Sciurus yucatanensis10:21103Peromyscus perilubus6:3646Marmosa mexicana10:12*a2104Sylvilagus floridanus6:2947Sphiggurus mexic	20	Sciurus contaet	11.95	1	80 87	Nustania surial verti	7.41	1
30Retundationality in tradits11.76136Dassaristis summeristi7.2531Baiomys musculus11.63°589Lepus flavigularis7.232Oryzomys alfaroi11.6190Peromyscus melanophrys7.1833Balantiopteryx plicata11.60°391Uroderma magnirostrum7.1634Dasyprocta punctata11.56192Sylvilagus cunicularius7.0935Carollia subrufa11.48°293Chiroderma salvini7.04°36Artibeus hirsutus11.31°294Idionycteris phyllotis6.8937Sciurus aureogaster11.04195Artibeus jamaicensis6.8638Liomys irroratus10.75°197Dermanura tolteca6.7740Peromyscus spicilegus10.72°298Sciurus ouclatus6.6641Cuniculus paca10.581100Cratogeomys fumosus6.5443Myotis carteri10.411101Mephitis macroura6.4344Megasorex gigas10.21103Peromyscus berfulvus6.3645Sciurus yucatanensis10.21103Peromyscus leifer6.3646Marmosa mexicana10.12°2104Sylvilagus floridanus6.2947Sphiggurus mexicanus10.111105Lasiurus blossevillii6.1748Dermanura azteca10.07	29	Deithre dont om this history	11.76	1	07	R goo grigorus sumichrasti B goo grigorus sumichrasti	7.33	1
1111103133512 pay flavoi1232Oryzonys alfaroi11.6190Peromyscus melanophrys7.1833Balantiopteryx plicata11.60°391Uroderma magnirostrum7.1634Dasyprocta punctata11.56192Sylvilagus cunicularius7.0935Carollia subrufa11.48°293Chiroderma salvini7.04°36Artibeus hirsutus11.31°294Idionycteris phyllotis6.8937Sciurus aureogaster11.04195Artibeus jamaicensis6.8638Liomys irroratus10.75197Dermanura tolteca6.7740Peromyscus spicilegus10.72°298Sciurus oculatus6.6741Cuniculus paca10.59199Glossophaga leachii6.6142Glossophaga soricina10.581100Cratgeomys fumosus6.5443Myotis carteri10.411101Mephitis macroura6.4344Megasorex gigas10.291102Myotis velifer6.37°45Sciurus yucatanensis10.111105Lasiurus blossevilli6.1748Dermanura azteca10.071106Sturnira hondurensis6.0449Alouatta palliata9.951107Peromyscus difficilis650Peromyscus levipes9.90°2108Deesmodus	30	Reinvoaoniomys nirsulus Baiomus musculus	11.62 <sup>a</sup>	5	00 80	Labus flavioularis	7.23	1
32Oryzoncy algor11190119011101133Balantiopteryx plicata11.60°391Uroderma magnirostrum7.1634Dasyprocta punctata11.56192Sylvilagus cunicularius7.0935Carollia subrufa11.48°293Chiroderma salvini7.04°36Artibeus hirsutus11.31°294Idionycteris phyllotis6.8937Sciurus aureogaster11.04195Artibeus jamaicensis6.8638Liomys irroratus10.88°396Cryptotis phillipsii6.8439Eptesicus furinalis10.72°298Sciurus oculatus6.6740Peromyscus spicilegus10.72°298Sciurus oculatus6.6741Cuniculus paca10.59199Glossophaga leachii6.6142Glossophaga soricina10.581100Cratogeomys fumosus6.5443Myotis carteri10.411101Mephitis macroura6.4344Megasorex gigas10.291102Myotis velifer6.3645Sciurus yucatanensis10.21103Peromyscus perfulvus6.3646Marmosa mexicanus10.111105Lasiurus blossevillii6.1747Sphiggurus mexicanus10.111105Lasiurus blossevillii6.1748Dermanura az	31	Orusomus alfaroi	11.6	5 1	00	Deromascus melanophras	7.18	1
33Data11 003910134Dasyprocta punctata11 06192Sylvilagus cunicularius70935Carollia subrufa11 06192Sylvilagus cunicularius70936Artibeus hirsutus11 0193Chiroderma salvini70436Artibeus hirsutus11 0495Artibeus jamaicensis60837Sciurus aureogaster11 0495Artibeus jamaicensis60838Liomys irroratus10 088°396Cryptotis phillipsii60839Eptesicus furinalis10 75197Dermanura tolteca67740Peromyscus spicilegus10 72°298Sciurus oculatus666741Cuniculus paca10 581100Cratogeomys fumosus65443Myotis carteri10 0411101Mephitis macroura64344Megasorex gigas10 21102Myotis velifer63645Sciurus yucatanensis10 21103Peromyscus perfulvus63646Marmosa mexicana10 12°2104Sylvilagus floridanus62947Sphiggurus mexicana10 111105Lasiurus blossevillii61748Dermanura azteca10 071106Sturnira hondurensis60250Peromyscus levipes9.90°2108Desmodus rotundus60251Notocitellus an	32	Balantiobteryy blicata	$11.60^{a}$	1	90	I eromyscus meiunophrys Uroderma magnirostrum	7.16	1
5.1Dasprot approximation11.36192Desprot approximation10035Carollia subrufa $11.48^{a}$ 293Chiroderma salvini7.04^{a}36Artibeus hirsutus $11.31^{a}$ 294Idionycteris phyllotis6.8937Sciurus aureogaster $11.04$ 195Artibeus jamaicensis6.8638Liomys irroratus $10.88^{a}$ 396Cryptotis phillipsii6.8439Eptesicus furinalis $10.75^{a}$ 197Dermanura tolteca6.7740Peromyscus spicilegus $10.72^{a}$ 298Sciurus oculatus6.6141Cuniculus paca $10.58$ 1100Cratogeomys fumosus6.5443Myotis carteri $10.41$ 1101Mephitis macroura6.4344Megasorex gigas $10.29$ 1 $102$ Myotis velifer6.3645Sciurus yucatanensis $10.2$ 1 $103$ Peromyscus perfulvus6.3646Marmosa mexicana $10.12^{a}$ 2 $104$ Sylvilagus floridanus6.2947Sphiggurus mexicanas $10.11$ 1 $105$ Lasiurus blossevillii $6.17$ 48Dermanura azteca $10.07$ 1 $106$ Sturnira hondurensis $6.02$ 50Peromyscus levipes $9.90^{a}$ 2 $108$ Desmodus rotundus $6.02$ 51Notocitellus annulatus $9.87^{a}$ 3 $109$ Peromyscus difficilis <td>34</td> <td>Dasvbrocta bunctata</td> <td>11.56</td> <td>1</td> <td>92</td> <td>Sylvilagus cunicularius</td> <td>7.09</td> <td>1</td>	34	Dasvbrocta bunctata	11.56	1	92	Sylvilagus cunicularius	7.09	1
5.5Curronic should11 10294Horoderins duction10 1036Artibeus hirsutus $11 \cdot 31^a$ 294Idionycteris phyllotis6.8937Sciurus aureogaster $11 \cdot 04$ 195Artibeus jamaicensis6.8638Liomys irroratus $10 \cdot 88^a$ 396Cryptotis phyllotis6.8439Eptesicus furinalis $10 \cdot 75$ 197Dermanura tolteca6.7740Peromyscus spicilegus $10 \cdot 72^a$ 298Sciurus oculatus6.6741Cuniculus paca $10 \cdot 59$ 199Glossophaga leachii6.6142Glossophaga soricina $10 \cdot 58$ 1100Cratogeomys fumosus6.5443Myotis carteri $10 \cdot 41$ 1101Mephitis macroura6.4344Megasorex gigas $10 \cdot 2$ 1102Myotis velifer6.3645Sciurus yucatanensis $10 \cdot 2$ 1103Peromyscus perfulvus6.6646Marmosa mexicana $10 \cdot 12^a$ 2104Sylvilagus floridanus6.2947Sphiggurus mexicanus $10 \cdot 11$ 1105Lasiurus blossevillii6.1748Dermanura azteca $10 \cdot 07$ 1106Sturnira hondurensis6.0449Alouatta palliata $9 \cdot 95$ 1 $107$ Peromyscus difficilis650Peromyscus levipes $9 \cdot 90^a$ 2108Desmodus rotundus6.02 <td< td=""><td>35</td><td>Carollia subrufa</td><td>11.30 <math>11.48^{a}</math></td><td>2</td><td>93</td><td>Chiroderma salvini</td><td><math>7.04^{a}</math></td><td>2</td></td<>	35	Carollia subrufa	11.30 $11.48^{a}$	2	93	Chiroderma salvini	$7.04^{a}$	2
50Interest mixtures11 01291Interest mixtures6.8737Sciurus aureogaster11 04195Artibeus jamaicensis6.8638Liomys irroratus10.88°396Cryptotis phillipsii6.8439Eptesicus furinalis10.75197Dermanura tolteca6.7740Peromyscus spicilegus10.72°298Sciurus oculatus6.6741Cuniculus paca10.59199Glossophaga leachii6.6142Glossophaga soricina10.581100Cratogeomys fumosus6.5443Myotis carteri10.411101Mephitis macroura6.4344Megasorex gigas10.21102Myotis velifer6.3645Sciurus yucatanensis10.21103Peromyscus perfulvus6.3646Marmosa mexicana10.11°1105Lasiurus blossevillii6.1748Dermanura azteca10.071106Sturnira hondurensis6.0449Alouatta palliata9.951107Peromyscus hylocetes6.0350Peromyscus levipes9.90°2108Desmodus rotundus6.0251Notocitellus annulatus9.84°2110Tadarida brasiliensis5.8653Spilogale gracilis9.84°2111Tlacuatzin canescens5.8254Peromyscus maniculatus9.74°2<	36	Artibeus hirsutus	11.10 $11.31^{a}$	2	94	Idionycteris phyllotis	6.89	1
$38$ Liomys irroratus $10.88^{a}$ $1$ $35$ Introduct of the phillipsii $6.84$ $39$ Eptesicus furinalis $10.75$ $1$ $97$ Dermanura tolteca $6.77$ $40$ Peromyscus spicilegus $10.72^{a}$ $2$ $98$ Sciurus oculatus $6.67$ $41$ Cuniculus paca $10.59$ $1$ $99$ Glossophaga leachii $6.61$ $42$ Glossophaga soricina $10.58$ $1$ $100$ Cratogeomys fumosus $6.54$ $43$ Myotis carteri $10.41$ $1$ $101$ Mephitis macroura $6.43$ $44$ Megasorex gigas $10.29$ $1$ $102$ Myotis velifer $6.37^{a}$ $45$ Sciurus yucatanensis $10.2$ $1$ $103$ Peromyscus perfulvus $6.36$ $46$ Marmosa mexicana $10.12^{a}$ $2$ $104$ Sylvilagus floridanus $6.29$ $47$ Sphiggurus mexicanus $10.11$ $1$ $105$ Lasiurus blossevillii $6.17$ $48$ Dermanura axteca $10.07$ $1$ $106$ Sturnira hondurensis $6.04$ $49$ Alouatta palliata $9.95$ $1$ $107$ Peromyscus hylocetes $6.03$ $50$ Peromyscus levipes $9.90^{a}$ $2$ $108$ Desmodus rotundus $6.02$ $51$ Notocitellus annulatus $9.87^{a}$ $3$ $109$ Peromyscus difficilis $6$ $52$ Glosophaga morenoi $9.84^{a}$ $2$ $111$ Tacataria brailiensis $5.86$ $5$	37	Sciurus aureogaster	11.04	1	95	Artibeus jamaicensis	6.86	1
39Eptesicus furinais10.05570Dermanura tolteca6.7740Peromyscus spicilegus $10.72^a$ 298Sciurus oculatus6.6741Cuniculus paca $10.59$ 199Glossophaga leachii6.6142Glossophaga soricina $10.58$ 1100Cratogeomys fumosus6.5443Myotis carteri $10.41$ 1101Mephitis macroura6.4344Megasorex gigas $10.29$ 1102Myotis velifer6.3645Sciurus yucatanensis $10.2$ 1103Peromyscus perfulvus6.3646Marmosa mexicana $10.12^a$ 2104Sylvilagus floridanus6.2947Sphiggurus mexicanus $10.11$ 1105Lasiurus blossevillii6.1748Dermanura azteca $10.07$ 1106Sturnira hondurensis6.0350Peromyscus levipes $9.90^a$ 2108Desmodus rotundus6.0251Notocitellus annulatus $9.87^a$ 3109Peromyscus difficilis652Glossophaga morenoi $9.84^a$ 110Tadarida brasiliensis5.8653Spilogale gracilis $9.74^a$ 2112Corvnerbinus mexicanus5.64	38	Liomvs irroratus	$10.88^{a}$	3	96	Cryptotis phillipsii	6.84	1
$40$ Peromyscus spicilegus $10\cdot72^a$ $2$ $98$ Sciurus outlatus $6\cdot67$ $41$ Cuniculus paca $10\cdot72^a$ $2$ $98$ Sciurus outlatus $6\cdot67$ $41$ Cuniculus paca $10\cdot59$ $1$ $99$ Glossophaga leachii $6\cdot61$ $42$ Glossophaga soricina $10\cdot58$ $1$ $100$ Cratogeomys fumosus $6\cdot54$ $43$ Myotis carteri $10\cdot41$ $1$ $101$ Mephitis macroura $6\cdot43$ $44$ Megasorex gigas $10\cdot29$ $1$ $102$ Myotis velifer $6\cdot37^a$ $45$ Sciurus yucatanensis $10\cdot2$ $1$ $103$ Peromyscus perfulvus $6\cdot36$ $46$ Marmosa mexicana $10\cdot12^a$ $2$ $104$ Sylvilagus floridanus $6\cdot29$ $47$ Sphiggurus mexicanus $10\cdot11$ $1$ $105$ Lasiurus blossevillii $6\cdot17$ $48$ Dermanura azteca $10\cdot07$ $1$ $106$ Sturnira hondurensis $6\cdot04$ $49$ Alouatta palliata $9\cdot95$ $1$ $107$ Peromyscus hylocetes $6\cdot03$ $50$ Peromyscus levipes $9\cdot90^a$ $2$ $108$ Desmodus rotundus $6\cdot02$ $51$ Notocitellus annulatus $9\cdot87^a$ $3$ $109$ Peromyscus difficilis $6$ $52$ Glossophaga morenoi $9\cdot84^a$ $2$ $110$ Tadarida brasiliensis $5\cdot86$ $53$ Spilogale gracilis $9\cdot84^a$ $2$ $111$ Tlacuatzin canescens $5\cdot82$ $54$ Peromyscus	39	Eptesicus furinalis	10.75	1	97	Dermanura tolteca	6.77	1
41Cuniculus paca10.59199Glossophaga leachii6.6142Glossophaga soricina10.581100Cratogeomys fumosus6.5443Myotis carteri10.411101Mephitis macroura6.4344Megasorex gigas10.291102Myotis velifer6.37°45Sciurus yucatanensis10.21103Peromyscus perfulvus6.3646Marmosa mexicana10.12°2104Sylvilagus floridanus6.2947Sphiggurus mexicanus10.111105Lasiurus blossevillii6.1748Dermanura azteca10.071106Sturnira hondurensis6.0449Alouatta palliata9.951107Peromyscus hylocetes6.0350Peromyscus levipes9.90°2108Desmodus rotundus6.0251Notocitellus annulatus9.87°3109Peromyscus difficilis652Glossophaga morenoi9.84°2110Tadarida brasiliensis5.8653Spilogale gracilis9.84°2111Tlacuatzin canescens5.8254Peromyscus maniculatus9.74°2112Corvnerbinus mexicanus5.64	40	Peromyscus spicilegus	$10.72^{a}$	2	98	Sciurus oculatus	6.67	1
42Glossophaga soricina10.581100Cratogeomys fumosus6.5443Myotis carteri10.411101Mephitis macroura6.4344Megasorex gigas10.291102Myotis velifer $6.37^a$ 45Sciurus yucatanensis10.21103Peromyscus perfulvus $6.36$ 46Marmosa mexicana10.12 <sup>a</sup> 2104Sylvilagus floridanus $6.29$ 47Sphiggurus mexicanus10.111105Lasiurus blossevillii $6.17$ 48Dermanura azteca10.071106Sturnira hondurensis $6.04$ 49Alouatta palliata9.951107Peromyscus hylocetes $6.03$ 50Peromyscus levipes9.90 <sup>a</sup> 2108Desmodus rotundus $6.02$ 51Notocitellus annulatus $9.87^a$ 3109Peromyscus difficilis $6$ 52Glossophaga morenoi $9.84^a$ 110Tadarida brasiliensis $5.86$ 53Spilogale gracilis $9.74^a$ 111Corvnerbinus mexicanus $5.64$	41	Cuniculus paca	10.59	1	99	Glossophaga leachii	6.61	1
43Myotis carteri10.411101Mephilis macroura $6.43$ 44Megasorex gigas $10.29$ 1 $102$ Myotis velifer $6.37^a$ 45Sciurus yucatanensis $10.2$ 1 $103$ Peromyscus perfulvus $6.36$ 46Marmosa mexicana $10.12^a$ 2 $104$ Sylvilagus floridanus $6.29$ 47Sphiggurus mexicanus $10.11$ 1 $105$ Lasiurus blossevillii $6.17$ 48Dermanura azteca $10.07$ 1 $106$ Sturnira hondurensis $6.04$ 49Alouatta palliata $9.95$ 1 $107$ Peromyscus hylocetes $6.03$ 50Peromyscus levipes $9.90^a$ 2 $108$ Desmodus rotundus $6.02$ 51Notocitellus annulatus $9.87^a$ 3 $109$ Peromyscus difficilis $6$ 52Glossophaga morenoi $9.84^a$ 2 $110$ Tadarida brasiliensis $5.86$ 53Spilogale gracilis $9.74^a$ 2 $112$ Corvnorbinus mexicanus $5.64$	42	Glossophaga soricina	10.58	1	100	Cratogeomys fumosus	6.54	1
44Megasorex gigas $10\cdot 29$ 1 $102$ Myotis velifer $6\cdot 37^{a}$ 45Sciurus yucatanensis $10\cdot 2$ 1 $103$ Peromyscus perfulvus $6\cdot 36$ 46Marmosa mexicana $10\cdot 12^{a}$ 2 $104$ Sylvilagus floridanus $6\cdot 29$ 47Sphiggurus mexicanus $10\cdot 11$ 1 $105$ Lasiurus blossevillii $6\cdot 17$ 48Dermanura azteca $10\cdot 07$ 1 $106$ Sturnira hondurensis $6\cdot 04$ 49Alouatta palliata $9\cdot 95$ 1 $107$ Peromyscus hylocetes $6\cdot 03$ 50Peromyscus levipes $9\cdot 90^{a}$ 2 $108$ Desmodus rotundus $6\cdot 02$ 51Notocitellus annulatus $9\cdot 87^{a}$ 3 $109$ Peromyscus difficilis $6$ 52Glossophaga morenoi $9\cdot 84^{a}$ 2 $110$ Tadarida brasiliensis $5\cdot 86$ 53Spilogale gracilis $9\cdot 84^{a}$ 2 $111$ Tlacuatzin canescens $5\cdot 82$ 54Peromyscus maniculatus $9\cdot 74^{a}$ 2 $112$ Corvnerbinus mericanus $5\cdot 64$	43	Myotis carteri	10.41	1	101	Mephitis macroura	6.43	1
45Sciurus yucatanensis $10\cdot2$ 1 $103$ Peromyscus perfulvus $6\cdot36$ 46Marmosa mexicana $10\cdot12^a$ 2 $104$ Sylvilagus floridanus $6\cdot29$ 47Sphiggurus mexicanus $10\cdot11$ 1 $105$ Lasiurus blossevillii $6\cdot17$ 48Dermanura azteca $10\cdot07$ 1 $106$ Sturnira hondurensis $6\cdot04$ 49Alouatta palliata $9\cdot95$ 1 $107$ Peromyscus hylocetes $6\cdot03$ 50Peromyscus levipes $9\cdot90^a$ 2 $108$ Desmodus rotundus $6\cdot02$ 51Notocitellus annulatus $9\cdot87^a$ 3 $109$ Peromyscus difficilis $6$ 52Glossophaga morenoi $9\cdot84^a$ 2 $110$ Tadarida brasiliensis $5\cdot86$ 53Spilogale gracilis $9\cdot84^a$ 2 $111$ Tlacuatzin canescens $5\cdot82$ 54Peromyscus maniculatus $9\cdot74^a$ 2 $112$ Corvnorbinus mexicanus $5\cdot64$	44	Megasorex gigas	10.29	1	102	Mvotis velifer	$6.37^{\mathrm{a}}$	3
46Marmosa mexicana10·12°2104Sylvilagus foridanus6·2947Sphiggurus mexicanus10·111105Lasiurus blossevillii6·1748Dermanura azteca10·071106Sturnira hondurensis6·0449Alouatta palliata9·951107Peromyscus hylocetes6·0350Peromyscus levipes9·90°2108Desmodus rotundus6·0251Notocitellus annulatus9·87°3109Peromyscus difficilis652Glossophaga morenoi9·84°2110Tadarida brasiliensis5·8653Spilogale gracilis9·84°2111Tlacuatzin canescens5·8254Peromyscus maniculatus9·74°2112Corvnorhimus mericanus5·64	45	Sciurus yucatanensis	10.2	1	103	Peromyscus perfulvus	6.36	1
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53Spilogale gracilis9.84ª2111Tlacuatzin canescens5.8254Peromuscus maniculatus9.74ª2112Conversitions mexicanus5.64	52	Glossophaga morenoi	$9.84^{\rm a}$	2	110	Tadarida brasiliensis	5.86	1
54 Peromyscus maniculatus 9.74 <sup>a</sup> 2 112 Corynorbinus mericanus 5.64	53	Spilogale gracilis	$9.84^{\mathrm{a}}$	2	111	Tlacuatzin canescens	5.82	1
51 Foromyseus municululus 571 2 112 Corynominus mexiculus 507	54	Peromyscus maniculatus	$9.74^{\mathrm{a}}$	2	112	Corynorhinus mexicanus	5.64	1
55Conepatus semistriatus9.661113Choeronycteris mexicana5.44	55	Conepatus semistriatus	9.66	1	113	Choeronycteris mexicana	5.44	1
56Diphylla ecaudata9.631114Pteronotus parnellii5.17	56	Diphylla ecaudata	9.63	1	114	Pteronotus parnellii	5.17	1
57Potos flavus $9.60^{a}$ 3115Peromyscus gratus $5.15$	57	Potos flavus	$9.60^{\mathrm{a}}$	3	115	Peromyscus gratus	5.15	1
58 Peromyscus megalops 9.55 1 116 Habromys lepturus 4.74	58	Peromyscus megalops	9.55	1	116	Habromys lepturus	4.74	1

*R*, relative rank of a mammal species, the lowest values of *R* being the most important;  $\varepsilon$ , epsilon values for mammal species estimated according to the level of co-occurrence with a given triatominae species; Tri., number of species of Triatominae potentially interacting with a given mammal species.

<sup>a</sup> Only the highest  $\varepsilon$  value is reported for a mammal species associated to two or more triatominae species.



Fig. 3. Hypotheses of ecological epidemiology of Chagas disease in Mexico. Hypotheses were examined considering the wild mammal species confirmed as *Trypanosoma cruzi* hosts in Mexico. (A) H1. The probability for a mammal species to be confirmed for *T. cruzi* is different among different quartiles. (B) H2. The probability for a mammal species to be confirmed is not correlated to the number of Triatominae species associated with a mammal species. (C) H3. The number of confirmed mammal species is correlated to the number of mammal species associated to a triatomine species. (D) H4. Percentages of confirmed mammals are different among triatomine species.

hand, checking three different  $\varepsilon$  thresholds ( $\varepsilon > 1.96$ ,  $\varepsilon > 4$  and  $\varepsilon > 6$ ), the percentage of true positives increased significantly in high quartiles as a function of the  $\varepsilon$  threshold, for example, with threshold  $\varepsilon > 4$ , the true positives percentage was higher in Q3 (27%), than with threshold  $\varepsilon > 1.96$  Q3 (17%) ( $\chi^2 = 6.7$ , P =0.03), and with threshold  $\varepsilon > 4$ , the true positives percentage was higher in Q4 (33%), than with threshold  $\varepsilon$ > 1.96 Q4 (29%) ( $\chi^2 = 24.2$ , P < 0.0001). These results are to be expected given that a higher threshold on  $\varepsilon$ means that we are restricting attention to a smaller subset of relations which are more statistically significant and therefore more likely to be associated with a true positive.

#### Ecological epidemiology of Chagas disease

Our literature survey uncovered 37 wild mammal species confirmed as hosts of T. cruzi in non-desert areas of Mexico (Table 2). Of these species 43% are of the order Rodentia, 38% are Chiroptera and the

other records are Carnivora, Didelphimorphia and Xenarthra (19%). Of these 37 species, 32 have been identified as positive using multiple diagnostic tests. We were not able to determine the used diagnostic test for T. cruzi for the remaining five species: Carollia perspicillata, Dasypus novemcinctus, Hodomys alleni, Ototylomys phyllotis and Tylomys nudicaudus.

With our list of mammals ranked by  $\varepsilon$ , and the list of confirmed hosts, we can test the network as a predictive model and also construct some simple hypotheses based on the overall structure of the network. More sophisticated hypotheses will potentially need additional data beyond just point collection data. Overall, all the confirmed mammal species were predicted by our analysis as being potentially associated in a statistically significant way to at least one triatomine species ( $\varepsilon > 1.96$ ). This allows us to formulate a first prediction: for a given mammal species, to be a host it must co-occur with the vector, we posit then that it is more likely to be

Table 2. Wild mammal species confirmed as *Trypanosoma cruzi* hosts in Mexico

	Confirmed mammal	OR	Method	Q
1	Artibeus lituratus	Ch	Р	4
2	Baiomys musculus	Rd	B,C,P	4
3	Carollia perspicillata	Ch	ND	4
4	Carollia sowelli (brevicauda)	Ch	Р	4
5	Dasyprocta punctata	Rd	A,B	4
6	Didelphis marsupialis	Dp	A,B,C,X	4
7	Didelphis virginiana	Dp	B,C,P,X	4
8	Glossophaga soricina	Cĥ	B,C	4
9	Heteromys desmarestianus	Rd	B,C,X	4
10	Heteromys gaumeri	Rd	PCR	4
11	Liomys irroratus	Rd	B,C,P	4
12	Otospermophilus (Spermophilus) variegatus	Rd	B,C	4
13	Ototylomys phyllotis	Ch	ND	4
14	Peromyscus leucopus	Rd	A.B	4
15	Peromyscus levipes	Rd	P	4
16	Peromyscus mexicanus	Rd	B.C.X	4
17	Peromyscus vucatanicus	Rd	A.B.P	4
18	Philander oposum	Dp	Ř	4
19	Reithrodontomvs fulvescens	Rd	Р	4
20	Sigmodon hispidus	Rd	B.C.P.X	4
21	Artibeus iamaicensis	Ch	B.C.P.X	3
22	Choeronvcteris mexicana	Ch	B.C	3
23	Desmodus rotundus	Ch	B.C.X	3
24	Hodomvs alleni	Rd	ND	3
25	Leptonvcteris verbabuenae (curasoae)	Ch	B.C	3
26	Myotis keavsi	Ch	-,- P	3
27	Nasua narica	Cr	P	3
28	Peromyscus melanophrys	Rd	B.C	3
29	Sturnira hondurensis (ludovici)	Ch	-,- P	3
30	Sturnira lilium	Ch	BCP	3
31	Tylomys mudicaudus	Rd	ND	3
32	Dasvous novemcinctus	Xn	ND	2
33	Dermanura phaeotis	Ch	P	2
34	Neotoma mexicana	Rd	BCP	2
35	Procvon lotor	Cr	Р.	2
36	Pteronotus parnellii	Ch	BC	2
37	Urocyon cinereoargenteus	Cr	A B	1
<u></u>	Crocyon emercourgenicus	01	11,12	1

OR, order of mammal species; Q, quartil of  $\varepsilon$  values, quartile 1 (Q1) being the lowest  $\varepsilon$  values and Q4 the highest; A, antibodies; B, blood smear; C, culture; Ch, Chiroptera; Cr, Carnivora; Dp, Didelphimorphia; P, polymerase chain reaction; R, random amplified polymorphic DNA; Rd, Rodentia; X, Xenodiagnosic; Xn, Xenarthra; ND, no data.

confirmed as a host of T. cruzi if it has a statistically significant overlap with a triatomine species (Table 3, H1). To test this hypothesis, we ranked all mammal species with significant co-occurrence associations ( $\varepsilon > 1.96$ ) with triatomine species according to their  $\varepsilon$  values and classified them in quartiles. So, quartile 1 (Q1) represents the mammals with the lowest  $\varepsilon$  values and Q4 the highest, ranking from low to high statistically significant associations between triatomine and mammal species. The corresponding quartiles of confirmed hosts were then assigned (Table 2). We found that the level of association between a mammal and a triatomine species correlated very well with the probability to be a confirmed host for T. cruzi ( $\chi^2 = 34.385$ , P = 0.0005, Fig. 3A). Thus, we can see that our inferred interaction network (Fig. 2) serves as a good prediction model for the vector-host system. Note that although only 29% of mammal species in Q4 have been

confirmed as hosts this serves only as a lower bound as many of the species in Q4 that have not been confirmed have either not been collected and tested for presence of T. cruzi or in such small numbers that a statistically significant rejection of them as hosts given a null hypothesis about the expected infection rate is not possible. The data in Fig. 3 were split into quartiles to facilitate the visual inspection of the relation between the true positive rate and the average value of  $\varepsilon$  in the quartiles in a way that presenting the regression coefficients and  $R^2$  value for the logistic regression does not. The coarse graining we use is not ad hoc. In the case of deciles rather than quartiles it is the standard grouping into by risk score used in the Hosmer-Lemeshow test often used with logistic regressions. We have also carried out a logistic regression at the species level. The associated relation is: Logit  $P = -3.648 + 0.235 \times \varepsilon$ , with a P-value < 0.001 on the regression coefficient. This

Hyp.	Data	Ho.	Hi.
H1	Mammal and triatominae species with significant values of $\varepsilon$ (Q1 to Q4)	The probability for a mammal species to be confirmed is independent of the level of co-occurrence with a triatomine species (Quartil)	The probability for a mammal species to be confirmed depends on the level of co-occurrence with a triatomine species (Quartil)
H2	Only mammal and triatomi- nae species with the highest significant values of $\varepsilon$ (Q4)	The probability for a mammal species to be confirmed is not correlated to the number of triatomine species co-occurring with the mammal species	The probability for a mammal species to be confirmed increases when the number of triatomine species co-occurring with the mammal species increases
Н3		The number of confirmed mammal species is not correlated with the number of mammal species co- occurring with a triatomine species	The number of confirmed mammal species increases when the number of mammal species co-occurring with a triatomine species increases
H4		The ability of triatomine species to transmit the parasite after a feeding interaction of its individuals is inde- pendent of the probability that a mammalian species is confirmed	The ability of triatomine species to transmit the parasite after a feeding interaction of its individuals explains the probability that a mammalian species is confirmed

Table 3. Hypotheses of ecological epidemiology of Chagas disease in Mexico

Hypotheses were formulated considering patterns of interaction among potential vectors and hosts of *Trypanosoma cruzi* ( $\varepsilon$  data).

confirms the statistically significant relation between  $\varepsilon$  as a statistical measure of geographical overlap and the probability to be a host of *T. cruzi*.

A second hypothesis is that mammal species that co-occur significantly with several triatomine species have a higher chance of being hosts of T. cruzi than mammal species that co-occur with few triatomine species (Table 3, H2). If this hypothesis is confirmed, then we expect an increment in the proportion of confirmed mammal host species as the number of associations increase. To test this hypothesis we assumed that all triatomine species have the same competence to transmit T. cruzi and the same population density. In this case, we cannot reject the null hypothesis at any level of statistical confidence and so we conclude that the probability to be a confirmed mammal host does not increase proportionally to the number of triatomine species for which these mammal species co-occur ( $r^2 = 0.72$ , P = 0.09, Fig. 3B).

A third hypothesis is that the transmission of T. *cruzi* is a more common process for triatomine species associated with many mammal species than for triatomine species associated with only a few (Table 3, H3). If this hypothesis is valid, then the number of confirmed mammal host species will increase as the number of mammal associated to a triatomine species increases. Again, to test this hypothesis we assumed that all triatomine species have the same competence to transmit T. *cruzi* and the same population density. Our statistical test indicated that when the number of mammal associated with a triatomine species increases the number of confirmed mammal host species also increases ( $r^2 = 0.72$ , P = 0.01, Fig. 3C).

The previous hypotheses explain the role of mammal species in the transmission of *T. cruzi*,

but ignore the explicit role of each vector. Also, we are implicitly considering the importance of a given mammal species for transmission of T. cruzi, without taking into account specific DTUs. In contrast to the above analyses, to explain the role of a specific vector we should ignore the role of the interaction in the transmissibility of the parasite. In other words, we assume that every triatomine species and their potential feeding resources are isolated from the other triatomine species or triatomine species that do not share any mammal species. In accordance with this setting, we can evaluate whether the transmission characteristics of distinct triatomine species are different (Table 3, H4). From the total set of potential feeding resources of a triatomine species, we compared the confirmed and non-confirmed mammal species percentages. If the transmissibility of triatomine species were the same, we would expect the percentages to be conserved among triatomine species. We found that the percentages of confirmed mammal hosts were different among triatomine species  $(\chi^2 = 70.419, P = 4.055 \times 10^{-12}),$ being the highest for T. barberi, T. dimidiata 2 and 3, and T. pallidipennis (Fig. 3D). Our results could be interpreted as showing that every triatomine species has a different competence to transmit T. cruzi and/or a different population density. It is interesting that in this way one can potentially infer indirectly vector competence in terms of the proportion of species it may infect.

Finally, to test the hypothesis that the statistical associations between vector and potential host do not simply reflect the relative range size of the different mammal (i.e., that a ranking by  $\varepsilon$  is different to a ranking by the distribution range), we show in Table S2 of the supporting information

that these distributions are quite distinct both *T. dimidiata* 2 (t = 2.53, P = 0.01) and *T. picturata* (t = -2.57, P = 0.01). Therefore, mammals' range sizes do not explain the observed co-occurrence patterns.

#### DISCUSSION

We inferred the potential vectors and hosts involved in the transmission of T. cruzi in non-desert ecoregions of Mexico and deduced the possible epidemiological consequences of triatomine-mammal interactions based on their geographic co-occurrence patterns. Certainly, transmission of T. cruzi could potentially occur in ways that do not directly involve a vector, e. g. maternal infection, feeding on infected mammals, etc. (Jansen et al. 2015). However, for a mammal species sharing most of its distribution with a triatomine species, we would expect that an important transmission route should be through vector interactions. An advantage of the type of analysis carried out here is that mammal occurrence data are much more complete and widely available than abundance data. Therefore, we posit that interaction networks inferred from co-occurrence patterns are efficient proxies with which to recognize potential hosts of T. cruzi and to understand their macro-level transmission dynamics in megadiverse countries.

As Mexico is a megadiverse country, there is a huge number of possible components of the vector-host system: 550 wild mammals and more than 30 Triatominae species (Ceballos and Arroyo, 2012; Ramsey et al. 2015). Complex Inference Networks allow us to recognize the most likely and most important wild hosts of T. cruzi, considering only those species with a significant co-occurrence. We predicted which were the most important mammal (116 species) and triatomine species involved in the ecological epidemiology of Chagas disease in Mexico. The high level of coincidence found between the predicted and confirmed hosts (Table 2), implies that many mammal species in our vector-host system can be considered as potential hosts or T. cruzi. Our results can help drive efforts for future experimental studies to confirm if the most probable predicted hosts are actually reservoirs of T. cruzi.

Interaction networks allow us to recognize patterns of transmissibility of T. cruzi. Testing our hypothesis 1 we observed the most of mammals confirmed positives to T. cruzi in the top quartile of our ranked list (Fig. 3A). This top quartile includes the most important spatial associations of triatomine and mammal species, relative to quartiles 1 to 3. Therefore, we rejected the null hypothesis of a vector-independent transmission of T. cruzi, which could exhibit a same number of confirmed mammals for all mammal species with a statistically significant geographical overlap with triatomines (Q1 to Q4). We concluded that hosting a T. cruzi can be correlated to mammal and vector co-occurrence and we interpreted this as an evidence of biotic interaction between mammals and triatomines.

Of course, the nature of this biotic interaction can itself be quite complex and multi-faceted. The most natural interaction, given that Triatominae are hematophagous, should be a feeding interaction, whereby a triatomine takes a bloodmeal from the mammal and the consequent triatomine defecation leads to an infection. This type of interaction can be confirmed with studies of blood meal origin at mammal species level. For example, our prediction of Mephitis macroura (Mephitidae: Carnivora), Reithrodontomys fulvescens and Sigmodon mascotensis (Cricetidae: Rodentia) as feeding resources of T. longipennis has been confirmed by a blood meal origin study (Bosseno et al. 2009). Similarly, B. musculus has been confirmed as a feeding resource of T. barberi, T. pallidipennis and T. phyllosoma (Mota et al. 2007). Given the scarcity of research about blood meal origin for triatomine species in Mexico (Mota et al. 2007; Bosseno et al. 2009; Ramsey et al. 2012), the interactions inferred by our model remain mostly as potential species. Another plausible type of interaction is that triatomine species are feeding resources for a given mammal species. However, this scenario seems less common in our network as most of the mammal species inferred are not insectivores (75%; González-Salazar et al. 2014) (Table S3, Supporting Information).

It is important to note that, although the detailed nature of the biotic interaction between an individual vector and an individual host may be important at some level, it does not affect our results, which are at a macro, ecosystemic level and therefore independent of the precise details of the interaction. That is not to say such details are unimportant. Moreover, this complexity extends further, considering the inclusion of the parasite itself, in that different mammals could have quite different competencies. Each species could deal in a different way with an infection by distinct T. cruzi DTUs. Similarly, the detailed behavioural traits of different mammal species can affect transmission probabilities. For instance, the confirmed host D. novemcinctus constructs burrows that upon abandonment are often used as shelter or breeding sites by other mammal species and triatominae thereby allowing for a Triatomine to feed on multiple mammal species. In principle, our methodology could take into account much more complexity if there were data to support it. For instance, there is no comprehensive database that lists the competencies of all mammal species with respect to all DTUs for instance.

The data that do exist for all species is where they are, at least as proxied by point collection data. Our research in that sense provides a first, crude but effective approximation to a very complex system: the vector-host ecosystem of Chagas disease in Mexico. In this respect, hypotheses 2 and 3 attempt to explain the role of potential interactions between mammal and triatomine species in the transmission of T. cruzi. We are acutely aware that transmissibility of T. cruzi involves many factors, not only mammal and triatomine co-occurrence. Here, the explicit role of every vector and mammal species, i.e. its competence and population density, was assumed similar in the absence of standard data of competence of triatomine species and available data on population densities. These are model assumptions. The approximate validity of those assumptions is tested by the results of the model. From the available data, we predict that transmissibility stays relatively constant for mammal species associated to a few or a lot of triatomine species (H2) with about 20% of mammals with significant  $\varepsilon$  values being confirmed hosts independent of the associated vector. Also, our data allow us to predict that the probability of transmission of T. cruzi increases for triatomine species associated from a few to a lot of mammal species (H3). This is not just a question of expecting that the more mammal species that are sampled the more positives one would expect. If mammals with low  $\varepsilon$  values were sampled then one would expect only a small number of confirmed mammals or none, no matter how many mammal species were sampled.

Testing differences in the transmissibility of T. cruzi among triatomine species from spatial data (Hypothesis 4), we observed that *Triatoma barberi*, T. dimidiata and T. pallidipennis have a particularly important role in T. cruzi transmission. This result might be expected as these species have the widest distribution in Mexico (Ramsey et al. 2015), but for the first time we were able to predict the epidemiological importance of some triatominae species by considering the number of potential feeding resources and confirmed mammal hosts. The transmission of T. cruzi is a process that usually occurs by contact between a mammal and a vector, such as via contaminated triatomine feces, a process that is assumed to be repeated in proportion to the populations and distribution sizes of the corresponding mammal and triatomine species. Due to the nature of the point collection data used, the population density of the species is unknown. However, we do know that all the interactions (links in our network) come from a large spatial overlap between mammal and triatomine distributions. Hence, a link in the network between mammal and triatomine species means a high potential for T. cruzi transmission. Therefore, we were able to test hypotheses of transmissibility with our vector-host system because potential interactions between species are quantifiable.

We note that differences in the transmissibility of T. cruzi between triatomine species are not only a result of mammal and vector densities and distributions, but also a result of different vector competences. For example, Triatoma barberi exhibits the best competence to transmit T. cruzi having the highest natural infection index, the highest frequency of trypomastigotes and the shortest time for defecation among the main vectors of Chagas disease in Mexico (Salazar-Schettino et al. 2005). Likewise, T. dimidiata and T. pallidipennis are recognized by their high degree of competence among the main vectors of Chagas disease in Mexico (Martínez-Ibarra and Novelo-López, 2004; Salazar-Schettino et al. 2005; Dorn et al. 2007). Even though there are no differences in the competence of distinct lineages of T. dimidiata, there are differences in their spatial dynamics (Herrera-Aguilar et al. 2009). Triatoma dimidiata 3 participates in the flow between sylvatic and domestic environments whereas T. dimidiata 2 does not, being restricted to only domestic habitats exclusively (Herrera-Aguilar et al. 2009).

Finally, we were able to recognize the epidemiological consequences of interactions between mammal and triatomine species, in spite of the limitations of our data and assumptions. Certainly, distribution datasets accumulate taxonomic and geographic sampling biases, for instance, accounting for the fact that some areas have been subject to intense field surveys while others have not. However, it is essential that we take advantage of the huge quantity of accumulated data (Varela et al. 2014). Although we did not analyse in depth the potential effect of sampling biases in our data, we believe that our coarse graining can reduce to some extent some collection bias by only counting once multiple collections in one grid square. In addition, Complex Inference Networks have been shown to be predictive in spite of sampling collection biases, as has been shown in the case of Leishmaniasis (Berzunza-Cruz et al. 2015; Stephens et al. 2016).

We are aware that the list of confirmed mammal species with T. cruzi has some limitations. In the first place, the list is not definitive and does not come from systematic samples of Mexico, but still represents the most complete knowledge available for the country today. Our list also reflects the diversity of mammal species by order in Mexico (Ceballos and Arroyo, 2012) and seems to not be biased for widely distributed mammal species. Secondly, some level of uncertainty is to be expected in T. cruzi determination depending on the diagnostic method used. Finally, we did not include information of DTU's of T. cruzi because this information is scarce in Mexico. However, one would expect that any pathogen that has a transmission cycle that involves triatomines and mammals will show a similar ecosystemic network. We would argue then that the list of confirmed mammal species with T. cruzi was sufficient to test our hypotheses.

With respect to our assumptions, we recognize that not only direct biotic interactions, such as feeding, but also other ecological interactions and evolutionary biogeographic processes can cause cooccurrence patterns between mammal and triatomine species (Morrone, 2009). In other words there may exist confounding factors such that a perceived pair correlation is really intermediated by another latent variable, such as climate. This can only be checked thoroughly by exhaustively including every potential confounding variable and checking if it is more predictive than its proxy. However, even if co-occurrence patterns had not been a direct consequence of feeding interactions, the fact that triatomines would feed more on those mammals that have a higher fraction of co-occurrences seems plausible because of their generalist habits. Only a few triatomine species have definite host preferences (Lent and Wygodzinsky, 1979) and they are not within the set of species studied herein (Dorn et al. 2007; Bosseno et al. 2009; Villalobos et al. 2011; Ramsey et al. 2012). We hope to examine some of the model assumptions within our vector-host system when more data are obtained. Finally, we hope that our findings will stimulate other researchers in the direction of making corresponding epidemiological hypotheses that can be verified by further experimental and ecological research.

Finally, we have emphasized throughout that cooccurrence, although an important necessary condition for a mammal to participate in the transmission cycle of Chagas disease, is not sufficient as the transmission cycle is highly multifactorial, with factors such as species abundance, phylogeny, sampling frequency, phenotypic characteristics of the potential host, to name but a few could play a significant role. Our formalism lends itself to the incorporation of such factors in a democratic fashion. What is lacking is to have databases that contain such information to integrate with the purely spatial data used here. This is an ongoing effort and will be reported in a future publication.

#### SUPPLEMENTARY MATERIAL

The supplementary material for this article can be found at https://doi.org/10.1017/S0031182016002468.

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