The Medical Benefit of Gnaphalium Luteoalbum-A Review

Ali Esmail Al-Snafi

Department of Pharmacology, College of Medicine, University of Thi qar, Iraq. Corresponding Author: Ali Esmail Al-Snafi

Abstract: The phytochemical screening of *Gnaphalium luteoalbum* revealed that the plant contained alkaloids, carbohydrates, phenols, flavonoids, saponins, tannins, glucoside resins, phytosterins, terpenoids and fixed oils. Pharmacoloical studies showed that *Gnaphalium luteoalbum* possessed antibacterial, antifungal, antioxidant anti- inflammatory and cytotoxic effects. The current review highlighted the chemical constituents and pharmacological effects of *Gnaphalium luteoalbum*.

Keywords: chemical constituents, pharmacology, Gnaphalium luteoalbum

Date of Submission: 25-04-2019 Date of acceptance: 06-05-2019

I. INTRODUCTION

Herbal medicine is the oldest form of medicine known to mankind. It was the mainstay of many earlycivilizations and still the most widely practiced form of medicine in the world today. Recent reviews revealed that the medicinal plants possessed wide range of pharmacological effect and represented a good alternatives in the treatment and prevention of human diseases⁽¹⁻²⁵⁾. The phytochemical screening of *Gnaphalium luteoalbum* revealed that the plant contained alkaloids, carbohydrates, phenols, flavonoids, saponins, tannins, glucoside resins, phytosterins, terpenoids and fixed oils. Pharmacolocical studies showed that *Gnaphalium luteoalbum* possessed antibacterial, antifungal, antioxidant anti-inflammatory and cytotoxic effects. The current review was designed to highlight the chemical constituents and pharmacological effects of *Gnaphalium luteoalbum*.

Synonyms:

Chrysocoma villosa, Dasyanthus conglobatus, Bubani, Gnaphalium dealbatum var luteo-fuscum, depressum Steud., Gnaphalium dichotomum, Gnaphalium helichrysoides, Gnaphalium Gnaphalium helichrysoides var helichrysoides, Gnaphalium luteo-fuscum, Gnaphalium luteoalbum var. compactum, Gnaphalium luteoalbum var incanum, Gnaphalium luteoalbum f. luteoalbum, Gnaphalium luteoalbum subsp Gnaphalium luteoalbum var. luteoalbum, luteoalbum luteoalbum. Gnaphalium var pallidum, Gnaphalium nanum, pallidum, Gnaphalium trifidum, Gnaphalium Laphangium luteoalbum, Pseudognaphalium luteoalbum, Pseudognaphalium luteoalbum subsp luteoalbum⁽²⁶⁻²⁷⁾.

Taxonomic classification:

Kingdom: Plantae, **Subkingdom**: Viridiplantae, **Infrakingdom**: Streptophyta, **Superdivision**: Embryophyta, **Division**: Tracheophyta, **Subdivision**: Spermatophytina, **Class**: Magnoliopsida, **Superorder**: Asteranae, **Order**: Asterales, **Family**: Asteraceae, **Genus**: *Gnaphalium*, **Species**: *Gnaphalium luteoalbum*⁽²⁸⁾. **Common names**:

Arabic: Kutaina, Ghwbira, râraâ, Sabon Efreet; **Chinese:** si mian cao; **English**: Jersey cudweed, red-tip rabbit-tobacco; **French:** Cotonnière blanc-jaunâtre, Gnaphale jaunâtre, Gnaphale jaune blanc; **German:** gelbliches, Scheinruhrkraut; **Italian:** Canapicchia pagliata, **Swedish:** vitnoppa⁽²⁹⁻³⁰⁾.

Distribution:

The plant was distributed in Africa, Asia, Europe, Australasia, Northern and Southern America. It was found in **Africa** (Kenya, Tanzania, Uganda, Kenya, Tanzania, Uganda, Eritrea, Ethiopia, Somalia, Sudan, Algeria, Egypt, Libya, Morocco, Tunisia, Angola, Malawi, Mozambique, Zambia, Zimbabwe, Botswana, Lesotho, Namibia, South Africa, Swaziland, Ghana, Mali, Nigeria, Senegal, Burundi, Cameroon, Equatorial Guinea, Comoros, Mauritius, Reunion); **Asia** (Oman; Yemen, Azerbaijan, Georgia, Russian Federation, China, Taiwan, Kazakhstan, Tajikistan, Turkmenistan, Afghanistan, Iraq, Iran, Palestine, Lebanon, Syria, Turkey, India, Pakistan, Laos, Thailand, Vietnam, Indonesia, Philippines); **Australasia** (Australia, New Zealand);

Europe (Belarus, Lithuania, Moldova, Russian Federation-European part, Ukraine, Austria, Belgium, Czech Republic, Germany, Hungary, Netherland, Poland, Slovakia, Switzerland, Sweden, United Kingdom, Bosnia and Herzegovina, Bulgaria, Croatia, Greece, Italy, Macedonia, Montenegro, Romania, Serbia, Slovenia, France, Portugal, Spain); Northern America (United states, Mexico) and Southern America (Argentina, Chile, Peru)⁽²⁹⁾.

Description:

Annual herb to 50 cm tall, all parts whitish-woolly. Stems usually several from the base, decumbent at first, later erect. Leaves sessile, up to 8×1 cm at base, oblanceolate, becoming smaller and lanceolate or linear above, greyish or whitish tomentose or arachnoid, on both sides; margin entire. Inflorescence consisting of dense clusters of capitula 1-several together in terminal corymbs. Involucres 3-4 mm in diameter; phyllaries in c. 3 rows, pale brown to whitish. Outer female florets very numerous, whitish. Bisexual disk florets cylindric. Achenes ellipsoid, papillose. Pappus of numerous soft bristles⁽³¹⁻³²⁾.

Traditional uses:

In the Punjab, the leaves of the plant were used as vulnerary and astringent. In Pakistan, It was used as anti-diarrheal, the infusion of aerial parts was used as emmenagogue. In Bangladesh, the plant was applied as a poultice to heal fractured bones. It also used in Bangladesh as tonic and for the treatment of tumor, gout, dermatitis^(26, 33-34). In Iraq, it was used traditionally as astringent, counterirritant, as vulnerary and for the treatment of gout⁽³⁵⁾. In the Punjab leaves were used as vulnerary and astringent. In Pakistan, it was used as anti-diarrheal. Infusion of aerial parts was used as emmenagogue. It was also used as a counter-irritant for gout. In Bangladesh, plant was used by the Garo tribe, crushed along with dried fish and applied as a poultice to heal fractured bones. Also, used by the Kavirajes of Chalna as tonic, and for tumor, gout, and dermatitis⁽²⁶⁾. In Belgium, it was used for the treatment of cancer (Breast)⁽³⁶⁾. The leaves of *G. luteo-album* were also used as astringent, cholagogue, diuretic, febrifuge, and haemostatic⁽³⁷⁾.

Chemical constituents:

The phytochemical screening of crude extract and its fractions revealed a wide range of phytoconstituents included: alkaloids, carbohydrates, phenols, flavonoids, saponins, tannins, glucoside resins, phytosterins, terpenoids and fixed oils⁽³⁸⁻⁴⁰⁾.

Many flavonoids were isolated from included apigenin, apigenin 7-O- β -D- glucopyranoside, luteolin, luteolin 4'-O- β -D-glucopyranoside, luteolin 7-O- β -D- glucopyranoside, Jaceosidin and gnaphalin⁽¹⁶⁾. 5, 7, 3, 4 tetrahydroxy flavone; 5, 3, 4 trihydroxy flavonol and 3, 5 dihydroxy flavonol were isolated from the aerial parts of Gnaphalium luteo-album⁽⁴¹⁻⁴²⁾.

5,4'-dihydroxy-6-methoxy-7-O- β -glucopyranosideflavone (hispidulin-7-O- gluco pyranoside) and stigmasterol-3-O- β -glucopyranoside were also isolated from the leaves of the plant⁽⁴³⁾.

Three flavonols isolated from the leaves of G. *luteo-album* were structurally related to each other (gnaphaliin, calycopterin and 3'-Methoxycalycopterin). Chlorophyll a, chlorophyll b and carotenoid were also isolated from G. *luteo-album* leaves. The leave flavonoids and pigments were increased when the plant exposed to UV-B radiation⁽⁴⁴⁾.

The essential oil of the herbal parts of *G. luteo-album* was analyzed by gas chromatography and gas chromatography/mass spectrometry. Forty-four compounds were identified in the oil of *G. luteo-album*, representing 70.6% of the total oil with 4.4% monoterpene hydrocarbons, 5.0% oxygenated monoterpenes, 14.7% sesquiterpene hydrocarbons, 3.6% oxygenated sesquiterpenes, 29.1% aliphatic compounds, 10.4% fatty acids and esters, and 3.4% others. The main constituents were found to be decanal (9.7%), β -caryophyllene (8.0%), and α -gurjunene (6.4%). However, the compounds identified in the essential oil of *Gnaphalium luteo-album* and thier percentage were: α -Pinene 2%, 3-Hexanone 1.9%, 2-Hexanone 2%, Hexanal 1%, Undecane 0.4, β -Pinene 0.3, δ -2-Carene trace, α -Phellandrene trace, Heptanal 0.9%, Limonene 1.5%, 1,8-Cineole 1.7%, 2-Hexanol 0.4%, 2-Pentyl furan 1.2%, γ –Terpinene 0.2%, *p*-Cymene 0.4%, Octanal 1%, Hexanol 0.2%, Nonanal 4.1%, Tetradecane 0.5%, (*E*)-2-Octenal 0.2%, Decanal 9.7%, Camphor trace, α -Gurjunene 6.4%, (*E*)-2-Nonenal 0.3%, Linalool 0.4%, β -Caryophyllene 8%, Undecanal 1.1%, α -Humulene 0.3, α -Terpineol trace, Dodecanal 2,5%, Naphthalene 0.7%, (*E*,*E*)-2,4-Decadienal 0.5%, Octyl hexanoate 0.6, (*E*)-Geranyl acetone 1.2%, Pentadecanal 2.4%, Carvacrol 1.7%, 1-Methylethyl hexadecanoic acid 3%, Dodecanoic acid 2.4%, Tetradecanoic acid 0.5%, Hexadecanoic acid 1.9%, Monoterpene hydrocarbons 4.4%,

Oxygenated monoterpenes 5%, Sesquiterpene hydrocarbons 14.7%, Oxygenated sesquiterpenes 3.6%, Aliphatic compounds 29.1% and Fatty acids and esters 10.4%⁽³⁷⁾.

However, the quantitative and qualitative analysis of the *Gnaphalium luteo-album* oil by GC and GC-MS carried out by Kushwaha, led to the identification of 14 constituents, constituting 96.13 % of the oil. The identified compounds (%) were: undecane: 2.51, indole: 8.12, 5-methyl undecane: 7.99, dodecane: 31.10, α -copaene: 2.36, limonene aldehyde: trace, isoledene 6.04, β -caryophyllene 4.12, E- β -fernecene 1.76, sesquisabinene: trace, γ -cuprenene 7.0, caryophyllene oxide 1.89, veridiflorol 4.98, 6,10,14-trimethyl 2-pentadecanone 18.23⁽⁴⁵⁾.

Pharmacological effects:

Antiinflammatory effect:

Several extracts from the aerial parts of *Gnaphalium luteo-album* possessed anti-inflammatory activity⁽³⁷⁾. **Antimicrobial effect:**

The *Gnaphalium luteo-album* oil showed good activity against *Klebsiella pneumoniae* with zone of inhibition of (ZOI) $15.00 \pm 0.00 \text{ mm}$ and 50 µl/ mL MIC value. Oil was found to be active against E. coli with $13.00 \pm 0.00 \text{ mm}$ ZOI and 50 µl/ mL MIC value. The oil has also shown significant activity against Pseudomonas aeruginosa and Salmonella enterica (ZOI = $12.00 \pm 0.00 \text{ and} 78 11.00 \pm 0.00 \text{ mm}$, respectively) with MIC value of 50 µl/ mL for each while least activity was recorded against Staphylococcus aureus (ZOI = $10.00 \pm 0.58 \text{ mm}$ and MIC = 100 µl/ml)⁽⁴⁶⁾.

The acetone leaf extract of the leaves was assayed for antifungal effect against plant pathogenic fungi in vitro (*Aspergillus parasiticus, Aspergillus niger, Colletotrichum gloeosporioides, Fusarium oxysporum, Penicillium expansum, Penicillium janthinellum, Phytophthora nicotiana, Pythium ultimum* and *Trichoderma harzianum*). The acetone leaves extract of possessed strong antifungal activity and showed excellent efficacy against *Phytophthora nicotiana* and *Fusarium oxysporum*, with MIC values of 20 and 160 µg/ml respectively. The isolated compounds (5,4'-dihydroxy-6-methoxy-7-O- β -glucopyranoside flavone (hispidulin-7-O-glucopyranoside) and stigmasterol-3-O- β -glucopyranoside) showed high activity against the selected fungal organisms with MIC values ranging from 0.02 to 1.25 mg/ml⁽⁴³⁾.

Cytotoxic effect:

The cytotoxic activity of crude methanol of the leaves of *Gnaphalium luteoalbum* was investigated against healthy mouse fibroblasts (NIH3T3), healthy monkey kidney (VERO) and four human cancer cell lines (gastric, AGS; colon, HT-29; and breast, MCF-7 and MDAMB-231) using MTT assay. The crude methanol of the leaves of *Gnaphalium luteoalbum* showed high cytotoxicity against AGS and MCF-7 cell lines with IC_{50} of 0.98 and 0.34 mg/ml respectively⁽⁴⁷⁾.

However, no cytotoxicity was recorded for two compounds isolated from the plant (5,4'-dihydroxy-6-methoxy-7-O- β -glucopyranosideflavone (hispidulin-7-O- glucopyranoside) against Vero kidney cells at 200 µg/ml, the highest concentration tested⁽⁴⁸⁾.

Antioxidant effect:

The methanol extracts *Gnaphalium luteoalbum* (50µg/ml) were subjected to preliminarily screening assay for their free radical scavenging potentialities against stable DPPH• (2, 2- diphenyl-1-picrylhydrazyl), using ascorbic acid as a positive control. DPPH• % at μ g/ml m was 90.4, EC₅₀ was 23.8±2.8 μ g/ml⁽⁴⁸⁾.

The antioxidant potentiality of crude methanol extract (CME), carbon tetrachloride fraction (CTF), petroleum ether fraction (PEF), chloroform fraction (CLF) and ethyl acetate fraction (EAF) of aerial parts of *Gnaphalium luteoalbum* (GL) was studied in vitro. The CME showed the highest scavenging activity (43.28%) with IC₅₀ of 398.49 µg/ml in the DPPH radical scavenging test. The IC₅₀ values of EAF, CME were statistically significant (P < 0.05, P < 0.01) with respect to ascorbic acid (ACA). In OH and NO radical scavenging tests, maximum scavenging (48.39%, 69.64%) was also reported for CME compared to CTF, PEF, CLF and EAF. Compared to ACA, in case of OH and NO radical scavenging activities the IC₅₀ values of CME were markedly significant (P < 0.01, P < 0.05). In the TAC test, CME showed the highest antioxidant activity (absorbance, 2.6 nm) related to other fractions. The total phenolic contents (TPC) was found to be the highest in the CME (115.96 mg of gallic acid equivalent/g of dried extract) rather than other fractions. The ranking order of CTF, PEF, CLF, EAF and CME for total flavonoids contents (TFC) was 48.67 < 55.75 < 65.29 < 71.35 < 82.29 mg quercetin equivalent/g of dried extract⁽³⁹⁾.

II. CONCLUSION

The current review discussed the chemical constituents and pharmacological effects of *Gnaphalium luteo-album* to encourage its uses in medical practice as a result of efficacy and safety.

REFERENCES

- [1]. Al Snafi AE, Al-Fartosi KG and Al-Yasiry ZQ. Study the effect of PG E1 and PGF2α on male rat reproductive functions. Immun Endoc & Metab Agents in Med Chem 2018; 18: 1-4.
- [2]. Al-Snafi AE. *Glycyrrhiza glabra*: A phytochemical and pharmacological review. IOSR Journal of Pharmacy 2018;8(6): 1-17.
- [3]. Al-Snafi AE. Therapeutic importance of *Hyoscyamus* species grown in Iraq (*Hyoscyamus albus*, *Hyoscyamus niger* and *Hyoscyamus reticulates*)- A review. IOSR Journal of Pharmacy 2018; 8(6): 18-32.
- [4]. Al-Snafi AE. Pharmacological and therapeutic activities of *Hedera helix* A review IOSR Journal of Pharmacy 2018; 8(5): 41-53.
- [5]. Al-Snafi AE. Pharmacological importance of *Haplophyllum* species grown in Iraq- A review. IOSR Journal of Pharmacy 2018;8(5): 54-62.
- [6]. Al-Snafi AE. Chemical constituents and pharmacological activities of *Gossypium herbaceum* and *Gossypium hirsutum* A review. IOSR Journal of Pharmacy 2018; 8(5): 64-80.
- [7]. Al-Snafi AE. The chemical constituents and pharmacological effects of *Foeniculum vulgare* A review. IOSR Journal of Pharmacy 2018; 8(5): 81-96.
- [8]. Al-Snafi AE. Arabian medicinal plants affected female fertility- plant based review (part 1). IOSR Journal of Pharmacy 2018; 8(7): 46-62.
- [9]. Al-Snafi AE. Arabian medicinal plants affected male fertility- plant based review (part 1). IOSR Journal of Pharmacy 2018; 8(7): 63-76.
- [10]. Al-Snafi AE. Arabian medicinal plants for the treatment of intestinal disorders- plant based review (part 1). IOSR Journal of Pharmacy 2018; 8(6): 53-66.
- [11]. Al-Snafi AE. Arabian medicinal plants possessed gastroprotective effects- plant based review (part 1). IOSR Journal of Pharmacy 2018; 8(7): 77-95.
- [12]. Al-Snafi AE. Arabian medicinal plants with analgesic and antipyretic effects- plant based review (Part 1). IOSR Journal of Pharmacy 2018; 8(6): 81-102.
- [13]. Al-Snafi AE. Arabian medicinal plants with antiurolithiatic and diuretic effects plant based review (Part 1). IOSR Journal of Pharmacy 2018; 8(6): 67-80.
- [14]. Al-Snafi AE. Pharmacological and therapeutic importance of *Hibiscus sabdariffa* A review. International Journal of Pharmaceutical Research 2018; 10(3): 451-475.
- [15]. Al-Snafi AE. Chemical constituents, pharmacological effects and therapeutic importance of *Hibiscus* rosa-sinensis- A review. Journal of Pharmacy 2018; 8 (7): 101-119.
- [16]. Al-Snafi AE. Arabian medicinal plants with antiinflammatory effects- plant based review (part 1). Journal of Pharmacy 2018; 8 (7): 55-100.
- [17]. 239-Al-Snafi AE and Thwaini MM. Nephro- protective effects of Arabian medicinal plants (part 1). Research Journal of Pharmaceutical, Biological and Chemical Sciences 2018; 9(5): 1504-1511.
- [18]. Al-Snafi AE and Thwaini MM. Arabian medicinal plants with hepatoprotective activity (part 1). Research Journal of Pharmaceutical, Biological and Chemical Sciences 2018; 9(5): 1469-1497.
- [19]. Al-Snafi AE. Traditional uses of Iraqi medicinal plants. IOSR Journal of Pharmacy 2018; 8 (8): 32-96.
- [20]. Al-Snafi AE. Arabian medicinal plants with dermatological effects- plant based review (part 1). IOSR Journal of Pharmacy 2018; 8(10): 44-73.
- [21]. Al-Snafi AE. Chemical constituents, nutritional, pharmacological and therapeutic importance of *Juglans regia* A review. IOSR Journal of Pharmacy 2018; 8(11): 1-21.
- [22]. 244- Al-Snafi AE. Medicinal plants affected contractility of smooth muscles- A review . IOSR Journal of Pharmacy 2018; 8(11): 22-35.
- [23]. Al-Snafi AE, Majid WJ and Talab TA. Medicinal plants with antidiabetic effects An overview (Part 1). IOSR Journal of pharmacy 2019, 9(3): 9-46.
- [24]. Al-Snafi AE. Fritillaria imperialis- A review. IOSR Journal of pharmacy 2019, 9(3): 47-51.
- [25]. Al-Snafi AE, Talab TA and Majid WJ. Medicinal plants with central nervous activity An overview (Part 1). IOSR Journal of pharmacy 2019, 9(3): 52-102.
- [26]. Philippine Medicinal Plants, Badok, *Laphangium luteoalbum* (L.), http://www.stuartxchange.org/Badok.html
- [27]. The plant list, a working list of all plant species, Laphangium luteoalbum (L.) Tzvelev

http://www.theplantlist.org/tpl1.1/record/gcc-32755

- [28]. IT IS, *Pseudognaphalium luteoalbum* (L.) Hilliard & B.L. Burtt, https://www.itis. gov/ servlet/SingleRpt/SingleRpt?search_topic=TSN&search_value=507655#null
- [29]. US National Plant Germplasm System, Taxon: *Pseudognaphalium luteoalbum*, https://npgsweb.ars-grin.gov/gringlobal/taxonomydetail.aspx?411038
- [30]. The Euro+Med Plantbase Project, Laphangium luteoalbum (L.) Tzvelev, http://ww2.bgbm. org/EuroPlusMed/PTaxonDetail.asp?NameCache=Laphangium+luteoalbum
- [31]. Jersey cudweed (Pseudognaphalium luteoalbum) http://bios.conabio.gob.mx/ especies/ 6061461.pdf
- [32]. Davis PH (Ed.). Flora of Turkey and The East Aegean Islands. Edinburgh University Press, Edinburgh 1975. Vol 5: 97-100..
- [33]. Rahmatullah M, Ferdausi D, Mollik MAH, Jahan R, Chowdhuy MH and Haque WM. A survey of medicinal plants used by Kavirajes of Chalna area, Khulna district, Bangladesh. Afr J Tradit Complement Altern Med 2010; 7(2): 91–97.
- [34]. Manzur-ul-Kadir M, Kadir MF, Hossan S, Rahmatullah M. Medicinal plants of the Garo tribe inhabiting the Madhupur forest region of Bangladesh. Am Eurasian J Sustain Agric 2009; 3(2): 165-171.
- [35]. Al-Rawi A. Medicinal Plants of Iraq. Tech. Bull. No. 15. Ministry of Agriculture, Directorate General of Agricultural Research Projects 1964.
- [36]. Hartwell JL. Plants used against cancer- A survey. Lloydia 1967; 71: 30-34.
- [37]. Demirci B, Baser KHC and Duman H. The essential oil composition of *Gnaphalium luteo-album*. Chem Nat Comp 2009; 45 (3): 446–447.
- [38]. Sofowora A. Medicinal plants and traditional medicinal in Africa. 3rd ed. Nigeria: John Wiley, 1982.
- [39]. Uddin S, Sala Uddin GM, Begum M, Begum Y, Herrera-Calderon O, Islam M and Abdel-Daim MM.Inspection of phytochemical content and in vitro antioxidant profile of Gnaphalium luteoalbum L.: An unexplored phytomedicine. Journal of Pharmacy and Nutrition Sciences, 2017; 7: 136-146.
- [40]. Zheng X, Wang W, Piao H, Xu W, Shi H, Zhao C. The genus Gnaphalium L. (Compositae): Phytochemical and pharmacological characteristics. Molecules 2013; 15;18(7): 8298-318.
- [41]. Mericli AH. Flavonoids from *Gnaphalium luteo-album* L. J Fac Pharm Istanbul Univ 1980; 16: 84–87.
- [42]. Hassan RA. Biochemical studies on Gnaphalium luteo-album L. 2.-Flavonoids content.Mansoura Univ. (Egypt). Faculty of Agriculture 1988., http://agris.fao.org/agris-search/ search.do?recordID=EG9000662
- [43]. Aderogba MA, McGaw Lj, Bagla VP, Eloff JN and Abegaz BM. *In vitro* antifungal activity of the acetone extract and two isolated compounds from the weed, *Pseudognaphalium luteoalbum*. South African Journal of Botany 2014; 94: 74-78.
- [44]. Guadra P, HarborneJB and Waterman PG. Increase in the surface flavonoids and photosynthetic pigments in *Gnaphalium luteo-album* in response to UV-B radiation. Phytochemistry 1997;45(7):1377-1383.
- [45]. Kushwaha P. Chemical investigation of *Gnaphalium luteo-album*, Sida Actua, Laggera aurita and Erigeron bonariensis. PhD thesis, Department of Chemistry D.S.B. Campus, Kumaun University Nainital-263002, India, 2015.
- [46]. Kushwaha P. Chemical investigation of *Gnaphalium luteo-album*, *Sida Actua*, *Laggera aurita* and *Erigeron bonariensis*. PhD thesis, Department of Chemistry D.S.B. Campus, Kumaun University Nainital-263002, India, 2015.
- [47]. Akter R, Uddin SJ, Grice ID and Tiralongo E. Cytotoxic activity screening of Bangladeshi medicinal plant extracts. J Nat Med 2014; 68:246–252.
- [48]. Moustafa SMA, Menshawi BM, Wassel GM, Mahmoud K and Mounier MM. Screening of some wild and cultivated Egyptian plants for their free radical scavenging activity. International Journal of PharmTech Research 2014; 6(4): 1271-1278.

IOSR Journal of Pharmacy (IOSR-PHR) is UGC approved Journal with Sl. No. 3365, Journal No-62875

Ali Esmail Al-Snafi. "The Medical Benefit of Gnaphalium Luteoalbum-A Review." IOSR Journal of Pharmacy (IOSRPHR), vol. 9, no. 5, 2019, pp. 40-44.