

Treatment of Multicentric Reticulohistiocytosis With Tocilizumab

César Pacheco-Tena, MD, MSc,* Greta Reyes-Cordero, MD,† Rosa Ochoa-Albíztegui, MD,*
Victor Ríos-Barrera, MD, PhD,‡ and Susana A. González-Chávez, MSc*

Abstract: Multicentric reticulohistiocytosis (MRH) is a rare multi-system, granulomatous debilitating disease. It affects the skin with a nodular diffuse dermatitis and the joints with a severe, potentially deforming, and handicapping arthritis. No standardized therapy exists, it is a disease with heterogeneous severity, and therefore, a diversity of therapeutic responses has been published.

Current experience with anti-tumor necrosis factor agents in disease-modifying antirheumatic drug-refractory MRH cases is encouraging, and other agents such as bisphosphonates have proven effective as well. Histological analysis of the granulomatous inflammatory lesions have shown the presence of cytokines including tumor necrosis factor α , interleukin 1, and interleukin 6; the presence of the latter makes tocilizumab a plausible alternative.

In this article, we report a 35-year-old woman with MRH refractory to a combined scheme of prednisone and methotrexate, both at high doses, and who received tocilizumab achieving remission on both cutaneous and articular symptoms. Our patient markedly improved by the second infusion (8 mg/kg monthly), and after 9 infusions, she remained asymptomatic; no toxicity was detected. Tocilizumab could be an alternative for disease-modifying antirheumatic drug-refractory MRH.

Key Words: tocilizumab, multicentric reticulohistiocytosis, biologic therapy

(*J Clin Rheumatol* 2013;19: 272–276)

Multicentric reticulohistiocytosis (MRH) is a chronic inflammatory disease of unknown origin. Its hallmark is the presence of spontaneous inflammatory nodular lesions that grow in the skin and in the synovial membrane.

It is often associated with arthritis, which may be severe, deforming, and handicapping (~60%); indeed, a mutilans variant is even more frequently observed (~45%) than in psoriatic arthritis.

Approximately 25% of MRH cases are paraneoplastic syndromes that have been linked to a diversity of neoplasms, including carcinomas (cervix, breast, lung, gastrointestinal, prostate, and others), leukemia, lymphoma, and melanoma.

The disease is often limited to the skin and joints; however, cases with notable systemic involvement affecting the lungs, heart, and kidney have been described as well.

Cases of MRH mimicking or coincidental with defined connective tissue diseases such as systemic lupus erythematosus,^{1,2} Sjögren syndrome,^{3,4} polymyositis, and anticitrullinated-positive rheumatoid arthritis (RA)-like disease⁵ have also been published, unveiling yet undefined relation.

Multicentric reticulohistiocytosis is a rare disease, with no standardized therapy; methotrexate (MTX) induces significant improvement or remission in some MRH patients according to several reports.^{6,7} Cyclophosphamide,⁸ leflunomide,⁹ azathioprine,¹⁰ and cyclosporin¹¹ have also proven effective; aside for immunosuppression, in several trials, bisphosphonates^{12–14} have also induced remission.

However, some reports present patients who were refractory to disease-modifying antirheumatic drugs, opening a door for alternative strategies such as biologic agents. Current experience includes cases successfully treated with etanercept,^{15,16} infliximab,^{17–20} or adalimumab²¹ as anti-tumor necrosis factor (anti-TNF) agents. To our knowledge, no MRH patient treated with tocilizumab (TCZ) has been reported.

In correlation with the presence of activated, committed macrophagic cells in the MRH nodules, interleukin 1 (IL-1), TNF- α , and IL-6 are overexpressed, and these could be related to the clinical presentation and also contribute to disease pathogenesis.

The intricate relationship between TNF- α and IL-6 creates redundant loops or signaling cycles where blocking either of them results on similar effects over inflammation.¹⁸ The blockade of IL-6 has successfully followed anti-TNF therapy in some of its indications.

Interleukin 6 is overexpressed in the rheumatoid synovium and other inflamed tissues from diseases where TCZ has proven effective; IL-6 was expressed in the MRH inflammatory infiltrate of our patient, in the giant multinucleated cells (GMCs); therefore, TCZ was proposed as a specific immunomodulator to treat this refractory patient.

In this article, we report the case of a 35-year-old woman with MRH, who was refractory to corticosteroids and MTX at top doses and who eventually received TCZ and achieved sustained remission.

CASE PRESENTATION

On September 2009, the patient presented with general malaise, progressive weight loss (15 kg/6 months), mild evening hyperthermia, and diffuse myoarthralgias. A primary care physician prescribed diclofenac; early symptoms initially improved, but in the upcoming months they got worse.

Eventually, an additive polyarthritis started and settled in, presenting itself as a RA in its pattern. It affected both wrists, metacarpophalangeal and proximal interphalangeal (and distal interphalangeal) on both hands and also on both elbows, knees, ankles, and metatarsophalangeal, but it was more severe than

From the *Facultad de Medicina, Universidad Autónoma de Chihuahua, Circuito Universitario Campus II, Chihuahua; †Departamento de Reumatología, Hospital General de Mexico, Mexico City; and ‡Facultad de Odontología, Universidad Autónoma de Chihuahua, Chihuahua, Mexico.

This article received no funding from commercial sources. The authors declare no conflict of interest.

Correspondence: César Pacheco-Tena, MD, MSc, Facultad de Medicina, Universidad Autónoma de Chihuahua, Circuito Universitario Campus II, Chihuahua, Chihuahua, Mexico, CP 31125. E-mail: dr.cesarpacheco@gmail.com.

Copyright © 2013 by Lippincott Williams & Wilkins
ISSN: 1076-1608/13/1905-0272

DOI: 10.1097/RHU.0b013e31829cf32b

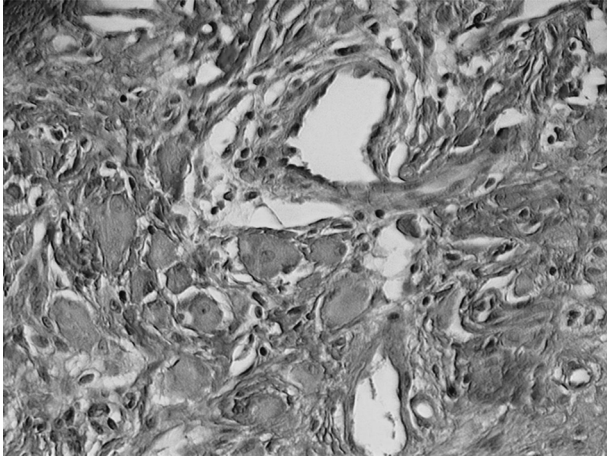


FIGURE 1. Hematoxylin-eosin stain micrograph (original magnification $\times 100$) showing the inflammatory infiltrate and multinucleated cells from one of the patient's nodules. Available in color online at www.jclinrheum.com.

that observed in an average RA patient; indeed, the patient's functional status plunged to functional class IV within days.

Concurrently, multiple vivid erythematous papular lesions appeared on the fingers' dorsa (periungual initially, as coral beds); they were neither painful nor pruriginous, and later on, they extended and raised up to become consistent, gummy nodules ranging in size from some millimeters up to 3 cm.

Nodules spread along the hands, forearms, feet, and forelegs, and in the upcoming weeks, they initially spread on the papulae, and shortly after, nodules were seen on the nose; the eyelids and ears were later involved.

The patient's referral to the hospital explained more the severity of the arthritis than by the notorious dermatitis. Multicentric reticulohistiocytosis was inferred at admission and confirmed with a biopsy days later (Fig. 1). Physical examination was unremarkable aside from joints and skin. No fever was detected; chest and abdomen showed neither abnormalities nor visceromegalies.

We scanned the patient looking for possible neoplasia; chest and abdomen computed tomographic scan, serology (CA-125 and carcinoembryonic antigen), and gynecologic assessment (breast and genitourinary tract) turned out negative, as the esophagogastroendoscopy did. Laboratory results included erythrocyte sedimentation rate of 47 mm/h; C-reactive protein, 16 mg/dL; rheumatoid factor, 1:160; negative anti-nuclear antibody and anti-cyclic citrullinated peptide antibody; hemoglobin, 9.6 g/dL; and normal liver function tests.

Our patient received 3 methylprednisolone IV pulses (1 g/d); afterward, oral prednisone (20 mg/d) and oral MTX (20 mg/wk) were prescribed. Dermatitis partially improved, but arthritis barely changed. One month after, the patient remained in functional class IV; prednisone was increased progressively up to 50 mg/d, achieving some improvement in the arthritis; but just as the dose was tapered down, intense relapse followed on 3 different attempts. Methotrexate dose was adjusted to 25 mg/wk without benefit. Multicentric reticulohistiocytosis is a potentially destructive disease; thus, aside from the handicapping picture, we were further concerned by the potential of structural damage.

After 4 months on top doses of MTX and prednisone with mild improvement, an alternative treatment was essential. Sadly, our patient had neither health insurance nor coverage for anti-TNF drugs, so these were not an option. Fortunately, a lot of

TCZ was available with no cost for 1 patient (likely with RA) as a donation for our hospital. The clinical picture of our patient was so desperate that we proposed to our authorities their approval to use the TCZ in our MRH patient. As a strategy to support the use of TCZ, immunofluorescence targeting IL-6 was performed on the nodule biopsy before the onset of the therapy (vide infra). It turned out positive.

After an initial approval, we submitted our petition to the ethics and research committee (it was clear in the submission that it was an off-label use); it was accepted. Finally, it was proposed to our patient, and she agreed. Initially, a 3-month treatment was permitted, and further use was reevaluated as by the results.

We screened the patient for latent tuberculosis with PPD testing and chest x-ray; eventually, a computed tomographic scan (vide supra) showed no evidence of prior or actual Tb infection.

Because the arthritis was so intense and refractory, we selected a high dose for TCZ (8 mg/kg). The patient was flawlessly infused, and after 2 weeks, a very significant improvement was observed in the dermatitis, and by the second dose, a complete absence of nodules was achieved (Fig. 2).

Arthritis improved promptly and sustainably; at the end of the first month, the patient was on functional class II and basically asymptomatic before the third dose. The MTX dose remained stable; prednisone was tapered in an unfixated, monthly



FIGURE 2. A, Patient's hand before starting TCZ (but after prednisone and MTX) showing incomplete resolution of dermatitis. B, Patient's hand after 4 months of treatment. Available in color online at www.jclinrheum.com.

reviewed scheme down to 7.5 mg/d; by the third dose of TCZ, neither cutaneous nor arthritic relapses occurred.

No adverse effects were observed throughout the course of the treatment, and laboratory results (including liver tests and complete blood count) remained unaffected; rheumatoid factor switched negative after the third month, and acute-phase reactants became normal after the second infusion. After 9 monthly infusions, the patient remained asymptomatic on skin and joints.

To carry out the IL-6 immunofluorescence staining, the slides were blocked for 1 hour at room temperature with phosphate-buffered saline at pH 7.4 (bovine fetal serum 10% and skimmed milk 5%). The blocking solution was removed, and approximately 100 μ L of the primary antibody in a 1:200 dilution was used to cover the tissue, for an overnight reaction at 4°C in a humid chamber, a negative control including everything in the solutions but the primary antibody, and was run in parallel. After incubation, slides were washed on phosphate-buffered saline, and a secondary antibody (fluorescein isothiocyanate-labeled goat anti-mouse antibody) was added in a 1:1000 dilution and incubated for an hour at room temperature.

The slides showed a positive detection of IL-6 in a variety of cells within the infiltrate (Fig. 3); of particular interest, GMCs showed an intense reaction. Negative control rendered only weak-background, nonspecific immunofluorescence. After the positive detection for IL-6, additional staining with macrophage (anti-CD68) and dendritic cell (anti-CD83) markers was done to further characterize GMCs. Those slides were reviewed under confocal microscopy. In both cases, these markers colocalized with IL-6-positive GMCs, suggesting a dendritic cell of monocyte origin lineage (Fig. 3). A fibroblast marker (ER-TR7) was used as a control and stained no GMCs. All antibodies used in the reactions came from Santa Cruz

Biotechnologies (Santa Cruz, CA), and the previously described protocol was used.

DISCUSSION

Pathogenesis of MRH is yet to be understood. Skin nodules and synovial membrane typically show an inflammatory infiltrate, with GMCs sparsely mixed with small histiocytes (indeed, MRH is a histiocytosis). This infiltrate includes also primed T cells (CD8 mainly) and B cells.

The lineage of the multinucleated cells (which resemble foreign-body type) has been debatable. Either a T cell, dendritic, or a monocyte lineage has been proposed, being this latter supported the most by current evidence, because morphologic and functional parallels link GMCs to osteoclasts.^{12,22}

The expression of markers for other lineages is, however, inconstant; that is, in the paper from Adamopoulos et al,²² GMCs were negative for CD3, CD20, and S100, excluding T cell, B cell, and dendritic lineage, respectively, but other cases with positive S100 GMCs have also been detected.²³ In our patient, markers for both macrophagic and dendritic cell lineages were present, suggesting a dendritic cell of macrophagic lineage.

These GMCs consistently express macrophage markers such as CD68, CD45 and are positive for tartrate-resistant acid phosphatase, Mac387, CD15, and others; frequently they express CD14.²⁴

The GMCs present a ground-glass aspect and contain deposits of an unspecific accumulation of neutral lipids (probably phospholipids)²⁴; this has been deduced based the staining affinity for this material rather than specific chemical characterization.²⁵ Images from electron microscopy²⁶ suggest that these lipid vacuoles (likely liposomes) result from a degenerative process more than from abnormal storage.

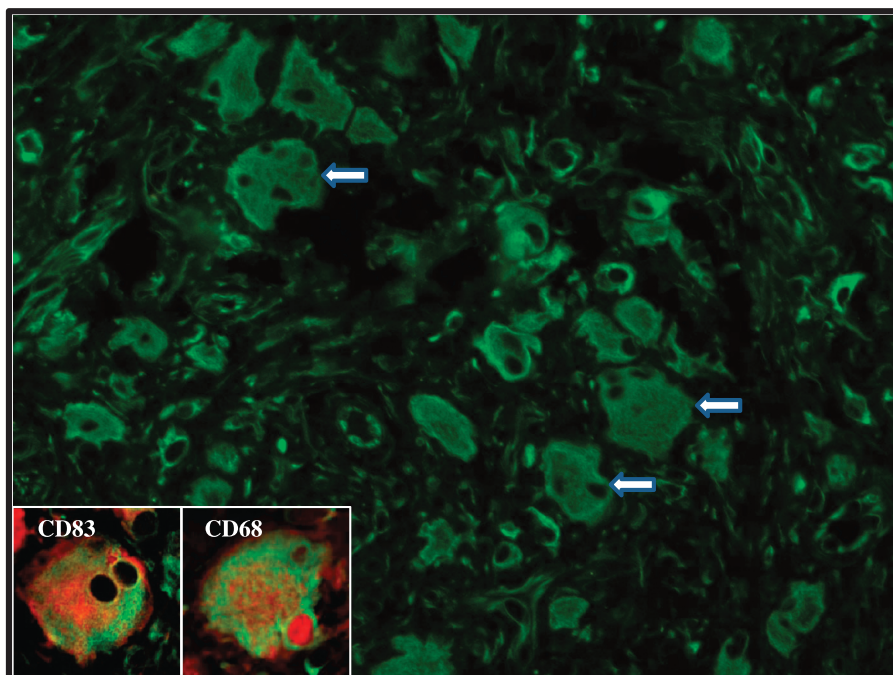


FIGURE 3. Immunofluorescence micrograph (original magnification $\times 100$) of a nodule biopsy stained with an anti-IL6 monoclonal antibody, showing a positive detection in a variety of cells in the infiltrate including GMCs (arrows). Corner: Confocal microscopy (original magnification $\times 100$) showing the positive detection of CD68, CD83, and its colocalization with IL-6 in GMCs.

Interleukin 6 is a pleiotropic cytokine and plays a role as a B-cell differentiation factor, induces T-cell expansion and differentiation to cytotoxic subsets, increases proliferation of hematopoietic precursors, induces macrophage differentiation at their terminal stages, and stimulates synthesis of acute-phase response proteins (such as C-reactive protein) in hepatocytes.

Interleukin 6 causes fever because it is able to cross the blood-brain barrier and stimulates hypothalamus to raise the body temperature. Interleukin 6 transgenic mice exhibit polyclonal hyperglobulinemia, splenomegaly, adenomegaly, and disperse inflammatory infiltrates. Some symptoms in MRH patients may be explained by the increased secretion of IL-6: fever, weight loss, arthritis, and bone resorption, and although the role for IL-6 as a trigger or nodule formation is not defined, its stimuli over macrophages could account for proliferation, activation, and differentiation.

Interleukin 6 may indeed play a role in the inflammatory structure of the nodules and account for some destructive features of MRH including osteoclastogenesis.²⁷ In our patient, IL-6 is clearly expressed in the cutaneous inflammatory nodule in a variety of cells including local histiocytes and GMCs. Interleukin 6 blockade with TCZ markedly improved her clinical picture.

Several bone-invasive neoplasms activate osteoclasts via IL-6,²⁷ which is expressed in stromal cells (mostly fibroblasts) within bone-tumor interface, and its expression correlates with that of RANKL (also in fibroblastic cells), conforming an osteoclastogenic environment in which bone invasiveness is the resultant.

Synovial fluid mononuclear cells from MRH arthritic patients are osteoclast precursors. The synovial fluid in patients with MRH contains a higher concentration of TNF- α (5 times) and lower concentrations of osteoprotegerin if compared with samples from osteoarthritis patients. In presence of TNF- α , RANKL is induced on osteoblasts and fibroblasts. Osteoclast differentiation among precursors is feasible.²²

In skin nodules, GMCs also express osteoclast surface markers, such as coincidental tartrate-resistant acid phosphatase and cathepsin K,²⁸ and interestingly, GMCs from MRH patients (in contrast to those from sarcoidosis, RA, and other granulomatous diseases) express CD10, a potential marker predicting its destructive and invasive nature; this marker is also known as neutral endopeptidase (among others) and is a zinc-dependent metalloproteinase present in the active invasive areas of several carcinomas (including lung and prostate).

Tocilizumab is a humanized immunoglobulin G1 antibody against IL-6 receptor (both soluble and membrane-attached) and blocks IL-6 proinflammatory actions. In 2009, TCZ was approved in Europe for moderate to severe RA and more recently in different countries.

Several trials have established the safety and efficacy of TCZ for MTX-refractory²⁹ anti-TNF-refractory³⁰ and naive early-RA patients, and aside from its efficacy to improve disease activity, long-term follow-up³¹ confirms a preventing effect on radiographic progression comparable to that of anti-TNF agents.^{32,33}

Tocilizumab represents a promising alternative and is currently being tested for additional indications (following anti-TNF trend); current reports include systemic lupus erythematosus and³⁴ spondylarthritis.^{35,36} Successfully treated case reports include patients with polymyositis,³⁷ renal amyloidosis,³⁸ polymyalgia rheumatica,³⁹ graft-versus-host disease,^{40,41} and Takayasu disease.^{42,43}

In this report, we describe the positive effect of TCZ in the skin and joints of a patient with MRH who was refractory to a combination of MTX and high-dose prednisone. Although

previous experience with anti-TNF agents in MRH is encouraging, the potential role of TCZ as an alternative agent is welcome especially in patients with contraindications to anti-TNF.

Further experience is required to ascertain the consistency of our findings.

REFERENCES

1. Badlissi F, Setty Y, Folzenlogen D. A case of multicentric reticulohistiocytosis initially misdiagnosed as lupus. *J Clin Rheumatol*. 2002;8:232–233.
2. Saito K, Fujii K, Awazu Y, et al. A case of systemic lupus erythematosus complicated with multicentric reticulohistiocytosis (MRH): successful treatment of MRH and lupus nephritis with cyclosporin A. *Lupus*. 2001;10:129–132.
3. Carey RN, Blotzer JW, Wolfe ID, et al. Multicentric reticulohistiocytosis and Sjögren's syndrome. *J Rheumatol*. 1985;12:1193–1195.
4. Morris-Jones R, Walker M, Hardman C. Multicentric reticulohistiocytosis associated with Sjögren's syndrome. *Br J Dermatol*. 2000;143:649–650.
5. Chauhan A, Mikulik Z, Hackshaw KV. Multicentric reticulohistiocytosis with positive anticyclic citrullinated antibodies. *J Natl Med Assoc*. 2007;99:678–680.
6. Cash JM, Tyree J, Recht M. Severe multicentric reticulohistiocytosis: disease stabilization achieved with methotrexate and hydroxychloroquine. *J Rheumatol*. 1997;24:2250–2253.
7. Franck N, Amor B, Ayril X, et al. Multicentric reticulohistiocytosis and methotrexate. *J Am Acad Dermatol*. 1995;33:524–525.
8. Liang GC, Granston AS. Complete remission of multicentric reticulohistiocytosis with combination therapy of steroid, cyclophosphamide, and low-dose pulse methotrexate. Case report, review of the literature, and proposal for treatment. *Arthritis Rheum*. 1996;39:171–174.
9. Lonsdale-Eccles AA, Haworth AE, McCrae FC, et al. Successful treatment of multicentric reticulohistiocytosis with leflunomide. *Br J Dermatol*. 2009;161:470–472.
10. Fedler R, Frantzman Y, Schwarze EW, et al. Multicenter reticulohistiocytosis. Therapy with azathioprine and prednisolone [in German]. *Hautarzt*. 1995;46:118–120.
11. Chalom EC, Rosenstein ED, Kramer N. Cyclosporine as a treatment for multicentric reticulohistiocytosis. *J Rheumatol*. 2000;27:556.
12. Goto H, Inaba M, Kobayashi K, et al. Successful treatment of multicentric reticulohistiocytosis with alendronate: evidence for a direct effect of bisphosphonate on histiocytes. *Arthritis Rheum*. 2003;48:3538–3541.
13. Satoh M, Oyama N, Yamada H, et al. Treatment trial of multicentric reticulohistiocytosis with a combination of prednisolone, methotrexate and alendronate. *J Dermatol*. 2008;135:168–171.
14. Mavragani CP, Batziou K, Aroni K, et al. Alleviation of polyarticular syndrome in multicentric reticulohistiocytosis with intravenous zoledronate. *Ann Rheum Dis*. 2005;64:1521–1522.
15. Alexis AF, Strober BE. Off-label dermatologic uses of anti-TNF- α therapies. *J Cutan Med Surg*. 2005;9:296–302.
16. Kovach BT, Calamia KT, Walsh JS, et al. Treatment of multicentric reticulohistiocytosis with etanercept. *Arch Dermatol*. 2004;140:919–921.
17. Kalajian AH, Callen JP. Multicentric reticulohistiocytosis successfully treated with infliximab: an illustrative case and evaluation of cytokine expression supporting anti-tumor necrosis factor therapy. *Arch Dermatol*. 2008;144:1360–1366.
18. Musacchio E, Valvason C, Botsios C, et al. The tumor necrosis factor- α -blocking agent infliximab inhibits interleukin 1beta (IL-1beta) and IL-6 gene expression in human osteoblastic cells. *J Rheumatol*. 2009;136:1575–1579.

19. Chiba E, Oda A, Tsutsumi T, et al. Case report; a case with multicentric reticulohistiocytosis successfully treated with infliximab [in Japanese]. *Nihon Naika Gakkai Zasshi*. 2011;100:483–486.
20. Sellam J, Deslandre CJ, Dubreuil F, et al. Refractory multicentric reticulohistiocytosis treated by infliximab: two cases. *Clin Exp Rheumatol*. 2005;123:97–99.
21. Shannon SE, Schumacher HR, Self S, et al. Multicentric reticulohistiocytosis responding to tumor necrosis factor- α inhibition in a renal transplant patient. *J Rheumatol*. 2005;132:565–567.
22. Adamopoulos IE, Wordsworth PB, Edwards JR, et al. Osteoclast differentiation and bone resorption in multicentric reticulohistiocytosis. *Hum Pathol*. 2006;37:1176–1185.
23. Bialynicki-Birula R, Sebastian-Rusin A, Maj J, et al. Multicentric reticulohistiocytosis with S100 protein positive staining: a case report. *Acta Dermatovenerol Croat*. 2010;18:35–37.
24. Gorman JD, Danning C, Schumacher HR, et al. Multicentric reticulohistiocytosis: case report with immunohistochemical analysis and literature review. *Arthritis Rheum*. 2000;43:930–938.
25. Campbell DA, Edwards NL. Multicentric reticulohistiocytosis: systemic macrophage disorder. *Baillieres Clin Rheumatol*. 1991;15:301–319.
26. Tani M, Hori K, Nakanishi T, et al. Multicentric reticulohistiocytosis. Electron microscopic and ultracytochemical studies. *Arch Dermatol*. 1981;117:495–499.
27. Kayamori K, Sakamoto K, Nakashima T, et al. Roles of interleukin-6 and parathyroid hormone-related peptide in osteoclast formation associated with oral cancers: significance of interleukin-6 synthesized by stromal cells in response to cancer cells. *Am J Pathol*. 2010;176:968–980.
28. Codriansky KA, Runger TM, Bhawan J, et al. Multicentric reticulohistiocytosis: a systemic osteoclastic disease? *Arthritis Rheum*. 2008;59:444–448.
29. Maini RN, Taylor PC, Szechinski J, et al. Double-blind randomized controlled clinical trial of the interleukin-6 receptor antagonist, tocilizumab, in European patients with rheumatoid arthritis who had an incomplete response to methotrexate. *Arthritis Rheum*. 2006;54:2817–2829.
30. Nakashima Y, Kondo M, Harada H, et al. Clinical evaluation of tocilizumab for patients with active rheumatoid arthritis refractory to anti-TNF biologics: tocilizumab in combination with methotrexate. *Mod Rheumatol*. 2010;120:343–352.
31. Nishimoto N, Ito K, Takagi N. Safety and efficacy profiles of tocilizumab monotherapy in Japanese patients with rheumatoid arthritis: meta-analysis of six initial trials and five long-term extensions. *Mod Rheumatol*. 2010;20:222–232.
32. Garnero P, Thompson E, Woodworth T, et al. Rapid and sustained improvement in bone and cartilage turnover markers with the anti-interleukin-6 receptor inhibitor tocilizumab plus methotrexate in rheumatoid arthritis patients with an inadequate response to methotrexate: results from a substudy of the multicenter double-blind, placebo-controlled trial of tocilizumab in inadequate responders to methotrexate alone. *Arthritis Rheum*. 2010;62:33–43.
33. Kremer JM, Blanco R, Brzosko M, et al. Tocilizumab inhibits structural joint damage in rheumatoid arthritis patients with inadequate responses to methotrexate: results from the double-blind treatment phase of a randomized placebo-controlled trial of tocilizumab safety and prevention of structural joint damage at one year. *Arthritis Rheum*. 2011;63:609–621.
34. Illei GG, Shirota Y, Yarboro CH, et al. Tocilizumab in systemic lupus erythematosus: data on safety, preliminary efficacy, and impact on circulating plasma cells from an open-label phase I dosage-escalation study. *Arthritis Rheum*. 2010;62:542–552.
35. Brulhart L, Nissen MJ, Chevallier P, et al. Tocilizumab in a patient with ankylosing spondylitis and Crohn's disease refractory to TNF antagonists. *Joint Bone Spine*. 2010;77:625–626.
36. Henes JC, Horger M, Guenaydin I, et al. Mixed response to tocilizumab for ankylosing spondylitis. *Ann Rheum Dis*. 2010;69:2217–2218.
37. Narazaki M, Hagihara K, Shima Y, et al. Therapeutic effect of tocilizumab on two patients with polymyositis. *Rheumatology (Oxford)*. 2011;50:1344–1346.
38. Hasegawa S, Sato A, Iesato K, et al. Case report; a case of rheumatoid arthritis with renal amyloidosis and nephrotic syndrome effectively treated with tocilizumab [in Japanese]. *Nihon Naika Gakkai Zasshi*. 2011;100:185–187.
39. Hagihara K, Kawase I, Tanaka T, et al. Tocilizumab ameliorates clinical symptoms in polymyalgia rheumatica. *J Rheumatol*. 2010;37:1075–1076.
40. Gergis U, Arnason J, Yantiss R, et al. Effectiveness and safety of tocilizumab, an anti-interleukin-6 receptor monoclonal antibody, in a patient with refractory GI graft-versus-host disease. *J Clin Oncol*. 2010;28:e602–e4.
41. Nishida S, Hagihara K, Shima Y, et al. Rapid improvement of AA amyloidosis with humanised anti-interleukin 6 receptor antibody treatment. *Ann Rheum Dis*. 2009;68:1235–1236.
42. Nishimoto N, Nakahara H, Yoshio-Hoshino N, et al. Successful treatment of a patient with Takayasu arteritis using a humanized anti-interleukin-6 receptor antibody. *Arthritis Rheum*. 2008;58:1197–1200.
43. Seitz M, Reichenbach S, Bonel HM, et al. Rapid induction of remission in large vessel vasculitis by IL-6 blockade. A case series. *Swiss Med Wkly*. 2011;141:w13156.