A3 adenosine receptor antagonists delay irreversible synaptic failure caused by oxygen and glucose deprivation in the rat CA1 hippocampus *in vitro*

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**Introduction**

Ischaemic episodes occurring in the mammalian central nervous system result in the impairment of neurotransmission and, with the duration of ischaemia, in increasingly severe tissue damage.

The impairment in neurotransmission is, however, not directly correlated with cell death and is reversible if the oxygen and glucose supply is restored within a narrow time window (Latini *et al.*, 1999; Pugliese *et al.*, 2003). While the disappearance of synaptic activity is the earliest detectable functional sign of tissue suffering, the absence of recovery after ischaemia interruption clearly indicates irreversible neurone damage.

Rapid anoxic depolarisation (AD) of a sizeable population of brain cells is observed with prolonged ischaemic episodes and its appearance is strictly correlated with neuronal and glial damage (see Somjen, 2001) during ischaemia, contributing also to the extension of cell damage to the so-called ‘ischaemic penumbral area’ (Touzani *et al.*, 2001). Therefore, it appears that pharmacological treatments directed to prevent or to delay AD would result in substantial neuroprotection (Obeidat & Andrew, 1998; Jarvis *et al.*, 2001; Somjen, 2001).

One of the early events occurring during oxygen and glucose deprivation (OGD) caused by an ischaemic episode is the release of substantial amounts of adenosine (Latini & Pedata, 2001) which, through the activation of specific receptors (see Fredholm *et al.*, 2001), is believed to exert important neuromodulatory effects relevant to the outcome of the ischaemic episode. Four subtypes of adenosine receptors, A1, A2A, A2B and A3, all coupled to the effector system through heterotrimeric G proteins, have been identified (Fredholm *et al.*, 2001) and suggest that selective A3 receptor block may substantially increase the resistance of the CA1 hippocampal region to ischaemic damage.

**Keywords:** Adenosine; A3 receptors; cerebral ischaemia; hippocampal slices; synaptic potential; anoxic depolarisation

**Abbreviations:** aCSF, artificial cerebral spinal fluid; AD, anoxic depolarisation; DMSO, dimethylsulphoxide; epsps, field excitatory post synaptic potential; MRS 1523, N-[9-chloro-2-(2-furanyl)[1,2,4]-triazolo[1,5-c]quinazolin-5-yl]benzeneacetamide; MRS 1220, N-(2-methoxyphenyl)-N’-[2-(3-pyrindinyl)-4-quinazolyl]-urea, (VUF 5574, 100nM, n = 3) and 5-[[4-pyridyl]amino]carbonyl][aminoo-8-methyl-2-(2-furyl)-pyrazolo[4,3-e]1,2,4-triazolo[1,5-c]pyrimidine hydrochloride (1 nM, n = 4), prevented the irreversible failure of neurotransmission induced by 7 min OGD (n = 45) and the development of AD in 20 out of 22 monitored slices.

When tested on OGD episodes of longer duration (8–10 min, n = 18), 100 nM MRS 1523 prevented or delayed the appearance of AD and exerted a protective effect on neurotransmission for episodes of up to 9 min duration. In the absence of AD, the epsps recovery was almost total, regardless of OGD episode duration. These findings support the notion that A3 receptor stimulation is deleterious during ischaemia and suggest that selective A3 receptor block may substantially increase the resistance of the CA1 hippocampal region to ischaemic damage.

The selective adenosine A3 antagonists 3-propyl-6-ethyl-5[(ethylthio)carbonyl]-2-phenyl-4-propyl-3-pyridinecarboxylate [MRS 1523, 1–100 nM, n = 31], N-[9-chloro-2-(2-furanyl)[1,2,4]-triazolo[1,5-c]quinazolin-5-yl]benzeneacetamide (MRS 1220, 100 nM, n = 7), N-(2-methoxyphenyl)-N’-[2-(3-pyrindinyl)-4-quinazolyl]-urea, (VUF 5574, 100 nM, n = 3) and 5-[[4-pyridyl]amino]carbonyl][aminoo-8-methyl-2-(2-furyl)-pyrazolo[4,3-e]1,2,4-triazolo[1,5-c]pyrimidine hydrochloride (1 nM, n = 4), prevented the irreversible failure of neurotransmission induced by 7 min OGD (n = 45) and the development of AD in 20 out of 22 monitored slices.

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et al., 2001) and are expressed in the brain (Dixon et al., 1996). Converging experimental data support the notion that activation of adenosine A₁ receptors during ischemic episodes results in neuroprotection of the brain tissue (see Abbracchio & Cattabeni, 1999; Phillis & Goshgarian, 2001) predominantly by reducing excitatory (glutamatergic) transmission. The role of the other adenosine receptors in cerebral ischemia, and in particular of A₃ receptors, is still controversial (see Von Lubitz, 1999). In fact, it has been demonstrated that mice lacking the A₃ adenosine receptors show an increase in neurodegeneration in response to repeated episodes of hypoxia (Fedorova et al., 2003). Consistent with these reports, Hentschel et al. (2003) demonstrate, in rat cortical neurons, that the selective activation of A₃ adenosine receptors during hypoxia is involved in the inhibition of excitatory neurotransmission indicating that the A₃ receptors also contribute to neuroprotective action of adenosine brought about by A₁ receptors. Similarly, at a cardiac level, most evidence indicates that A₂ receptors are involved in protection of the ischemic heart (Fredholm et al., 2005). On the other hand, acute A₃ receptor stimulation has been shown to exacerbate the damage caused by a concomitant ischemic episode in vivo (Von Lubitz et al., 1994), indicating a deleterious role of these adenosine receptors during cerebral ischemia. The observation that A₃ receptor activation during OGD limits the beneficial effects of ischemic preconditioning on the resistance of synaptic transmission to ischemia-like insults in hippocampal slices (Pugliese et al., 2003) is consistent with this hypothesis.

The role of A₃ receptors in the brain under normoxic conditions appears to be equally controversial. In fact, in different brain regions in vitro, selective A₃ receptor stimulation induces either an inhibitory (Brand et al., 2001) or a facilitatory effect (Dunwiddie et al., 1997; Fleming & Mogul, 1997; Macek et al., 1998; Costenla et al., 2001; Laudadio & Psarropoulos, 2004) on excitatory neurotransmission. These opposite effects may sustain a protective or a deleterious role of A₃ receptors, respectively, during ischemia. Therefore, the overall modulatory effects of A₃ receptors on neurotransmission during cerebral ischemia are not well defined.

In the present work, we used selective A₃ receptor antagonists to investigate the role of A₃ adenosine receptors on synaptic transmission during severe (7 min or longer duration) OGD episodes aimed at reproducing in vitro the consequences of interruption of blood flow following cardiac arrest or occlusion of intracranial vessels. A preliminary account of this work has been communicated (Pugliese et al., 2004).

Methods

All animal procedures were carried out according to the European Community Guidelines for Animal Care, DL 116/92, application of the European Communities Council Directive (86/609/EEC). Experiments were carried out on rat hippocampal slices, prepared as previously described (Pugliese et al., 2003).

Slice preparation

Male Wistar rats (Harlan, Italy; Udine, Italy, 150–200 g body weight) were deeply anaesthetised with ether and decapitated with a guillotine. The hippocampi were rapidly removed and placed in ice-cold oxygenated (95% O₂–5% CO₂) artificial cerebral spinal fluid (aCSF) of the following composition (mM): NaCl 124, KCl 3.33, KH₂PO₄ 1.25, MgSO₄ 2, CaCl₂ 2, NaHCO₃ 25 and d-glucose 10. Slices (400 μm nominal thickness) were cut with a McIlwain tissue chopper (The Mickle Lab. Engineering, Co. Ltd, Gomshall, U.K.) and kept in oxygenated aCSF for at least 1 h at room temperature. A single slice was then placed on a nylon mesh, completely submerged in a small chamber (0.8 ml) and superfused with oxygenated aCSF (30–32°C) at a constant flow rate of 2 ml min⁻¹. The treated solutions reached the preparation in 90 s and this delay was taken into account in our calculations.

Extracellular recording

Test pulses (80 μs, 0.066 Hz) were delivered through a bipolar nichrome electrode positioned in the stratum radiatum. Evoked potentials were recorded with glass microelectrodes (2–10 MΩ, Clark Electromedical Instruments, Pangbourne, U.K.) filled with 150 mM NaCl, and placed in the CA1 region of the stratum radiatum. The negative shifts were recorded in direct current (d.c.) mode. Responses were amplified (BM 622, Mangoni, Pisa, Italy), digitised (sample rate, 33.33 kHz), low-pass filtered (10 kHz), and stored for later analysis using LTP software facilities (version 2.30D, Anderson & Collingridge, 2001, www.ltp-program.com). Stimulus–response curves were obtained by gradual increases in stimulus strength at the beginning of each experiment, when a stable baseline of evoked response was reached. The test stimulus pulse was then adjusted to produce field excitatory postsynaptic potentials (fepsps) whose amplitude was 40–50% of the maximum and was kept constant throughout the experiment. The amplitude of fepsps was routinely measured and expressed as the percentage of the average amplitude of the potentials measured during the 5 min preceding exposure of the hippocampal slices to in vitro ischemia. In all the experiments, both the amplitude and initial slope of fepsps were measured, but since no appreciable differences between these two parameters were observed in the effect of drugs and of in vitro ischemia, only the measure of the amplitude was expressed in the figures. The amplitude of AD was measured as the integral of tissue depolarisation in the first 2 min after AD peak.

Application of drugs and OGD

In vitro OGD was obtained by perfusing the slice for 7 min with glucose-free aCSF gassed with nitrogen (95% N₂–5% CO₂). At the end of the ischemic period, slices were superfused with normal, glucose-containing, oxygenated aCSF.

All the A₃ adenosine receptor antagonists used in our experiments were applied 10 or 20 min before, during OGD and 5 min after the end of ischemic episode. On each experimental day, data were obtained either in the absence or in the presence of the adenosine antagonists, in slices taken from the same rat. The concentration used for each of the selective adenosine A₃ receptor antagonists was chosen on the basis of Kᵢ values on rat or human A₃ receptors. 3-Propyl-6-ethyl-5[(ethylthio) carbonyl]-2-phenyl-4-propyl-3-pyridinecarboxylate
(MRS 1523), among all A3 antagonists used in our experiments, is the most potent and selective antagonist for the rat A3 receptors actually available in commerce (Ki value of 113 nM; Li et al., 1998; Muller, 2003).

**Drugs**

MRS 1523 was purchased from Sigma (Milano, Italy). MRS 1220 (N-[9-chloro-2-(2-furanyl)[1,2,4]-triazolo[1,5-c]quinazolin-5-yl]benzene acetamide) and VUF 5574 (N-(2-methoxyphenyl)-N-[2-(3-pyridinyl)-4-quinazolyl]urea) were from Tocris (Bristol, U.K.). MRS 1523, MRS 1220 and VUF 5574 were dissolved in dimethylsulphoxide (DMSO) and stock solutions were made to obtain concentrations in DMSO of 0.05 and 0.01% in aCSF, respectively. Control experiments, carried out in parallel, showed that this concentration of DMSO did not affect either fepsp amplitude before OGD or the depression of synaptic potential induced by the following OGD. The hydrophilic A3 antagonist 5-[(4-pyridyl)amino]-carbonyl amino-8-methyl-2-(2-furyl)-pyrazolo[4,3-e]1,2,4-triazolo[1,5-c]pyrimidine hydrochloride (WSAB) was provided by Dr G. Spalluto.

**Statistical analysis**

Data were analysed using Prism 3.02 software (Graphpad Software, San Diego, CA, U.S.A.). All numerical data are expressed as the mean ± s.e. Data were tested for statistical significance with two-tailed Mann–Whitney test or by analysis of variance (one-way ANOVA), as appropriate. When significant differences were observed, the Newman–Keuls multiple comparison test (one-way ANOVA) was inferred. A value of P<0.05 was considered significant.

**Results**

The role of A3 adenosine receptor stimulation by endogenous adenosine released during in vitro severe ischaemia-like episodes on synaptic transmission was investigated using selective A3 adenosine receptor antagonists. Electrically evoked fepsp were extracellularly recorded in the CA1 region of 124 hippocampal slices taken from 55 rats for monitoring the time course of the effects of OGD episodes of different duration on synaptic responses, both in control and treated slices. In a subset of experiments (n=53), the d.c. shift produced by AD was simultaneously recorded.

Selective block of A3 adenosine receptors prevents the irreversible impairment of neurotransmission induced by 7 min OGD

In a first series of experiments, we characterised the response of synaptic excitatory transmission to 7 min OGD, an ischaemia-like insult that in our experimental conditions has been shown to consistently produce an irreversible loss of synaptic transmission, but to be sensitive to the protective effects of ischaemic preconditioning (Pugliese et al., 2003).

Figure 1a illustrates the effects of 7 min OGD on the amplitude of the synaptic responses evoked by stimulation of the CA1 stratum radiatum and recorded from the apical dendrite region of pyramidal cells. One 7 min OGD episode induced the disappearance of fepsp, which did not recover after prolonged superfusion with oxygenated, glucose-containing aCSF (up to 80 min, n=7; data not shown). The effect of 7 min OGD on fepsp was similar in 42 slices examined and the mean recovery of fepsp amplitude, after 7 min OGD episode, was 5±1%, n=42 (see also Figure 3).

The presence of selective adenosine A3 receptor antagonists prevented the irreversible disappearance of synaptic potentials...
induced by 7 min OGD. Thus, in the presence of MRS 1523 (100 nM, Figure 1b), a total recovery of synaptic response was observed within 10 min from OGD interruption. The mean recovery of fepsp amplitude after 7 min OGD in the presence of 100 nM MRS 1523 was 83 ± 6%, (n = 17, Figure 3a). Furthermore, if compared with those obtained in control conditions, in MRS 1523-treated slices the transient recovery of fepsp amplitude was delayed and the afferent fibre volley did not disappear at the end of 7-min OGD (Figure 1b, trace 2, see also Table 1).

In a subset of experiments, we investigated the effects of 100 nM MRS 1523 on AD by comparing the time of peak and the magnitude of depolarising d.c. shifts caused by 7-min OGD in treated slices and in matched control slices from the same rats. As illustrated in Figure 2a, in control conditions, 7-min OGD episodes always caused AD, recorded as negative d.c.

### Table 1

<table>
<thead>
<tr>
<th>Treatment with MRS 1523 produces a delay in the effects of OGD in the CA1 region of hippocampal slices</th>
<th>(n)</th>
<th>Control</th>
<th>(n)</th>
<th>MRS 1523</th>
</tr>
</thead>
<tbody>
<tr>
<td>Initial fepsp disappearance time (s)</td>
<td>(50/50)</td>
<td>177.9 ± 7.2</td>
<td>(41/41)</td>
<td>254.3 ± 10.9</td>
</tr>
<tr>
<td>Transient fepsp recovery peak time (s)</td>
<td>(31/50)</td>
<td>357 ± 17</td>
<td>(34/41)</td>
<td>480 ± 18</td>
</tr>
<tr>
<td>Transient fepsp recovery duration (s)</td>
<td>(31/50)</td>
<td>59 ± 7</td>
<td>(34/41)</td>
<td>46 ± 7</td>
</tr>
<tr>
<td>Transient fepsp recovery amplitude (%)</td>
<td>(31/50)</td>
<td>18.0 ± 3.3</td>
<td>(34/41)</td>
<td>15.5 ± 2.4</td>
</tr>
<tr>
<td>Fibre volley disappearance time (s)</td>
<td>(34/50)</td>
<td>372 ± 18</td>
<td>(19/41)</td>
<td>501 ± 26</td>
</tr>
<tr>
<td>AD peak time (s)</td>
<td>(20/20)</td>
<td>436 ± 17</td>
<td>(22/33)</td>
<td>520 ± 23</td>
</tr>
</tbody>
</table>

Data are from slices receiving 7 min or 30 min OGD in control (n = 50) and 7, 8, 9, 10 or 30 min OGD in the presence of 100 nM MRS 1523 (n = 41). Numbers in parentheses (n/n) indicate number of observations out of investigated slices. Time is calculated from OGD initiation. The amplitude of the transient fepsp recovery is expressed as per cent of baseline fepsp recorded before OGD application. Statistical significance was assessed by Mann–Whitney test.

* Fibre volley did not disappear in any of the slices receiving 7 min OGD in the presence of MRS 1523.

*AD was absent in 11 slices receiving 7–8 min OGD in the presence of MRS 1523.

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**Figure 2** The A₃ adenosine receptor antagonist MRS 1523 minimises AD and protects CA1 hippocampus from irreversible fepsp depression induced by 7-min OGD. (a) AD was recorded as the negative d.c. shift in response to 7-min OGD (solid bars) in control conditions (n = 8) and in the presence of 100 nM MRS 1523 (open bar, n = 8). MRS 1523 significantly prevented AD in seven out of eight slices. (b) Graph shows the time course of 7-min OGD effect on fepsp amplitude, expressed as per cent of baseline, in control aCSF (filled circles; mean ± s.e., n = 8) and in the presence of 100 nM MRS 1523 (unfilled circles, mean ± s.e., n = 8). Note that in the presence of the A₃ antagonist, the time course of fepsp depression during OGD was significantly delayed (see Table 1) in comparison to corresponding times in the absence of the drug (control).
shifts, with a mean peak latency of about 6.5 min (390 ± 20 s) from the beginning of ischaemia and a peak amplitude of 8.9 ± 0.6 mV (n = 8). The duration of d.c. shifts was variable (range 5–15 min) and was always accompanied by complete and irreversible disappearance of fepsps. In the presence of MRS 1523 (100 nM, n = 8), AD was virtually absent in seven out of eight preparations and, as shown in Figure 2b, the mean recovery of fepsps amplitude was significantly greater than in control slices taken from the same rats (85.8 ± 13.3%, n = 8 vs 33.2 ± 2.5%; P < 0.001). Interestingly, in one experiment where a sizeable AD (peak: −7.6 mV) was recorded, the recovery from OGD-evoked impairment in neurotransmission was only 22%.

The protective effect of MRS 1523 on OGD-evoked irreversible depression of fepsps was detectable at concentrations as low as 0.1 nM and the recovery of fepsps amplitude became statistically significant with concentrations of 1–100 nM of the antagonist (Figure 3a). The apparent EC50 value for MRS 1523 was 0.25 nM (95% CI 0.05–1.2 nM). However, since within the time of the antagonist application (10–20 min) it might have not reached equilibrium at the receptor level in slices, the EC50 value is likely to be underestimated. Similar beneficial effects on the recovery of fepsps from 7-min OGD were exerted by the selective adenosine A3 receptor antagonists MRS 1220, VUF 5574 and WSAB, all chemically different from MRS 1523 (Kim et al., 1996; van Muijlwijk-Koezen et al., 2000a; Maconi et al., 2002). As shown in Figure 3a, all A3 antagonists prevented synaptic impairment and allowed for complete synaptic recovery within 15 min from OGD interruption. In addition, all the A3 antagonists tested prevented or significantly delayed AD after 7-min OGD (data not shown).

The overall effect of A3 antagonism on fepsps recovery after 7-min OGD, compared with that observed in control preparations, is summarised in Figure 3b. As illustrated in the frequency histogram, 39 out of 45 slices treated with any of the A3 antagonists at effective concentrations (all except 0.1 nM MRS 1523), had a substantial (>50%) recovery of fepsps amplitude after 7-min OGD, while synaptic activity in controls (n = 42) never recovered beyond 20% of responses recorded before OGD.

**Figure 3.** Effects of selective A3 adenosine receptor antagonists on recovery of fepsps amplitude after 7-min OGD. (a) Column bars indicate the average recovery (mean ± s.e.) of fepsps after 7-min OGD, recorded in hippocampal slices at 15 min reperfusion in normal, oxygenated ACSF. n indicates the number of slices tested and asterisks indicate P < 0.05, one-way ANOVA, Newman–Keuls multiple comparison post hoc test, vs control and 0.1 nM MRS 1523-treated slices. (b) Distribution analysis of fepsps recovery in 87 slices receiving 7-min OGD episodes either in control or in the presence of A3 receptor antagonists (see a, 0.1 nM MRS 1523 excluded). Bars indicate the number of cells (ordinate) that showed a given recovery of fepsps (abscissa) from 7-min OGD episodes in control ACSF (n = 42; white bars) or in the presence of A3 antagonists (n = 45; black bars). Note that the large majority of treated slices (38 out of 45) show more than 50% recovery of fepsps.

Selective block of A3 adenosine receptors delays the effects of prolonged OGD application on synaptic transmission

In the experiments performed with 7-min OGD, the inhibitory effect of A3 antagonists on the development of AD seemed to be in relation to the significant recovery of fepsps amplitude induced by these drugs.

Furthermore, as illustrated in Figures 1 and 2, OGD episodes produced a typical sequence of neurophysiological effects that followed the initial disappearance of fepsps and that comprised a transient recovery of fepsps response, the disappearance of the afferent fibre volley and the development of an AD. All these phenomena were significantly delayed and/or reduced in the presence of A3 receptor antagonists, as shown in Table 1, where the effects of 100 nM MRS 1523 on OGD of different duration are summarised.

The possible correlation of these effects of A3 receptor antagonists with the recovery of the fepsps after OGD interruption and the time window in which A3 receptor block may play a role in limiting the deleterious effects of severe ischaemia were investigated by applying OGD episodes longer than 7 min.

To delimit the time window in which A3 antagonists could delay the appearance of AD, we applied 30-min OGD in the presence of 100 nM MRS 1523 (Figure 4a). Compared to the effects observed in matched control slices, the A3 receptor antagonist significantly increased the latency of AD peak from 7.28 min (437 ± 31 s) in control to 9.5 min (571 ± 70 s) in MRS 1523 (P < 0.01, Mann–Whitney test, two-tailed), without significantly affecting either the average magnitude (Figure 4) or the peak amplitude of AD (6.2 ± 1.2 mV, n = 6 vs 7.3 ± 0.3 mV, n = 8 in control, P = 0.49, two-tailed Mann–Whitney test). No recovery of fepsps was recorded after interruption of 30-min OGD (not shown).

In a final set of experiments, we monitored the recovery of fepsps while recording AD latency and amplitude following OGDs of different duration (8–10 min) in the presence of 100 nM MRS 1523.

As summarised in Figure 4b, a significant recovery of fepsps amplitude was obtained after OGD insults of duration up to

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**Table 1.** Effects of MRS 1523 on OGD-evoked synaptic depression.

<table>
<thead>
<tr>
<th>Concentration</th>
<th>n</th>
<th>Recovery of fepsps amplitude at 15 min post OGD (mean ± s.e.)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>8</td>
<td>33.2 ± 2.5%</td>
</tr>
<tr>
<td>0.1 nM MRS 1523</td>
<td>8</td>
<td>85.8 ± 13.3%</td>
</tr>
<tr>
<td>1 nM MRS 1523</td>
<td>8</td>
<td>77.8 ± 10.2%</td>
</tr>
<tr>
<td>10 nM MRS 1523</td>
<td>8</td>
<td>77.7 ± 9.6%</td>
</tr>
<tr>
<td>200 nM MRS 1523</td>
<td>8</td>
<td>77.7 ± 9.6%</td>
</tr>
</tbody>
</table>

**Figure 1.** The protective effect of MRS 1523 on OGD-evoked irreversible synaptic failure. Shifts in AD latency and amplitude (mean ± s.e.) following 7-min OGD interruption (n = 8), show a statistically significant (t-test, vs control and 0.1 nM MRS 1523-treated slices) delay of AD latency (P < 0.01, t-test, two-tailed) and a significant (t-test, vs control and 0.1 nM MRS 1523-treated slices) reduction of AD amplitude (P < 0.05, t-test, two-tailed). All these phenomena were significantly delayed and/or reduced in the presence of A3 receptor antagonists, as shown in Table 1, where the effects of 100 nM MRS 1523 on OGD of different duration are summarised.

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**Figure 2.** Effects of selective A3 adenosine receptor antagonists on synaptic depression in response to 7-min OGD. (a) Comparison of the latency and amplitude (mean ± s.e.) of AD following 7-min OGD interruption (n = 8), shows a statistically significant (t-test, vs control and 0.1 nM MRS 1523-treated slices) delay of AD latency (P < 0.01, t-test, two-tailed) and a significant (t-test, vs control and 0.1 nM MRS 1523-treated slices) reduction of AD amplitude (P < 0.05, t-test, two-tailed). All these phenomena were significantly delayed and/or reduced in the presence of A3 receptor antagonists, as shown in Table 1, where the effects of 100 nM MRS 1523 on OGD of different duration are summarised.

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**Figure 3a.** The protective effect of MRS 1523 on OGD-evoked irreversible synaptic failure. Shifts in AD latency and amplitude (mean ± s.e.) following 7-min OGD interruption (n = 8), show a statistically significant (t-test, vs control and 0.1 nM MRS 1523-treated slices) delay of AD latency (P < 0.01, t-test, two-tailed) and a significant (t-test, vs control and 0.1 nM MRS 1523-treated slices) reduction of AD amplitude (P < 0.05, t-test, two-tailed). All these phenomena were significantly delayed and/or reduced in the presence of A3 receptor antagonists, as shown in Table 1, where the effects of 100 nM MRS 1523 on OGD of different duration are summarised.

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**Figure 3b.** The protective effect of MRS 1523 on OGD-evoked irreversible synaptic failure. Shifts in AD latency and amplitude (mean ± s.e.) following 7-min OGD interruption (n = 8), show a statistically significant (t-test, vs control and 0.1 nM MRS 1523-treated slices) delay of AD latency (P < 0.01, t-test, two-tailed) and a significant (t-test, vs control and 0.1 nM MRS 1523-treated slices) reduction of AD amplitude (P < 0.05, t-test, two-tailed). All these phenomena were significantly delayed and/or reduced in the presence of A3 receptor antagonists, as shown in Table 1, where the effects of 100 nM MRS 1523 on OGD of different duration are summarised.
9 min. No significant recovery was observed in five slices after 10 min OGD. Analysing the fepsps recovery in relation to AD appearance after OGD, it appeared that the degree of fepsps recovery after OGD in A3 antagonist-treated slices depended on the appearance of AD and not on the duration of OGD episodes. In fact, in the absence of AD a full recovery of fepsps was found in most preparations receiving 7 or 8 min OGD episodes. In one slice after 9 min OGD.

Interestingly, treatment with MRS 1523 allowed for a significantly better recovery of neurotransmission also in those preparations in which AD peaked after interruption of the OGD episode. In our experiments, the pretreatment of hippocampal slices with selective A3 adenosine receptor antagonists considerably delays the occurrence of AD and significantly protects from the irreversible disruption of excitatory neurotransmission caused by 7-min OGD episodes. A3 receptor antagonists exert a protective effect on OGD episodes of ≤ 9 min duration, showing that other cellular mechanisms are implicated and predominate when AD takes place.

We used four selective A3 adenosine receptor antagonists with different chemical structure and lipid solubility (Kim et al., 1996; Jacobson et al., 1997; Li et al., 1998; van Muijlwijk-Koezen et al., 2000; Maconi et al., 2002; Muller, 2003) to assess the role of A3 receptor stimulation by endogenous adenosine released during OGD episodes. All compounds show similar protective effects on the 7 min OGD at nm concentration, ensuring specific involvement of A3 receptors in the observed effects.

MRS 1220, VUF 5574 and WSAB were reported to have higher affinity for the human A3 receptors (Kim et al., 1996; Jacobson et al., 1997; van Muijlwijk-Koezen et al., 2000; Maconi et al., 2002) and their effectiveness on native rat A3 receptors at very low concentration may be surprising. We do not have any straightforward explanation for this phenomenon. The tissue accumulation of the lipophilic MRS 1523, MRS 1220 and VUF 5574 is an unlikely explanation since WSAB is a hydrophilic compound (Maconi et al., 2002). On the other hand, receptor affinity of these compounds may also depend

**Discussion**

The main finding of the present work is that the selective antagonism of A3 adenosine receptors reduces the deleterious effect induced by OGD on CA1 hippocampal neurotransmission and pyramidal cell survival.

In our experiments, the pretreatment of hippocampal slices with selective A3 adenosine receptor antagonists considerably delays the occurrence of AD and significantly protects from the irreversible disruption of excitatory neurotransmission caused by 7-min OGD episodes. A3 receptor antagonists exert a protective effect on OGD episodes of ≤ 9 min duration, showing that other cellular mechanisms are implicated and predominate when AD takes place.

The main finding of the present work is that the selective antagonism of A3 adenosine receptors reduces the deleterious effect induced by OGD on CA1 hippocampal neurotransmission and pyramidal cell survival.
on specific conformations assumed by $A_3$ receptors in the integer cell membrane and may differ from that assessed in receptor binding experiments on disrupted membranes. Alternatively, the paucity of $A_3$ receptors in native tissue (Ji et al., 1994; von Lubitz, 1999) allows the speculation that occupancy of a substantial fraction of $A_3$ receptors is required for evoking cell response(s). In this case, the block of a relatively small fraction of $A_3$ receptors may be sufficient to greatly antagonise the effects of endogenous adenosine released during OGD (see also below).

**Role of adenosine receptors in depression and disruption of CA1 hippocampal excitatory synaptic transmission during in vitro ischaemia**

The depression of synaptic responses caused by OGD episodes of short duration (up to 5 min) in the CA1 region of the hippocampus is fully reversible (Latini et al., 1999; Pugliese et al., 2003). In contrast, the application of 7-min OGD elicits a complete and irreversible block of hippocampal neurotransmission, that persists upon slice reperfusion with normally oxygenated and glucose-containing aCSF (Pugliese et al., 2003).

A constant sequence of changes in neurotransmission occurs during the application of long OGD episodes and comprises (i) the early depression of evoked fepsps, (ii) a transient recovery of fepsps followed by (iii) the disappearance of synaptic responses and afferent fibre volley and (iv) AD. The whole sequence of events lasts about 6 min and therefore is typically recorded within the application of 7-min OGD.

Our results show that the block of $A_3$ receptor-mediated effects by selective antagonists results in a significant delay of the sequence of electrophysiological changes, including disappearance of the afferent fibre volley and AD that are considered early electrophysiological signs of tissue suffering.

The earliest event observed during OGD was the disappearance of the electrically evoked fepsps that reflect the currents generated by the inflow of cations into CA1 pyramidal cell dendrites produced by activation of synaptic glutamate receptors. The predominant, although not exclusive, mechanism that accounts for the reduction in fepsps during the first 4–5 min of ischaemia is a decrease in glutamatergic neurotransmission caused by activation of adenosine $A_1$ presynaptic receptors (Fowler, 1990; Gribkoff et al., 1990; Pedata et al., 1993; Latini et al., 1999).

$A_3$ receptor stimulation per se does not produce any harmful effects in normoxic tissue. In fact, neither $A_3$ receptor agonists nor adenosine disrupt CA1 neurotransmission in normally oxygenated slices (Dunwiddie et al., 1997). This implies that the main role of $A_3$ receptor activation during OGD is to hasten the processes that lead to AD and that removal of these mechanisms prolongs the period of tissue survival to OGD, but cannot fully block the consequences of ischaemia.

In order to explain the mechanism by which $A_3$ receptors may be contributing to failure of synaptic transmission during OGD, we can postulate that stimulation of $A_3$ receptors by adenosine released during prolonged and severe ischaemia may enhance excitatory transmission on CA1 pyramidal neurones, accounting for increased neuronal excitability and consequent lack of protection of the ischaemic tissue. Stimulation of $A_3$ receptors in the hippocampus may in fact: (i) counteract the inhibitory action of $A_1$ adenosine receptors on excitatory neurotransmission (Dunwiddie et al., 1997); (ii) inhibit the presynaptic metabotropic glutamate receptor inhibitory function on excitatory transmission (Macek et al., 1998). Furthermore, $A_1$ receptor-mediated stimulation of phospholipase C could contribute to neuronal damage through mobilisation of intracellular calcium (Abbracchio et al., 1995) and/or activation of PKC, resulting in an increase in excitability of CA1 neurones (Hu et al., 1987).

Consistently, our data show that the action of $A_3$ receptor antagonists is limited to a time window that extends survival to about 9 min of OGD. This time window seems to be related to the maximal delay allowed for the appearance of AD in the absence of $A_3$ receptor stimulation.

The generation of AD is complex and multifactorial (see Somjen, 2001) and the mechanisms responsible for the delay in AD remain elusive. Interestingly, the time window of $A_3$ receptor-mediated effects overlaps with the delay that can be obtained by treating the slices with glutamate receptor antagonists (Tanaka et al., 1997; Yamamoto et al., 1997; see also in Somjen, 2001). It is appealing to suggest that removal of the $A_3$ receptor-mediated impairment of the feedback inhibition of glutamate release exerted by specific metabotropic glutamate receptor subtypes (Macek et al., 1998) may substantially decrease/delay the participation of the excitatory neurotransmitter in triggering the AD.

Our results are in contrast with those obtained in transgenic mice with a deletion of the $A_3$ receptor. It has been demonstrated that after repeated brief exposure to carbon monoxide, mice lacking the $A_3$ receptors are more vulnerable than control animals to hippocampal damage following hypoxia (Fedorova et al., 2003), suggesting a neuroprotective role of $A_3$ receptors. The discrepancy about the functional role of $A_3$ receptors in the brain during hypoxia or ischaemia could be due to the diversity of both the experimental conditions (hypoxia/ischaemia) and pharmacological profiles of these receptors across species.

The delay of the initial depression of fepsps caused by $A_3$ antagonists indicates that $A_3$ receptors are activated within the first 2 min of OGD despite the reported low affinity of adenosine for these receptors (about 5 $\mu M$: Zhou et al., 1992). However, as the estimated concentration of adenosine at the receptor level approaches 5 $\mu M$ at the second minute of OGD and reaches 30–40 $\mu M$ within the fifth minute of OGD (Latini et al., 1999), a substantial activation of $A_3$ receptors may be achieved since the beginning of ischaemia. According to the Hill–Langmuir equation, implemented with the above reported values, the estimated occupancy of $A_3$ receptors by endogenous adenosine would approach 50% within the second minute of OGD and be almost 90% at 5 min of OGD.

It is therefore conceivable that all the changes in neurophysiological parameters observed with $A_3$ receptor antagonists result from block of cell mechanisms activated by $A_3$ receptors and that their stimulation by adenosine released during OGD contributes to the development of ischaemia effects from the beginning of OGD, thus hastening the deleterious effects of ischaemia on neurotransmission.

A modest, transient, recovery of fepsps was observed in most preparations and fading of the recovery was accompanied by disappearance of the presynaptic fibre volley. This sequence of events has been ascribed to progressive
increase in the extracellular K⁺ concentration that initially produces hyperexcitability of pyramidal cells followed by a depolarisation block of neurotransmission when extracellular K⁺ reaches 10 mM or higher concentration (Sick et al., 1987). After fibre volley disappearance and in the absence of any synaptic response, the large efflux of potassium into the extracellular space combined with activation of sodium and calcium channels, triggers sustained depolarisation of hippocampal cells that coincides with AD recorded in the CA1 region. Although similar to the spreading depression described by Leao (1951) and known to be harmless to the cerebral cortex under normoxic conditions, AD has been suggested to contribute to cell damage during ischaemia (see Somjen, 2001). Increased intracellular calcium and/or massive glutamate receptor activation are additional mechanisms that concur with potassium redistribution to produce AD (Tanaka et al., 1997; Yamamoto et al., 1997) and have been suggested to contribute to cell damage during ischaemia (see Somjen, 2001).

Therapeutic implications

In the brain, spreading depression is a phenomenon characterised by a slow transient cellular depolarisation moving at 3–4 mm min⁻¹ over the surface of the cortex (Leao, 1951). A large efflux of potassium into the extracellular space coincides with the shift in the d.c. potential. The changes in brain homeostasis are transient and do not cause visible injury in normoxic conditions (Hansen & Nedergaard, 1988), but are correlated with tissue damage during ischaemia (see Somjen, 2001). Within 2 min of stroke onset, neurons and glia suddenly depolarise in the brain area where cerebral blood flow falls to 10% of control (Macdonald & Stoodley, 1998). In these conditions, the generation of AD may contribute to the extent and severity of neuronal damage. In particular, the propagation of the AD to the hypoxic/hypoglycemic region (penumbral area) surrounding the ischaemic core may extend the damage. Consistently, it has been demonstrated that one major factor contributing to neuronal death in the penumbra is the propagation of spreading depression waves (Koroleva & Bures, 1996). Because the penumbra constitutes potentially salvageable tissue, the molecular responses of the perifocal neurons to focal ischaemia and AD are of interest (Obeidat et al., 2000).

In our experiments, a substantial field depolarisation was recorded for several minutes (see e.g. Figures 2 and 4) after the AD peak and therefore even when OGD is interrupted immediately after the AD peak, hypoxia persists for few minutes after AD. Indeed, in our experimental conditions the recovery of pO₂ to normal levels takes about 3–4 min (Pugliese et al., 2003).

This is important for the possible therapeutic outcome during ischaemia in vivo. It may be envisaged that the block of A₃ receptors may increase the resistance of the brain tissue not only in the ischaemic core but also in the surrounding 'penumbral' region. However, while the action of A₁ block is of limited effectiveness in the ischaemic core (depending on the duration of the episode), more effective neuroprotection can occur in the surrounding regions, where the damage can be ascribed to the concomitant hypoxia/hypoglycaemia and appearance of AD. The observation that a statistically significant recovery of fepsps occurred when AD peaked in concomitance or after interruption of OGD in the presence of A₁ antagonists but not in control slices, supports the notion that A₁ receptor block increases the resistance to the deleterious effect of AD in conditions of milder hypoglycaemia/hypoxia. The causal association of A₃ receptor block, delayed AD appearance and better recovery from ischaemic episodes needs, however, further investigation.

Regardless of the exact mechanisms exerted by A₃ receptors at a cellular level, it appears that the activation of these adenosine receptor subtypes during an ischaemic episode produces deleterious consequences for the survival of neuronal cells and that the block of A₃ receptors may substantially increase the resistance of brain tissue to OGD occurring in ischaemia.

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References


MACK, T.A., SCHAFFHAUSER, H. & CONN, P.J. (1998). Protein kinase C and A3 adenosine receptor activation inhibit presynaptic metabolotropic glutamate receptor (mGlur) function and uncouple mGluRs from GTP-binding proteins. *J. Neurosci.*, 18, 6138–6146.


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