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**RESEARCH ARTICLE** 

# Phytochemical Analysis of Canna indica L.

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### ABSTRACT

The medical plant *Canna indica* L. was selected for Isolation and Characterization of its medicinal value based on phytochemical studies. This plant was collected from Sacred grove forest of Aanaivari village, Rayavaram Panchayat Arimalam block, Pudukkottai District, Tamil Nadu, India during the month of February 2010. The collected plants and their parts were cleaned with tap water and dried under shade, then ground well to find powder. About 100g of dry leaf powder of *Canna indica L.* was extracted with solvent ethanol using Soxhlet apparatus at 60-70 degree temperature. Greenish black waxy residue was obtained. The residue was used for phytochemical and Characterization studies. Phytochemical constituent like alkaloids, flavonoids, carbohydrates, glycosides, phytosterols, fixed oil and fats, proteins, phenolic compounds, tannins and saponins of ethanol solvent extract of *Canna indica L.* were analysed qualitatively. In conclusion, these results suggest that *Canna indica L.* might be a source of large amount of proto alkaloids it is an antibiotic and antioxidant. Therefore, this result may suggest that *Canna indica L.*. extracts posses' compounds with antioxidant and antimicrobial properties which can be used as Phyto cancer agents in new drugs for therapy of diseases in human.

Key words : Canna indica L., phytochemicals, antioxidant, antimicrobial.

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# INTRODUCTION

The use of traditional medicine and medicinal plants in most developing countries, as a normative basis for the maintenance of good health, has been widely observed. Furthermore, an increasing reliance on the use of medicinal plants in the industrialized societies has been traced to the extraction & development of several drugs and chemotherapeutics from these plants as well as from traditionally used rural herbal remedies have become more popular in the treatment of minor ailments, & also on account of the increasing costs of personal health maintenance [1-20]. Indeed, the market & public demand has been so great that there is a great risk that many medicinal plants today, face either extinction or loss of genetic diversity. The World Health Organization (WHO) estimated that 80% of the population of developing countries relies on traditional medicines, mostly plant drugs, for their primary health care needs [21-30]. Also, modern pharmacopoeia still contains at least 25% drugs derived from plants and many others which are synthetic analogues built on prototypes compounds isolated from plants. Demand for medicinal plant is increasing in both developing and developed countries due to growing recognition of natural products, being non-narcotic, having no side effects, easily available at affordable prices and sometimes the only source of health care available to the poor. Medicinal plant sector has traditionally occupied an important position in the socio cultural, spiritual & medicinal arena of rural & tribal lives of India[31-40]. A number of compounds extracted from various species of higher plants have shown antiviral activity. Examples included tannins, flavones, alkaloids, that displayed in vitro against numerous viruses. It has been suggested that selection of plant on the basis of ethno medical considerations gives a higher hit rate than screening programmes of general synthetic products [41-45].

### Plant description

*Canna indica* L.(also known as saka siri, Indian shot, canna, bandera, chancle, coyol, or platanillo, Kardal in Marathi) is a species of the Canna genus, belonging to the family Cannaceae, a native of the Caribbean and tropical Americas that is also widely cultivated as a garden plant. It is a perennial growing from 0.5m to 2.5m, depending on the variety. It is hardy to zone 10 and is frost tender. In the northern latitudes it is in flower from August to October, and the seeds ripen in October. The flowers are hermaphrodite. The seeds are small, globular, black pellets, hard and heavy enough to sink in water. They resemble shotgun pellets giving rise to the plant's common name of Indian Shot.

They are widely used for jewellery. In the last three decades of the 20th century, Canna species have been categorised by two different taxonomists, Paulus Johannes Maria Maas from the Netherlands and Nobuyuki Tanaka from Japan. Maas regards *C. coccinea, C. compacta, C. discolor, C. patens* and *C. speciosa* as synonyms or varieties of *C. indica,* while Tanaka recognises several additional varieties of *C. indica. Canna indica* var. *indica* L.A medium sized species; green foliage, oblong shaped, spreading habit; triangular stems, coloured green; spikes of flowers are erect, self-coloured red, staminodes are long and narrow, edges regular, petals red, partial self-cleaning; fertile both ways, self-pollinating and also true to type, capsules globose; rhizomes are thick, up to 3 cm in diameter, coloured purple; tillering is prolific. Introduced by Linnaeus.[46].

#### Aims & Objectives

- To Identify and collect the Canna indica L. plant material with specific review of literature.
- To extract the secondary plant metabolites of the plant stem material of the species.
- To isolate, characterize and find out the bioactive metabolites through UV-VIS and FTIR analysis method.
- To correlate the identified metabolites with pharmacological applications.

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# MATERIALS AND METHODS

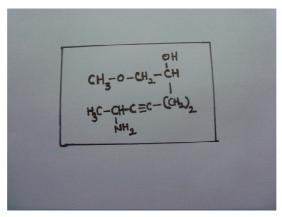
On the basis of its medicinal value which are available in the literature *Canna indica L*. was selected for phytochemical studies. The medicinal plant *Canna indica L*. was collected from Arimalam Village, Pudukkottai district during the month of January – 2010. The collected medicinal plant was brought into the TNSRO laboratory for phytochemical studies.

### Phytochemical analysis

About 100 g of dry leaf powder of was Canna indica L. extracted with ethanol at 60°c to 70°c by continuous hot percolation using soxhlet apparatus. The extraction was filtered and kept in over at 50°c for 24 hours to evaporate the extracts from them. A greenish black waxy residue was obtained. These extracts were used for phytochemical analysis qualitatively through UV-VIS and FT-IR. Phytochemical analysis for major phyto constituents of the plant extract was undertaken using standard qualitative methods as described by various authors (Vogel 1958, Van 1997, Williams 1949). The plant extracts were screened for the presence of biologically active compounds like alkaloids, flavonoids, glycosides, carbohydrates, phytosteriods and fatty acids, proteins, phenolics, tannins and saponins.[47-52]

## **RESULTS AND DISCUSSION**

Phytochemical constituents like alkaloids, flavoniods, carbohydrates, glycosides, phytosterols, fixed oil and fats, proteins, phenolic compounds, and saponins of *Canna indica* L. were analyzed by qualitatively and reported in Table - 1. On the basis of UV-Vis and FT-IR spectral analysis on *Canna indica* we have found the following data UV-Vis shown in figure 1 yielded 4 elevations (382.89nm, 1.6344 422.36,0.64656nm, 669.72,0.28462 and 980.66,0.079267) and the values were interpreted with table values and confirm the presence of **proto alkaloids** in the given sample. FT-IR result is yielded Maximum peak level 3930.13 cm<sup>-1</sup> and Minimum peak level 537.48 cm<sup>-1</sup>. So that the compound may be **proto alkaloids** The compound may be **9-amino 3,4 dihydroxy 2 methoxy non-6-yne** having molecular weight : GMM, structure is given below



9-amino 3,4 dihydroxy 2 methoxy non-6-yne Molecular weight: 171 GMM

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# CONCLUSION

From the above mentioned results this study reveals the presence of various alkaloids compounds including quoins, iso quoins, etc. in *Canna indica*. In the present study, UV-Vis and FT-IR procedure was applied for the identification of secondary metabolites. In the effort to study plant of *Canna indica* from the identified localities subjected to alkaloids screening. From this total samples, 78% gave positive result for proto alkaloids, from this 33% gave a positive reaction for iso quenolins. So the final study of the result is indicating the identified plant may be used for antimicrobial, anti helminthetic and anti-inflammatory agent in phyto-pharmaceutical applications.

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SI.No	Metabolites	Results
1	Alakloids	+++
2	Flavonoids	++
3	Terpenoids	++
4	Fixed oils	++
5	Phytosterols	+
6	Saponins	-
7	Phenolic compounds	++
8	Fats	++
9	Carbohydrates	++
10	Proteins	++
11	Glycosides	++
12	Tannins	+

### Table 1: Depicts various phytochemicals analyzed qualitatively

+++ present in high concentration, ++ present in medium concentration, + present in low concentration

- not present in the sample

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# REFERENCES

- 1. Albert L Sajem, (2006). Traditional use of medicinal plants by the Jaintia tribes in North Cachar Hills district of Assam, northeast India, *Journal of Ethnobiology Ethnomedicine*,5(2):85-88.
- 2. Atiqur Rahman M, (2007). "Medicinal Plants used by Chakma tribe in Hill Tracts District of Bangladesh. Indian Journal of Traditional Knowledge,9(3):122-125.
- 3. Ashish Ghosh,(2008). "Ethnomedicinal Plants used in West Rarrh region or West Bengal" Indian Journal of Traditional Knowledge- Natural Product Radiance, 7 (5):125-128.
- 4. Anon. (1986). *The useful plants of India. Publications & Information Directorate*, CSIR, New Delhi, India.1(1):27-33.
- 5. Beentje HJ, (1994). Kenya trees, shrubs and lianas. National Museums of Kenya. 1(1):27-33.
- 6. Coates-Palgrave K, (1988). Trees of southern Africa. C.S. Struik Publishers Cape Town. 1(1):18-33.
- 7. Chaisawadi,S,Thongbute,D.,(1990).Plants, Ethnopharmacology, 1:7-12.
- 8. Chevallier, (1996). Andrew. Encyclopedia of Medicinal Plants. New York: DK Publishing, Inc.,7(2):56-62.
- 9. Datta BK, (2006). "Medicinal plants prescribed by different tribal and non-tribal medicine in Tripura State". Indian Journal of Traditional Knowledge, 3(2):36-40.
- 10. .Dinesh Jadhav MP, (2006). "Ethnomedicinal Plants used by Bhil tribe of Bibdod" Indian Journal of Traditional Knowledge April, 3(2):24-56.
- 11. Dewan., and S.Bhakuni, (1995). Science reporter, CSIR, New Delhi.8(3):74-82.
- 12. David, (1998). Complete Guide to Chinese Herbal Medicine. New York: Berkeley Books, 1(1):156-160.
- 13. Dale IR, Greenway PJ, (1961). Kenya trees and shrubs. Buchanan's Kenya Estates Ltd, 1(2):80-88.
- 14. Delorit RJ, Gunn CR, (1986). Seeds of continental United States; Legumes (Fabaceae). Agronomy Publications, 1(2):78-82.
- 15. Duker-Eshun, G., Joroszewski, J.W., Asomaning, W.A., Oppong-Boachie, F. and Christensen, S.B., (2004). Antiplasmodial constituents of *Cajanus cajan*. Phytother. Res. 18 (2): 128-130.
- 16. Finar I.L, (1995). Organic Chemistry, Vol.II, Pearson Education, New Delhi, 1(2):88-95.
- 17. Hsieh, T. C., X. Lu, J. Guo, (2002). "Effects of Herbal Preparation Equiguard on Hormone-Responsive and Hormone-Refractory Prostate Carcinoma Cells: Mechanistic Studies. "*International Journal of Oncology*", 1(1):102-110.
- 18. Hilary A. Sandler, (2010). Managing Cuscuta gronovii (Swamp Dodder) in Cranberry
- 19. Requires an Integrated Approach, Sustainability, 1(3):660-683.
- 20. Hibberd J.M, (1998). Localization of photosynthetic metabolism in the parasitic angiosperm Cuscuta reflexa, *Planta*, 205: 506±513.
- 21. Hong TD, Linington S, Ellis RH, (1996). Seed storage behaviour: a compendium. Handbooks for Gene banks,4:37-45.
- 22. Harborne J.B,(1984). Phytochemical Methods, Second Edition, Newdelhi
- 23. Jian Bo Xiao, (2007). Chromatographia, 65:185-190.
- 24. Joffe, J, (2001). Creative Gardening with Indigenous Plants. A South African Guide. Briza, Pretoria. 1(2):56-62.
- 25. Katewa SS, (2008). "Poisionus Plants of the Southern Aravalli Hills of Rajasthan. Indian Journal of Traditional Knowledge,1(1):224-228.
- 26. Krishna, K.L., K. Mruthunjaya and J.A. Patel, (2009). Antioxidant and hepatoprotective activity of leaf extract of *Justicia gendarussa* burm. Int. J. Biol. Chemi; 3: 99-110.
- 27. Lawal I. O, (2010). Ethno medicinal information on collation and identification of some medicinal plants in Research Institutes of South-west Nigeria, African Journal of Pharmacy and Pharmacology , 4(1). 001-007.
- 28. Mahajan SK, (2007). "Traditional herbal remedies among the tribes Bijagarah of West Nimar Dist. MP. Indian Journal of Traditional Knowledge, 5(3):28-30.

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#### Antony Joseph Velanganni et al

- 29. Mahanta J, (2006). "Traditional Medicine in the Treatment of GIT Disease in Upper Assam" Indian Journal of Traditional Knowledge, 5(2):50-54.
- 30. Mukherjee KR, (2009). "Some abortifacient plants used by the tribal people of West Bengal" Indian Journal of Traditional Knowledge - Natural Product Radiance,8 (2):27:29.
- 31. Misra MK, (2006). "Medicinal Plants used by the Kandhar of Kandhamal" Dist Orissa, Indian Journal of Traditional Knowledge,5(2):58-61.
- 32. Montvale .NJ, (1998). PDR for Herbal Medicines. Medical Economics Company, New Delhi, 3(2):7-10.
- 33. Mohammed Rahmatullah, (2010). Effect of *cuscuta reflexa* stem and *calotropis procera* leaf extracts on Glucose tolerance in glucose-induced hyperglycemic rats and mice. *Afr. J. Trad. CAM* 7 (2): 109 112.
- 34. Noad T, Birnie A, (1989). Trees of Kenya. General Printers, Nairobi,1(2):174-178.
- 35. Pande PC, (2007). "Ethnovetrinary Plants of Uttaranchal", Indian Journal of Traditional Knowledge, 5(3):80-85.
- 36. Phytochemical studies on Cuscuta reflexa, (1999). H E J Research Institute of Chemistry, University of Karachi, Karachi, Pak. *Pakistan Journal of Scientific and Industrial Research*, 42(4), 170-172.
- 37. Patil Amol, (2009). In vitro free radicals scavenging activity of stems of Cuscuta reflexa, *Journal of Pharmacy Research*, 2 (1): 67-77
- 38. Pooley.E, (1993). A complete field guide to Trees of Natal, Zululand & Transkei.Natal Flora Publications Trust c/o Natal Herbarium. Durban, 1(1):98-102.
- 39. Palmer E, Pitman .N,(1972). Trees of Southern Africa, A.A. BalKema Cape Town, 1(1):55-58.
- 40. Schmidt . E, Lotter. M., McCleland. W, (2002). Trees and shrubs of Mpumalanga and Kruger National Park . Jacana. Johannesburg,1(2):178-183.
- 41. Saroj verma, (2007). "Indigenous Medicinal Plants Knowledge of Kunihar forest Division Solan District-HP". Indian Journal of Traditional Knowledge,7(3):88-97.
- 42. Sakshy Sharma, (2010). Comparative morpho-anatomical and preliminary phytochemical studies of *cuscuta reflexa*, *cassytha filiformis*,*I.J.of.Pharmacy and Phar.Sciences*, 2(1): 43-47.
- 43. Sajjad Khan1.M, (2009). Chromatographic estimation of maturity based phytochemical profiling of *Ipomoea mauritiana, International Journal of Phytomedicine*, 1(7):22-30.
- 44. SCORE (2010). Abstracts and Scientific papers, TNSRO, Arimalam, 1(1):45-55.
- 45. Teegaurden, Ron. (1998). The Ancient Wisdom of the Chinese Tonic Herbs. New York: Warner Books,2(2):78-80.
- 46. Tripathi S.M. Singh D.K. Molluscicidal, (2000). Activity of *Punica granatum* bark and *Canna indica* root, Brazilian Journal of Medical and Biological Research, 33: 1351-1355.
- 47. Vogel, A.I, (1958). A text book of practical organic chemistry Longman, London, 90-92.
- 48. Van Wyk, B., Gericke, N, (2000). People's plants: A guide to Useful Plants of Southern Africa. Briza, Pretoria,1(1):36-39.
- 49. Van Wyk, B, Van Wyk, P, (1997). Field guide to trees of Southern Africa . Struik, Cape Town, 1(2):27-35.
- 50. Vijikumar.S, (2010). Abstracts and Scientific papers of, SCORE, TNSRO, Arimalam, 1(1):30-39.
- 51. Venter F, Venter J-A, (1996). Making the most of Indigenous trees. Briza Publications, 1(1):122-125.
- 52. Williams R.O & OBE, (1949). The useful and ornamental plants in Zanzibar and Pemba. Zanzibar Protectorate,1(1): 88-99.