



# Pomegranate (*Punica granatum*) Fruit Extract Suppresses Cancer Progression and Tumor Angiogenesis of Pancreatic and Colon Cancer in Chick Chorioallantoic Membrane Model

Thangirala Sudha<sup>a</sup> , Deena S. Mousa<sup>b</sup> , Ali H. El-Far<sup>c</sup> , and Shaker A. Mousa<sup>a</sup>

<sup>a</sup>Pharmaceutical Research Institute, Albany College of Pharmacy and Health Sciences, Albany, New York, USA; <sup>b</sup>Yale University, New Haven, Connecticut, USA; <sup>c</sup>Department of Biochemistry, Faculty of Veterinary Medicine, Damanhour University, Damanhour, Egypt

## ABSTRACT

Pomegranate fruit extract contains many polyphenols and flavonoids of diverse biological importance including anticancer potential. In cancer, the angiogenesis process facilitates solid cancer growth and metastasis. Here, the antiangiogenic effect of pomegranate fruit extract against human pancreatic cancer (Suit-2) and colon (colo205) cell lines in the chick chorioallantoic membrane (CAM) model was studied along with the effect of pomegranate fruit extract on fibroblast growth factor (FGF2). Pomegranate fruit extract significantly reduced the tumor weight and hemoglobin content in CAM models of pancreatic Suit-2 and colon colo205.

**Abbreviation:** EDTA: ethylene diamine tetraacetate; PBS: phosphate buffered saline; DMEM: Dulbecco's Modified Eagle's medium; ELISA: enzyme linked immunosorbent assay; CAM model: chick chorioallantoic membrane model; ANOVA: analysis of variance

## ARTICLE HISTORY

Received 5 December 2019  
Accepted 6 July 2020

## Introduction

Pancreatic cancer is a malignant disease and represents the fourth leading cause of cancer death in the US, where it is predicted to become the second most frequent cause of cancer-related death by 2030 (1). The incidence rate of pancreatic cancer in both sexes increases with age, therefore, it can be defined as a disease of elderly populations (2).

Histological, morphological, and genetic changes are associated with colon cancer development that accumulate over time (3). Nowadays, colon cancer is the fourth most deadly cancer worldwide with about 900,000 deaths annually. Besides an ageing population and dietary habits, risk factors like obesity, lack of physical exercise, and smoking increase the risk of colorectal cancer (4).

Tumor angiogenesis has been thought to be a valid target for many solid tumors (5). Because angiogenesis is an important factor in tumor progression, inhibition of angiogenesis can lead to tumor growth inhibition (6). Hypoxia-inducing factor-1 (HIF-1) can be expressed in tumors often under hypoxic conditions (7). As a result, HIF-1 induces the expression of

endothelial growth factor (VEGF), epidermal growth, fibroblast growth factor (FGF), and hepatocyte growth factor and thus promotes hypervascularization (8). FGF signaling regulates several processes including angiogenesis. Therefore, agents targeting FGF signaling potentially target both the VEGF and FGF pathways and inhibit angiogenesis (9).

Pomegranate (*Punica granatum*) is a natural product that has been shown to be anti-cancerous due to its large concentration of polyphenols, including ellagitannins, ellagic acid, and other flavonoids like quercetin, kaempferol, and luteolin glycosides (10). After patients with colorectal cancer consumed pomegranate extract at 900 mg/day for 15 days, their colon tissues were found to have significantly high levels of ellagic acid and derivatives, suggesting a potential prevention role by pomegranates against colorectal cancer (11). Furthermore, punicalagin and ellagic acid induced significant inhibition of benzo[a]pyrene-induced DNA adducts (12). Pomegranate induced its anticancer effect through cell-cycle arrest, induction of apoptosis, and inhibition of angiogenesis and metastasis (13).

Numerous cancer studies have investigated antiangiogenesis in either in vitro or in vivo models

(14–17). The current study investigated the antiangiogenic effect of pomegranate against pancreatic and colon cancer cells using the chick chorioallantoic membrane (CAM) model.

## Materials and Methods

### Materials

Human pancreatic cancer (Suit-2) and colon (colo205) cell lines were obtained from ATCC (Manassas, VA, USA). Cell culture reagents and hemoglobin (Hb) standard, Drabkin's reagent, and other common reagents were purchased from Sigma (St. Louis, MO, USA). Matrigel was purchased from BD Bioscience (San Jose, CA, USA).

POMx is a pomegranate (*P. granatum*) derivative of fruits (Pom Wonderable, Los Angeles, CA, USA). POMx capsule contains polyphenolics of about 753 mg that are expressed as mg gallic acid equivalents (18).

### Cells and Cell Culture

Cell lines were grown in DMEM supplemented with 10% fetal bovine serum, 1% penicillin, and 1% streptomycin. Cells were cultured at 37°C with a humidifier atmosphere of 5% CO<sub>2</sub> to sub-confluence and treated with 0.25% (w/v) trypsin/EDTA to effect cell release from culture flask. After washing cells with culture medium, they were suspended in DMEM (free of phenol red and fetal bovine serum) and counted.

### Chick Chorioallantoic Membrane (CAM) Cancer Implant Model

#### Tumor Growth in the CAM Cancer Implant Model

Seven-day old chick embryos were purchased from Spafas, Inc. (Preston, CT, USA) and incubated at 37°C with 55% relative humidity. A hypodermic needle was used to make a small hole in the shell at the air sac and a second hole was made on the broadside of the egg, directly over an avascular portion of the embryonic membrane that was identified by candling. A false air sac was created beneath the second hole by the application of negative pressure at the first hole, causing the CAM to separate from the shell. A window, approximately 1.0 cm<sup>2</sup> was made in the shell over the dropped CAM using a small craft grinding wheel (Dremel, Racine, WI, USA), allowing direct access to the underlying CAM.

The pancreatic (Suit-2) and colon cancer (colo205) cells in exponential growth phase were harvested using 0.25% trypsin-EDTA, washed and suspended in medium. Only suspensions of single cells with a

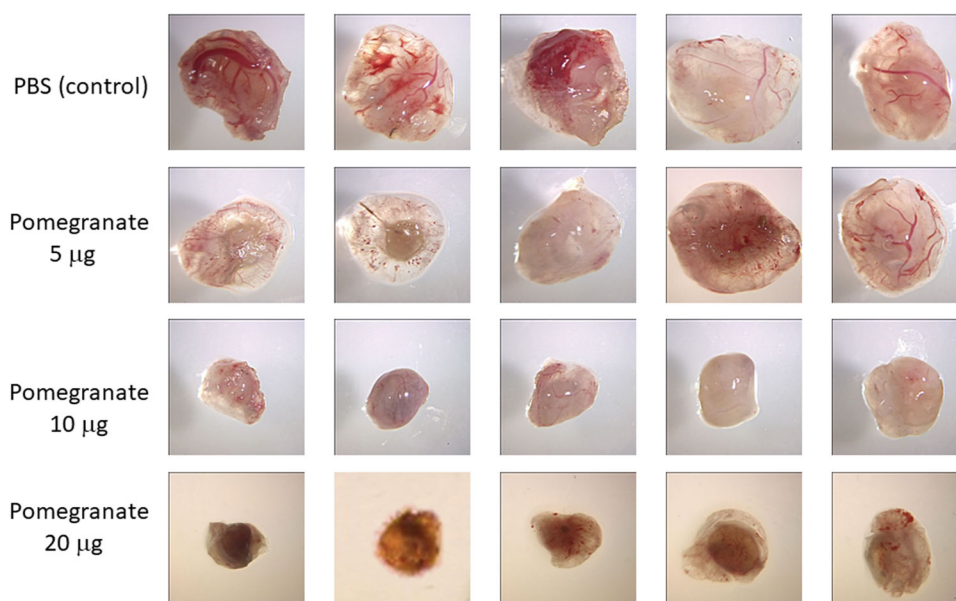
viability exceeding 95% were inoculated. Approximately  $1 \times 10^6$  cells in 30  $\mu$ l of medium were mixed with the same volume (30  $\mu$ l) of Matrigel® (BD Bioscience) and implanted on the chorioallantoic membrane. Matrigel containing different treatments were also inoculated to determine their effect on tumor growth and tumor angiogenesis at day 7 after tumor cell implantation. Three sets of CAM experiments were done, all using the Matrigel and PBS as the control as described next.

In experiment 1, pancreatic cancer cells (all in Matrigel) were treated with PBS, pancreatic cancer (Suit-2) + pomegranate (5  $\mu$ g/CAM), pancreatic cancer (Suit-2) + pomegranate (10  $\mu$ g/CAM), and pancreatic cancer (Suit-2) + pomegranate (20  $\mu$ g/CAM). In experiment 2, the treatment groups (all in Matrigel) were PBS, DMSO, colon cancer (colo205) + pomegranate (20  $\mu$ g/CAM), and colon cancer (colo205) + resveratrol (a promising anticancer drug, 20  $\mu$ g/CAM). In experiment 3, CAMs were inoculated with PBS, FGF2 (40 ng/CAM), and FGF2 (40 ng/CAM) + pomegranate (20  $\mu$ g/CAM), all in Matrigel.

Hemoglobin (Hb) content of tumor masses was determined with Drabkin's reagent. Data represent mean tumor weight (mg) and tumor Hb (mg/ml)  $\pm$  SEM per treatment group ( $n = 5-8$  per group). Tumor mass Hb content was measured as described previously Yalcin et al. (19) to index vascularity of the tumor. For this purpose, each tumor mass was placed into a 0.5 ml tube of double-distilled H<sub>2</sub>O and homogenized for 5–10 min. The samples were then centrifuged at 3,000 g for 10 min, and the supernatants were collected for Hb determination. Fifty microliters of supernatant were mixed with 50  $\mu$ l Drabkin's reagent and allowed to sit at room temperature for 15–30 min; 100  $\mu$ l of this mixture was then placed in a 96-well plate, and absorbance was measured at 540 nm with a Microplate Manager ELISA reader (BioRad Laboratories, Hercules, CA). Hb concentration (mg/ml) was determined by comparison with a standard curve.

#### CAM Growth Factors-Mediated Angiogenesis Assay

The antiangiogenesis potency of pomegranate was examined in the CAM of angiogenesis using 10-day-old chick embryos (20–22). FGF2 (40 ng, dissolved in PBS) was used as a standard pro-angiogenic agent to induce new blood vessel branches on the CAM. Sterile 1.0 cm diameter disks of #1 filter paper were pretreated with 3 mg/mL cortisone acetate and air dried under sterile conditions. PBS (control) and FGF2 with pomegranate (20  $\mu$ g/CAM) were then applied to the pretreated disks and dried. The disks were placed on



**Figure 1.** Dose–response reduction of pancreatic tumor weight as a function of pomegranate dose in CAM tumor model.

growing CAMs in an area between preexisting vessels. After incubation at 37°C with 55% relative humidity for 3 days, the CAM tissue directly beneath each filter disk was resected from the control and treated CAM samples. Tissues were washed three times with PBS, placed in 35-mm Petri dishes and examined under an SV6 stereomicroscope (Karl Zeiss, Thornwood, NY) at 50× magnification. Digital images of the CAM sections were collected using a 3-CCD color video camera system and analyzed with Image-Pro software (Media Cybernetics, Silver Spring, MD, USA). The numbers of vessel branch points contained in a circular region equal to the area of each filter disk were counted. One image was counted in each CAM preparation, and findings from 5 to 8 CAM preparations/group were used for each treatment condition.

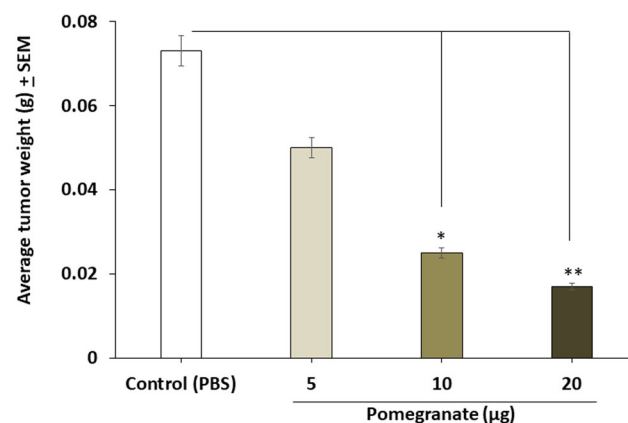
### Statistical Analysis

Statistical analysis was performed using one-way ANOVA and comparing the mean  $\pm$  SEM of branch points from each experimental group with its respective control group. Statistical differences approaching  $P < 0.05$  were considered statistically significant.

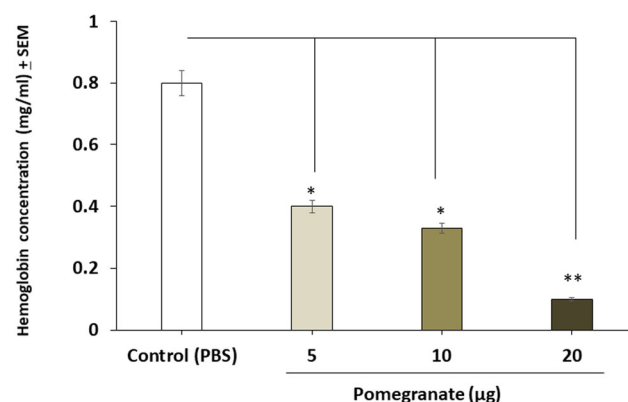
## Results

### Effect of Pomegranate on Pancreatic Tumor Weight and Hemoglobin Concentration

The effect of different pomegranate extract concentrations (5, 10, and 20  $\mu$ g/ml) on the morphology, tumor weight, and Hb concentration are presented in Figures

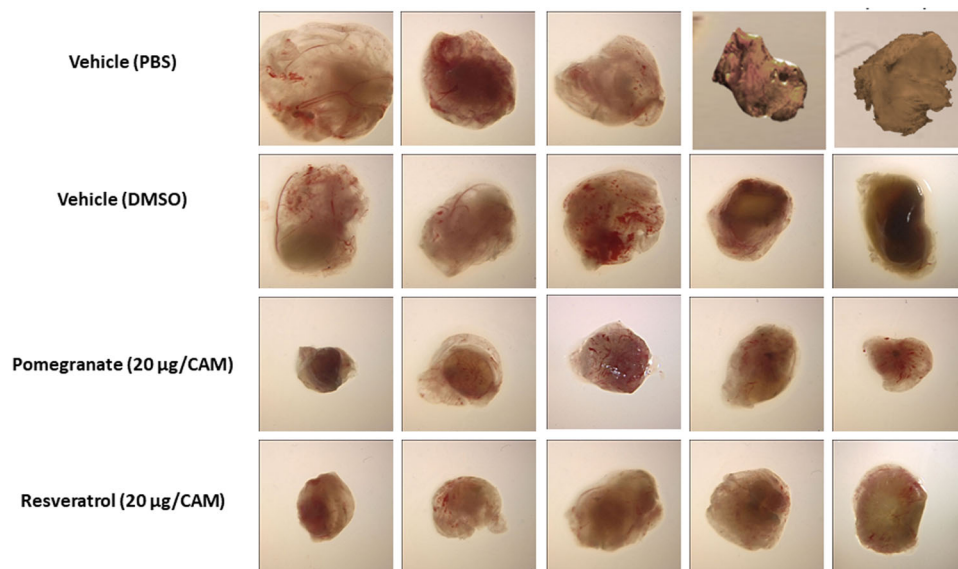


**Figure 2.** Effect of pomegranate extract on pancreatic tumor growth (g) in CAM tumor model.  $n = 5-8$ , \* $P < 0.05$ ; \*\* $P < 0.01$ .

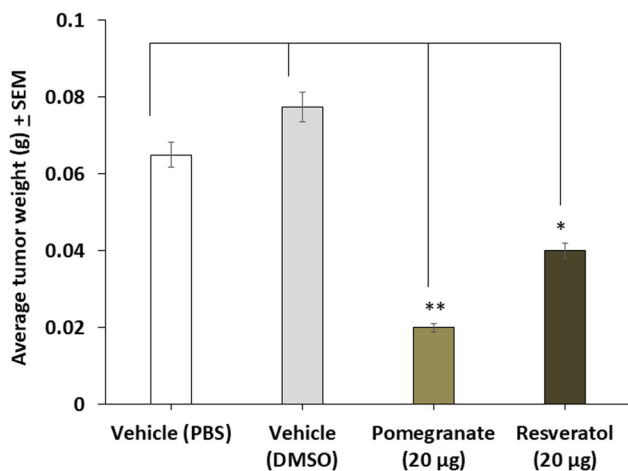


**Figure 3.** Effect of pomegranate extract on pancreatic tumor-mediated angiogenesis (hemoglobin concentration, mg/ml) in CAM tumor model.  $n = 5-8$ , \* $P < 0.05$ ; \*\* $P < 0.01$ .

1–3. Compared with control, pomegranate extract significantly decreased tumor weight ( $P < 0.05$ ) at 5–10  $\mu$ g/ml and  $P < 0.01$  at 20  $\mu$ g/ml.



**Figure 4.** Anti-carcinogenic effect of pomegranate and resveratrol on colo205 tumors in CAM tumor model.



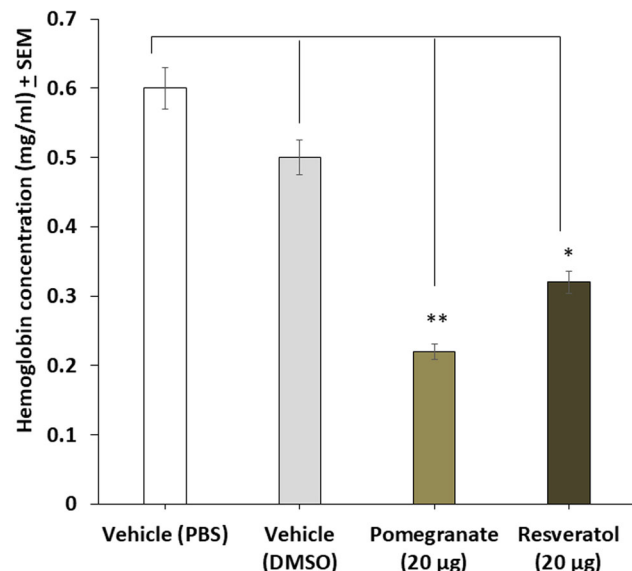
**Figure 5.** Reduction of colon cancer (colo205) tumor weight by pomegranate or resveratrol in CAM tumor model.  $n = 5-8$ , \* $P < 0.05$ ; \*\* $P < 0.01$ .

Hb concentrations were significantly decreased in CAM groups treated with 5 µg/ml ( $P < 0.05$ ), 10 µg/ml ( $P < 0.05$ ), and 20 µg/ml ( $P < 0.01$ ) pomegranate extract.

### Effect of Pomegranate on Colon Tumor Weight and Hemoglobin Concentration

Data illustrated in Figure 4 show the morphological reduction in the tumor of colo205 cancer size due to pomegranate extract (20 µg/ml) in comparison with control and resveratrol-treated (20 µg/ml) CAM.

Pomegranate extract at 20 µg/ml significantly decreased ( $P < 0.01$ ) tumor weight in comparison with positive control group (Figure 5). In comparison with the resveratrol-treated group, pomegranate also induced significant reduction in tumor size ( $P < 0.05$ ).

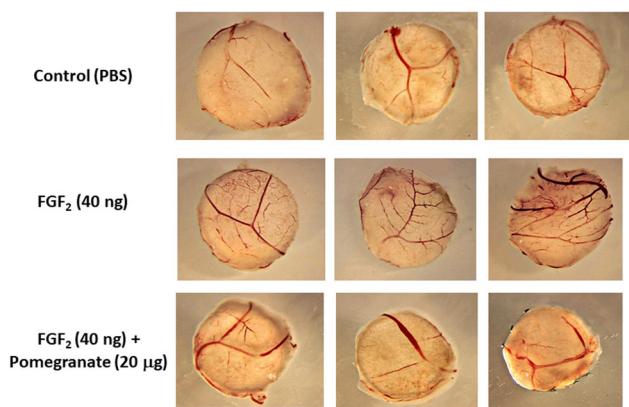


**Figure 6.** Reduction of colon (colo205) tumor weight by pomegranate or resveratrol in CAM tumor model.  $n = 5-8$ , \* $P < 0.05$ ; \*\* $P < 0.01$ .

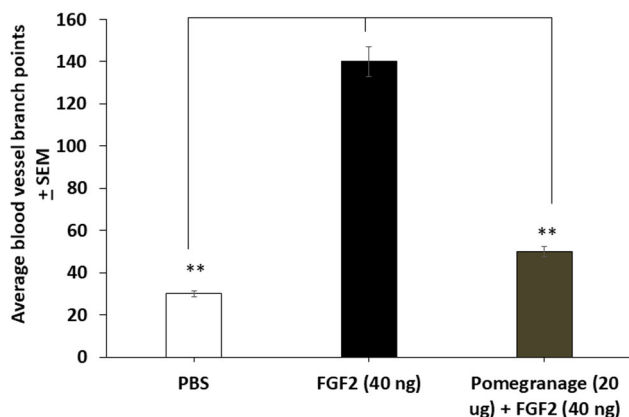
Results shown in Figure 6 revealed a significant decrease in Hb concentrations in pomegranate ( $P < 0.01$ ) and resveratrol-treated ( $P < 0.05$ ) groups compared with positive control CAM. Also, pomegranate significantly decreased ( $P < 0.05$ ) the Hb concentration in comparison with resveratrol treated.

### Effect of Pomegranate on FGF2-Induced Angiogenesis

Results shown in Figures 7 and 8 are the effect of FGF2 (40 ng/ml) and pomegranate extract (20 µg/ml) on the average blood vessel branch count in CAMs.



**Figure 7.** Effect of pomegranate on FGF2-mediated angiogenesis in CAM tumor model.



**Figure 8.** Reduction of hemoglobin content in colon (colo205) tumors by pomegranate CAM tumor model.  $n = 5-8$ ,  $**P < 0.01$ .

FGF2 significantly increased the average blood vessel branch count compared with control CAM, while pomegranate-treated CAM significantly decreased the average blood vessel branch count compared with FGF2-treated CAM.

## Discussion

Angiogenesis is a process that is pivotal for cancer progression (23). Therefore, many studies have investigated the antiangiogenesis effect of various synthetic and natural products (24,25). Pomegranate is a natural product with an anticancer effect against different cancer types including breast (26), colorectal (27), pancreatic (28), prostate (29), lung (30), ovarian (31), and liver (32) cancers.

In the present study, pomegranate extract significantly decreased the tumor weight and Hb concentrations of CAM-implanted pancreatic and colon cancers. These results indicated the anticancer effect of pomegranate, especially through attenuation of angiogenesis.

Sartippour et al. (15) revealed that the ellagitannin-rich pomegranate extract induced antiangiogenic effect on human prostate cancer cells (LNCaP) through induction of HIF-1 $\alpha$  and VEGF. Also, pomegranate seed oil or fermented juice polyphenols downregulated VEGF in MCF-7 breast cancer cells and upregulated migration inhibitory factor (MIF) in MDA-MB-231 triple negative breast cancer cells (14). Oral consumption of pomegranate fruit extract inhibits growth and progression of primary lung tumors in mice through downregulation of nuclear factor-kappa B (NF- $\kappa$ B), phosphatidylinositol 3-kinase (PI3K), protein kinase B (Akt), mammalian target of rapamycin (mTOR), and VEGF (17). Ellagic acid, a constituent of pomegranate, induced a significant decrease in VEGF level in LNCaP at concentrations of 25 and 50  $\mu$ M (33). Also, ellagic acid reduced VEGF receptor type 2 (VEGFR-2) expression in human bladder cancer cells (34). Moreover, ellagic acid inhibited MDA-MB-231 cancer growth and VEGFR-2 expression in a breast cancer xenograft study (35).

Overexpression of FGF2 and its receptors has a wide range of effects in various cancers and many human tumor cell lines (36) including cellular proliferation, cellular invasiveness, increased angiogenesis, and enhanced metastasis (37). In the present study pomegranate extract defeated the angiogenic effect of FGF2. Therefore, pomegranate extract is a promising natural product that can effectively control tumor angiogenesis.

## Conclusion

Pomegranate extract attenuated the angiogenesis of pancreatic (Suit-2) and colon (colo205) cancer cells through significant reduction in tumor weight and small blood vessels' count in the CAM model. In addition, pomegranate extract successfully reduced angiogenesis in the CAM that was induced by FGF2. Therefore, we can conclude that pomegranate and its active constituents can reduce cancer progression and angiogenesis in the CAM model.

## Disclosure Statement

The authors declare no conflict of interest.

## Funding

This study was funded by the Pharmaceutical Research Institute (PRI) and its Center of Excellence for Translation Research (CETR).

## ORCID

Thangirala Sudha  <http://orcid.org/0000-0002-6428-1804>

Deena S. Mousa  <http://orcid.org/0000-0002-9294-015X>

Ali H. El-Far  <http://orcid.org/0000-0001-9721-4360>

## References

- Rahib L, Smith BD, Aizenberg R, et al. Projecting cancer incidence and deaths to 2030: The unexpected burden of thyroid, liver, and pancreas cancers in the United States. *Cancer Res.* 2014;74:2913–2921.
- Bosetti C, Bertuccio P, Negri E, La Vecchia C, Zeegers MP, Boffetta P. Pancreatic cancer: overview of descriptive epidemiology. *Mol Carcinog.* 2012; 51(1):3–13. doi:10.1002/mc.20785
- Balchen V, Simon K. Colorectal cancer development and advances in screening. *Clin Interventions Aging.* 2016; 11:967–976. doi:10.2147/CIA.S109285
- Dekker E, Tanis PJ, Vleugels JLA, Kasi PM, Wallace MB. Colorectal cancer. *Lancet.* 2019;394(10207): 1467–1480. doi:10.1016/S0140-6736(19)32319-0
- Marmé D. Tumor angiogenesis: a key target for cancer therapy. *Oncol Res Treat.* 2018;41(4):164. doi:10.1159/000488340
- Melegh Z, Oltean S. Targeting angiogenesis in prostate cancer. *IJMS.* 2019;20(11):2676. doi:10.3390/ijms20112676
- Itatani Y, Kawada K, Yamamoto T, Sakai Y. Resistance to antiangiogenic therapy in cancer—alterations to anti-VEGF pathway. *Int J Mol Sci.* 2018; 19(4):1232. doi:10.3390/ijms19041232
- Bergers G, Benjamin LE. Tumorigenesis and the angiogenic switch. *Nat Rev Cancer* 2003;3(6):401–410. doi:10.1038/nrc1093
- Lieu C, Heymach J, Overman M, Tran H, Kopetz S. Beyond VEGF: inhibition of the fibroblast growth factor pathway and antiangiogenesis. *Clin Cancer Res.* 2011;17(19):6130–6139. doi:10.1158/1078-0432.CCR-11-0659
- Seeram NP, Adams LS, Henning SM, Niu Y, Zhang Y, Nair MG, Heber D. In vitro antiproliferative, apoptotic and antioxidant activities of punicalagin, ellagic acid and a total pomegranate tannin extract are enhanced in combination with other polyphenols as found in pomegranate juice. *J Nutr Biochem.* 2005; 16(6):360–367. doi:10.1016/j.jnutbio.2005.01.006
- Nuñez-Sánchez MA, García-Villalba R, Monedero-Saiz T, García-Talavera NV, Gómez-Sánchez MB, Sánchez-Álvarez C, García-Albert AM, Rodríguez-Gil FJ, Ruiz-Marín M, Pastor-Quirante FA, et al. Targeted metabolic profiling of pomegranate polyphenols and urolithins in plasma, urine and colon tissues from colorectal cancer patients. *Mol Nutr Food Res.* 2014; 58(6):1199–1211. doi:10.1002/mnfr.201300931
- Zahin M, Ahmad I, Gupta RC, Aqil F. Punicalagin and ellagic acid demonstrate antimutagenic activity and inhibition of benzo[a]pyrene induced DNA adducts. *Biomed Res Int.* 2014;2014:1–10. doi:10.1155/2014/467465
- Turrini E, Ferruzzi L, Fimognari C. Potential effects of pomegranate polyphenols in cancer prevention and therapy. *Oxid Med Cell Longev.* 2015;2015:938475. doi:10.1155/2015/938475
- Toi M, Bando H, Ramachandran C, Melnick SJ, Imai A, Fife RS, Carr RE, Oikawa T, Lansky EP. Preliminary studies on the antiangiogenic potential of pomegranate fractions in vitro and in vivo. *Angiogenesis* 2003;6(2):121–128. doi:10.1023/B:AGEN.0000011802.81320.e4
- Sartippour MR, Seeram NP, Rao JY, Moro A, Harris DM, Henning SM, Firouzi A, Rettig MB, Aronson WJ, Pantuck AJ, et al. Ellagitannin-rich pomegranate extract inhibits angiogenesis in prostate cancer in vitro and in vivo. *Int J Oncol.* 2008;32(2):475–480. <http://www.ncbi.nlm.nih.gov/pubmed/18202771> doi:10.3892/ijo.32.2.475
- Labrecque L, Lamy S, Chapus A, Mihoubi S, Durocher Y, Cass B, Bojanowski MW, Gingras D, Béliveau R. Combined inhibition of PDGF and VEGF receptors by ellagic acid, a dietary-derived phenolic compound. *Carcinogenesis* 2005;26(4):821–826. doi:10.1093/carcin/bgi024
- Khan N, Afaq F, Kweon M-H, Kim K, Mukhtar H. Oral consumption of pomegranate fruit extract inhibits growth and progression of primary lung tumors in mice. *Cancer Res.* 2007;67(7):3475–3482. doi:10.1158/0008-5472.CAN-06-3941
- Basu A, Newman ED, Bryant AL, Lyons TJ, Betts NM. Pomegranate polyphenols lower lipid peroxidation in adults with type 2 diabetes but have no effects in healthy volunteers: a pilot study. *J Nutr Metab.* 2013;2013:1–7. doi:10.1155/2013/708381
- Yalcin M, Dyskin E, Lansing L, Bharali DJ, Mousa SS, Bridoux A, Hercbergs AH, Lin HY, Davis FB, Glinesky GV, et al. Tetraiodothyroacetic acid (tetrac) and nanoparticulate tetrac arrest growth of medullary carcinoma of the thyroid. *J Clin Endocrinol Metab.* 2010; 95(4):1972–1980. doi:10.1210/jc.2009-1926
- Deryugina EI, Quigley JP. Chapter 2. Chick embryo chorioallantoic membrane models to quantify angiogenesis induced by inflammatory and tumor cells or purified effector molecules. *Methods Enzymol.* 2008; 444:21–41. doi:10.1016/S0076-6879(08)02802-4
- Marcinkiewicz C, Weinreb PH, Calvete JJ, Kisiel DG, Mousa SA, Tuszynski GP, Lobb RR. Obtustatin: a potent selective inhibitor of  $\alpha 1 \beta 1$  integrin in vitro and angiogenesis in vivo. *Cancer Res.* 2003;63(9): 2020–2023.
- Bridoux A, Cui H, Dyskin E, Yalcin M, Mousa SA. Semisynthesis and pharmacological activities of tetrac analogs: angiogenesis modulators. *Bioorg Med Chem Lett.* 2009;19(12):3259–3263. doi:10.1016/j.bmcl.2009.04.094
- Bielenberg DR, Zetter BR. The contribution of angiogenesis to the process of metastasis. *Cancer J.* 2015; 21(4):267–273. doi:10.1097/PPO.0000000000001138
- Al-Abd AM, Alamoudi AJ, Abdel-Naim AB, Neamatallah TA, Ashour OM. Antiangiogenic agents for the treatment of solid tumors: potential pathways, therapy and current strategies—a review. *J Adv Res.* 2017;8(6):591–605. doi:10.1016/J.JARE.2017.06.006
- Wang T-Y, Chen J-X. Effects of curcumin on vessel formation insight into the pro- and antiangiogenesis

- of curcumin. *Evid Based Complement Alternat Med.* 2019;2019:1390795. doi:10.1155/2019/1390795
26. Adams LS, Zhang Y, Seeram NP, Heber D, Chen S. Pomegranate ellagitannin-derived compounds exhibit antiproliferative and antiaromatase activity in breast cancer cells in vitro. *Cancer Prev Res.* 2010;3(1):108–113. doi:10.1158/1940-6207.CAPR-08-0225
27. Nuñez-Sánchez MA, Dávalos A, González-Sarrías A, Casas-Agustench P, Visioli F, Monedero-Saiz T, García-Talavera NV, Gómez-Sánchez MB, Sánchez-Álvarez C, García-Albert AM, et al. MicroRNAs expression in normal and malignant colon tissues as biomarkers of colorectal cancer and in response to pomegranate extracts consumption: critical issues to discern between modulatory effects and potential artefacts. *Mol Nutr Food Res.* 2015;59(10):1973–1986. doi:10.1002/mnfr.201500357
28. Nair V, Dai Z, Khan M, Ciolino HP. Pomegranate extract induces cell cycle arrest and alters cellular phenotype of human pancreatic cancer cells. *Anticancer Res.* 2011;31(9):2699–2704. <http://www.ncbi.nlm.nih.gov/pubmed/21868510>
29. Malik A, Afaq F, Sarfaraz S, Adhami VM, Syed DN, Mukhtar H. Pomegranate fruit juice for chemoprevention and chemotherapy of prostate cancer. *Proc Natl Acad Sci USA.* 2005;102(41):14813–14818. doi:10.1073/pnas.0505870102
30. Khan N, Hadi N, Afaq F, Syed DN, Kweon M-HH, Mukhtar H. Pomegranate fruit extract inhibits prosurvival pathways in human A549 lung carcinoma cells and tumor growth in athymic nude mice. *Carcinogenesis* 2007;28(1):163–173. doi:10.1093/carcin/bgl145
31. Liu H, Zeng Z, Wang S, Li T, Mastriani E, Li Q-H, Bao H-X, Zhou Y-J, Wang X, Liu Y, et al. Main components of pomegranate, ellagic acid and luteolin, inhibit metastasis of ovarian cancer by down-regulating MMP2 and MMP9. *Cancer Biol Ther.* 2017;18(12):990–999. doi:10.1080/15384047.2017.1394542
32. Bishayee A, Bhatia D, Thoppil RJ, Darvesh AS, Nevo E, Lansky EP. Pomegranate-mediated chemoprevention of experimental hepatocarcinogenesis involves Nrf2-regulated antioxidant mechanisms. *Carcinogenesis* 2011;32(6):888–896. doi:10.1093/carcin/bgr045
33. Vanella L, Di Giacomo C, Acquaviva R, Barbagallo I, Li Volti G, Cardile V, Abraham N, Sorrenti V. Effects of ellagic acid on angiogenic factors in prostate cancer cells. *Cancers.* 2013;5(4):726–738. doi:10.3390/cancers5020726
34. Ceci C, Tentori L, Atzori M, Lacal P, Bonanno E, Scimeca M, Cicconi R, Mattei M, de Martino M, Vespasiani G, et al. Ellagic acid inhibits bladder cancer invasiveness and in vivo tumor growth. *Nutrients* 2016;8(11):744. doi:10.3390/nu8110744
35. Wang N, Wang Z-Y, Mo S-L, Loo TY, Wang D-M, Luo H-B, Yang D-P, Chen Y-L, Shen J-G, Chen J-P, et al. Ellagic acid, a phenolic compound, exerts anti-angiogenesis effects via VEGFR-2 signaling pathway in breast cancer. *Breast Cancer Res Treat.* 2012;134(3):943–955. doi:10.1007/s10549-012-1977-9
36. Chandler LA, Sosnowski BA, Greenlees L, Aukerman SL, Baird A, Pierce GF. Prevalent expression of fibroblast growth factor (FGF) receptors and FGF2 in human tumor cell lines. *Int J Cancer* 1999;81(3):451–458. doi:10.1002/(sici)1097-0215(19990505)81:3 < 451::aid-ijc20 > 3.0.co;2-h
37. Kwabi-Addo B, Ozen M, Ittmann M. The role of fibroblast growth factors and their receptors in prostate cancer. *Endocr Relat Cancer.* 2004;11:709–724. doi:10.1677/erc.1.00535