



Nanotechnology Based Approach for Hepatocellular Carcinoma Targeting

Abdulsalam Alhalmi¹, Sarwar Beg^{1,*}, Kanchan Kohli^{1,*}, Md. Waris² and Tanuja Singh³

¹Department of Pharmaceutics, School of Pharmaceutical Education and Research, Jamia Hamdard, New Delhi, 110062, India, ²Department of Botany, Thakur Prasad Singh College, Patna, Magadh University, Bodh Gaya, India, ³University Department of Botany, Patliputra University, Patna, Bihar, India

ARTICLE HISTORY

Received: July 13, 2020
Revised: October 20, 2020
Accepted: October 20, 2020

DOI:
10.2174/1389450121999201209194524



Abstract: Hepatocellular carcinoma (HCC) is the primary liver cancer that has shown a high incidence and mortality rate worldwide among several types of cancers. A large variety of chemotherapeutic agents employed for the treatment have a limited success rate owing to their limited site-specific drug targeting ability. Thus, there is a demand to develop novel approaches for the treatment of HCC. With advancements in nanotechnology-based drug delivery approaches, the challenges of conventional chemotherapy have been continuously decreasing. Nanomedicines constituted of lipidic and polymeric composites provide a better platform for delivering and opening new pathways for HCC treatment. A score of nanocarriers such as surface-engineered liposomes, nanoparticles, nanotubes, micelles, quantum dots, *etc.*, has been investigated in the treatment of HCC. These nanocarriers are considered to be highly effective clinically for delivering chemotherapeutic drugs with high site-specificity ability and therapeutic efficiency. The present review highlights the current focus on the application of nanocarrier systems using various ligand-based receptor-specific targeting strategies for the treatment and management of HCC. Moreover, the article has also included information on the current clinically approved drug therapy for hepatocellular carcinoma treatment and updates of regulatory requirements for approval of such nanomedicines.

Keywords: Hepatocellular carcinoma, chemotherapy, nanocarriers, ligands, tumor targeting, quantum dots.

1. INTRODUCTION

Liver diseases are one of the leading causes of illness and death worldwide. Each year, 2 million deaths occur due to liver diseases [1], including liver fibrosis, hepatitis (A, B, and C), fatty liver, autoimmune hepatitis, and hepatocellular carcinoma (HCC) [2-4]. Liver tumors are frequent in occurrence and third in the most leading cause of cancer-related death worldwide [5]. Amongst various types of liver cancers, hepatic carcinoma is the most common, which is originated from the hepatocytes [6]. In other cases, secondary liver cancers are not originated from the liver but are formed due to metastasis from other parts of the body. Moreover, intrahepatic cholangiocarcinoma [7] and hepatoblastoma are other less common types of hepatic cancers reported in the literature [8, 9].

HCC is a very common form of malignant liver cancer. It is the sixth most common cancer in the world, accounting for more than 8,40,000 case deaths annually [10-12]. About 90% of HCC developed in patients with major risk factors are primarily infected with chronic hepatitis (type B and C viruses), liver cirrhosis, heavy alcohol consumption, smoking, non-alcoholic fatty liver, obesity, tobacco consumption,

and diabetes [13-15]. There are several conventional therapies available for HCC [16], surgical resection [17], ablation [18], transarterial chemoembolization [19], liver transplantation [20], radiation therapy [16], chemotherapy and combinatorial approaches [21].

Surgical interventions facilitating tumor recurrence by local metastasis [22], heat sink effect of ablation [23], complications of transarterial chemoembolization [24], immunosuppressive therapy side effect due to transplantation [25], hepatic toxicity of radiotherapy [26], and chemoresistance in HCC toward chemotherapy [27] are just a few examples of the current conventional strategies used for the treatment of HCC. Besides, these conventional therapies are also associated with many drawbacks like high treatment cost, lack of safety, poor patient compliance, and chances of tumor recurrence.

Conventional chemotherapy treatment, in particular, has several disadvantages, such as the inability to provide a sufficient concentration of therapeutic agents for liver disease, low targeting efficiency, poor tumor penetration, and/or the contribution to undesirable effects with systemic toxicity [28]. In order to avoid the serious and intolerable side-effects of the chemotherapy on normal tissues, the idea of exploration of novel tumor-targeting systems has typically taken momentum and greatly encourages the development of nanocarriers with targeting ability to achieve better efficacy with negligible undesirable effects [9]. The administration of a liver-specific drug delivery system helps in reducing the

*Address correspondence to these authors at the Department of Pharmaceutics, School of Pharmaceutical Education and Research, Jamia Hamdard, New Delhi 110062, India; ; Tel: +91 8447120434;
E-mail: sarwar.beg@gmail.com (S. Beg); Tel: +91 1126059860;
E-mail: prof.kanchankohli@gmail.com (K. Kohli)

side effects by reducing the distribution of the drugs to the non-target organs and increases the therapeutic efficacy by simultaneously increasing the drug levels in the target cells [29, 30].

Recently, with the rapid progress in nanotechnology development, it has been confirmed that drug delivery systems based on nanocarriers such as liposomes, polymeric micelle, quantum dote, dendrimers, carbon nanotube, nanoshells, and nanoparticles (Fig. 1). These systems have demonstrated great potential in the treatment of cancer by increasing the effectiveness of the drugs, reducing systemic toxicity, improving dissolution of the drugs, increasing stability and release behavior in order to achieve the best therapeutic efficiency [31-33].

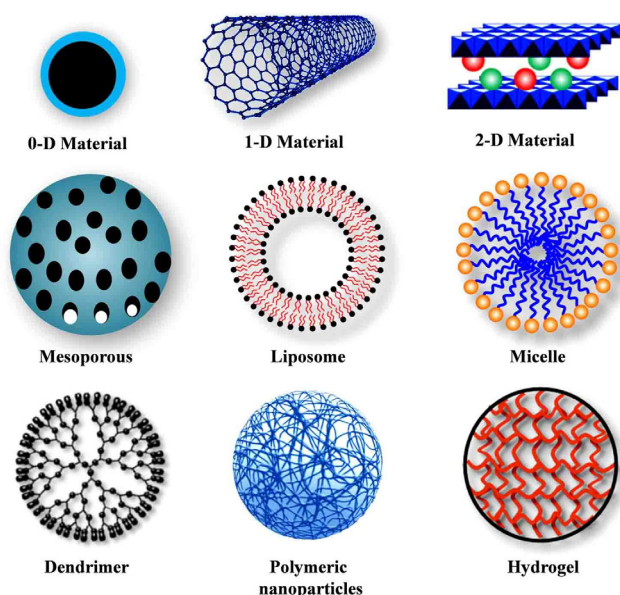


Fig. (1). Examples of nanocarriers drug delivery system for cancer targeting [34]. (A higher resolution / colour version of this figure is available in the electronic copy of the article).

The present review highlights various pathways used for targeting drug-loaded nanocarriers to liver carcinoma with some of the receptors specifically overexpressed on the surface of hepatocytes. Besides, the review focuses on the recent developments in the domain of nanocarriers with various functional modifications for drug targeting to HCC.

2. Various approaches used for liver targeting

The effective delivery of therapeutics to the liver can be obtained by passive targeting and active targeting approaches that increase the accumulation of the drugs at the targeted site, consequently, it may limit the adverse effects and improves the therapeutic efficacy of drug therapy [35] (Fig. 2). Passive targeting only increases the local concentration of the drugs within the tumor tissues by the enhanced permeability and retention effect (EPR) [36, 37]. Active targeting can be achieved by surface modification of the nanocarriers with specific targeting ligands such as proteins, antibodies,

peptides, and carbohydrates, which has the affinity to bind a specific-site on the liver cells and facilitates endocytotic uptake into the liver cells [4].

2.1. Passive Drug Targeting

Accumulation of nanocarriers at specific body sites is possible due to certain key features of the tumor microenvironment. Hence, such targeting is also known as passive drug targeting. The tumor microenvironment differs from normal tissue by features like the presence of highly vascular structure, oxygenation, pH, perfusion, and metabolic activity, which facilitate the accumulation of the drugs and nanocarriers in it [39]. These characteristic features facilitate the passive accumulation of nanocarrier therapeutics. The presence of fenestration in the endothelial wall of sinusoids capillaries of the liver and the absence of basal lamina favors the passive accumulation of nanocarriers therapeutics [40]. Nanocarriers with a size less than 200 nm can release through the sinusoidal fenestrations and facilitate passive liver targeting. Tumor-specific accumulation, also called the EPR effect, plays a significant role in the passive accumulation of the drugs and nanocarriers due to their extravasation through the leaky vasculature of the tumor (Fig. 3) [41]. The permeability and extravasation of macromolecules through the leaky tumor vasculature are enhanced by the EPR effect [42], and drainage of tumor tissues through an impaired lymphatic system is favored by retention of the nanostructured therapeutic carriers [43].

2.2. Active Drug Targeting

Drug delivery to the liver by an active targeting approach is a promising strategy for localizing the drugs to the tumor site. Active drug targeting is achieved by surface engineering of the nanocarriers with receptor-specific ligands such as peptides [45], carbohydrates [46], proteins [47], and antibodies [48], which specifically bind to the overexpressed receptors on the tumor cells [49]. Various surface receptors expressed on hepatocytes include asialoglycoprotein, glycyrrhizic acid, transferrin, folate, and integrin receptors [40, 50]. The targeting ligands facilitate the endocytotic uptake of drugs by receptors into the liver tumor cells, therefore, increase selective targeting of the chemotherapeutics to the tumor by avoiding undesirable side-effects [51].

3. LIGAND-RECEPTOR BASED ACTIVE TARGETING OF HCC TREATMENT

Ligand-receptor active targeting plays a critical role in the internalization of the drugs to the hepatocyte cells and subsequent endocytosis of anticancer drugs. It is one of the most common strategies used for targeting HCC, which helps improve the targeting ability. Some receptors that are overexpressed on HCC cells include asialoglycoprotein receptor, folate receptor, transferrin receptor, glycyrrhizic acid receptor and, integrin receptor, thus various ligands that can be attached to such receptors on the surface of hepatoma cells were used to design nanocarrier systems for effective targeting [52]. In this part of the review, a summary of the latest investigations carried out by the researchers for utili-

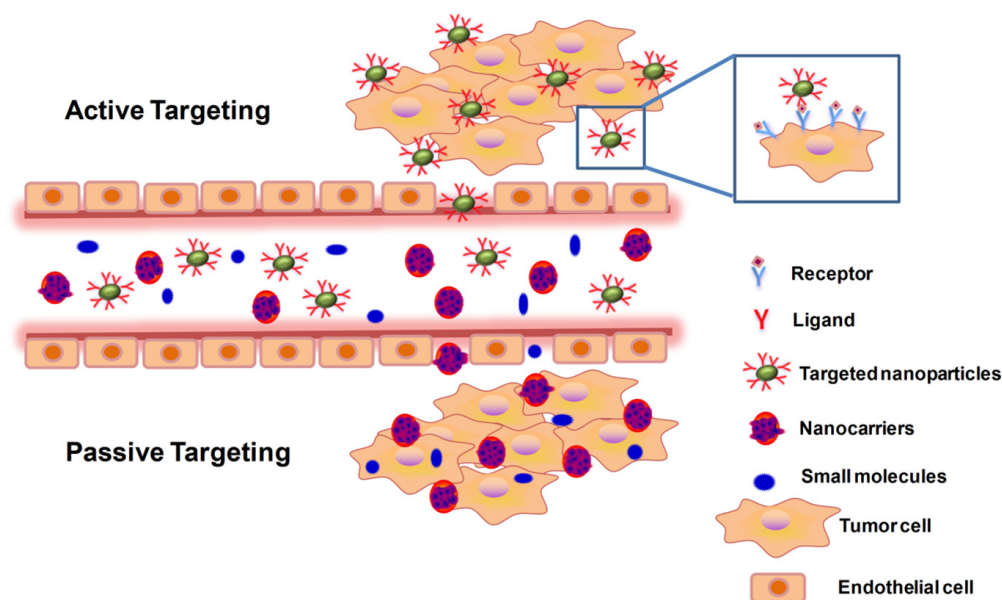


Fig. (2). Illustrative diagram for passive and active targeting to tumor tissues [38]. (A higher resolution / colour version of this figure is available in the electronic copy of the article).

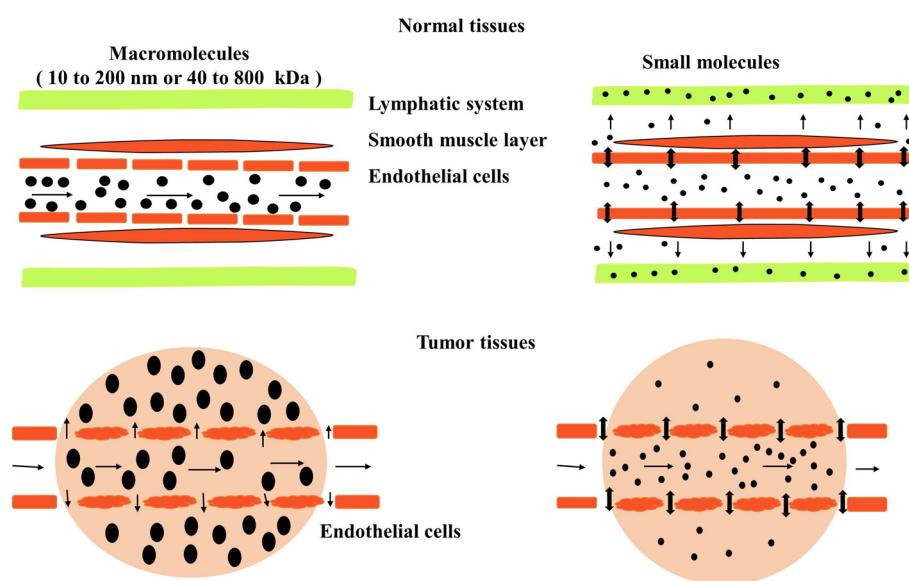


Fig. (3). Enhanced permeability and retention effect on tumor cells (EPR effect) [44]. (A higher resolution / colour version of this figure is available in the electronic copy of the article).

zing ligand-receptor mediated active targeting of chemotherapies for targeting of HCC is given in Table 1.

3.1. Asialoglycoprotein Receptors (ASGPR)

Since asialoglycoprotein receptors are present on hepatocytes and other non-hepatic cells, it is strongly expressed by the hepatocytes [4]. Various ligands such as asialofetuin, glycoproteins, carbohydrates, pullulan, and galactoside have been used to achieve the specific liver ASGPR targeting

[53]. Yousef *et al.* [54] reported the ability of galactosamine-anchored polyamidoamine dendrimers (PAMAM-s) loaded with a potent anticancer agent, curcumin, to achieve highly selective cellular uptake through the ASGPR-mediated endocytosis process, which improved the delivery of curcumin into the HCC cells. In another report, Xu *et al.* [55] prepared the solid lipid nanoparticles (SLN) of docetaxel-loaded with galactosylated dioleoylphosphatidylethanolamine, which showed higher cytotoxicity of SLN on the BEL7402 cell line over plain docetaxel (Taxotere[®]) and

Table 1. Summary of receptor-ligand based active targeting delivery systems for HCC to enhance the targeting effect.

Receptor	Ligands	Delivery	Drug	References
Asialoglyco-protein receptor	Galactosamine	Dendrimer	Curcumin	[54]
	Galactosylated-di-oleoylphosphatidyl- ethanolamine	Solid lipid nanoparticles	Docetaxel	[55]
	Galactosamine	Nanoparticles	Paclitaxel	[56]
Folate receptor	Folate	PEGylated-PLGA nanoparticles	Sorafenib	[59]
	Folate	Micelles	Doxorubicin	[60]
	Folate	Superparamagnetic iron oxide nanoparticles	Sorafenib	[61]
Transferrin receptor	Transferrin	Polymeric nanoparticles	Cisplatin/ Doxorubicin	[65]
	Transferrin	Conjugate	Doxorubicin	[66]
Glycyrrhetic acid receptor	Glycyrrhetic acid-dextran moiety	Dextran-based nanoscale	Curcumin	[70]
	Glycyrrhetic acid	Supramolecular gel	Curcumin	[71]
	Glycyrrhetic acid	Alginate nanoparticles	Doxorubicin	[72]
	Glycyrrhetic acid	Micelles.	Doxorubicin	[73]
Integrin receptor	RGD peptide (Arg-Gly-Asp)	Liposome	Paclitaxel	[78]

enhanced cellular uptake and accumulation of the drug in the hepatoma cells. Similarly, Liang *et al.* [56] developed the paclitaxel-loaded self-assembled nanoparticles conjugated with galactosamine (Gal-P/NPs). *In vitro* cell culture studies of Gal-P/NP on the HepG2 cells revealed comparative inhibition ($p < 0.05$) in cell growth as compared to the plain paclitaxel (Phyxsols).

3.2. Folate Receptors

These receptors are highly overexpressed on the surface of liver carcinoma cells, and their natural ligand is folic acid that has been used to target these receptors. Folate-conjugated drugs bind specifically to folate receptors and promote internalization of the drugs that bind with the folate receptors and uptake by receptor-mediated endocytosis mechanism [57]. The attached drug molecules can be released into the target tumor cells, where they can induce their cytotoxic activity [58]. Li *et al.* [59] developed the folate-PEGylated PLGA nanoparticles co-encapsulated with sorafenib (SRF/-FA-PEG-PLGA NP) for targeting HCC. The nanoparticles showed sustained release and improved cellular uptake of the drug during the *in vitro* study on Bel-7420 cancer cells. Besides, these nanoparticles effectively suppressed the proliferation of tumor cells and improved anticancer activity as compared to the free drug. Another study by Niu *et al.* [60] developed the doxorubicin-loaded polymeric micelles functionalized with folate ligand. *In vitro* cellular uptake study showed a controlled release profile of doxorubicin release and enhanced cytotoxicity of micelles on the Bel-7402 cells. Similarly, Zhang *et al.* [61] developed folic acid functionalized polymeric micelles-loaded with superparamagnetic iron oxide nanoparticles and sorafenib for enhanced anticancer activity against HCC. The developed nanoparticles exhibited superior inhibitory activity and *in vitro* apoptosis rate on HepG2 cells than nontargeted micelles.

3.3. Transferrin Receptor (TfR)

These receptors are the cell surface receptors overexpressed on many types of cancers, including HCC [62].

Therefore, this carrier protein can be utilized as a component of several carrier systems for chemotherapeutic agents [63]. TfR receptor expression on HCC is 100-times higher than the normal cells [64]. Hepatoma cells overexpressed with transferrin receptors have become promising targets for effective chemotherapy against HCC. In a study, Zhang *et al.* [65] prepared transferrin (Tf) modified polymeric nanoparticles for co-administration of cisplatin (DDP) and doxorubicin (DOX) for the treatment of hepatic carcinoma. The nanoparticles cytotoxicity assessed on the HepG2 cell line showed a better antitumor effect. Tf-DDP/DOX-NPs showed exceptional antitumor activity due to the combined action of two drugs and the ability to actively target the tumor cells through the Tf ligand. Similarly, Szwed *et al.* [66] demonstrated that doxorubicin-transferrin conjugated nanoparticles showed higher cytotoxicity on HepG2 cells as compared to the free doxorubicin and induced greater oxidative stress.

3.4. Glycyrrhetic Acid Receptor (GaR)

These receptors are overexpressed on the surface of hepatocytes and their ligand glycyrrhetic acid has been widely used to target drugs by different nanocarrier delivery systems, including micelles, nanoparticles, and liposomes [47, 67, 68]. Tian *et al.* [69] reviewed the role of GA and nanocarriers modified with GA as an efficient tool for hepatocyte targeted delivery for the treatment of HCC. Anirudhan and Binusreejayan [70] developed a dextran-based nanoscale drug carrier (GHDx) for curcumin delivery. Liver-directed curcumin is loaded in GHDx. *In vitro* cytotoxicity study on HepG2 and L929 cells demonstrated that GHDx-loaded with curcumin exhibited high toxicity with sustained drug release profile to liver cells. Chen *et al.* [71] formulated a glycyrrhetic acid-modified curcumin supramolecular gel, which exhibited good water solubility and sustained release delivery of curcumin in buffer solution under *in vitro* studies. *In vivo* studies showed enhanced cellular uptake and better inhibition ability on HepG2 cells. Zhang *et al.* [72] prepared doxorubicin-loaded glycyrrhetic acid-modified alginate nano-

particles, which revealed significantly higher concentration in the liver tumor induced in mice than nonglycyrrhetic acid-modified doxorubicin nanoparticles and plain doxorubicin. Similarly, in another study, Tain *et al.* [73] prepared doxorubicin-loaded glycyrrhetic acid-modified sulfated chitosan micelles, which demonstrated excellent *in vivo* inhibitory effect against HepG2 cells. The antitumor effect was extremely high with doxorubicin-loaded with the micelles than surface unmodified micelles.

3.5. Integrin Receptor (IgR)

These receptors are found in most types of human cancer, including HCC. Various types of integrins, in particular, $\alpha 1\beta 1$, $\alpha 5\beta 1$, and $\alpha 9\beta 1$, are expressed on the surface of normal hepatocyte to maintain a normal cell-matrix connection [74, 75]. In hepatocyte tumor cells, integrins $\alpha 3\beta 1$ and $\alpha 6\beta 4$ are overexpressed [76]. The RGD peptide (Arg-Gly-Asp) acts as a targeting ligand on the surface of nanocarrier systems to deliver an antitumor drug to hepatocytes [77]. Chen *et al.* [78] developed integrin receptor-targeted RGD-modified liposomal paclitaxel formulation by conjugating a specific Arg-Gly-Asp (RGD) ligand with 1,2-distearoyl-phosphatidylethanol-aminepolyethyleneglycol-2000. The study demonstrated the high efficacy of RGD-LP-PTX being easily uptaken by HepG2 cells than plain liposomes without RGD. *In vitro* evaluation of the formulation indicated inhibition of tumor growth in HepG2-bearing mice by RGD-LP-PTX formulation than LP-PTX or free PTX.

4. DIFFERENT NANOTECHNOLOGY-BASED CARRIERS FOR HCC TARGETING

Recently, innovation in the field of nanotechnology has exploited different novel nanotechnology approaches for the diagnosis and management of the HCC [79]. Novel nanocarriers are highly helpful to overcome the unwanted side-effects of chemotherapeutic agents by improving the pharmacokinetic profile of the drug by specific accumulation in the tumor site for enhancing the treatment effectiveness [80, 81]. In this part of the review, we provide a brief overview of the most recent examples of novel targeted delivery systems using various types of nanocarriers for delivering chemotherapeutic agents for HCC treatment [82]. Some of the extensively investigated nanocarriers for cancer treatment include nanoparticles, polymeric micelles, liposomes, carbon nanotubes, dendrimers, quantum dots, nanofibers, and lipid nanoparticulate carriers. Such nanosystems have shown great potential in liver cancer chemotherapy by enhancing the performance of the existing drugs, reducing their systemic side-effects, and increasing therapeutic efficacy [83-85]. Selective instances of the nanocarriers used for drug targeting to the HCC in literature is reported in this section of the manuscript (Table 2).

4.1. Nanoparticle-based Nanocarriers

Nanoparticles are small colloidal particles with a size range of 1 to 100 nm [86]. A wide range of NPs have been developed to target drugs, especially polymeric nanoparti-

cles, ceramic nanoparticles, metal nanoparticles, lipid nanoparticles, and carbon-based nanoparticles [87, 88]. Antitumor agents are either captured in or and adsorbed on the surface of NPs in order to efficiently transport the anticancer agent to hepatocytoma cells [89]. Modifying the surface of NPs can provide specific targeting ligands that allow NPs to control drug delivery to HCC with better therapeutic efficacy. Nanoparticles based delivery of anticancer drugs can improve solubility, reduce the dose and frequency of therapy and, above all, reduce the undesirable toxicities accompanied by antitumor drugs [90]. In addition, the delivery of nanoparticles, a combination of different anticancer drugs, can be loaded, making it a promising tool for the treatment of HCC.

Toma *et al.* [91] prepared superparamagnetic iron oxide nanoparticles (SPIONs) coated with polyvinyl alcohol (PVP) for delivery of sorafenib, which exhibited a higher loading capacity for sorafenib and long-term drug effect. The cytotoxicity of sorafenib with PVA/SPIONs has shown greater efficacy against cancer than that of free sorafenib alone. Karimia *et al.* [92] developed κ -carrageenan-cross-linked magnetic chitosan nanoparticles of sunitinib with high drug loading efficiency and a controlled release profile for effective management of HCC. Gao *et al.* [93] evaluated hollow alumina nanoparticles functionalized with hyaluronic acid loaded with paclitaxel (PAC) (HMHA-NP). *In vitro* cellular uptake of PAC-HMHA-NP was significantly high and *in vivo* studies have shown better antitumor activity by PAC-HMHA-NP than nonfunctionalized PAC-MHA-NP and pure PAC.

Zhao R *et al.* [94] prepared a pH-sensitive mesoporous silica nanoparticle for co-administration of sorafenib and ursolic acid. The prepared nanoparticles were decorated with chitosan and lactobionic acid (MSN-CS-LA nanocarriers) to target ASGPR in hepatocellular carcinoma cells. The study showed better bioavailability of the drug and effective targeting and synergistic cytotoxicity. *In vivo*, compared with UA or SO alone, the nanocomplex significantly reduced the tumor burden in hepatocellular carcinoma (HCC). Mathilde *et al.* [95] developed nanoparticles of human serum albumin loaded with doxorubicin with high loading capacity (88%) to inhibit the *in vivo* growth of human hepatocarcinoma cells; the study showed significant growth inhibition. W. Ni *et al.* [96] prepared nanoparticle of biotin/lactobionic acid modified poly (ethylene glycol)-poly (lactic-co-glycolic acid)-poly (ethylene glycol) (BLPP) copolymer for co-delivery of curcumin and 5-fluorouracil to enhance the treatment of hepatocellular carcinoma. The cytotoxicity study on animals and the hepatoma Hep G2 cell line showed higher cellular uptake and a synergistic anticancer effect. W. Gao *et al.* [97] prepared human serum albumin (HAS) nanoparticle surface modified with grafted folic acid for loading sorafenib (FA-HAS-SRF-NPs). *In vitro* study in the hepatocellular BEL-7402 showed enhanced cytotoxicity and increased safety in the normal liver LO2 cells. *In vivo*, the prepared nanoparticles showed effective antitumor activity toward nude mice bearing xenograft tumors without systemic toxicity.

Table 2. Nanocarrier system-based drug delivery for targeting HCC.

Nanocarrier System	Drug	Finding	References
Superparamagnetic iron oxide nanoparticles	Sorafenib	Higher encapsulation efficiency, sustained release and better anticancer efficiency.	[91]
Magnetic chitosan nanoparticles	Sunitinib	Sustained and controlled release.	[92]
Mesoporous hollow alumina nanoparticles	Paclitaxel	High cellular uptake and better antitumor effect.	[93]
Mesoporous silica nanoparticles	Sorafenib and ursolic acid,	Enhanced bioavailability of the drugs and efficient targeting and synergistic cytotoxicity.	[94]
Human serum albumin nanoparticles	Doxorubicin	Significant <i>in vivo</i> growth inhibition in multicellular tumor spheroid models (MCTS) of human hepatocarcinoma cells.	[95]
Nanoparticles	Curcumin and 5- fluorouracil	Higher cellular uptake and synergistic anticancer effects on the hepatoma cell line Hep G2.	[96]
Human serum albumin nanoparticle	Sorafenib	Enhanced cytotoxicity against hepatocellular BEL-7402 cells and increased the safety for normal liver LO2 cells.	[97]
Liposomes	Doxorubicin	Better cytotoxicity and pharmacokinetics profile were obtained.	[105]
Liposomes	Doxorubicin and lovastatin,	Higher growth inhibition and reduced toxicity in H22 mice hepatoma cell.	[106]
Single walled carbon nanotubes	Doxorubicin	Enhanced cytotoxicity in the HCC cell line SMMC-7721.	[110]
Carbon nanotubes	Doxorubicin	More efficient tumor targeting and higher cellular uptake in HepG2 cells.	[111]
PEGylated solid lipid nanoparticles	Sorafenib	Superior cytotoxicity, intracellular uptake and apoptotic activities on HepG2 cells. <i>In vivo</i> studies in BALB/c mice show superior pharmacokinetic profile and better targeting of the liver.	[117]
Nanostructured lipid carriers	Paclitaxel	Better cytotoxicity and pharmacokinetics profile were obtained.	[118]
Nanostructured lipid carriers	Sorafenib	Enhanced <i>in vitro</i> growth inhibition.	[119]
Polymeric micelles	Doxorubicin	Higher cellular uptake and cytotoxicity in HepG2 cell lines. <i>In vivo</i> studies in orthotopic H22 tumor-bearing mice show stronger tumor inhibition of GA-GEL-2 micelles.	[123]
Micelles	Sorafenib	Improved water solubility, sufficient uptake of sorafenib by Hep G2-Luc tumor-bearing mice and higher tumor growth inhibition.	[124]
Poliamidoamine dendrimer	Sorafenib	Higher uptake ability of dendrimer in ASGPR expressing the hepatoma cell line HepG2. Superior and long-lasting antitumor activity.	[128]
Dendrimers	Curcumin	Selective high cellular uptake <i>via</i> ASGPR mediated endocytosis and significantly enhanced the delivery of curcumin into the HCC cell lines.	[129]

4.2. Liposome Based Nanocarriers

Liposomes are a colloidal nanovesicle with phospholipid bilayer membrane, which have the ability to encapsulate various hydrophilic anticancer agents in their aqueous core and hydrophobic cytotoxic agents in their hydrophobic outer membrane [98]. Liposomes are effective nanocarriers for delivering many therapeutic drugs. They are biocompatible, biodegradable, and, because of their non-immunogenic properties, have a safe and effective therapeutic potential for clinical applications [99]. Many liposomal formulations of anti-neoplastic chemotherapy drugs have been approved for clinical use and are commercially available on the market, such as, Doxil® doxorubicin encapsulated in PEG-liposome, which is the first nano-drug product approved by FDA for clinical use [100]. PEGylated liposome has been widely used as a nanocarrier to improve the effectiveness of chemotherapy and is clinically effective with reduced toxicity [89]. Recently, research works focus on surface engineering by modifying the surface with ligands with different functional groups to achieve ligand binding. Targeted ligands enable specific targeting of tumor sites by targeting the lipo-

some towards specific receptors that are overexpressed in hepatoma cells, like folate receptor [101], CD-44 receptor [102], and transferrin receptor [103, 104].

Shah *et al.* [105] prepared doxorubicin-loaded palmitoylated arabinogalactan (PAG) liposomes. *In vitro* cytotoxicity study in HepG2 cell lines showed higher antitumor activity by PAG liposomes as compared to the non-PAG liposomes. A better pharmacokinetic profile was observed by PAG liposomes as compared to the non-PAG liposomes. T. Wang *et al.* [106] prepared liposome for co-delivery of doxorubicin and lovastatin. The *in vivo* study on H22 mice model mice hepatoma demonstrated that the co-loaded Doxorubicin-Lovastatin liposomes effectively inhibit the growth of the tumor with reducing toxicity.

4.3. Carbon Nanotube-based Nanocarriers

Carbon nanotubes are cylindrical hydrophobic tubes made of carbon atoms with a diameter of approximately 1-4 nm and length 1-100 nm. Depending on the number of graphene layers, nanotube can be single-walled nanotube or multiwalled carbon nanotubes [107]. Carbon nanotubes are

Table 3. FDA approved drug for liver cancer.

Drug	Developed by	Line Therapy	Target	References
Sorafenib (Nexavar)	Bayer	1	Multiple tyrosine kinase inhibitor, PDGF- α , β , VEGFR-1, 2, and 3.	[135]
Cabozantinib (Cabometyx)	Exelixis Ini.	2	Multiple tyrosine kinase inhibitor, c-Met, VEGFR2, AXL and RET.	[136]
Regorafenib (Stivarga)	Bayer	2	Multikinase inhibitor VEGFR2-TIE2.	[137, 138]
Lenvatinib (Lenvima)	Eisai	1	Multiple kinase inhibitors against the VEGFR1, 2, and 3.	[139]
Ramucirumab (Cyramza)	Eli Lilly	2	VEGFR2 inhibitor.	[140]
Nivolumab (Opdivo)	Bristol-Myers Squibb	2	PD-1 immune checkpoint inhibitor.	[141, 142]
Pembrolizumab (Keytruda)	Merck	2	PD-1 immune checkpoint inhibitor.	[143]

Table 4. Drugs ongoing development in phase 3 trials for HCC.

Drugs	Developed by	Phase Trials	Target	References
Durvalumab (Imfinzi)	AstraZeneca	3	Block the interaction of (PD-L1) with PD-1 (CD279).	[144]
Tremelimumab	Pfizer	3	CTLA-4, immune checkpoint inhibitor.	[145]
Atezolizumab (Tecentriq)	Genentech	3	PD-L1	[146]
Bevacizumab (Avastin)	Genentech	3	VEGF-A inhibitor	[147]
Nintedanib (Ofev)	Boehringer	3	VEGFR 1-3, FGFR and PDGFR	[148]
Tivantinib (ARQ197)	Arqule, Inc	3	Met inhibitor	[149]

widely applied for cancer diagnosis and therapy due to their unique features [108]. Moreover, carbon nanotubes have a unique physicochemical architecture that can be functionalized chemically on their surface by modifications or bounding with different targeting ligands to make them a promising platform for active targeting of tumor cells [109].

Z. Ji *et al.* [110] prepared chitosan modified single-walled carbon nanotubes loaded with doxorubicin; chitosan layer was bounded with folic acid for targeting folate receptor, highly expressed in cancer liver cells. The *in vitro* and *in vivo* studies in the HCC cell line SMMC-7721 showed that the DOX/FA/CHI/ SWNTs are much more effective in inhibiting cancer cells than free DOX. X. Qi *et al.* [111] developed galactosylated chitosan-grafted oxidized carbon nanotubes loaded doxorubicin. The *in vitro* studies on HepG2 cells showed that the prepared doxorubicin carbon nanotubes were more efficient in tumor targeting and higher cellular uptake.

4.4. Lipid Nanoparticulate Carrier

Particulate carriers (solid lipid nanoparticles, and nanostructured lipid carriers) have gained much attention for the loading of antitumor drugs for the treatment of various types of cancers [112]. Nanoparticulates are desirable as carriers of active drugs because they have a high carrying capacity, longer circulation time and facilitate the selective accumula-

tion of tumors due to the effect of increased permeability and retention (EPR) or active targeting [113]. Lipid nanoparticulate can improve oral bioavailability, control the release, and, target the anticancer with better physical stability [114]. Lipid nanoparticulate carriers are a promising candidate for anticancer targeting of the liver by lymphatic delivery [115]. NLCs show superior stability and loading capacity profile to overcome the possible drawbacks and limitations of SLNs [116]. Various anticancer drugs have been encapsulated either in SLN or in NLC.

L. Tunki *et al.* [117] prepared sorafenib loaded solid lipid nanoparticle conjugated with polyethylene glycol (PEGylated) galactose as a delivery carrier for HCC. Sorafenib loaded ligand conjugated nanoparticles show superior cytotoxicity, intracellular uptake and, apoptotic activities on HepG2 cells when compared with the free drug or non-ligand nanoparticle. *In vivo* studies on BALB/c mice show ligand conjugated SLN resulted in superior pharmacokinetic profile and better targeting of the liver by nanoparticles.

Harshita *et al.* [118] prepared paclitaxel-loaded nanostructured lipid carrier (PTX-NLC). PTX-NLCs showed higher antitumor activity than commercial formulation (Intaxel[®]) on the HepG2 cell line. The bioavailability of paclitaxel from PTX-NLCs was better than from PTX suspension. In another study, M.L. Bondi *et al.* [119] prepared nanostructured lipid carriers for delivery of sorafenib. The *in vitro*

studies showed that sorafenib loaded into NLC had more growth inhibition than that of free drug.

4.5. Polymeric Micelles Based Nanocarriers

Polymeric micelles are colloidal structures that contain amphiphilic copolymers. They have a hydrophobic core responsible for the uptake of water-insoluble drugs and a hydrophilic shell that ensures good stability drugs from the physiological environment [120]. The diameter of the polymeric micelles is less than 100 nm. Due to their range of nanometer sizes, their ability to self-assemble, stability, their ability to dissolve and transport hydrophobic drugs, polymeric micelles offer an attractive option for delivery of cytotoxic drugs to HCC [121]. High stability, low toxicity, and sustained release of the incorporated drug are the major advantages of polymeric micelles over surfactant-based micelles [122].

Fan *et al.* [123] prepared polymeric micelles-based gelatin functionalized with glycyrrhetic acid for delivery of doxorubicin (DOX-GA-GEL) polymeric micelles. The *in vivo* studies on HepG2 cell lines have shown higher cellular uptake and cytotoxicity than DOX-HCl. *In vivo* studies on mice with orthotopic H22 tumor have demonstrated the targeted ability and stronger tumor inhibition of GA-GEL-2 micelles to liver tissue compared with the free DOX. Su *et al.* [124] formulated micelles loaded with sorafenib for improved water solubility and enhanced anticancer activity, as observed through inhibition of tumor growth in the HepG2 tumor cells *in vivo*.

4.6. Dendrimer Based Nanocarriers

Dendrimers are highly branched three-dimensional synthetic macromolecules of various sizes (10-100 nm) [125]. The typical architectural structure of dendrimers includes a core, monomer branches, and functional surface groups, in which branching units are arranged around the central core, so dendrimers are candidates for different ligands and allow transport of a wide variety of drugs [126]. The modification of the chemical synthesis of the dendrimers improves the pharmacokinetics and the biocompatibility of the carrier and gives it promising properties for its use as a new carrier in the treatment of cancer [89, 127]. Maria *et al.* [128] prepared polyamidoamine dendrimer (PAMAM) loaded with sorafenib to target asialoglycoprotein receptor (ASGP-R). The prepared dendrimer functionalized with lactobionic acid as a ligand. *In vitro* studies conducted on HepG2 and HLE cell lines have shown a higher uptake ability of dendrimer in ASGPR expressing the hepatoma cell line HepG2 than in non-expressing HEL cells. *In vivo* cytotoxicity studies have shown that sorafenib loaded with dendrimer exhibits superior and long-lasting antitumor activity due to the kinetic release with delayed-release. Kuruvilla *et al.* [129] fabricated PAMAM dendrimers coupled with N-acetylgalactosamine ligands for targeting doxorubicin into hepatic cancer tissue. The result demonstrated that the targeted dendrimers show controlled drug release with improved therapeutic efficacy against tumors in mice as compared to free doxorubicin.

5. RECENT UPDATES ON THE DRUGS APPROVED FOR HCC TREATMENT

US Food and Drug Administration (FDA) has approved several drugs for use in patients with liver cancer [130]. In this clinical-stage, the systemic treatment for HCC with the multikinase inhibitor sorafenib is the most common treatment option [131]. Also, several immune checkpoint drugs are under development in phase 1, phase 2, and phase 3 trials, such as durvalumab, tremelimumab, atezolizumab, bevacizumab and tivantinib have shown significant positive results in clinical phase 1 and 2. However, clinical studies in phase 3 trials are required to confirm their efficacy for use in HCC [132-134] (Tables 3 and 4).

6. CHALLENGES WITH HCC TREATMENT AND FUTURE OPPORTUNITIES

Morbidity and mortality rates of HCC are significantly higher due to complexity that demands the development of an effective targeting therapeutic approach for treatment and prevention of HCC [150]. Despite this, the design of an effective nanocarrier system for HCC targeting faces challenges and only a few nanotherapeutic formulations have entered clinical trials [151, 152]. Despite advances in nanotechnologies for targeting of nanocarrier containing chemotherapeutic agents, yet many challenges and limitations are remaining. Toxicity is a major safety concern for applications of nanocarriers in clinical trials [153, 154]. In addition, the accumulation of nanocarriers in the liver and their poor clearance rate causes high toxicity. The discovery of new ligands or targeting molecules needed to deliver nanocarriers to hepatoma cells is a major challenge [155, 156]. For active targeting, the selection of the most suitable targeting agents, “ligands”, which are capable of binding the specific receptors expressed on the tumor cell surface, is the prerequisite for the successful transport of nanocarriers to tumorous liver tissues for avoiding systemic toxicity [153, 157, 158].

7. AUTHORS' INSIGHT ON THE TOPIC

Currently, the growing interest in the field of hepatocellular carcinoma diagnosis and nanocarrier based chemotherapy demonstrates a potential future scope for human application. Nanocarriers such as surface-engineered liposomes, nanoparticles, nanotubes, micelles, quantum dots, *etc.*, are some of the nanocarriers that are considered potentially useful as drug delivery agents in the treatment of HCC. The incidence of HCC is related to many sophisticated factors and molecular mechanisms, so we should comprehensively consider when to fabricate and investigate novel nanocarriers loaded therapy for HCC targeting. The design of an ideal drug carrier still needs more research and continuous efforts to understand the exact molecular mechanism of various nanocarrier materials, their possible long-term hazards, to provide a safe and reliable treatment for HCC. The perfect HCC targeted nanocarrier based drug delivery system should be able to maintain the drug in the liver tissue, specifically identifying the hepatocarcinoma cells. Thus, ligand-based hepatic receptor targeted drug delivery systems

are expected to play a significant role in HCC diagnosis and treatment. In the present, nanocarrier-based cancer-targeting therapy will face many challenges, such as surface engineered modification, multireceptor targeting, and drug loading efficacy, toxicology, immunotoxicology, biocompatibility testing, and, stability testing. The emerging nanocarrier chemotherapy targeting techniques will be theranostic with a multifunctional capability of simultaneous diagnosis and therapy.

CONCLUSION

Most traditional strategies for treating hepatocellular carcinoma experience poor targeting ability. Thus, it has gained increasing attention by the researchers for the exploration of new targeting receptors, ligands, and nanostructured systems to ensure efficient delivery of chemotherapeutic agents for the HCC treatment. Several studies in the literature reported mainly on animal or cell line models have shown the HCC-selective targeting ability of the nanocarriers based on their binding affinity to the target ligands-receptors, which further require exploration of their safety and efficacy through clinical studies in patients with HCC.

LIST OF ABBREVIATIONS

C-Met	= Tyrosine-protein kinase Met or hepatocyte growth factor receptor (HGFR)
CD274	= Cluster of differentiation 274
CTLA-4	= Cytotoxic T-lymphocyte-associated protein 4
FGFR	= Fibroblast growth factor receptors
PD-1	= Programmed cell death protein 1
PD-L1	= Programmed death-ligand 1
PDGF-R	= Platelet-derived growth factor receptors
VEGF-A	= Vascular endothelial growth factor A
VEGFR	= Vascular endothelial growth factor receptor

CONSENT FOR PUBLICATION

Not applicable.

FUNDING

None.

CONFLICT OF INTEREST

The authors declare no conflict of interest, financial or otherwise.

ACKNOWLEDGEMENTS

Declared none.

REFERENCES

- [1] Asrani SK, Devarbhavi H, Eaton J, Kamath PS. Burden of liver diseases in the world. *J Hepatol* 2019; 70(1): 151-71. <http://dx.doi.org/10.1016/j.jhep.2018.09.014> PMID: 30266282

- [2] Beg S, Kawish SM, Panda SK, *et al.* Nanomedicinal strategies as efficient therapeutic interventions for delivery of cancer vaccines. *Semin Cancer Biol* 2019. <http://dx.doi.org/10.1016/j.semcancer.2019.10.005> PMID: 31618687
- [3] Pandey P, Rahman M, Bhatt PC, *et al.* Implication of nano-antioxidant therapy for treatment of hepatocellular carcinoma using PL-GA nanoparticles of rutin. *Nanomedicine (Lond)* 2018; 13(8): 849-70. <http://dx.doi.org/10.2217/nnm-2017-0306> PMID: 29565220
- [4] Shilpi S. Drug targeting strategies for liver cancer and other liver diseases. *MOJ Drug Des Dev Ther* 2018; 2(4): 171-7. <http://dx.doi.org/10.15406/mojddt.2018.02.00044>
- [5] Siegel RL, Miller KD, Jemal A. Cancer statistics, 2019. *CA Cancer J Clin* 2019; 69(1): 7-34. <http://dx.doi.org/10.3322/caac.21551> PMID: 30620402
- [6] Marengo A, Rosso C, Bugianesi E. Liver cancer: Connections with obesity, fatty liver, and cirrhosis. *Annu Rev Med* 2016; 67(12): 103-17. <http://dx.doi.org/10.1146/annurev-med-090514-013832> PMID: 26473416
- [7] Rizvi S, Khan SA, Hallemeier CL, Kelley RK, Gores GJ. Cholangiocarcinoma - evolving concepts and therapeutic strategies. *Nat Rev Clin Oncol* 2018; 15(2): 95-111. <http://dx.doi.org/10.1038/nrclinonc.2017.157> PMID: 28994423
- [8] Sharma D, Subbarao G, Saxena R. Hepatoblastoma. *Semin Diagn Pathol* 2017; 34(2): 192-200. <http://dx.doi.org/10.1053/j.semmp.2016.12.015> PMID: 28126357
- [9] Usmani A, Mishra A, Ahmad M. Nanomedicines: a theranostic approach for hepatocellular carcinoma. *Artif Cells Nanomed Biotechnol* 2018; 46(4): 680-90. <http://dx.doi.org/10.1080/21691401.2017.1374282> PMID: 28884605
- [10] Duan W, Liu Y. Targeted and synergistic therapy for hepatocellular carcinoma: monosaccharide modified lipid nanoparticles for the co-delivery of doxorubicin and sorafenib. *Drug Des Devel Ther* 2018; 12: 2149-61. <https://dx.doi.org/10.2147/2FDD-DT.S166402> <http://dx.doi.org/10.2147/DDDT.S166402> PMID: 30034219
- [11] Greten TF, Lai CW, Li G, Staveley-O'Carroll KF. Targeted and immune-based therapies for hepatocellular carcinoma. *Gastroenterology* 2019; 156(2): 510-24. <http://dx.doi.org/10.1053/j.gastro.2018.09.051> PMID: 30287171
- [12] Pittman ME. Hepatocellular carcinoma: A practical review for the surgical pathologist. *Diagn Histopathol* 2018; 24(12): 500-7. <http://dx.doi.org/10.1016/j.mpdhp.2018.09.005>
- [13] Kar P. Risk factors for hepatocellular carcinoma in India. *J Clin Exp Hepatol* 2014; 4(S3) (Suppl. 3): S34-42. <http://dx.doi.org/10.1016/j.jceh.2014.02.155> PMID: 25755609
- [14] Ho BN, Pfeffer CM, Singh ATK. Update on nanotechnology-based drug delivery systems in cancer treatment. *Anticancer Res* 2017; 37(11): 5975-81. <http://dx.doi.org/10.21873/anticancerres.12044> PMID: 29061776
- [15] Lu C, Rong D, Zhang B, *et al.* Current perspectives on the immunosuppressive tumor microenvironment in hepatocellular carcinoma: challenges and opportunities. *Mol Cancer* 2019; 18(1): 130. <http://dx.doi.org/10.1186/s12943-019-1047-6> PMID: 31464625
- [16] Sapir E, Tao Y, Schipper M J, *et al.* Stereotactic body radiotherapy as an alternative to transarterial chemoembolization for hepatocellular carcinoma. *Int J Radiat Oncol Biol Phys* 2017; 100(1): 122-30.
- [17] Santambrogio R, Barabino M, Bruno S, *et al.* Surgical resection vs. Ablative therapies through a laparoscopic approach for hepatocellular carcinoma: A comparative study. *J Gastrointest Surg* 2018; 22(4): 650-60. <http://dx.doi.org/10.1007/s11605-017-3648-y> PMID: 29235004
- [18] Kang TW, Lim HK, Cha DI. Aggressive tumor recurrence after radiofrequency ablation for hepatocellular carcinoma. *Clical Mol Hepatol* 2017; 23(1): 95-101.
- [19] Silva JP, Berger NG, Tsai S, *et al.* Transarterial chemoembolization in hepatocellular carcinoma with portal vein tumor thrombosis: a systematic review and meta-analysis. *HPB (Oxford)* 2017; 19(8): 659-66.

- <http://dx.doi.org/10.1016/j.hpb.2017.04.016> PMID: 28552299
- [20] Sapisochin G, Bruix J. Liver transplantation for hepatocellular carcinoma: outcomes and novel surgical approaches. *Nat Rev Gastroenterol Hepatol* 2017; 14(4): 203-17. <http://dx.doi.org/10.1038/nrgastro.2016.193> PMID: 28053342
- [21] Gans JH, Lipman J, Golowa Y, Kinkhabwala M, Kaubisch A. Hepatic cancers overview: Surgical and chemotherapeutic options, how do Y-90 microspheres fit in? *Semin Nucl Med* 2019; 49(3): 170-81. <http://dx.doi.org/10.1053/j.semnuclmed.2019.01.001> PMID: 30954182
- [22] Belghiti J, Kianmanesh R. Surgical treatment of hepatocellular carcinoma. *HPB (Oxford)* 2005; 7(1): 42-9. <http://dx.doi.org/10.1080/13651820410024067> PMID: 18333160
- [23] Rhim H, Lim HK. Radiofrequency ablation of hepatocellular carcinoma: pros and cons. *Gut Liver* 2010; 4(1) (Suppl. 1): S113-8. <http://dx.doi.org/10.5009/gnl.2010.4.S1.S113> PMID: 21103289
- [24] Tu J, Jia Z, Ying X, *et al.* The incidence and outcome of major complication following conventional TAE/TACE for hepatocellular carcinoma. *Medicine (Baltimore)* 2016; 95(49): e5606. <https://dx.doi.org/10.1097%2FMD.0000000000005606> <http://dx.doi.org/10.1097/MD.0000000000005606> PMID: 27930585
- [25] Khorsandi SE, Heaton N. Optimization of immunosuppressive medication upon liver transplantation against HCC recurrence. *Transl Gastroenterol Hepatol* 2016; 1(80): 25. <http://dx.doi.org/10.21037/tgh.2016.03.18> PMID: 28138592
- [26] Kalogeridi M, Zygogianni A, Kyrgias G, Kouvaris J, Chatziioannou S, Kouloulis V. Role of radiotherapy in the management of hepatocellular carcinoma: A systematic review. *world. J Hepatol* 2015; 7(1): 101-12. <http://dx.doi.org/10.4254/wjh.v7.i1.101> PMID: 25135862
- [27] Lohitesh K, Chowdhury R, Mukherjee S. Resistance a major hindrance to chemotherapy in hepatocellular carcinoma: an insight. *Cancer Cell Int* 2018; 18(44): 44. <http://dx.doi.org/10.1186/s12935-018-0538-7> PMID: 29568237
- [28] Kydd J, Jadia R, Velpurisiva P, Gad A, Paliwal S, Rai P. Targeting strategies for the combination treatment of cancer using drug delivery systems. *Pharmaceutics* 2017; 9(4): 1-26. <http://dx.doi.org/10.3390/pharmaceutics9040046> PMID: 29036899
- [29] Baig B, Halim SA, Farrukh A, Greish Y, Amin A. Current status of nanomaterial-based treatment for hepatocellular carcinoma. *Biomed Pharmacother* 2019; 116: 108852. <http://dx.doi.org/10.1016/j.biopha.2019.108852> PMID: 30999152
- [30] Barkat MA, Beg S, Pottou FH, Ahmad FJ. Nanopaclitaxel therapy: an evidence based review on the battle for next-generation formulation challenges. *Nanomedicine (Lond)* 2019; 14(10): 1323-41. <http://dx.doi.org/10.2217/nmm-2018-0313> PMID: 31124758
- [31] Qiao W, Wang B, Wang Y, Yang L, Zhang Y, Shao P. Cancer therapy based on nanomaterials and nanocarrier systems. *J Nanomater* 2010; 2010: 1-9. <http://dx.doi.org/10.1155/2010/796303>
- [32] Rahman M, Beg S, Ahmed A, Swain S. Emergence of functionalized nanomedicines in cancer chemotherapy: recent advancements, current challenges and toxicity considerations. *Recent Pat Nanomed* 2013; 2(3): 128-39. <http://dx.doi.org/10.2174/18779123113036660002>
- [33] Rahman M, Beg S, Verma A, *et al.* Therapeutic applications of liposomal based drug delivery and drug targeting for immune linked inflammatory maladies: A contemporary view point. *Curr Drug Targets* 2017; 18(13): 1558-71. <http://dx.doi.org/10.2174/1389450118666170414113926> PMID: 28413980
- [34] Senapati S, Mahanta AK, Kumar S, Maiti P. Controlled drug delivery vehicles for cancer treatment and their performance. *Signal Transduct Target Ther* 2018; 3(1): 7. <http://dx.doi.org/10.1038/s41392-017-0004-3> PMID: 29560283
- [35] Zafar S, Beg S, Panda SK, *et al.* Novel therapeutic interventions in cancer treatment using protein and peptide-based targeted smart systems. *Semin Cancer Biol* 2019; (July): 1-19. <http://dx.doi.org/10.1016/j.semcancer.2019.08.023> PMID: 31442570
- [36] Zhou F, Teng F, Deng P, Meng N, Song Z, Feng R. Recent progress of nano-drug delivery system for liver cancer treatment. *Anticancer Agents Med Chem* 2018; 17(14): 1884-97. <http://dx.doi.org/10.2174/1871520617666170713151149> PMID: 28707574
- [37] Sun Y, Ma W, Yang Y, *et al.* Cancer nanotechnology: Enhancing tumor cell response to chemotherapy for hepatocellular carcinoma therapy. *Asian J Pharm Sci* 2019; 14(6): 581-94. <http://dx.doi.org/10.1016/j.ajps.2019.04.005> PMID: 32104485
- [38] Bhushan B, Khanadeev V, Khlebtsov B, Khlebtsov N, Gopinath P. Impact of albumin based approaches in nanomedicine: Imaging, targeting and drug delivery. *Adv Colloid Interface Sci* 2017; 246: 13-39. <http://dx.doi.org/10.1016/j.cis.2017.06.012> PMID: 28716187
- [39] Hirsjärvi S, Passirani C, Benoit JP. Passive and active tumour targeting with nanocarriers. *Curr Drug Discov Technol* 2011; 8(3): 188-96. <http://dx.doi.org/10.2174/157016311796798991> PMID: 21513482
- [40] Barkat A, Beg S, Panda SKS, S Alharbi K, Rahman M, Ahmed FJ. Functionalized mesoporous silica nanoparticles in anticancer therapeutics. *Semin Cancer Biol* 2019; 1-32. <http://dx.doi.org/10.1016/j.semcancer.2019.08.022> PMID: 31442571
- [41] Bazak R, Houry M, Achy SE, Hussein W, Refaat T. Passive targeting of nanoparticles to cancer: A comprehensive review of the literature. *Mol Clin Oncol* 2014; 2(6): 904-8. <http://dx.doi.org/10.3892/mco.2014.356> PMID: 25279172
- [42] Dutta R, Mahato RI. Recent advances in hepatocellular carcinoma therapy. *Pharmacol Ther* 2017; 173: 106-17. <http://dx.doi.org/10.1016/j.pharmthera.2017.02.010> PMID: 28174094
- [43] Fang J, Nakamura H, Maeda H. The EPR effect: Unique features of tumor blood vessels for drug delivery, factors involved, and limitations and augmentation of the effect. *Adv Drug Deliv Rev* 2011; 63(3): 136-51. <http://dx.doi.org/10.1016/j.addr.2010.04.009> PMID: 20441782
- [44] Li R, Zheng K, Yuan C, Chen Z, Huang M. Be active or not: The relative contribution of active and passive tumor targeting of nanomaterials. *Nanotheranostics* 2017; 1(4): 346-57. <http://dx.doi.org/10.7150/ntno.19380> PMID: 29071198
- [45] Goodman SL, Hölzemann G, Sulyok GA, Kessler H. Nanomolar small molecule inhibitors for α v β 6, α v β 5, and α v β 3 integrins. *J Med Chem* 2002; 45(5): 1045-51. <http://dx.doi.org/10.1021/jm0102598> PMID: 11855984
- [46] Seymour LW, Ferry DR, Anderson D, *et al.* Cancer Research Campaign Phase I/II Clinical Trials Committee. Hepatic drug targeting: phase I evaluation of polymer-bound doxorubicin. *J Clin Oncol* 2002; 20(6): 1668-76. <http://dx.doi.org/10.1200/JCO.2002.20.6.1668> PMID: 11896118
- [47] Choi CHJ, Alabi CA, Webster P, Davis ME. Mechanism of active targeting in solid tumors with transferrin-containing gold nanoparticles. *Proc Natl Acad Sci USA* 2010; 107(3): 1235-40. <http://dx.doi.org/10.1073/pnas.0914140107> PMID: 20080552
- [48] Bazak R, Houry M, El Achy S, Kamel S, Refaat T. Cancer active targeting by nanoparticles: a comprehensive review of literature. *J Cancer Res Clin Oncol* 2015; 141(5): 769-84. <http://dx.doi.org/10.1007/s00432-014-1767-3> PMID: 25005786
- [49] Lammers T, Kiessling F, Hennink WE, Storm G. Drug targeting to tumors: principles, pitfalls and (pre-) clinical progress. *J Control Release* 2012; 161(2): 175-87. <http://dx.doi.org/10.1016/j.jconrel.2011.09.063> PMID: 21945285
- [50] Mohamed NK, Hamad MA, Hafez MZE, Wooley KL, Elsabahy M. Nanomedicine in management of hepatocellular carcinoma: Challenges and opportunities. *Int J Cancer* 2017; 140(7): 1475-84. <http://dx.doi.org/10.1002/ijc.30517> PMID: 27861850
- [51] Yoo J, Park C, Yi G, Lee D, Koo H. Active Targeting strategies using biological ligands for nanoparticle drug delivery systems. *Cancers (Basel)* 2019; 11(5): 11050640. <https://dx.doi.org/10.3390%2Fcancers11050640> <http://dx.doi.org/10.3390/cancers11050640> PMID: 31072061
- [52] Li M, Zhang W, Wang B, Gao Y, Song Z, Zheng QC. Ligand-based targeted therapy: a novel strategy for hepatocellular

- carcinoma. *Int J Nanomedicine* 2016; 11: 5645-69.
<http://dx.doi.org/10.2147/IJN.S115727> PMID: 27920520
- [53] D'Souza AA, Devarajan PV. Asialoglycoprotein receptor mediated hepatocyte targeting - strategies and applications. *J Control Release* 2015; 203: 126-39.
<http://dx.doi.org/10.1016/j.jconrel.2015.02.022> PMID: 25701309
- [54] Yousef S, Alsaab HO, Sau S, Iyer AK. Development of asialoglycoprotein receptor directed nanoparticles for selective delivery of curcumin derivative to hepatocellular carcinoma. *Heliyon* 2018; 4(12): e01071.
<http://dx.doi.org/10.1016/j.heliyon.2018.e01071> PMID: 30603704
- [55] Xu Z, Chen L, Gu W, *et al.* The performance of docetaxel-loaded solid lipid nanoparticles targeted to hepatocellular carcinoma. *Biomaterials* 2009; 30(2): 226-32.
<http://dx.doi.org/10.1016/j.biomaterials.2008.09.014> PMID: 18851881
- [56] Liang HF, Chen CT, Chen SC, *et al.* Paclitaxel-loaded poly(gamma-glutamic acid)-poly(lactide) nanoparticles as a targeted drug delivery system for the treatment of liver cancer. *Biomaterials* 2006; 27(9): 2051-9.
<http://dx.doi.org/10.1016/j.biomaterials.2005.10.027> PMID: 16307794
- [57] Rahman M, Kumar V, Beg S, Sharma G, Katare OP, Anwar F. Emergence of liposome as targeted magic bullet for inflammatory disorders: current state of the art. *Artif Cells Nanomed Biotechnol* 2016; 44(7): 1597-608.
<http://dx.doi.org/10.3109/21691401.2015.1129617> PMID: 26758815
- [58] Gonen N, Assaraf YG. Antifolates in cancer therapy: structure, activity and mechanisms of drug resistance. *Drug Resist Updat* 2012; 15(4): 183-210.
<http://dx.doi.org/10.1016/j.drug.2012.07.002> PMID: 22921318
- [59] Li YJ, Dong M, Kong FM, Zhou JP. Folate-decorated anticancer drug and magnetic nanoparticles encapsulated polymeric carrier for liver cancer therapeutics. *Int J Pharm* 2015; 489(1-2): 83-90.
<http://dx.doi.org/10.1016/j.ijpharm.2015.04.028> PMID: 25888801
- [60] Niu C, Sun Q, Zhou J, Cheng D, Hong G. Folate-functionalized polymeric micelles based on biodegradable PEG-PDLLA as a hepatic carcinoma-targeting delivery system. *Asian Pac J Cancer Prev* 2011; 12(8): 1995-9.
 PMID: 22292640
- [61] Zhang L, Gong F, Zhang F, Ma J, Zhang P, Shen J. Targeted therapy for human hepatic carcinoma cells using folate-functionalized polymeric micelles loaded with superparamagnetic iron oxide and sorafenib *in vitro*. *Int J Nanomedicine* 2013; 8(1): 1517-24.
<http://dx.doi.org/10.2147/IJN.S43263> PMID: 23620667
- [62] Daniels TR, Bernabeu E, Rodríguez JA, *et al.* The transferrin receptor and the targeted delivery of therapeutic agents against cancer. *Biochim Biophys Acta* 2012; 1820(3): 291-317.
<http://dx.doi.org/10.1016/j.bbagen.2011.07.016> PMID: 21851850
- [63] Pascale R M, Miglio M R, De , Muroli R R, Simile M M. Transferrin and transferrin receptor gene expression and iron uptake in hepatocellular carcinoma in the rat. *Hepatology* 1998; 27(2): 452-61.
- [64] Sciort R, Paterson AC, van Eyken P, Callea F, Kew MC, Desmet VJ. Transferrin receptor expression in human hepatocellular carcinoma: an immunohistochemical study of 34 cases. *Histopathology* 1988; 12(1): 53-63.
<http://dx.doi.org/10.1111/j.1365-2559.1988.tb01916.x> PMID: 2836292
- [65] Zhang X, Li J, Yan M. Targeted hepatocellular carcinoma therapy: transferrin modified, self-assembled polymeric nanomedicine for co-delivery of cisplatin and doxorubicin. *Drug Dev Ind Pharm* 2016; 42(10): 1590-9.
<http://dx.doi.org/10.3109/03639045.2016.1160103> PMID: 26942448
- [66] Szwed M, Wrona D, Kania KD, Koceva-Chyla A, Marczak A. Doxorubicin-transferrin conjugate triggers pro-oxidative disorders in solid tumor cells. *Toxicol In Vitro* 2016; 31: 60-71.
<http://dx.doi.org/10.1016/j.tiv.2015.11.009> PMID: 26607004
- [67] Zhang J, Zhang M, Ji J, *et al.* Glycyrrhetic acid-mediated polymeric drug delivery targeting the acidic microenvironment of hepatocellular carcinoma. *Pharm Res* 2015; 32(10): 3376-90.
<http://dx.doi.org/10.1007/s11095-015-1714-2> PMID: 26148773
- [68] Cai Y, Xu Y, Chan HF, Fang X, He C, Chen M. Glycyrrhetic acid mediated drug delivery carriers for hepatocellular carcinoma therapy. *Mol Pharm* 2016; 13(3): 699-709.
<http://dx.doi.org/10.1021/acs.molpharmaceut.5b00677> PMID: 26808002
- [69] Tian Q, Wang X, Wang W, Zhang C, Liu Y, Yuan Z. Insight into glycyrrhetic acid: the role of the hydroxyl group on liver targeting. *Int J Pharm* 2010; 400(1-2): 153-7.
<http://dx.doi.org/10.1016/j.ijpharm.2010.08.032> PMID: 20813176
- [70] Anirudhan TS. Binusreejayan. Dextran based nanosized for the controlled and targeted delivery of curcumin to liver cancer cells. *Int J Biol Macromol* 2016; 88: 222-35.
<http://dx.doi.org/10.1016/j.ijbiomac.2016.03.040> PMID: 27012895
- [71] Chen G, Li J, Cai Y, Zhan J, Gao J, Song M. Y. S. & Z. Y. A glycyrrhetic acid-modified curcumin supramolecular hydrogel for liver tumor targeting therapy. *Sci Rep* 2017; 7(44210): 1-8.
<https://dx.doi.org/10.1038%2Fsrep44210>
<http://dx.doi.org/10.1038/srep44210>
- [72] Zhang C, Wang W, Liu T, *et al.* Doxorubicin-loaded glycyrrhetic acid-modified alginate nanoparticles for liver tumor chemotherapy. *Biomaterials* 2012; 33(7): 2187-96.
<http://dx.doi.org/10.1016/j.biomaterials.2011.11.045> PMID: 22169820
- [73] Tian Q, Wang XH, Wang W, Zhang CN, Wang P, Yuan Z. Self-assembly and liver targeting of sulfated chitosan nanoparticles functionalized with glycyrrhetic acid. *Nanomedicine (Lond)* 2012; 8(6): 870-9.
<http://dx.doi.org/10.1016/j.nano.2011.11.002> PMID: 22100756
- [74] Azzariti A, Mancarella S, Porcelli L, Quatrala A, Caligiuri A, Lupo L. Hepatic stellate cells induce hepatocellular carcinoma cell resistance to sorafenib through the laminin-332/a3 integrin axis recovery of focal adhesion kinase ubiquitination. *Hepatology* 2016; 64(6): 2103-17.
- [75] Wu Y, Qiao X, Qiao S, Yu L. Targeting integrins in hepatocellular carcinoma. *Expert Opin Ther Targets* 2011; 15(4): 421-37.
<http://dx.doi.org/10.1517/14728222.2011.555402> PMID: 21332366
- [76] Bergamini C, Sgarra C, Trerotoli P, *et al.* Laminin-5 stimulates hepatocellular carcinoma growth through a different function of alpha6beta4 and alpha3beta1 integrins. *Hepatology* 2007; 46(6): 1801-9.
<http://dx.doi.org/10.1002/hep.21936> PMID: 17948258
- [77] Danhier F, Le Breton A, Préat V. RGD-based strategies to target alpha(v) beta(3) integrin in cancer therapy and diagnosis. *Mol Pharm* 2012; 9(11): 2961-73.
<http://dx.doi.org/10.1021/mp3002733> PMID: 22967287
- [78] Chen L, Liu Y, Wang W, Liu K. Effect of integrin receptor-targeted liposomal paclitaxel for hepatocellular carcinoma targeting and therapy. *Oncol Lett* 2015; 10(1): 77-84.
<http://dx.doi.org/10.3892/ol.2015.3242> PMID: 26170980
- [79] Rahman M, Akhter S, Ahmad MZ, *et al.* Emerging advances in cancer nanotheranostics with graphene nanocomposites: opportunities and challenges. *Nanomedicine (Lond)* 2015; 10(15): 2405-22.
<http://dx.doi.org/10.2217/nnm.15.68> PMID: 26252175
- [80] Din FU, Aman W, Ullah I, *et al.* Effective use of nanocarriers as drug delivery systems for the treatment of selected tumors. *Int J Nanomedicine* 2017; 12: 7291-309.
<https://dx.doi.org/10.2147%2FIJN.S146315>
<http://dx.doi.org/10.2147/IJN.S146315> PMID: 29042776
- [81] Truong DH, Hoa Le VK, Pham TT, Dao AH, Dung Pham TP, Tran TH. Delivery of erlotinib for enhanced cancer treatment: An update review on particulate systems. *J Drug Deliv Sci Technol* 2019; 55: 101348.
<http://dx.doi.org/10.1016/j.jddst.2019.101348>
- [82] Peng Y, Bariwal J, Kumar V, Tan C, Mahato RI. Organic nanocarriers for delivery and targeting of therapeutic agents for cancer treatment. *Adv Ther* 2020; 3(2): 1900136.
<http://dx.doi.org/10.1002/adtp.201900136>
- [83] Chi X, Liu K, Luo X, Yin Z, Lin H, Gao J. Recent advances of nanomedicines for liver cancer therapy. *J Mater Chem B Mater Biol Med* 2020; 8(17): 3747-71.
<http://dx.doi.org/10.1039/C9TB02871D> PMID: 32215381

- [84] Rahman M, Kazmi I, Beg S, *et al.* Functionalized graphene-based nanomaterials for drug delivery and biomedical applications in cancer chemotherapy. *Nanoparticles in Pharmacotherapy* 2019; pp. 429-60.
<http://dx.doi.org/10.1016/B978-0-12-816504-1.00011-9>
- [85] Swain S, Sahu PK, Beg S, Babu SM. Nanoparticles for cancer targeting: current and future directions. *Curr Drug Deliv* 2016; 13(8): 1290-302.
<http://dx.doi.org/10.2174/1567201813666160713121122> PMID: 27411485
- [86] Pérez-Herrero E, Fernández-Medarde A. Advanced targeted therapies in cancer: Drug nanocarriers, the future of chemotherapy. *Eur J Pharm Biopharm* 2015; 93: 52-79.
<http://dx.doi.org/10.1016/j.ejpb.2015.03.018> PMID: 25813885
- [87] Sur S, Rathore A, Dave V, Reddy KR, Chouhan RS, Sadhu V. Recent developments in functionalized polymer nanoparticles for efficient drug delivery system. *Nano-Structures and Nano-Objects* 2019; 20: 100397.
<http://dx.doi.org/10.1016/j.nanoso.2019.100397>
- [88] Ma Z, Zhang B, Fan Y, *et al.* Traditional Chinese medicine combined with hepatic targeted drug delivery systems: A new strategy for the treatment of liver diseases. *Biomed Pharmacother* 2019; 117: 109128.
<http://dx.doi.org/10.1016/j.biopha.2019.109128> PMID: 31234023
- [89] Mishra N, Yadav NP, Rai VK, *et al.* Efficient hepatic delivery of drugs: novel strategies and their significance. *BioMed Res Int* 2013; 2013: 382184.
<http://dx.doi.org/10.1155/2013/382184> PMID: 24286077
- [90] Dang Y, Guan J. Nanoparticle-based drug delivery systems for cancer therapy. *Smart Mater Med* 2020; 1: 10-9.
<http://dx.doi.org/10.1016/j.smaim.2020.04.001>
- [91] Tom G, Philip S, Isaac R, Praseetha PK, Jiji SG, Asha VV. Preparation of an efficient and safe polymeric-magnetic nanoparticle delivery system for sorafenib in hepatocellular carcinoma. *Life Sci* 2018; 206: 10-21.
<http://dx.doi.org/10.1016/j.lfs.2018.04.046> PMID: 29709652
- [92] Karimi MH, Mahdavinia GR, Massoumi B. pH-controlled sunitinib anticancer release from magnetic chitosan nanoparticles crosslinked with κ -carrageenan. *Mater Sci Eng C* 2018; 91: 705-14.
<http://dx.doi.org/10.1016/j.msec.2018.06.019> PMID: 30033305
- [93] Gao Y, Hu L, Liu Y, Xu X, Wu C. Targeted delivery of paclitaxel in liver cancer using hyaluronic acid functionalized mesoporous hollow alumina nanoparticles. *BioMed Res Int* 2019; 2019: 2928507.
<http://dx.doi.org/10.1155/2019/2928507> PMID: 31119162
- [94] Zhao R, Li T, Zheng G, Jiang K, Fan L, Shao J. Simultaneous inhibition of growth and metastasis of hepatocellular carcinoma by co-delivery of ursolic acid and sorafenib using lactobionic acid modified and pH-sensitive chitosan-conjugated mesoporous silica nanocomplex. *Biomaterials* 2017; 143: 1-16.
<http://dx.doi.org/10.1016/j.biomaterials.2017.07.030> PMID: 28755539
- [95] Ménard M, Meyer F, Parkhomenko K, *et al.* Mesoporous silica templated-albumin nanoparticles with high doxorubicin payload for drug delivery assessed with a 3-D tumor cell model. *Biochim Biophys Acta, Gen Subj* 2019; 1863(2): 332-41.
<http://dx.doi.org/10.1016/j.bbagen.2018.10.020> PMID: 30391506
- [96] Ni W, Li Z, Liu Z, *et al.* Dual-targeting nanoparticles: Codelivery of curcumin and 5-fluorouracil for synergistic treatment of hepatocellular carcinoma. *J Pharm Sci* 2019; 108(3): 1284-95.
<http://dx.doi.org/10.1016/j.xphs.2018.10.042> PMID: 30395829
- [97] Gao W, Jia X, Wu J, *et al.* Preparation and evaluation of folate-decorated human serum albumin nanoparticles for the targeted delivery of sorafenib to enhance antihepatocarcinoma efficacy. *J Drug Deliv Sci Technol* 2019; 54(October): 101349.
<http://dx.doi.org/10.1016/j.jddst.2019.101349>
- [98] He H, Lu Y, Qi J, Zhu Q, Chen Z, Wu W. Adapting liposomes for oral drug delivery. *Acta Pharm Sin B* 2019; 9(1): 36-48.
<http://dx.doi.org/10.1016/j.apsb.2018.06.005> PMID: 30766776
- [99] Kiaie SH, Mojarad-Jabali S, Khalesh F, *et al.* Axial pharmaceutical properties of liposome in cancer therapy: Recent advances and perspectives. *Int J Pharm* 2020; 581(January): 119269.
<http://dx.doi.org/10.1016/j.ijpharm.2020.119269> PMID: 32234427
- [100] Barenholz Y. Doxil®--the first FDA-approved nano-drug: lessons learned. *J Control Release* 2012; 160(2): 117-34.
<http://dx.doi.org/10.1016/j.jconrel.2012.03.020> PMID: 22484195
- [101] Monteiro LOF, Fernandes RS, Oda CMR, *et al.* Paclitaxel-loaded folate-coated long circulating and pH-sensitive liposomes as a potential drug delivery system: A biodistribution study. *Biomed Pharmacother* 2018; 97(97): 489-95.
<http://dx.doi.org/10.1016/j.biopha.2017.10.135> PMID: 29091899
- [102] Wang L, Su W, Liu Z, *et al.* CD44 antibody-targeted liposomal nanoparticles for molecular imaging and therapy of hepatocellular carcinoma. *Biomaterials* 2012; 33(20): 5107-14.
<http://dx.doi.org/10.1016/j.biomaterials.2012.03.067> PMID: 22494888
- [103] Wei M, Xu Y, Zou Q, *et al.* Hepatocellular carcinoma targeting effect of PEGylated liposomes modified with lactoferrin. *Eur J Pharm Sci* 2012; 46(3): 131-41.
<http://dx.doi.org/10.1016/j.ejps.2012.02.007> PMID: 22369856
- [104] Fu J, Li W, Xin X, Chen D, Hu H. Transferrin modified nano-liposome co-delivery strategies for enhancing the cancer therapy. *J Pharm Sci* 2020; 109(8): 2426-36.
<http://dx.doi.org/10.1016/j.xphs.2019.11.013> PMID: 31760084
- [105] Shah SM, Goel PN, Jain AS, *et al.* Liposomes for targeting hepatocellular carcinoma: use of conjugated arabinogalactan as targeting ligand. *Int J Pharm* 2014; 477(1-2): 128-39.
<http://dx.doi.org/10.1016/j.ijpharm.2014.10.014> PMID: 25311181
- [106] Wang T, Jiang Y, Chu H, Liu X, Dai Y, Wang D. Doxorubicin and lovastatin co-delivery liposomes for synergistic therapy of liver cancer. *J Drug Deliv Sci Technol* 2019; 52(January): 452-9.
<http://dx.doi.org/10.1016/j.jddst.2019.04.045>
- [107] Peretz S, Regev O. Carbon nanotubes as nanocarriers in medicine. *Curr Opin Colloid Interface Sci* 2012; 17(6): 360-8.
<http://dx.doi.org/10.1016/j.cocis.2012.09.001>
- [108] Badea N, Craciun MM, Dragomir AS, *et al.* Systems based on carbon nanotubes with potential in cancer therapy. *Mater Chem Phys* 2020; 241: 122435.
<http://dx.doi.org/10.1016/j.matchemphys.2019.122435>
- [109] Eatemadi A, Daraee H, Karimkhanloo, H; Kouhi, M; Zarghami, N; Akbarzadeh, A; Mozhgan, A; Younes, H; Sang, W.J. Carbon nanotubes: Properties, synthesis, purification, and medical applications. *Nanoscale Res Lett* 2014; 9(1): 1-13.
<http://dx.doi.org/10.1186/1556-276X-9-393> PMID: 24380376
- [110] Ji Z, Lin G, Lu Q, *et al.* Targeted therapy of SMMC-7721 liver cancer in vitro and in vivo with carbon nanotubes based drug delivery system. *J Colloid Interface Sci* 2012; 365(1): 143-9.
<http://dx.doi.org/10.1016/j.jcis.2011.09.013> PMID: 21974923
- [111] Qi X, Rui Y, Fan Y, Chen H, Ma N, Wu Z. Galactosylated chitosan-grafted multiwall carbon nanotubes for pH-dependent sustained release and hepatic tumor-targeted delivery of doxorubicin in vivo. *Colloids Surf B Biointerfaces* 2015; 133: 314-22.
<http://dx.doi.org/10.1016/j.colsurfb.2015.06.003> PMID: 26123852
- [112] García-Pinel B, Porras-Alcalá C, Ortega-Rodríguez A, *et al.* Lipid-based nanoparticles: application and recent advances in cancer treatment. *Nanomaterials (Basel)* 2019; 9(4): 1-23.
<http://dx.doi.org/10.3390/nano9040638> PMID: 31010180
- [113] Talluri SV, Kuppasamy G, Karri VVSR, Tummala S, Madhupantula SV. Lipid-based nanocarriers for breast cancer treatment - comprehensive review. *Drug Deliv* 2016; 23(4): 1291-305.
<http://dx.doi.org/10.3109/10717544.2015.1092183> PMID: 26430913
- [114] Lim SB, Banerjee A, Önyüksel H. Improvement of drug safety by the use of lipid-based nanocarriers. *J Control Release* 2012; 163(1): 34-45.
<http://dx.doi.org/10.1016/j.jconrel.2012.06.002> PMID: 22698939
- [115] Ali Khan A, Mudassir J, Mohtar N, Darwis Y. Advanced drug delivery to the lymphatic system: lipid-based nanoformulations. *Int J Nanomedicine* 2013; 8: 2733-44.
<http://dx.doi.org/10.2174/IJN.S41521> PMID: 23926431
- [116] Mishra DK, Shandilya R, Mishra PK. Lipid based nanocarriers: a translational perspective. *Nanomedicine (Lond)* 2018; 14(7): 2023-50.
<http://dx.doi.org/10.1016/j.nano.2018.05.021> PMID: 29944981

- [117] Tunki L, Kulhari H, Vadithe LN, *et al.* Modulating the site-specific oral delivery of sorafenib using sugar-grafted nanoparticles for hepatocellular carcinoma treatment. *Eur J Pharm Sci* 2019; 137: 104978. <http://dx.doi.org/10.1016/j.ejps.2019.104978> PMID: 31254645
- [118] Harshita; Barkat, A.; Beg, S.; Pottoo, F. H.; Siddiqui, S.; Ahmad, F. J. Paclitaxel-loaded nanolipidic carriers with improved oral bioavailability and anticancer activity against human liver carcinoma. *AAPS PharmSciTech* 2019; 20(2): 1-14. <http://dx.doi.org/10.1208/s12249-019-1304-4>
- [119] Bondi ML, Botto C, Amore E, *et al.* Lipid nanocarriers containing sorafenib inhibit colonies formation in human hepatocarcinoma cells. *Int J Pharm* 2015; 493(1-2): 75-85. <http://dx.doi.org/10.1016/j.ijpharm.2015.07.055> PMID: 26211902
- [120] Shi H, van Steenberg MJ, Lou B, Liu Y, Hennink WE, Kok RJ. Folate decorated polymeric micelles for targeted delivery of the kinase inhibitor dactolisib to cancer cells. *Int J Pharm* 2020; 582: 119305. <http://dx.doi.org/10.1016/j.ijpharm.2020.119305> PMID: 32278056
- [121] Hanafy NAN, Quarta A, Ferraro MM, *et al.* Polymeric nano-micelles as novel cargo-carriers for LY2157299 liver cancer cells delivery. *Int J Mol Sci* 2018; 19(3): 1-13. <http://dx.doi.org/10.3390/ijms19030748> PMID: 29509706
- [122] Yokoyama M. Clinical applications of polymeric micelle carrier systems in chemotherapy and image diagnosis of solid tumors. *J Exp Clin Med* 2011; 3(4): 151-8. <http://dx.doi.org/10.1016/j.jecm.2011.06.002>
- [123] Fan D, Yu J, Yan R, *et al.* Preparation and evaluation of doxorubicin-loaded micelles based on glycyrrhetic acid modified gelatin conjugates for targeting hepatocellular carcinoma. *J Chin Pharm Sci* 2018; 27(8): 530-9. <http://dx.doi.org/10.5246/jcps.2018.08.054>
- [124] Su Y, Wang K, Li Y, *et al.* Sorafenib-loaded polymeric micelles as passive targeting therapeutic agents for hepatocellular carcinoma therapy. *Nanomedicine (Lond)* 2018; 13(9): 1009-23. <http://dx.doi.org/10.2217/nmm-2018-0046> PMID: 29630448
- [125] Ambekar RS, Choudhary M, Kandasubramanian B. Recent advances in dendrimer-based nanoplatform for cancer treatment: A review. *Eur Polym J* 2020; 126: 109546. <http://dx.doi.org/10.1016/j.eurpolymj.2020.109546>
- [126] Pedziwiatr-Werbicka E, Milowska K, Dzimtruk V, Ionov M, Shcharbin D, Bryszewska M. Dendrimers and hyperbranched structures for biomedical applications. *Eur Polym J* 2019; 119(April): 61-73. <http://dx.doi.org/10.1016/j.eurpolymj.2019.07.013>
- [127] Jędrzak A, Grzeszkowiak BF, Coy E, *et al.* Dendrimer based theranostic nanostructures for combined chemo- and photothermal therapy of liver cancer cells *in vitro*. *Colloids Surf B Biointerfaces* 2019; 173(173): 698-708. <http://dx.doi.org/10.1016/j.colsurfb.2018.10.045> PMID: 30384266
- [128] Iacobazzi RM, Porcelli L, Lopodota AA, *et al.* Targeting human liver cancer cells with lactobionic acid-G(4)-PAMAM-FITC sorafenib loaded dendrimers. *Int J Pharm* 2017; 528(1-2): 485-97. <http://dx.doi.org/10.1016/j.ijpharm.2017.06.049> PMID: 28624661
- [129] Kuruvilla SP, Tiruchinapally G, Crouch AC, El Sayed MEH, Greve JM. Dendrimer-doxorubicin conjugates exhibit improved anticancer activity and reduce doxorubicin-induced cardiotoxicity in a murine hepatocellular carcinoma model. *PLoS One* 2017; 12(8): 1-24. <http://dx.doi.org/10.1371/journal.pone.0181944>
- [130] Ikeda M, Morizane C, Ueno M, Okusaka T, Ishii H, Furue J. Chemotherapy for hepatocellular carcinoma: current status and future perspectives. *Jpn J Clin Oncol* 2018; 48(2): 103-14. <http://dx.doi.org/10.1093/jcco/hyx180> PMID: 29253194
- [131] Baidoo SA, Wang Z, Sarkodie EK, Kesse S. Nanomedicinal delivery systems for intelligent treatment of hepatocellular carcinoma. *J Drug Deliv Sci Technol* 2019; 53: 101152. <http://dx.doi.org/10.1016/j.jddst.2019.101152>
- [132] Liu X, Qin S. Immune checkpoint inhibitors in hepatocellular carcinoma: opportunities and challenges. *Oncologist* 2019; 24(1) (Suppl. 1): S3-S10. <http://dx.doi.org/10.1634/theoncologist.2019-io-s1-s01> PMID: 30819826
- [133] Deng GL, Zeng S, Shen H. Chemotherapy and target therapy for hepatocellular carcinoma: New advances and challenges. *World J Hepatol* 2015; 7(5): 787-98. <http://dx.doi.org/10.4254/wjh.v7.i5.787> PMID: 25914779
- [134] Rimassa L. [135] Lorenza Rimassa. Drugs in development for hepatocellular carcinoma. *Gastroenterol Hepatol (N Y)* 2018; 14(9): 542-4. PMID: 30364332
- [135] Tak WY, Ryoo BY, Lim HY, *et al.* Phase I/II study of first-line combination therapy with sorafenib plus resminostat, an oral HDAC inhibitor, versus sorafenib monotherapy for advanced hepatocellular carcinoma in east Asian patients. *Invest New Drugs* 2018; 36(6): 1072-84. <http://dx.doi.org/10.1007/s10637-018-0658-x> PMID: 30198057
- [136] Abou-Alfa GK, Meyer T, Cheng AL, *et al.* Cabozantinib in patients with advanced and progressing hepatocellular carcinoma. *N Engl J Med* 2018; 379(1): 54-63. <http://dx.doi.org/10.1056/NEJMoa1717002> PMID: 29972759
- [137] Pelosof L, Lemery S, Casak S, *et al.* Benefit risk summary of regorafenib for the treatment of patients with advanced hepatocellular carcinoma that has progressed on sorafenib. *Oncologist* 2018; 23(4): 496-500. <http://dx.doi.org/10.1634/theoncologist.2017-0422> PMID: 29386313
- [138] Finn RS, Merle P, Granito A, *et al.* Outcomes of sequential treatment with sorafenib followed by regorafenib for HCC: Additional analyses from the phase III RESORCE trial. *J Hepatol* 2018; 69(2): 353-8. <http://dx.doi.org/10.1016/j.jhep.2018.04.010> PMID: 29704513
- [139] Kudo M, Finn RS, Qin S, *et al.* Lenvatinib versus sorafenib in first-line treatment of patients with unresectable hepatocellular carcinoma: a randomised phase 3 non-inferiority trial. *Lancet* 2018; 391(10126): 1163-73. [http://dx.doi.org/10.1016/S0140-6736\(18\)30207-1](http://dx.doi.org/10.1016/S0140-6736(18)30207-1) PMID: 29433850
- [140] Chau I, Peck-Radosavljevic M, Borg C, *et al.* Ramucirumab as second-line treatment in patients with advanced hepatocellular carcinoma following first-line therapy with sorafenib: Patient-focused outcome results from the randomised phase III REACH study. *Eur J Cancer* 2017; 81: 17-25. <http://dx.doi.org/10.1016/j.ejca.2017.05.001> PMID: 28591675
- [141] Scognamiglio G, De Chiara A, Parafioriti A, *et al.* Patient-derived organoids as a potential model to predict response to PD-1/PD-L1 checkpoint inhibitors. *Br J Cancer* 2019; 121(11): 979-82. <http://dx.doi.org/10.1038/s41416-019-0616-1> PMID: 31666667
- [142] Chiew Woon L, Joycelyn Jie Xin L, Su Pin C. Nivolumab for the treatment of hepatocellular carcinoma. *Expert Opin Biol Ther* 2020; 20(7): 687-93. <http://dx.doi.org/10.1080/14712598.2020.1749593> PMID: 32249635
- [143] Zhu AX, Finn RS, Edeline J, *et al.* KEYNOTE-224 investigators. Pembrolizumab in patients with advanced hepatocellular carcinoma previously treated with sorafenib (KEYNOTE-224): a non-randomised, open-label phase 2 trial. *Lancet Oncol* 2018; 19(7): 940-52. [http://dx.doi.org/10.1016/S1470-2045\(18\)30351-6](http://dx.doi.org/10.1016/S1470-2045(18)30351-6) PMID: 29875066
- [144] Akinleye A, Rasool Z. Immune checkpoint inhibitors of PD-L1 as cancer therapeutics. *J Hematol Oncol* 2019; 12(1): 92. <http://dx.doi.org/10.1186/s13045-019-0779-5> PMID: 31488176
- [145] Duffy AG, Ulahannan SV, Makorova-Rusher O, *et al.* Tremelimumab in combination with ablation in patients with advanced hepatocellular carcinoma. *J Hepatol* 2017; 66(3): 545-51. <http://dx.doi.org/10.1016/j.jhep.2016.10.029> PMID: 27816492
- [146] Karamchandani DM, Chetty R. Immune checkpoint inhibitor-induced gastrointestinal and hepatic injury: pathologists' perspective. *J Clin Pathol* 2018; 71(8): 665-71. <http://dx.doi.org/10.1136/jclinpath-2018-205143> PMID: 29703758
- [147] Mawalla B, Yuan X, Luo X, Chalya PL. Treatment outcome of anti-angiogenesis through VEGF-pathway in the management of gastric cancer: a systematic review of phase II and III clinical trials.

- BMC Res Notes 2018; 11(1): 21.
<http://dx.doi.org/10.1186/s13104-018-3137-8> PMID: 29329598
- [148] Roskoski R Jr. Properties of FDA-approved small molecule protein kinase inhibitors. *Pharmacol Res* 2019; 144(March): 19-50.
<http://dx.doi.org/10.1016/j.phrs.2019.03.006> PMID: 30877063
- [149] Rimassa L, Assenat E, Peck-Radosavljevic M, *et al.* Tivantinib for second-line treatment of MET-high, advanced hepatocellular carcinoma (METIV-HCC): a final analysis of a phase 3, randomised, placebo-controlled study. *Lancet Oncol* 2018; 19(5): 682-93.
[http://dx.doi.org/10.1016/S1470-2045\(18\)30146-3](http://dx.doi.org/10.1016/S1470-2045(18)30146-3) PMID: 29625879
- [150] Rahman M, Al-Ghamdi SA, Alharbi KS, *et al.* Ganoderic acid loaded nano-lipidic carriers improvise treatment of hepatocellular carcinoma. *Drug Deliv* 2019; 26(1): 782-93.
<http://dx.doi.org/10.1080/10717544.2019.1606865> PMID: 31357897
- [151] Hossen S, Hossain MK, Basher MK, Mia MNH, Rahman MT, Uddin MJ. Smart nanocarrier-based drug delivery systems for cancer therapy and toxicity studies: A review. *J Adv Res* 2018; 15: 1-18.
<http://dx.doi.org/10.1016/j.jare.2018.06.005> PMID: 30581608
- [152] Navya PN, Kaphle A, Srinivas SP, Bhargava SK, Rotello VM, Daima HK. Current trends and challenges in cancer management and therapy using designer nanomaterials. *Nano Converg* 2019; 6(1): 23.
<http://dx.doi.org/10.1186/s40580-019-0193-2> PMID: 31304563
- [153] Jiang W, Kim BYS, Rutka JT, Chan WCW. Advances and challenges of nanotechnology-based drug delivery systems. *Expert Opin Drug Deliv* 2007; 4(6): 621-33.
<http://dx.doi.org/10.1517/17425247.4.6.621> PMID: 17970665
- [154] Rahman M, Beg S. Hitting the target – refining anticancer nanomedicine development. *Eur Pharm Rev* 2019; 24(4): 1-4.
- [155] Ruman U, Fakurazi S, Masarudin MJ, Hussein MZ. Nanocarrier-based therapeutics and theranostics drug delivery systems for next generation of liver cancer nanodrug modalities. *Int J Nanomedicine* 2020; 15: 1437-56.
<http://dx.doi.org/10.2147/IJN.S236927> PMID: 32184597
- [156] Chen S, Cao Q, Wen W, Wang H. Targeted therapy for hepatocellular carcinoma: Challenges and opportunities. *Cancer Lett* 2019; 460: 1-9.
<http://dx.doi.org/10.1016/j.canlet.2019.114428> PMID: 31207320
- [157] Hare JI, Lammers T, Ashford MB, Puri S, Storm G, Barry ST. Challenges and strategies in anti-cancer nanomedicine development: An industry perspective. *Adv Drug Deliv Rev* 2017; 108: 25-38.
<http://dx.doi.org/10.1016/j.addr.2016.04.025> PMID: 27137110
- [158] Chen C, Wang G. Mechanisms of hepatocellular carcinoma and challenges and opportunities for molecular targeted therapy. *World J Hepatol* 2015; 7(15): 1964-70.
<http://dx.doi.org/10.4254/wjh.v7.i15.1964> PMID: 26244070