

The yeast
genome
directory



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Residents of a particular city notoriously have never visited most sights shown to tourists. In the same way, the complete genomic sequence of the brewer's yeast, *Saccharomyces cerevisiae*, has now been available for over a year, proving its usefulness to molecular biologists in a hundred quiet ways, without attracting the concerted attention that is its due. This *Directory* is intended to remedy that situation, by presenting papers on all the hitherto undescribed chromosomal sequences alongside an overview of the entire genomic sequence.

Like many a monument ignored by locals, familiarity has obscured what an achievement it represents. For one thing, it is still almost three times as large as any other genome sequenced so far. Then, too, this size necessitated a unique international collaboration which will surely inspire related projects in the future. Most importantly, it represents the first complete sequence of a eukaryotic genome, the last of the three superkingdoms of life to have a fully sequenced representative.

As such, it will be a rich mine of information for everyone concerned with all kinds of eukaryotic genomes, up to and including those of human beings. The opportunity this represents is equalled only by the challenge: dauntingly, of the 5,800 open reading frames in the sequence, the function of 45% cannot even be guessed. For now, we can only be grateful to the European Commission (DG XII – Life Sciences) for initiating and supporting, since 1989, the international project whose success has allowed *Nature* to present a unified account of this vital resource, and look forward to the flood of information that its study will surely release.

Nicholas Short

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The Yeast Genome Directory

This directory was made possible by a unique international collaboration between the 633 scientists whose names appear below. It represents both the first published description of the complete sequence of most chromosomes from *Saccharomyces cerevisiae*, and the first published overview of the entire sequence. As such, the authors would like future papers referring to the entire sequence and/or its contents to cite this directory; future papers referring to the sequence of individual chromosomes should refer to the papers listed at the head of page 9. The authors' affiliations appear in the papers describing the individual chromosomes.

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Zimmermann, M. Zimmermann, W.-W. Zhong, A. Zollner, E. Zumstein.

Overview of the yeast genome

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The collaboration of more than 600 scientists from over 100 laboratories to sequence the *Saccharomyces cerevisiae* genome was the largest decentralised experiment in modern molecular biology and resulted in a unique data resource representing the first complete set of genes from a eukaryotic organism. 12 million bases were sequenced in a truly international effort involving European, US, Canadian and Japanese laboratories. While the yeast genome represents only a small fraction of the information in today's public sequence databases, the complete, ordered and non-redundant sequence provides an invaluable resource for the detailed analysis of cellular gene function and genome architecture. In terms of throughput, completeness and information content, yeast has always been the lead eukaryotic organism in genomics; it is still the largest genome to be completely sequenced.

The *Yeast Genome Directory* presents the basic features of this sequence: the arrangement of the 6,000 genes on 16 chromosomes; a summary of the function of the encoded proteins; and a view of the genome's architecture, based on an exhaustive intra-genomic sequence comparison¹. The complete yeast sequence can be retrieved from a number of public databases, as well as from specialized World-Wide Web sites, which provide sophisticated query interfaces (see Box). These data are maintained and updated continuously. Although the form of the genome directory shown in this printed volume must present static information, its intention is to document the interpretation of the yeast genome shortly after its completion. To present the genome in a printable form, we have had to include a very limited selection of fact-oriented data. The *Yeast Genome Directory* cannot answer the question "What's in the yeast genome?" exhaustively, but summarizes what is known to be in it.

The sequence The final sequence was assembled from roughly 300,000 independent sequence reads, with error rates from 0.5 to 1%, resulting in an estimated error rate of the final sequence of less than 3 errors in 10,000 bases (0.03%). For the European Union part of the sequencing effort, a central database and informatics used to assemble, verify and analyse contiguous sequences submitted by the participating laboratories was provided by the Martinsrieder Institut für Protein Sequenzen (MIPS). The complete sequence was made available to the public on 24 April 1996. The first map represents the open reading frames (ORFs) of each of the successive chromosomes, at a uniform scale of 5 mm per kilobase. ORFs are named according to the location of the gene, using the convention Y (for yeast) followed by a letter denoting the chromosome (A for I, B for II, and so forth), a letter denoting the arm (R or L), a three-digit code ordering the ORFs from the centromere, and a letter denoting the coding strand (w or c).

Duplications The availability of the complete sequence of yeast allows us for the first time to examine the evolution of a eukaryotic species in a truly comprehensive manner. The footprints that indicate the evolutionary path taken by the yeast genome may be recognized by internal similarities between distinct regions of the present-day genome. Our approach to the inspection of these relationships is based on an all-against-all comparison of the genomic sequence data, applying local sequence alignments. Investigation of the yeast genome involved more than 24,000 blocks of 500 nucleotides. For each block, the six-frame translation into protein sequences was also generated, to allow for the concurrent comparison of DNA and amino-acid sequences².

Once an all-against-all matching of the yeast genome had been accomplished, duplication patterns within the genome could be investigated in a systematic way. The frequent, collinear block duplications found by our method seem to be an important consequence of the evolutionary development of *S. cerevisiae*. We have systematically inspected the genome for clusters of genes that have been produced by local duplication events. This involved evaluating the parameters that describe a cluster: its size; the degree of similarity of the duplicated ORFs; and the compactness of the cluster. Scanning this parameter-space at a sensitivity just above noise level (50% identity on the DNA level), we found that a window size of 25

kb and an enforced compactness of 10% of the coding region in that window generated an optimal representation of gene clusters. These criteria allowed us to identify 53 regions of clustered gene duplications, not including the well-known high level of similarity in the telomeric and subtelomeric regions. The second map shows the set of all collinear clusters of genes in the genome as a two-page fold-out. The significant number of gene duplications in yeast must reflect an evolutionarily successful strategy: gene duplications allow for evolutionary modifications in one of the copies without disturbing possibly vital functions of the other.

Gene function The computational analysis of the yeast genome is a challenging task³. The scientific assessment of raw sequence data aims to connect genetic entities to biological knowledge, either by computational analysis or by linking sequence-deduced information to other experimental evidence (such as a genetic locus). Sophisticated data modelling is required to reflect the correct relationship between the sequence and any associated information and to allow for the consistent integration of complex, heterogeneous biological data. For example, the precursor of the ADP/ATP carrier protein AAC2p is represented five times in the EMBL Nucleic Acid Data Library, although four of the coding sequences reported are identical. The current nucleic acid databases (EBI/NCBI/DBJ) contain 2,678 entries covering the yeast genome with a high degree of redundancy and inconsistency. Thus the traditional model of collecting

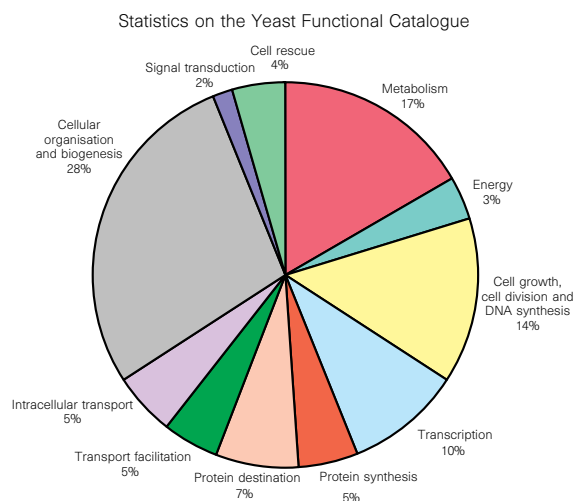


Figure 1 This shows the relative number of ORFs assigned to individual categories in the Gazetteer. There are eleven functional categories. Only proteins with a known function, or similarity or strong similarity to known proteins were assigned to one of the categories (similarities were measured by FASTA scores). In total, 3167 ORFs were assigned to at least one category. A single ORF can be assigned to more than one category.

database entries and presenting them as individual reports does not seem to be suitable for coping with the data analysis of complete genomes. Like the printed page, the traditional layout of the sequence databases is static, incorporating information from scientific annotation at the time of publication. Additional information extracted from the literature must be translated manually into the formal framework of a database entry. The knowledge of the functional properties of an uncharacterized gene may be published independently, and family members found in other organisms may allow for characterization by homology. Suitable models must thus be developed to cope with the data analysis of a complete genome, and to enable integrated views of the genomic sequence and associated information.

Only 43.3 % of the yeast genes are currently classified as 'functionally characterized', having experimentally well-investigated properties, being members of well-defined protein families, or displaying strong homology to proteins with known biochemical functions. The systematic functional analysis of these genes, currently in progress⁴, will identify the functions of many of the uncharacterized 'orphans'. Various experimental methods, including improved *in silico* analysis, will also increase dramatically the information content of the biological databases. Previously, individual attempts have been made to analyse specific chromosomes systematically by sequence analysis^{5,6}, and the results of an automated software system, 'Genecrunch', to 'crunch' the complete yeast genome have been offered as an Internet service (<http://genecrunch.sgi.com>).

To provide information on the biochemical and physiological context of protein function, we have compiled a gazetteer listing all the ORFs that can be related to well-understood functions. Similarities between biological sequences were measured by FASTA scores⁷: a FASTA score between 100 and 200 was defined as a 'weak similarity'; between 200 and a third of the self-score (the score of the protein when aligned with itself) for the protein was defined as a 'similarity'; and higher than a third of the self-score was defined as a 'strong similarity'. Weak similarities were not considered. In addition to the similarity scores, we have used pattern data⁸, including experimental data from the literature combined with genetic data, to characterize ORFs.

Each entry in the gazetteer lists the ORF name (defined as above), the gene name (if any), and the name or a short description of the protein. Entries are divided into 11 categories representing the cellular function of the individual ORFs (such as metabolism, energy or transcription), and each category is divided into sub-categories⁹. The yeast genome encodes about 5,800 proteins, less than half of which are 'known' in the sense that they have been genetically and biochemically well characterized. For about 20% of the remaining proteins, the experimental data are heterogeneous and provide only some indication of their functions *in vivo*. The remaining 38% either show similarities to other uncharacterized proteins or show no similarities at all. Defined categories provoke redundant entries, such as the classification of multifunctional enzymes. As gene names are used elsewhere in a very inconsistent way, we have used the name from the *Saccharomyces* genome database (see Box) register whenever possible. Gene names that are used twice for different ORF names are written in brackets.

Sequences of common evolutionary origin (homologous *sensu stricto*) reflect their relatedness by sequence similarities, and the organization of related primary, secondary and tertiary structures into groups ('families') remains the most powerful principle in sequence data analysis. We have used this principle to cluster proteins into families and superfamilies, following a previous scheme¹⁰, allowing us to cope with the many taxonomic complications inherent in protein evolution.

The World-Wide Web site of the Martinsrieder Institut für Protein Sequenzen yeast resource combines information generated by automated procedures with the results of systematic analysis by yeast researchers. Users can: (1) visualize chromosomes and selected regions to inspect genetic elements, such as ORFs, Ty's, tRNAs etc.; (2) receive detailed information on a yeast gene by searching with accession numbers, systematic codes, or gene names; (3) browse yeast genes according to their functional classification; (4) search for human homologues; (5) obtain information on functional properties; (6) download nucleic-acid or protein sequence data; (7) inspect up-to-date sequence homologies and alignments (FASTA database); (8) browse the family and superfamily classification of yeast proteins; (9) search

the yeast genome interactively for sequence patterns and sequence similarities; and (10) inspect the yeast genome for gene redundancy.

As a complement to the printed information in the *Yeast Genome Directory*, a CD-ROM compiled by Martinsrieder Institut für Protein Sequenzen, is available on request to subscribers of *Nature* and *Science* for the exploration of the yeast genome on a local, network-independent installation. The basic functionality of the retrieval software and the databases incorporated is equivalent to our World-Wide Web resource. The CD-ROM can be installed on Windows95, WindowsNT and Power-Macintosh operating systems, and is accompanied by a detailed description of its functionality, installation procedures and system requirements. □

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BOX Useful World-Wide Web addresses

Yeast databases

Munich Information Centre for Protein Sequences (MIPS)
<http://www.mips.biochem.mpg.de/mips/yeast/>

Yeast Protein Database (YPD)
<http://quest7.proteome.com/YPDhome.html>

Saccharomyces Genome Database (SGD)
<http://genome-www.stanford.edu/Saccharomyces/>

Specialized Yeast Databases

Sacch3D – Structural information for yeast proteins
<http://genome-www.stanford.edu/Sacch3D/>

Yeast Gene Duplications
<http://acer.gen.tcd.ie/~khwolfe/yeast/topmenu.html>

Related human disease genes (NIH XREFdb)
<http://www.ncbi.nlm.nih.gov/XREFdb/>

Genetic and physical maps (hyperlinked to biological information)
<http://genome-www.stanford.edu/cgi-bin/SGD/pgMAP/pgMap>

NIH yeast information page
<http://www.ncbi.nlm.nih.gov/Yeast/budding.html>

Schizosaccharomyces pombe
<http://www2.bio.uva.nl/pombe/>

Candida albicans information
<http://alces.med.umn.edu/Candida.html>

Common DNA and protein databases

European Bioinformatics Institute (EBI)
<http://www.ebi.ac.uk/services/services.html>

National Center for Biotechnology Information (NCBI)
<http://www.ncbi.nlm.nih.gov/>

DNA Data Bank of Japan (DDBJ)
<http://www.ddbj.nig.ac.jp/>

PIR-International
<http://www.mips.biochem.mpg.de>

SwissProt
<http://expasy.hcuge.ch/sprot/sprot-top.html>

References to be cited for the complete sequence and individual chromosomes

Complete sequence, The Yeast Genome Directory Goffeau, A. *et al. Nature* **387** (suppl.), 1–105 (1997)

Chromosome 1, The nucleotide sequence of chromosome I from *Saccharomyces cerevisiae* Bussey, H. *et al. Proc. Natl Acad. Sci. USA* **92**, 3809–3813 (1995)

EMBL accession numbers: U12980, L20125, L05146, L22015, L28920

Chromosome 2, Complete DNA sequence of yeast chromosome II

Feldmann, H. *et al. EMBO J.* **13**, 5795–5809 (1994)

EMBL accession numbers: Y08934, Z35762-Z35869, Z35870-Z36171

Chromosome 3, The complete DNA sequence of yeast chromosome III

Oliver, S. *et al. Nature* **357**, 38–46 (1992)

EMBL accession number: X59720

Chromosome 4, The nucleotide sequence of *Saccharomyces cerevisiae* chromosome IV

Jacq, C. *et al. Nature* **387** (suppl.), 75–78 (1997)

EMBL accession numbers: Z74051-Z74296, Z48008, Z74305-Z74385, Z49770, Z47814, Z68196, Z54075, Z49812, Z49209, Z46796, Z50111, Z47746, Z48758, Z48179, Z54139, Z50046, Z47813, Z46727, Z48784, Z68194, Z68195, Z48612, Z49701, Z68329, Z70202, Z68290, U51030, U51031, U28374, U32517, U51032, U28372, U28373, U32274, U33050, U33057, Z74389

Chromosome 5, The nucleotide sequence of *Saccharomyces cerevisiae* chromosome V

Dietrich, F. S. *et al. Nature* **387** (suppl.), 78–81 (1997)

EMBL accession numbers: U73806, U18795, L10830, U18779, U18530, U18778, U18796, U18813, U18814,

U18839, U18916, U18917, L10718, U18922

Chromosome 6, Analysis of the nucleotide sequence of chromosome VI from *Saccharomyces cerevisiae*

Murakami, Y. *et al. Nature Genet.* **10**, 261–268 (1995)

EMBL accession number: D50617

Chromosome 7, The nucleotide sequence of *Saccharomyces cerevisiae* chromosome VII

Tettelin, H. *et al. Nature* **387** (suppl.), 81–84 (1997)

EMBL accession numbers: X94357, Z72785-Z73081

Chromosome 8, Complete nucleotide sequence of *Saccharomyces cerevisiae* Chromosome VIII

Johnston, M. *et al. Science* **265**, 2077–2082 (1994)

EMBL accession numbers: U11583, U11582, U11581, U10555, U10400, U10399, U00062, U00061, U10556, U00060, U00059, U10398, U10397, U00027, U00028, U00030, U00029

Chromosome 9, The nucleotide sequence of *Saccharomyces cerevisiae* chromosome IX

Barrell, B. G. *et al. Nature* **387** (suppl.), 84–87 (1997)

EMBL accession number: Z47047

Chromosome 10, Complete nucleotide sequence of *Saccharomyces cerevisiae* Chromosome X

Galibert, F. *et al. EMBO J.* **15**, 2031–2049 (1996)

EMBL accession numbers: Z34098, Z49276-Z49662

Chromosome 11, Complete DNA sequence of yeast chromosome XI

Dujon, B. *et al. Nature* **369**, 371–378 (1994)

EMBL accession numbers: Z28001-Z28330

Chromosome 12, The nucleotide sequence of *Saccharomyces cerevisiae* chromosome XII

Johnston, M. *et al. Nature* **387** (suppl.), 87–90 (1997)

EMBL accession numbers: Z73106-Z73327, U53879, U51921, U17246, U14913, U19027, U20865, U17244, U17245, U17243, U17247, U20618, U19028, U19102, U19103, U19104, U19729, U20162, U20939, U21094, U22382, U22383

Chromosome 13, The nucleotide sequence of *Saccharomyces cerevisiae* chromosome XIII

Barrell, B. *et al. Nature* **387** (suppl.), 90–93 (1997)

EMBL accession numbers: Z50178, Z49218, Z49210, X80835, Z46660, Z46373, Z38114, Z46729, Z47816, Z48430, Z46659, Z49810, Z48613, Z49211, Z49213, Z48502, Z49703, Z48952, Z49259, Z49807, Z50179, Z49702, Z49273, Z48622, Z47071, Z49700, Z49808, Z47815, Z48755, Z49809, Z49939, Z48756, Z48639, Z49260, Z49704, X80836, Z49212, Z54141

Chromosome 14, The nucleotide sequence of *Saccharomyces cerevisiae* chromosome XIV and its evolutionary implications

Philippsen, P. *et al. Nature* **387** (suppl.), 93–98 (1997)

EMBL accession numbers: Z71277-Z71692

Chromosome 15, The nucleotide sequence of *Saccharomyces cerevisiae* chromosome XV

Dujon, B. *et al. Nature* **387** (suppl.), 98–102 (1997)

EMBL accession numbers: Z74743-Z75302

Chromosome 16, The nucleotide sequence of *Saccharomyces cerevisiae* chromosome XVI

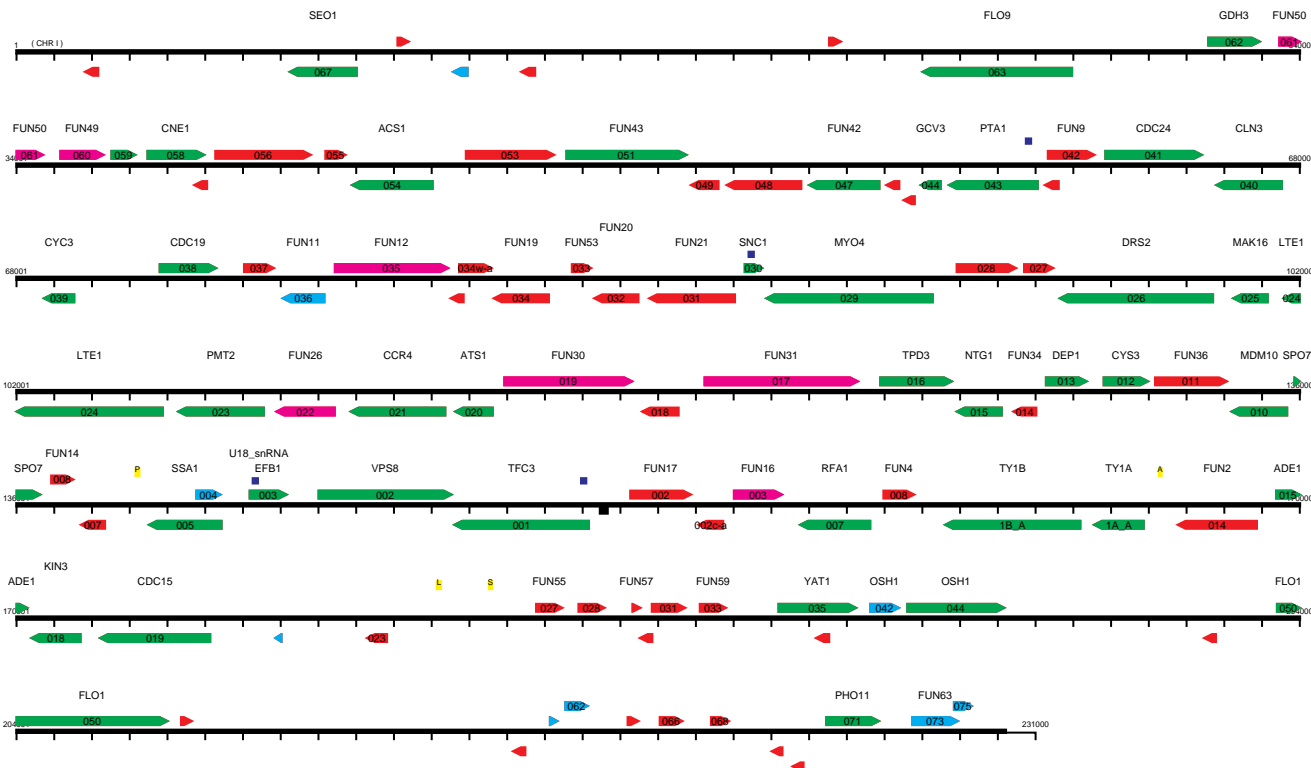
Bussey, H. *et al. Nature* **387** (suppl.), 103–105 (1997)

EMBL accession numbers: Z73521, Z73499-Z73638, U43703, U43503, U43281, U41849, U39205, U44030, U36624, U33335, Z48951, Z49919, Z49274, Z68111, Z49219, U51033, U32445, U40828, U40829, U28371, U25840, U25842, U25841, Z73537, Z73541

Legend of the chromosome displays

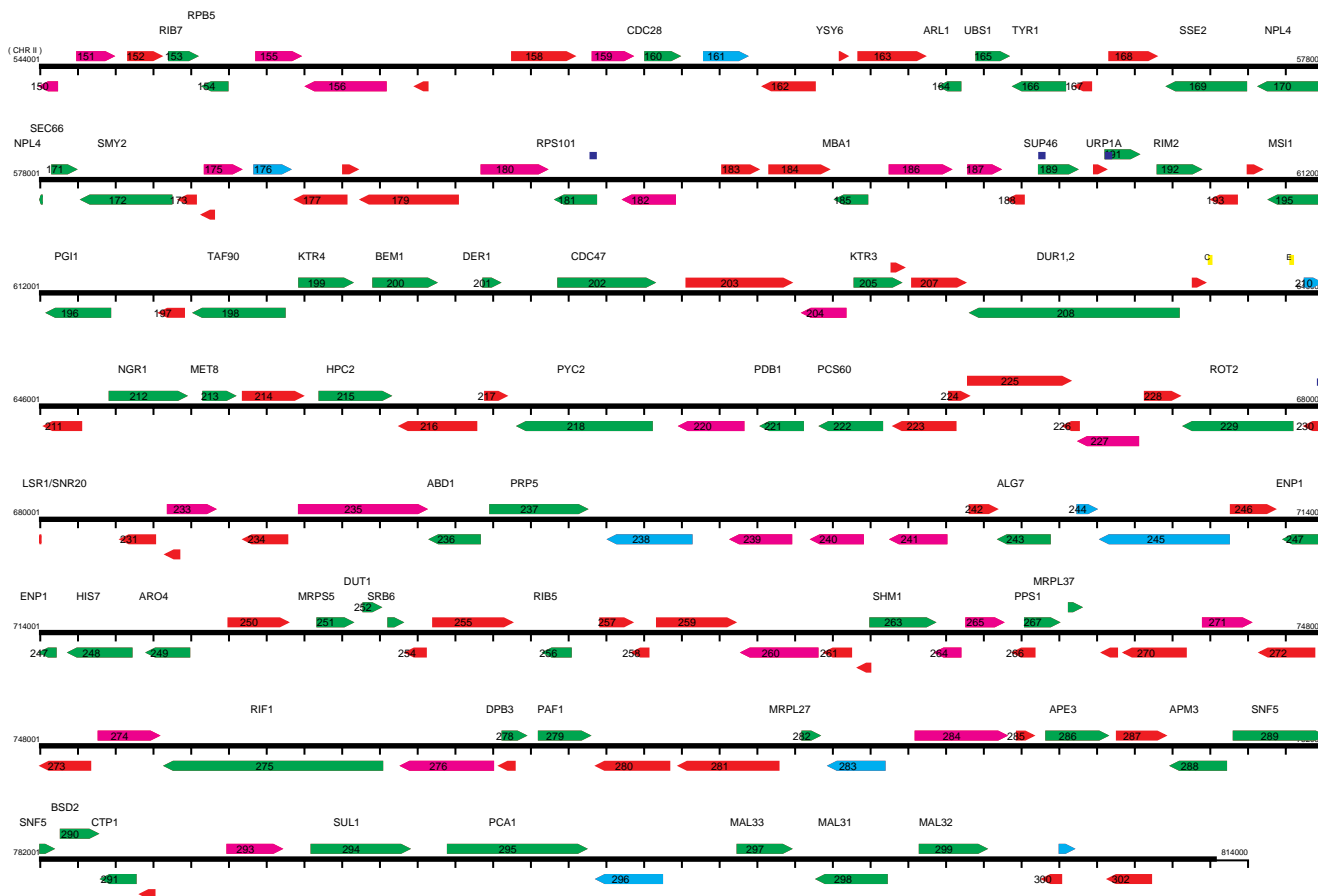
- ▶ ORF (Open Reading Frame)
- ▶ proteins of known function
- ▶ similarity to proteins of known function (more than one third of self-score)
- ▶ weak similarity to proteins of known function (lower than one third of self-score)
- Intron
- tRNA
- Centromere

Chromosome 1

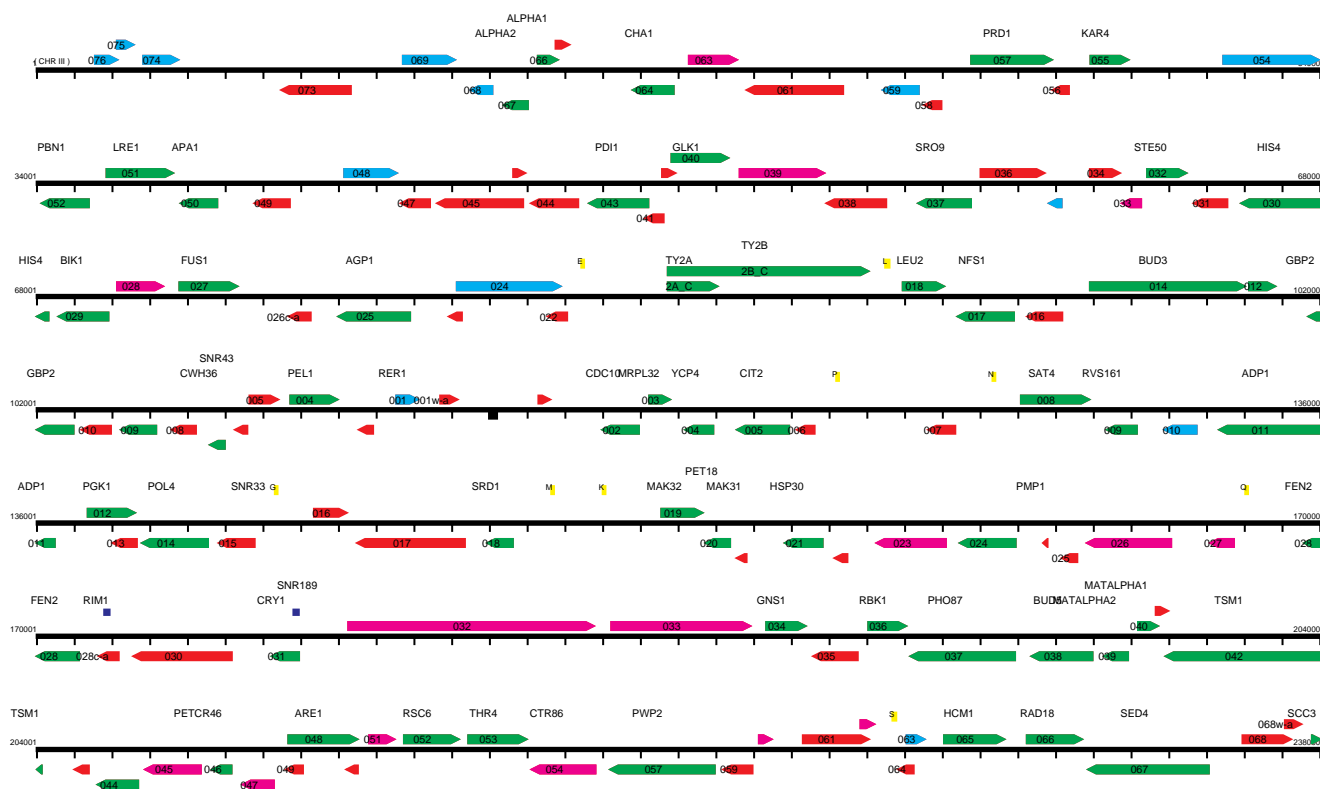


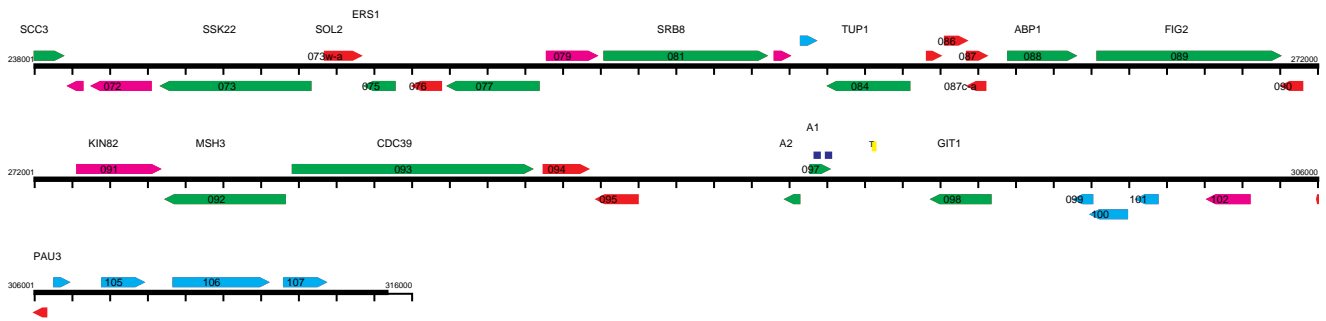
Chromosome 2



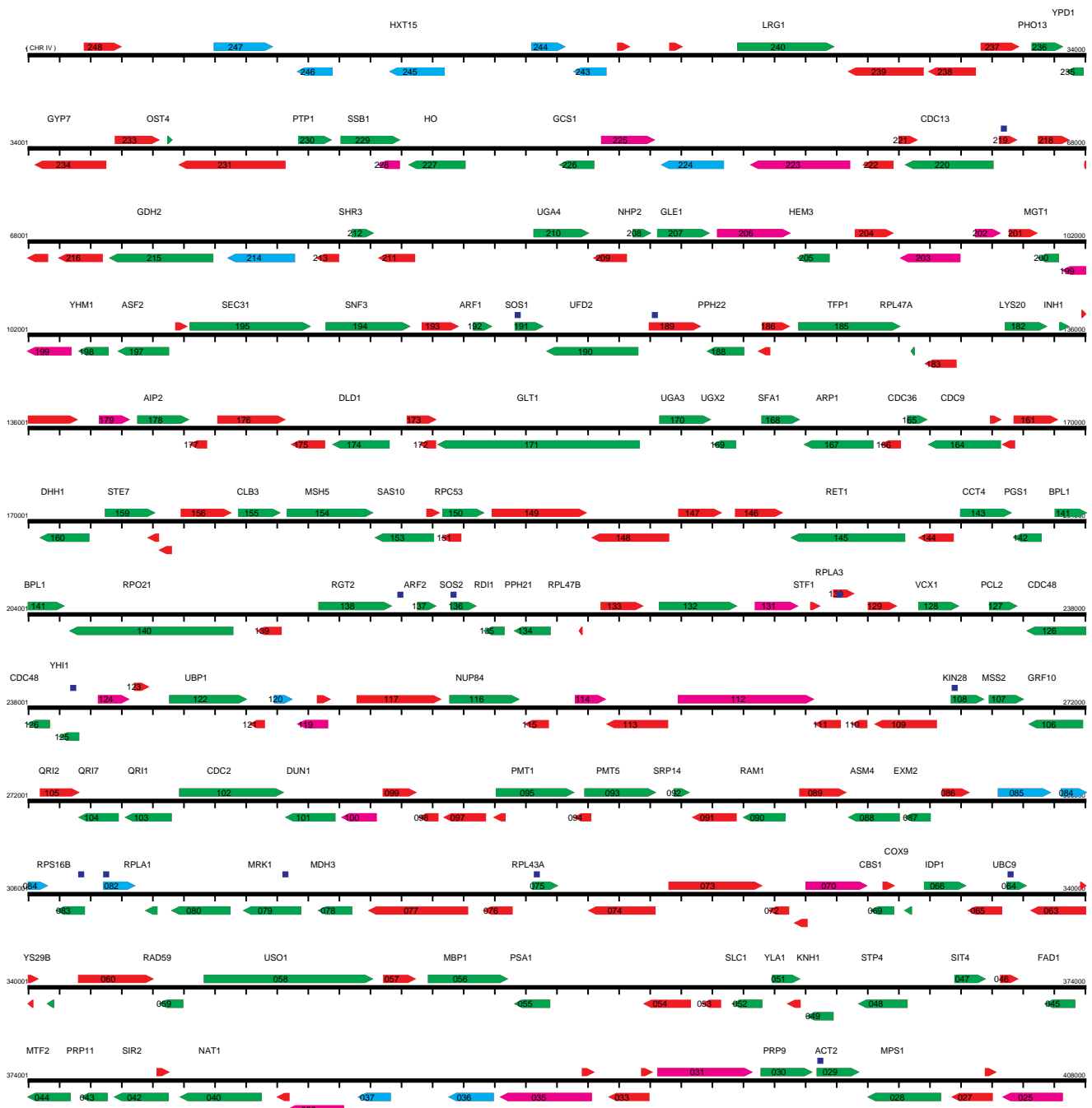


Chromosome 3

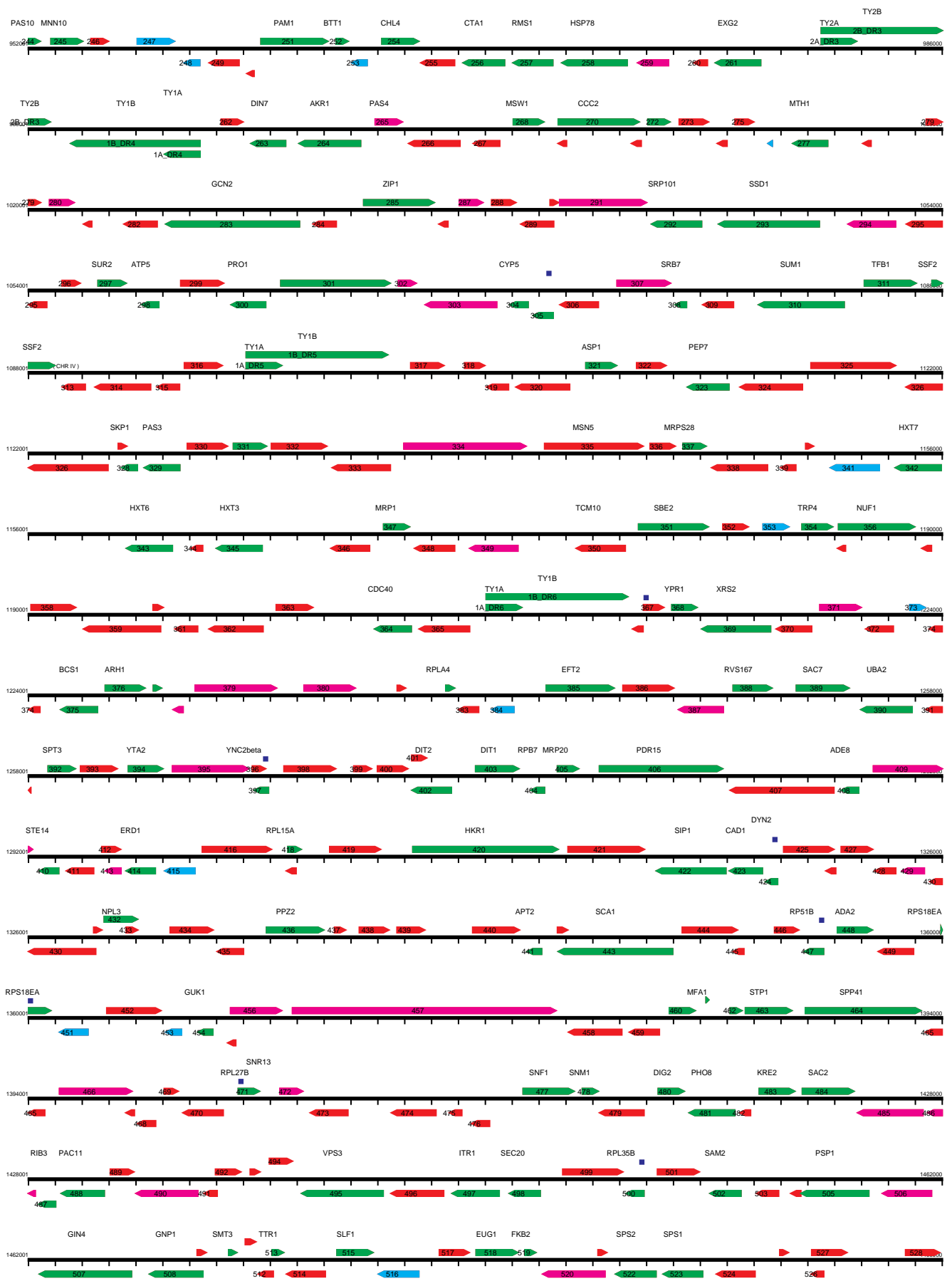


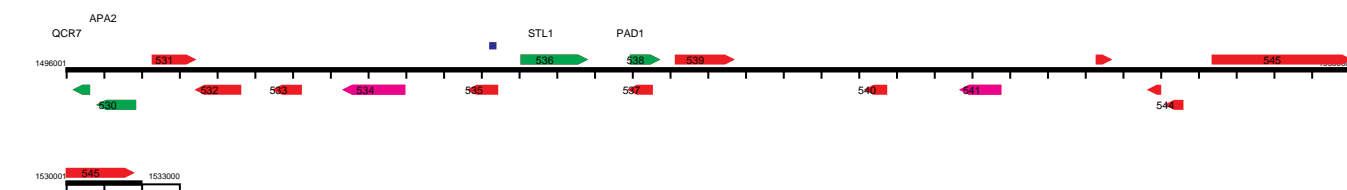


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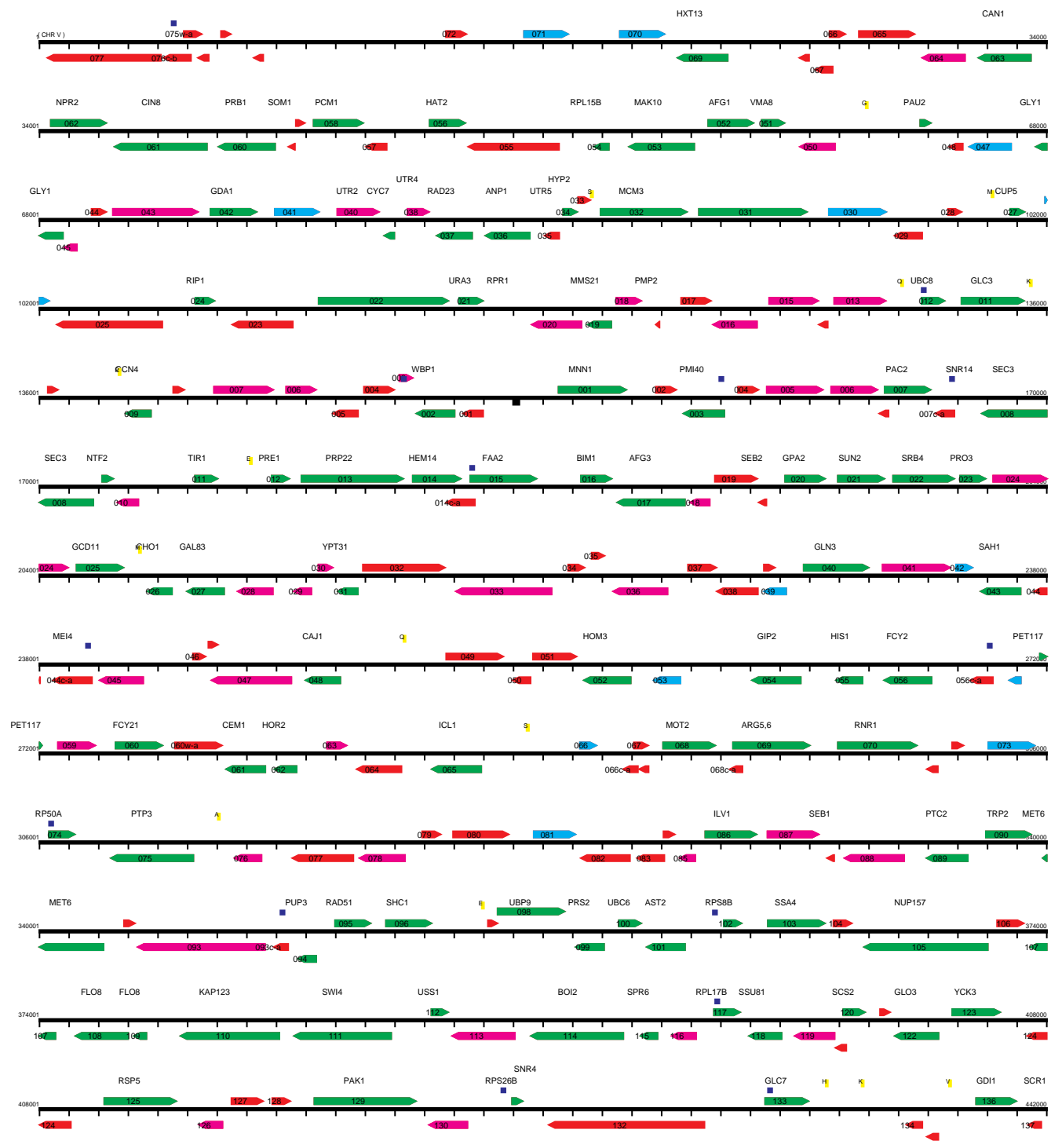


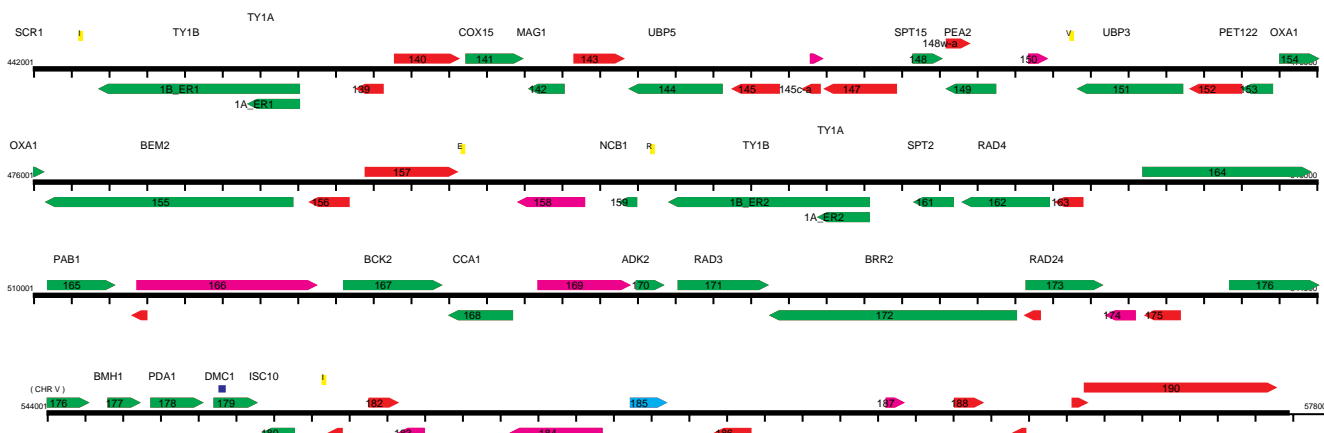




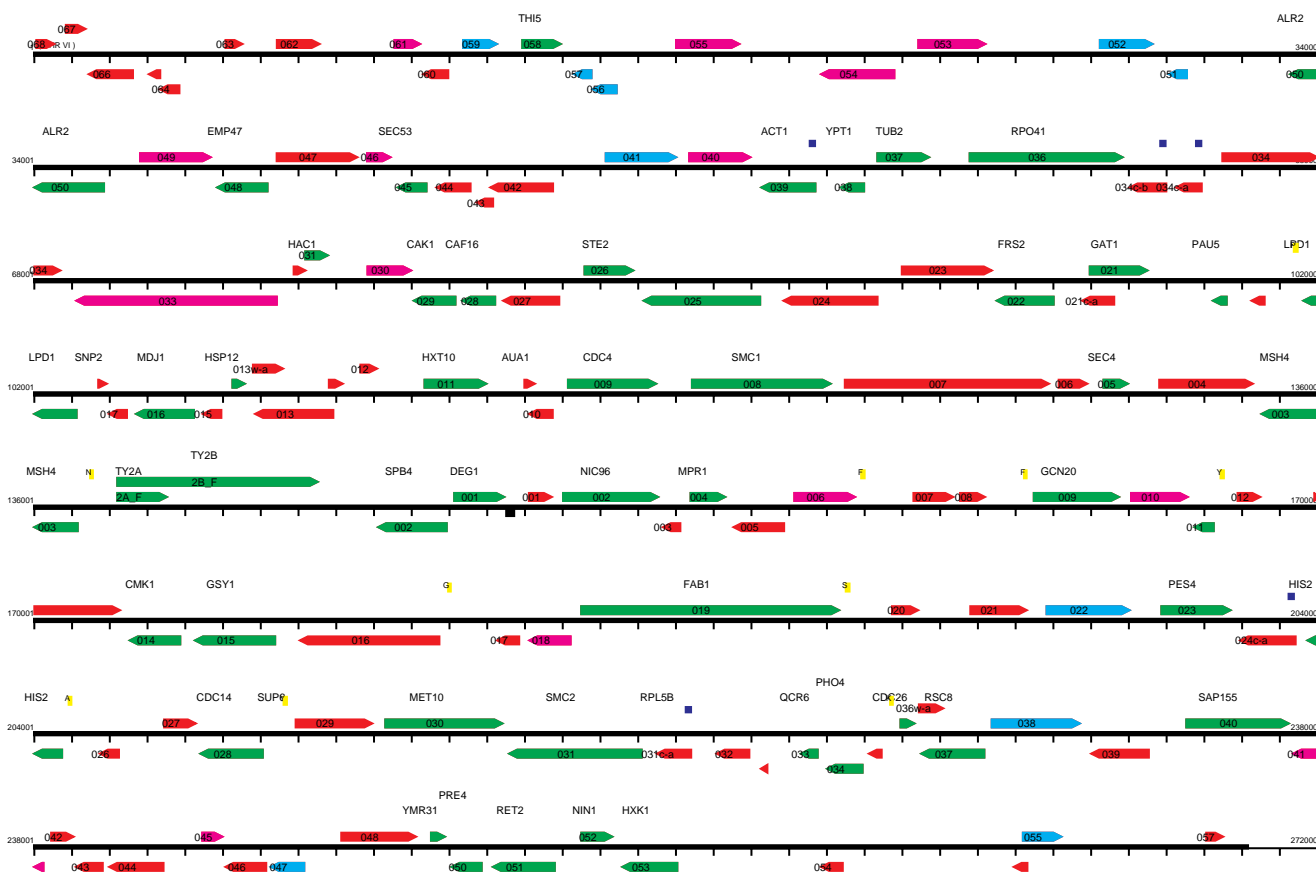


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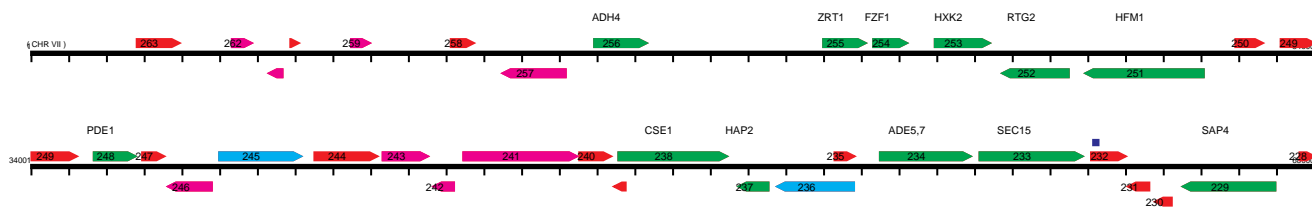


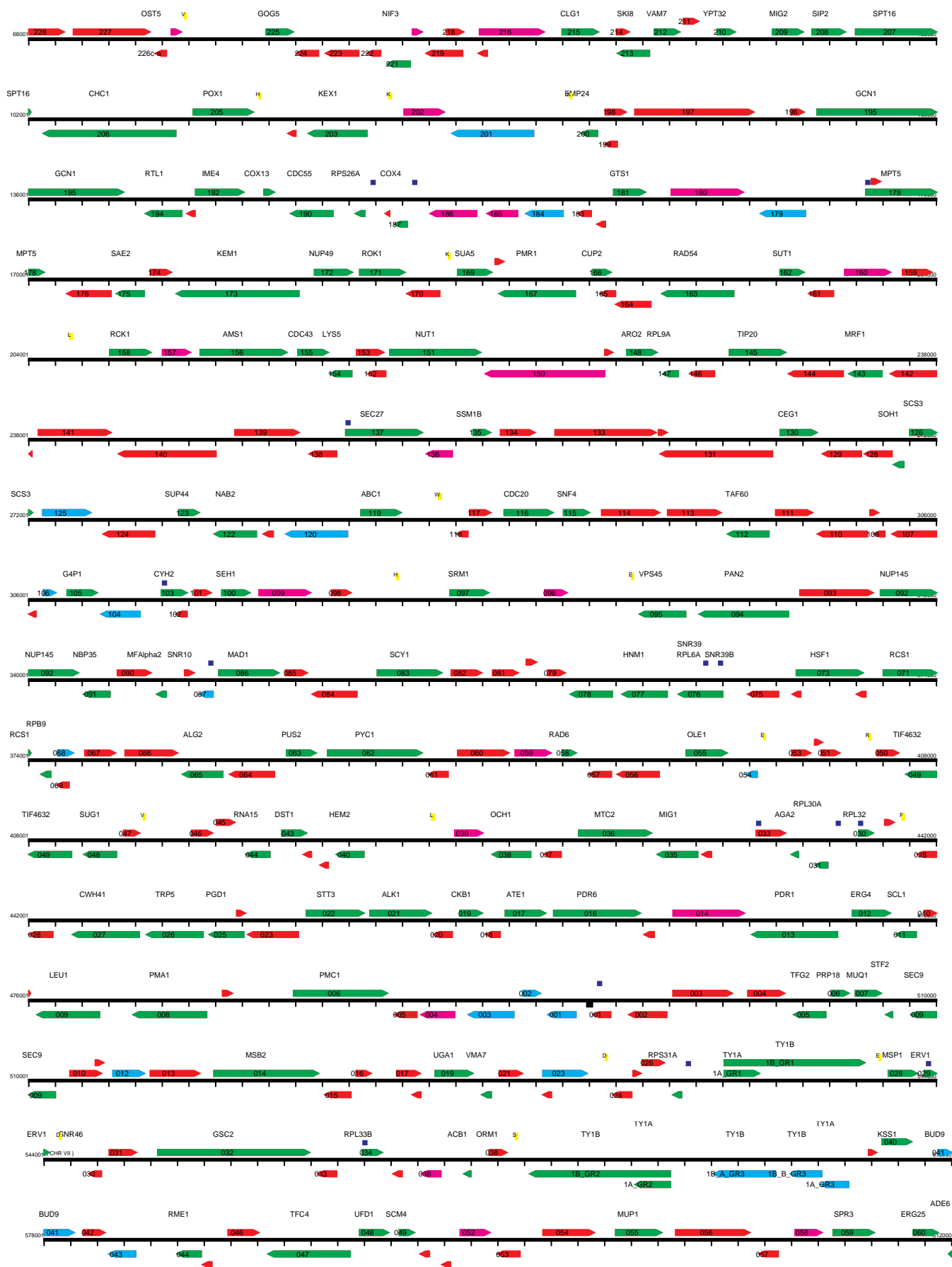


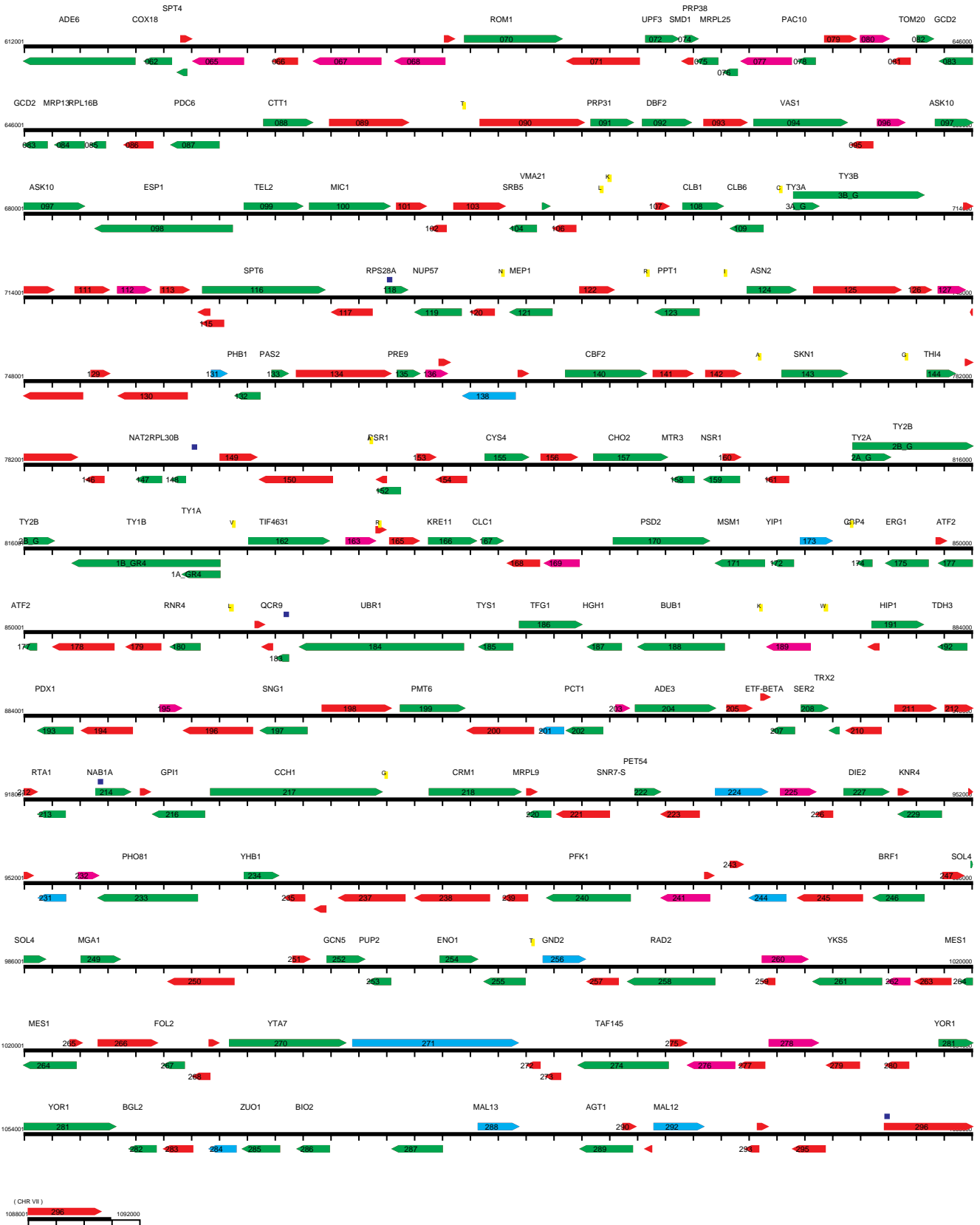
Chromosome 6



Chromosome 7

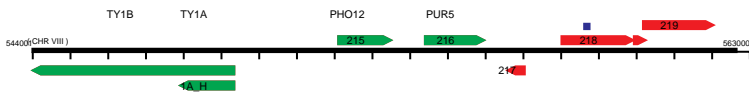




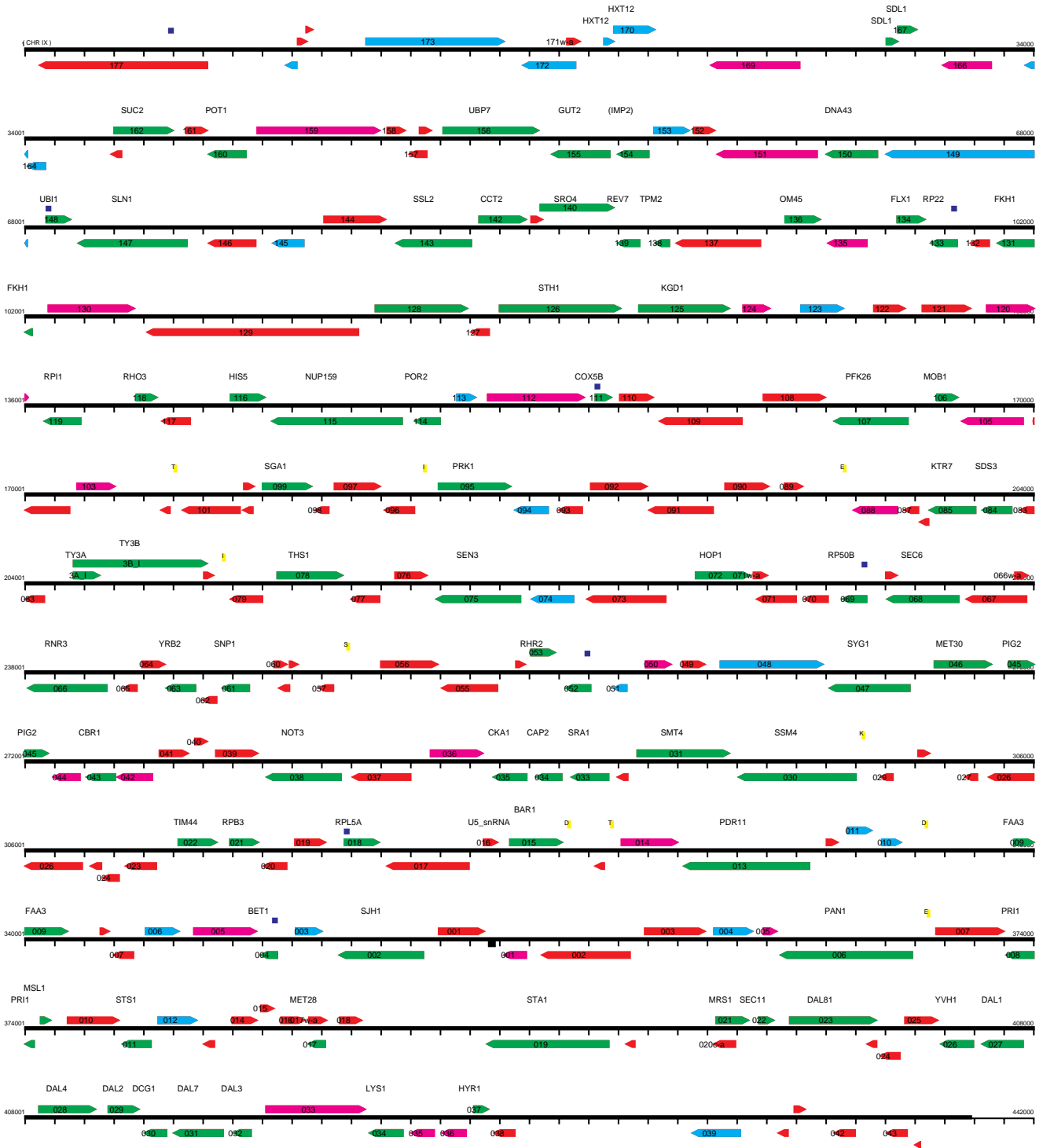


Chromosome 8



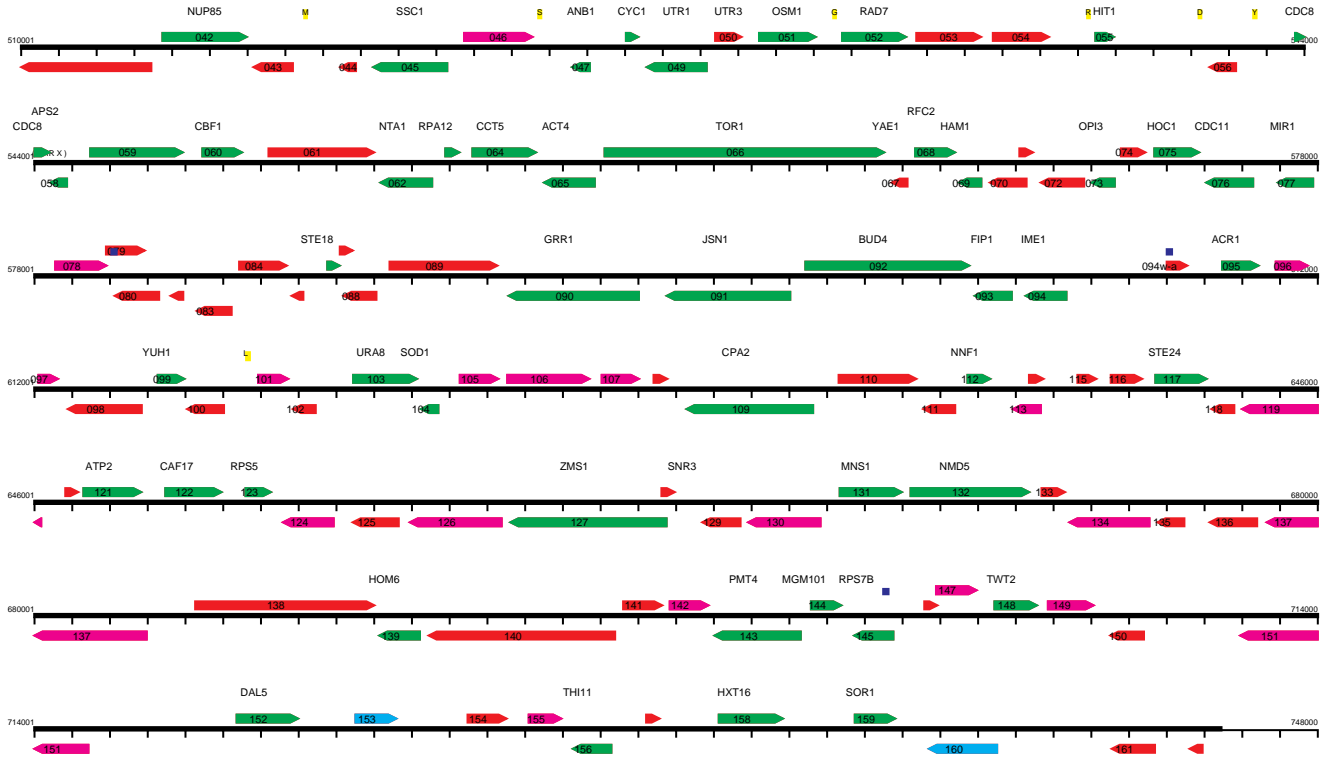


Chromosome 9

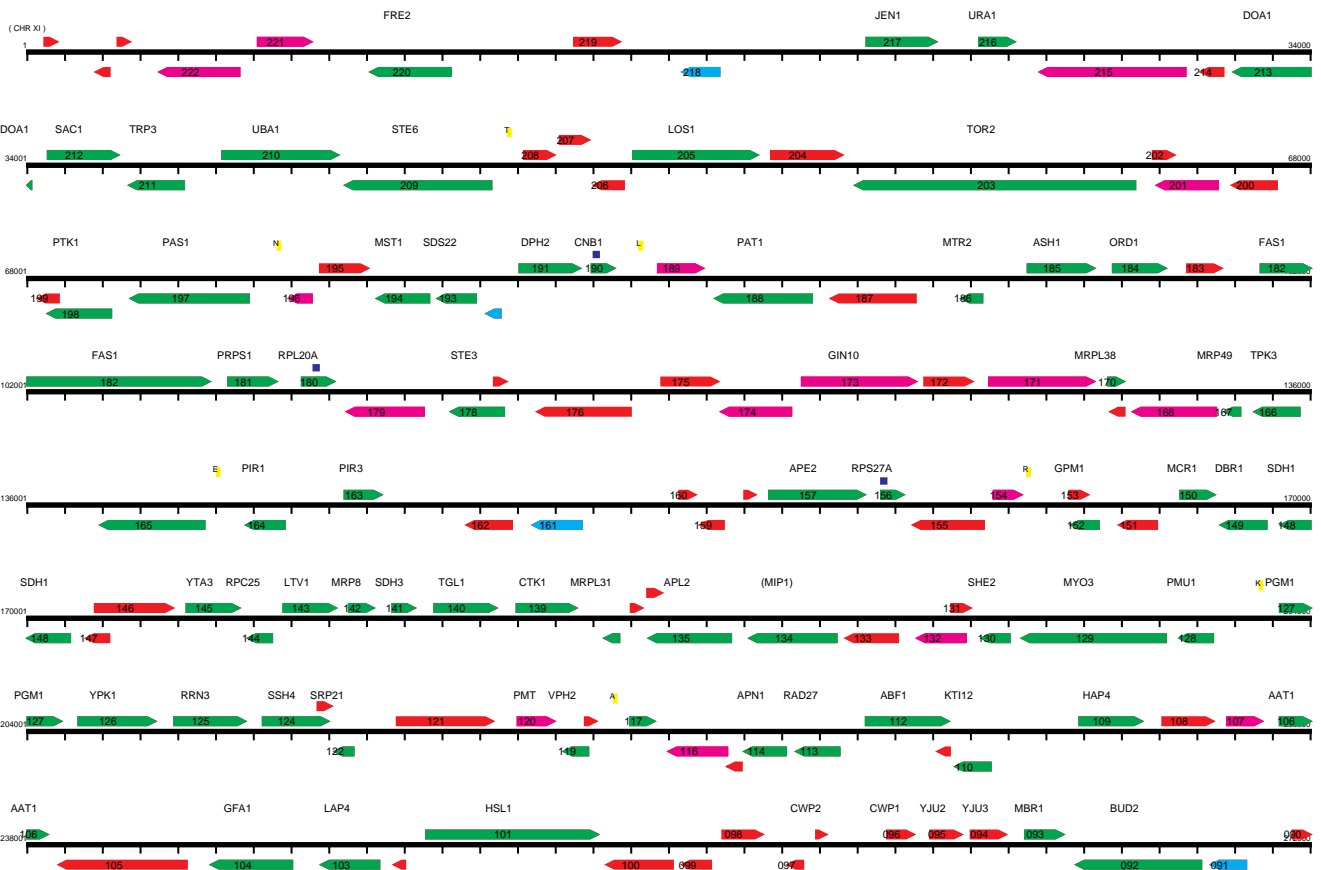


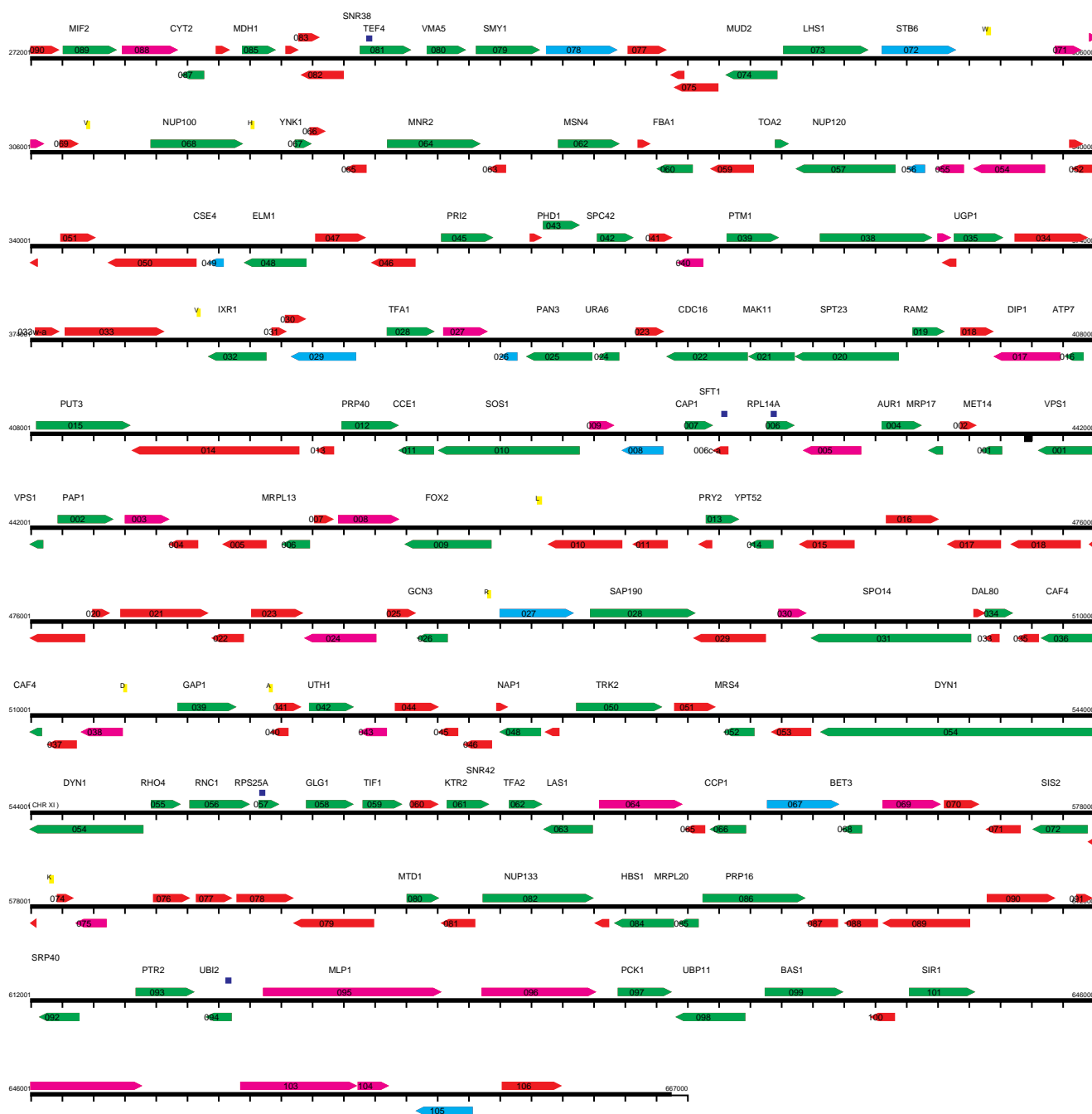
Chromosome 10



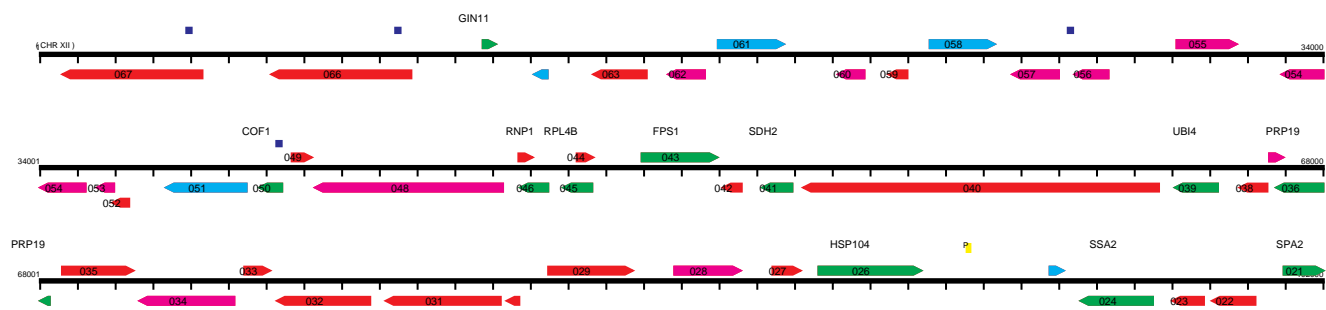


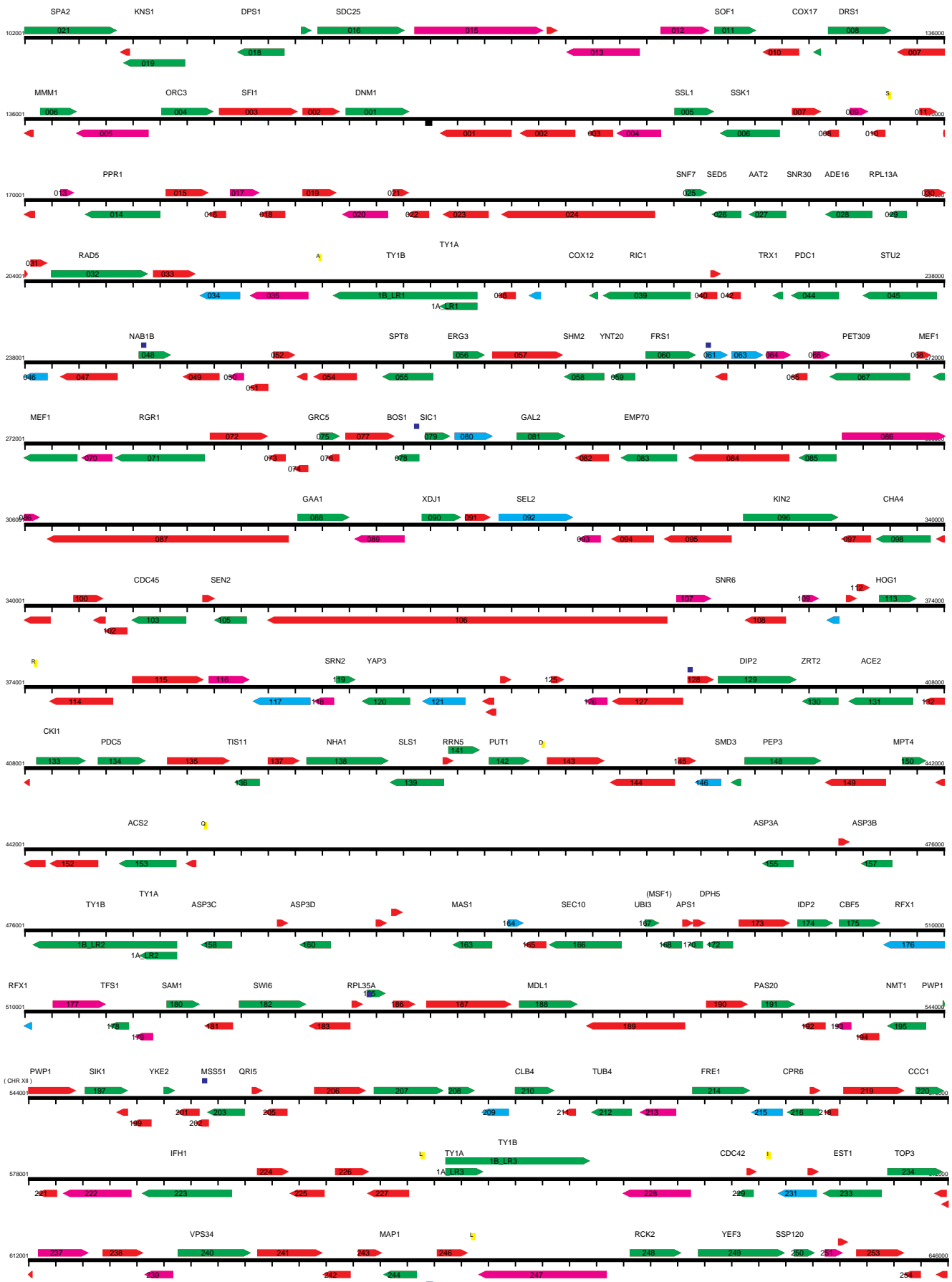
Chromosome 11

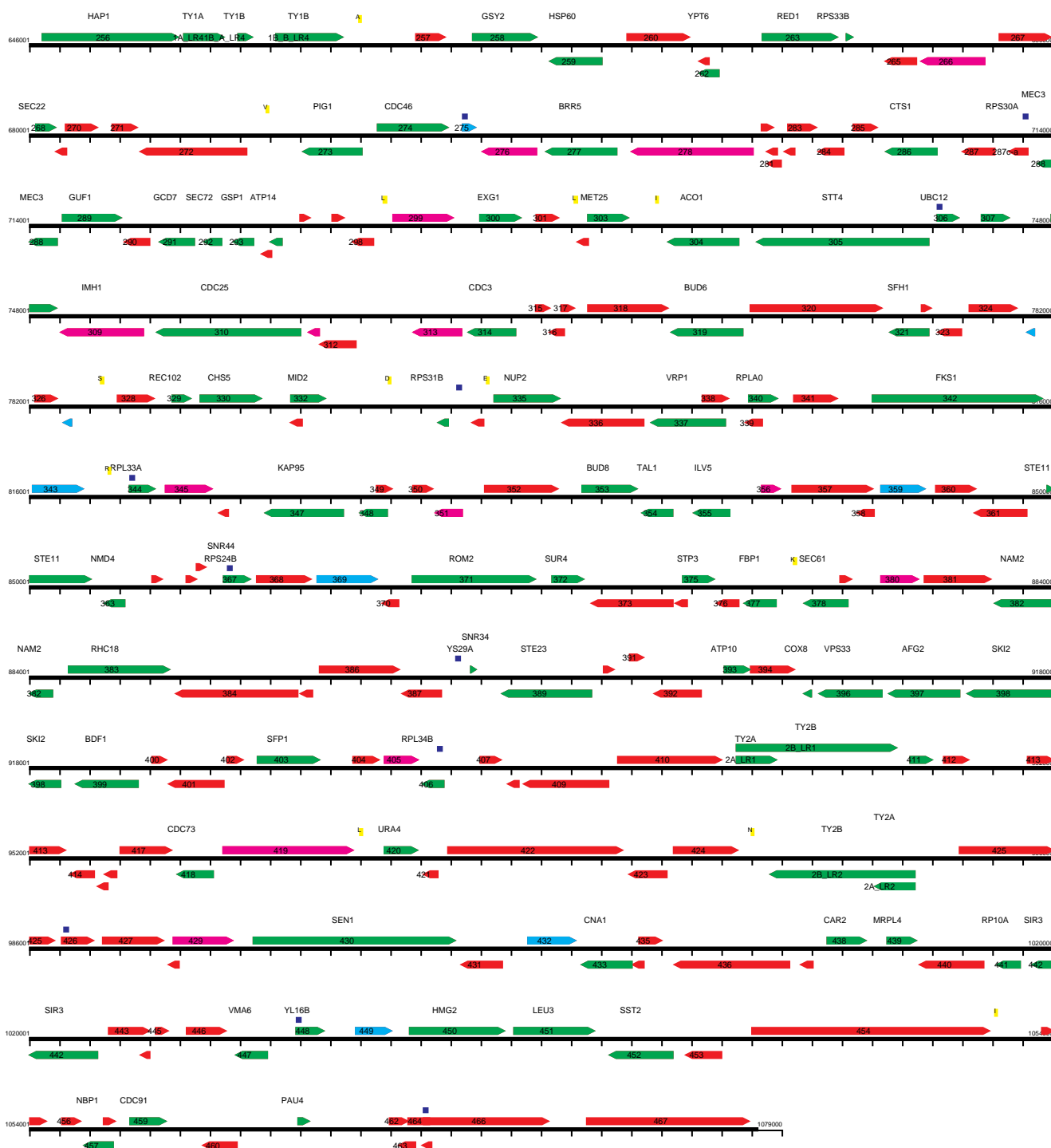




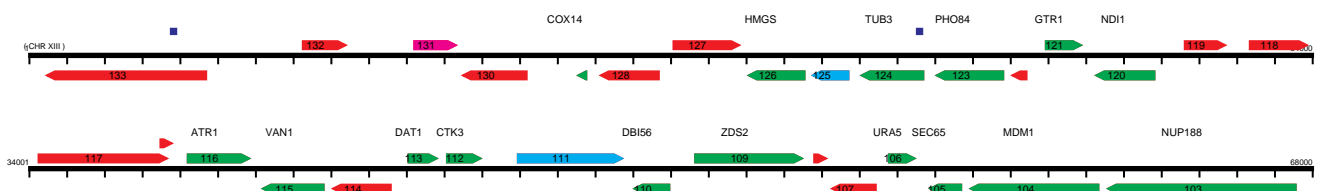
Chromosome 12



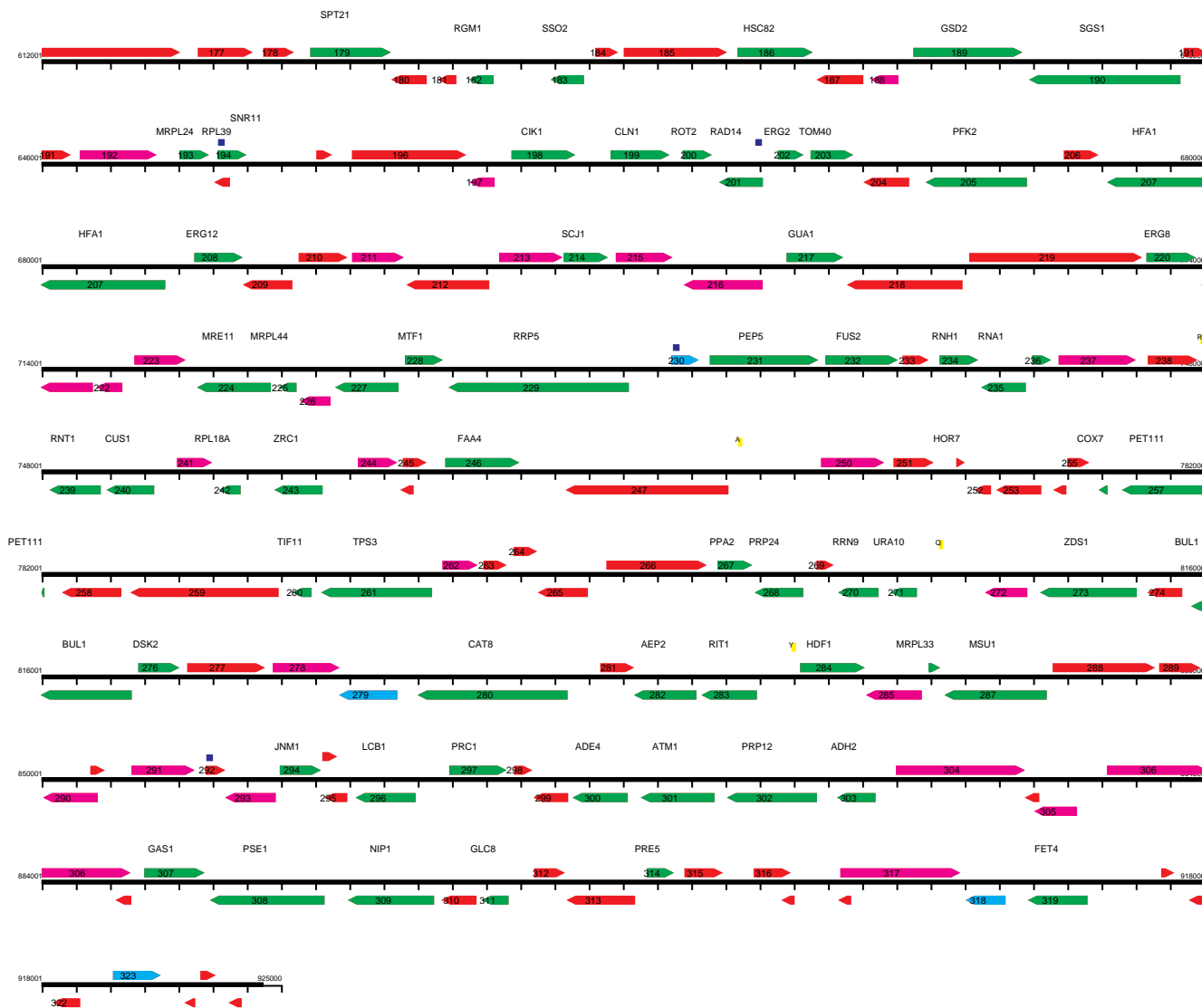




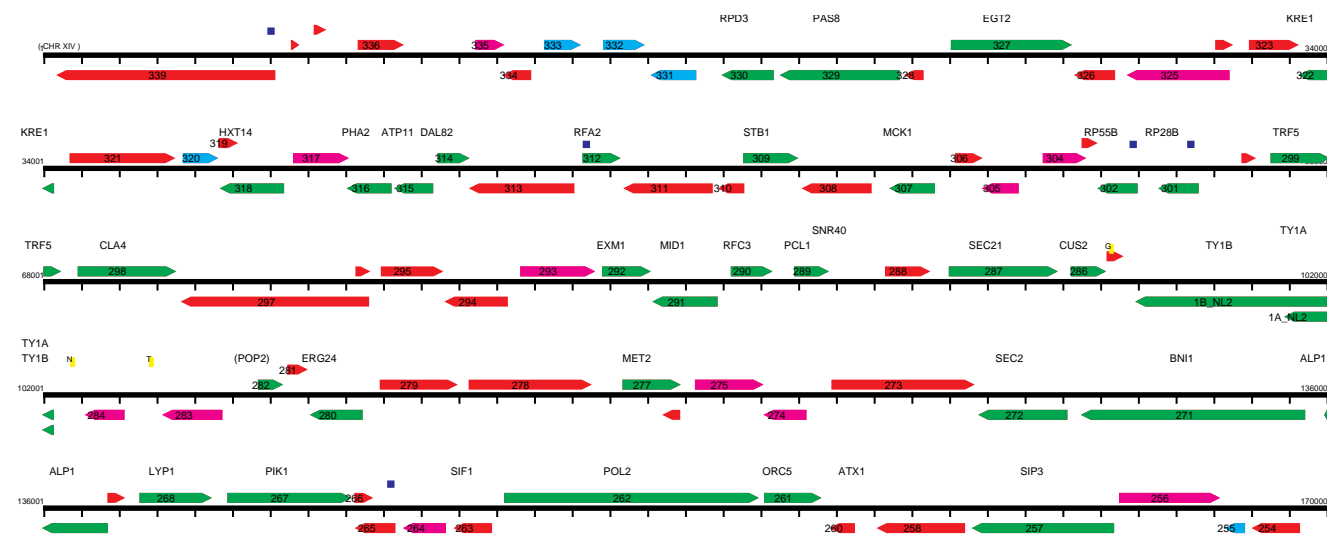
Chromosome 13

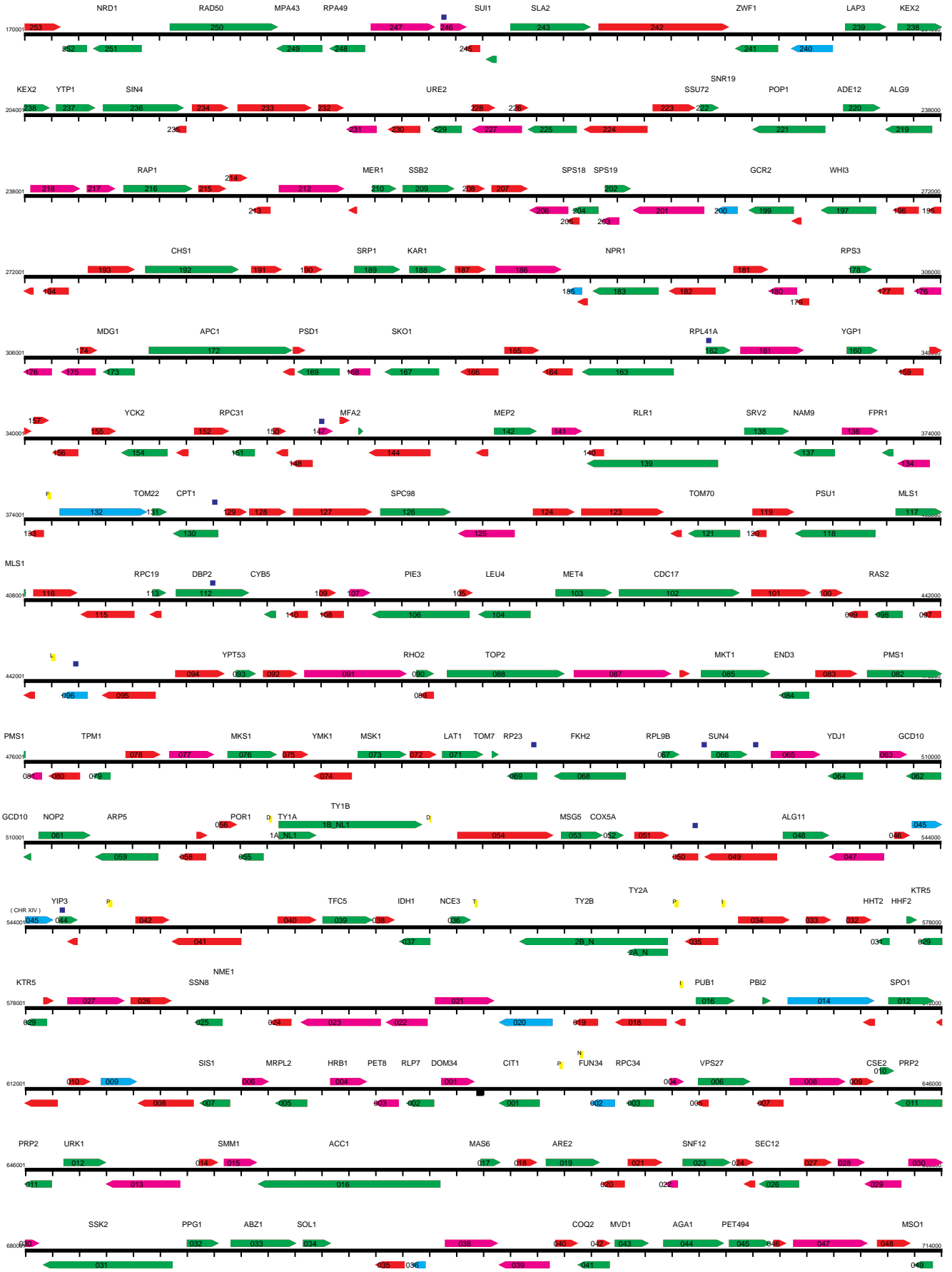


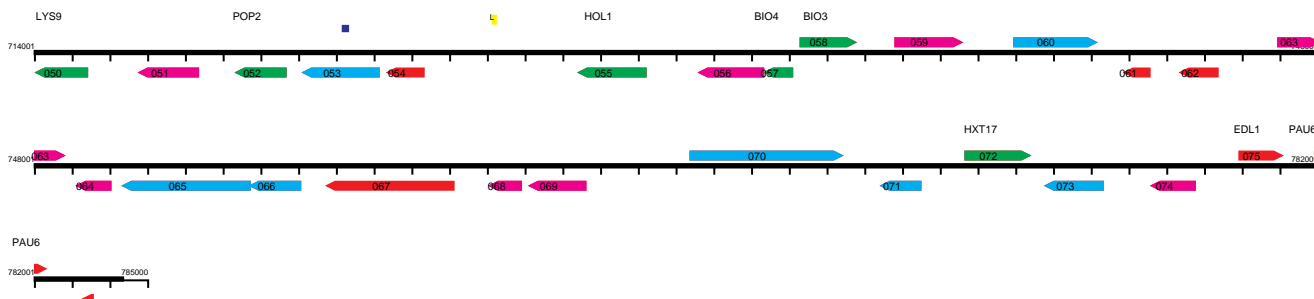




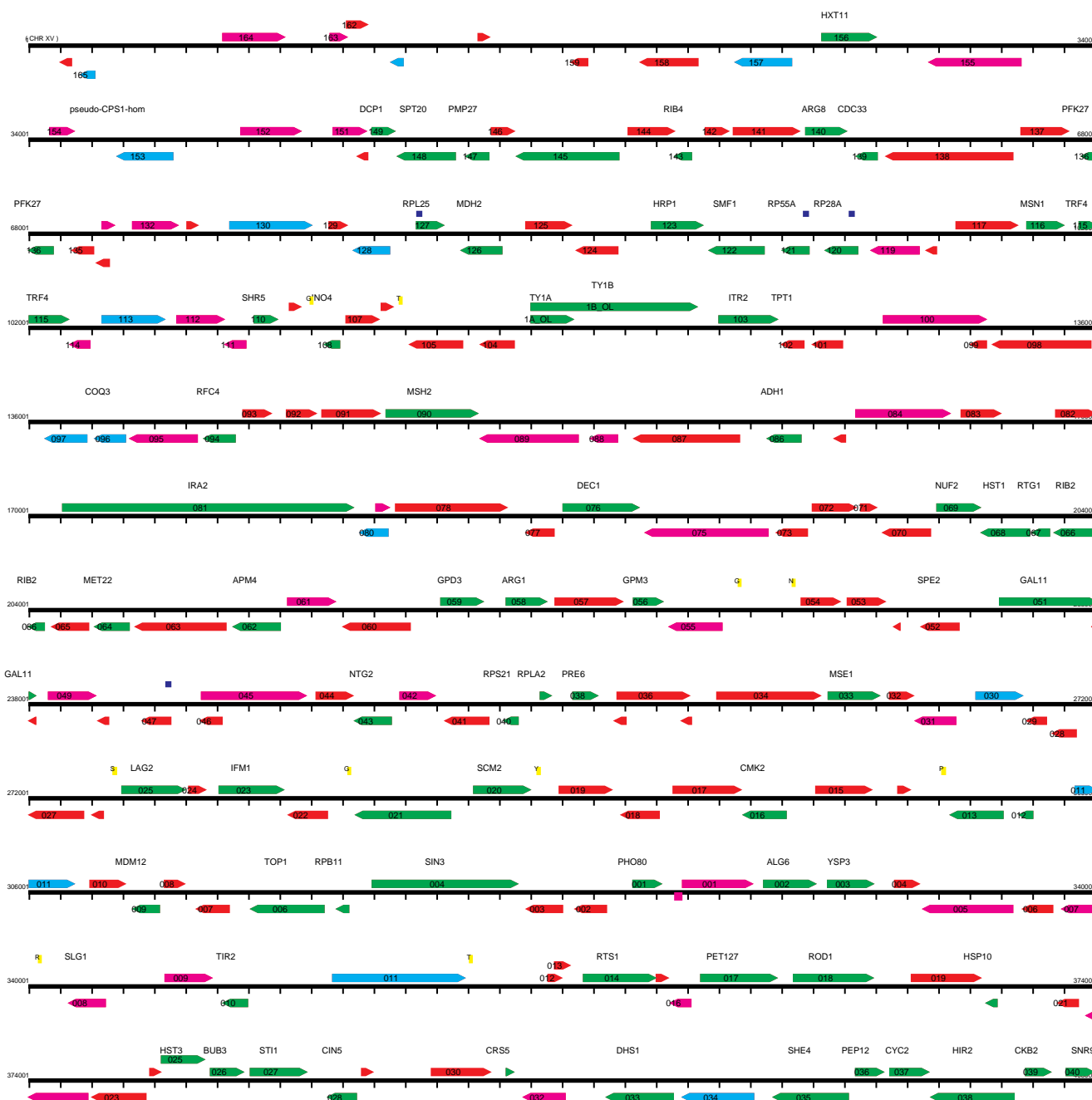
Chromosome14



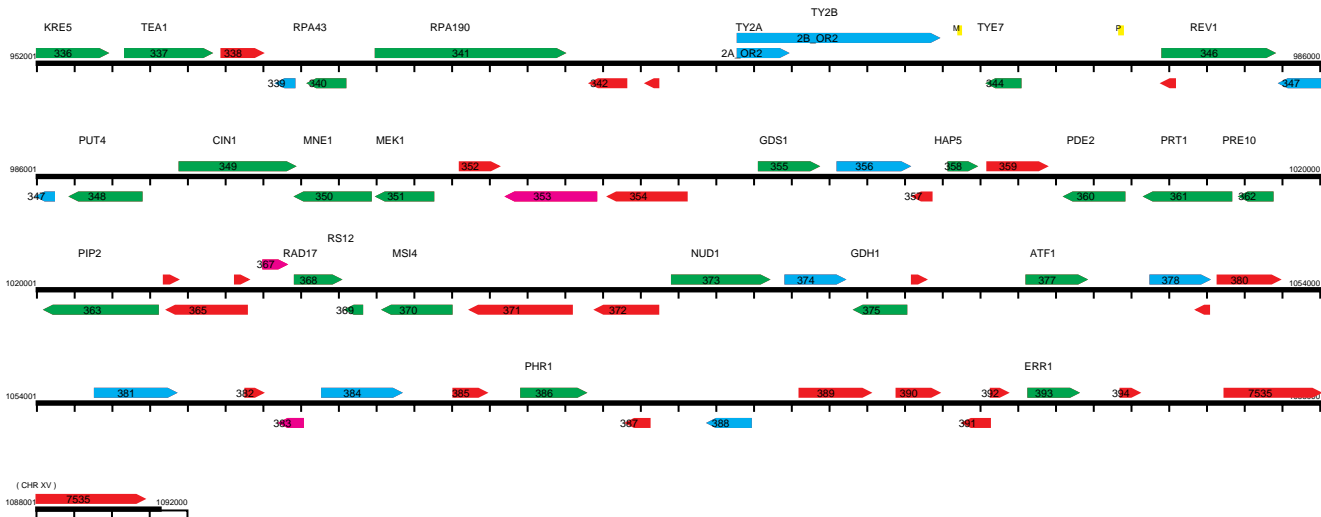




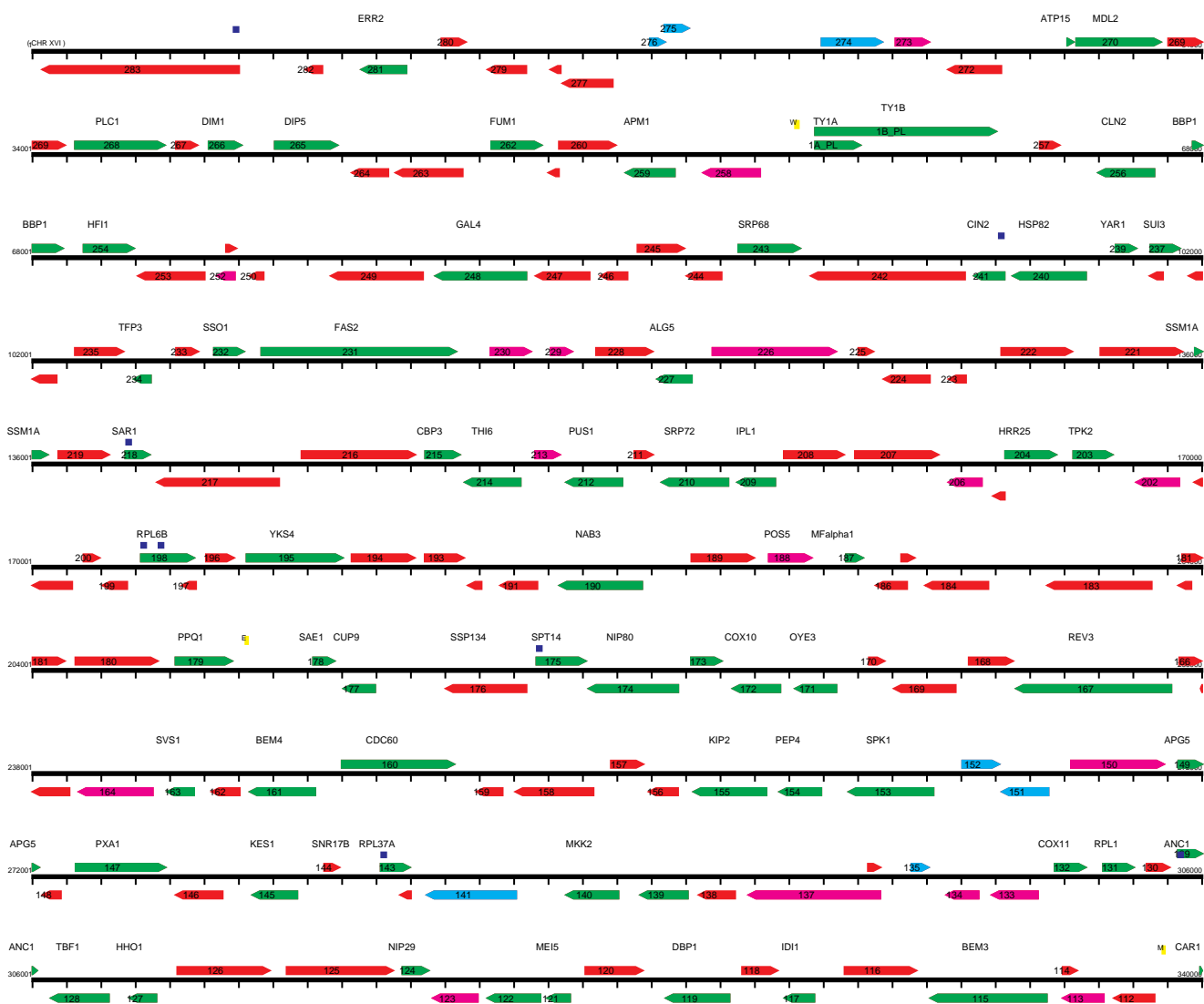
Chromosome 15





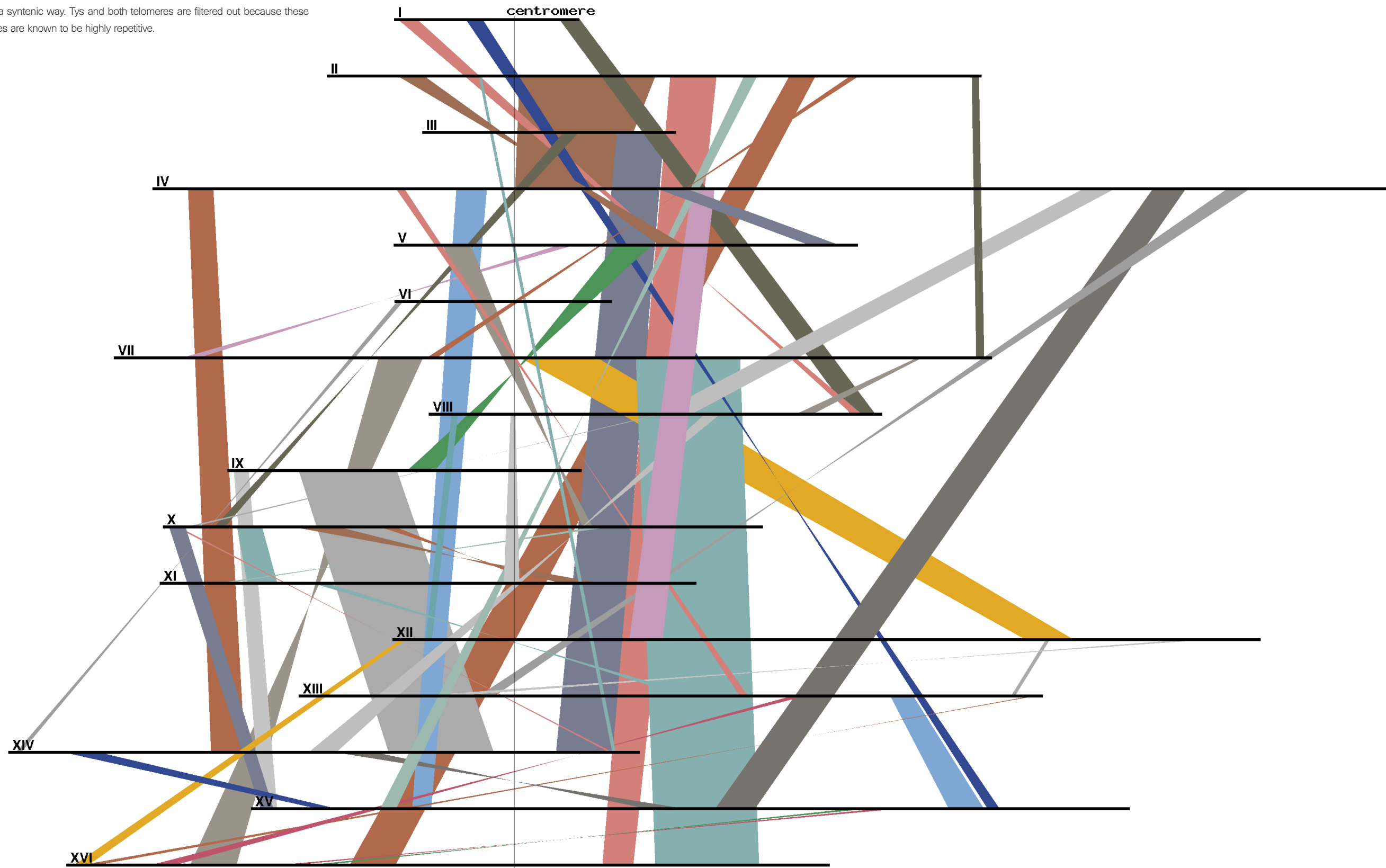


Chromosome 16





View of 53 clustered gene duplications between the 16 chromosomes of yeast. Each chromosome is represented by a bold horizontal line. All chromosomes are aligned according to the centromere, shown as a vertical line. Gene clusters are represented by coloured polygons. A cluster is detected if, within a window of 25 kb, at least 5 blocks of coding DNA (block size 500 nucleotides) are duplicated in a syntenic way. Tys and both telomeres are filtered out because these sequences are known to be highly repetitive.



scale 50000 bases

| | | | |
|---|-----------|---|---|
| Metabolism | 35 | tRNA modification other tRNA-transcription activities mRNA synthesis mRNA processing (splicing) mRNA processing (5'-end, 3'-end processing and mRNA degradation) other mRNA-transcription activities RNA transport other transcription activities | mitochondrial transport vesicular transport (Golgi network, etc.) peroxisomal transport vacuolar transport extracellular transport cellular import other intracellular-transport activities |
| Energy | 40 | Protein synthesis ribosomal proteins translation (initiation, elongation and termination) translational control tRNA synthetases other protein-synthesis activities | Cellular organization and biogenesis 54 organization and biogenesis of cell wall and plasma membrane organization and biogenesis of cytoskeleton organization and biogenesis of endoplasmic reticulum and Golgi organization and biogenesis of chromosome structure mitochondrial organization and biogenesis peroxisomal organization and biogenesis endosomal organization and biogenesis vacuolar and lysosomal organization and biogenesis other cellular organization and biogenesis activities |
| Cell growth, cell division and DNA synthesis | 41 | Protein destination 49 protein folding and stabilization protein targeting, sorting and translocation protein modification (glycosylation, acylation, myristylation, palmitoylation, farnesylation and processing) assembly of protein complexes proteolysis other protein-destination activities | Signal transduction 58 pheromone response generation morphogenesis osmosensing nutritional response other signal-transduction activities |
| Transcription | 45 | Transport facilitation 51 ion channels ion transporters sugar and carbohydrate transporters amino-acid transporters lipid transporters purine and pyrimidine transporters allantoin and allantate transporters transport ATPases ABC transporters drug transporters other transport-facilitators | Cell rescue 59 stress response generation DNA repair (direct repair, base excision repair and nucleotide excision repair) detoxification cell death and ageing degradation of exogenous polynucleotides other cell-rescue activities |
| | | Intracellular transport 53 nuclear transport | Unclassified proteins 60 |

Metabolism

amino-acid metabolism

amino-acid biosynthesis

| [ORF] | [Gene] | [Encoded or related protein] |
|---------|---------------|--|
| YHR037w | <i>PUT2</i> | 1-pyrroline-5-carboxylate dehydrogenase |
| YDR035w | <i>ARO3</i> | 2-dehydro-3-deoxyphosphoheptonate aldolase, phenylalanine-inhibited |
| YBR249c | <i>ARO4</i> | 2-dehydro-3-deoxyphosphoheptonate aldolase, tyrosine-inhibited |
| YNL104c | <i>LEU4</i> | 2-isopropylmalate synthase |
| YGL009c | <i>LEU1</i> | 3-isopropylmalate dehydratase |
| YPR167c | <i>MET16</i> | 3'-phosphoadenylylsulphate reductase |
| YGR019w | <i>UGA1</i> | 4-aminobutyrate aminotransferase (GABA transaminase) |
| YER091c | <i>MET6</i> | 5-methyltetrahydropteroyltryglutamate-homocysteine methyltransferase |
| YMR108w | <i>ILV2</i> | acetolactate synthase |
| YCL009c | <i>ILV6</i> | acetolactate synthase, regulatory subunit |
| YER069w | <i>ARG5,6</i> | acetylglutamate kinase and acetylglutamyl-phosphate reductase |
| YJL071w | <i>ARG2</i> | acetylglutamate synthase |
| YOL140w | <i>ARG8</i> | acetylornithine aminotransferase |
| YDR234w | <i>LYS4</i> | aconitate hydratase |
| YER086w | <i>ILV1</i> | anabolic serine and threonine dehydratase |
| YDR354w | <i>TRP4</i> | anthranilate phosphoribosyltransferase |
| YER090w | <i>TRP2</i> | anthranilate synthase component I |
| YKL211c | <i>TRP3</i> | anthranilate synthase component II |
| YJR109c | <i>CPA2</i> | arginine-specific carbamoylphosphate synthase, large subunit |
| YOR303w | <i>CPA1</i> | arginine-specific carbamoylphosphate synthase, small subunit |
| YOL058w | <i>ARG1</i> | argininosuccinate synthetase |
| YHR018c | <i>ARG4</i> | arginosuccinate lyase |
| YDR127w | <i>ARO1</i> | arom pentafunctional enzyme |
| YPR145w | <i>ASN1</i> | asparagine synthetase |
| YGR124w | <i>ASN2</i> | asparagine synthetase |
| YLR027c | <i>AAT2</i> | aspartate aminotransferase, cytosolic |
| YKL106w | <i>AAT1</i> | aspartate transaminase, mitochondrial |
| YDR158w | <i>HOM2</i> | aspartate-semialdehyde dehydrogenase |
| YFR030w | <i>MET10</i> | assimilatory sulphite reductase flavin-binding subunit |
| YER055c | <i>HIS1</i> | ATP phosphoribosyltransferase |
| YCL018w | <i>LEU2</i> | β-isopropyl-malate dehydrogenase |
| YJR148w | <i>TWT2</i> | branched-chain amino acid aminotransferase, cytosolic |
| YHR208w | <i>TWT1</i> | branched-chain amino acid aminotransferase, mitochondrial |
| YGR204w | <i>ADE3</i> | C1-tetrahydrofolate synthase, cytoplasmic |

| | | |
|---------|--------------|---|
| YBR084w | <i>MIS1</i> | C1-tetrahydrofolate synthase, mitochondrial |
| YPR060c | <i>ARO7</i> | chorismate mutase |
| YGL148w | <i>ARO2</i> | chorismate synthase |
| YGR155w | <i>CYS4</i> | cystathionine β-synthase |
| YAL012w | <i>CYS3</i> | cystathionine γ-lyase |
| YER023w | <i>PRO3</i> | δ 1-pyrroline-5-carboxylate reductase |
| YHR068w | <i>DYS1</i> | deoxyhypusine synthase |
| YOR236w | <i>DFR1</i> | dihydrofolate reductase |
| YFL018c | <i>LPD1</i> | dihydroliipoamide dehydrogenase |
| YJR016c | <i>ILV3</i> | dihydroxy-acid dehydratase |
| YLR172c | <i>DPH5</i> | diphthamide methyltransferase |
| YKL191w | <i>DPH2</i> | diphtheria toxin resistance protein |
| YOR323c | <i>PRO2</i> | γ-glutamyl phosphate reductase |
| YDR300c | <i>PRO1</i> | glutamate 5-kinase |
| YDL171c | <i>GLT1</i> | glutamate synthase (NAPDPH) (GOGAT) |
| YPR035w | <i>GLN1</i> | glutamate-ammonia ligase |
| YBR248c | <i>HIS7</i> | glutamine amidotransferase/cyclase |
| YFR025c | <i>HIS2</i> | histidinol phosphatase |
| YIL116w | <i>HIS5</i> | histidinol-phosphate aminotransferase |
| YMR038c | <i>LYS7</i> | homocitrate dehydrogenase |
| YDL182w | <i>LYS20</i> | homocitrate synthase |
| YJR139c | <i>HOM6</i> | homoserine dehydrogenase |
| YHR025w | <i>THR1</i> | homoserine kinase |
| YNL277w | <i>MET2</i> | homoserine O-acetyltransferase |
| YOR202w | <i>HIS3</i> | imidazoleglycerol-phosphate dehydratase |
| YLR355c | <i>ILV5</i> | ketol-acid reducto-isomerase |
| YBR115c | <i>LYS2</i> | L-aminoacidipate-semialdehyde dehydrogenase, large subunit |
| YGL154c | <i>LYS5</i> | L-aminoacidipate-semialdehyde dehydrogenase, small subunit |
| YER052c | <i>HOM3</i> | L-aspartate 4-P-transferase |
| YLR303w | <i>MET25</i> | O-acetylhomoserine sulphhydrylase |
| YLR438w | <i>CAR2</i> | ornithine aminotransferase |
| YIL088w | <i>ARG3</i> | ornithine carbamoyltransferase |
| YCL030c | <i>HIS4</i> | phosphoribosyl-AMP cyclohydrolase/phosphoribosyl-ATP pyrophosphatase/histidinol dehydrogenase |
| YDR007w | <i>TRP1</i> | phosphoribosylanthranilate isomerase |
| YGR208w | <i>SER2</i> | phosphoserine phosphatase |
| YOR184w | <i>SER1</i> | phosphoserine transaminase |
| YNL316c | <i>PHA2</i> | prephenate dehydratase |
| YBR165c | <i>TRY1</i> | prephenate dehydrogenase (NADP ⁺) |
| YLR142w | <i>PUT1</i> | proline oxidase |
| YOL064c | <i>MET22</i> | protein ser/thr phosphatase |
| YCR054c | <i>CTR86</i> | putative threonine biosynthesis pathway protein |
| YEL046c | <i>GLY1</i> | required for glycine prototrophy in SHMT1 and SHMT2 double mutant |
| YIR034c | <i>LYS1</i> | saccharopine dehydrogenase |
| YNR050c | <i>LYS9</i> | saccharopine dehydrogenase (NADP ⁺ , L-glutamate forming) |

| | | |
|--|--------------|--|
| YCL009c | | similarity to acetolactate synthase III small subunit |
| YLR089c | | similarity to alanine transaminases |
| YGL184c | | similarity to cystathionine β-lyase |
| YHR112c | | similarity to cystathionine γ-synthases |
| YIL074c | | similarity to <i>E. coli</i> phosphoglycerate dehydrogenase |
| YGR012w | | similarity to <i>E. nidulans</i> cysteine synthase |
| YMR250w | | similarity to glutamate decarboxylases |
| YMR062c | | similarity to glutamate N-acetyltransferase |
| YDL131w | | similarity to homocitrate synthases and isopropylmalate synthases |
| YIL094c | | similarity to isopropyl malate and tartrate dehydrogenases |
| YEL038w | <i>UTR4</i> | similarity to <i>K. oxytoca</i> enolase-phosphatase E-1 |
| YML082w | | similarity to <i>N. crassa</i> O-succinylhomoserine (thiol)-lyase |
| YJR130c | | similarity to O-succinylhomoserine (thiol)-lyase |
| YKL215c | | similarity to <i>P. aeruginosa</i> huYA and huYB |
| YOR280c | | similarity to <i>S. pombe</i> dihydrofolate reductase |
| YFL030w | | similarity to several transaminases |
| YOR108w | | strong similarity to 2-isopropylmalate synthase |
| YAL004w | | strong similarity to <i>A. klebsiana</i> glutamate dehydrogenase |
| YDR111c | | strong similarity to alanine transaminase |
| YFR055w | | strong similarity to β-cystathionases |
| YHR033w | | strong similarity to glutamate 5-kinase |
| YHR070w | | strong similarity to <i>N. crassa</i> met-10 ⁺ protein |
| YLL058w | | strong similarity to <i>N. crassa</i> O-succinylhomoserine (thiol)-lyase |
| YER081w | | strong similarity to phosphoglycerate dehydrogenases |
| YJR010w | <i>MET3</i> | sulphate adenylyltransferase |
| YCR053w | <i>THR4</i> | threonine synthase (o-p-homoserine p-lyase) |
| YPR074c | <i>TKL1</i> | transketolase 1 |
| YBR117c | <i>TKL2</i> | transketolase 2 |
| YGL026c | <i>TRP5</i> | tryptophan synthase |
| YML096w | | weak similarity to asparagine synthases |
| regulation of amino-acid metabolism | | |
| YDR173c | <i>ARG82</i> | arginine metabolism transcription factor |
| YKL112w | <i>ABF1</i> | ARS-binding factor |
| YER055c | <i>HIS1</i> | ATP phosphoribosyltransferase |
| YBR253w | <i>SRB6</i> | DNA-directed RNA polymerase II suppressor protein |
| YNL236w | <i>SIN4</i> | global regulator protein |

YFL010w-a *AUA1* involved in ammonia regulation of amino-acid transport
 YIL046w *MET30* involved in regulation of sulphur assimilation genes
 YJR060w *CBF1* kinetochore protein
 YDR159w *SAC3* leucine permease transcriptional regulator
 YDR207c *UME6* negative transcriptional regulator
 YKL015w *PUT3* positive activator of the proline utilisation pathway
 YCR028c *FEN2* similarity to allantoin permease transporter
 YMR116c strong similarity to *N. crassa* CPC2 protein
 YKR099w *BAS1* transcription factor
 YLR098c *CHA4* transcription factor
 YLR451w *LEU3* transcription factor
 YMR042w *ARG80* transcription factor involved in arginine metabolism
 YML099c *ARG81* transcription factor involved in arginine metabolism
 YMR043w *MCM1* transcription factor of the MADS box family
 YIR023w *DAL81* transcriptional activator for allantoin and GABA catabolic genes
 YEL009c *GCN4* transcriptional activator of amino acid biosynthetic genes
 YDR034c *LYS14* transcriptional activator of lysine pathway genes
 YIR017c *MET28* transcriptional activator of sulphur amino acid metabolism
 YNL103w *MET4* transcriptional activator of sulphur metabolism

amino-acid transport

YBR068c *BAP2* amino-acid permease
 YEL063c *CAN1* amino-acid permease
 YBR069c *VAP1* amino-acid permease
 YCL025c *AGP1* asparagine and glutamine permease
 YPL265w *DIP5* dicarboxylic amino-acid permease
 YKR039w *GAP1* general amino-acid permease
 YDR508c *GNP1* high-affinity glutamine permease
 YGR055w *MUP1* high-affinity methionine permease
 YNL270c *ALP1* high-affinity permease for basic amino acids
 YOL020w *SCM2* high-affinity tryptophan transport protein
 YGR191w *HIP1* histidine permease
 YLR375w *STP3* involved in pre-tRNA splicing and in uptake of branched-chain amino acids
 YDL048c *STP4* involved in pre-tRNA splicing and in uptake of branched-chain amino acids
 YNL268w *LYP1* lysine-specific high affinity permease
 YDR130c *ARG11* member of the mitochondrial carrier family (MCF)
 YHL036w *MUP3* methionine permease
 YOR348c *PUT4* proline and γ -aminobutyrate permease
 YFL055w similarity to Gap1p and other amino-acid permeases
 YDR160w similarity to lysine transport protein LYP1
 YLL061w strong similarity to amino-acid transport protein Gap1p
 YPL274w strong similarity to amino-acid transport proteins
 YDR046c (*PAP1*) strong similarity to amino-acid transport proteins
 YBR132c strong similarity to amino-acid permeases

amino-acid degradation

YPL111w *CAR1* arginase
 YDR321w *ASP1* asparaginase
 YOR375c *GDH1* glutamate dehydrogenase (NADP⁺)
 YMR189w *GSD2* glycine decarboxylase subunit
 YDR019c *GCV1* glycine decarboxylase T subunit
 YDR272w *GLO2* glyoxalase II
 YOR040w *GLO4* glyoxalase II
 YLR155c *ASP3A* L-asparaginase II
 YLR157c *ASP3B* L-asparaginase II
 YLR158c *ASP3C* L-asparaginase II
 YLR160c *ASP3D* L-asparaginase II
 YCL064c *CHA1* L-serine/L-threonine deaminase
 YAL062w *GDH3* NADP-glutamate dehydrogenase
 YDL215c *GDH2* NAD-specific glutamate dehydrogenase (NAD)
 YKL184w *ORD1* ornithine decarboxylase
 YLR142w *PUT1* proline oxidase
 YLR180w *SAM1* S-adenosylmethionine synthetase 1
 YDR502c *SAM2* S-adenosylmethionine synthetase 2
 YIL167w *SDL1* serine dehydratase
 YDR294c similarity to glutamate decarboxylases
 YJR078w similarity to indoleamine 2,3-dioxygenase
 YIL042c similarity to rat branched-chain α -ketoacid dehydrogenase kinase
 YGL202w similarity to rat kynurenine/ α -aminoacidate aminotransferase
 YHR137w similarity to rat kynurenine/ α -aminoacidate aminotransferase
 YFL030w similarity to several transaminases
 YDR111c strong similarity to alanine transaminase
 YBR006w strong similarity to *E. coli* succinate semialdehyde dehydrogenase
 YAL044c *GCV3* strong similarity to human glycine cleavage system protein H
 YIL168w *SDL1* strong similarity to L-serine dehydratase Cha1p
 YLR231c strong similarity to rat kynureninase
 YKL218c strong similarity to threonine dehydratase
 YBR208c *DUR1,2* urea amidolyase

other amino-acid metabolism activities

YKL157w *APE2* aminopeptidase yscII
 YGL017w *ATE1* arginyl tRNA transferase
 YPR068c *SPE3* putrescine aminopropyltransferase (spermidine synthase)
 YFR018c similarity to human glutamyl-peptide cyclotransferase

nitrogen and sulphur metabolism

nitrogen and sulphur utilization

YPR167c *MET16* 3'-phosphoadenylylsulphate reductase
 YIR027c *DAL1* allantoinase
 YIR029w *DAL2* allantoinase
 YDR242w *AMD2* amidase
 YOL058w *ARG1* argininosuccinate synthetase
 YLR027c *AAT2* aspartate aminotransferase, cytosolic
 YFR030w *MET10* assimilatory sulphite reductase flavin-binding subunit
 YKL001c *MET14* ATP adenosine-5'-phosphosulphate 3'-phosphotransferase
 YHR176w *FMO* flavin-containing monooxygenase
 YOR375c *GDH1* glutamate dehydrogenase (NADP⁺)
 YDL171c *GLT1* glutamate synthase (NAPDPH) (GOGAT)
 YPR035w *GLN1* glutamate-ammonia ligase
 YIL172w *CPS1* Gly-X carboxypeptidase YSCS
 YAL062w *GDH3* NADP-glutamate dehydrogenase
 YDL215c *GDH2* NAD-specific glutamate dehydrogenase (NAD)
 YLR438w *CAR2* ornithine aminotransferase
 YJR149w similarity to 2-nitropropane dioxygenase
 YLR089c similarity to alanine transaminases
 YMR293c similarity to amidases
 YFL061w similarity to *M. verrucaria* cyanamide hydratase
 YDR111c similarity to several transaminases
 YER057c similarity to thiol sulphurtransferases
 YIL051c strong similarity to alanine transaminase
 YLR089c strong similarity to *Azotobacter* nitrogen fixation vnfA protein
 YJL060w strong similarity to *Azotobacter* nitrogen fixation vnfA protein
 YPL135w strong similarity to kynurenine aminotransferase
 YOR226c strong similarity to nitrogen fixation protein (nifU)
 YJR010w *MET3* sulphate adenylyltransferase
 YJR137c sulphite reductase
 YBR208c *DUR1,2* urea amidolyase
 YIR032c *DAL3* ureidoglycolate hydrolase
 YJL035c weak similarity to *B. japonicum* nitrogen fixation protein
 YKL040c weak similarity to nitrogen fixation protein nifU

regulation of nitrogen and sulphur utilization

YGR019w *UGA1* 4-aminobutyrate aminotransferase (GABA transaminase)
 YPL111w *CAR1* arginase
 YKL112w *ABF1* ARS-binding factor
 YNL216w *RAP1* DNA-binding protein with repressor and activator activity
 YFL010w-a *AUA1* involved in ammonia regulation of amino acid transport
 YIR030c *DCG1* involved in nitrogen-catabolite metabolism
 YBR213w *MET8* involved in the expression of PAPS reductase and sulphite reductase
 YDR207c *UME6* negative transcriptional regulator
 YNL229c *URE2* nitrogen catabolite repression regulator
 YEL062w *NPR2* nitrogen permease regulator
 YNL183c *NPR1* ser/thr protein kinase
 YCR028c *FEN2* similarity to allantoin permease transporter
 YLR013w similarity to nitrogen regulatory proteins
 YGL254w *FZF1* sulphite resistance protein
 YFL021w *GAT1* transcription factor for nitrogen regulation
 YER040w *GLN3* transcription factor for positive nitrogen regulation
 YMR042w *ARG80* transcription factor involved in arginine metabolism
 YML099c *ARG81* transcription factor involved in arginine metabolism
 YMR043w *MCM1* transcription factor of the MADS box family
 YIR023w *DAL81* transcriptional activator for allantoin and GABA catabolic genes
 YDL170w *UGA3* transcriptional activator for GABA catabolic genes
 YNL103w *MET4* transcriptional activator of sulphur metabolism
 YKR034w *DAL80* transcriptional repressor for allantoin and GABA catabolic genes

nitrogen and sulphur transport

YGR121c *MEP1* ammonia permease of high capacity and moderate affinity
 YNL142w *MEP2* high-affinity low-capacity ammonia permease
 YBR294w *SUL1* high-affinity sulphate transport protein
 YPR003c similarity to sulphate transporter proteins
 YPR138c *MEP3* strong similarity to ammonium transport proteins
 YLR092w *SEL2* strong similarity to Sul1p
 YHL061c *DUR3* urea transport protein

nucleotide metabolism

purine-ribonucleotide metabolism

YLR028c *ADE16* 5-aminoimidazole-4-carboxamide ribotide transformylase
 YGR061c *ADE6* 5'-phosphoribosylformyl glycinamide synthetase
 YML022w *APT1* adenine phosphoribosyltransferase
 YDR441c *APT2* adenine phosphoribosyltransferase
 YNL220w *ADE12* adenylosuccinate synthetase
 YIR027c *DAL1* allantoinase
 YIR029w *DAL2* allantoinase
 YMR300c *ADE4* amidophosphoribosyltransferase
 YML035c *AMD1* AMP deaminase
 YGR204w *ADE3* C1-tetrahydrofolate synthase, cytoplasmic
 YBR084w *MIS1* C1-tetrahydrofolate synthase, mitochondrial
 YOR236w *DFR1* dihydrofolate reductase
 YMR217w *GUA1* GMP synthase (glutamine-hydrolyzing)
 YHR216w *PUR5* IMP dehydrogenase
 YIR031c *DAL7* malate synthase 2
 YKR080w *MTD1* methylentetrahydrofolate dehydrogenase (NAD⁺)
 YAR015w *ADE1* phosphoribosylamidoimidazole-succinocarboxamide synthase
 YGL234w *ADE5,7* phosphoribosylamine-glycine ligase and phosphoribosylformylglycinamide cyclo-ligase
 YOR128c *ADE2* phosphoribosylamidoimidazole carboxylase
 YDR408c *ADE8* phosphoribosylglycinamide formyltransferase (GART)
 YKL181w *PRPS1* ribose-phosphate pyrophosphokinase
 YER099c *PRS2* ribose-phosphate pyrophosphokinase
 YHL011c *PRS3* ribose-phosphate pyrophosphokinase
 YBL068w *PRS4* ribose-phosphate pyrophosphokinase
 YLR058c *SHM2* serine hydroxymethyltransferase, cytoplasmic
 YBR263w *SHM1* serine hydroxymethyltransferase, mitochondrial
 YNL141w similarity to adenosine deaminase
 YBR284w similarity to AMP deaminase
 YJL070c similarity to AMP deaminases
 YLR017w similarity to human 5'-methylthioadenosine phosphorylase
 YOL061w similarity to ribose-phosphate pyrophosphokinases
 YOR280c similarity to *S. pombe* dihydrofolate reductase
 YDR020c similarity to uridine kinases and phosphoribulokinases
 YLR359w strong similarity to adenylosuccinate lyase
 YMR120c strong similarity to chicken purH bifunctional enzyme
 YJR105w strong similarity to human adenosine kinase
 YAR075w strong similarity to IMP dehydrogenases
 YML056c strong similarity to IMP dehydrogenases
 YAR073w *FUN63* strong similarity to IMP dehydrogenases
 YLR432w strong similarity to IMP dehydrogenases, Pur5p and YML056c
 YLR209c strong similarity to purine-nucleoside phosphorylases
 YBR208c *DUR1,2* urea amidolyase
 YIR032c *DAL3* ureidoglycolate hydrolase

pyrimidine-ribonucleotide metabolism

YBL039c *URA7* CTP synthase 1
 YJR103w *URA8* CTP synthase 2
 YHR144c *DCD1* deoxycytidylate deaminase
 YLR420w *URA4* dihydroorotase
 YKL216w *URA1* dihydroorotate dehydrogenase
 YJR057w *CDC8* dTMP kinase
 YBR252w *DUT1* dUTP pyrophosphatase, mitochondrial
 YJL130c *URA2* multifunctional pyrimidine biosynthesis protein
 YML106w *URA5* orotate phosphoribosyltransferase 1
 YMR271c *URA10* orotate phosphoribosyltransferase 2
 YEL021w *URA3* orotidine-5'-phosphate decarboxylase
 YFL058w *THI5* pyrimidine biosynthesis protein
 YKL181w *PRPS1* ribose-phosphate pyrophosphokinase
 YER099c *PRS2* ribose-phosphate pyrophosphokinase
 YHL011c *PRS3* ribose-phosphate pyrophosphokinase
 YBL068w *PRS4* ribose-phosphate pyrophosphokinase
 YOL061w similarity to ribose-phosphate pyrophosphokinases
 YOR280c similarity to *S. pombe* dihydrofolate reductase
 YLR245c strong similarity to *B. subtilis* cytidine deaminase
 YNL332w strong similarity to Thi5p, YJR156c, YDL244w and *A. parasiticus*, *S. pombe* Nmt1p
 YDL244w strong similarity to Thi5p, YJR156c, YNL332w and *A. parasiticus*, *S. pombe* nmt1 protein
 YJR156c *THI11* thiamine regulated gene, homologous to nmt1a in *S. pombe*
 YHR128w *FUR1* uracil phosphoribosyltransferase
 YNR012w *URK1* uridine kinase
 YKL024c *URA6* uridine-monophosphate kinase

deoxyribonucleotide metabolism

YDR513w *TRR1* glutaredoxin
 YER070w *RNR1* ribonucleoside-diphosphate reductase, large subunit

YJL026w *RNR2* ribonucleoside-diphosphate reductase, small subunit
 YGR180c *RNR4* ribonucleotide reductase small subunit
 YJL066c *RNR3* ribonucleotide reductase, repair inducible large subunit
 YBR014c similarity to glutaredoxin
 YPL059w similarity to glutaredoxins
 YOR269w *PAC1* similarity to human LJS-1 protein
 YDL010w similarity to hypothetical protein YBR014c and glutaredoxins
 YCL035c strong similarity to glutaredoxin
 YDR353w strong similarity to thioredoxin reductase (NADPH)
 YHR106w strong similarity to thioredoxin reductases
 YOR074c *CDC21* thymidylate synthase

metabolism of cyclic and unusual nucleotides

YJL005w *CYR1* adenylate cyclase
 YJL050c *APA1* ATP adenyltransferase I
 YDR530c *APA2* ATP adenyltransferase II
 YOL081w *IRA2* GTPase-activating protein for RAS proteins
 YOR380c *PDE2* high affinity 3',5'-cyclic-nucleotide phosphodiesterase
 YPL212c *PUS1* pseudouridine synthase 1
 YGL063w *PUS2* pseudouridine synthase 2

regulation of nucleotide metabolism

YOL081w *IRA2* GTPase-activating protein for RAS proteins
 YOR101w *RAS1* GTP-binding protein
 YNL098c *RAS2* GTP-binding protein
 YDL106c *GRF10* homeodomain protein
 YGL248w *PDE1* low-affinity 3',5'-cyclic-nucleotide phosphodiesterase
 YNL076w *MKS1* negative regulator of RAS-cAMP pathway
 YJL096w putative regulator of purine and/or pyrimidine biosynthesis
 YOL110w *SHR5* RAS suppressor
 YKR099w *BAS1* transcription factor
 YLR014c *PPR1* transcription factor regulating pyrimidine pathway
 YIR023w *DAL81* transcriptional activator for allantoin and GABA catabolic genes
 YNL314w *DAL82* transcriptional activator for allantoin catabolic genes

polynucleotide degradation

YMR287c *MSU1* 3'-5' exonuclease for RNA 3' ss-tail, mitochondrial
 YPL029w *SUV3* ATP-dependent RNA helicase, mitochondrial
 YOR033c *DHS1* exonuclease, interacting with Msh2p
 YKL149c *DBR1* lariat-debranching enzyme
 YGL173c *KEM1* multifunctional nuclease
 YLR363c *NMD4* Nam7p/Upf1p-interacting protein
 YJR132w *NMD5* Nam7p/Upf1p-interacting protein
 YMR080c *NAM7* nonsense-mediated mRNA decay protein
 YGR072w *UPF3* nonsense-mediated mRNA decay protein
 YHR077c *NMD2* nonsense-mediated mRNA decay protein 2
 YJL208c *NUC1* nuclease, mitochondrial
 YMR234w *RNH1* ribonuclease H
 YPL123c similarity to ribonucleases
 YGR195w weak similarity to *P. aeruginosa* RNase PH

nucleotide transport

YMR056c *AAC1* ADP/ATP carrier protein (MCF)
 YBL030c *AAC2* ADP/ATP carrier protein (MCF)
 YBR085w *AAC3* ADP/ATP carrier protein (MCF)
 YER056c *FCY2* purine-cytosine permease
 YER060w *FCY21* purine-cytosine permease
 YOR222w similarity to ADP/ATP carrier proteins
 YPR011c similarity to ADP/ATP carrier proteins and Graves disease carrier protein
 YOR071c similarity to allantoin or uracil transport proteins
 YOR192c similarity to allantoin or uracil transport proteins
 YLR237w similarity to allantoin transport protein
 YGR096w similarity to bovine Graves disease carrier protein
 YHR002w similarity to bovine mitochondrial carrier protein/Grave's disease carrier protein
 YGL186c similarity to hypothetical protein YER060w and weak similarity to Fcy2p
 YBL042c strong similarity to allantoin and uracil transport proteins
 YER060w-a *FCY22* strong similarity to Fcy2p
 YBR021w *FUR4* uracil permease

other nucleotide-metabolism activities

YDR226w *ADK1* adenylate kinase, cytosolic
 YER170w *ADK2* adenylate kinase, mitochondrial
 YDR454c *GUK1* guanylate kinase
 YKL067w *YNK1* nucleoside diphosphate kinase
 YDL125c *HNT1* similarity to protein kinase C inhibitor-I
 YDR305c *HNT2* strong similarity to *S. pombe* diadenosine 5',5''-P₁P₄-tetraphosphate asymmetrical hydrolase

phosphate metabolism

phosphate utilization

YHR044c *DOG1* 2-deoxyglucose-6-phosphate phosphatase

YHR043c *DOG2* 2-deoxyglucose-6-phosphate phosphatase
 YMR282c *AEP2* 2'-O-ribosyl phosphate transferase
 YBR092c *PHO3* constitutive acid phosphatase
 YHR201c *PPX1* exopolyphosphatase
 YBR011c *IPP1* inorganic pyrophosphatase, cytoplasmic
 YMR267w *PPA2* inorganic pyrophosphatase, mitochondrial
 YBR093c *PHO5* repressible acid phosphatase
 YAR071w *PHO11* secreted acid phosphatase
 YHR215w *PHO12* secreted acid phosphatase
 YDL024c strong similarity to acid phosphatase
 YNL330c *RPD3* transcription modifier protein
 YBR243c *ALG7* UDP-N-acetylglucosamine-1-phosphate transferase

regulation of phosphate utilization

YOL001w *PHO80* cyclin
 YGR233c *PHO81* cyclin-dependent kinase inhibitor
 YPL031c *PHO85* cyclin-dependent protein kinase
 YNL121w *GTR1* GTP-binding protein
 YDL106c *GRF10* homeodomain protein
 YHR106w *PHO88* involved in phosphate transport
 YFR034c *PHO4* transcription factor

phosphate transport

YLR348c dicarboxylate carrier protein
 YML123c *PHO84* high-affinity inorganic phosphate/H⁺ symporter
 YJL117w *PHO86* inorganic phosphate transporter
 YBR106w *PHO88* involved in phosphate transport
 YCR037c *PHO87* member of the phosphate permease family
 YJR077c *MIR1* phosphate transport protein, mitochondrial (MCF)
 similarity to membrane protein Pho87p and hypothetical protein YJL198w
 YCR098c *GIT1* similarity to phosphate transporter proteins
 YER053c strong similarity to mitochondrial phosphate carrier protein
 YJL198w strong similarity to Pho87p
 YBR296c strong similarity to phosphate-repressible phosphate permease

other phosphate-metabolism activities

YDL236w *PHO13* 4-nitrophenylphosphatase
 YOR008c *SLG1* weak similarity to *L. mexicana* Imsap2 gene

carbohydrate metabolism

carbohydrate utilization

YLR342w *FKS1* 1,3-D-glucan synthase, catalytic subunit
 YGR032w *GSC2* 1,3-D-glucan synthase, subunit
 YEL011w *GLC3* 1,4-glucan branching enzyme (glycogen branching enzyme)
 YHR044c *DOG1* 2-deoxyglucose-6-phosphate phosphatase
 YHR043c *DOG2* 2-deoxyglucose-6-phosphate phosphatase
 YJL125w *KGD1* 2-oxoglutarate dehydrogenase complex E1 component
 YDR148c *KGD2* 2-oxoglutarate dehydrogenase complex E2 component
 YGR240c *PFK1* 6-phosphofructokinase, α subunit
 YMR205c *PFK2* 6-phosphofructokinase, β subunit
 YIL107c *PFK26* 6-phosphofructose-2-kinase, isoenzyme 1
 YOL136c *PFK27* 6-phosphofructose-2-kinase, isoenzyme 2
 YHR183c *GND1* 6-phosphogluconate dehydrogenase
 YBR001c *NTH2* α,α-trehalase
 YDR074w *TPS2* α,α-trehalose-phosphate synthase, 105K subunit
 YMR261c *TPS3* α,α-trehalose-phosphate synthase, 115K subunit
 YML100w *TSL1* α,α-trehalose-phosphate synthase, 123K subunit
 YBR126c *TPS1* α,α-trehalose-phosphate synthase, 56K subunit
 YJR131w *MNS1* α-1,2-mannosidase
 YDR483w *KRE2* α-1,2-mannosyltransferase
 YER001w *MNN1* α-1,3-mannosyltransferase
 YGL038c *OCH1* α-1,6-mannosyltransferase
 YAL054c *ACS1* acetyl-CoA synthetase
 YLR153c *ACS2* acetyl-CoA synthetase
 YPR026w *ATH1* acid trehalase, vacuolar
 YBR299w *MAL32* α-glucosidase
 YGR292w *MAL12* α-glucosidase of the *MAL1* locus
 YOR377w *ATF1* alcohol acetyltransferase
 YOL086c *ADH1* alcohol dehydrogenase I
 YMR303c *ADH2* alcohol dehydrogenase II
 YMR083w *ADH3* alcohol dehydrogenase III
 YGL256w *ADH4* alcohol dehydrogenase IV
 YBR145w *ADH5* alcohol dehydrogenase V
 YGR177c *ATF2* alcohol O-acetyltransferase
 YCL040w *GLK1* aldehyde-specific glucokinase
 YGL156w *AMS1* α-mannosidase
 YGL027c *CWH41* β-1,6-galactan assembly protein
 YHR101c *BIG1* big cells phenotype
 YBR110w *ALG1* β-mannosyltransferase
 YBR084w *MIS1* C1-tetrahydrofolate synthase, mitochondrial
 YNL322c *KRE1* cell-wall protein
 YJL174w *KRE9* cell-wall synthesis protein
 YJL099w *CHS6* chitin biosynthesis protein
 YNL192w *CHS1* chitin synthase I
 YBR038w *CHS2* chitin synthase II
 YBR023c *CHS3* chitin synthase III
 YNR001c *CIT1* citrate (si)-synthase, mitochondrial

YPR001w *CIT3* citrate (si)-synthase, mitochondrial
 YCR005c *CIT2* citrate (si)-synthase, peroxisomal
 YML086c *ALO* D-arabinino-1,4-lactone oxidase
 YNL071w *LAT1* dihydroloipoamide S-acetyltransferase
 YDL174c *DLD1* D-lactate ferricytochrome c oxidoreductase (D-LCR)
 YER062c *HOR2* DL-glycerol phosphatase
 YJL053w *RHR2* DL-glycerol phosphatase
 YJL227c *ALG5* dolichol-P-glucose synthetase
 YPR183w *DPM1* dolichyl-phosphate β-D-mannosyltransferase
 YJR143c *PMT4* dolichyl-phosphate-mannose-protein O-mannosyl transferase
 YOR095c *RK11* D-ribose-5-phosphate ketol-isomerase
 YJL121c *POS18* D-ribose-5-phosphate 3-epimerase
 YGR282c *BGL2* endo-1,3—glucanase of the cell wall
 YLR286c *CTS1* endochitinase
 YGR254w *ENO1* enolase I (2-phosphoglycerate dehydratase)
 YHR174w *ENO2* enolase II (2-phosphoglycerate dehydratase)
 YOR393w *ERR1* enolase-related protein
 YPL281c *ERR2* enolase-related protein
 YPR190w *SPR1* exo-1,3—glucanase
 YDR261c *EXG2* exo-1,3—glucanase minor isoform
 YLR300w *EXG1* exo-1,3—glucanase (I/II), major isoform
 YIR019c *STA1* extracellular α-1,4-glucan glucosidase
 YLR377c *FBP1* fructose-1,6-bisphosphatase
 YJL155c *FBP26* fructose-2,6-bisphosphatase
 YKL060c *FBA1* fructose-bisphosphate aldolase
 YDL049c *KNH1* functional homologue of Kre9p
 YBR020w *GAL1* galactokinase
 YDR009w *GAL3* galactokinase
 YOR120w *GCY1* galactose-induced protein of aldo/keto reductase family
 YPR159w *KRE6* glucan synthase subunit
 YGR143w *SKN1* glucan synthase subunit
 YKL104c *GFA1* glucosamine-fructose-6-phosphate transaminase
 YNL241c *ZWF1* glucose-6-phosphate dehydrogenase
 YBR196c *PGI1* glucose-6-phosphate isomerase
 YBR229c *ROT2* glucosidase II, catalytic subunit
 YOR002w *ALG6* glucosyltransferase
 YOR067c *ALG8* glucosyltransferase
 YJL052w *TDH1* glyceraldehyde-3-phosphate dehydrogenase 1
 YJR009c *TDH2* glyceraldehyde-3-phosphate dehydrogenase 2
 YGR192c *TDH3* glyceraldehyde-3-phosphate dehydrogenase 3
 YHL032c *GUT1* glycerol kinase
 YDL022w *GPD1* glycerol-3-phosphate dehydrogenase (NAD⁺), cytoplasmic
 YOL059w *GPD3* glycerol-3-phosphate dehydrogenase (NAD⁺), mitochondrial
 YJL155c *GUT2* glycerol-3-phosphate dehydrogenase, mitochondrial
 YPR160w *GPH1* glycogen phosphorylase
 YFL014w *HSP12* heat-shock protein
 YFR053c *HXK1* hexokinase I
 YGL253w *HXK2* hexokinase II
 YDL013w *HEX3* hexose metabolism-related protein
 YDL182w *LYS20* homocitrate synthase
 YOR126c *EST2* isoamyl acetate hydrolytic enzyme
 YNL037c *IDH1* isocitrate dehydrogenase (NAD⁺) subunit 1, mitochondrial
 YOR136w *IDH2* isocitrate dehydrogenase (NAD⁺) subunit 2, mitochondrial
 YDL066w *IDP1* isocitrate dehydrogenase (NADP⁺), mitochondrial
 YLR174w *IDP2* isocitrate dehydrogenase, cytosolic
 YER065c *ICL1* isocitrate lyase
 YOR336w *KRE5* killer toxin-resistance protein
 YML054c *CYB2* lactate dehydrogenase cytochrome b2
 YDL168w *SFA1* long-chain alcohol dehydrogenase
 YOL126c *MDH2* malate dehydrogenase, cytoplasmic
 YKL085w *MDH1* malate dehydrogenase, mitochondrial
 YDL078c *MDH3* malate dehydrogenase, peroxisomal
 YNL117w *MLS1* malate synthase 1
 YJR031c *DAL7* malate synthase 2
 YDL055c *PSA1* mannose-1-phosphate guanylyltransferase
 YER003c *PM40* mannose-6-phosphate isomerase
 YGL065c *ALG2* mannosyltransferase
 YNL219c *ALG9* mannosyltransferase
 YKR061w *KTR2* mannosyltransferase
 YDL095w *PMT1* mannosyltransferase
 YAL023c *PMT2* mannosyltransferase
 YOR321w *PMT3* mannosyltransferase
 YBL082c *RHK1* mannosyltransferase
 YJL139c *YUR1* mannosyltransferase
 YJL153c *INO1* myo-inositol-1-phosphate synthase
 YPL175w *SPT14* N-acetylglucosaminyltransferase
 YDR001c *NTH1* neutral trehalase (α,α-trehalase)
 YPR006c *ICL2* non-functional isocitrate lyase
 YJL002c *OST1* oligosaccharyltransferase α subunit
 YEL002c *WBP1* oligosaccharyltransferase β subunit
 YMR149w *SWP1* oligosaccharyltransferase δ subunit
 YOR103c *OST2* oligosaccharyltransferase ε subunit
 YOR085w *OST3* oligosaccharyltransferase γ subunit
 YDL232w *OST4* oligosaccharyltransferase subunit
 YGL022w *STT3* oligosaccharyltransferase subunit
 YEL058w *PCM1* phosphoacetylglucosamine mutase
 YKR097w *PKC1* phosphoenolpyruvate carboxylkinase
 YMR105c *PGM2* phosphoglucomutase, major isoform
 YKL127w *PGM1* phosphoglucomutase, minor isoform
 YGR256w *GND2* phosphogluconate dehydrogenase
 YCR012w *PGK1* phosphoglycerate kinase
 YKL152c *GPM1* phosphoglycerate mutase

| | | | | | | | |
|---------|--------------|--|---------|--|---------|---------------|--|
| YOL056w | <i>GPM3</i> | phosphoglycerate mutase | YJL218w | strong similarity to <i>E. coli</i> galactoside O-acetyltransferase | YCR081w | <i>SRB8</i> | DNA-directed RNA polymerase II holoenzyme and Kornberg's mediator (SRB) subcomplex subunit |
| YFL045c | <i>SEC53</i> | probable 1,3-glucan synthase subunit | YDR248c | strong similarity to <i>E. coli</i> thermoresistant gluconokinase | YNL025c | <i>SSN8</i> | DNA-directed RNA polymerase II holoenzyme and Kornberg's mediator (SRB) subcomplex subunit, cyclin C homologue |
| YCR034w | <i>GNS1</i> | putative α -1,2-mannosyltransferase | YIL172c | strong similarity to Fsp2p | YLR071c | <i>RGR1</i> | DNA-directed RNA polymerase II holoenzyme subunit |
| YIL045w | <i>PIG2</i> | Protein interacting with Gsy2p | YDL037c | strong similarity to glucan 1,4-glucosidase | YBR253w | <i>SRB6</i> | DNA-directed RNA polymerase II suppressor protein |
| YIL085c | <i>KTR7</i> | putative α -1,2-mannosyltransferase | YDR516c | strong similarity to glucokinase | YOR047c | <i>STD1</i> | dosage-dependent modulator of glucose repression |
| YNL029c | <i>KTR5</i> | putative mannosyltransferase | YOR299w | strong similarity to hypothetical protein YMR237w and similarity to Chs6p | YIL155c | <i>FBP28</i> | fructose-2,6-bisphosphatase |
| YGR199w | <i>PMT6</i> | putative mannosyltransferase | YMR237w | strong similarity to hypothetical protein YOR299w and similarity to CHS6 protein | YPL037c | <i>EGD1</i> | GAL4 DNA-binding enhancer protein |
| YGL062w | <i>PYC1</i> | pyruvate carboxylase, isozyme 1 | YNL009w | strong similarity to phosphopyruvate hydratases | YDR009w | <i>GAL3</i> | galactokinase |
| YBR218c | <i>PYC2</i> | pyruvate carboxylase, isozyme 2 | YJL216c | strong similarity to polygalacturonases | YLL016w | <i>SDC25</i> | GDP/GTP exchange factor |
| YDR081c | <i>PDC2</i> | pyruvate decarboxylase regulatory protein | YGR287c | strong similarity to pyruvate decarboxylases | YLR310c | <i>CDC25</i> | GDP/GTP exchange factor for Ras1p and Ras2p |
| YDL044c | <i>PDC1</i> | pyruvate decarboxylase, isozyme 1 | YOR099w | strong similarity to pyruvate kinase | YGR070w | <i>ROM1</i> | GDP/GTP exchange protein for Rho1p |
| YLR134w | <i>PDC5</i> | pyruvate decarboxylase, isozyme 2 | YDR368w | strong similarity to rumen fungus β -succinyl-CoA synthetase | YLR371w | <i>ROM2</i> | GDP/GTP exchange protein for Rho1p |
| YGR087c | <i>PDC6</i> | pyruvate decarboxylase, isozyme 3 | YPR1 | strong similarity to members of the aldo/keto reductase family | YDR176w | <i>NGG1</i> | general transcriptional adaptor or co-activator |
| YER178w | <i>PDA1</i> | pyruvate dehydrogenase (lipoaamide) α subunit | YDL021w | strong similarity to phosphoglycerate mutase Gpm1p | YER054c | <i>GIP2</i> | Glc7p-interacting protein |
| YBR221c | <i>PDB1</i> | pyruvate dehydrogenase (lipoaamide) β subunit | YMR323w | strong similarity to phosphopyruvate hydratases | YNL236w | <i>SIN4</i> | global regulator protein |
| YGR193c | <i>PDX1</i> | pyruvate dehydrogenase complex protein X | YR153w | strong similarity to succinate dehydrogenase flavoprotein | YHL025w | <i>SNF6</i> | global transcription activator |
| YAL038w | <i>CDC19</i> | pyruvate kinase | YDL080c | strong similarity to succinate dehydrogenase membrane anchor subunit for Sdh2p | YER027c | <i>GAL83</i> | glucose repression protein |
| YNL048w | <i>ALG11</i> | required for asparagine-linked glycosylation | YOR347c | strong similarity to succinate dehydrogenase flavoprotein | YNL199c | <i>GCR2</i> | glycolytic genes transcriptional activator |
| YCR036w | <i>RBK1</i> | ribokinase | YGR244c | strong similarity to rumen fungus β -succinyl-CoA synthetase | YOR101w | <i>RAS1</i> | GTP-binding protein |
| YJL137c | <i>GLG2</i> | self-glucosylating initiator of glycogen synthesis | YKL029c | strong similarity to <i>S. pombe</i> malate oxidoreductase | YNL098c | <i>RAS2</i> | GTP-binding protein |
| YMR306w | | similarity to 1,3-glucan synthases | YBL001c | strong similarity to <i>S. xylosus</i> glucose kinase | YPR165w | <i>RHO1</i> | GTP-binding protein of the RHO subfamily of RAS-like proteins |
| YAL060w | <i>FUN49</i> | similarity to alcohol/sorbitol dehydrogenase | YLR164w | strong similarity to Sdh4p | YNL090w | <i>RHO2</i> | GTP-binding protein of the RHO subfamily of RAS-like proteins |
| YAL061w | <i>FUN50</i> | similarity to alcohol/sorbitol dehydrogenase | YDL246c | strong similarity to Sor1p | YDR420w | <i>HKR1</i> | <i>Hansenula</i> MrakII k9 killer toxin-resistance protein |
| YMR318c | | similarity to alcohol-dehydrogenase | YJL045w | strong similarity to succinate dehydrogenase flavoprotein | YGL253w | <i>HKK2</i> | hexokinase II |
| YDL124w | | similarity to aldose reductases | YGR043c | strong similarity to transaldolase | YPL002c | <i>SNF8</i> | involved in glucose derepression |
| YHR204w | | similarity to α -mannosidases | YNR071c | strong similarity to UDP-glucose 4-epimerase Gal10p | YOR125c | <i>CAT5</i> | involved in glucose repression |
| YPL088w | | similarity to aryl-alcohol dehydrogenases | YDR384c | strong similarity to <i>Y. lipolytica</i> GPR1 gene | YML048w | <i>EFF2</i> | involved in glucose repression |
| YML070w | | similarity to <i>C. freundii</i> dihydroxyacetone kinase | YHR210c | strong similarity to UDP-glucose-4-epimerase | YNL201c | | involved in regulation of carbon metabolism |
| YIL124w | | similarity to <i>C. perfringens</i> nanH protein | YKL148c | succinate dehydrogenase flavoprotein | YGR227w | <i>DIE2</i> | ITR1 expression promoting protein |
| YDR371w | | similarity to chitinases | YDR178w | succinate dehydrogenase membrane anchor subunit for Sdh2p | YBR297w | <i>MAL33</i> | maltose fermentation regulatory protein |
| YIR036c | | similarity to <i>E. coli</i> 3-ketoacyl-ACP reductase | YIL162w | sucrose hydrolyzing enzyme | YGR288w | <i>MAL13</i> | maltose pathway regulatory protein |
| YBR149w | | similarity to Gcy1p and aldose reductases | YJR075w | suppressor of <i>pkc1</i> | YLR131c | <i>ACE2</i> | metallothionein expression activator |
| YBR056w | | similarity to glucan 1,3-glucosidase | YPR074c | transketolase 1 | YDR422c | <i>SIP1</i> | multicopy suppressor of <i>snf1</i> |
| YIL169c | | similarity to glucan 1,4-glucosidase | YBR117c | transketolase 2 | YML051w | <i>GAL80</i> | negative regulator for expression of galactose-induced genes |
| YOL155c | | similarity to glucan 1,4-glucosidase Mal5p | YDR060c | triose-phosphate isomerase | YNL076w | <i>MKS1</i> | negative regulator of RAS-cAMP pathway |
| YNL274c | | similarity to glycerate- and formate-dehydrogenases | YBR019c | UDP-glucose 4-epimerase | YIL119c | <i>RPI1</i> | negative regulator of RAS-cAMP pathway |
| YPL113c | | similarity to glycerate dehydrogenases | YBR018c | UDP-glucose-hexose-1-phosphate uridylyltransferase | YLR025w | <i>SNF7</i> | nuclear protein |
| YPR184w | | similarity to glycogen debranching enzymes | YFR015c | UDP-glucose-starch glucosyltransferase, isoform 1 | YGL115w | <i>SNF4</i> | nuclear regulatory protein |
| YDL131w | | similarity to homocitrate synthases and isopropylmalate synthases | YLR258w | UDP-glucose-starch glucosyltransferase, isoform 2 | YLR273c | <i>PIG1</i> | protein interacting with Gsy2p |
| YJR096w | | similarity to <i>Leishmania</i> reductase | YBR243c | UDP-N-acetylglucosamine-1-phosphate transferase | YIL045w | <i>PIG2</i> | protein interacting with Gsy2p |
| YKR096w | | similarity to mitochondrial aldehyde dehydrogenase Ald1p | YKL035w | UTP-glucose-1-phosphate uridylyltransferase | YDL006w | <i>PTC1</i> | protein ser/thr phosphatase 2c |
| YGL257c | | similarity to Mnn1p | YKR043c | weak similarity to phosphoglycerate mutase | YDL134c | <i>PPH21</i> | protein ser/thr phosphatase PP2A-1 |
| YIL014w | | similarity to Mnn1p | YOR283w | weak similarity to phosphoglycerate mutases | YDL188c | <i>PPH22</i> | protein ser/thr phosphatase PP2A-2 |
| YEL020c | | similarity to <i>O. formigenes</i> oxalyl-CoA decarboxylase | YGR240c | 6-phosphofructokinase, α subunit | YBL061c | <i>SKT5</i> | protoplast regeneration and killer toxin resistance protein |
| YDL093w | <i>PMT5</i> | similarity to O-mannosyltransferases Pmt1p-Pmt4p | YMR205c | 6-phosphofructokinase, β subunit | YOL110w | <i>SHR5</i> | RAS suppressor |
| YMR278w | | similarity to phosphomannomutases | YIL107c | 6-phosphofructokinase, β subunit | YJR095w | <i>ACR1</i> | regulator of acetyl-CoA synthetase activity |
| YDR307w | | similarity to Pmt1p | YOL120c | 6-phosphofructose-2-kinase, isoenzyme 1 | YKL038w | <i>RG1</i> | regulator of glucose induced genes |
| YDR380w | | similarity to pyruvate decarboxylases | YOL136c | 6-phosphofructose-2-kinase, isoenzyme 2 | YDR028c | <i>REG1</i> | regulatory subunit for protein phosphatase Glc7p |
| YDR245w | <i>MNN10</i> | similarity to <i>S. pombe</i> galactosyltransferase | YBR126c | α -D-trehalose-phosphate synthase, 56K subunit | YMR311c | <i>GLC8</i> | regulatory subunit for protein ser/thr phosphatase Glc7p |
| YLR070c | | similarity to sugar dehydrogenases | YHR047c | alanine/arginine aminopeptidase | YBR050c | <i>REG2</i> | regulatory subunit of type I protein phosphatase |
| YNR059w | | similarity to α -1,3-mannosyltransferase | YGR229c | β -1,3-glucan synthesis protein | YDR277c | <i>MTH1</i> | repressor of hexose transport genes |
| YHL012w | | similarity to UDP glucose pyrophosphorylase | YOR344c | basic helix-loop-helix transcription factor | YNR052c | <i>POP2</i> | required for glucose derepression |
| YJR159w | <i>SOR1</i> | sorbitol dehydrogenase | YOR344c | basic helix-loop-helix transcription factor | YJR090c | <i>GRR1</i> | required for glucose repression and for glucose and cation transport |
| YIL099w | <i>SGA1</i> | sporulation specific glucan 1,4-glucosidase | YGR166w | β -glucan synthesis-associated protein | YGL252c | <i>RTG2</i> | retrograde regulation protein |
| YER096w | <i>SHC1</i> | sporulation-specific protein | YBL103c | bHLH/zip transcription factor | YJR076c | <i>CDC11</i> | septin |
| YLR307w | <i>CDA1</i> | sporulation-specific chitin deacetylase | YPL161c | bud emergence protein | YHR107c | <i>CDC12</i> | septin |
| YLR308w | <i>CDA2</i> | sporulation-specific chitin deacetylase | YGL209w | C2H2 zinc-finger protein | YER133w | <i>GLC7</i> | ser/thr phosphoprotein phosphatase 1, catalytic subunit |
| YNL331c | | strong similarity to aryl-alcohol reductase | YIL033c | cAMP dependent protein kinase, regulatory subunit | YOR178c | <i>GAC1</i> | ser/thr phosphoprotein phosphatase 1, regulatory subunit |
| YCR107w | | strong similarity to dihydroliipoamide dehydrogenases | YDR477w | carbon catabolite derepressing ser/thr protein kinase | YLR113w | <i>HOG1</i> | ser/thr protein kinase of MAP kinase (MAPK) family |
| YPL017c | | strong similarity to dihydroliipoamide dehydrogenases | YCR002c | cell division control protein | YLR345w | | similarity to 6-phosphofructo-2-kinase |
| YBR205w | <i>KTR3</i> | strong similarity to α -1,2-mannosyltransferase | YLR330w | chitin synthesis protein | YCR028c | <i>FEN2</i> | similarity to allantoin permease transporter |
| YBR199w | <i>KTR4</i> | strong similarity to α -1,2-mannosyltransferase | YDR073w | component of SWI/SNF transcription activator complex | YFL053w | | similarity to <i>C. freundii</i> dihydroxyacetone kinase |
| YPL053c | <i>KTR6</i> | strong similarity to α -1,2-mannosyltransferase | YOR290c | component of SWI/SNF transcription activator complex | YDL225w | | similarity to Cdc11p, Cdc3p and human CDC10 protein |
| YJL221c | <i>FSP2</i> | strong similarity to α -D-glucosidase | YBR289w | component of SWI/SNF transcription activator complex | YHR193c | <i>EGD2</i> | similarity to human α -NAC |
| YOL157c | | strong similarity to α -glucosidases | YPL016w | component of SWI/SNF transcription activator complex | YDL203c | | similarity to Skt5p |
| YCR105w | | strong similarity to alcohol dehydrogenases | YPL042c | component of SWI/SNF transcription activator complex | YNL257c | <i>SIP3</i> | Snf1p protein kinase interacting protein |
| YDL243c | | strong similarity to aryl-alcohol dehydrogenase | YNL216w | component of SWI/SNF transcription activator complex | YLR150w | <i>MPT4</i> | specific affinity for guanine-rich quadruplex nucleic acids |
| YJR155w | | strong similarity to aryl-alcohol dehydrogenase | YPL051w | DNA-directed RNA polymerase II holoenzyme and Kornberg's mediator (SRB) subcomplex subunit | YFL062w | | strong similarity to Mal63p, Mal23p and Mal33p |
| YFL056c | | strong similarity to aryl-alcohol dehydrogenases | YDR443c | DNA-directed RNA polymerase II holoenzyme and Kornberg's mediator (SRB) subcomplex subunit | YHR155w | | strong similarity to Snf1p-interacting protein Sip3p |
| YFL057c | | strong similarity to aryl-alcohol dehydrogenases | YCR002c | cell division control protein | YNR002c | <i>FUN34</i> | strong similarity to <i>Y. lipolytica</i> glyoxylate pathway regulator GPR1 |
| YOL165c | | strong similarity to aryl-alcohol dehydrogenases | YLR330w | chitin synthesis protein | YCR010c | | strong similarity to <i>Y. lipolytica</i> GPR1 protein and Fun34p |
| YKR027w | | strong similarity to Chs6p | YDR073w | component of SWI/SNF transcription activator complex | YIL154c | <i>(IMP2)</i> | sugar utilization regulatory protein |
| YHR104w | | strong similarity to D-xylose 1-dehydrogenase | YOR290c | component of SWI/SNF transcription activator complex | YPL026c | <i>SKS1</i> | suppressor kinase of <i>snf3</i> |
| YEL070w | | strong similarity to <i>E. coli</i> D-mannonate oxidoreductase | YBR289w | component of SWI/SNF transcription activator complex | YPL129w | <i>ANC1</i> | TFIIIF subunit (transcription initiation factor), 30K |
| YNR073c | | strong similarity to <i>E. coli</i> D-mannonate oxidoreductase, identical to YEL070w | YPL016w | component of SWI/SNF transcription activator complex | YPL248c | <i>GAL4</i> | transcription factor |

YMR280c *CAT8* transcription factor involved in gluconeogenesis
 YMR043w *MCM1* transcription factor of the MADS box family
 YJL176c *SWI3* transcription regulatory protein
 YPL075w *GCR1* transcriptional activator
 YOL116w *MSN1* transcriptional activator
 YKL062w *MSN4* transcriptional activator
 YGL035c *MIG1* transcriptional repressor
 YIL147c *SLN1* two-component signal transducer
 YLR006c *SSK1* two-component signal transducer
 YIL128c *PBS2* tyrosine protein kinase of the MAP kinase kinase family
 YLR258w *GSY2* UDP-glucose-starch glucosyltransferase, isoform 2
 YDR216w *ADR1* zinc-finger transcription factor

carbohydrate transport

YCL040w *GLK1* aldohexose specific glucokinase
 YKL217w *JEN1* carboxylic acid transporter protein
 YBR291c *CTP1* citrate transport protein, mitochondrial (MCF)
 YLR348c *GAL2* dicarboxylate carrier protein
 YLR081w *GAL2* galactose (and glucose) permease
 YGR289c *AGT1* general α-glucoside permease
 YLL043w *FPS1* glycerol channel protein
 YNL318c *HXT14* hexose transport protein
 YJL214w *HXT8* hexose transport protein
 YJL219w *HXT9* hexose transport protein
 YFL011w *HXT10* hexose transporter
 YDL194w *SNF3* high-affinity glucose transporter
 YEL069c *HXT13* high-affinity hexose transporter
 YMR011w *HXT2* high-affinity hexose transporter
 YDR343c *HXT6* high-affinity hexose transporter
 YDR342c *HXT7* high-affinity hexose transporter
 YOL156w *HXT11* low-affinity glucose transporter
 YHR094c *HXT1* low-affinity hexose transporter
 YDR345c *HXT3* low-affinity hexose transporter
 YDR497c *ITR1* major myo-inositol permease
 YBR298c *MAL31* maltose permease
 YHR092c *HXT4* moderate- to low-affinity glucose transporter
 YOL103w *ITR2* myo-inositol transport protein
 YIR035c similarity to *C. lanceolata* 3-oxoacyl-[acyl-carrier-protein] reductase
 YBR241c similarity to glucose transport proteins
 YGL104c similarity to glucose transport proteins
 YRP021c similarity to human citrate transporter protein
 YDR387c similarity to Itr1p and Itr2p
 YDL199c similarity to sugar transporter proteins
 YFL040w similarity to yeast glucose transport proteins
 YHR096c *HXT5* strong similarity to hexose transporters
 YDL245c *HXT15* strong similarity to Hxt17p and Hxt7p
 YJR160c strong similarity to Mal3Tp
 YDR536w *STL1* strong similarity to members of the sugar permease family
 YDL247w strong similarity to sugar transport proteins
 YIL170w *HXT12* strong similarity to sugar transport proteins
 YIL171w *HXT12* strong similarity to sugar transport proteins
 YJR158w *HXT16* strong similarity to sugar transport proteins
 YNR072w *HXT17* sugar transport protein
 YDL138w *RGT2* suppressor of *snf3* mutant

lipid, fatty-acid and sterol metabolism

lipid, fatty-acid and sterol biosynthesis

YML075c *HMG1* 3-hydroxy-3-methylglutaryl-CoA reductase 1
 YLR450w *HMG2* 3-hydroxy-3-methylglutaryl-CoA reductase 2
 YML126c *HMG5* 3-hydroxy-3-methylglutaryl-CoA synthase
 YPL028w *ERG10* acetyl-CoA C-acetyltransferase, cytosolic
 YNR016c *ACC1* acetyl-CoA carboxylase
 YCR048w *ARE1* acyl-CoA sterol acyltransferase
 YNR019w *ARE2* acyl-CoA sterol acyltransferase
 YIL009w *FAA3* acyl-CoA synthase
 YGR037c *ACB1* acyl-CoA-binding protein (diazepam-binding inhibitor)
 YBR222c *PCS60* AMP-binding protein, peroxisomal
 YER061c *CEM1* β-keto-acyl-ACP synthase
 YNL280c *ERG24* C-14 sterol reductase
 YMR015c *ERG5* C-22 sterol desaturase
 YGR060w *ERG25* C-4 sterol methyl oxidase
 YLR056w *ERG3* C-5 sterol desaturase
 YMR202w *ERG2* C-8 sterol isomerase
 YML042w *CAT2* carnitine O-acetyltransferase
 YPR113w *PIS1* CDP diacylglycerol-inositol 3-phosphatidyltransferase
 YER026c *CHO1* CDP-diacylglycerol serine O-phosphatidyltransferase
 YBR029c *CDS1* CDP-diacylglycerol synthase
 YLR133w *CK1* choline kinase
 YGR007w *MUQ1* choline phosphate cytidyltransferase
 YGR202c *PCT1* cholinephosphate cytidyltransferase
 YNL111c *CYB5* cytochrome b5
 YHR007c *ERG11* cytochrome P450 lanosterol 14α-demethylase
 YHR190w *ERG9* farnesyl-diphosphate farnesyltransferase
 YJL167w *ERG20* farnesyl-pyrophosphate synthetase
 YDL052c *SLC1* fatty acyltransferase
 YDL231w *FAS2* fatty-acyl-CoA synthase, α subunit
 YKL182w *FAS1* fatty-acyl-CoA synthase, β subunit

YPL069c *BTS1* geranylgeranyl diphosphate synthase
 YGL126w *SCS3* inositol phospholipid synthesis protein
 YOR237w *HES1* involved in ergosterol biosynthesis
 YPL145c *KE51* involved in ergosterol biosynthesis
 YPL117c *ID1* isopentenyl-diphosphate δ-isomerase
 YHR072w *ERG7* lanosterol synthase
 YMR246w *FAA4* long-chain-fatty-acid-CoA ligase
 YNL219c *ALG9* mannosyltransferase
 YJR073c *OPI3* methylene-fatty-acyl-phospholipid synthase
 YNR043w *MVD1* mevalonate pyrophosphate decarboxylase
 YPL076w *GPI2* N-acetylglucosaminyl-phosphatidylinositol biosynthetic protein
 YPL175w *SPT14* N-acetylglucosaminyltransferase
 YGR157w *CHO2* phosphatidylethanolamine N-methyltransferase
 YDL142c *PGS1* phosphatidylglycerophosphate synthase
 YNL169c *PSD1* phosphatidylserine decarboxylase 1
 YGR170w *PSD2* phosphatidylserine decarboxylase 2
 YCL004w *PEL1* phosphatidylserine synthase
 YMR220w *ERG8* phosphomevalonate kinase
 YCR034w *GNS1* probable 1,3–glucan synthase subunit
 YGL001c putative 3–hydroxysteroid dehydrogenase
 YJL196c *ELO1* required for elongation of fatty acids
 YGR216c *GPI1* required for N-acetylglucosaminyl phosphatidylinositol synthesis
 YML008c *ERG6* S-adenosyl-methionine 8-24-sterol-c-methyltransferase
 YMR296c *LCB1* serine C-palmitoyltransferase subunit
 YDR062w *LCB2* serine C-palmitoyltransferase subunit
 YLR058c *SHM2* serine hydroxymethyltransferase, cytoplasmic
 YBR263w *SHM1* serine hydroxymethyltransferase, mitochondrial
 YMR207c *HFA1* similarity to acetyl-CoA carboxylase
 YDR147w similarity to choline kinase
 YBR159w similarity to human 17–hydroxysteroid dehydrogenase
 YDR376w *ARH1* similarity to human adrenodoxin reductase
 YML131w similarity to human leukotriene b4 12-hydroxydehydrogenase
 YAR042w *OSH1* similarity to human oxysterol binding protein
 YDR208w *MSS4* similarity to human PI 5-kinase
 YHR001w similarity to Kes1p
 YKR003w similarity to Kes1p, Hes1p and Osh1p
 YDL019c similarity to Osh1p
 YHR073w similarity to Osh1p, Hes1p, Kes1p
 YGR175c *ERG1* squalene monooxygenase
 YGL055w *OLE1* stearyl-CoA desaturase
 YGL012w *ERG4* sterol C-24 reductase
 YLR372w *SUR4* sterol isomerase
 YKL192c strong similarity to acyl-carrier proteins
 YNL045w strong similarity to human leukotriene-A4 hydrolase

breakdown of lipids and phospholipids

YPL288w *PLC1* 1-phosphatidylinositol-4,5-bisphosphate phosphodiesterase
 YGL205w *POX1* acyl-CoA oxidase
 YMR008c *PLB1* phospholipase B (lysophospholipase)
 YKR031c *SPO14* phospholipase D
 YLR020c similarity to triacylglycerol lipase
 YLL012w similarity to triacylglycerol lipases
 YJL068c strong similarity to human esterase D
 YOL011w strong similarity to phospholipases
 YMR006c strong similarity to P1b1p
 YKL140w *TGL1* triacylglycerol lipase
 YDR058c *TGL2* triacylglycerol lipase
 YJR107w weak similarity to acylglycerol lipase
 YBR204c weak similarity to peroxisomal serine-acyl lipase

lipid, fatty-acid and sterol utilization

YIL160c *POT1* acetyl-CoA C-acyltransferase, peroxisomal
 YIL009w *FAA3* acyl-CoA synthase
 YMR013c *SEC59* dolichol kinase
 YDR331w *GPI8* essential for GPI-anchor attachment
 YDR410c *STE14* farnesyl cysteine carboxyl-methyltransferase
 YPL172c *COX10* farnesyl transferase
 YOR370c *MSI4* geranylgeranyltransferase regulatory subunit
 YGL155w *CDC43* geranylgeranyltransferase type I β subunit
 YPR176c *BET2* geranylgeranyltransferase type II β subunit
 YJL031c *BET4* geranylgeranyltransferase, α subunit
 YFL014w *HSP12* heat-shock protein
 YBR003w *COQ1* hexaprenyl pyrophosphate synthetase
 YKR009c *FOX2* hydratase-dehydrogenase-epimerase, peroxisomal
 YOR317w *FAA1* long-chain-fatty-acid-CoA ligase
 YER015w *FAA2* long-chain-fatty-acid-CoA ligase
 YMR246w *FAA4* long-chain-fatty-acid-CoA ligase
 YDL078c *MDH3* malate dehydrogenase, peroxisomal
 YLR195c *NMT1* N-myristoyltransferase
 YJR066w *TOR1* phosphatidylinositol 3-kinase
 YKL019w *RAM2* protein farnesyltransferase, α subunit
 YDL090c *RAM1* protein farnesyltransferase, β subunit
 YHR013c *ARD1* protein N-acetyltransferase subunit
 YDL040c *NAT1* protein N-acetyltransferase subunit

regulation of lipid, fatty-acid and sterol biosynthesis

YBL015w *ACH1* acetyl-CoA hydrolase
 YDR123c *INO2* basic helix-loop-helix (BHLH) transcription factor
 YOL108c *INO4* basic helix-loop-helix transcription factor

YGL162w *SUT1* hypoxic protein involved in sterol uptake
 YMR208w *ERG12* mevalonate kinase
 YHL020c *OPI1* negative regulator of phospholipid biosynthesis pathway
 YDR207c *UME6* negative transcriptional regulator
 YAL051w *OAF1* peroxisome proliferating transcription factor
 YOR363c *PIP2* peroxisome proliferating transcription factor
 YNL169c *PSD1* phosphatidylserine decarboxylase 1
 YAL013w *DEP1* regulator of phospholipid metabolism
 YMR207c *HFA1* similarity to acetyl-CoA carboxylase
 YCR028c *FEN2* similarity to allantoate permease transporter
 YMR043w *MCM1* transcription factor of the MADS box family
 YOL004w *SIN3* transcription regulatory protein

lipid and fatty-acid transport

YGR037c *ACB1* acyl-CoA-binding protein (diazepam-binding inhibitor)
 YNL130c *CPT1* diacylglycerol cholinephosphotransferase
 YBR041w *FAT1* fatty-acid transporter
 YKL188c *PAT1* long-chain-fatty-acid transporter
 YPL147w *PXA1* long-chain-fatty-acid transporter
 YOR317w *FAA1* long-chain-fatty-acid-CoA ligase
 YER015w *FAA2* long-chain-fatty-acid-CoA ligase
 YMR246w *FAA4* long-chain-fatty-acid-CoA ligase
 YAR035w *YAT1* outer carnitine acetyltransferase, mitochondrial
 YCL043c *PDI1* protein disulphide-isomerase
 YER024w similarity to carnitine O-acetyltransferase
 YKL174c similarity to choline transport protein
 Ctr1p
 YNR056c similarity to choline transport protein
 Ctr1p
 YHR123w *EPT1* sn-1,2-diacylglycerol ethanolamine- and cholinephosphotransferase

lipid and fatty-acid binding

YHR001w similarity to Kes1p
 YMR020w *FMS1* suppressor of fenpropimorph resistance mutation *fen2*

other lipid, fatty-acid and sterol metabolism activities

YBR036c *CSG2* calcium dependent regulatory protein
 YMR079w *SEC14* phosphatidylinositol/phosphatidylcholine transfer protein
 YNL284c similarity to Sec14p
 YKL091c strong similarity to Sec14p
 YIL002c *SIH1* synaptotagmin homologue 1
 YDR302w weak similarity to human GPI-anchor biosynthesis protein

biosynthesis of vitamins, cofactors and prosthetic groups

metabolism of vitamins, cofactors and prosthetic groups

YOL096c *COQ3* 3,4-dihydroxy-5-hexaprenylbenzoate methyltransferase
 YDR232w *HEM1* 5-aminolevulinatase synthase
 YOL143c *RIB4* 6,7-dimethyl-8-ribityllumazine synthase
 YGR286c *BI02* biotin synthetase
 YGR204w *AD53* C1-tetrahydrofolate synthase, cytoplasmic
 YBR084w *MIS1* C1-tetrahydrofolate synthase, mitochondrial
 YDR044w *HEM13* coproporphyrinogen III oxidase
 YPL132w *COX11* cytochrome c oxidase assembly protein
 YNR058w *BI03* DAPA aminotransferase
 YDR487c *RIB3* DBP synthase
 YOL066c *RIB2* DRAP deaminase
 YPL172c *COX10* farnesyl transferase
 YOR176w *HEM15* ferrochelatase
 YDL045c *FAD1* flavin adenine dinucleotide (FAD) synthetase
 YGR267c *FOL2* GTP cyclohydrolase I
 YBL033c *RIB1* GTP cyclohydrolase II
 YBR003w *COQ1* hexaprenyl pyrophosphate synthetase
 YBR153w *RIB7* HTP reductase
 YOR196c *LIP5* lipoidic acid synthase
 YHR042w *NCP1* NADPH-cytochrome P450 reductase
 YOR209c *NPT1* nicotinate phosphoribosyltransferase
 YNR033w *ABZ1* para-aminobenzoate synthase
 YNR041c *COQ2* para-hydroxybenzoate-polyprenyltransferase
 YOR184w *SER1* phosphoserine transaminase
 YDL205c *HEM3* porphobilinogen deaminase
 YGL040c *HEM2* porphobilinogen synthase
 YER014w *HEM14* protoporphyrinogen oxidase, mitochondrial
 YNR057c *BI04* putative dethiobiotin synthetase
 YBR035c *PDX3* pyridoxamine-phosphate oxidase
 YBR256c *RIB5* riboflavin synthase, α subunit
 YER043c *SAH1* S-adenosyl-L-homocysteine hydrolase
 YNL256w similarity to bacterial dithydropterote synthase
 YKL027w similarity to *E. coli* molybdopterin-converting factor chN
 YIL145c similarity to *E. coli* pantothenate synthetase
 YGR255c similarity to *E. coli* ubiH protein
 YMR113w similarity to folylpolyglutamate synthetases and strong similarity to hypothetical protein YKL132c
 YGL125w similarity to human methylenetetrahydrofolate reductase

YPL023c similarity to human methylenetetrahydrofolate reductase
 YFR047c similarity to human quinolinate phosphoribosyltransferase
 YHR111w similarity to molybdopterin biosynthesis proteins
 YOL151w similarity to plant dihydroflavonol-4-reductases
 YMR222c similarity to *S. pombe* dihydrofolate reductase
 YOR241w similarity to tetrahydrofolylpolyglutamate synthase
 YJR142w similarity to thiamin pyrophosphokinase
 YKR069w similarity to uroporphyrinogen methyltransferases
 YGL039w similarity to *V. vinifera* dihydroflavonol reductase
 YBR176w strong similarity to *E. coli* 3-methyl-2-oxobutanoate hydroxymethyltransferase
 YHR003c similarity to molybdopterin-converting factor homologue YKL027w
 YDL036c strong similarity to Rib2p
 YOL049w strong similarity to *S. pombe* Gsa1p
 YOR143c *THI80* thiamin pyrophosphokinase
 YPL214c *THI6* thiamin-phosphate pyrophosphorylase and hydroxyethylthiazole kinase
 YDR047w *HEM12* uroporphyrinogen decarboxylase
 YOR278w *HEM4* uroporphyrinogen III synthase

utilization of vitamins, cofactors and prosthetic groups

YDL141w *BPL1* biotin holocarboxylase synthetase
 YHR092c *PHO3* constitutive acid phosphatase
 YAL039c *CYC3* holo-cytochrome c synthase (cytochrome c haem lyase)
 YKL087c *CYT2* holo-cytochrome c1 synthase
 YML120c *NDI1* NADH-ubiquinone-6 oxidoreductase
 YBR093c *PHO5* repressible acid phosphatase
 YKL132c similarity to *B. subtilis* foIC protein and strong similarity to hypothetical protein YMR113w

regulation of vitamins, cofactors and prosthetic groups

YPR065w *ROX1* haem-dependent transcriptional repressor of hypoxic genes
 YOR125c *CAT5* involved in glucose repression

transport of vitamins, cofactors and prosthetic groups

YIL134w *FLX1* FAD carrier protein, mitochondrial (MCF)
 YIL006w similarity to Flx1p

other vitamin, cofactor and prosthetic-group activities

YDR541c similarity to dihydroflavonol-4-reductases
 YAL157w similarity to *E. gunnii* cinnamyl alcohol dehydrogenase
 YER183c similarity to formyltetrahydrofolate cyclo-ligase
 YHL018w similarity to human pterin-4-carbinolamine dehydratase
 YGR169c similarity to Rib2p
 YGR144w *THI4* thiamine-repressed protein

ionic homeostasis

homeostasis of metal ions

YNL259c *ATX1* antioxidant protein and metal homeostasis factor
 YJR049c *UTR1* associated with ferric reductase activity
 YMR058w *FET3* cell-surface ferroxidase
 YOR316c *COT1* cobalt accumulation protein
 YPL177c *CUP9* copper homeostasis protein
 YDR515w *SLF1* copper homeostasis protein
 YPR124w *CTR1* copper transport protein
 YHR175w *CTR2* copper transport protein
 YLR411w *CTR3* copper transport protein
 YGL166w *CUP2* copper-dependent transcription factor
 YLR214w *FRE1* ferric (and cupric) reductase
 YKL220c *FRE2* ferric (and cupric) reductase
 YOR176w *HEM15* ferrochelatase
 YGL255w *ZRT1* high-affinity zinc-transport protein
 YLL009c *COX17* interacts genetically with *SCO1* and *SCO2* in cytochrome oxidase assembly
 YBR037c *SCO1* involved in stabilization of Cox1p and Cox2p
 YGL071w *RCS1* iron-regulated transcriptional repressor
 YLR130c *ZRT2* low-affinity zinc transporter
 YMR319c *FET4* low-affinity Fe(II) iron transport protein
 YOL122c *SMF1* manganese transporter
 YBR290w *BSD2* metal homeostasis protein
 YKL064w *MNR2* overexpression overcomes manganese toxicity
 YDR270w *CCC2* probable copper-transporting ATPase
 YBR295w *PCA1* P-type Cu²⁺-transporting ATPase
 YGL096w similarity to copper homeostasis protein Cup9p
 YDR506c similarity to Fet3p
 YOL152w similarity to Fre1p and Fre2p
 YGL160w similarity to hypothetical protein YLR047c and Fre2p
 YJR126c similarity to transferrin receptor protein
 YFL041w strong similarity to cell-surface ferroxidase Fet3p
 YLL051c strong similarity to ferric reductase Fre2p
 YOR381w strong similarity to ferric reductase Fre2p
 YOR384w strong similarity to ferric reductase Fre2p
 YNR060w strong similarity to Fre2p and hypothetical protein YOR381w, and similarity to Fre1p
 YBR024w *SCO2* strong similarity to Sco1p

YOR031w *CRS5* suppressor of *cup1* deletion, metallothionein-like protein
 YCL037c *SRO9* suppressor of *rho3*
 YMR243c *ZRC1* zinc- and cadmium-resistance protein

homeostasis of other ions

YDL128w *VCX1* Ca²⁺-transport (H⁺/Ca²⁺ exchange) protein, vacuolar
 YGL006w *PMC1* Ca²⁺-transporting P-type ATPase
 YGL167c *PMR1* Ca²⁺-transporting P-type ATPase
 YGR217w *COH1* calcium channel protein
 YLR348c dicarboxylate carrier protein
 YKL016c *ATP7* F1F0-ATPase complex, F0 D subunit
 YBL099w *ATP1* F1F0-ATPase complex, F1 α subunit
 YJR121w *ATP2* F1F0-ATPase complex, F1 β subunit
 YDL004w *ATP16* F1F0-ATPase complex, F1 δ subunit
 YPL078c *ATP4* F1F0-ATPase complex, F1 ϵ subunit
 YPL271w *ATP15* F1F0-ATPase complex, F1 ζ subunit
 YBR039w *ATP3* F1F0-ATPase complex, F1 γ subunit
 YDR298c *ATP5* F1F0-ATPase complex, OSCP subunit
 YLR295c *ATP14* F1F0-ATPase complex, subunit h
 YCR024c-a *PMP1* H⁺-ATPase subunit, plasma membrane
 YEL017c-a *PMP2* H⁺-ATPase subunit, plasma membrane
 YMR054w *STV1* H⁺-ATPase V0 domain 102K subunit, vacuolar
 YHR039c-a *VMA10* H⁺-ATPase V0 domain 13K subunit, vacuolar
 YEL027w *CUP5* H⁺-ATPase V0 domain 17K subunit, vacuolar
 YPL234c *TFP3* H⁺-ATPase V0 domain 17K subunit, vacuolar
 YLR447c *VMA6* H⁺-ATPase V0 domain 36K subunit, vacuolar
 YOR270c *VPH1* H⁺-ATPase V0 domain 95K subunit, vacuolar
 YGR020c *VMA7* H⁺-ATPase V1 domain 14K subunit, vacuolar
 YOR332w *VMA4* H⁺-ATPase V1 domain 27K subunit, vacuolar
 YEL051w *VMA8* H⁺-ATPase V1 domain 32K subunit, vacuolar
 YKL080w *VMA5* H⁺-ATPase V1 domain 42K subunit, vacuolar
 YPR036w *VMA13* H⁺-ATPase V1 domain 54K subunit, vacuolar
 YBR127c *VMA2* H⁺-ATPase V1 domain 60K subunit, vacuolar
 YDL185w *TFP1* H⁺-ATPase V1 domain 69K subunit, vacuolar
 YGL008c *PMA1* H⁺-transporting P-type ATPase
 YPL036w *PMA2* H⁺-transporting P-type ATPase 2
 YML123c *PHO84* high affinity inorganic phosphate/H⁺ symporter
 YJL129c *TRK1* high-affinity potassium transport protein
 YBR294w *SUL1* high-affinity sulphate transport protein
 YJL117w *PHO86* inorganic phosphate transporter
 YLR220w *CCC1* involved in calcium regulation
 YCR037c *PHO87* member of the phosphate permease family
 YJL093c *TOK1* outward-rectifier potassium channel
 YJR077c *MIR1* phosphate transport protein, mitochondrial (MCF)
 YEL031w *SPF1* P-type ATPase
 YDR040c *ENA1* P-type ATPase involved in Na⁺ and Li⁺ efflux
 YDR039c *ENA2* P-type ATPase involved in Na⁺ efflux
 YDR038c *ENA5* P-type ATPase involved in Na⁺ efflux
 YLR138w *NHA1* putative Na⁺/H⁺ antiporter
 YBR235w similarity to bumetanide-sensitive Na⁺-K⁺-Cl⁻ cotransport protein
 YJL094c similarity to *E. hirae* Na⁺/H⁺-antiporter NapA
 YNR013c similarity to membrane protein Pho87p and hypothetical protein YIL198w
 YDR456w similarity to Na⁺/H⁺ antiporters
 YPR003c similarity to sulphate transporter proteins
 YER053c strong similarity to mitochondrial phosphate carrier protein
 YJL198w strong similarity to Pho87p
 YBR296c strong similarity to phosphate-repressible phosphate permease
 YLR092w *SEL2* strong similarity to Sul1p
 YHL016c *DUR3* urea transport protein
 YJR040w *GEF1* voltage-gated chloride channel protein

Energy

glycolysis

YGR240c *PFK1* 6-phosphofructokinase, α subunit
 YMR205c *PFK2* 6-phosphofructokinase, β subunit
 YIL107c *PFK26* 6-phosphofructose-2-kinase, isoenzyme 1
 YOL136c *PFK27* 6-phosphofructose-2-kinase, isoenzyme 2
 YNL071w *LAT1* dihydroloipoamide S-acetyltransferase
 YGR254w *ENO1* enolase I (2-phosphoglycerate dehydratase)
 YHR174w *ENO2* enolase II (2-phosphoglycerate dehydratase)
 YOR393w *ERR1* enolase-related protein
 YPL281c *ERR2* enolase-related protein
 YKL060c *FBA1* fructose-bisphosphate aldolase
 YBR196c *PGI1* glucose-6-phosphate isomerase
 YJL052w *TDH1* glyceraldehyde-3-phosphate dehydrogenase 1
 YJR009c *TDH2* glyceraldehyde-3-phosphate dehydrogenase 2

YGR192c *TDH3* glyceraldehyde-3-phosphate dehydrogenase 3
 YOL059w *GPD3* glycerol-3-phosphate dehydrogenase (NAD⁺), mitochondrial
 YFR053c *HXK1* hexokinase I
 YGL253w *HXK2* hexokinase II
 YCR012w *PGK1* phosphoglycerate kinase
 YKL152c *GPM1* phosphoglycerate mutase
 YOL056w *GPM3* phosphoglycerate mutase
 YER178w *PDA1* pyruvate dehydrogenase (lipoamide) α subunit
 YBR221c *PDB1* pyruvate dehydrogenase (lipoamide) β subunit
 YGR193c *PDX1* pyruvate dehydrogenase complex protein X
 YAL038w *CDC19* pyruvate kinase
 YLR345w similarity to 6-phosphofructo-2-kinase
 YDL021w *GPM2* strong similarity to phosphoglycerate mutase Gpm1p
 YMR323w strong similarity to phosphopyruvate hydratases
 YOR347c strong similarity to pyruvate kinase
 YKL029c strong similarity to *S. pombe* malate oxidoreductase
 YMR125w *GCR3* transcription factor for glycolytic genes
 YDR050c *TP11* triose-phosphate isomerase
 YKR043c weak similarity to phosphoglycerate mutase
 YOR283w weak similarity to phosphoglycerate mutases

gluconeogenesis

YOR393w *ERR1* enolase-related protein
 YPL281c *ERR2* enolase-related protein
 YLR377c *FBP1* fructose-1,6-bisphosphatase
 YBR196c *PGI1* glucose-6-phosphate isomerase
 YER065c *ICL1* isocitrate lyase
 YKR097w *PKC1* phosphoenolpyruvate carboxykinase
 YCR012w *PGK1* phosphoglycerate kinase
 YGL062w *PYC1* pyruvate carboxylase 1
 YBR218c *PYC2* pyruvate carboxylase 2
 YIL167w *SDL1* serine dehydratase
 YOR347c strong similarity to pyruvate kinase
 YMR280c *CAT8* transcription factor involved in gluconeogenesis
 YDR050c *TP11* triose-phosphate isomerase

pentose-phosphate pathway

YHR183w *GND1* 6-phosphogluconate dehydrogenase
 YOR095c *RK11* D-ribose-5-phosphate ketol-isomerase
 YJL121c *POS18* D-ribulose-5-phosphate 3-epimerase
 YNL241c *ZWF1* glucose-6-phosphate dehydrogenase
 YGR256w *GND2* phosphogluconate dehydrogenase
 YGR043c strong similarity to transaldolase
 YLR354c *TAL1* transaldolase
 YPR074c *TKL1* transketolase 1
 YBR117c *TKL2* transketolase 2

tricarboxylic-acid pathway

YIL125w *KGD1* 2-oxoglutarate dehydrogenase complex E1 component
 YDR148c *KGD2* 2-oxoglutarate dehydrogenase complex E2 component
 YLR304c *ACO1* aconitate hydratase
 YNR001c *CIT1* citrate (si)-synthase, mitochondrial
 YPR001w *CIT3* citrate (si)-synthase, mitochondrial
 YFL018c *LPD1* dihydroloipoamide dehydrogenase
 YPL262w *FUM1* fumarate hydratase
 YNL009w homology to isocitrate dehydrogenase
 YNL037c *IDH1* isocitrate dehydrogenase (NAD⁺) subunit 1, mitochondrial
 YOR136w *IDH2* isocitrate dehydrogenase (NAD⁺) subunit 2, mitochondrial
 YLR174w *IDP2* isocitrate dehydrogenase, cytosolic
 YOL126c *MDH2* malate dehydrogenase, cytoplasmic
 YOR297c similarity to SDH4 protein
 YJL200c strong similarity to aconitate hydratase
 YGR244c strong similarity to rumen fungus β -succinyl-CoA synthetase
 YLR164w strong similarity to Sdh4p
 YMR118c strong similarity to succinate dehydrogenase
 YJL045w strong similarity to succinate dehydrogenase flavoprotein
 YOR142w strong similarity to succinate-CoA ligase α subunit
 YKL148c *SDH1* succinate dehydrogenase flavoprotein
 YLL041c *SDH2* succinate dehydrogenase iron-sulphur protein subunit
 YDR178w *SDH4* succinate dehydrogenase membrane anchor subunit for Sdh2p

respiration

YMR282c *AEP2* 2'-O-ribosyl phosphate transferase
 YGR008c *STF2* ATPase stabilizing factor
 YDL130w-a *STF1* ATPase stabilizing factor, 10K
 YDR377w *ATP17* ATPase synthase subunit f
 YER061c *CEM1* β -keto-acyl-ACP synthase
 YIL043c *CBR1* cytochrome b5 reductase
 YKL150w *MCR1* cytochrome b5 reductase
 YKL141w *SDH3* cytochrome b560 subunit of respiratory complex II
 YJR048w *CYC1* cytochrome c isoform 1
 YEL039c *CYC2* cytochrome c isoform 2

YML129c *COX14* cytochrome c oxidase assembly protein
 YDR079w *PET100* cytochrome c oxidase assembly protein
 YGL187c *COX4* cytochrome c oxidase subunit IV
 YNL052w *COX5A* cytochrome c oxidase subunit VA
 YIL111w *COX5B* cytochrome c oxidase subunit Vb
 YHR051w *COX6* cytochrome c oxidase subunit VI
 YGL191w *COX13* cytochrome c oxidase subunit VIa
 YLR038c *COX12* cytochrome c oxidase subunit VIb
 YMR256c *COX7* cytochrome c oxidase subunit VII
 YDL067c *COX9* cytochrome c oxidase subunit VIIA
 YLR395c *COX8* cytochrome c oxidase subunit VIII
 YOR065w *CYT1* cytochrome c1
 YER154w *OXA1* cytochrome oxidase biogenesis protein
 YGR207c *ETF-β* electron-transferring flavoprotein, β subunit
 YKL016c *ATP7* F1F0-ATPase complex, F0 D subunit
 YBL099w *ATP1* F1F0-ATPase complex, F1 α subunit
 YJR121w *ATP2* F1F0-ATPase complex, F1 β subunit
 YDL004w *ATP16* F1F0-ATPase complex, F1 δ subunit
 YLR078c *ATP4* F1F0-ATPase complex, F1 ε subunit
 YPL271w *ATP15* F1F0-ATPase complex, F1 ε subunit
 YBR039w *ATP3* F1F0-ATPase complex, F1 γ subunit
 YDR298c *ATP5* F1F0-ATPase complex, OSCP subunit
 YLR295c *ATP14* F1F0-ATPase complex, subunit h
 YEL053c *MAK10* glucose-repressible protein
 YLL009c *COX17* interacts genetically with *SCO1* and *SCO2* in cytochrome oxidase assembly
 YJR034w *PET191* involved in assembly of cytochrome oxidase
 YMR165c *SMP2* involved in plasmid maintenance, respiration and cell proliferation
 YML054c *CYB2* lactate dehydrogenase cytochrome b2
 YBR192w *RIM2* mitochondrial carrier protein (MCF)
 YMR089c *YTA12* protease of the *SEC18/ CDC48/ PAS1* family of ATPases (AAA)
 YLR044c *PDC1* pyruvate decarboxylase, isozyme 1
 YGR062c *COX18* required for activity of mitochondrial cytochrome oxidase
 YBR185c *MBA1* respiratory chain assembly protein
 YDL107w *MSS2* ser/thr protein kinase
 YPR048w similarity to *M. domestica* NADPH-ferritinoprotein reductase and mammalian nitric-oxide synthases
 YGL226w similarity to *N. crassa* cytochrome c oxidase subunit V
 YMR244w similarity to NCA3 and SUN4 protein
 YML087c strong similarity to cytochrome b5- and nitrate reductases
 YML125c strong similarity to cytochrome b5- and nitrate reductases
 YPR004c strong similarity to electron transfer flavoproteins α subunit
 YOR356w strong similarity to human electron transfer flavoprotein-ubiquinone oxidoreductase
 YDL085w strong similarity to NADH dehydrogenase (ubiquinone)
 YMR145c strong similarity to NADH dehydrogenase (ubiquinone)
 YDL080c strong similarity to pyruvate decarboxylases
 YDR178w *SDH4* succinate dehydrogenase membrane anchor subunit for Sdh2p
 YNL118c *PSU1* suppressor protein of a yeast *pet* mutant
 YFR033c *QCR6* ubiquinol-cytochrome c reductase 17K protein
 YPR191w *QCR2* ubiquinol-cytochrome c reductase 40K subunit II
 YBL045c *COR1* ubiquinol-cytochrome c reductase 44K core protein
 YHR001w-a *QCR10* ubiquinol-cytochrome c reductase 8.5K subunit
 YGR174c *CBP4* ubiquinol-cytochrome c reductase assembly factor
 YGL119w *ABC1* ubiquinol-cytochrome c reductase complex assembly protein
 YEL024w *RIP1* ubiquinol-cytochrome c reductase iron-sulphur protein
 YDR529c *QCR7* ubiquinol-cytochrome c reductase subunit 7
 YGR183c *QCR9* ubiquinol-cytochrome c reductase subunit 9
 YJL166w *QCR8* ubiquinol-cytochrome c reductase subunit VIII
 YMR073c weak similarity to C-terminal part of cytochrome b5 and b2
 YNL237w *YTP1* weak similarity to mitochondrial electron transport proteins

fermentation

YOL086c *ADH1* alcohol dehydrogenase I
 YMR303c *ADH2* alcohol dehydrogenase II
 YMR083w *ADH3* alcohol dehydrogenase III
 YGL256w *ADH4* alcohol dehydrogenase IV
 YBR145w *ADH5* alcohol dehydrogenase V
 YER073w aldehyde dehydrogenase (NAD⁺)
 YPL061w *ALD6* aldehyde dehydrogenase, cytosolic
 YDL174c *DLD1* D-lactate ferriocytocrome c oxidoreductase (D-LCR)
 YML004c *GLO1* glyoxalase I
 YOR126c *EST2* isoamyl acetate hydrolytic enzyme
 YPL275w putative formate dehydrogenase/putative pseudogene
 YPL276w putative formate dehydrogenase/putative pseudogene
 YDR081c *PDC2* pyruvate decarboxylase regulatory protein
 YLR044c *PDC1* pyruvate decarboxylase, isozyme 1

YLR134w *PDC5* pyruvate decarboxylase, isozyme 2
 YGR087c *PDC6* pyruvate decarboxylase, isozyme3
 YAL060w *FUN49* similarity to alcohol/sorbitol dehydrogenase
 YAL061w *FUN50* similarity to alcohol/sorbitol dehydrogenase
 YMR318c similarity to alcohol-dehydrogenase
 YHR039c similarity to aldehyde dehydrogenases
 YPL088w similarity to aryl-alcohol dehydrogenases
 YMR285c similarity to Ccr4p
 YHL008c similarity to *M. formicicum* formate dehydrogenase
 YKR096w similarity to mitochondrial aldehyde dehydrogenase Ald1p
 YDR380w similarity to pyruvate decarboxylases
 YNL331c strong similarity aryl-alcohol reductase
 YCR107w strong similarity aryl-alcohol reductases
 YCR105w strong similarity to alcohol dehydrogenases
 YMR169c *ALD4* strong similarity to aldehyde dehydrogenase
 YDL243c strong similarity to aryl-alcohol dehydrogenase
 YJR155w strong similarity to aryl-alcohol dehydrogenase
 YFL056c strong similarity to aryl-alcohol dehydrogenases
 YFL057c strong similarity to aryl-alcohol dehydrogenases
 YOL165c strong similarity to aryl-alcohol dehydrogenases
 YOR388c strong similarity to *H. polymorpha* formate dehydrogenase

metabolism of energy reserves (glycogen and trehalose)

YEL011w *GLC3* 1,4-glucan branching enzyme (glycogen branching enzyme)
 YBR001c *NTH2* α,α-trehalase
 YDR074w *TPS2* α,α-trehalose-phosphate synthase, 105K subunit
 YMR261c *TPS3* α,α-trehalose-phosphate synthase, 115K subunit
 YML100w *TSL1* α,α-trehalose-phosphate synthase, 123K subunit
 YBR126c *TPS1* α,α-trehalose-phosphate synthase, 56K subunit
 YPR026w *ATH1* acid trehalase, vacuolar
 YBR299w *MAL32* α-glucosidase
 YHR047c *AAP1* alanine/arginine aminopeptidase
 YPL031c *PHO85* cyclin-dependent protein kinase
 YLR071c *RGR1* DNA-directed RNA polymerase II holoenzyme subunit
 YER054c *GIP2* Glc7p-interacting protein
 YBR229c *ROT2* glucosidase II, catalytic subunit
 YPR160w *GPH1* glycogen phosphorylase
 YPL240c *HSP82* heat-shock protein
 YKL128c *PMU1* high copy suppressor of ts *tps2* mutant phenotype
 YDR001c *NTH1* neutral trehalase (α,α-trehalase)
 YMR105c *PGM2* phosphoglucumutase, major isoform
 YKL127w *PGM1* phosphoglucumutase, minor isoform
 YNR032w *PPG1* phosphoprotein phosphatase PPG catalytic subunit
 YBL058w *SHP1* potential regulatory subunit for Glc7p
 YLR273c *PIG1* protein interacting with Gsy2p
 YIL045w *PIG2* protein interacting with Gsy2p
 YJL137c *GLG2* self-glycosylating initiator of glycogen synthesis
 YKR058w *GLG1* self-glycosylating initiator of glycogen synthesis
 YER133w *GLC7* ser/thr phosphoprotein phosphatase 1, catalytic subunit
 YOR178c *GAC1* ser/thr phosphoprotein phosphatase 1, regulatory subunit
 YPR184w similarity to glycogen debranching enzymes
 YJL221c *FSP2* strong similarity to α-D-glucosidase
 YOL157c strong similarity to α-glucosidases
 YIL172c strong similarity to Fsp2p
 YJL216c strong similarity to Mal62p
 YFR015c *GSY1* UDP glucose-starch glucosyltransferase, isoform 1
 YLR258w *GSY2* UDP-glucose-starch glucosyltransferase, isoform 2
 YKL035w *UGP1* UTP-glucose-1-phosphate uridylyltransferase

other energy-generation activities

YIL160c *POT1* acetyl-CoA C-acyltransferase, peroxisomal
 YGL205w *POX1* acyl-CoA oxidase
 YMR170c *ALD5* aldehyde dehydrogenase 2 (NAD⁺), mitochondrial
 YML035c *AMD1* AMP deaminase
 YCR005c *CIT2* citrate (si)-synthase, peroxisomal
 YKR009c *FOX2* hydratase-dehydrogenase-epimerase, peroxisomal
 YMR267w *PPA2* inorganic pyrophosphatase, mitochondrial
 YDL066w *IDP1* isocitrate dehydrogenase (NADP⁺), mitochondrial
 YER065c *ICL1* isocitrate lyase
 YKL085w *MDH1* malate dehydrogenase, mitochondrial
 YDL078c *MDH3* malate dehydrogenase, peroxisomal
 YNL117w *MLS1* malate synthase 1
 YIR031c *DAL7* malate synthase 2

YHR179w *OYE2* NADPH dehydrogenase (old yellow enzyme), isoform 1
 YPL171c *OYE3* NADPH dehydrogenase (old yellow enzyme), isoform 3
 YPR006c *ICL2* non-functional isocitrate lyase
 YOL079w similarity to NADH dehydrogenase
 YEL020c similarity to *O. formigenes* oxalyl-CoA decarboxylase
 YDR384c strong similarity to *Y. lipolytica* *GPR1* gene

Cell growth, cell division and DNA synthesis

cell growth

YPL268w *PLC1* 1-phosphatidylinositol-4,5-bisphosphate phosphodiesterase
 YCR088w *ABP1* actin-binding protein
 YDL029w *ACT2* actin-like protein
 YOL052c *SPE2* adenosylmethionine decarboxylase
 YNL138w *SRV2* adenylate cyclase-associated protein, 70K
 YER170w *ADK2* adenylate kinase, mitochondrial
 YBR109c *CMD1* calmodulin
 YHR135c *YCK1* casein kinase I isoform
 YNL154c *YCK2* casein kinase I isoform
 YJL174w *KRE9* cell-wall synthesis protein
 YNL327w *EGT2* cell-cycle regulation protein
 YLR175w *CBF5* centromere/microtubule binding protein
 YGR167w *CLC1* clathrin light chain
 YDR251w *PAM1* coiled-coil protein multiplicity suppressor of loss of PP2A
 YBL007c *SLA1* cytoskeleton assembly control protein
 YFL001w *DEG1* depressed growth-rate protein
 YLR300w *EXG1* exo-1,3-glucanase (I/II), major isoform
 YDR261c *EXG2* exo-1,3-glucanase minor isoform
 YPR159w *KRE6* glucan synthase subunit
 YBR212w *NGR1* glucose-repressible RNA-binding protein
 YOR043w *WHI2* growth regulation protein
 YKL021c *MAK11* involved in cell growth and replication of M1 dsRNA virus
 YMR165c *SMP2* involved in plasmid maintenance, respiration and cell proliferation
 YNL197c *WHI3* involved in regulation of cell size
 YDR293c *SSD1* involved in the tolerance to high concentration of Ca²⁺
 YOR336w *KRE5* killer toxin-resistance protein
 YDR480w *DIG2* MAP kinase-associated protein
 YOR032w *CPB7* member of the cytopliphin family
 YGR029w *ERV1* mitochondrial biogenesis and cell viability protein
 YBR089c-a *NHP6B* nonhistone chromosomal protein
 YPR052c *NHP6A* nonhistone chromosomal protein related to mammalian HMG1
 YKL203c *TOR2* phosphatidylinositol 3-kinase
 YCR034w *GNS1* probable 1,3-glucan synthase subunit
 YOR149c *SMP3* protein kinase C pathway protein
 YPL140c *MKK2* protein kinase of the MAP kinase kinase (MEK) family
 YDR151c *CTH1* protein of the inducible CCCH zinc-finger family
 YDR075w *PPH3* protein ser/thr phosphatase
 YDL006w *PTC1* protein ser/thr phosphatase 2c
 YDL134c *PPH21* protein ser/thr phosphatase PP2A-1
 YDL188c *PPH22* protein ser/thr phosphatase PP2A-2
 YDR137w *RGP1* reduced growth phenotype protein
 YJR090c *GRR1* required for glucose repression and for glucose and cation transport
 YGL190c *CDC55* ser/thr phosphatase 2A regulatory subunit B
 YAL016w *TPD3* ser/thr phosphatase 2A, regulatory subunit A
 YOR231w *MKK1* ser/thr protein kinase
 YPR161c *SGV1* ser/thr protein kinase
 YHR030c *SLT2* ser/thr protein kinase of MAP kinase family
 YJL095w *BCK1* ser/thr protein kinase of the MEKK family
 YER167w *BCK2* ser/thr protein kinase of the protein kinase C pathway
 YNL180c similarity to *S. pombe* Cdc42p and other GTP-binding proteins
 YOR027w *STI1* stress-induced protein
 YGL106w strong similarity to calmodulins
 YER177w *BMH1* strong similarity to mammalian 14-3-3 proteins
 YPL032c strong similarity to Pam1p
 YKL161c strong similarity to ser/thr-specific protein kinase Sit2p
 YDL224c strong similarity to WHI3 protein
 YDR099w *BMH2* suppressor of clathrin deficiency
 YJR075w *HOC1* suppressor of *pkc1*
 YOR075w *UFE1* syntaxin (T-SNARE) of the ER
 YHR206w *SKN7* transcription factor with similarity to Hsf1p
 YNL079c *TPM1* tropomyosin 1
 YLR337w *VRP1* verplogin
 YLR403w *SFP1* zinc-finger protein

budding, cell polarity and filament formation

YFL039c *ACT1* actin
 YDR129c *SAC6* actin filament bundling protein, fimbrin
 YCR088w *ABP1* actin-binding protein
 YDL029w *ACT2* actin-like protein
 YNL138w *SRV2* adenylate cyclase-associated protein, 70K
 YOR304c-a *BAT2* AIP3 binding protein

YBL085w *BOI1* BEM1 protein-binding protein
 YBR200w *BEM1* bud emergence mediator
 YPL161c *BEM4* bud emergence protein
 YDR351w *SBE2* bud growth protein
 YLR319c *BUD6* bud site selection protein
 YNL271c *BN11* budding protein
 YER114c *BOI2* budding protein
 YJR092w *BUD4* budding protein
 YLR353w *BUD8* budding protein
 YGR041w *BUD9* budding protein
 YBR109c *CMD1* calmodulin
 YHR135c *YCK1* casein kinase I isoform
 YNL154c *YCK2* casein kinase I isoform
 YPL255w *BBP1* cell division control protein
 YDR182w *CDC1* cell division control protein
 YLR314c *CDC3* cell division control protein
 YFL009w *CDC4* cell division control protein
 YJL174w *KRE9* cell-wall synthesis protein
 YBR038w *CHS2* chitin synthase II
 YBR023c *CHS3* chitin synthase III
 YLR330w *CHS5* chitin synthesis protein
 YLL050c *COF1* cofillin, actin binding and severing protein
 YDR251w *PAM1* coiled-coil protein multicopy suppressor of loss of PP2A
 YPL256c *CLN2* cyclin, G1/S specific
 YBR160w *CDC28* cyclin-dependent protein kinase
 YNL243w *SLA2* cytoskeleton assembly control protein
 YLR071c *RGR1* DNA-directed RNA polymerase II holoenzyme subunit
 YKL007w *CAP1* F-actin capping protein, α subunit
 YLJ034c *CAP2* F-actin capping protein, β subunit
 YJL157c *FAR1* factor arrest protein
 YAL041w *CDC24* GDP/GTP exchange factor for Cdc42p
 YCR038c *BUD5* GDP/GTP exchange factor for Rsr1p/Bud1p
 YGR070w *ROM1* GDP/GTP exchange protein for Rho1p
 YLR371w *ROM2* GDP/GTP exchange protein for Rho1p
 YGL155w *CDC43* geranylgeranyltransferase type I β subunit
 YER155c *BEM2* GTPase-activating protein
 YLR092c *BUD2* GTPase-activating protein for Bud1p/Rsr1p
 YPL115c *BEM3* GTPase-activating protein for Cdc42p and Rho1p
 YGR152c *RSR1* GTP-binding protein
 YLR229c *CDC42* GTP-binding protein of RAS superfamily
 YLJ118w *RHO3* GTP-binding protein of the RHO family
 YKR055w *RHO4* GTP-binding protein of the RHO family
 YPR165w *RHO1* GTP-binding protein of the RHO subfamily of RAS-like proteins
 YNL090w *RHO2* GTP-binding protein of the RHO subfamily of RAS-like proteins
 YOR156c *NF1* interacts with Cdc12p in 2-hybrid assay
 YBR063c *LAS1* involved in cell morphogenesis, cytoskeletal regulation and bud formation
 YLL021w *SPA2* involved in cell polarity
 YDR085c *AFR1* involved in morphogenesis of the mating projection
 YMR273c *ZDS1* involved in negative regulation of cell polarity
 YER149c *PEA2* involved in oriented growth towards mating partner
 YDR293c *SSD1* involved in the tolerance to high concentration of Ca^{2+}
 YKL079w *SMY1* kinesin-related protein
 YOR198c *BFR1* maintenance of normal ploidy
 YJL158c member of the Pir1p/Hsp150p/Pir3p family
 YJL159w member of the Pir1p/Hsp150p/Pir3p family
 YJL160c member of the Pir1p/Hsp150p/Pir3p family
 YOR188w *MSB1* morphogenesis-related protein
 YGR014w *MSB2* multicopy suppressor of a *cdc24* bud emergence defect
 YHR023w *MYO1* myosin I isoform (type II myosin) heavy chain
 YOR326w *MYO2* myosin heavy chain
 YAL029c *MYO4* myosin heavy chain, unconventional (class V) isoform
 YMR109w *MYO5* myosin I
 YKL129c *MYO3* myosin type I
 YBR089c-a *NHP6B* nonhistone chromosomal protein
 YPR052c *NHP6A* nonhistone chromosomal protein related to mammalian HMG1
 YCR057c *PWP2* periodic tryptophan protein
 YBL058w *SHP1* potential regulatory subunit for Gic7p
 YCR034w *GNS1* probable 1,3-glucan synthase subunit
 YER108c *FLO8* probable transcriptional activator of Flo1p
 YOR122c *PFY1* profilin
 YPR122w *AXL1* protease
 YOR149c *SMF3* protein kinase C pathway protein
 YPL140c *MKK2* protein kinase of the MAP kinase kinase (MEK) family
 YDL047w *SIT4* protein ser/thr phosphatase
 YDL006w *PTC1* protein ser/thr phosphatase 2c
 YDL134c *PPH21* protein ser/thr phosphatase PP2A-1
 YDL188c *PPH22* protein ser/thr phosphatase PP2A-2
 YDR388w *RVS167* reduced viability upon starvation protein
 YMR016c *SOK2* regulatory protein in the PKA signal transduction pathway
 YCL014w *BUD3* required for axial budding
 YJL140w *SRO4* required for axial pattern of budding
 YER109c *FLO8* required for diploid filamentous growth
 YNL084c *END3* required for endocytosis and cytoskeletal organization
 YJR090c *GRR1* required for glucose repression and for glucose and cation transport

YDL135c *RDI1* RHO GDP dissociation inhibitor with activity toward Rho1p
 YOR127w *RGA1* RHO-type GTPase-activating protein for Cdc42p
 YKL181w *PRPS1* ribose-phosphate pyrophosphokinase
 YJR076c *CDC11* septin
 YHR107c *CDC12* septin
 YGL190c *CDC55* ser/thr phosphatase 2A regulatory subunit B
 YAL016w *TPD3* ser/thr phosphatase 2A, regulatory subunit A
 YER133w *GLC7* ser/thr phosphoprotein phosphatase 1, catalytic subunit
 YOR178c *GAC1* ser/thr phosphoprotein phosphatase 1, regulatory subunit
 YKL048c *ELM1* ser/thr protein kinase
 YOR231w *MKK1* ser/thr protein kinase
 YBL105c *PKC1* ser/thr protein kinase
 YHR030c *SLT2* ser/thr protein kinase of MAP kinase family
 YJL095w *BCK1* ser/thr protein kinase of the MEKK family
 YLR362w *STE11* ser/thr protein kinase of the MEKK family
 YER167w *BCK2* ser/thr protein kinase of the protein kinase C pathway
 YDL159w *STE7* ser/thr/tyr protein kinase of MAP kinase kinase family
 YDL225w similarity to Cdc11p, Cdc3p and human CDC10 protein
 YGL054c similarity to *D. melanogaster* cni protein
 YCR009c *RVS161* similarity to human amphiphysin and Rvs167p
 YDR208w *MSS4* similarity to human PI 5-kinase
 YDR409w similarity to Nfl1p
 YDL125c *YHI1* similarity to protein kinase C inhibitor-I
 YDR245w *MNN10* similarity to *S. pombe* galactosyltransferase
 YFR040w *SAP155* Sit4p-associated protein
 YKR028w *SAP190* Sit4p-associated protein
 YJL098w *SAP185* Sit4p-associating protein
 YLR372w *SUR4* sterol isomerase
 YHR103w *SBE22* strong similarity to budding protein Sbe2p
 YGL106w strong similarity to calmodulins
 YBR210w strong similarity to *D. melanogaster* cornichon protein
 YPL032c strong similarity to Pam1p
 YNL320w strong similarity to *S. pombe* Bem46p
 YKL161c strong similarity to ser/thr-specific protein kinase Sltp2p
 YBR161w strong similarity to Sur1p
 YJR075w *HOC1* suppressor of *pkc1*
 YDR297w *SUR2* suppressor of *rvs161* and *rvs167* mutations
 YPL057c *SUR1* suppressor of *rvs161*, *rvs167*, and *cls2* mutations
 YKL043w *PHD1* transcription factor
 YGL181w *GTS1* transcription factor of the Gcs1p/Glc3p/Sps18p family
 YHR084w *STE12* transcriptional activator
 YNL079c *TPM1* tropomyosin 1
 YJL138c *TPM2* tropomyosin 2
 YBR083w *TEC1* Ty transcription activator
 YGL095c *VPS45* vacuolar protein sorting-associated protein
 YLR337w *VRP1* verprolin

pheromone response and mating-type determination

YJR004c *SAG1* α -agglutinin
 YNR044w *AGA1* α -agglutinin anchor subunit
 YGL032c *AGA2* α -agglutinin binding subunit
 YDR264c *AKR1* ankyrin repeat-containing protein
 YKL209c *STE6* ATP-binding cassette transporter protein
 YJL015w *BAR1* barrier pepsin
 YBR200w *BEM1* bud emergence mediator
 YPL161c *BEM4* bud emergence protein
 YNL057w *CMF2* calcineurin B, catalytic subunit
 YLR433c *CNA1* calcineurin B, catalytic subunit
 YKL190w *CNB1* calcineurin B, regulatory subunit
 YBR109c *CMD1* calmodulin
 YCR002c *CDC10* cell division control protein
 YLR314c *CDC3* cell division control protein
 YNL188w *KAR1* cell fusion protein
 YCL027w *FUS1* cell-wall synthesis protein
 YJL174w *KRE9* cell-wall synthesis protein
 YNL192w *CHS1* chitin synthase I
 YBR023c *CHS3* chitin synthase III
 YLR330w *CHS5* chitin synthesis protein
 YDR073w *SNF11* component of SWI/SNF transcription activator complex
 YOR290c *SNF2* component of SWI/SNF transcription activator complex
 YBR289w *SNF5* component of SWI/SNF transcription activator complex
 YPL016w *SWI1* component of SWI/SNF transcription activator complex
 YPL256c *CLN2* cyclin, G1/S specific
 YNL243w *SLA2* cytoskeleton assembly control protein
 YER095w *RAD51* DNA repair protein
 YDR076w *RAD55* DNA repair protein
 YDR004w *RAD57* DNA repair protein
 YAR007c *RFA1* DNA replication factor A, 69K subunit
 YOL051w *GAL11* DNA-directed RNA polymerase II holoenzyme and Kornberg's mediator (SRB) subcomplex subunit
 YLR071c *RGR1* DNA-directed RNA polymerase II holoenzyme subunit

YBR253w *SRB6* DNA-directed RNA polymerase II suppressor protein
 YNL053w *MSG5* dual-specificity protein phosphatase
 YNL238w *KEK2* endopeptidase of late Golgi compartment
 YLR300w *EXG1* exo-1,3-glucanase (I/II), major isoform
 YDR261c *EXG2* exo-1,3-glucanase minor isoform
 YKL007w *CAP1* F-actin capping protein, α subunit
 YJL034c *CAP2* F-actin capping protein, β subunit
 YJL157c *FAR1* factor arrest protein
 YAL041w *CDC24* GDP/GTP exchange factor for Cdc42p
 YGL097w *SRM1* GDP/GTP exchange factor for Gsp1p/Gsp2p
 YNL236w *SIN4* global regulator protein
 YHL025w *SNF6* global transcription activator
 YKL104c *GFA1* glucosamine-fructose-6-phosphate transaminase
 YPL115c *BEM3* GTPase-activating protein for Cdc42p and Rho1p
 YHR005c *GPA1* GTP-binding protein α subunit of the pheromone pathway
 YOR212w *STE4* GTP-binding protein β subunit of the pheromone pathway
 YJR086w *STE18* GTP-binding protein γ subunit of the pheromone pathway
 YLR229c *CDC42* GTP-binding protein of RAS superfamily
 YNL173c *MDG1* GTP-binding protein of the pheromone pathway
 YER020w *GPA2* guanine nucleotide-binding regulatory protein
 YDL227c *HO* homothallic switching endonuclease
 YNL291c *MID1* involved in Ca^{2+} influx during mating
 YLL021w *SPA2* involved in cell polarity
 YLR452c *SST2* involved in desensitization to α -factor pheromone
 YDR085c *AFR1* involved in morphogenesis of the mating projection
 YER149c *PEA2* involved in oriented growth toward mating partner
 YMR052w *FAR3* involved in pheromone-mediated cell cycle arrest
 YMR127c *SAS2* involved in silencing at HMR
 YPR141c *KAR3* kinesin-related protein
 YPL187w *MFA1* mating pheromone α -1 factor
 YGL089c *MFA2* mating pheromone α -2 factor
 YDR461w *MFA1* mating pheromone α -factor 1
 YNL145w *MFA2* mating pheromone α -factor 2
 YLR332w *MID2* mating process protein
 YHR066w *SSF1* mating protein
 YDR143c *SAN1* mating type regulatory protein
 YCR040w *MAT α 1* mating type regulatory protein (expressed copy at MAT locus)
 YCR039c *MAT α 2* mating type regulatory protein (expressed copy at MAT locus)
 YCL066w $\alpha 1$ mating type regulatory protein (silenced copy at HML locus)
 YCL067c $\alpha 2$ mating type regulatory protein (silenced copy at HML locus)
 YCR097w *A1* mating type regulatory protein (silenced copy at HMR locus)
 YCR096c *A2* mating type regulatory protein (silenced copy at HMR locus)
 YIL047c *SYG1* member of the major facilitator superfamily
 YBL016w *FUS3* mitogen-activated protein kinase (MAP kinase)
 YGL151w *NUT1* negative regulator of HO endonuclease
 YPR168w *NUT2* negative regulator of HO endonuclease
 YKL185w *ASH1* negative regulator of HO expression
 YCL029c *BIK1* nuclear fusion protein
 YML065w *ORC1* origin recognition complex, 104K subunit
 YNL261w *ORC5* origin recognition complex, 50K subunit
 YHR118c *ORC6* origin recognition complex, 50K subunit
 YPR162c *ORC4* origin recognition complex, 56K subunit
 YLL004w *ORC3* origin recognition complex, 62K subunit
 YBR060c *RRR1* origin recognition complex, 72K subunit
 YFL026w *STE2* pheromone α -factor receptor
 YKL178c *STE3* pheromone α -factor receptor
 YCL032w *STE50* pheromone response pathway protein
 YDR103w *STE5* pheromone signal transduction pathway protein
 YCR034w *GNS1* probable 1,3-glucan synthase subunit
 YLR389c *STE23* protease involved in α -factor processing
 YML032c *RAD52* recombination and DNA repair protein
 YDR392w *SPT3* regulatory protein
 YCL055w *KAR4* regulatory protein required for pheromone induction of karyogamy genes
 YMR232w *FUS2* required for cell fusion during mating
 YBR040w *FIG1* required for efficient mating
 YCR089w *FIG2* required for efficient mating
 YNL084c *END3* required for endocytosis and cytoskeletal organization
 YKL130c *SHE2* required for mother cell-specific expression of HO
 YOR035c *SHE4* required for mother cell-specific gene expression
 YOR127w *RGA1* RHO-type GTPase-activating protein for Cdc42p
 YLR441c *RP10A* ribosomal protein S3a.e
 YJR076c *CDC11* septin
 YHR107c *CDC12* septin
 YPR161c *SGV1* ser/thr protein kinase
 YGR040w *KSS1* ser/thr protein kinase of the MAP kinase family
 YLR362w *STE11* ser/thr protein kinase of the MEKK family
 YHL007c *STE20* ser/thr protein kinase of the pheromone pathway

| | | | | | | | | | | |
|------------------------------------|--------|--|----------------|---|--|-----------|---|---|------|-----------------------|
| YDL159w | STE7 | ser/thr/tyr protein kinase of MAP kinase kinase family | YBR078w | strong similarity to sporulation-specific Sps2p | YJR094c | IME1 | transcription factor required for sporulation | | | |
| YOL068c | HST1 | silencing protein | YCL048w | strong similarity to sporulation-specific Sps2p | YGR044c | RME1 | zinc-finger transcription factor | | | |
| YBL052c | SAS3 | silencing protein | YDR055w | strong similarity to Sps2p | DNA synthesis and replication | | | | | |
| YKR101w | SIR1 | silencing regulatory protein | YOR075w | syntaxin (T-SNARE) of the ER | YKL112w | ABF1 | ARS-binding factor | | | |
| YDL042c | SIR2 | silencing regulatory protein | YMR043w | transcription factor of the MADS box family | YFL009w | CDC4 | cell division control protein | | | |
| YLR442c | SIR3 | silencing regulatory protein | YJR094c | transcription factor required for sporulation | YDR364c | CDC40 | cell division control protein | | | |
| YDR227w | SIR4 | silencing regulatory protein | YNL330c | transcription modifier protein | YLR274w | CDC46 | cell division control protein | | | |
| YNL283c | | similarity to a-agglutinin core protein Aga1p | YOL004w | transcription regulatory protein | YBR202w | CDC47 | cell division control protein | | | |
| YDL225w | | similarity to Cdc11p, Cdc3p and human CDC10 protein | YNL012w | transcriptional regulator involved in sporulation | YJL194w | CDC6 | cell division control protein | | | |
| YDR191w | HST4 | similarity to Hst3p, Hst1p and Sir2p | YLL039c | ubiquitin | YML102w | CAC2 | chromatin assembly complex, subunit | | | |
| YGR023w | | similarity to Mid2p | YGL058w | ubiquitin-conjugating enzyme | YBR195c | MSI1 | chromatin assembly complex, subunit p50 | | | |
| YHR146w | | similarity to pheromone-response G-protein YNL173c | YDR177w | ubiquitin-conjugating enzyme | YPR018w | RLF2 | chromatin assembly complex, subunit p90 | | | |
| YKL117w | | Ste5p-associated protein | YER125w | ubiquitin-protein ligase | YPR120c | CLB5 | cyclin, B-type | | | |
| YGL106w | | strong similarity to calmodulins | YML115c | vanadate resistance protein | YGR109c | CLB6 | cyclin, B-type | | | |
| YJL212c | | strong similarity to <i>S. pombe</i> isp4 protein | meiosis | | | | | YPL256c | CLN2 | cyclin, G1/S specific |
| YDR312w | SSF2 | strong similarity to SSF1 protein | YPR103w | PRE2 | 26S proteasome subunit | YPR160w | CDC28 | cyclin-dependent protein kinase | | |
| YPL129w | ANC1 | TFIIIF subunit (transcription initiation factor), 30K | YGR253c | PUP2 | 26S proteasome subunit | YER176w | | DNA dependent ATPase/DNA helicase B | | |
| YDR146c | SWI5 | transcription factor | YKL112w | ABF1 | ARS-binding factor | YHR164c | DNA2 | DNA helicase | | |
| YMR043w | MCM1 | transcription factor of the MADS box family | YFL037w | TUB2 | β -tubulin | YDL164c | CDC9 | DNA ligase | | |
| YNL330c | RPD3 | transcription modifier protein | YOR265w | RBL2 | β -tubulin binding protein | YJL173c | RFA3 | DNA replication factor A, 13K subunit | | |
| YOL004w | SIN3 | transcription regulatory protein | YPL204w | HRR25 | casein kinase I, ser/thr/tyr protein kinase | YNL312w | RFA2 | DNA replication factor A, 36K subunit | | |
| YJL176c | SWI3 | transcription regulatory protein | YBR136w | ESR1 | cell cycle checkpoint protein | YAR007c | RFA1 | DNA replication factor A, 69K subunit | | |
| YHR084w | STE12 | transcriptional activator | YPL255w | BBP1 | cell division control protein | YOL094c | RFC4 | DNA replication factor C, 37K subunit | | |
| YER068w | MOT2 | transcriptional repressor | YDL220c | CDC13 | cell division control protein | YNL290w | RFC3 | DNA replication factor C, 40K subunit | | |
| YER068w | TPM1 | tropomyosin 1 | YPL042c | SSN3 | cyclin-dependent ser/thr protein kinase | YBR087w | RFC5 | DNA replication factor C, 40K subunit | | |
| YOR219c | STE13 | type IV dipeptidyl aminopeptidase | YOR368w | RAD17 | DNA damage checkpoint control protein | YJR068w | RFC2 | DNA replication factor C, 41K subunit | | |
| YMR022w | QR18 | ubiquitin-conjugating enzyme | YMR224c | MRE11 | DNA repair and meiotic recombination protein | YOR217w | RFC1 | DNA replication factor C, 95K subunit | | |
| YBR082c | UBC4 | ubiquitin-conjugating enzyme | YNL250w | RAD50 | DNA repair protein | YJR006w | HUS2 | DNA replication protein | | |
| YDR059c | UBC5 | ubiquitin-conjugating enzyme | YDR076w | RAD55 | DNA repair protein | YOL006c | TOP1 | DNA topoisomerase I | | |
| YER100w | UBC6 | ubiquitin-conjugating enzyme | YDR004w | RAD57 | DNA repair protein | YNL088w | TOP2 | DNA topoisomerase II (ATP-hydrolysing) | | |
| YJR117w | STE24 | zinc metallo-protease | YDR369c | XRS2 | DNA repair protein | YNL102w | CDC17 | DNA-directed DNA polymerase α , 180K subunit | | |
| sporulation and germination | | | YLR234w | TOP3 | DNA topoisomerase III | YIR008c | PRI1 | DNA-directed DNA polymerase α , 48K subunit (DNA primase) | | |
| YGR032w | GSC2 | 1,3-D-glucan synthase, subunit | YGL163c | RAD54 | DNA-dependent ATPase of the Snf2p family | YKL045w | PRI2 | DNA-directed DNA polymerase α , 58K subunit (DNA primase) | | |
| YPR103w | PRE2 | 26S proteasome subunit | YNL025c | SSN8 | DNA-directed RNA polymerase II holoenzyme and Kornberg's mediator (SRB) subcomplex subunit, cyclin C homolog | YPR135w | POB1 | DNA-directed DNA polymerase α -binding protein | | |
| YGR253c | PUP2 | 26S proteasome subunit | YKL017c | DIP1 | Dom34p Interacting Protein | YDL102w | CDC2 | DNA-directed DNA polymerase δ , catalytic 125K subunit | | |
| YBL015w | ACH1 | acetyl-CoA hydrolase | YLR129w | DIP2 | Dom34p-interacting protein | YNL262w | POL2 | DNA-directed DNA polymerase ϵ , catalytic subunit A | | |
| YCR048w | ARE1 | acyl-CoA sterol acyltransferase | YBR045c | GIP1 | Glc7-interacting protein | YPR175w | DPB2 | DNA-directed DNA polymerase ϵ , subunit B | | |
| YNR019w | ARE2 | acyl-CoA sterol acyltransferase | YPL240c | HSP82 | heat-shock protein | YBR278w | DPB3 | DNA-directed DNA polymerase ϵ , subunit C | | |
| YOL052c | SPE2 | adenosylmethionine decarboxylase | YJL146w | IDS2 | IME2-dependent signaling protein | YOR330c | MIP1 | DNA-directed DNA polymerase γ , catalytic subunit, mitochondrial | | |
| YDR173c | ARG82 | arginine metabolism transcription factor | YOR198c | BFR1 | maintenance of normal ploidy | YPL167c | REV3 | DNA-directed DNA polymerase ζ | | |
| YIL033c | SRA1 | cAMP dependent protein kinase, regulatory subunit | YHL022c | SPO11 | meiosis-specific protein | YFL036w | RPO41 | DNA-directed RNA polymerase, mitochondrial | | |
| YDR182w | CDC1 | cell division control protein | YER179w | DMC1 | meiosis-specific protein | YMR072w | ABF2 | high-mobility group protein | | |
| YFL009w | CDC4 | cell division control protein | YIL072w | HOP1 | meiosis-specific protein | YJL090c | DPB11 | involved in DNA replication and S-phase checkpoint | | |
| YNL192w | CHS1 | chitin synthase I | YER180c | ISC10 | meiosis-specific protein | YDR068w | DOS1 | involved in genome stability | | |
| YBR023c | CHS3 | chitin synthase III | YER044c-a | MEI4 | meiosis-specific protein | YMR011c | CDC5 | involved in regulation of DNA replication | | |
| YDR402c | DIT2 | cytochrome P450 5ll | YDL154w | MSH5 | meiosis-specific protein | YPR019w | CDC54 | member of the Cdc46p/Mcm2p/Mcm3p family | | |
| YML110c | DBI56 | DBF2 interacting protein | YHR124w | NDT80 | meiosis-specific protein | YBL023c | MCM2 | member of the Mcm2p, Mcm3p, Cdc46p family | | |
| YOL051w | GAL11 | DNA-directed RNA polymerase II holoenzyme and Kornberg's mediator (SRB) subcomplex subunit | YHR157w | REC104 | meiosis-specific protein | YML065w | ORC1 | origin recognition complex, 104K subunit | | |
| YBR253w | SRB6 | DNA-directed RNA polymerase II suppressor protein | YLR263w | RED1 | meiosis-specific protein | YNL261w | ORC5 | origin recognition complex, 50K subunit | | |
| YMR013c | SEC59 | dolichol kinase | YHR079c-a | SAE3 | meiosis-specific protein | YHR118c | ORC6 | origin recognition complex, 50K subunit | | |
| YOR190w | SPR1 | exo-1,3-glucanase | YHR014w | SPO13 | meiosis-specific protein | YPR162c | ORC4 | origin recognition complex, 56K subunit | | |
| YLR300w | EXG1 | exo-1,3-glucanase (I/II), major isoform | YPL121c | MEI5 | meiotic protein | YLL004w | ORC3 | origin recognition complex, 62K subunit | | |
| YDR261c | EXG2 | exo-1,3-glucanase minor isoform | YBR057c | MUM2 | meiotic protein | YBR060c | RRR1 | origin recognition complex, 72K subunit | | |
| YBR045c | GIP1 | Glc7p-interacting protein | YAL009w | SPO7 | meiotic protein | YGR132c | PHB1 | prohibitin, antiproliferative protein | | |
| YNL236w | SIN4 | global regulator protein | YHR025w | SNF7 | nuclear protein | YBR088c | POL30 | proliferating cell nuclear antigen (PCNA) protein kinase | | |
| YDL240w | LRG1 | GTPase-activating protein of the RHO/RAC family | YKR031c | SPO14 | phospholipase D | YDL017w | CDC7 | protein-tyrosine-phosphatase | | |
| YNL098c | RAS2 | GTP-binding protein | YGL192w | IME4 | positive transcription factor for IME2 | YFR028c | CDC14 | regulatory subunit for Cdc7p protein kinase | | |
| YPL240c | HSP82 | heat-shock protein | YGR034w | GNS1 | probable 1,3-glucan synthase subunit | YEL032w | MCM3 | replication initiation protein | | |
| YJL146w | IDS2 | IME2-dependent signaling protein | YIR026c | YVH1 | protein tyrosine phosphatase | YIL139c | REV7 | required for DNA damage-induced mutagenesis | | |
| YER044c-a | MEI4 | meiosis-specific protein | YOR208w | PTP2 | protein-tyrosine phosphatase | YLR103c | CDC45 | required for minichromosome maintenance and initiation of chromosomal DNA replication | | |
| YBR057c | MUM2 | meiotic protein | YBR073w | RDH54 | required for glucose derepression | YIL150c | DNA43 | required for S-phase initiation or completion | | |
| YAL009w | SPO7 | meiotic protein | YER133w | GLC7 | putative transcription factor | YER070w | RNR1 | ribonucleoside-diphosphate reductase, large subunit | | |
| YLR025w | SNF7 | nuclear protein | YOR178c | GAC1 | required for glucose derepression | YJL026w | RNR2 | ribonucleoside-diphosphate reductase, small subunit | | |
| YKR031c | SPO14 | phospholipase D | YJL106w | IME2 | positive transcription factor for IME2 | YIL066c | RNR3 | ribonucleotide reductase, repair inducible | | |
| YGL192w | IME4 | positive transcription factor for IME2 | YMR139w | MDS1 | probable 1,3-glucan synthase subunit | YJL065c | | similarity to DNA-directed DNA polymerase II subunit C | | |
| YGR034w | GNS1 | probable 1,3-glucan synthase subunit | YOR351c | MEK1 | protein tyrosine phosphatase | YDR206w | | similarity to EST1 protein | | |
| YIR026c | YVH1 | protein tyrosine phosphatase | YDR523c | SPS1 | protein tyrosine phosphatase | YOR005c | | similarity to human mRNA for DNA ligase IV | | |
| YOR208w | PTP2 | protein-tyrosine phosphatase | YNL307c | MCK1 | required for glucose derepression | YOL095c | | similarity to <i>S. aureus</i> DNA helicase | | |
| YBL066c | SEF1 | putative transcription factor | YDL028c | MPS1 | required for glucose derepression | YGL201c | | similarity with rat intestinal DNA replication protein | | |
| YNR052c | POP2 | required for glucose derepression | YNL001w | DOM34 | similarity to <i>D. melanogaster</i> pelota protein | YKL113c | RAD27 | ssDNA endonuclease and 5'-3' exonuclease | | |
| YDR218c | SPR28 | septin-related sporulation protein | YFL003c | MSH4 | similarity to MSH proteins | YCR028c-a | RIM1 | ssDNA-binding protein, mitochondrial | | |
| YJL106w | IME2 | IME2-dependent signaling protein | YHR152w | SPO12 | sporulation protein | YGR231c | | strong similarity to prohibitins | | |
| YDR523c | SPS1 | ser/thr protein kinase | YHR153c | SPO16 | sporulation protein | YBL035c | POL12 | subunit of DNA polymerase α -primase complex | | |
| YNL307c | MCK1 | ser/thr/tyr protein kinase | YDR108w | GGG1 | sporulation-specific protein | YLR233c | EST1 | telomere elongation protein | | |
| YDR403w | DIT1 | spore-wall maturation protein | YNL202w | SPS19 | sporulation-specific protein | YCR077c | (PAT1) | topoisomerase II-associated protein | | |
| YBR148w | YSW1 | spore-specific protein | YDR522c | SPS2 | sporulation-specific protein | YOL115w | TRF4 | topoisomerase II-related protein | | |
| YKL165c | | sporulation protein | YNL204c | SPS18 | sporulation-specific protein | | | | | |
| YLR399c | BDF1 | sporulation protein | YBR078w | | strong similarity to sporulation-specific Sps2p | | | | | |
| YHR152w | SPO12 | sporulation protein | YCL048w | | strong similarity to sporulation-specific Sps2p | | | | | |
| YHR153c | SPO16 | sporulation protein | YDR055w | | strong similarity to Sps2p | | | | | |
| YIL099w | SGA1 | sporulation specific glucan 1,4-glucosidase | YGR258c | RAD2 | structure-specific nuclease of the nucleotide excision repairosome | | | | | |
| YDR108w | GSG1 | sporulation-specific protein | YDR285w | ZIP1 | synaptonemal complex protein | | | | | |
| YER096w | SHC1 | sporulation-specific protein | | | | | | | | |
| YLR307w | CDA1 | sporulation-specific chitin deacetylase | | | | | | | | |
| YLR308w | CDA2 | sporulation-specific chitin deacetylase | | | | | | | | |
| YPR054w | SMK1 | sporulation-specific MAP kinase | | | | | | | | |
| YER115c | SPR6 | sporulation-specific protein | | | | | | | | |
| YNL202w | SPS19 | sporulation-specific protein | | | | | | | | |
| YDR522c | SPS2 | sporulation-specific protein | | | | | | | | |
| YOR313c | SPS4 | sporulation-specific protein | | | | | | | | |
| YGR059w | SPR3 | sporulation-specific septin | | | | | | | | |
| YHR139c | SPS100 | sporulation-specific wall maturation protein | | | | | | | | |
| YNL204c | SPS18 | sporulation-specific zinc-finger protein | | | | | | | | |

YLR182w *SWI6* transcription factor
 YDR054c *CDC34* ubiquitin-conjugating enzyme
 YGL068w *RAD6* ubiquitin-conjugating enzyme

recombination and DNA repair

YGL127c *SOH1* allows *hpr1* null mutant to grow at 37°C
 YBR136w *ESR1* cell-cycle checkpoint protein
 YER173w *RAD24* cell-cycle checkpoint protein
 YDR182w *CDC1* cell-division control protein
 YMR106c *HDF2* component of DNA end-joining repair pathway
 YKL011c *CCE1* cruciform-cutting endonuclease, mitochondrial
 YER176w DNA-dependent ATPase/DNA helicase B
 YLR032w *RAD5* DNA helicase
 YMR190c *SGS1* DNA helicase
 YML061c *PIF1* DNA helicase involved in mitochondrial DNA repair and telomere length
 YDL164c *CDC9* DNA ligase
 YMR167w *MLH1* DNA mismatch repair protein
 YOL090w *MSH2* DNA mismatch repair protein
 YNL082w *PMS1* DNA mismatch repair protein
 YHR120w *MSH1* DNA mismatch repair protein, mitochondrial
 YCR014c *POL4* DNA polymerase
 YMR224c *MRE11* DNA repair and meiotic recombination protein

YJR035w *RAD26* DNA repair and recombination protein
 YEL019c *MMS21* DNA repair protein
 YML095c *RAD10* DNA repair protein
 YCR066w *RAD18* DNA repair protein
 YNL250w *RAD50* DNA repair protein
 YER095w *RAD51* DNA repair protein
 YDR076w *RAD55* DNA repair protein
 YDR004w *RAD57* DNA repair protein
 YDR369c *XRS2* DNA repair protein
 YAR007c *RFA1* DNA replication factor A, 69k subunit
 YOL006c *TOP1* DNA topoisomerase I
 YNL088w *TOP2* DNA topoisomerase II (ATP-hydrolysing)
 YGL163c *RAD54* DNA-dependent ATPase of the Snp2p family
 YFR023w *PES4* DNA-directed DNA polymerase ε suppressor
 YPL167c *REV3* DNA-directed DNA polymerase ζ
 YCR092c *MSH3* DNA-repair protein
 YKR056w *RNC1* endo-exonuclease
 YOR033c *DHS1* exonuclease, interacting with Msh2p
 YDR288c *MEC3* G2-specific checkpoint protein
 YMR284w *HDF1* high-affinity DNA-binding protein
 YDR138w *HPR1* hyperrecombination protein related to Top1p

YHL022c *SPO11* meiosis-specific protein
 YER179w *DMC1* meiosis-specific protein
 YL072w *HOP1* meiosis-specific protein
 YHR157w *REC104* meiosis-specific protein
 YLR263w *RED1* meiosis-specific protein
 YPL121c *MEI5* meiotic protein
 YNL210w *MER1* meiotic recombination protein
 YHR086w *NAM8* meiotic recombination protein
 YLR329w *REC102* meiotic recombination protein
 YJR021c *REC107* meiotic recombination protein
 YGL175c *SAE2* meiotic recombination protein
 YBR114w *RAD16* nucleotide excision repair protein
 YGL025c *PGD1* probable transcription factor
 YBR088c *POL30* proliferating cell nuclear antigen (PCNA)
 YML032c *RAD52* recombination and DNA repair protein
 YDL059c *RAD59* recombination and DNA repair protein
 YLR383w *RHC18* recombination repair protein
 YBR073w *RDH54* required for meiosis
 YL066c *RNR3* ribonucleotide reductase, repair inducible
 YMR228w *MTF1* RNA polymerase specific factor, mitochondrial
 YOR351c *MEK1* ser/thr protein kinase
 YPL164c similarity to mismatch repair protein Mlh1p

YOR077w *RTS2* similarity to mouse KIN17 protein
 YFL003c *MSH4* similarity to MSH proteins
 YDR097c *MSH6* similarity to MSH proteins
 YHR031c similarity to Pif1p
 YGL043w *DST1* TFIIIS (transcription elongation factor)
 YCR077c *(PAT1)* topoisomerase II-associated protein
 YGR063c *SPT4* transcription initiation protein
 YHR116w *SPT6* transcription initiation protein
 YML021c *UNG1* uracil-DNA glycosylase

cell-cycle control and mitosis

YFR052w *NIN1* 26S proteasome regulatory subunit
 YPR103w *PRE2* 26S proteasome subunit
 YGL048c *SUG1* 26S proteasome subunit
 YKL145w *YTA3* 26S proteasome subunit
 YJL005w *CYR1* adenylate cyclase
 YPL239w *YAR1* ankyrin repeat-containing protein
 YHR101c *BIG1* big cells phenotype
 YHR208w *TWT1* branched-chain amino-acid aminotransferase, mitochondrial
 YFL037w *TUB2* β-tubulin
 YBR200w *BEM1* bud emergence mediator
 YBR109c *CMD1* calmodulin
 YOR061w *CKA2* casein kinase II α' subunit
 YL035c *CKA1* casein kinase II, catalytic α subunit
 YER123w *YCK3* casein kinase, isoform 3
 YLR178c *TF51* cdc25-dependent nutrient- and ammonia-response cell-cycle regulator
 YGL049w *SCM4* cdc4 suppressor
 YFL029c *CAK1* cdk-activating protein kinase
 YMR055c *BUB2* cell-cycle arrest protein

YOR026w *BUB3* cell-cycle arrest protein
 YPL178w *SAE1* cell-cycle block in meiotic prophase
 YBR136w *ESR1* cell-cycle checkpoint protein
 YPR111w *DBF20* cell-cycle protein kinase related to Dbf2p
 YDR113c *PDS1* cell-cycle regulator
 YBR215w *HPC2* cell-cycle regulatory protein
 YOR373w *NUD1* cell-cycle regulatory protein
 YPL255w *BBP1* cell division control protein
 YDR182w *CDC1* cell division control protein
 YCR002c *CDC10* cell division control protein
 YDL220c *CDC13* cell division control protein
 YGL116w *CDC20* cell division control protein
 YLR314c *CDC3* cell division control protein
 YDR168w *CDC37* cell division control protein
 YFL009w *CDC4* cell division control protein
 YDR364c *CDC40* cell division control protein
 YLR274w *CDC46* cell division control protein
 YBR202w *CDC47* cell division control protein
 YJL194w *CDC6* cell division control protein
 YNL188w *KAR1* cell division control protein
 YDL226c *GCS1* cell proliferation zinc-finger protein
 YNL327w *EGT2* cell-cycle regulation protein
 YHR129c *ACT5* centractin
 YLR175w *CBF5* centromere/microtubule binding protein
 YDR254w *CHL4* chromosome segregation protein
 YOR349w *CIN1* chromosome segregation protein
 YFL008w *SMC1* chromosome segregation protein
 YFR031c *SMC2* chromosome segregation protein
 YMR028w *TAP42* component of the Tor signalling pathway
 YL106w *MOB1* conditional mutants arrest in late mitosis
 YDL132w *CDC53* controls G1/S transition
 YPR120c *CLB5* cyclin, B-type
 YGR109c *CLB6* cyclin, B-type
 YMR199w *CLN1* cyclin, G1/S specific
 YPL256c *CLN2* cyclin, G1/S specific
 YAL040c *CLN3* cyclin, G1/S specific
 YNL289w *PCL1* cyclin, G1/S specific
 YDL127w *PCL2* cyclin, G1/S specific
 YGR108w *CLB1* cyclin, G2/M-specific
 YPR119w *CLB2* cyclin, G2/M-specific
 YDL155w *CLB3* cyclin, G2/M-specific
 YLR210w *CLB4* cyclin, G2/M-specific
 YBR135w *CKS1* cyclin-dependent kinase regulatory subunit

YBR160w *CDC28* cyclin-dependent protein kinase
 YPL031c *PHO85* cyclin-dependent protein kinase
 YDL108w *KIN28* cyclin-dependent ser/thr protein kinase
 YGL215w *CLG1* cyclin-like protein
 YOR368w *RAD17* DNA damage checkpoint control protein
 YDR217c *RAD9* DNA repair checkpoint protein
 YOR217w *RFC1* DNA replication factor C, 95k subunit
 YLR234w *TOP3* DNA topoisomerase III
 YKL017c *DIP1* Dom34p-interacting protein
 YLR129w *DIP2* Dom34p-interacting protein
 YNL053w *MSG5* dual-specificity protein phosphatase
 YKR054c *DYN1* dynein heavy chain, cytosolic
 YDR172w *SUP35* eukaryotic peptide chain release factor
 YNL292w *EXM1* exit from mitosis
 YDL087c *EXM2* exit from mitosis
 YJL157c *FAR1* factor arrest protein
 YLR212c *TUB4* γ tubulin
 YLR288c *MEC3* G2-specific checkpoint protein
 YAL024c *LTE1* GDP/GTP exchange factor
 YL016w *SDC25* GDP/GTP exchange factor
 YAL041w *CDC24* GDP/GTP exchange factor for Cdc42p
 YLR310c *CDC25* GDP/GTP exchange factor for Ras1p and Ras2p

YGL207w *SPT16* general chromatin factor
 YGL155w *CDC43* geranylgeranyltransferase type I β subunit
 YER155c *BEM2* GTPase-activating protein
 YMR138w *CIN4* GTP-binding protein
 YNL098c *RAS2* GTP-binding protein
 YLR229c *CDC42* GTP-binding protein of RAS superfamily
 YML064c *TEM1* GTP-binding protein of the RAS superfamily
 YNL007c *SIS1* heat-shock protein
 YBR133c *HSL7* histone synthetic lethality
 YJL080c *SCP160* histone-like protein
 YOL076w *DEC1* interacts genetically with *CIN8*
 YOR156c *NF1* interacts with Cdc12p in 2-hybrid assay
 YNL010w *CSE2* interacts with centromeric element CDEII
 YML104c *MDM1* intermediate filament protein
 YKR072c *SIS2* involved in cell cycle-specific gene expression
 YKR063c *LAS1* involved in cell morphogenesis, cytoskeletal regulation and bud formation
 YPL241c *CIN2* involved in chromosome segregation
 YPL018w *CTF19* involved in chromosome segregation
 YJL090c *DPB11* involved in DNA replication and S-phase checkpoint
 YMR273c *ZDS1* involved in negative regulation of cell polarity
 YMR052w *FAR3* involved in pheromone-mediated cell cycle arrest
 YMR052w *FAR3* involved in pheromone-mediated cell cycle arrest
 YMR001c *CDC5* involved in regulation of DNA replication
 YMR127c *SAS2* involved in silencing at HMR
 YKR042w *UTH1* involved in the aging process
 YDR293c *SSD1* involved in the tolerance to high concentration of Ca²⁺
 YNL189w *SRP1* karyopherin-α or importin
 YEL061c *CIN8* kinesin-related protein
 YPR141c *KAR3* kinesin-related protein
 YBL063w *KIP1* kinesin-related protein
 YJR060w *CBF1* kinetochore protein

YGR140w *CBF2* kinetochore protein complex CBF3, 110k subunit
 YMR094w *CTF13* kinetochore protein complex CBF3, 58k subunit
 YMR168c *CEP3* kinetochore protein complex CBF3, 71k subunit
 YDR328c *SKP1* kinetochore protein complex CBF3, subunit D
 YOR198c *BFR1* maintenance of normal ploidy
 YLR332w *MID2* mating process protein
 YER179w *DMC1* meiosis-specific protein
 YHR079c *SAE3* meiosis-specific protein
 YPR019w *CDC54* member of the Cdc46p/Mcm2p/Mcm3p family
 YDL126c *CDC48* microosomal protein
 YOR058c *ASE1* CDC48/PAS1/SEC18 family of ATPases
 YNL064c *YDI1* microtubule-associated protein
 YGR029w *ERV1* mitochondrial and ER import protein
 YBL016w *FUS3* mitochondrial biogenesis and regulation of cell cycle
 YMR294w *JNM1* mitogen-activated protein kinase (MAP kinase)
 YMR036c *MIH1* mitosis protein, involved in nuclear migration
 YGL178w *MPT5* M-phase inducing protein tyrosine phosphatase
 YML109w *ZDS2* multicopy suppressor of *pop2*
 YGL173c *KEM1* multicopy suppressor of *sin4*
 YDR207c *UME6* multifunctional nuclease
 YBL020w *RFT1* negative transcriptional regulator
 YNL299w *TRF5* nuclear division protein
 YML031w *NDC1* nuclear envelope protein
 YJR112w *NNF1* nuclear envelope protein
 YCL029c *BIK1* nuclear fusion protein
 YJL034w *KAR2* nuclear fusion protein
 YDR150w *NUM1* nuclear migration protein
 YAL025c *MAK16* nuclear viral propagation protein
 YKR048c *NAP1* nucleosome assembly protein I
 YLR079w *SIC1* p40 inhibitor of Cdc28p-Clnb protein kinase complex
 YJR066w *TOR1* phosphatidylinositol 3-kinase
 YKL203c *TOR2* phosphatidylinositol 3-kinase
 YLR305c *STT4* phosphatidylinositol-4-kinase
 YCL004w *PEL1* phosphatidylserine synthase
 YBL058w *SHP1* potential regulatory subunit for Glc7p
 YGL238w *CSE1* probable kinetochore protein
 YGR132c *PHB1* prohibitin, antiproliferative protein
 YDL017w *CDC7* protein kinase
 YOR149c *SMP3* protein kinase C pathway protein
 YPL140c *MKK2* protein kinase of the MAP kinase kinase (MEK) family
 YAR019c *CDC15* protein kinase of the MAP kinase kinase kinase family
 YHR013c *ARD1* protein N-acetyltransferase subunit
 YPL008w *CHL1* protein of the DEAH box family
 YDL047w *SIT4* protein ser/thr phosphatase
 YDL134c *PPH21* protein ser/thr phosphatase PP2A-1
 YDL188c *PPH22* protein ser/thr phosphatase PP2A-2
 YBR267w *PPS1* protein tyrosine phosphatase
 YFR028c *CDC14* protein-tyrosine-phosphatase
 YOL021c *DIS3* Ran binding protein
 YDR137w *RGP1* reduced growth phenotype protein
 YDR052c *DBF4* regulatory subunit for Cdc7p protein kinase
 YKL193c *SDS22* regulatory subunit for the mitotic function of type I protein phosphatase
 YMR078c *CHL12* required for accurate chromosome transmission in mitosis and maintenance of normal telomere length
 YOL145c *CTR9* required for G1 cyclin expression
 YLR103c *CDC45* required for minichromosome maintenance and initiation of chromosomal DNA replication
 YKL089w *MIF2* required for normal chromosome segregation and spindle integrity
 YGR098c *ESP1* required for normal spindle structure
 YIL150c *DNA43* required for S-phase initiation or completion
 YJL074c *SMC3* required for structural maintenance of chromosomes
 YGR078c *PAC10* required in the absence of Cin8p
 YDR488c *PAC11* required in the absence of Cin8p
 YLR075w *GRC5* ribosomal protein
 YJR076c *CDC11* septin
 YHR107c *CDC12* septin
 YAL016w *TPD3* ser/thr phosphatase 2A, regulatory subunit A
 YER133w *GLC7* ser/thr phosphoprotein phosphatase 1, catalytic subunit
 YOR178c *GAC1* ser/thr phosphoprotein phosphatase 1, regulatory subunit
 YGR188c *BUB1* ser/thr protein kinase
 YKL048c *ELM1* ser/thr protein kinase
 YJL106w *IME2* ser/thr protein kinase
 YPL209c *IPL1* ser/thr protein kinase
 YOR231w *MKK1* ser/thr protein kinase
 YBL105c *PKC1* ser/thr protein kinase
 YPR161c *SGV1* ser/thr protein kinase
 YJL141c *YAK1* ser/thr protein kinase
 YMR104c *YPK2* ser/thr protein kinase
 YHR030c *SLT2* ser/thr protein kinase of MAP kinase family
 YJL095w *BCK1* ser/thr protein kinase of the MEKK family
 YER167w *BCK2* ser/thr protein kinase of the protein kinase C pathway
 YGR092w *DBF2* ser/thr protein kinase related to Dbf20p

| | | |
|---------|---------------|---|
| YKL101w | <i>HSL1</i> | ser/thr protein kinase that interacts genetically with histone mutations |
| YHR102w | <i>NRK1</i> | ser/thr protein kinase that interacts with Cdc31p |
| YNL307c | <i>MCK1</i> | ser/thr/tyr protein kinase |
| YDL028c | <i>MPS1</i> | ser/thr/tyr protein kinase |
| YPL153c | <i>SPK1</i> | ser/thr/tyr protein kinase |
| YJL187c | <i>SWE1</i> | ser/tyr dual-specificity protein kinase |
| YGL003c | | similarity to <i>C. elegans</i> CDC20 protein and human p55CDC |
| YDL225w | | similarity to Cdc11p, Cdc3p and human CDC10 protein |
| YDL179w | | similarity to cyclin G1 homologue HCS26 |
| YNL001w | <i>DOM34</i> | similarity to <i>D. melanogaster</i> pelota protein |
| YNL246w | | similarity to <i>D. melanogaster</i> SET protein |
| YAR003w | <i>FUN16</i> | similarity to human RB protein binding protein |
| YKL179c | | similarity to Nuf1p |
| YDL125c | <i>YH11</i> | similarity to protein kinase C inhibitor-1 |
| YMR262w | | similarity to <i>S. pombe</i> scn1 protein |
| YLR356w | | similarity to SCM4 protein |
| YJL095w | <i>PRK1</i> | similarity to ser/thr protein kinase |
| YLR179c | | similarity to Tis1p and Nsp1p |
| YOL080c | | similarity to <i>X. laevis</i> XPMC2 protein |
| YFR040w | <i>SAP155</i> | Sit4p-associated protein |
| YKR028w | <i>SAP190</i> | Sit4p-associated protein |
| YGL229c | <i>SAP4</i> | Sit4p-associated protein |
| YJL098w | <i>SAP185</i> | Sit4p-associated protein |
| YGL086w | <i>MAD1</i> | spindle assembly checkpoint protein |
| YMR198w | <i>CIK1</i> | spindle pole body associated protein |
| YDR356w | <i>NUF1</i> | spindle pole body component |
| YKL042w | <i>SPC42</i> | spindle pole body component |
| YNL126w | <i>SPC38</i> | spindle pole body component |
| YOR257w | <i>CDC31</i> | spindle pole body component, centrin |
| YOL069w | <i>NUF2</i> | spindle pole body protein |
| YJL030w | <i>MAD2</i> | spindle-assembly checkpoint protein |
| YJL013c | <i>MAD3</i> | spindle-assembly checkpoint protein |
| YGL106w | | strong similarity to calmodulins |
| YHR205w | <i>SCH9</i> | strong similarity to cAMP-dependent protein kinase |
| YER066w | | strong similarity to cell division control protein Cdc4p |
| YLR117c | | strong similarity to <i>D. melanogaster</i> putative cell-cycle control protein crn |
| YIR012w | | strong similarity to Dbf8p |
| YBR238c | | strong similarity to general chromatin factor Spt16p |
| YKL049c | <i>CSE4</i> | strong similarity to histone H3 |
| YGR215c | | strong similarity to prohibitins |
| YLR215c | | strong similarity to rat cell cycle progression related D123 protein |
| YKL161c | | strong similarity to ser/thr-specific protein kinase Sit2p |
| YNL172w | <i>APC1</i> | subunit of anaphase-promoting complex (cyclosome) |
| YKL022c | <i>CDC16</i> | subunit of anaphase-promoting complex (cyclosome) |
| YHR166c | <i>CDC23</i> | subunit of anaphase-promoting complex (cyclosome) |
| YFR036w | <i>CDC26</i> | subunit of anaphase-promoting complex (cyclosome) |
| YBL084c | <i>CDC27</i> | subunit of anaphase-promoting complex (cyclosome) |
| YOR213c | | subunit of the RSC complex |
| YCR052w | <i>RSC6</i> | subunit of the RSC complex |
| YFR037c | <i>RSC8</i> | subunit of the RSC complex |
| YLR321c | <i>SFH1</i> | subunit of the RSC complex |
| YJL126w | <i>STH1</i> | subunit of the RSC complex |
| YJR091c | <i>JSN1</i> | suppresses the high-temperature lethality of tub2-150 |
| YIL031w | <i>SMT4</i> | suppressor of <i>mi12</i> temperature-sensitive mutation |
| YDR285w | <i>ZIP1</i> | synaptonemal complex protein |
| YPL129w | <i>ANC1</i> | TFIIF subunit (transcription initiation factor), 30K |
| YPR025c | <i>CCL1</i> | TFIIH subunit (transcription initiation factor), cyclin C component |
| YLR043c | <i>TRX1</i> | thioredoxin I |
| YGR209c | <i>TRX2</i> | thioredoxin II |
| YOL115w | <i>TRF4</i> | topoisomerase I-related protein |
| YDL165w | <i>CDC36</i> | transcription factor |
| YER111c | <i>SWI4</i> | transcription factor |
| YLR182w | <i>SWI6</i> | transcription factor |
| YGL181w | <i>GTS1</i> | transcription factor of the Gcs1p/Glc3p/Sps18p family |
| YMR043w | <i>MCM1</i> | transcription factor of the MADS box family |
| YHR206w | <i>SKN7</i> | transcription factor with similarity to Hsf1p |
| YDL056w | <i>MBP1</i> | transcription factor, subunit of the MBF factor |
| YOR028c | <i>CIN5</i> | transcriptional activator |
| YNL012w | <i>SPO1</i> | transcriptional regulator involved in sporulation |
| YOR361c | <i>PRT1</i> | translation initiation factor elf3 subunit |
| YHR165c | <i>PRP8</i> | U5 snRNP protein, pre-mRNA splicing factor |
| YDR054c | <i>CDC34</i> | ubiquitin-conjugating enzyme |
| YDL064w | <i>UBC9</i> | ubiquitin-conjugating enzyme |
| YMR276w | <i>DSK2</i> | ubiquitin-like protein |
| YML115c | <i>VAN1</i> | vanadate resistance protein |
| YHR071w | | weak similarity to cyclin G1 homolog HCS26 protein |
| YER122c | <i>GLO3</i> | zinc-finger protein |
| YLR403w | <i>SFP1</i> | zinc-finger protein |
| YGR044c | <i>RME1</i> | zinc-finger transcription factor |

cytokinesis

| | | |
|---------|--------------|--|
| YDL029w | <i>ACT2</i> | actin-like protein |
| YPL255w | <i>BBP1</i> | cell division control protein |
| YCR002c | <i>CDC10</i> | cell division control protein |
| YLR314c | <i>CDC3</i> | cell division control protein |
| YNL327w | <i>EGT2</i> | cell-cycle regulation protein |
| YNL192w | <i>CHS1</i> | chitin synthase I |
| YLR286c | <i>CTS1</i> | endochitinase |
| YOR156c | <i>NFI1</i> | interacts with Cdc12p in 2-hybrid assay |
| YNL225c | | involved in cytokinesis |
| YLR131c | <i>ACE2</i> | metallothionein expression activator |
| YAL029c | <i>MYO4</i> | myosin heavy chain, unconventional (class V) isoform |
| YHR023w | <i>MYO1</i> | myosin-1 isoform (type II myosin) heavy chain |
| YCR057c | <i>PWP2</i> | periodic tryptophan protein |
| YNL267w | <i>PIK1</i> | phosphatidylinositol 4-kinase |
| YJR076c | <i>CDC11</i> | septin |
| YHR107c | <i>CDC12</i> | septin |
| YGL190c | <i>CDC55</i> | ser/thr phosphatase 2A regulatory subunit B |
| YAL016w | <i>TPD3</i> | ser/thr phosphatase 2A, regulatory subunit A |
| YNL298w | <i>CLA4</i> | ser/thr protein kinase |
| YDL225w | | similarity to Cdc11p, Cdc3p and human CDC10 protein |
| YMR197c | | similarity to Nuf1p |

other cell-growth, cell-division and DNA-synthesis activities

| | | |
|---------|--------------|---|
| YLR248w | <i>RCK2</i> | Ca ²⁺ /calmodulin-dependent ser/thr protein kinase |
| YOR101w | <i>RAS1</i> | GTP-binding protein |
| YNL098c | <i>RAS2</i> | GTP-binding protein |
| YBR138c | <i>HDR1</i> | high-dosage reductional segregation defective |
| YHL003c | <i>LAG1</i> | longevity-assurance protein |
| YDL003w | <i>MCD1</i> | mitotic chromosome determinant |
| YGL091c | <i>NBP35</i> | nucleotide-binding protein |
| YOL110w | <i>SHR5</i> | RAS suppressor |
| YGL158w | <i>RCK1</i> | ser/thr protein kinase |
| YKL189w | | similarity to mouse MO25 gene |

Transcription

rRNA synthesis

| | | |
|-----------|---------------|---|
| YJR063w | <i>RPA12</i> | DNA-directed RNA polymerase I, 13.7K |
| YPR010c | <i>RPA135</i> | DNA-directed RNA polymerase I, 135K subunit |
| YOR341w | <i>RPA190</i> | DNA-directed RNA polymerase I, 190K α subunit |
| YOR340c | <i>RPA43</i> | DNA-directed RNA polymerase I, 36K subunit |
| YNL248c | <i>RPA49</i> | DNA-directed RNA polymerase I, 49K α subunit |
| YDR156w | <i>RPA14</i> | DNA-directed RNA polymerase I, A14 subunit |
| YOR224c | <i>RPB8</i> | DNA-directed RNA polymerase I, II, III 16K subunit |
| YPR187w | <i>RPO26</i> | DNA-directed RNA polymerase I, II, III 18K subunit |
| YBR154c | <i>RPB5</i> | DNA-directed RNA polymerase I, II, III 25K subunit |
| YHR143w-a | <i>RPC10</i> | DNA-directed RNA polymerase I, II, III 7.7K subunit |
| YOR210w | <i>RPB10</i> | DNA-directed RNA polymerase I, II, III 8.3K subunit |
| YPR110c | <i>RPC40</i> | DNA-directed RNA polymerase I, III 40K subunit |
| YNL113w | <i>RPC19</i> | DNA-directed RNA polymerase I, III, 16K subunit |
| YOR207c | <i>RPC128</i> | DNA-directed RNA polymerase III, 130K subunit |
| YOR116c | <i>RPO31</i> | DNA-directed RNA polymerase III, 160K subunit |
| YKL144c | <i>RPC25</i> | DNA-directed RNA polymerase III, 25K subunit |
| YNL151c | <i>RPC31</i> | DNA-directed RNA polymerase III, 31K subunit |
| YNR003c | <i>RPC34</i> | DNA-directed RNA polymerase III, 34K subunit |
| YDL150w | <i>RPC53</i> | DNA-directed RNA polymerase III, 47K subunit |
| YPR190c | <i>RPC82</i> | DNA-directed RNA polymerase III, 82K subunit |
| YLR039c | <i>RIC1</i> | involved in transcription of ribosomal proteins and ribosomal RNA |
| YKL125w | <i>RRN3</i> | RNA polymerase I specific transcription initiation factor |
| YBL025w | <i>RRN10</i> | RNA polymerase I specific transcription initiation factor |
| YML043c | <i>RRN11</i> | RNA polymerase I specific transcription initiation factor |
| YLR141w | <i>RRN5</i> | RNA polymerase I specific transcription initiation factor |
| YBL014c | <i>RRN6</i> | RNA polymerase I specific transcription initiation factor |
| YJL025w | <i>RRN7</i> | RNA polymerase I specific transcription initiation factor |
| YMR270c | <i>RRN9</i> | RNA polymerase I specific transcription initiation factor |
| YER148w | <i>SPT15</i> | TFIID and TFIIIB subunit |

| | | |
|---------|-------------|--|
| YPR186c | <i>TFC2</i> | TFIIA subunit (transcription initiation factor) |
| YGR246c | <i>BRF1</i> | TFIIIB subunit, 70K |
| YNL039w | <i>TFC5</i> | TFIIIB subunit, 90K |
| YGR047c | <i>TFC4</i> | TFIIIC subunit (transcription initiation factor), 131K |
| YAL001c | <i>TFC3</i> | TFIIIC subunit (transcription initiation factor), 138K |
| YBR123c | <i>TFC1</i> | TFIIIC subunit (transcription initiation factor), 95K |
| YBR049c | <i>REB1</i> | transcription factor |

rRNA processing

| | | |
|---------|--------------|---|
| YOR048c | <i>RAT1</i> | 5'-3' exoribonuclease |
| YBR142w | <i>MAK5</i> | ATP-dependent RNA helicase |
| YNL112w | <i>DBP2</i> | ATP-dependent RNA helicase of DEAD box family |
| YFL002c | <i>SPB4</i> | ATP-dependent RNA helicase of DEAH box family |
| YPL029w | <i>SUV3</i> | ATP-dependent RNA helicase, mitochondrial |
| YJL033w | <i>HCA4</i> | can suppress the U14 snoRNA rRNA processing function |
| YMR239c | <i>RNT1</i> | double-stranded ribonuclease |
| YDL014w | <i>NOP1</i> | fibrillar |
| YGL097w | <i>SRM1</i> | GDP/GTP exchange factor for Gsp1p/Gsp2p |
| YMR235c | <i>RNA1</i> | GTPase-activating protein |
| YLL011w | <i>SOF1</i> | involved in 18S pre-rRNA production |
| YGR158c | <i>MTR3</i> | involved in mRNA transport |
| YGL173c | <i>KEM1</i> | multifunctional nuclease |
| YGR159c | <i>NSR1</i> | nuclear localization sequence binding protein |
| YPL043w | <i>NOP4</i> | nucleolar protein |
| YDR432w | <i>NPL3</i> | nucleolar protein |
| YOR018c | <i>SRD1</i> | nucleolar protein |
| YHR089c | <i>GAR1</i> | nucleolar rRNA processing protein |
| YLR223c | <i>IFH1</i> | pre-rRNA processing machinery control protein |
| YMR229c | <i>RRP5</i> | processing of pre-ribosomal RNA |
| YNL221c | <i>POP1</i> | protein component of ribonuclease P and ribonuclease MRP |
| YHR065c | <i>RRP3</i> | required for maturation of the 35S primary transcript |
| YLL008w | <i>DRS1</i> | RNA helicase of the DEAD box family |
| YPL266w | <i>DIM1</i> | rRNA (adenosine-N ₆ ,N ₆ -) dimethyltransferase |
| YOR201c | <i>SET52</i> | rRNA (guanosine-2'-O-)-methyltransferase |
| YPL021w | <i>SRD2</i> | similarity to Srd1p |
| YHL034c | <i>SBP1</i> | single-strand nucleic acid binding protein |
| YLR059c | <i>YNT20</i> | suppressor of <i>mat12 yme2</i> |

other rRNA-transcription activities

| | | |
|---------|--------------|---|
| YNL216w | <i>RAP1</i> | DNA-binding protein with repressor and activator activity |
| YMR302c | <i>PRP12</i> | involved in early maturation of pre-rRNA |

tRNA synthesis

| | | |
|-----------|---------------|--|
| YGL019w | <i>CKB1</i> | casein kinase II, β subunit |
| YOR224c | <i>RPB8</i> | DNA-directed RNA polymerase I, II, III 16K subunit |
| YPR187w | <i>RPO26</i> | DNA-directed RNA polymerase I, II, III 18K subunit |
| YBR154c | <i>RPB5</i> | DNA-directed RNA polymerase I, II, III 25K subunit |
| YHR143w-a | <i>RPC10</i> | DNA-directed RNA polymerase I, II, III 7.7K subunit |
| YOR210w | <i>RPB10</i> | DNA-directed RNA polymerase I, II, III 8.3K subunit |
| YPR110c | <i>RPC40</i> | DNA-directed RNA polymerase I, III 40K subunit |
| YNL113w | <i>RPC19</i> | DNA-directed RNA polymerase I, III, 16K subunit |
| YOR207c | <i>RPC128</i> | DNA-directed RNA polymerase III, 130K subunit |
| YOR116c | <i>RPO31</i> | DNA-directed RNA polymerase III, 160K subunit |
| YKL144c | <i>RPC25</i> | DNA-directed RNA polymerase III, 25K subunit |
| YNL151c | <i>RPC31</i> | DNA-directed RNA polymerase III, 31K subunit |
| YNR003c | <i>RPC34</i> | DNA-directed RNA polymerase III, 34K subunit |
| YDL150w | <i>RPC53</i> | DNA-directed RNA polymerase III, 47K subunit |
| YPR190c | <i>RPC82</i> | DNA-directed RNA polymerase III, 82K subunit |
| YER148w | <i>SPT15</i> | TFIID and TFIIIB subunit |
| YPR186c | <i>TFC2</i> | TFIIA subunit (transcription initiation factor) |
| YGR246c | <i>BRF1</i> | TFIIIB subunit, 70K |
| YNL039w | <i>TFC5</i> | TFIIIB subunit, 90K |
| YGR047c | <i>TFC4</i> | TFIIIC subunit (transcription initiation factor), 131K |
| YAL001c | <i>TFC3</i> | TFIIIC subunit (transcription initiation factor), 138K |
| YBR123c | <i>TFC1</i> | TFIIIC subunit (transcription initiation factor), 95K |

tRNA processing

| | | |
|---------|-------------|---|
| YIL075c | <i>SEN3</i> | 26S proteasome regulatory subunit |
| YGL097w | <i>SRM1</i> | GDP/GTP exchange factor for Gsp1p/Gsp2p |
| YMR235c | <i>RNA1</i> | GTPase activating protein |

YHR006w *STP2* involved in pre-tRNA splicing
 YLR375w *STP3* involved in pre-tRNA splicing and in uptake of branched-chain amino acids
 YDL048c *STP4* involved in pre-tRNA splicing and in uptake of branched-chain amino acids
 YCL017c *NFS1* involved in tRNA processing and mitochondrial metabolism
 YNR034w *SOL1* multicopy suppressor of *los1-1*
 YCR073w-a *SOL2* multicopy suppressor of *los1-1*
 YMR047c *NUP116* nuclear pore protein
 YGL092w *NUP145* nuclear pore protein
 YJR042w *NUP85* nuclear pore protein
 YLR430w *SEN1* positive effector of tRNA-splicing endonuclease
 YAL043c *PTA1* pre-tRNA processing protein
 YKL205w *LOS1* pre-tRNA splicing protein
 YDR463w *STP1* pre-tRNA splicing protein
 YNL221c *POP1* protein component of ribonuclease P and ribonuclease MRP
 YML091c *RPM2* ribonuclease P, mitochondrial
 YFR004w *MPR1* strong similarity to *S. pombe* pad1 protein
 YGR248w *SOL4* strong similarity to Sol3p
 YLJ087c *TRL1* tRNA ligase
 YLR105c *SEN2* tRNA splicing endonuclease β subunit
 YHR163w *SOL3* weak multicopy suppressor of *los1-1*

tRNA modification

YGR204w *ADE3* C1-tetrahydrofolate synthase, cytoplasmic
 YMR283c *RTI1* initiator tRNA phosphoribosyl-transferase
 YDR120c *TRM1* N₂,N₂-dimethylguanine tRNA methyltransferase
 YGL105w *G4P1* protein with specific affinity for G4 quadruplex nucleic acids
 YPL212c *PUS1* pseudouridine synthase 1
 YGL063w *PUS2* pseudouridine synthase 2
 YFR010w similarity to *C. elegans* tRNA-guanine transglycosylase
 YBL013w similarity to methionyl-tRNA formyltransferase
 YOR274w *MOD5* tRNA isopentenyltransferase
 YER168c *CCA1* tRNA nucleotidyltransferase

other tRNA-transcription activities

YOR061w *CKA2* casein kinase II α' subunit
 YOR039w *CKB2* casein kinase II β' subunit
 YLJ041w *NSP1* nuclear pore protein

mRNA synthesis

general transcription activities

YDL108w *KIN28* cyclin-dependent ser/thr protein kinase
 YIL143c *SSL2* DNA helicase
 YER171w *RAD3* DNA helicase/ATPase
 YOR224c *RPB8* DNA-directed RNA polymerase I, II, III 16K subunit
 YPR187w *RPO26* DNA-directed RNA polymerase I, II, III 18K subunit
 YBR154c *RPB5* DNA-directed RNA polymerase I, II, III 25K subunit
 YHR143w-a *RPC10* DNA-directed RNA polymerase I, II, III 7.7K subunit
 YOR210w *RPB10* DNA-directed RNA polymerase I, II, III 8.3K subunit
 YOL051w *GAL11* DNA-directed RNA polymerase II holoenzyme and Kornberg's mediator (SRB) subcomplex subunit
 YDR443c *SCA1* DNA-directed RNA polymerase II holoenzyme and Kornberg's mediator (SRB) subcomplex subunit
 YHR041c *SRB2* DNA-directed RNA polymerase II holoenzyme and Kornberg's mediator (SRB) subcomplex subunit
 YER022w *SRB4* DNA-directed RNA polymerase II holoenzyme and Kornberg's mediator (SRB) subcomplex subunit
 YGR104c *SRB5* DNA-directed RNA polymerase II holoenzyme and Kornberg's mediator (SRB) subcomplex subunit
 YDR308c *SRB7* DNA-directed RNA polymerase II holoenzyme and Kornberg's mediator (SRB) subcomplex subunit
 YCR081w *SRB8* DNA-directed RNA polymerase II holoenzyme and Kornberg's mediator (SRB) subcomplex subunit
 YNL025c *SSN8* DNA-directed RNA polymerase II holoenzyme and Kornberg's mediator (SRB) subcomplex subunit, cyclin C homologue
 YLR071c *RGR1* DNA-directed RNA polymerase II holoenzyme subunit
 YBR253w *SRB6* DNA-directed RNA polymerase II suppressor protein
 YOL005c *RPB11* DNA-directed RNA polymerase II, 13.6K subunit
 YGL070c *RPB9* DNA-directed RNA polymerase II, 14.2K subunit
 YOR151c *RPB2* DNA-directed RNA polymerase II, 140K subunit
 YDR404c *RPB7* DNA-directed RNA polymerase II, 19K subunit
 YDL140c *RPO21* DNA-directed RNA polymerase II, 215K subunit
 YLJ140w *RPB4* DNA-directed RNA polymerase II, 32K subunit
 YFL036w *RPO41* DNA-directed RNA polymerase, mitochondrial

YIL021w *RPB3* DNA-directed RNA-polymerase II, 45K
 YLJ28w *MMS19* involved in repair and RNA polymerase transcription
 YMR228w *MTF1* RNA polymerase specific factor, mitochondrial
 YPR056w similarity to human transcription factor BTF2/TFIIH subunit p34
 YPL046c strong similarity to human DNA-directed RNA polymerase II elongation factor SIII, p15
 YDR045c strong similarity to *S. acidocaldarius* transcription elongation factor tfs
 YKL058w *TOA2* TFIIA subunit (transcription initiation factor), 13.5K
 YOR194c *TOA1* TFIIA subunit (transcription initiation factor), 32K
 YPR086w *SUA7* TFIIB subunit (transcription initiation factor), factor E
 YER148w *SPT15* TFIID and TFIIB subunit
 YGR274c *TAF145* TFIID subunit (TBP-associated factor), 145K
 YMR236w *TAF17* TFIID subunit (TBP-associated factor), 17K
 YML098w *TAF19* TFIID subunit (TBP-associated factor), 19K
 YDR167w *TAF23* TFIID subunit (TBP-associated factor), 23K
 YML015c *TAF40* TFIID subunit (TBP-associated factor), 40K
 YGL112c *TAF60* TFIID subunit (TBP-associated factor), 60K
 YDR145w *TAF61* TFIID subunit (TBP-associated factor), 61K
 YMR227c *TAF67* TFIID subunit (TBP-associated factor), 67K
 YBR198c *TAF90* TFIID subunit (TBP-associated factor), 90K
 YKR062w *TFA2* TFIIE subunit (transcription initiation factor), 43K
 YKL028w *TFA1* TFIIE subunit (transcription initiation factor), 66K
 YGR186w *TFG1* TFIIF subunit (transcription initiation factor), 105K
 YPL129w *ANC1* TFIIF subunit (transcription initiation factor), 30K
 YGR005c *TFG2* TFIIF subunit (transcription initiation factor), 54K
 YDR311w *TFB1* TFIIF subunit (transcription initiation factor), 75K
 YPR025c *CCL1* TFIIF subunit (transcription initiation factor), cyclin C component
 YLR005w *SSL1* TFIIF subunit (transcription initiation factor), factor B
 YPL122c *TFB2* TFIIF subunit (transcription/repair factor)
 YDR460w *TFB3* TFIIF subunit (transcription/repair factor)
 YGL043w *DST1* TFIIS (transcription elongation factor)
 YGR063c *SPT4* transcription initiation protein
 YML010w *SPT5* transcription initiation protein
 YGR116w *SPT6* transcription initiation protein

transcriptional control

YOR259c *CRL13* 26S proteasome subunit
 YLJ115w *ASF1* anti-silencing protein
 YDL197c *ASF2* anti-silencing protein
 YDR173c *ARG82* arginine metabolism transcription factor
 YKL112w *ABF1* ARS-binding factor
 YDR123c *INO2* basic helix-loop-helix (BHLH) transcription factor
 YOL108c *INO4* basic helix-loop-helix transcription factor
 YOL067c *RTG1* basic helix-loop-helix transcription factor
 YOR344c *TYE7* basic helix-loop-helix transcription factor
 YBL103c *RTG3* bHLH/zip transcription factor
 YHL009c bZip DNA binding protein
 YGL209w *MIG2* C2H2 zinc-finger protein
 YKL190w *CNB1* calcineurin B, regulatory subunit
 YML112w *CTK3* carboxy terminal domain (CTD) kinase, γ subunit
 YLJ006c *CTK2* carboxy-terminal domain (CTD) kinase, β subunit
 YKL139w *CTK1* carboxy-terminal domain (CTD) kinase, α subunit
 YIL035c *CKA1* casein kinase II, catalytic α subunit
 YGL237c *HAP2* CCAAT-binding factor subunit
 YBL021c *HAP3* CCAAT-binding factor subunit
 YLJ109w *HAP4* CCAAT-binding factor subunit
 YOR358w *HAP5* CCAAT-binding factor subunit
 YJR122w *CAF17* CCR4-associated factor
 YKR036c *CAF4* CCR4-associated factor
 YBR215w *HPC2* cell-cycle regulatory protein
 YDL220c *CDC13* cell division control protein component of SWI/SNF transcription activator complex
 YDR073w *SNF11* component of SWI/SNF transcription activator complex
 YNR023w *SNF12* component of SWI/SNF transcription activator complex
 YOR290c *SNF2* component of SWI/SNF transcription activator complex
 YBR289w *SNF5* component of SWI/SNF transcription activator complex
 YPL016w *SWI1* component of SWI/SNF transcription activator complex
 YCR042c *TSM1* component of TAF(II) complex
 YLJ177c *CUP9* copper homeostasis protein
 YGL166w *CUP2* copper-dependent transcription factor
 YNL167c *SKO1* cre-binding bzip protein
 YPL042c *SSN3* cyclin-dependent ser/thr protein kinase
 YLR176c *RFX1* DNA binding protein
 YDR217c *RAD9* DNA repair checkpoint protein
 YNL216w *RAP1* DNA-binding protein with repressor and activator activity
 YLR418c *CDC73* DNA-directed RNA polymerase II accessory protein
 YBR279w *PAF1* DNA-directed RNA polymerase II regulator
 YGL208w *SIP2* dominant suppressor of some ts mutations *rpo21* and *prp4*

YOR047c *STD1* dosage-dependent modulator of glucose repression
 YER159c *NCB1* functional homologue of human NC2 α
 YDR397c *YNC2B* functional homologue of human NC2 β /Dr1
 YPL037c *EGD1* GAL4 DNA-binding enhancer protein
 YLJ110c *GZF3* GATA zinc-finger protein 3
 YIL038c *NOT3* general negative regulator of transcription, subunit 3
 YBR112c *CYC8* general repressor of transcription
 YCR084c *TUP1* general transcription repressor
 YDR176w *NGG1* general transcriptional adaptor or co-activator
 YBR045c *GIP1* Glc7p-interacting protein
 YNL239w *SIN4* global regulator protein
 YHL025w *SNF6* global transcription activator
 YER027c *GAL83* glucose repression protein
 YNL199c *GOR2* glycolytic genes transcriptional activator
 YGL073w *HSF1* heat shock transcription factor
 YPR065w *YAP1* heme-dependent transcriptional repressor of hypoxic genes
 YDR006c *SOX1* high copy suppressor of a cyclic AMP-dependent protein kinase mutant
 YMR070w *HMS1* high-copy suppressor of *mot1 spt3* synthetic lethality
 YGR252w *GCN5* histone acetyltransferase
 YDR225w *HTA1* histone H2A
 YBL003c *HTA2* histone H2A.2
 YDR224c *HTB1* histone H2B
 YBL002w *HTB2* histone H2B.2
 YBR010w *HHT1* histone H3
 YNL031c *HHT2* histone H3
 YBR009c *HHF1* histone H4
 YNL030w *HHF2* histone H4
 YBL008w *HIR1* histone transcription regulator
 YOR038c *HIR2* histone transcription regulator
 YER161c *SPT2* HMG-like chromatin protein
 YDL106c *GRF10* homeodomain protein
 YDR138w *HPR1* hyperrecombination protein related to Top1p
 YLJ089w *SIP4* interacts with SNF1 protein kinase
 YKL032c *IXR1* intrastand crosslink recognition protein and transcription factor
 YBR081c *SPT7* involved in alteration of transcription start site selection
 YKR072c *SIS2* involved in cell cycle-specific gene expression
 YNL251c *NRD1* involved in regulation of nuclear pre-mRNA abundance
 YIL046w *MET30* involved in regulation of sulphur assimilation genes
 YDL153c *SAS10* involved in silencing
 YMR127c *SAS2* involved in silencing at HMR
 YGR097w *ASK10* involved in Skn7p-dependent transcription
 YLR039c *RIC1* involved in transcription of ribosomal proteins and ribosomal RNA
 YGL071w *RCS1* iron-regulated transcriptional repressor
 YJR060w *CBF1* kinetochore protein
 YDR159w *SAC3* leucine permease transcriptional regulator
 YGR100w *MIC1* Mac1p interacting protein
 YBR297w *MAL33* maltose fermentation regulatory protein
 YGR288w *MAL13* maltose pathway regulatory protein
 YCR040w *MAT α 1* mating type regulatory protein (expressed copy at MAT locus)
 YCR039c *MAT α 2* mating type regulatory protein (expressed copy at MAT locus)
 YCL067c *α 2* mating type regulatory protein (silenced copy at HML locus)
 YHL027w *RIM101* meiotic regulatory protein
 YLR216c *CPB6* member of the cyclophilin family
 YJR032w *CPB7* member of the cyclophilin family
 YLR136c *TIS11* member of the inducible cox zinc-finger family
 YOL148c *SPT20* member of the TBP class of SPT proteins that alter transcription site selection
 YMR021c *MAC1* metal binding activator
 YLR131c *ACE2* metallothionein expression activator
 YBR026c *(MRP1)* mitochondrial respiratory function protein
 YML051w *GAL80* negative regulator for expression of galactose-induced genes
 YGL151w *NUT1* negative regulator of HO endonuclease
 YPR168w *NUT2* negative regulator of HO endonuclease
 YKL185w *ASH1* negative regulator of HO expression
 YHL020c *OPI1* negative regulator of phospholipid biosynthesis pathway
 YDR464w *SPP41* negative regulator of PRP3 and PRP4 gene expression
 YNL076w *MKS1* negative regulator of RAS-cAMP pathway
 YDR207c *UME6* negative transcriptional regulator
 YGL221c *NIF3* Ngg1p-interacting factor 3
 YGL115w *SNF4* nuclear regulatory protein
 YCR093w *CDC39* nuclear transmembrane protein
 YML065w *ORC1* origin recognition complex, 104K subunit
 YNL261w *ORC5* origin recognition complex, 50K subunit
 YHR118c *ORC6* origin recognition complex, 50K subunit
 YPR162c *ORC4* origin recognition complex, 56K subunit
 YLL004w *ORC3* origin recognition complex, 62K subunit
 YBR060c *RRR1* origin recognition complex, 72K subunit
 YAL051w *OAF1* peroxisome proliferating transcription factor
 YOR363c *PIP2* peroxisome proliferating transcription factor
 YBL005w *PDR3* pleiotropic drug resistance regulatory protein
 YKL015w *PUT3* positive activator of the proline utilization pathway
 YGL192w *IME4* positive transcription factor for *IME2*

| | | | | | | | | |
|---------|--------------|--|---------|--------------|--|-------------------------------|---------------|--|
| YDR017c | <i>KCS1</i> | potential transcription factor of the BZIP type | YFR037c | <i>RSC8</i> | subunit of the RSC complex | chromatin modification | | |
| YGL025c | <i>PGD1</i> | probable transcription factor | YLR321c | <i>SFH1</i> | subunit of the RSC complex | YBR195c | <i>MSI1</i> | chromatin assembly complex, subunit p50 |
| YER108c | <i>FLO8</i> | probable transcriptional activator of Flo1p | YIL126w | <i>STH1</i> | subunit of the RSC complex | YML102w | <i>CAC2</i> | chromatin assembly complex, subunit p60 |
| YGL194c | <i>RTL1</i> | putative deacetylase | YGL224w | <i>ZZF1</i> | sulphite resistance protein | YPR018w | <i>RLF2</i> | chromatin assembly complex, subunit p90 |
| YJL103c | | putative regulatory protein | YNL252w | <i>SSU72</i> | suppressor of cs mutant <i>suaf7</i> | | | |
| YBL066c | <i>SEF1</i> | putative transcription factor | YDR310c | <i>SUM1</i> | suppressor of <i>mar1-1</i> | | | |
| YHR056c | | putative transcription regulator | YKR099w | <i>BAS1</i> | transcription factor | YGL207w | <i>SPT16</i> | general chromatin factor |
| YBR275c | <i>RIF1</i> | Rap1p-interacting factor 1 | YDL165w | <i>CDC36</i> | transcription factor | YGR252w | <i>GCN5</i> | histone acetyltransferase |
| YKL038w | <i>RG1</i> | regulator of glucose induced genes | YGL098c | <i>CHA4</i> | transcription factor | YGL194c | <i>RTL1</i> | putative deacetylase |
| YDR257c | <i>RMS1</i> | regulatory protein | YPL248c | <i>GAL4</i> | transcription factor | YHR119w | <i>YTX1</i> | regulatory protein |
| YDR392w | <i>SPT3</i> | regulatory protein | YFL031w | <i>HAC1</i> | transcription factor | YBR238c | | strong similarity to general chromatin factor Spt16p |
| YMR016c | <i>SOK2</i> | regulatory protein in the PKA signal transduction pathway | YLR256w | <i>HAP1</i> | transcription factor | | | |
| | | | YCR065w | <i>HCM1</i> | transcription factor | YFR004w | <i>MPR1</i> | strong similarity to <i>S. pombe</i> pad1 protein |
| YCL055w | <i>KAR4</i> | regulatory protein required for pheromone induction of karyogamy genes | YLR451w | <i>LEU3</i> | transcription factor | YOR213c | | subunit of the RSC complex |
| | | | YGL013c | <i>PDR1</i> | transcription factor | YCR052w | <i>RSC6</i> | subunit of the RSC complex |
| YNR052c | <i>POP2</i> | required for glucose depression | YGL043w | <i>PHD1</i> | transcription factor | YFR037c | <i>RSC8</i> | subunit of the RSC complex |
| YMR179w | <i>SPT21</i> | required for normal transcription at a number of loci | YFR034c | <i>PHO4</i> | transcription factor | YLR321c | <i>SFH1</i> | subunit of the RSC complex |
| | | | YBR049c | <i>REB1</i> | transcription factor | YIL126w | <i>STH1</i> | subunit of the RSC complex |
| YKL093w | <i>MBR1</i> | required for optimal growth on glycerol | YBL093c | <i>ROX3</i> | transcription factor | YGR274c | <i>TAF145</i> | TFIID subunit (TBP-associated factor), 145K |
| YOL068c | <i>HST1</i> | silencing protein | YOR140w | <i>SFL1</i> | transcription factor | | | |
| YOR025w | <i>HST3</i> | silencing protein | YLR055c | <i>SPT8</i> | transcription factor | YGR063c | <i>SPT4</i> | transcription initiation protein |
| YBL052c | <i>SAS3</i> | silencing protein | YER111c | <i>SWI4</i> | transcription factor | YML010w | <i>SPT5</i> | transcription initiation protein |
| YKR101w | <i>SIR1</i> | silencing regulatory protein | YDR146c | <i>SWI5</i> | transcription factor | YGR116w | <i>SPT6</i> | transcription initiation protein |
| YDL042c | <i>SIR2</i> | silencing regulatory protein | YLR182w | <i>SWI6</i> | transcription factor | | | |
| YLR442c | <i>SIR3</i> | silencing regulatory protein | YFL021w | <i>GAT1</i> | transcription factor for nitrogen regulation | | | |
| YDR227w | <i>SIR4</i> | silencing regulatory protein | YER040w | <i>GLN3</i> | transcription factor for positive nitrogen regulation | | | |
| YKL070w | | similarity to <i>B. subtilis</i> transcriptional regulatory protein | YMR042w | <i>ARG80</i> | transcription factor involved in arginine metabolism | | | |
| YDL070w | | similarity to bromodomain protein BDF1 | YML099c | <i>ARG81</i> | transcription factor involved in arginine metabolism | | | |
| YIL131c | <i>FKH1</i> | similarity to <i>D. melanogaster</i> fork head protein | YMR280c | <i>CAT8</i> | transcription factor involved in gluconeogenesis | | | |
| YIL056c | | similarity to developmental control proteins | YGL181w | <i>GTS1</i> | transcription factor of the Gcs1p/Glc3p/Sps18p family | | | |
| YBR061c | | similarity to <i>E. coli</i> fliJ protein | YMR043w | <i>MCM1</i> | transcription factor of the MADS box family | | | |
| YJR147w | | similarity to heat-shock transcription factors | YPL089c | <i>RLM1</i> | transcription factor of the MADS box family | | | |
| YGR249w | <i>MGA1</i> | similarity to heat-shock transcription factors | YLR014c | <i>PPR1</i> | transcription factor regulating pyrimidine pathway | | | |
| YPL015c | <i>HST2</i> | similarity to Hst1p and Sir2p | YJR094c | <i>IME1</i> | transcription factor required for sporulation | | | |
| YDR191w | <i>HST4</i> | similarity to Hst3p, Hst1p and Sir2p | YHR206w | <i>SKN7</i> | transcription factor with similarity to Hsf1p | | | |
| YNL107w | | similarity to human AF-9 protein | YDL056w | <i>MBP1</i> | transcription factor, subunit of the MBF factor | | | |
| YHR193c | <i>EGD2</i> | similarity to human α -NAC | YNL330c | <i>RPD3</i> | transcription modifier protein | | | |
| YAR003w | <i>FUN16</i> | similarity to human RB protein binding protein | YOL004w | <i>SIN3</i> | transcription regulatory protein | | | |
| YER169w | | similarity to human retinoblastoma binding protein 2 | YJL127c | <i>SPT10</i> | transcription regulatory protein | | | |
| YJR119c | | similarity to human retinoblastoma binding protein 2 | YJL176c | <i>SWI3</i> | transcription regulatory protein | | | |
| YLR116w | | similarity to human ZFM1 protein and mouse CW17R protein | YPL082c | <i>MOT1</i> | transcriptional accessory protein | | | |
| YJL206c | | similarity to hypothetical protein YIL130p and Put3p | YDR423c | <i>CAD1</i> | transcriptional activator | | | |
| YOL133w | | similarity to Lotus RING-finger protein | YOR028c | <i>CIN5</i> | transcriptional activator | | | |
| YER130c | | similarity to Msn2p and Msn4p | YPL075w | <i>GCR1</i> | transcriptional activator | | | |
| YER184c | | similarity to multidrug resistance protein PDR3 | YOL116w | <i>MSN1</i> | transcriptional activator | | | |
| YIL050w | | similarity to <i>N. crassa</i> regulatory protein preg(+) | YKL062w | <i>MSN4</i> | transcriptional activator | | | |
| YLR013w | | similarity to nitrogen regulatory proteins | YHR084w | <i>STE12</i> | transcriptional activator | | | |
| YDR334w | | similarity to nuclear Sth1p, Snf2p and related proteins | YIR023w | <i>DAL81</i> | transcriptional activator for allantoin and GABA catabolic genes | | | |
| YPR115w | | similarity to probable transcription factor Ask10p, and to hypothetical proteins YNL047c and YIL105c | YNL314w | <i>DAL82</i> | transcriptional activator for allantoin catabolic genes | | | |
| YIL130w | | similarity to Put3p and to hypothetical protein YJL206c | YDL170w | <i>UGA3</i> | transcriptional activator for GABA catabolic genes | | | |
| YCR020c | <i>PET18</i> | similarity to regulatory protein | YML007w | <i>YAP1</i> | transcriptional activator involved in oxidative stress response | | | |
| YBR182c | | similarity to Rlm1p, Mcm1p, and hMEF2 | YEL009c | <i>GCN4</i> | transcriptional activator of amino acid biosynthetic genes | | | |
| YKR008w | | similarity to <i>S. pombe</i> bromodomain protein | YDR034c | <i>LYS14</i> | transcriptional activator of lysine pathway genes | | | |
| YGL150c | | similarity to Snf2p and human SNF2 α | YIR017c | <i>MET28</i> | transcriptional activator of sulphur amino acid metabolism | | | |
| YLR228c | | similarity to transcription activator Lys14p | YNL103w | <i>MET4</i> | transcriptional activator of sulphur metabolism | | | |
| YLL054c | | similarity to transcription factor Pip2p | YPR104c | <i>FHL1</i> | transcriptional activator of the forkhead/hnf3 family | | | |
| YLR266c | | similarity to transcription factors | YDR448w | <i>ADA2</i> | transcriptional adaptor | | | |
| YLR278c | | similarity to transcription factors | YMR039c | <i>SUB1</i> | transcriptional coactivator | | | |
| YDR303c | | similarity to transcriptional regulator proteins | YAL021c | <i>CCR4</i> | transcriptional regulator | | | |
| YMR019w | <i>STB4</i> | SIN3 binding protein | YER164w | <i>CHD1</i> | transcriptional regulator | | | |
| YHR178w | <i>STB5</i> | SIN3 binding protein | YIL084c | <i>SDS3</i> | transcriptional regulator | | | |
| YKL072w | <i>STB6</i> | SIN3 binding protein | YNL012w | <i>SPO1</i> | transcriptional regulator involved in sporulation | | | |
| YNL309w | <i>STB1</i> | Sin3p binding protein | YGL035c | <i>MIG1</i> | transcriptional repressor | | | |
| YMR053c | <i>STB2</i> | Sin3p binding protein | YER068w | <i>MOT2</i> | transcriptional repressor | | | |
| YLR399c | <i>BDF1</i> | sporulation protein | YKR034w | <i>DAL80</i> | transcriptional repressor for allantoin and GABA catabolic genes | | | |
| YNL204c | <i>SPS18</i> | sporulation-specific zinc-finger protein | YMR182c | <i>RGM1</i> | transcriptional repressor protein | | | |
| YMR037c | | stress responsive regulatory protein | YCL054w | | transcriptional silencing protein | | | |
| YDR026c | | strong similarity to DNA-binding protein Reb1p | YOR083w | <i>TEC1</i> | Ty transcription activator | | | |
| YDR252w | <i>BTT1</i> | strong similarity to Egd1p and to human Btf3 protein | YOR337w | <i>TEA1</i> | TY1 enhancer activator | | | |
| YHR211w | <i>FLO5</i> | strong similarity to Flo1p | YER151c | <i>UBP3</i> | ubiquitin-specific proteinase | | | |
| YOR304w | | strong similarity to human Snf2p homologue | YER088c | | weak similarity human transforming proteins (B-myb) | | | |
| YOL055c | | strong similarity to hypothetical proteins YPL258c, YPR121w, similarity to <i>B. subtilis</i> transcriptional activator tenA | YBR240c | | weak similarity to regulatory proteins | | | |
| YFL052w | | strong similarity to Mal63p, Mal23p and Mal33p | YML076c | | weak similarity to transcription factor | | | |
| YPR196w | | strong similarity to regulatory protein Mal63p | YCR106w | | weak similarity to transcription factor Pip2p | | | |
| YBR245c | | strong similarity to SNF2/SWI2 DNA binding regulatory protein | YBR033w | | weak similarity to transcription factors | | | |
| YDR253c | | strong similarity to zinc-finger proteins | YKR064w | | weak similarity to transcription factors | | | |
| YML081w | | strong similarity to ZMS1 protein | YOL089c | | weak similarity to transcription factors, similarity to finger proteins YOR162c, YOR172w and YLR266c | | | |
| YEL056w | <i>HAT2</i> | strong similarity to ZMS1 protein subunit of the major yeast histone acetyltransferase | YKL222c | | weak similarity to transcription factors | | | |
| YOR213c | | subunit of the RSC complex | YBL054w | | weak similarity transforming protein (B-myb) | | | |
| YCR062w | <i>RSC6</i> | subunit of the RSC complex | YER122c | <i>GLO3</i> | zinc-finger protein | | | |
| | | | YDR216w | <i>ADR1</i> | zinc-finger transcription factor | | | |
| | | | YGR044c | <i>RME1</i> | zinc-finger transcription factor | | | |

YPR178w *PRP4* U4/U6 snRNP 52K protein
 YGR006w *PRP18* U5 snRNA-associated protein
 YHR165c *PRP8* U5 snRNP protein, pre-mRNA splicing factor
 YER112w *USS1* U6 snRNA associated protein

mRNA processing (5'-end, 3'-end processing and mRNA degradation)

YJL209w *CBP1* apo-cytochrome b pre-mRNA processing protein
 YBR120c *CBP6* apo-cytochrome b pre-mRNA processing protein
 YPL178w *SAE1* cell cycle block in meiotic prophase
 YGL094c *PAN2* component of Pab1p-stimulated poly(A) ribonuclease
 YMR061w *RNA14* component of pre-mRNA 3'-end processing factor
 YGL044c *RNA15* component of pre-mRNA 3'-end processing factor
 YJR093c *FIP1* component of pre-mRNA polyadenylation factor
 YKL025c *PAN3* component of the Pab1p-dependent poly(A)
 YOL149w *DCP1* component of the yeast decapping enzyme
 YGL097w *SRM1* GDP/GTP exchange factor for Gsp1p/Gsp2p
 YGR158c *MTR3* involved in mRNA transport
 YBR236c *ABD1* methyltransferase
 YGL130w *CEG1* mRNA guanylyltransferase (mRNA capping enzyme, α subunit)
 YER165w *PAB1* mRNA polyadenylate-binding protein
 YLR277c *BRR5* mRNA processing protein
 YKR002w *PAP1* poly(A) polymerase
 YGL122c *NAB2* poly(A)-binding protein
 YPL190c *NAB3* polyadenylated RNA-binding protein
 YDR301w *CFT1* pre-mRNA 3'-end processing factor
 YJR017c *ESS1* processing/termination factor 1
 YDR195w *REF2* RNA 3'-end formation protein
 YOR179c *REF2* similarity to BRR5 protein
 YOR319w *HSH49* similarity to human SAP49 and RNA-binding proteins
 YHR015w similarity to PES4 PAB-like protein
 YOR159c *SME1* strong similarity to human small nuclear ribonucleoprotein E

other mRNA-transcription activities

YGL171w *ROK1* ATP-dependent RNA helicase
 YOL006c *TOP1* DNA topoisomerase I
 YGL251c *HFM1* DNA/RNA helicase
 YNL037c *IDH1* isocitrate dehydrogenase (NAD⁺) subunit 1, mitochondrial
 YOR136w *IDH2* isocitrate dehydrogenase (NAD⁺) subunit 2, mitochondrial
 YNL016w *PUB1* major polyadenylated RNA-binding protein of nucleus and cytoplasm
 YHR170w *NMD3* nonsense-mediated mRNA decay protein
 YMR064w *AEP1* nuclear control of ATPase messenger RNA expression protein
 YOL123w *HRP1* polyadenylated RNA-binding protein
 YLR067c *PET309* required for stability and translation of COX1 mRNA
 YAL016w *TPD3* ser/thr phosphatase 2A, regulatory subunit A
 YOL042w similarity to CCR4 protein
 YER146w similarity to human snRNP E
 YER029c similarity to human snRNP-associated protein B
 YER028c similarity to Mig1p
 YDR429c similarity to nuclear RNA binding proteins
 YJR127c similarity to regulatory protein ADR1
 YDL031w similarity to RNA helicases
 YNL147w similarity to snRNP proteins
 YNL021w similarity to transcription factor Rpd3p
 YPR068c similarity to transcription factor Rpd3p
 YPL213w similarity to U2 snRNP protein A'
 YBL026w *SNP3* snRNP-related protein
 YDL160c *DHH1* strong similarity to RNA helicases of the DEAD box family
 YLR147c *SMD3* strong similarity to small nuclear ribonucleoprotein D3
 YLR139c *SLS1* suppresses lethality of SSM4 deletion

RNA transport

YGL097w *SRM1* GDP/GTP exchange factor for Gsp1p/Gsp2p
 YMR235c *RNA1* GTPase activating protein
 YOR185c *GSP2* GTP-binding protein
 YLR293c *GSP1* GTP-binding protein of the ras superfamily
 YOR160w *MTR10* involved in mRNA transport
 YGR158c *MTR3* involved in mRNA transport
 YJL050w *MTR4* involved in nucleocytoplasmic transport of mRNA
 YKL186c *MTR2* mRNA transport protein
 YKL069w *NUP100* nuclear pore protein
 YKL057c *NUP120* nuclear pore protein
 YKR082w *NUP133* nuclear pore protein
 YGL092w *NUP145* nuclear pore protein
 YDR192c *NUP42* nuclear pore protein
 YGL172w *NUP49* nuclear pore protein
 YJL061w *NUP82* nuclear pore protein
 YDR432w *NPL3* nucleolar protein
 YGL122c *NAB2* poly(A)-binding protein
 YKL205w *LOS1* pre-tRNA splicing protein

YER110c *KAP123* RAN-binding protein
 YDR002w *YRB1* ran-specific GTPase-activating protein
 YER107c *GLE2* required for nuclear pore complex structure and function
 YIR011c *STS1* required for transport of Rna15p from the cytoplasm to the nucleus
 YDL207w *GLE1* RNA export mediator
 YLR119w *SRN2* suppressor of *ma1-1* mutation

other transcription activities

YGL127c *SOH1* allows *hpr1* null mutant to grow at 37°C
 YOR113w *AZF1* asparagine-rich zinc-finger protein
 YPR198w *SGE1* drug resistance protein
 YBR212w *NGR1* glucose-repressible RNA-binding protein
 YML027w *YOX1* homoeodomain protein
 YNL068c *FKH2* homology to *D. melanogaster* forkhead protein
 YIL030c *SSM4* involved in mRNA turnover
 YNL282w *(POP2)* involved in processing RNAs
 YCR087c-a *YLA1* nucleic acid-binding protein
 YDL051w *YLA1* RNA binding protein
 YMR164c similarity to CYC8 protein
 YGL014w similarity to *D. melanogaster* pumilio protein and HTR1 protein
 YIR001c similarity to *D. melanogaster* RNA binding protein
 YNL004w *HRB1* similarity to Gbp2p
 YBR233w similarity to human hnRNP-E1 protein
 YPR031w similarity to human zinc-finger protein BR140
 YCL033c similarity to *M. capricolum* transcription repressor
 YIR005w similarity to RNA-binding proteins
 YMR213w similarity to *S. pombe* putative transcription factor cdc5
 YNL175c similarity to *S. pombe* Rnp24p
 YOR244w similarity to SAS2 and SAS3 protein
 YPR008w similarity to transcription factor
 YPL230w similarity to transcription factors
 YPR013c similarity to transcription factors
 YPR015c similarity to transcription factors
 YNL027w similarity to zinc-finger proteins
 YPL038w similarity to zinc-finger proteins
 YDR169c *STB3* SIN3 protein-binding protein
 YPR107c strong similarity to *D. melanogaster* zinc-finger protein
 YIL105c strong similarity to hypothetical protein YNL047c, similarity to hypothetical protein YNR115w and to probable transcription factor Ask10p
 YNL255c strong similarity to nucleic acid-binding proteins
 YHR169w strong similarity to RNA helicase
 YGL078c strong similarity to RNA helicase DBP2 protein
 YCR004c *YCP4* strong similarity to *S. pombe* protein obr1
 YFL017w-a *SNP2* strong similarity to snRNP E
 YDR451c strong similarity to Yox1p
 YML017w *PSP2* suppressor of DNA polymerase α mutation
 YKR092c *SRP40* suppressor of mutant AC40 of RNA polymerase I and III
 YKL005c weak similarity to *D. melanogaster* transcription elongation factor Dms-II
 YNR063w weak similarity to CYC1/CYP3 transcription activator
 YJL124c weak similarity to human Sm protein G
 YDR043c weak similarity to *K. marxianus* Mig1 and other regulatory proteins
 YIR018w weak similarity to transcription activator Pdr4p
 YPR199c weak similarity to transcription activator Yap1p
 YBR239c weak similarity to transcription factor Put3p
 YBR150c weak similarity to transcription factors
 YGR067c weak similarity to transcription factors
 YPL133c weak similarity to transcription factors
 YER116c zinc-finger protein

Protein synthesis

ribosomal proteins

YGR214w *NAB1A* 40S ribosomal protein p40 homologue A
 YLR048w *NAB1B* 40S ribosomal protein p40 homologue B
 YCR031c *CRY1* 40S ribosomal protein S14.e
 YGL189c *RPS26A* 40S ribosomal protein S26.e.c7
 YML009c *MRPL39A* 60S ribosomal protein, mitochondrial
 YDL081c *RPLA1* acidic ribosomal protein a1
 YLR340w *RPLA0* acidic ribosomal protein L10.e
 YDL130w *RPLA3* acidic ribosomal protein L44prime
 YDR382w *RPLA4* acidic ribosomal protein L45
 YOL039w *RPLA2* acidic ribosomal protein P2, β
 YOR369c *RS12* acidic ribosomal protein S12
 YGL069w probable ribosomal protein L12
 YLR325c putative ribosomal protein L38
 YFL034c-a ribosomal protein
 YFR032c-a ribosomal protein
 YLR061w ribosomal protein
 YMR142c ribosomal protein
 YPL183w-a ribosomal protein
 YLR075w *GRG5* ribosomal protein
 YIL133c *RP22* ribosomal protein
 YNL069c *RP23* ribosomal protein

YOR096w *RP30* ribosomal protein
 YKL008w *RPL14A* ribosomal protein
 YHL001w *RPL14B* ribosomal protein
 YMR242c *RPL18A* ribosomal protein
 YOR312c *RPL18B* ribosomal protein
 YDR471w *RPL27B* ribosomal protein
 YLR344w *RPL33A* ribosomal protein
 YMR194w *RPL39A* ribosomal protein
 YDL184c *RPL47A* ribosomal protein
 YDL133c-a *RPL47B* ribosomal protein
 YPL198w *RPL6B* ribosomal protein
 YOL040c *RPS21* ribosomal protein
 YLR287c-a *RPS21A* ribosomal protein
 YDL191w *RPS21B* ribosomal protein
 YDL136w *RPS22* ribosomal protein
 YPL220w *SSM1A* ribosomal protein
 YGL135w *SSM1B* ribosomal protein
 YGL123w *SUP44* ribosomal protein
 YIL148w *UBI1* ribosomal protein
 YKR094c *UBI2* ribosomal protein
 YHL015w *URP2* ribosomal protein
 YML073c *YL16A* ribosomal protein
 YLR448w *YL16B* ribosomal protein
 YDR500c *RPL35B* ribosomal protein L37.e
 YPR102c *RPL16A* ribosomal protein L11.e
 YDR418w *RPL15A* ribosomal protein L12.e
 YEL054c *RPL15B* ribosomal protein L12.e
 YDL082w ribosomal protein L13
 YKL170w *MRPL38* ribosomal protein L14, mitochondrial
 YLR029c *RPL13A* ribosomal protein L15.e.c12
 YMR121c *RPL13B* ribosomal protein L15.e.c13
 YJL063c *MRPL8* ribosomal protein L17, mitochondrial
 YKL180w *RPL20A* ribosomal protein L17.e
 YJL177w *RPL20B* ribosomal protein L17.e
 YNL301c *RPT28B* ribosomal protein L18.e
 YBL027w *RPL19A* ribosomal protein L19.e
 YBR084c-a *RPL19B* ribosomal protein L19.e
 YPL079w *URP1B* ribosomal protein L21
 YBR191w *URP1A* ribosomal protein L21.e
 YBL087c *RPL17A* ribosomal protein L23.e
 YER117w *RPL17B* ribosomal protein L23.e
 YOL127w *RPL25* ribosomal protein L23.a.e
 YGL031c *RPL30A* ribosomal protein L24.e
 YGR148c *RPL30B* ribosomal protein L24.e.B
 YGR034w *RPL33B* ribosomal protein L26
 YHR010w *RPL27A* ribosomal protein L27.e
 YGL103w *CYH2* ribosomal protein L27.a.e
 YBR031w *RPL2A* ribosomal protein L2A
 YOR063w *TCM1* ribosomal protein L3.0.e
 YGL030w *RPL32* ribosomal protein L30.e
 YDL075w *RPL43A* ribosomal protein L31.e
 YLR406c *RPL34B* ribosomal protein L31.e.c12
 YBL092w ribosomal protein L32.e
 YIL052c ribosomal protein L34.e
 YOR234c *RPL37B* ribosomal protein L35.a.e.c15
 YPL143w *RPL37A* ribosomal protein L35.a.e.c16
 YNL162w *RPL41A* ribosomal protein L36.a.e
 YHR141c *RPL41B* ribosomal protein L36.a.e
 YLR185w *RPL35A* ribosomal protein L37.e
 YPR043w ribosomal protein L37.a.e
 YJL189w *RPL46* ribosomal protein L39.e
 YDR012w *RPL2B* ribosomal protein L4.e.B
 YPL131w *RPL1* ribosomal protein L5.e
 YNL002c *RLP7* ribosomal protein L7.e
 YGL076c *RPL6A* ribosomal protein L7.e.A
 YHL033c *RPL4A* ribosomal protein L7.a.e.A
 YLL045c *RPL4B* ribosomal protein L7.a.e.B
 YIL018w *RPL5A* ribosomal protein L8.e
 YFR031c-a *RPL5B* ribosomal protein L8.e
 YGL147c *RPL9A* ribosomal protein L9.e
 YNL067w *RPL9B* ribosomal protein L9.e.c14
 YGR076c *MRPL25* ribosomal protein (YML25), mitochondrial
 YJR094w-a ribosomal protein of the large subunit
 YOR293w ribosomal protein S10.e
 YDR025w *RPS18A* ribosomal protein S11.e
 YBR048w *RPS18B* ribosomal protein S11.e.B
 YPR166c *MRP2* ribosomal protein S14
 YJL191w *CRY2* ribosomal protein S14.e.B
 YJL190c *RPS24A* ribosomal protein S15.a.e.c10
 YLR367w *RPS24B* ribosomal protein S15.a.e.c12
 YPL013c ribosomal protein S16, mitochondrial
 YMR143w *RPS16A* ribosomal protein S16.e
 YDL083c *RPS16B* ribosomal protein S16.e
 YML024w *RP51A* ribosomal protein S17.e.A
 YDR447c *RP51B* ribosomal protein S17.e.B
 YOL120c *RP28A* ribosomal protein S18.e
 YML026c *RPS18EB* ribosomal protein S18.e.c13
 YDR450w *RPS18EA* ribosomal protein S18.e.c4
 YOL121c *RP55A* ribosomal protein S19.e
 YNL302c *RP55B* ribosomal protein S19.e
 YKR057w *RPS25A* ribosomal protein S21.e
 YJL136c *RPS25B* ribosomal protein S21.e
 YGR118w *RPS28A* ribosomal protein S23.e
 YPR132w *RPS28B* ribosomal protein S23.e
 YER074w *RP50A* ribosomal protein S24.e
 YIL069c *RP50B* ribosomal protein S24.e
 YLR333c *RPS31B* ribosomal protein S25.e.c12
 YGR027c *RPS31A* ribosomal protein S25.e.c7
 YER131w *RPS26A* ribosomal protein S26.e-c5
 YKL156w *RPS27A* ribosomal protein S27.e
 YHR021c *RPS27B* ribosomal protein S27.e
 YDR064w *YS15* ribosomal protein S27.e
 YLR264w *RPS33B* ribosomal protein S28.e.c12
 YOR167c *RPS33A* ribosomal protein S28.e.c15
 YLR388w *YS29A* ribosomal protein S29.e.A
 YDL061c *YS29B* ribosomal protein S29.e.B
 YNL178w *RPS3* ribosomal protein S3.e

YLR441c *RP10A* ribosomal protein S3a.e
 YML063w *RP10B* ribosomal protein S3a.e
 YJR145c *RPS7B* ribosomal protein S4.e.c10
 YHR203c *RPS7A* ribosomal protein S4.e.c8
 YBR251w *MRPS55* ribosomal protein S5, mitochondrial
 YJR123w *RPS5* ribosomal protein S5.e
 YBR181c *RPS101* ribosomal protein S6.e
 YPL090c *RPS10B* ribosomal protein S6.e
 YBL072c *RPS8A* ribosomal protein S8.e
 YER102w *RPS8B* ribosomal protein S8.e
 YBR146w *MRPS9* ribosomal protein S9, mitochondrial
 YPL081w *RPS13B* ribosomal protein S9.e.A
 YBR189w *SUP46* ribosomal protein S9.e.B
 YGR085c *RPL16B* ribosomal protein YmL16.B
 YKR006c *MRPL13* ribosomal protein YmL13, mitochondrial
 YNL005c *MRPL2* ribosomal protein YmL2, mitochondrial
 YKR085c *MRPL20* ribosomal protein YmL20, mitochondrial
 YBR282w *MRPL27* ribosomal protein YmL27, mitochondrial
 YDR462w *MRPL31* ribosomal protein YmL28, mitochondrial
 YNL252c *MRPL32* ribosomal protein YmL30, mitochondrial
 YKL138c *MRPL31* ribosomal protein YmL31, mitochondrial
 YCR003w *MRPL32* ribosomal protein YmL32, mitochondrial
 YBR122c *MRPL36* ribosomal protein YmL36, mitochondrial
 YBR268w *MRPL37* ribosomal protein YmL37, mitochondrial
 YPL173w *MRPL44* ribosomal protein YmL44, mitochondrial
 YMR225c *RPL39B* ribosomal protein, cytoplasmic
 YPL249c-a *MRP1* ribosomal protein, mitochondrial
 YML025c *MRP13* ribosomal protein, mitochondrial
 YDR347w *MRP17* ribosomal protein, mitochondrial
 YGR084c *MRP17* ribosomal protein, mitochondrial
 YKL003c *MRP20* ribosomal protein, mitochondrial
 YDR405w *MRP4* ribosomal protein, mitochondrial
 YHL004w *MRP4* ribosomal protein, mitochondrial
 YKL167c *MRP49* ribosomal protein, mitochondrial
 YKL142w *MRP8* ribosomal protein, mitochondrial
 YLR312w-a *MRPL15* ribosomal protein, mitochondrial
 YBL038w *MRPL16* ribosomal protein, mitochondrial
 YMR193w *MRPL24* ribosomal protein, mitochondrial
 YMR024w *MRPL3* ribosomal protein, mitochondrial
 YMR286w *MRPL33* ribosomal protein, mitochondrial
 YLR439w *MRPL4* ribosomal protein, mitochondrial
 YHR147c *MRPL6* ribosomal protein, mitochondrial
 YGR220c *MRPL9* ribosomal protein, mitochondrial
 YDR337w *MRPS28* ribosomal protein, mitochondrial
 YNL137c *NAM9* ribosomal protein, mitochondrial
 YOR158w *PET123* ribosomal protein, mitochondrial
 YCR046c *PETCR46* ribosomal protein, mitochondrial
 YFR049w *YMR31* ribosomal protein, mitochondrial
 YJR113c similarity to ribosomal protein S7
 YHR148w similarity to ribosomal protein L1
 YDR116c similarity to ribosomal protein L13
 YOR150w similarity to ribosomal protein L15
 YNL284c similarity to ribosomal protein L2
 YEL050c similarity to ribosomal protein L24.e.B
 YLR009w similarity to ribosomal protein L34
 YDR115w similarity to ribosomal protein L5
 YDR237w similarity to ribosomal protein S13
 YNL081c similarity to ribosomal proteins
 YDR041w similarity to Rpl10p and S.solfataricus
 YKL009w ribosomal protein L10
 YDL208w *NHP2* strong similarity to high-mobility group (HMG) family
 YEL026w strong similarity to high-mobility group-like protein Nhp2p
 YOR182c *RPS30B* strong similarity to human ubiquitin-like protein/ribosomal protein S30
 YNR037c strong similarity to *Mycoplasma* ribosomal protein S19
 YNL185c strong similarity to ribosomal protein L11
 YER056c-a strong similarity to ribosomal protein L34.e
 YMR230w strong similarity to ribosomal protein S10
 YNR036c strong similarity to ribosomal protein S12
 YNL096c strong similarity to ribosomal protein S7
 YLR167w *UBI3* ubiquitin/ribosomal protein S27a
 YMR188c weak similarity to 30S ribosomal protein S17
 YDL202w weak similarity to ribosomal protein

translation (initiation, elongation and termination)

YDL081c *RPLA1* acidic ribosomal protein a1
 YMR309c *NIP1* associated with 40s ribosomal subunit
 YGL094c *PAN2* component of Pab1p-stimulated poly(A) ribonuclease
 YDR172w *SUP35* eukaryotic peptide chain release factor
 YNL007c *SIS1* GTP-binding subunit
 YBL091c *MAP2* heat-shock protein
 YMR023c *MSS1* methionine aminopeptidase, isoform 2
 mitochondrial GTPase involved in expression of *COX1*
 YGL143c *MRF1* mitochondrial peptide chain release factor
 YGL049c *TIF4632* mRNA cap-binding protein (eIF4F), 130K subunit
 YGR162w *TIF4631* mRNA cap-binding protein (eIF4F), 150K subunit
 YOR276w *CAF20* mRNA CAP-binding protein (eIF4F), 20K subunit
 YER165w *PAB1* mRNA polyadenylate-binding protein
 YKL173w *GIN10* similarity to elongation factor 2 eEF1
 YHR015w similarity to PES4 PAB-like protein
 YGR201c similarity to Tef3p and Tef4p
 YPL226w similarity to translation elongation factor eEF3

YLR289w *GUF1* strong similarity to *E. coli* elongation factor-type GTP-binding protein Iepa
 YDL084w strong similarity to nuclear RNA helicase (DEAD family)
 YDR021w strong similarity to translation initiation factor eIF4A
 YKL010c *SOS1* suppressor of *sis1*
 YLR005w *SSL1* TFIH subunit (transcription initiation factor), factor B
 YKR084c *HBS1* translation elongation factor eEF1
 YPR080w *TEF1* α -A subunit, cytosolic
 YBR118w *TEF2* translation elongation factor eEF1
 YPL048w *CAM1* translation elongation factor eEF1, α subunit
 YKL081w *TEF4* translation elongation factor eEF1, γ subunit
 YAL003w *EFB1* translation elongation factor eEF1 β
 YOR133w *EFT1* translation elongation factor eEF2
 YDR385w *EFT2* translation elongation factor eEF2
 YLR249w *YEF3* translation elongation factor eEF3
 YNL014w translation elongation factor eEF3 homologue
 YNL163c translation elongation factor eEF4
 YLR069c *MEF1* translation elongation factor G, mitochondrial
 YOR187w *TUF1* translation elongation factor TU, mitochondrial
 YJL102w *MEF2* translation elongation factor, mitochondrial
 YOL023w *IFM1* translation initiation factor 2, mitochondrial
 YNL244c *SUI1* translation initiation factor 3 (eIF3)
 YKR059w *TIF1* translation initiation factor 4A
 YMR260w *TIF11* translation initiation factor eIF1A
 YPL237c *SUI3* translation initiation factor eIF2 β subunit
 YJR007w *SUI2* translation initiation factor eIF2, α subunit
 YER025w *GCD11* translation initiation factor eIF2, γ subunit
 YKR026c *GNC3* translation initiation factor eIF2 β , 34K, α subunit
 YLR291c *GCD7* translation initiation factor eIF2 β , 43K subunit
 YGR083c *GCD2* translation initiation factor eIF2 β , 71K (δ) subunit
 YDR211w *GCD6* translation initiation factor eIF2 β , ϵ 81K subunit
 YOR260w *GCD1* translation initiation factor eIF2 β , γ subunit
 YOR361c *PR11* translation initiation factor eIF3 subunit
 YMR146c *TIF34* translation initiation factor eIF3, P39 subunit
 YNL062c *GCD10* translation initiation factor eIF3, RNA-binding subunit
 YJL138c *TIF2* translation initiation factor eIF4A
 YPR163c *TIF3* translation initiation factor eIF4B
 YOL139c *CDC33* translation initiation factor eIF4E
 YPR041w *TIF5* translation initiation factor eIF5
 YEL034w *HYP2* translation initiation factor eIF5A.1
 YJR047c *ANB1* translation initiation factor eIF5A.2
 YGL169w *SUA5* translation initiation protein
 YBR143c *SUP45* translational release factor

translational control

YMR282c *AEP2* 2'-O-ribosyl phosphate transferase
 YBR120c *CBP6* apo-cytochrome b pre-mRNA processing protein
 YPL029w *SUV3* ATP-dependent RNA helicase, mitochondrial
 YOR017w *PET127* component of mitochondrial translation system
 YMR028w *TAP42* component of the Tor signalling pathway
 YDR197w *CBS2* cytochrome b translational activator protein
 YNL216w *RAP1* DNA-binding protein with repressor and activator activity
 YER054c *GIP2* Glc7p-interacting protein
 YMR080c *NAM7* nonsense-mediated mRNA decay protein
 YGR072w *UPF3* nonsense-mediated mRNA decay protein
 YHR077c *NMD2* nonsense-mediated mRNA decay protein 2
 YMR064w *AEP1* nuclear control of ATPase messenger RNA expression protein
 YPL179w *PPQ1* phosphoprotein phosphatase
 YFR009w *GCN20* positive effector of Gcn2p
 YLR203c *MSS51* possibly involved in translational activation of COX1 and COB mRNA
 YBL058w *SHP1* potential regulatory subunit for Glc7p
 YNL139c *RLR1* regulatory protein
 YLR067c *PET309* required for stability and translation of COX1 mRNA
 YMR257c *PET111* required for translation of COX2 mRNA
 YER133w *GLC7* ser/thr phosphoprotein phosphatase 1, catalytic subunit
 YOR178c *GAC1* ser/thr phosphoprotein phosphatase 1, regulatory subunit
 YDR283c *GCN2* ser/thr protein kinase
 YGR222w *PET54* splicing protein and translational activator, mitochondrial
 YBR024w *SCO2* strong similarity to Sco1p
 YGL195w *GCN1* translational activator
 YDL069c *CBS1* translational activator of cob mRNA
 YER153c *PET122* translational activator of cytochrome c oxidase subunit III
 YNR045w *PET494* translational activator, mitochondrial
 YJL125c *GCD14* translational repressor of GCN4

tRNA synthetases

YOR335c *ALA1* alanyl-tRNA synthetase, cytosolic
 YHR091c *MSR1* arginyl-tRNA synthetase, mitochondrial
 YCR024c asn-tRNA synthetase, mitochondrial
 YHR019c *MSD1* asparaginyl-tRNA synthetase
 YPL104w *DED81* aspartate-tRNA ligase, mitochondrial
 YLL018c *DPS1* aspartyl-tRNA synthetase, cytosolic
 YOR168w *GLN4* glutamyl-tRNA synthetase
 YOL033w *MSE1* glutamyl-tRNA synthetase, mitochondrial
 YBR121c *GRS1* glycine-tRNA ligase
 YPR033c *HTS1* histidine-tRNA ligase, mitochondrial
 YPL040c *ISM1* isoleucine-tRNA ligase, mitochondrial
 YBL076c *ILS1* isoleucyl-tRNA synthetase
 YPL160w *CDC60* leucine-tRNA ligase, cytosolic
 YLR382c *NAM2* leucine-tRNA ligase, mitochondrial
 YDR037w *KRS1* lysyl-tRNA synthetase, cytosolic
 YNL073w *MSK1* lysyl-tRNA synthetase, mitochondrial
 YGR264c *MES1* methionyl-tRNA synthetase
 YGR171c *MSM1* methionyl-tRNA synthetase
 YPR047w *MSF1* phenylalanine-tRNA ligase α subunit, mitochondrial
 YFL022c *FRS2* phenylalanine-tRNA ligase β subunit, cytosolic
 YLR060w *FRS1* phenylalanyl-tRNA synthetase, α subunit, cytosolic
 YHR011w seryl-tRNA synthetase
 YDR023w *SES1* seryl-tRNA synthetase, cytosolic
 YNL247w similarity to cysteinyl-tRNA synthetases
 YER087w similarity to *E. coli* prolyl-tRNA synthetase
 YDR341c strong similarity to arginine-tRNA ligase and mitochondrial arginyl-tRNA synthetase
 YGL245w strong similarity to glutamine-tRNA ligase
 YPR081c strong similarity to glycyl-tRNA synthetases
 YHR020w strong similarity to human glutamyl-prolyl-tRNA synthetase
 YKL194c *MST1* threonine-tRNA ligase, mitochondrial
 YL078w *THS1* threonyl-tRNA synthetase, cytosolic
 YOL097c *WRS1* tryptophan-tRNA ligase
 YDR268w *MSW1* tryptophanyl-tRNA synthetase, mitochondrial
 YPL097w *MSY1* tyrosyl-tRNA synthetase
 YGR185c *TYS1* tyrosyl-tRNA synthetase
 YGR094w *VAS1* valyl-tRNA synthetase

other protein-synthesis activities

YDL229w *SSB1* heat-shock protein of HSP70 family
 YNL209w *SSB2* heat-shock protein of HSP70 family, cytosolic
 YGR147c *NAT2* N-acetyltransferase for N-terminal methionine
 YJR066w *TOR1* phosphatidylinositol 3-kinase
 YDL044c *MTF2* protein involved in mRNA splicing and protein synthesis, mitochondrial
 YDL040c *NAT1* protein N-acetyltransferase subunit
 YJL023c *PET130* protein synthesis protein, mitochondrial
 YMR005w *MPT1* required for protein synthesis
 YBL080c *PET112* required to maintain RHO⁺ mitochondrial DNA
 YHR189w similarity to peptidyl-tRNA hydrolases
 YLL039c *UBI4* ubiquitin

Protein destination

protein folding and stabilization

YEL030w heat-shock protein of HSP70 family
 YKL073w *LHS1* chaperone of the ER lumen
 YIL142w *CCT2* chaperonin of the TCP1 ring complex, cytosolic
 YJL014w *CCT3* chaperonin of the TCP1 ring complex, cytosolic
 YDR212w *CCT1* component of chaperonin-containing T-complex
 YDL143w *CCT4* component of chaperonin-containing T-complex
 YJL111w *CCT7* component of chaperonin-containing T-complex
 YJL008c *CCT8* component of chaperonin-containing T-complex
 YDR188w *CCT6* component of chaperonin-containing T-complex (ζ subunit)
 YDR155c *CPH1* cyclophilin (peptidylprolyl isomerase)
 YML078w *CPH3* cyclophilin (peptidylprolyl isomerase), mitochondrial
 YOR288c *MPD1* disulphide isomerase related protein
 YER048c *CAJ1* dnaJ homologue
 YDR519w *FKB2* FK506/rapamycin-binding protein of the ER
 YFL016c *MDJ1* heat-shock protein - chaperone
 YOR232w *MGE1* heat-shock protein - chaperone
 YLR259c *HSP60* heat-shock protein - chaperone, mitochondrial
 YJR045c *SSC1* heat-shock protein 70-related protein, mitochondrial
 YAL005c *SSA1* heat-shock protein of HSP70 family
 YLL024c *SSA2* heat-shock protein of HSP70 family, cytosolic
 YDR171w *HSP42* heat-shock protein with similarity to Hsp26p
 YLR216c *CPR6* member of the cyclophilin family
 YJR032w *CPR7* member of the cyclophilin family
 YJL034w *KAR2* nuclear fusion protein

YHR057c *CYP2* peptidyl-prolyl *cis-trans* isomerase
 YCR069w *SCC3* peptidyl-prolyl *cis-trans* isomerase
 YDR304c *CYP5* peptidyl-prolyl *cis-trans* isomerase D
 (cyclophilin D) of the ER
 YNL135c *FPR1* peptidyl-prolyl *cis-trans* isomerase, FK506-
 binding protein
 YML074c *NPI46* proline *cis-trans* isomerase
 YDR518w *EUG1* protein disulphide isomerase
 YCL043c *PDI1* protein disulphide-isomerase
 YHR097w similarity to Caj1p
 YBR044c similarity to chaperonin HSP60 proteins
 YOL088c similarity to disulphide isomerases and
 ER60 proteases
 YNL077w similarity to dnaJ protein homologue YDJ1
 YMR161w *HLJ1* similarity to dnaJ proteins
 YIR004w similarity to DNAJ-like proteins
 YLR090w *XDJ1* similarity to *E. coli* dnaJ
 YJL073w similarity to heat-shock proteins
 YNR028w similarity to peptidylprolyl isomerase
 Sec3p
 YIL005w similarity to protein disulphide isomerases
 YLR369w strong similarity to heat-shock protein 70-
 related proteins
 YLR449w strong similarity to peptidylprolyl
 isomerase FRP3
 YIR064w *CCT5* T-complex protein 1, ϵ subunit
 YFR041c weak similarity to dnaJ-like heat-shock
 proteins
 YNL227c weak similarity to dnaJ-like proteins

protein targeting, sorting and translocation

YKL073w *LHS1* chaperone of the ER lumen
 YGL206c *CHC1* clathrin heavy chain
 YOL062c *APM4* clathrin-associate protein YAP54
 YPL259c *APM1* clathrin-associated protein
 YBR288c *APM3* clathrin-associated protein complex,
 medium subunit
 YDL212w *SHR3* endoplasmic reticulum membrane protein
 YLR040c *ERD2* ER lumen protein-retaining receptor
 YER019c-a *SEB2* ER protein-translocation complex subunit
 YLR378c *SEC61* ER protein-translocation complex subunit
 YPL094c *SEC62* ER protein-translocation complex subunit
 YOR254c *SEC63* ER protein-translocation complex subunit
 YBR171w *SEC66* ER protein-translocation complex subunit
 YLR292c *SEC72* ER protein-translocation complex subunit
 YDR086c *SSS1* ER protein-translocation complex subunit
 YOR089c *VPS21* GTP-binding protein
 YKR014c *YPT52* GTP-binding protein of the RAB family
 YNL093w *YPT53* GTP-binding protein of the RAB family
 (RAS superfamily)
 YOR270c *VPH1* H⁺-ATPase V0 domain 95K subunit,
 vacuolar
 YAL005c *SSA1* heat-shock protein of HSP70 family
 YCR075c *ERS1* intracellular protein transport protein
 YGR028w *MSP1* intra-mitochondrial sorting protein
 YOR069w *VPS5* involved in Golgi retention and vacuolar
 sorting
 YLR347c *KAP95* karyopherin- β
 YOL122c *SMF1* manganese transporter
 YKR001c *VPS1* member of the dynamin family of
 GTPases
 YOR020c *HSP10* mitochondrial chaperonin
 YNR017w *MAS6* mitochondrial inner membrane import
 translocase subunit
 YJL143w *TIM17* mitochondrial inner membrane import
 translocase subunit
 YIL022w *TIM44* mitochondrial inner membrane import
 translocase subunit
 YMR035w *IMP2* mitochondrial inner membrane protease
 subunit
 YNL131w *TOM22* mitochondrial outer membrane import
 receptor subunit
 YGR082w *TOM20* mitochondrial outer membrane import
 receptor subunit, 20K
 YMR060c *TOM37* mitochondrial outer membrane import
 receptor subunit, 37K
 YMR203w *TOM40* mitochondrial outer membrane import
 receptor subunit, 40K
 YOR045w *TOM6* mitochondrial outer membrane import
 receptor subunit, 6K
 YNL070w *TOM7* mitochondrial outer membrane import
 receptor subunit, 7K
 YNL121c *TOM70* mitochondrial outer membrane
 specialized import receptor
 YMR150c *IMP1* mitochondrial protease
 YJL034w *KAR2* nuclear fusion protein
 YMR091c *NPL6* nuclear protein localization factor
 YBR170c *NPL4* nuclear protein localization factor and ER
 translocation component
 YDR432w *NPL3* nucleolar protein
 YLR191w *PAS20* peroxisomal protein involved in protein
 import
 YPR047w *MSF1* phenylalanine-tRNA ligase α subunit,
 mitochondrial
 YNL106c *PIE3* phosphatidylinositol phosphate
 phosphatase
 YBL069w *AST1* PMA1 protein targeting protein
 YLR168c *(MSF1)* probably involved in intramitochondrial
 protein sorting
 YDR244w *PAS10* putative peroxisomal targeting signal
 receptor
 YCL001w *RER1* required for correct localization of Sec12p
 YDR414c *ERD1* required for retention of luminal ER
 proteins
 YDR005c *MAF1* required for sorting of Mod5p
 YMR004w *MVP1* required for vacuolar protein sorting
 YDR498c *SEC20* secretory pathway protein

YBR162w-a *YSY6* secretory pathway protein
 YBR097w *VSP15* ser/thr protein kinase
 YPL243w *SRP68* signal recognition particle protein
 YPL210c *SRP72* signal recognition particle protein
 YDR292c *SRP101* signal recognition particle receptor, α
 subunit
 YML105c *SEC65* signal recognition particle subunit
 YDL092w *SRP14* signal recognition particle subunit
 YKL122c *SRP21* signal recognition particle subunit
 YPR088c *SRP54* signal recognition particle subunit
 YOR285w similarity to *D. melanogaster* heat-shock
 protein 67B2
 YOR286w similarity to *D. melanogaster* heat-shock
 protein 67B2
 YHR110w similarity to mammalian gp25L protein
 YAR002c-a similarity to mouse signal recognition
 YKL154w particle receptor β subunit
 similarity to Sec22p
 YKL196c similarity to tetratricopeptide-repeat
 YMR018w protein PAS10
 YMR214w *SCJ1* similarity to to *E. coli* dnaJ
 YNL304w similarity to Ypt1p and other GTP-binding
 proteins
 YER101c *AST2* strong similarity to Ast1p
 YOR016c strong similarity to FUN54 protein,
 similarity to hamster COOP-coated vesicle
 membrane protein
 YGL002w strong similarity to human gp25L2 protein
 YER087c-a *SEB1* strong similarity to mammalian Sec61 β
 subunit
 YCR099c strong similarity to Pep1p
 YCR100c strong similarity to Pep1p
 YCR101c strong similarity to Pep1p
 YIL173w strong similarity to Pep1p
 YJL222w strong similarity to Pep1p
 YBR283c strong similarity to Sec61p
 YOR327c *SNC2* strong similarity to synaptobrevin
 YOR329c *SCD5* suppressor of clathrin deficiency
 YHR050w *SMF2* suppressor of mitochondrial matrix
 mutant
 YOR036w *PEP12* syntaxin (T-SNARE)
 YLR148w *PEP3* vacuolar membrane protein
 YBL017c *PEP1* vacuolar protein sorting/targeting protein
 YJL053w *PEP8* vacuolar protein sorting/targeting protein
 YOR132w *VPS17* vacuolar protein sorting-associated
 protein
 YNR006w *VPS27* vacuolar protein sorting-associated
 protein
 YGL095c *VPS45* vacuolar protein sorting-associated
 protein
 YJL154c *VPS35* vacuolar protein-sorting protein
 YDR323c *PEP7* vacuolar segregation protein
 YPL045w *VPS16* vacuolar sorting protein
 YDR495c *VPS3* vacuolar sorting protein
 YLR396c *VPS33* vacuolar sorting protein
 YAL002w *VPS8* vacuolar sorting protein
 YML097c *VPS9* vacuolar sorting protein

protein modification (glycosylation, acylation, myristylation, palmitoylation, farnesylation and processing)

YJR131w *MNS1* α -1,2-mannosidase
 YDR483w *KRE2* α -1,2-mannosyltransferase
 YER001w *MNN1* α -1,3-mannosyltransferase
 YGL038c *OCH1* α -1,6-mannosyltransferase
 YBR164c *ARL1* ADP-ribosylation factor
 YKL157w *APE2* aminopeptidase yscil
 YJR062c *NTA1* amino-terminal amidase
 YGL017w *ATE1* arginyl tRNA transferase
 YPL154c *PEP4* aspartyl protease
 YLR120c *YAP3* aspergillopepsin
 YDL141w *BPL1* biotin holocarboxylase synthetase
 YBR110w *ALG1* β -mannosyltransferase
 YGL203c *KEX1* carboxypeptidase (YSC- α)
 YDL227c *ALG5* dolicho-P-glucose synthetase
 YJR143c *PMT4* dolichyl-phosphate-mannose-protein
 O-mannosyl transferase
 YDL212w *SHR3* endoplasmic reticulum membrane protein
 YNL238w *KEX2* endoprotease of late Golgi
 compartment
 YDR331w *GPI8* essential for GPI-anchor attachment
 YDR410c *STE14* farnesyl cysteine carboxyl-
 methyltransferase
 YOR370c *MSI4* geranylgeranyltransferase regulatory
 subunit
 YGL155w *CDC43* geranylgeranyltransferase type I β subunit
 YPR176c *BET2* geranylgeranyltransferase type II β subunit
 YJL031c *BET4* geranylgeranyltransferase, α subunit
 YOR002w *ALG6* glucosyltransferase
 YOR067c *ALG8* glucosyltransferase
 YEL042w *GDA1* guanosine diphosphatase
 YOR252w *GON5* histone acetyltransferase
 YPL001w *HAT1* histone acetyltransferase subunit
 YBR034c *HMT1* hnRNP methyltransferase
 YAL039c *CYC3* holocytochrome c synthase (cytochrome
 c heme lyase)
 YKL087c *CYT2* holocytochrome c1 synthase
 YGL065c *ALG2* mannosyltransferase
 YDL095w *PMT1* mannosyltransferase
 YAL023c *PMT2* mannosyltransferase
 YOR321w *PMT3* mannosyltransferase
 YBL082c *RHK1* mannosyltransferase
 YLR244c *MAP1* methionine aminopeptidase, isoform 1
 YBL091c *MAP2* methionine aminopeptidase, isoform 2
 YMR035w *IMP2* mitochondrial inner membrane protease
 subunit

YKL134c *(MIP1)* mitochondrial intermediate peptidase
 YLR163c *MAS1* mitochondrial processing peptidase
 YHR024c *MAS2* mitochondrial processing peptidase,
 catalytic 53K (α) subunit
 YMR150c *IMP1* mitochondrial protease
 YPR051w *MAK3* N-acetyltransferase
 YGR147c *NAT2* N-acetyltransferase for N-terminal
 methionine
 YBR247c *ENP1* N-glycosylation protein
 YLR195c *NMT1* N-myristoyltransferase
 YJL002c *OST1* oligosaccharyltransferase α subunit
 YEL002c *WBP1* oligosaccharyltransferase β subunit
 YMR149w *SWP1* oligosaccharyltransferase δ subunit
 YOR103c *OST2* oligosaccharyltransferase ϵ subunit
 YOR085w *OST3* oligosaccharyltransferase γ subunit
 YDL232w *OST4* oligosaccharyltransferase subunit
 YGL022w *STT3* oligosaccharyltransferase subunit
 YFL045c *SEC53* phosphomannomutase
 YPR122w *AXL1* protease
 YEL060c *PRB1* protease B, vacuolar
 YLR389c *STE23* protease involved in a-factor processing
 YCL043c *PDI1* protein disulphide-isomerase
 YKL019w *RAM2* protein farnesyltransferase, α subunit
 YDL090c *RAM1* protein farnesyltransferase, β subunit
 YHR013c *ARD1* protein N-acetyltransferase subunit
 YDL040c *NAT1* protein N-acetyltransferase subunit
 YIL085c *KTR7* putative α -1,2-mannosyltransferase
 YGL194c *RTL1* putative deacetylase
 YNL029c *KTR5* putative mannosyltransferase
 YGR199w *PMT6* putative mannosyltransferase
 YKL193c *SDS22* regulatory subunit for the mitotic function
 of type I protein phosphatase
 YNL048w *ALG11* required for asparagine-linked
 glycosylation
 YLR088w *GAA1* required for attachment of GPI anchor
 onto proteins
 YPL050c *MNN9* required for complex N-glycosylation
 YJR010c-a *SPC1* signal peptidase 10.8K subunit
 YML055w *SPC2* signal peptidase 18K subunit
 YIR022w *SEC11* signal sequence processing protein
 YDL042c *SIR2* silencing regulatory protein
 YER005w similarity to Gda1p
 YMR223w similarity to human putative ubiquitin
 carboxyl-terminal hydrolase
 YDR098c similarity to *Legionella* glutaredoxin-like
 protein
 YER174c similarity to *Legionella* glutaredoxin-like
 protein
 YGL257c similarity to Mnn1p
 YIL014w similarity to Mnn1p
 YPR131c similarity to N-acetyltransferases
 YDL093w *PMT5* similarity to O-mannosyltransferases
 Pmt1p-Pmt4p
 YDR307w similarity to Pmt1p
 YDR245w *MNN10* similarity to *S. pombe*
 galactosyltransferase
 YLR066w similarity to signal peptidase
 YNR059w similarity to to α -1,3-mannosyltransferase
 YPL003w similarity to ubiquitin-activating enzymes
 YPR180w similarity to ubiquitin-activating enzymes
 YGL087c similarity to ubiquitin-protein ligase
 YIR039c similarity to Yap3p
 YBR205w *KTR3* strong similarity to α -1,2-
 mannosyltransferase
 YBR199w *KTR4* strong similarity to α -1,2-
 mannosyltransferase
 YPL053c *KTR6* strong similarity to α -1,2-
 mannosyltransferase Kre2p
 YPL051w strong similarity to ADP-ribosylation
 factors
 YLR121c strong similarity to aspartylproteases
 YOR099w *KTR1* strong similarity to mannosyltransferases
 YCR083w strong similarity to thioredoxin
 YOR339c strong similarity to ubiquitin conjugating
 enzymes
 YPR066w strong similarity to ubiquitin-activating
 enzymes
 YML111w strong similarity to ubiquitination protein
 Bul1p
 YGL226c-a *OST5* subunit of N-oligosaccharyltransferase,
 ζ subunit
 YEL056w *HAT2* subunit of the major yeast histone
 acetyltransferase
 YJR075w *HOC1* suppressor of *pkc1*
 YLR043c *TRX1* thioredoxin I
 YGR209c *TRX2* thioredoxin II
 YOR219c *STE13* type IV dipeptidyl aminopeptidase
 YMR275c *BUL1* ubiquitination pathway protein
 YDR054c *CDC34* ubiquitin-conjugating enzyme
 YGR133w ubiquitin-conjugating enzyme
 YMR022w *QR18* ubiquitin-conjugating enzyme
 YGL058w *RAD6* ubiquitin-conjugating enzyme
 YDR177w *UBC1* ubiquitin-conjugating enzyme
 YLR306w *UBC12* ubiquitin-conjugating enzyme
 YDR092w *UBC13* ubiquitin-conjugating enzyme
 YDR059c *UBC5* ubiquitin-conjugating enzyme
 YER100w *UBC6* ubiquitin-conjugating enzyme
 YEL012w *UBC8* ubiquitin-conjugating enzyme
 YDL064w *UBC9* ubiquitin-conjugating enzyme
 YDR139c ubiquitin-like protein
 YER125w *RSP5* ubiquitin-protein ligase
 YKL210w *UBA1* ubiquitin-protein ligase
 YJR099w *YUH1* ubiquitin-specific protease
 YBR243c *ALG7* UDP-N-acetylglucosamine-1-phosphate
 transferase
 YKL035w *UGP1* UTP-glucose-1-phosphate
 uridylyltransferase
 YDR495c *VPS3* vacuolar sorting protein

YJR117w *STE24* zinc metallo-protease

assembly of protein complexes

YGR214w *NAB1A* 40S ribosomal protein p40 homologue A
 YLR048w *NAB1B* 40S ribosomal protein p40 homologue B
 YPL195w *YKS4* α - or γ - adaptin, large subunit of the clathrin-associated protein(AP) complex
 YBL037w *APL3* β -adaptin, large subunit of the clathrin-associated protein(AP) complex
 YOR094w *ARF3* ADP-ribosylation factor 3
 YBL022c *PIM1* ATP-dependent protease, mitochondrial
 YKL135c *APL2* β -adaptin
 YGR261c *YKS5* β -adaptin, large subunit of the clathrin-associated protein(AP) complex
 YBR195c *MSI1* chromatin assembly complex, subunit p50
 YPR018w *RLF2* chromatin assembly complex, subunit p90
 YLR170c *APS1* clathrin-associated protein (AP) complex, small subunit AP19
 YJR058c *APS2* clathrin-associated protein 17, small subunit
 YJR005w *YAP80* clathrin-associated protein complex, β subunit
 YJL024c *APS3* clathrin-associated protein(AP) complex, small subunit
 YER058w *PET117* cytochrome c oxidase assembly factor
 YPL132w *COX11* cytochrome c oxidase assembly protein
 YML129c *COX14* cytochrome c oxidase assembly protein
 YDR079w *PET100* cytochrome c oxidase assembly protein
 YLR038c *COX12* cytochrome c oxidase subunit VIB
 YMR256c *COX7* cytochrome c oxidase subunit VII
 YDL067c *COX9* cytochrome c oxidase subunit VIIA
 YER141w *COX15* cytochrome oxidase assembly factor
 YER154w *OXA1* cytochrome oxidase biogenesis protein
 YBL007c *SLA1* cytoskeleton assembly control protein
 YNL243w *SLA2* cytoskeleton assembly control protein
 YLR393w *ATP10* F1F0 ATPase complex assembly protein
 YNL315c *ATP11* F1F0-ATPase complex assembly protein
 YJL180c *ATP12* F1F0-ATPase complex assembly protein
 YPL172c *COX10* farnesyl transferase
 YDL231w *FAS2* fatty-acyl-CoA synthase, α subunit
 YPR029c *APL4* γ -adaptin, large subunit of the clathrin-associated protein(AP) complex
 YDL192w *ARF1* GTP-binding protein of the ARF family
 YDL137w *ARF2* GTP-binding protein of the ARF family
 YKL119c *VPH2* H⁺-ATPase assembly protein, vacuolar
 YPL234c *TFP3* H⁺-ATPase V0 domain 17K subunit, vacuolar
 YLR447c *VMA6* H⁺-ATPase V0 domain 36K subunit, vacuolar
 YOR270c *VPH1* H⁺-ATPase V0 domain 95K subunit, vacuolar
 YOR332w *VMA4* H⁺-ATPase V1 domain 27K subunit, vacuolar
 YKL080w *VMA5* H⁺-ATPase V1 domain 42K subunit, vacuolar
 YLL009c *COX17* interacts genetically with *SCO1* and *SCO2* in cytochrome oxidase assembly
 YJR034w *PET191* involved in assembly of cytochrome oxidase
 YBR037c *SCO1* involved in stabilization of Cox1p and Cox2p
 YDR375c *BCS1* mitochondrial protein of the CDC48/PAS1/SEC18 (AAA) family of ATPases
 YPL085w *SEC16* multidomain vesicle coat protein
 YKR048c *NAP1* nucleosome assembly protein I
 YML102w *CAC2* p60 subunit of the chromatin assembly factor-I (CAF-I)
 YKL197c *PAS1* peroxisomal assembly protein
 YDR265w *PAS4* peroxisomal assembly protein
 YNL329c *PAS8* peroxisomal assembly protein
 YBR237w *PRP5* pre-mRNA processing RNA-helicase
 YJL203w *PRP21* pre-mRNA splicing factor
 YER017c *AFG3* protease of the SEC18/CDC48/PAS1 family of ATPases (AAA)
 YPR024w *YME1* protease of the SEC18/CDC48/PAS1 family of ATPases (AAA)
 YMR089c *YTA12* protease of the SEC18/CDC48/PAS1 family of ATPases (AAA)
 YPL215w *CBP3* required for assembly of cytochrome bc1 complex
 YBR185c *MBA1* respiratory chain assembly protein
 YLL008w *DRS1* RNA helicase of the DEAD box family
 YBR227c similarity to *E. coli* ATP-binding protein clpX
 YGR241c similarity to rat clathrin assembly protein
 YHR161c similarity to rat clathrin assembly protein AP180
 YGR074w *SMD1* snRNA-associated protein
 YBR024w *SCO2* strong similarity to Sco1p
 YLR327c strong similarity to STF2p
 YOR134w *BAG7* structural homologue of Sac7p
 YDR389w *SAC7* suppressor of actin mutation
 YOR106w *VAM3* syntaxin related protein
 YDR460w *TFB3* TFIIF subunit (transcription/repair factor)
 YPR178w *PRP4* U4/U6 snRNP 52K protein
 YGL119w *ABC1* ubiquinol cytochrome c reductase complex assembly protein
 YPR191w *QCR2* ubiquinol-cytochrome c reductase 40K subunit II
 YGR174c *CBP4* ubiquinol-cytochrome c reductase assembly factor
 YLL039c *UBI4* ubiquitin
 YGR133w *PAS2* ubiquitin-conjugating enzyme

YGR105w *VMA21* vacuolar ATPase assembly integral membrane protein
 YHR060w *VMA22* vacuolar ATPase assembly protein
 YDR495c *VPS3* vacuolar sorting protein

proteolysis

YPL149w *APG5* involved in autophagy and nutrient starvation
 YOL013c *HRD1* involved in degradation of Hmg2p
 YLR207w *HRD3* involved in degradation of Hmg2p
 YMR174c *PAI3* protease A (ysca) inhibitor IA3
 YNL015w *PBI2* protease B inhibitor 2
 YCL052c *PBN1* required for Prb1p expression
 YHR132c similarity to carboxypeptidase
 YNR069c similarity to central part of Bul1p
 YBR227c similarity to *E. coli* ATP-binding protein clpX
 YNL186w similarity to human putative ubiquitin carboxy-terminal hydrolase
 YJL137c similarity to *M. musculus* aminopeptidase
 YKR038c similarity to Qri7p
 YER047c similarity to regulatory subunit Yta6p of 26S proteasome
 YCR045c similarity to serin proteases
 YER078c similarity to X-Pro aminopeptidase II
 YBR139w strong similarity to carboxypeptidase
 YBL067c ubiquitin carboxy-terminal hydrolase
 YOL111c weak similarity to human ubiquitin-like protein GDx

cytoplasmic degradation

YFR052w *NIN1* 26S proteasome regulatory subunit
 YJL075c *SEN3* 26S proteasome regulatory subunit
 YOR259c *CRL13* 26S proteasome subunit
 YPR103w *PPE2* 26S proteasome subunit
 YFR050c *PPE4* 26S proteasome subunit
 YOR157c *PUP1* 26S proteasome subunit
 YGR253c *PUP2* 26S proteasome subunit
 YER094c *PUP3* 26S proteasome subunit
 YGL048c *SUG1* 26S proteasome subunit
 YHR200w *SUN1* 26S proteasome subunit
 YER021w *SUN2* 26S proteasome subunit
 YHR117w *YTA1* 26S proteasome subunit
 YDR394w *YTA2* 26S proteasome subunit
 YKL145w *YTA3* 26S proteasome subunit
 YGR270w *YTA7* 26S proteasome subunit
 YOR362c *PRE10* 26S proteasome subunit C1
 YER012w *PRE1* 26S proteasome subunit C11
 YGR135w *PRE9* 26S proteasome subunit Y13
 YML092c *PRE8* 26S proteasome subunit Y7
 YGL011c *SCL1* 26S proteasome subunit YC7 α /Y8
 YMR314w *PRE5* 26S proteasome subunit, α -type
 YNL239w *LAP3* aminopeptidase of cysteine protease family
 YJR062c *NTA1* amino-terminal amidase
 YGL017w *ATE1* arginyl tRNA transferase
 YDL132w *CDC53* controls G1/S transition
 YLR452c *SST2* involved in desensitization to α -factor pheromone
 YKL213c *DOA1* involved in ubiquitin-dependent proteolysis
 YDL126c *CDC48* microosomal protein of CDC48/PAS1/SEC18 family of ATPases
 YJL001w *PRE3* multicatalytic endopeptidase complex subunit
 YOL038w *PRE6* multicatalytic endopeptidase complex subunit
 YBL041w *PRE7* multicatalytic endopeptidase complex subunit
 YDL020c *SON1* nuclear protein
 YBR165w *UBS1* positive regulator of Cdc34p
 YDL007w *YTA5* probable component of 26S proteasome complex
 YJL148w *UBI1* ribosomal protein
 YKR094c *UBI2* ribosomal protein
 YMR223w similarity to human putative ubiquitin carboxyl-terminal hydrolase
 YDR390c *UBA2* similarity to Uba1p
 YPL003w similarity to ubiquitin-activating enzymes
 YPR180w similarity to ubiquitin-activating enzymes
 YGL087c similarity to ubiquitin-protein ligase
 YPL074w *YTA6* similarity to Vps4p and Yta4p
 YOR261c strong similarity to human 26S proteasome regulatory subunit, p40
 YOR339c strong similarity to ubiquitin conjugating enzymes
 YPR066w strong similarity to ubiquitin-activating enzymes
 YML111w strong similarity to ubiquitination protein Bul1p
 YHR027c *HRD2* subunit of 26S proteasome
 YKL022c *CDC16* subunit of anaphase-promoting complex (cytosome)
 YHR166c *CDC23* subunit of anaphase-promoting complex (cytosome)
 YBL084c *CDC27* subunit of anaphase-promoting complex (cytosome)
 YLL039c *UBI4* ubiquitin
 YJL156w *UBP7* ubiquitin carboxy terminal hydrolase
 YER098w *UBP9* ubiquitin carboxyl-terminal hydrolase
 YKR098c *UBP11* ubiquitin C-terminal hydrolase
 YJL197w *UBP12* ubiquitin C-terminal hydrolase
 YGR048w *UFD1* ubiquitin fusion degradation protein
 YDL190c *UFD2* ubiquitin fusion degradation protein
 YBR058c *UBP14* ubiquitin specific protease
 YLR167w *UBI3* ubiquitin/ribosomal protein S27a
 YMR275c *BUL1* ubiquitination pathway protein

YDR054c *CDC34* ubiquitin-conjugating enzyme
 YMR022w *QR8* ubiquitin-conjugating enzyme
 YDR177w *UBC1* ubiquitin-conjugating enzyme
 YLR306w *UBC12* ubiquitin-conjugating enzyme
 YDR092w *UBC13* ubiquitin-conjugating enzyme
 YBR082c *UBC4* ubiquitin-conjugating enzyme
 YDR059c *UBC5* ubiquitin-conjugating enzyme
 YER100w *UBC6* ubiquitin-conjugating enzyme
 YDL064w *UBC9* ubiquitin-conjugating enzyme
 YDR139c ubiquitin-like protein
 YER125w *RSP5* ubiquitin-protein ligase
 YKL210w *UBA1* ubiquitin-protein ligase
 YGR184c *UBR1* ubiquitin-protein ligase
 YDR069c *DOA4* ubiquitin-specific protease
 YDL122w *UBP1* ubiquitin-specific protease
 YER144c *UBP5* ubiquitin-specific protease
 YJR099w *YUH1* ubiquitin-specific protease
 YOR124c *UBP2* ubiquitin-specific proteinase
 YER151c *UBP3* ubiquitin-specific proteinase

lysosomal and vacuolar degradation

YBR286w *APE3* aminopeptidase Y, vacuolar
 YKL103c *LAP4* aminopeptidase yscl, vacuolar
 YPL154c *PEP4* aspartyl protease
 YMR297w *PRC1* carboxypeptidase y, serine-type protease
 YHR028c *DAP2* dipeptidyl aminopeptidase B
 YJL172w *CP51* Gly-X carboxypeptidase YSCS
 YEL060c *PRB1* protease B, vacuolar
 YOR003w *YSP3* subtilisin-like protease III

other subcellular degradation

YDR144c *MKC7* aspartyl protease of the periplasmic space
 YBL022c *PIM1* ATP-dependent protease, mitochondrial
 YJL015w *BAR1* barrierpepsin
 YBR201w *DER1* involved in degradation proteins in the ER
 YER017c *AFG3* protease of the SEC18/CDC48/PAS1 family of ATPases (AAA)
 YPR024w *YME1* protease of the SEC18/CDC48/PAS1 family of ATPases (AAA)
 YMR089c *YTA12* protease of the SEC18/CDC48/PAS1 family of ATPases (AAA)
 YCL057w *PRD1* proteinase yscD
 YHR113w similarity to vacuolar aminopeptidase Lap4p/Ape1p
 YFR006w similarity to X-Pro dipeptidases

other protein-destination activities

YDR258c *HSP78* heat-shock protein of clpb family of ATP-dependent proteases, mitochondrial
 YDL104c *QR17* similarity to *H. influenzae* sialoglycoprotease (gcp)
 YDR415c strong similarity to bacterial leucyl aminopeptidase

Transport facilitation

ion channels

YGR217w *CCH1* calcium channel protein
 YLL052c member of mip family transmembrane channels
 YJL093c *TOK1* outward-rectifier potassium channel
 YPR192w similarity to plasma membrane and water channel proteins
 YLL053c similarity to water channel proteins
 YJL114c *POR2* voltage-dependent anion channel (YVDAC2)
 YJR040w *GEF1* voltage-gated chloride channel protein

ion transporters

YNR055c *HOL1* member of major facilitator superfamily multidrug-resistance protein subfamily 1
 YJL048w similarity to amino-phospholipids-ATPase Drs2p

metal ion transporters (Cu, Fe, etc.)

YOR316c *COT1* cobalt accumulation protein
 YPR124w *CTR1* copper transport protein
 YHR175w *CTR2* copper transport protein
 YLR411w *CTR3* copper transport protein
 YGL255w *ZRT1* high-affinity zinc transport protein
 YLR130c *ZRT2* low-affinity zinc transporter
 YMR319c *FET4* low-affinity Fe(II) iron transport protein
 YOL122c *SMF1* manganese transporter
 YBR290w *BSD2* metal homeostasis protein
 YDR270w *CCO2* probable copper-transporting ATPase
 YBR295w *PCA1* P-type Cu²⁺-transporting ATPase
 YMR243c *ZRC1* zinc- and cadmium resistance protein

other cation transporters (Na, K, Ca, NH₄, etc.)

YGR121c *MEP1* ammonia permease of high capacity and moderate affinity
 YDL128w *VCX1* Ca²⁺-transport (H⁺/Ca²⁺ exchange) protein, vacuolar
 YGL006w *PMC1* Ca²⁺-transporting P-type ATPase
 YGL167c *PMR1* Ca²⁺-transporting P-type ATPase
 YER024c-a *PMP1* H⁺-ATPase subunit, plasma membrane
 YCL017c-a *PMP2* H⁺-ATPase subunit, plasma membrane
 YMR054w *STV1* H⁺-ATPase V0 domain 102K subunit, vacuolar
 YHR039c-a *VMA10* H⁺-ATPase V0 domain 13K subunit, vacuolar

YEL027w *CUP5* H⁺-ATPase V0 domain 17K subunit, vacuolar
 YPL234c *TFP3* H⁺-ATPase V0 domain 17K subunit, vacuolar
 YLR447c *VMA6* H⁺-ATPase V0 domain 36K subunit, vacuolar
 YOR270c *VPH1* H⁺-ATPase V0 domain 95K subunit, vacuolar
 YGR020c *VMA7* H⁺-ATPase V1 domain 14K subunit, vacuolar
 YOR332w *VMA4* H⁺-ATPase V1 domain 27K subunit, vacuolar
 YEL051w *VMA8* H⁺-ATPase V1 domain 32K subunit, vacuolar
 YKL080w *VMA5* H⁺-ATPase V1 domain 42K subunit, vacuolar
 YPR036w *VMA13* H⁺-ATPase V1 domain 54K subunit, vacuolar
 YBR127c *VMA2* H⁺-ATPase V1 domain 60K subunit, vacuolar
 YDL185w *TFP1* H⁺-ATPase V1 domain 69K subunit, vacuolar
 YGL008c *PMA1* H⁺-transporting P-type ATPase
 YPL038w *PMA2* H⁺-transporting P-type ATPase 2
 YNL142w *MEP2* high-affinity low-capacity ammonia permease
 YJL129c *TRK1* high-affinity potassium transport protein
 YAL026c *DRS2* membrane-spanning P-type amino-phospholipids-ATPase
 YKR050w *TRK2* moderate-affinity potassium transport protein
 YHR026w *PPA1* proteolipid protein of proton-transporting ATPase
 YEL031w *SPF1* P-type ATPase
 YLR138w *NHA1* putative Na⁺/H⁺ antiporter
 YBR235w similarity to bumetanide-sensitive Na⁺-K⁺-Cl⁻ cotransport protein
 YJL094c similarity to *E. hirae* Na⁺/H⁺-antiporter NapA
 YDR456w similarity to Na⁺/H⁺ antiporters
 YPR138c *MEP3* strong similarity to ammonium transport proteins
 YHL016c *DUR3* urea transport protein

anion transporters (Cl, SO₄, PO₄, etc)
 YLR348c dicarboxylate carrier protein
 YML123c *PHO84* high-affinity inorganic phosphate/H⁺ symporter
 YBR294w *SUL1* high-affinity sulphate transport protein
 YJL117w *PHO86* inorganic phosphate transporter
 YCR037c *PHO87* member of the phosphate permease family
 YJR077c *MIR1* phosphate transport protein, mitochondrial (MCF)
 YBR235w similarity to bumetanide-sensitive Na⁺-K⁺-Cl⁻ cotransport protein
 YJL094c similarity to *E. hirae* Na⁺/H⁺-antiporter NapA
 YNR013c similarity to membrane protein Pho87p and hypothetical protein YJL198w
 YCR098c *GIT1* similarity to phosphate transporter proteins
 YPR003c similarity to sulphate transporter proteins
 YER053c strong similarity to mitochondrial phosphate carrier protein
 YJL198w strong similarity to Pho87p
 YBR296c strong similarity to phosphate-repressible phosphate permease
 YLR092w *SEL2* strong similarity to Sul1p

sugar and carbohydrate transporters
 YPR021c similarity to human citrate transporter protein
 YKL217w *JEN1* carboxylic-acid transporter protein
 YBR291c *CTP1* citrate transport protein, mitochondrial (MCF)
 YLR348c dicarboxylate carrier protein
 YLR081w *GAL2* galactose (and glucose) permease
 YGR289c *AGT1* general α-glucoside permease
 YLL043w *FPS1* glycerol channel protein
 YNL318c *HXT14* hexose transport protein
 YJL214w *HXT8* hexose transport protein
 YJL219w *HXT9* hexose transport protein
 YFL011w *HXT10* hexose transporter
 YDL194w *SNF3* high-affinity glucose transporter
 YEL069c *HXT13* high-affinity hexose transporter
 YMR011w *HXT2* high-affinity hexose transporter
 YDR343c *HXT6* high-affinity hexose transporter
 YDR342c *HXT7* high-affinity hexose transporter
 YML123c *PHO84* high-affinity inorganic phosphate/H⁺ symporter
 YOL156w *HXT11* low-affinity glucose transporter
 YHR094c *HXT1* low-affinity hexose transporter
 YDR345c *HXT3* low-affinity hexose transporter
 YDR497c *ITR1* major myo-inositol permease
 YBR298c *MAL31* maltose permease
 YDL198c *YHM1* member of the mitochondrial carrier family (MCF)
 YHR092c *HXT4* moderate- to low-affinity glucose transporter
 YOL103w *ITR2* myo-inositol transport protein
 YBR241c similarity to glucose transport proteins
 YGL104c similarity to glucose transport proteins
 YDR387c similarity to Itr1p and Itr2p
 YFR045w similarity to mitochondrial citrate transport proteins

YCR098c *GIT1* similarity to phosphate transporter proteins
 YDL199c similarity to sugar transporter proteins
 YFL040w similarity to yeast glucose transport proteins
 YHR096c *HXT5* strong similarity to hexose transporters
 YDL245c *HXT15* strong similarity to Hxt17p and Hxt7p
 YJR160c strong similarity to Mal3Tp
 YDR536w *STL1* strong similarity to members of the sugar permease family
 YOR271c strong similarity to *Rattus* tricarboxylate carrier
 YDL247w strong similarity to sugar transport proteins
 YIL171w strong similarity to sugar transport proteins
 YIL170w *HXT12* strong similarity to sugar transport proteins
 YJR158w *HXT16* strong similarity to sugar transport proteins
 YNR072w *HXT17* sugar transport protein
 YDL139w *RG72* suppressor of *snf3* mutant

amino-acid transporters
 YBR068c *BAP2* amino-acid permease
 YEL063c *CAN1* amino-acid permease
 YBR069c *VAP1* amino-acid permease
 YCL025c *AGP1* asparagine and glutamine permease
 YGL077c *HNM1* choline permease
 YPL265w *DIP5* dicarboxylic amino-acid permease
 YDL210w *UGA4* GABA-specific high-affinity permease
 YKR039w *GAP1* general amino-acid permease
 YDR508c *GNP1* high-affinity glutamine permease
 YGR055w *MUP1* high-affinity methionine permease
 YNL270c *ALP1* high-affinity permease for basic amino acids
 YOL020w *SCM2* high-affinity tryptophan transport protein
 YGR191w *HIP1* histidine permease
 YNL268w *LYP1* lysine-specific high-affinity permease
 YOR130c *ARG11* member of the mitochondrial carrier family (MCF)
 YHL036w *MUP3* methionine permease
 YOR348c *PUT4* proline and γ-aminobutyrate permease
 YKL174c similarity to choline transport protein
 YNR056c similarity to choline transport protein
 YFL055w similarity to Gap1p and other amino-acid permeases
 YDR160w similarity to lysine transport protein LYP1
 YLL061w strong similarity to amino-acid transport protein Gap1p
 YDR046c (*PAP1*) strong similarity to amino-acid transport proteins
 YBR132c strong similarity to amino-acid permeases
 YPL274w strong similarity to amino-acid transport proteins

lipid transporters
 YBR041w *FAT1* fatty-acid transporter
 YKL188c *PAT1* long-chain-fatty-acid transporter
 YPL147w *PXA1* long-chain-fatty-acid transporter
 YKL174c similarity to choline transport protein
 YNR056c similarity to choline transport protein
 YHR123w *EPT1* sn-1,2-diacylglycerol ethanalamine- and cholinephosphotransferase

purine and pyrimidine transporters
 YMR056c *AAC1* ADP/ATP carrier protein (MCF)
 YBL030c *AAC2* ADP/ATP carrier protein (MCF)
 YBR085w *AAC3* ADP/ATP carrier protein (MCF)
 YBR192w *RIM2* mitochondrial carrier protein (MCF)
 YER056c *FCY2* purine-cytosine permease
 YOR060w purine-cytosine permease
 YOR222w similarity to ADP/ATP carrier proteins
 YPL134c similarity to ADP/ATP carrier proteins
 YPR011c similarity to ADP/ATP carrier proteins and Graves disease carrier protein
 YOR071c similarity to allantoin or uracil transport proteins
 YOR192c similarity to allantoin or uracil transport proteins
 YGR096w similarity to bovine Graves disease carrier protein
 YHR002w similarity to bovine mitochondrial carrier protein/Grave's disease carrier protein
 YGL186c similarity to hypothetical protein YER060w and weak similarity to FCY2 protein
 YBL042c strong similarity to allantoin and uracil transport proteins
 YER060w-a *FCY22* strong similarity to Fcy2p
 YBR021w *FUR4* uracil permease

allantoin and allantoate transporters
 YJR152w *DAL5* allantoate permease
 YIR028w *DAL4* allantoin permease
 YIL166c similarity to allantoin permease
 YCR028c *FEN2* similarity to allantoate permease transporter
 YGR260w similarity to allantoate transport protein
 YLR004c similarity to allantoate transport protein

YOR071c similarity to allantoin or uracil transport proteins
 YOR192c similarity to allantoin or uracil transport proteins
 YLR237w similarity to allantoin transport protein
 YLL056w similarity to Dal5p
 YBL042c strong similarity to allantoin and uracil transport proteins
 YAL067c *SEO1* suppressor of sulphoxyde ethionine resistance

transport ATPases
 YGL006w *PMC1* Ca²⁺-transporting P-type ATPase
 YGL167c *PMR1* Ca²⁺-transporting P-type ATPase
 YKL016c *ATP7* F1F0-ATPase complex, F0 D subunit
 YBL099w *ATP1* F1F0-ATPase complex, F1 α subunit
 YJR121w *ATP2* F1F0-ATPase complex, F1 β subunit
 YDL004w *ATP16* F1F0-ATPase complex, F1 δ subunit
 YPL078c *ATP4* F1F0-ATPase complex, F1 ε subunit
 YPL271w *ATP15* F1F0-ATPase complex, F1 ζ subunit
 YBR039w *ATP3* F1F0-ATPase complex, F1 η subunit
 YDR298c *ATP5* F1F0-ATPase complex, OSCP subunit
 YLR295c *ATP14* F1F0-ATPase complex, subunit h
 YCR024c-a *PMP1* H⁺-ATPase subunit, plasma membrane
 YEL017c-a *PMP2* H⁺-ATPase subunit, plasma membrane
 YMR054w *STV1* H⁺-ATPase V0 domain 102K subunit, vacuolar
 YHR039c-a *VMA10* H⁺-ATPase V0 domain 13K subunit, vacuolar
 YEL027w *CUP5* H⁺-ATPase V0 domain 17K subunit, vacuolar
 YPL234c *TFP3* H⁺-ATPase V0 domain 17K subunit, vacuolar
 YLR447c *VMA6* H⁺-ATPase V0 domain 36K subunit, vacuolar
 YOR270c *VPH1* H⁺-ATPase V0 domain 95K subunit, vacuolar
 YGR020c *VMA7* H⁺-ATPase V1 domain 14K subunit, vacuolar
 YOR332w *VMA4* H⁺-ATPase V1 domain 27K subunit, vacuolar
 YEL051w *VMA8* H⁺-ATPase V1 domain 32K subunit, vacuolar
 YKL080w *VMA5* H⁺-ATPase V1 domain 42K subunit, vacuolar
 YPR036w *VMA13* H⁺-ATPase V1 domain 54K subunit, vacuolar
 YBR127c *VMA2* H⁺-ATPase V1 domain 60K subunit, vacuolar
 YDL185w *TFP1* H⁺-ATPase V1 domain 69K subunit, vacuolar
 YGL008c *PMA1* H⁺-transporting P-type ATPase
 YPL036w *PMA2* H⁺-transporting P-type ATPase 2
 YAL026c *DRS2* membrane-spanning P-type amino-phospholipids-ATPase
 YDR270w *CCC2* probable copper-transporting ATPase
 YHR026w *PPA1* proteolipid protein of proton-transporting ATPase
 YEL031w *SPF1* P-type ATPase
 YDR040c *ENA1* P-type ATPase involved in Na⁺ and Li⁺ efflux
 YDR039c *ENA2* P-type ATPase involved in Na⁺ efflux
 YDR038c *ENA5* P-type ATPase involved in Na⁺ efflux
 YBR295w *PCA1* P-type Cu²⁺-transporting ATPase
 YLR048w similarity to amino-phospholipids-ATPase Drs2p
 YER166w similarity to ATPase *P. falciparum* ATPase 2
 YMR162c similarity to ATPases
 YDR093w similarity to *P. falciparum* ATPase 2

ABC transporters
 YDR406w *PDR15* ATP-binding cassette protein family member
 YLR188w *MDL1* ATP-binding cassette transporter family member
 YPL270w *MDL2* ATP-binding cassette transporter family member
 YKL209c *STE6* ATP-binding cassette transporter protein
 YGR281w *YOR1* ATP-binding cassette transporter protein required for oligomycin resistance
 YMR301c *ATM1* ATP-binding cassette transporter protein, mitochondrial
 YCR011c *ADP1* ATP-dependent permease
 YDR135c *YCF1* glutathione S-conjugate transporter, vacuolar
 YKL188c *PAT1* long-chain-fatty-acid transporter
 YPL147w *PXA1* long-chain-fatty-acid transporter
 YDR011w *SNQ2* multidrug resistance protein
 YPL058c *PDR12* multidrug resistance transporter
 YOR153w *PDR5* pleiotropic drug resistance protein
 YLR075c similarity to *A. gambiae* ATP-binding-cassette protein
 YER036c similarity to members of the ABC transporter family
 YLL015w similarity to metal resistance proteins
 YHL035c similarity to multidrug resistance proteins
 YKR103w similarity to multidrug resistance proteins
 YKR104w similarity to multidrug resistance proteins
 YLL048c similarity to rat organic anion transporter
 YIL013c *PDR11* similarity to Snq2p and other ATP-dependent permeases
 YOR328w *PDR10* strong similarity to ABC transporter proteins
 YOR311w strong similarity to ATP-dependent permeases

YDR091c strong similarity to human RNase L inhibitor and to *M. jannaschii* ABC transporter

YNR070w strong similarity to Snq2p

drug transporters

YML116w *ATR1* aminotriazole and 4-nitroquinoline resistance protein

YGR281w *YOR1* ATP-binding cassette transporter protein required for oligomycin resistance

YGR197c *SNG1* involved in nitroguanidine resistance

YEL065w probably multidrug resistance protein

YMR123w *PKR1* resistance against *Pichia farinosa* killer toxin (SMK toxin) when expressed by a multicopy plasmid

YMR088c similarity multidrug resistance proteins

YDR119w similarity to *B. subtilis* tetracycline resistance

YBR293w similarity to multidrug resistance proteins

YGR138c similarity to multidrug resistance proteins

YHR048w similarity to multidrug resistance proteins

YPR156c similarity to multidrug resistance proteins

YNL065w similarity to resistance proteins

YIL013c *PDR11* similarity to Snq2p and other ATP-dependent permeases

YMR279c strong similarity to aminotriazole resistance protein

YGR224w strong similarity to drug resistance protein SGE1

YBR052c strong similarity to *S. pombe* brefeldin A resistance protein obr1

YAL067c *SEO1* suppressor of sulphoxyde ethionine resistance

other transport-facilitators

YFL028c *CAF16* ATP-binding cassette transporter protein family member

YER019c-a *SEB2* ER protein-translocation complex subunit

YLR378c *SEC61* ER protein-translocation complex subunit

YPL094c *SEC62* ER protein-translocation complex subunit

YOR254c *SEC63* ER protein-translocation complex subunit

YBR171w *SEC66* ER protein-translocation complex subunit

YLR292c *SEC72* ER protein-translocation complex subunit

YDR086c *SSS1* ER protein-translocation complex subunit

YIL134w *FLX1* FAD carrier protein, mitochondrial (MCF)

YCR023c member of major facilitator superfamily

YKL039w *PTM1* multidrug-resistance protein family 2 member of the major facilitator superfamily

YPR058w *YMC1* mitochondrial carrier protein (MCF)

YNR017w *MAS6* mitochondrial inner membrane import translocase subunit

YIL143w *TIM17* mitochondrial inner membrane import translocase subunit

YIL022w *TIM44* mitochondrial inner membrane import translocase subunit

YNL131w *TOM22* mitochondrial outer membrane import receptor subunit

YGR082w *TOM20* mitochondrial outer membrane import receptor subunit, 20K

YMR060c *TOM37* mitochondrial outer membrane import receptor subunit, 37K

YMR203w *TOM40* mitochondrial outer membrane import receptor subunit, 40K

YOR045w *TOM6* mitochondrial outer membrane import receptor subunit, 6K

YNL070w *TOM7* mitochondrial outer membrane import receptor subunit, 7K

YNL055c *POR1* mitochondrial outer membrane porin

YNL121c *TOM70* mitochondrial outer membrane specialized import receptor

YAR035w *YAT1* outer carnitine acetyltransferase, mitochondrial

YKR093w *PTR2* peptide transporter

YIL120w similarity to antibiotic resistance proteins

YIL121w similarity to antibiotic resistance proteins

YCL038c similarity to bacterial membrane transporter

YBR043c similarity to benomyl/methotrexate resistance protein

YHL040c similarity to *C. carbonum* toxin pump

YHL047c similarity to *C. carbonum* toxin pump

YER024w similarity to carnitine O-acetyltransferase Yat1p

YOR291w similarity to cation translocating ATPases

YFL054c similarity to channel proteins

YBR180w similarity to drug resistance proteins

YIL006w similarity to Flx1p

YOR306c similarity to human X-linked PEST-containing transporter

YIL056w similarity to hypothetical protein YER064c

YNL003c *PET8* similarity to mitochondrial rat tricarboxylate transport protein

YOL119c similarity to monocarboxylate transporter proteins

YGR065c similarity to *P. putida* phthalate transporter

YOL163w similarity to *P. putida* phthalate transporter

YEL006w similarity to peroxisomal membrane and mitochondrial carrier proteins

YJR124c similarity to *Staphylococcus* multidrug resistance protein

YNL125c similarity to YKL221w and human X-linked PEST-containing transporter

YBR008c strong similarity to benomyl/methotrexate resistance protein

YCL069w strong similarity to drug resistance protein SGE1

YER087c-a *SEB1* strong similarity to mammalian Sec61 β subunit

YKR067w strong similarity to Sct1p

YBL011w *SC11* suppresses a choline-transport mutant

YBL089w weak similarity to *A. thaliana* aminoacid permease AAP3

YIL088c weak similarity to *A. thaliana* aminoacid permease AAP4

YJR001w weak similarity to *A. thaliana* aminoacid permease AAP4

YER119c weak similarity to *E. herbicola* tyrosine permease

YKL221w weak similarity to human X-linked PEST-containing transporter

YEL064c weak similarity to members of the major facilitator superfamily

YJR106w weak similarity to Na⁺/H⁺ antiporter

YDL206w weak similarity to transporter proteins

YKL146w weak similarity to transporter proteins

Intracellular transport

nuclear transport

YIL075c *SEN3* 26S proteasome regulatory subunit

YOR048c *RA11* 5'-3' exoribonuclease

YGL097w *SRM1* GDP/GTP exchange factor for Gsp1p/Gsp2p

YOR185c *GSP2* GTP-binding protein

YLR293c *GSP1* GTP-binding protein of the RAS superfamily

YAL005c *SSA1* heat-shock protein of HSP70 family

YOR160w *MTR10* involved in mRNA transport

YIL050w *MTR4* involved in nucleocytoplasmic transport of mRNA

YBR017c *KAP104* karyopherin

YNL189w *SRP1* karyopherin- α or importin

YLR347c *KAP95* karyopherin- β

YKL186c *MTR2* mRNA transport protein

YPL124w *NIP29* nuclear import protein

YPL174c *NIP80* nuclear import protein

YMR129w *POM152* nuclear pore membrane glycoprotein

YFR002w *NIC96* nuclear pore protein

YIL041w *NSP1* nuclear pore protein

YOR098c *NUP1* nuclear pore protein

YKL068w *NUP100* nuclear pore protein

YMR047c *NUP116* nuclear pore protein

YKL057c *NUP120* nuclear pore protein

YKR082w *NUP133* nuclear pore protein

YGL092w *NUP145* nuclear pore protein

YER105c *NUP157* nuclear pore protein

YIL115c *NUP159* nuclear pore protein

YML079w *NUP170* nuclear pore protein

YML103c *NUP188* nuclear pore protein

YLR335w *NUP2* nuclear pore protein

YDR192c *NUP42* nuclear pore protein

YGL172w *NUP49* nuclear pore protein

YGR119c *NUP57* nuclear pore protein

YIL061w *NUP82* nuclear pore protein

YDL116w *NUP84* nuclear pore protein

YJR042w *NUP85* nuclear pore protein

YGL100w *SEH1* nuclear pore protein

YBR170c *NPL4* nuclear protein localization factor and ER translocation component

YER009w *NTF2* nuclear transport factor

YDR432w *NPL3* nucleolar protein

YER002w *YRB1* ran-specific GTPase-activating protein

YER107c *GLE2* required for nuclear pore complex structure and function

YIR011c *STS1* required for transport of Rna15p from the cytoplasm to the nucleus

YDL207w *GLE1* RNA export mediator

YIL063c *YRB2* similarity to Yrp1p and Nup2p

YLR119w *SRN2* suppressor of *mat1-1* mutation

YFL049w weak similarity to Npl6p

mitochondrial transport

YMR056c *AAC1* ADP/ATP carrier protein (MCF)

YBL030c *AAC2* ADP/ATP carrier protein (MCF)

YBR085w *AAC3* ADP/ATP carrier protein (MCF)

YMR301c *ATM1* ATP-binding cassette transporter protein, mitochondrial

YML042w *CAT2* carnitine O-acetyltransferase

YBR291c *CTP1* citrate transport protein, mitochondrial (MCF)

YOR316c *COT1* cobalt accumulation protein

YOR037w *CYC2* cytochrome c mitochondrial import factor

YLR348c dicarboxylate carrier protein

YKL016c *ATP7* F1F0-ATPase complex, F0 D subunit

YBL099w *ATP1* F1F0-ATPase complex, F1 α subunit

YJR121w *ATP2* F1F0-ATPase complex, F1 β subunit

YDL004w *ATP16* F1F0-ATPase complex, F1 δ subunit

YPL078c *ATP4* F1F0-ATPase complex, F1 δ subunit

YPL271w *ATP15* F1F0-ATPase complex, F1 ϵ subunit

YBR039w *ATP3* F1F0-ATPase complex, F1 γ subunit

YDR298c *ATP5* F1F0-ATPase complex, OSCP subunit

YLR295c *ATP14* F1F0-ATPase complex, subunit h

YIL134w *FLX1* FAD carrier protein, mitochondrial (MCF)

YOR232w *MGE1* heat-shock protein - chaperone

YLL024c *SSA2* heat-shock protein of HSP70 family, cytosolic

YGR028w *MSP1* intra-mitochondrial sorting protein

YOR130c *ARG11* member of the mitochondrial carrier family (MCF)

YDL198c *YHM1* member of the mitochondrial carrier family (MCF)

YNL064c *YDJ1* mitochondrial and ER import protein

YBR192w *RIM2* mitochondrial carrier protein (MCF)

YPR058w *YMC1* mitochondrial carrier protein (MCF)

YBR104w *YMC2* mitochondrial carrier protein (MCF)

YML062c *MFT1* mitochondrial fusion target protein

YNR017w *MAS6* mitochondrial inner membrane import translocase subunit

YIL143w *TIM17* mitochondrial inner membrane import translocase subunit

YIL022w *TIM44* mitochondrial inner membrane import translocase subunit

YNL131w *TOM22* mitochondrial outer membrane import receptor subunit

YGR082w *TOM20* mitochondrial outer membrane import receptor subunit, 20K

YMR060c *TOM37* mitochondrial outer membrane import receptor subunit, 37K

YMR203w *TOM40* mitochondrial outer membrane import receptor subunit, 40K

YOR045w *TOM6* mitochondrial outer membrane import receptor subunit, 6K

YNL070w *TOM7* mitochondrial outer membrane import receptor subunit, 7K

YNL055c *POR1* mitochondrial outer membrane porin

YNL121c *TOM70* mitochondrial outer membrane specialized import receptor

YAR035w *YAT1* outer carnitine acetyltransferase, mitochondrial

YJR077c *MIR1* phosphate transport protein, mitochondrial (MCF)

YBR091c *MRS5* regulator of mitochondrial intron splicing

YPL134c similarity to ADP/ATP carrier proteins

YER024w similarity to carnitine O-acetyltransferase Yat1p

YPR021c similarity to human citrate transporter protein

YER053c strong similarity to mitochondrial phosphate carrier protein

YLR034c strong similarity to SMF2 protein

YHR117w *TOM71* strong similarity to Tom70p/Mas70p

YHR050w *SMF2* suppressor of mitochondrial matrix mutant

YIL114c *POR2* voltage dependent anion channel (YVDAC2)

vesicular transport (Golgi network, etc.)

YBR164c *ARL1* ADP-ribosylation factor

YOR094w *ARF3* ADP-ribosylation factor 3

YGL167c *PMR1* Ca²⁺-transporting P-type ATPase

YGR167w *CLC1* clathrin light chain

YOL062c *APM4* clathrin-associate protein YAP54

YPL259c *APM1* clathrin-associated protein

YBR288c *APM3* clathrin-associated protein complex, medium subunit

YPL010w *RET3* coatamer complex ζ subunit

YDL145c *RET1* coatamer complex α subunit of secretory pathway vesicles

YGL137w *SEC27* coatamer complex β' subunit (β' -cop) of secretory pathway vesicles

YDR238c *SEC26* coatamer complex β subunit of secretory pathway vesicles

YFR051c *RET2* coatamer complex δ subunit

YNL287w *SEC21* coatamer complex γ subunit (γ -COP) of secretory pathway vesicles

YDR170c *SEC7* component of non-clathrin vesicle coat

YPR181c *SEC23* component of the COPII coat of ER-Golgi vesicles

YDL195w *SEC31* component of the COPII coat of ER-Golgi vesicles

YGL200c *EMP24* component of the COPII-coated vesicles, 24K

YML012w *ERV25* component of the COPII-coated vesicles, 25K

YMR017w *DBI9* Dbl2p interacting protein

YKR054c *DYN1* dynein heavy chain, cytosolic

YLR083c *EMP70* endosomal protein

YBL040c *ERD2* ER lumen protein-retaining receptor

YJR031c *GEA1* GDP/GTP exchange factor for ARF

YEL022w *GEA2* GDP/GTP exchange factor for ARF

YNR026c *SEC12* GDP/GTP exchange factor for Sar1p

YPR017c *DSS4* GDP/GTP exchange factor for Sec4p

YDL192w *ARF1* GTP-binding protein of the ARF family

YDL137w *ARF2* GTP-binding protein of the ARF family

YPL218w *SAR1* GTP-binding protein of the ARF family

YFL038c *YPT71* GTP-binding protein of the RAB family

YER031c *YPT31* GTP-binding protein of the RAB family

YLR262c *YPT6* GTP-binding protein of the RAB family

YML001w *YPT7* GTP-binding protein of the RAB family

YFL005w *SEC4* GTP-binding protein of the RAS superfamily

YLL024c *SSA2* heat-shock protein of HSP70 family, cytosolic

YLR268w *SEC22* high copy suppressor of *ypt1* null mutation

YDR189w *SLY1* hydrophilic suppressor of *ypt1* and member of the Sec1p family

YDL058w *USO1* intracellular protein transport protein involved in ergosterol biosynthesis

YPL145c *KES1* involved in targeting and fusion of ER to Golgi transport vesicles

YKR068c *BET3* involved in vesicle transport from Golgi to plasma membrane

YMR183c *SSO2* mitochondrial and ER import protein

YNL064c *YDJ1* mitochondrial and ER import protein

YIL004c *SYS1* multicopy suppressor of *ypt6*
 YPL085w *SEC16* multidomain vesicle coat protein
 YOR326w *MYO2* myosin heavy chain
 YFL025c *BST1* negative regulator of COPII vesicle formation
 YMR079w *SEC14* phosphatidylinositol/phosphatidylcholine transfer protein
 YCR067c *SED4* protein of the endoplasmic reticulum
 YIL004c *BET1* protein transport protein
 YLR208w *SEC13* protein transport protein
 YNL272c *SEC2* protein transport protein
 YGR009c *SEC9* protein transport protein
 YGL145w *TIP20* required for ER to Golgi transport
 YDR498c *SEC20* secretory pathway protein
 YBR097w *VPS15* ser/thr protein kinase
 YBR264c similarity to GTP-binding proteins
 YKL196c similarity to Sec22p
 YIL193w similarity to Sly41p
 YGL210w *YPT32* small GTP-binding protein
 YKL006c-a *SFT1* SNARE-like protein
 YPL051w strong similarity to ADP-ribosylation factors
 YDR107c strong similarity to EMP70 protein
 YAL030w *SNC1* strong similarity to synaptic vesicle-associated membrane protein
 YOR327c *SNC2* strong similarity to synaptobrevin
 YER039c strong similarity to vanadate resistance protein Van2p
 YLR026c *SED5* syntaxin (T-SNARE)
 YOR075w *UFE1* syntaxin (T-SNARE) of the ER
 YPL232w *SSO1* syntaxin-related protein
 YBL050w *SEC17* transport vesicle fusion protein
 YLR396c *VPS33* vacuolar sorting protein
 YML097c *VPS9* vacuolar sorting protein
 YGL225w *GOG5* vanadate-resistance protein
 YGL233w *SEC15* vesicular traffic control protein
 YBR080c *SEC18* vesicular-fusion protein, functional homologue of NSF
 YLR093c weak similarity to synaptobrevin
 YLR078c *BOS1* weak similarity to synaptobrevin (V-SNARE)

peroxisomal transport

YML042w *CAT2* carnitine O-acetyltransferase
 YKL188c *PAT1* long-chain-fatty-acid transporter
 YPL147w *PXA1* long-chain-fatty-acid transporter
 YER015w *FAA2* long-chain-fatty-acid-CoA ligase
 YDR329c *PAS3* peroxisomal assembly protein
 YIL210w *PAS5* peroxisomal assembly protein
 YDR142c *PAS7* peroxisomal import protein
 YLR191w *PAS20* peroxisomal protein involved in protein import
 YDR244w *PAS10* putative peroxisomal targeting signal receptor

vacuolar transport

YDL128w *VCX1* Ca²⁺-transport (H⁺/Ca²⁺ exchange) protein, vacuolar
 YGL006w *PMC1* Ca²⁺-transporting P-type ATPase
 YDR135c *YCF1* glutathione S-conjugate transporter, vacuolar
 YOR089c *VPS21* GTP-binding protein
 YER031c *YPT31* GTP-binding protein of the RAB family
 YKR014c *YPT52* GTP-binding protein of the RAB family
 YNL093w *YPT53* GTP-binding protein of the RAB family (RAS superfamily)
 YMR054w *STV1* H⁺-ATPase V0 domain 102K subunit, vacuolar
 YHR039c-a *VMA10* H⁺-ATPase V0 domain 13K subunit, vacuolar
 YEL027w *CUP5* H⁺-ATPase V0 domain 17K subunit, vacuolar
 YPL234c *TFP3* H⁺-ATPase V0 domain 17K subunit, vacuolar
 YLR447c *VMA6* H⁺-ATPase V0 domain 36K subunit, vacuolar
 YOR270c *VPH1* H⁺-ATPase V0 domain 95K subunit, vacuolar
 YGR020c *VMA7* H⁺-ATPase V1 domain 14K subunit, vacuolar
 YOR332w *VMA4* H⁺-ATPase V1 domain 27K subunit, vacuolar
 YEL051w *VMA8* H⁺-ATPase V1 domain 32K subunit, vacuolar
 YKL080w *VMA5* H⁺-ATPase V1 domain 42K subunit, vacuolar
 YPR036w *VMA13* H⁺-ATPase V1 domain 54K subunit, vacuolar
 YBR127c *VMA2* H⁺-ATPase V1 domain 60K subunit, vacuolar
 YDL185w *TFP1* H⁺-ATPase V1 domain 69K subunit, vacuolar
 YPL066w *VPS28* involved in vacuolar traffic
 YMR004w *MVP1* required for vacuolar protein sorting
 YBR097w *VPS15* ser/thr protein kinase
 YIL222w strong similarity to Pep1p
 YOR036w *PEP12* syntaxin (T-SNARE)
 YNR006w *VPS27* vacuolar protein sorting-associated protein
 YGL095c *VPS45* vacuolar protein sorting-associated protein
 YDR323c *PEP7* vacuolar segregation protein
 YPL045w *VPS16* vacuolar sorting protein
 YPR173c *VPS4* vacuolar sorting protein
 YAL002w *VPS8* vacuolar sorting protein
 YML097c *VPS9* vacuolar sorting protein

extracellular transport

YIL085w *EXO70* 70K exocyst component protein
 YDR129c *SAC6* actin filament bundling protein, fimbrin
 YKL209c *STE6* ATP-binding cassette transporter protein
 YDL226c *GCS1* cell proliferation zinc-finger protein
 YER136w *GDI1* GDP dissociation inhibitor
 YLL043w *FPS1* glycerol channel protein
 YHR044c *GYP6* GTPase-activating protein
 YLR262c *YPT6* GTP-binding protein of the RAB family
 YIL205c-a *NCE1* involved in non-classical protein export pathway
 YPR149w *NCE2* involved in non-classical protein export pathway
 YNL036w *NCE3* involved in non-classical protein export pathway
 YOR326w *MYO2* myosin heavy chain
 YJL093c *TOK1* outward-rectifier potassium channel
 YMR308c *PSE1* protein secretion enhancer
 YDR164c *SEC1* protein transport protein
 YNL272c *SEC2* protein transport protein
 YIL068c *SEC6* protein transport protein
 YPR055w *SEC8* protein transport protein
 YGR009c *SEC9* protein transport protein
 YKL212w *SAC1* recessive suppressor of secretory defect
 YLR166c *SEC10* required for exocytosis
 YDR166c *SEC5* required for exocytosis
 YNR049c *MSO1* secretion protein, multicopy suppressor of *sec1*
 YER008c *SEC3* secretory pathway protein
 YOR307c *SLY41* secretory pathway protein
 YBR162w-a *YSY6* secretory pathway protein
 YLR250w *SSP120* secretory protein
 YCR032w similarity to human CDC4L protein
 YER009w similarity to *P. polycyphalum* myosin-related protein mIpA
 YGR131w strong similarity to Nce2p
 YNL325c suppressor of *sac1* mutation
 YPL232w *SSO1* syntaxin-related protein
 YGL233w *SEC15* vesicular traffic control protein
 YBR080c *SEC18* vesicular-fusion protein, functional homologue of NSF

cellular import

YPL195w *YKS4* α- or γ- adaptin, large subunit of the clathrin-associated protein(AP) complex
 YBL037w *APL3* α-adaptin, large subunit of the clathrin-associated protein(AP) complex
 YFL039c *ACT1* actin
 YDR129c *SAC6* actin filament bundling protein, fimbrin
 YDL029w *ACT2* actin-like protein
 YGL040w *GLK1* aldohexose specific glucokinase
 YJR152w *DAL5* allantoate permease
 YIR028w *DAL4* allantoin permease
 YBR068c *BAP2* amino-acid permease
 YEL063c *CAN1* amino-acid permease
 YBR069c *VAP1* amino-acid permease
 YGR121c *MEP1* ammonia permease of high capacity and moderate affinity
 YKL135c *APL2* β-adaptin
 YGR261c *YKS5* β-adaptin, large subunit of the clathrin-associated protein(AP) complex
 YBR109c *CMD1* calmodulin
 YDL226c *GCS1* cell proliferation zinc-finger protein
 YMR058w *FET3* cell-surface ferroxidase
 YGL077c *HNM1* choline permease
 YGL206c *CHC1* clathrin heavy chain
 YGR167w *CLC1* clathrin light chain
 YLR170c *APS1* clathrin-associated protein (AP) complex, small subunit AP19
 YJR058c *APS2* clathrin-associated protein 17, small subunit
 YJR005w *YAP80* clathrin-associated protein complex, β subunit
 YJL024c *APS3* clathrin-associated protein(AP) complex, small subunit
 YPR124w *CTR1* copper transport protein
 YHR175w *CTR2* copper transport protein
 YNL243w *SLA2* cytoskeleton assembly control protein
 YLL001w *DNM1* dynamin-related protein
 YDL210w *UGA4* GABA-specific high-affinity permease
 YPR029c *APL4* γ-adaptin, large subunit of the clathrin-associated protein(AP) complex
 YLR081w *GAL2* galactose (and glucose) permease
 YNR026c *SEC12* GDP/GTP exchange factor for Sar1p
 YKR039w *GAP1* general amino-acid permease
 YLL043w *FPS1* glycerol channel protein
 YOR089c *VPS21* GTP-binding protein
 YER031c *YPT31* GTP-binding protein of the RAB family
 YKR014c *YPT52* GTP-binding protein of the RAB family
 YML001w *YPT7* GTP-binding protein of the RAB family
 YNL093w *YPT53* GTP-binding protein of the RAB family (RAS superfamily)
 YNL318c *HXT14* hexose transport protein
 YJL214w *HXT8* hexose transport protein
 YIL219w *HXT9* hexose transport protein
 YFL011w *HXT10* hexose transporter
 YDL194w *SNF3* high-affinity glucose transporter
 YEL069c *HXT13* high-affinity hexose transporter
 YMR011w *HXT2* high-affinity hexose transporter
 YDR343c *HXT6* high-affinity hexose transporter
 YDR342c *HXT7* high-affinity hexose transporter
 YML123c *PHO84* high-affinity inorganic phosphate/H⁺ symporter
 YNL142w *MEP2* high-affinity low capacity ammonia permease
 YGR055w *MUP1* high-affinity methionine permease

YNL270c *ALP1* high-affinity permease for basic amino acids
 YJL129c *TRK1* high-affinity potassium transport protein
 YBR294w *SUL1* high-affinity sulphate transport protein
 YOL020w *SCM2* high-affinity tryptophan transport protein
 YGR191w *HIP1* histidine permease
 YMR319c *FET4* low-affinity Fe(II) iron transport protein
 YOL156w *HXT11* low-affinity glucose transporter
 YHR094c *HXT1* low-affinity hexose transporter
 YDR345c *HXT3* low-affinity hexose transporter
 YNL268w *LYP1* lysine-specific high-affinity permease
 YDR497c *ITR1* major myo-inositol permease
 YBR298c *MAL31* maltose permease
 YOL122c *SMF1* manganese transporter
 YKR050w *TRK2* moderate-affinity potassium transport protein
 YHR092c *HXT4* moderate to low-affinity glucose transporter
 YOL103w *ITR2* myo-inositol transport protein
 YLR240w *VPS34* phosphatidylinositol 3-kinase
 YOR348c *PUT4* proline and γ-aminobutyrate permease
 YER056c *FCY2* purine-cytosine permease
 YLR088w *GAA1* required for attachment of GPI anchor onto proteins
 YNL084c *END3* required for endocytosis and cytoskeletal organization
 YJR090c *GRR1* required for glucose repression and for glucose and cation transport
 YCR009c *RVS161* similarity to human amphiphysin and Rvs167p
 YDR046c *(PAP1)* strong similarity to amino-acid transport proteins
 YHR096c *HXT5* strong similarity to hexose transporters
 YIL170w *HXT12* strong similarity to sugar transport proteins
 YJR158w *HXT16* strong similarity to sugar transport proteins
 YNR072w *HXT17* sugar transport protein
 YOR329c *SCD5* suppressor of clathrin deficiency
 YPR129w *SCD6* suppressor of clathrin deficiency
 YBR021w *FUR4* uracil permease
 YMR231w *PEP5* vacuolar biogenesis protein
 YLR337w *VRC1* verprolin
 YBR080c *SEC18* vesicular-fusion protein, functional homologue of NSF
 YJR040w *GEF1* voltage-gated chloride channel protein
 YMR243c *ZRC1* zinc- and cadmium resistance protein

other intracellular-transport activities

YKR054c *DYN1* dynein heavy chain, cytosolic
 YER019c-a *SEB2* ER protein-translocation complex subunit
 YLR378c *SEC61* ER protein-translocation complex subunit
 YPL094c *SEC62* ER protein-translocation complex subunit
 YOR254c *SEC63* ER protein-translocation complex subunit
 YBR171w *SEC66* ER protein-translocation complex subunit
 YLR292c *SEC72* ER protein-translocation complex subunit
 YDR086c *SSS1* ER protein-translocation complex subunit
 YCR075c *ERS1* intracellular protein transport protein
 YHL019c *APM2* involved in clathrin-dependent transport processes
 YJL034w *KAR2* nuclear fusion protein
 YBR170c *NPL4* nuclear protein localization factor and ER translocation component
 YKR093w *PTR2* peptide transporter
 YKL198c *PTK1* polyamine transport enhancing protein
 YDR040c *ENA1* P-type ATPase involved in Na⁺ and Li⁺ efflux
 YER060w *FCY21* purine-cytosine permease
 YNL183c *NPR1* ser/thr protein kinase
 YOR100c similarity to mitochondrial carrier protein YMC1
 YGL216w similarity to mouse kinesin-related protein KIF3
 YPR009w similarity to sterol uptake protein Sut1p
 YHR123w *EPT1* sn-1,2-diaclyglycerol ethanolamine- and cholinephosphotransferase
 YER060w-a *FCY22* strong similarity to Fcy2p
 YER087c-a *SEB1* strong similarity to mammalian Sec61β subunit

Cellular organization and biogenesis

organization and biogenesis of cell wall and plasma membrane

YLR342w *FKS1* 1,3—D-glucan synthase, catalytic subunit
 YGR032w *GSC2* 1,3—D-glucan synthase, subunit
 YOR362c *PRE10* 26S proteasome subunit C1
 YJR004c *SAG1* α-agglutinin
 YNR044w *AGA1* α-agglutinin anchor subunit
 YGL032c *AGA2* α-agglutinin binding subunit
 YDR077w *SED1* abundant cell-surface glycoprotein
 YJL005w *CYR1* adenylyate cyclase
 YCL007c *CWH36* affects the mannoprotein layer of the cell wall
 YOR335c *ALA1* alanyl-tRNA synthetase, cytosolic
 YPL061w *ALD6* aldehyde dehydrogenase, cytosolic
 YJR152w *DAL5* allantoin permease
 YIR028w *DAL4* allantoin permease
 YBR068c *BAP2* amino-acid permease
 YEL063c *CAN1* amino-acid permease
 YBR069c *VAP1* amino-acid permease

YDL195w *SEC31* component of the COPII coat of ER-Golgi vesicles
 YGL200c *EMP24* component of the COPII-coated vesicles, 24K
 YML012w *ERV25* component of the COPII-coated vesicles, 25K
 YHR007c *ERG11* cytochrome P450 lanosterol 14a-demethylase
 YNL130c *CPT1* diacylglycerol cholinephosphotransferase
 YMR013c *SEC59* dolichol kinase
 YPL227c *ALG5* dolichol-P-glucose synthetase
 YPR183w *DPM1* dolichyl-phosphate
 YJR143c *PMT4* β-D-mannosyltransferase
 YDL212w *SHR3* endoplasmic reticulum membrane protein
 YNL238w *KEX2* endoprotease of late Golgi compartment
 YBL040c *ERD2* ER lumen protein-retaining receptor
 YLR378c *SEC61* ER protein-translocation complex subunit
 YPL094c *SEC62* ER protein-translocation complex subunit
 YOR254c *SEC63* ER protein-translocation complex subunit
 YBR171w *SEC66* ER protein-translocation complex subunit
 YLR292c *SEC72* ER protein-translocation complex subunit
 YDR086c *SSS1* ER protein-translocation complex subunit
 YDR331w *GPI8* essential for GPI-anchor attachment
 YHR190w *ERG9* farnesyl-diphosphate farnesyltransferase
 YDR519w *FKB2* FK506/rapamycin-binding protein of the ER
 YJR031c *GEA1* GDP/GTP exchange factor for ARF
 YDEL022w *GEA2* GDP/GTP exchange factor for ARF
 YNR026c *SEC12* GDP/GTP exchange factor for Sar1p
 YPR159w *KRE6* glucan synthase subunit
 YGR143w *SKN1* glucan synthase subunit
 YOR067c *ALG8* glucosyltransferase
 YFL048c *EMP47* Golgi membrane protein
 YDL192w *ARF1* GTP-binding protein of the ARF family
 YPL218w *SAR1* GTP-binding protein of the ARF family
 YFL038c *YPT1* GTP-binding protein of the RAB family
 YLR262c *YPT6* GTP-binding protein of the RAB family
 YPR165w *RHO1* GTP-binding protein of the RHO subfamily of RAS-like proteins
 YNL090w *RHO2* GTP-binding protein of the RHO subfamily of RAS-like proteins
 YEL042w *GDA1* guanoinic diphosphatase
 YDR189w *SLY1* hydrophilic suppressor of *ypt1* and member of the Sec1p family
 YCR075c *ERS1* intracellular protein transport protein
 YDL058w *USO1* intracellular protein transport protein
 YLR220w *CCC1* involved in calcium regulation
 YBR201w *DER1* involved in degradation proteins in the ER
 YOR069w *VPS5* involved in Golgi retention and vacuolar sorting
 YOR336w *KRE5* killer toxin-resistance protein
 YHR072w *ERG7* lanosterol synthase
 YKR061w *KTR2* mannosyltransferase
 YDL095w *PMT1* mannosyltransferase
 YAL023c *PMT2* mannosyltransferase
 YOR321w *PMT3* mannosyltransferase
 YBL082c *RHK1* mannosyltransferase
 YLR139c *YUR1* mannosyltransferase
 YKR001c *VPS1* member of the dynamin family of GTPases
 YJR073c *OPI3* methylene-fatty-acyl-phospholipid synthase
 YNL064c *YDJ1* mitochondrial and ER import protein
 YPL085w *SEC16* multidomain vesicle coat protein
 YHR042w *NCP1* NADPH-cytochrome P450 reductase
 YFL025c *BST1* negative regulator of COPII vesicle formation
 YJL034w *KAR2* nuclear fusion protein
 YJL002c *OST1* oligosaccharyltransferase α subunit
 YEL002c *WBP1* oligosaccharyltransferase β subunit
 YMR149w *SWP1* oligosaccharyltransferase δ subunit
 YOR103c *OST2* oligosaccharyltransferase ε subunit
 YOR085w *OST3* oligosaccharyltransferase γ subunit
 YGL022w *STT3* oligosaccharyltransferase subunit
 YHR057c *CYP2* peptidyl-prolyl *cis-trans* isomerase
 YCR069w *SCC3* peptidyl-prolyl *cis-trans* isomerase
 YDR304c *CYP5* peptidyl-prolyl *cis-trans* isomerase D (cyclophilin D) of the ER
 YGR157w *CHO2* phosphatidylethanolamine N-methyltransferase
 YGR170w *PSD2* phosphatidylserine decarboxylase 2
 YDR518w *EUG1* protein disulphide isomerase
 YCL043c *PDI1* protein disulphide-isomerase
 YHR079c *IRE1* protein kinase
 YEL036c *ANP1* protein of the endoplasmic reticulum
 YOR067c *SED4* protein of the endoplasmic reticulum
 YIL004c *BET1* protein transport protein
 YDR040c *ENA1* P-type ATPase involved in Na⁺ and Li⁺ efflux
 YKL212w *SAC1* recessive suppressor of secretory defect
 YLR088w *GAA1* required for attachment of GPI anchor onto proteins
 YPL050c *MNN9* required for complex N-glycosylation
 YCL001w *RER1* required for correct localization of Sec12p
 YGL145w *TIP20* required for ER to Golgi transport
 YDR414c *ERD1* required for retention of luminal ER proteins
 YMR004w *MVP1* required for vacuolar protein sorting
 YDR498c *SEC20* secretory pathway protein
 YJR010c-a *SPC1* signal peptidase 10.8K subunit
 YML055w *SPC2* signal peptidase 18K subunit
 YDR292c *SRP101* signal recognition particle receptor, α subunit
 YIR022w *SEC11* signal sequence processing protein
 YAL058w *CNE1* similarity to calnexin

YDR245w *MNN10* similarity to *S. pombe* galactosyltransferase
 YMR214w *SCJ1* similarity to *E. coli* dnaJ
 YHR123w *EPT1* sn-1,2-diacylglycerol ethanolamine- and cholinephosphotransferase
 YKL006c-a *SFT1* SNARE-like protein
 YGR175c *ERG1* squalene monoxygenase
 YGL055w *OLE1* stearoyl-CoA desaturase
 YLR080w strong similarity to Emp47p
 YER087c-a *SEB1* strong similarity to mammalian Sec61β subunit
 YOR099w *KTR1* strong similarity to mannosyltransferases
 YGL226c-a *OST5* subunit of N-oligosaccharyltransferase, ζ subunit
 YDR297w *SUR2* suppressor of *rvs161* and *rvs167* mutations
 YBL102w *SFT2* suppressor of *sed5* ts mutants
 YLR026c *SED5* syntaxin (T-SNARE)
 YOR075w *UFE1* syntaxin (T-SNARE) of the ER
 YOR219c *STE13* type IV dipeptidyl aminopeptidase
 YMR022w *QRI8* ubiquitin-conjugating enzyme
 YER100w *UBC6* ubiquitin-conjugating enzyme
 YBR243c *ALG7* UDP-N-acetylglucosamine-1-phosphate transferase
 YGR105w *VMA21* vacuolar ATPase assembly integral membrane protein
 YBL017c *PEP1* vacuolar protein sorting/targeting protein
 YGL225w *GOG5* vanadate-resistance protein
organization and biogenesis of chromosome structure
 YJL081c *ACT3* actin-related protein
 YDL220c *CDC13* cell division control protein
 YBR195c *MSI1* chromatin assembly complex, subunit p50
 YML102w *CAC2* chromatin assembly complex, subunit p60
 YPR018w *RLF2* chromatin assembly complex, subunit p90
 YGR218w *CRM1* chromosome region maintenance protein
 YNL216w *RAP1* DNA-binding protein with repressor and activator activity
 YER159c *NCB1* functional homologue of human NC2α
 YMR072w *ABF2* high-mobility group protein
 YPL127c *HHO1* histone H1 protein
 YDR225w *HTA1* histone H2A
 YBL003c *HTA2* histone H2A.2
 YDR224c *HTB1* histone H2B
 YBL002w *HTB2* histone H2B.2
 YBR010w *HHT1* histone H3
 YNL031c *HHT2* histone H3
 YBR009c *HHF1* histone H4
 YNL030w *HHF2* histone H4
 YER161c *SPT2* HMG-like chromatin protein
 YPL254w *HFI1* interacts functionally with histone H2A
 YNR010w *CSE2* interacts with centromeric element CDEII
 YGR099w *TEL2* involved in controlling telomere length and position effect
 YJR060w *CBF1* kinetochore protein
 YGR140w *CBF2* kinetochore protein complex CBF3, 110K subunit
 YMR094w *CTF13* kinetochore protein complex CBF3, 58K subunit
 YMR168c *CEP3* kinetochore protein complex CBF3, 71K subunit
 YDR328c *SKP1* kinetochore protein complex CBF3, subunit D
 YIL072w *HOP1* meiosis-specific protein
 YLR263w *RED1* meiosis-specific protein
 YLR457c *NBP1* Nap1p-binding protein
 YDR162c *NBP2* Nap1p-binding protein
 YBR089c-a *NHP6B* nonhistone chromosomal protein
 YPR052c *NHP6A* nonhistone chromosomal protein related to mammalian HMG1
 YDR174w *HMO1* nonhistone protein
 YDL002c *HMO2* nonhistone protein
 YKR048c *NAP1* nucleosome assembly protein I
 YCL011c *GBP2* potential telomere-associated protein
 YGL238w *CSE1* probable kinetochore protein
 YGL194c *RTL1* putative deacetylase
 YBR275c *RIF1* Rap1p-interacting factor 1
 YKL089w *MIF2* required for normal chromosome segregation and spindle integrity
 YJL074c *SMC3* required for structural maintenance of chromosomes
 YLR086w similarity to chromosome condensation proteins
 YML069w *POB3* similarity to HMG proteins
 YOR053w similarity to protamines
 YNL206c similarity to structure-specific recognition proteins
 YDL208w *NHP2* strong similarity to high mobility group (HMG) family
 YEL026w strong similarity to high mobility group-like protein Nhp2p
 YOL012c *HTA3* strong similarity to histone H2A protein
 YKL049c *CSE4* strong similarity to histone H3
 YOR213c subunit of the RSC complex
 YCR052w *RSC6* subunit of the RSC complex
 YFR037c *RSC8* subunit of the RSC complex
 YLR321c *SFH1* subunit of the RSC complex
 YIL126w *STH1* subunit of the RSC complex
 YDR285w *ZIP1* synaptonemal complex protein
 YLR233c *EST1* telomere elongation protein
 YBL088c *TEL1* telomere length control protein
 YPL128c *TBF1* telomere TTAGGG repeat-binding factor 1

YGR274c *TAF145* TFIID subunit (TBP-associated factor), 145K
 YGR187c *HGH1* weak similarity to human Hmg1p and Hmg2p
 YGR285c *ZUO1* zotuin (Z-DNA binding protein)
mitochondrial organization and biogenesis
 YHR037w *PUT2* 1-pyrroline-5-carboxylate dehydrogenase
 YNL104c *LEU4* 2-isopropylmalate synthase
 YMR282c *AEP2* 2'-O-ribosyl phosphate transferase
 YIL125w *KGD1* 2-oxoglutarate dehydrogenase complex E1 component
 YDR148c *KGD2* 2-oxoglutarate dehydrogenase complex E2 component
 YOL096c *COQ3* 3,4-dihydroxy-5-hexaprenylbenzoate methyltransferase
 YMR287c *MSU1* 3'-5' exonuclease for RNA 3' ss-tail, mitochondrial
 YDR232w *HEM1* 5-aminolevulinic synthase
 YML009c *MRPL39* 60S ribosomal protein, mitochondrial
 YMR108w *ILV2* acetylacetyl synthase
 YCL009c *ILV6* acetylacetyl synthase, regulatory subunit
 YAL054c *ACS1* acetyl-CoA synthetase
 YER069w *ARG5,6* acetylglutamate kinase and acetylglutamyl-phosphate reductase
 YOL140w *ARG8* acetylornithine aminotransferase
 YLR304c *ACO1* aconitate hydratase
 YDR234w *LYS4* aconitate hydratase
 YER170w *ADK2* adenylate kinase, mitochondrial
 YMR056c *AAC1* ADP/ATP carrier protein (MCF)
 YBL030c *AAC2* ADP/ATP carrier protein (MCF)
 YBR085w *AAC3* ADP/ATP carrier protein (MCF)
 YMR083w *ADH3* alcohol dehydrogenase III
 YER073w aldehyde dehydrogenase (NAD⁺)
 YMR170c *ALD5* aldehyde dehydrogenase 2 (NAD⁺), mitochondrial
 YER086w *ILV1* anabolic serine and threonine dehydratase
 YJL209w *CBP1* apo-cytochrome b pre-mRNA processing protein
 YBR120c *CBP6* apo-cytochrome b pre-mRNA processing protein
 YHL038c *CBP2* apo-cytochrome b pre-mRNA processing protein 2
 YHR091c *MSR1* arginyl-tRNA synthetase, mitochondrial
 YCR024c aspartate synthetase, mitochondrial
 YKL106w *AAT1* aspartate transaminase, mitochondrial
 YPL104w *MSD1* aspartate-tRNA ligase, mitochondrial
 YGR008c *SFD2* ATPase stabilizing factor
 YDL130w-a *STF1* ATPase stabilizing factor, 10K
 YDR377w *ATP17* ATPase synthase subunit f
 YMR301c *ATM1* ATP-binding cassette transporter protein, mitochondrial
 YBL022c *PIM1* ATP-dependent protease, mitochondrial
 YPL029w *SUV3* ATP-dependent RNA helicase, mitochondrial
 YER061c *CEM1* β-keto-acyl-ACP synthase
 YHR208w *TWT1* branched chain amino-acid aminotransferase, mitochondrial
 YBR084w *MIS1* C1-tetrahydrofolate synthase, mitochondrial
 YML042w *CAT2* carnitine O-acetyltransferase
 YER026c *CHO1* CDP-diacylglycerol serine O-phosphatidyltransferase
 YNR001c *CIT1* citrate (si)-synthase, mitochondrial
 YPR001w *CIT3* citrate (si)-synthase, mitochondrial
 YBR291c *CTP1* citrate transport protein, mitochondrial (MCF)
 YOR316c *COT1* cobalt accumulation protein
 YOR017w *PET127* component of mitochondrial translation system
 YPR155c *NCA2* control of mitochondrial synthesis of Atp6p and Atp8p
 YKL011c *CCE1* cruciform-cutting endonuclease, mitochondrial
 YML078w *CPR3* cyclophilin (peptidylprolyl isomerase), mitochondrial
 YDR197w *CBS2* cytochrome b translational activator protein
 YIL043c *CBR1* cytochrome b5 reductase
 YKL150w *MCR1* cytochrome b560 subunit of respiratory complex II
 YKL141w *SDH3* cytochrome c isoform 1
 YJR048w *CYC1* cytochrome c isoform 2
 YEL039c *CYC2* cytochrome c mitochondrial import factor
 YOR037w *CYC2* cytochrome c oxidase assembly factor
 YER058w *PET117* cytochrome c oxidase assembly protein
 YPL132w *COX11* cytochrome c oxidase assembly protein
 YML129c *COX14* cytochrome c oxidase assembly protein
 YDR079w *PET100* cytochrome c oxidase assembly protein
 YGL187c *COX4* cytochrome c oxidase subunit IV
 YNL052w *COX5A* cytochrome c oxidase subunit V/A
 YIL111w *COX5B* cytochrome c oxidase subunit V/B
 YHR051w *COX6* cytochrome c oxidase subunit VI
 YGL191w *COX13* cytochrome c oxidase subunit VIa
 YLR038c *COX12* cytochrome c oxidase subunit VIb
 YMR256c *COX7* cytochrome c oxidase subunit VII
 YDL067c *COX9* cytochrome c oxidase subunit VIIa
 YLR395c *COX8* cytochrome c oxidase subunit VIII
 YKR066c *COP1* cytochrome c peroxidase
 YOR065w *CYT1* cytochrome c1
 YER141w *COX15* cytochrome oxidase assembly factor
 YER154w *OXA1* cytochrome oxidase biogenesis protein
 YOR386w *PHR1* deoxyribodipyrimidine photo-hyase
 YLR348c dicarboxylate carrier protein
 YFL018c *LPD1* dihydrolipoamide dehydrogenase
 YNL071w *LAT1* dihydrolipoamide S-acetyltransferase

| | | | | | | | | |
|---------|-------------------------------|--|-----------|---------------|--|--|----------------|---|
| YJR016c | <i>ILV3</i> | dihydroxy-acid dehydratase | YNL131w | <i>TOM22</i> | mitochondrial outer membrane import receptor subunit | YBL038w | <i>MRPL16</i> | ribosomal protein, mitochondrial |
| YDL174c | <i>DLD1</i> | D-lactate ferriocytochrome c oxidoreductase (D-LCR) | YGR082w | <i>TOM20</i> | mitochondrial outer membrane import receptor subunit, 20K | YMR193w | <i>MRPL24</i> | ribosomal protein, mitochondrial |
| YML061c | <i>PIF1</i> | DNA helicase involved in mitochondrial DNA repair and telomere length | YMR060c | <i>TOM37</i> | mitochondrial outer membrane import receptor subunit, 37K | YMR024w | <i>MRPL3</i> | ribosomal protein, mitochondrial |
| YHR120w | <i>MSH1</i> | DNA mismatch repair protein, mitochondrial | YMR203w | <i>TOM40</i> | mitochondrial outer membrane import receptor subunit, 40K | YMR286w | <i>MRPL33</i> | ribosomal protein, mitochondrial |
| YOR330c | <i>MIP1</i> | DNA-directed DNA polymerase γ , catalytic subunit, mitochondrial | YOR045w | <i>TOM6</i> | mitochondrial outer membrane import receptor subunit, 6K | YLR439w | <i>MRPL4</i> | ribosomal protein, mitochondrial |
| YFL036w | <i>RPO41</i> | DNA-directed RNA polymerase, mitochondrial | YNL070w | <i>TOM7</i> | mitochondrial outer membrane import receptor subunit, 7K | YHR147c | <i>MRPL6</i> | ribosomal protein, mitochondrial |
| YBR252w | <i>DUT1</i> | dUTP pyrophosphatase, mitochondrial | YNL055c | <i>POR1</i> | mitochondrial outer membrane porin | YGR220c | <i>MRPL9</i> | ribosomal protein, mitochondrial |
| YOR211c | <i>MGM1</i> | dynamatin-like protein | YNL121c | <i>TOM70</i> | mitochondrial outer membrane specialized import receptor | YDR337w | <i>MRPS28</i> | ribosomal protein, mitochondrial |
| YGR207c | <i>ETF-β</i> | electron-transferring flavoprotein, β subunit | YGL143c | <i>MRF1</i> | mitochondrial peptide chain release factor | YNL137c | <i>NAM9</i> | ribosomal protein, mitochondrial |
| YLR393w | <i>ATP10</i> | F1F0 ATPase complex assembly protein | YLR163c | <i>MAS1</i> | mitochondrial processing peptidase | YOR158w | <i>PET123</i> | ribosomal protein, mitochondrial |
| YNL315c | <i>ATP11</i> | F1F0-ATPase complex assembly protein | YLR024c | <i>MAS2</i> | mitochondrial processing peptidase, catalytic 53K (α) subunit | YCR046c | <i>PETCR46</i> | ribosomal protein, mitochondrial |
| YIL180c | <i>ATP12</i> | F1F0-ATPase complex assembly protein | YMR150c | <i>IMP1</i> | mitochondrial protease | YFR049w | <i>YMR31</i> | ribosomal protein, mitochondrial |
| YKL016c | <i>ATP7</i> | F1F0-ATPase complex, F0 D subunit | YDR375c | <i>BCS1</i> | mitochondrial protease | YDR194c | <i>MSS116</i> | RNA helicase of the DEAD box family, mitochondrial |
| YBL099w | <i>ATP1</i> | F1F0-ATPase complex, F1 α subunit | YDR120c | <i>TRM1</i> | CDC48/PAS1/SEC18 (AAA) family of ATPases | YMR228w | <i>MTF1</i> | RNA polymerase specific factor, mitochondrial |
| YJR121w | <i>ATP2</i> | F1F0-ATPase complex, F1 β subunit | YML120c | <i>ND1</i> | N2,N2-dimethylguanine tRNA methyltransferase | YOR334w | <i>MRS2</i> | RNA splicing protein and member of the mitochondrial carrier family (MCF) |
| YDL004w | <i>ATP16</i> | F1F0-ATPase complex, F1 δ subunit | YOR355w | <i>GDS1</i> | NADH-ubiquinone-6 oxidoreductase <i>nam9-1</i> suppressor | YJL133w | <i>MRS3</i> | RNA splicing protein and member of the mitochondrial carrier family (MCF) |
| YPL078c | <i>ATP4</i> | F1F0-ATPase complex, F1 δ subunit | YJL208c | <i>NUC1</i> | nuclease, mitochondrial | YKR052c | <i>MRS4</i> | RNA splicing protein and member of the mitochondrial carrier family (MCF) |
| YPL271w | <i>ATP15</i> | F1F0-ATPase complex, F1 ϵ subunit | YAR035w | <i>YAT1</i> | outer carnitine acetyltransferase, mitochondrial | YOR201c | <i>PET66</i> | rRNA (guanosine-2'-O)-methyltransferase |
| YBR039w | <i>ATP3</i> | F1F0-ATPase complex, F1 γ subunit | YNR041c | <i>COQ2</i> | para-hydroxybenzoate-polypropenyltransferase | YBR263w | <i>SHM1</i> | serine hydroxymethyltransferase, mitochondrial |
| YDR298c | <i>ATP5</i> | F1F0-ATPase complex, OSCP subunit | YPR047w | <i>MSF1</i> | phenylalanine-tRNA ligase α subunit, mitochondrial | YPR037c | | similarity to Erv1p and rat ALR protein |
| YLR296c | <i>ATP14</i> | F1F0-ATPase complex, subunit h | YJR077c | <i>MIR1</i> | phosphate transport protein, mitochondrial (MCF) | YDR376w | <i>ARH1</i> | similarity to human adrenodoxin reductase |
| YIL134w | <i>FLX1</i> | FAD carrier protein, mitochondrial (MCF) | YNL169c | <i>PSD1</i> | phosphatidylserine decarboxylase 1 | YBL013w | | similarity to methionyl-tRNA formyltransferase |
| YPL172c | <i>COX10</i> | farnesyl transferase | YLR203c | <i>MSS51</i> | possibly involved in translational activation of COX1 and COB mRNA | YJR113c | | similarity to mitochondrial ribosomal protein S7 |
| YOR176w | <i>HEM15</i> | ferrochelatase | YGL068w | | probable ribosomal protein L12 | YKL120w | <i>PMT</i> | similarity to mitochondrial uncoupling protein (MCF) |
| YPL262w | <i>FUM1</i> | fumarate hydratase | YLR168c | <i>(MSF1)</i> | probably involved in intramitochondrial protein sorting | YMR244w | | similarity to NCA3 and SUN4 protein |
| YOL033w | <i>MSE1</i> | glutamyl-tRNA synthetase, mitochondrial | YLR142w | <i>PUT1</i> | proline oxidase | YOR150w | | similarity to ribosomal protein L13 |
| YOL059w | <i>GPD3</i> | glycerol-3-phosphate dehydrogenase (NAD ⁺), mitochondrial | YER017c | <i>AFG3</i> | protease of the SEC18/CDC48/PAS1 family of ATPases (AAA) | YPR134w | <i>MSS18</i> | splicing protein |
| YIL155c | <i>GUT2</i> | glycerol-3-phosphate dehydrogenase, mitochondrial | YPR024w | <i>YME1</i> | protease of the SEC18/CDC48/PAS1 family of ATPases (AAA) | YGR222w | <i>PET54</i> | splicing protein and translational activator, mitochondrial |
| YMR189w | <i>GSD2</i> | glycine decarboxylase subunit | YMR089c | <i>YTA12</i> | protease of the SEC18/CDC48/PAS1 family of ATPases (AAA) | YCR028c-a | <i>RIM1</i> | ssDNA-binding protein, mitochondrial |
| YDR019c | <i>GCV1</i> | glycine decarboxylase T subunit | YDL044c | <i>MTF2</i> | protein involved in mRNA splicing and protein synthesis, mitochondrial | YKL192c | | strong similarity to acyl-carrier proteins |
| YFL016c | <i>MDJ1</i> | heat-shock protein - chaperone | YIL136w | <i>OM45</i> | protein of the outer mitochondrial membrane | YBR024w | <i>SCO2</i> | strong similarity to Scp1p |
| YOR232w | <i>MGE1</i> | heat-shock protein - chaperone | YAL011w | <i>FUN36</i> | protein of unknown function | YHR117w | <i>TOM71</i> | strong similarity to Tom70p/Mas70p |
| YLR259c | <i>HSP60</i> | heat-shock protein - chaperone, mitochondrial | YJL023c | <i>PET130</i> | protein synthesis protein, mitochondrial | YKL148c | <i>SDH1</i> | succinate dehydrogenase flavoprotein |
| YJR045c | <i>SSC1</i> | heat-shock protein 70-related protein, mitochondrial | YER014w | <i>HEM14</i> | protoporphyrinogen oxidase, mitochondrial | YLL041c | <i>SDH2</i> | succinate dehydrogenase iron-sulphur protein subunit |
| YDR258c | <i>HSP78</i> | heat-shock protein of clpb family of ATP-dependent proteases, mitochondrial | YER178w | <i>PDA1</i> | pyruvate dehydrogenase (lipoamide) α subunit | YDR178w | <i>SDH4</i> | succinate dehydrogenase membrane anchor subunit for Sdh2p |
| YBR003w | <i>COQ1</i> | hexaprenyl pyrophosphate synthetase | YBR221c | <i>PDB1</i> | pyruvate dehydrogenase (lipoamide) β subunit | YHR008c | <i>SOD2</i> | superoxide dismutase (Mn), mitochondrial |
| YMR072w | <i>ABF2</i> | high mobility group protein | YGR193c | <i>PDX1</i> | pyruvate dehydrogenase complex protein X | YHR050w | <i>SMF2</i> | suppressor of mitochondrial matrix mutant |
| YAL039c | <i>CYC3</i> | holocytochrome c synthase (cytochrome c haem lyase) | YJR095w | <i>ACR1</i> | regulator of acetyl-CoA synthetase activity | YKL194c | <i>MST1</i> | threonine-tRNA ligase, mitochondrial |
| YKL087c | <i>CYT2</i> | holocytochrome c1 synthase | YBR091c | <i>MRS5</i> | regulator of mitochondrial intron splicing | YLR069c | <i>MEF1</i> | translation elongation factor G, mitochondrial |
| YMR038c | <i>LYS7</i> | homocitrate dehydrogenase | YPL215w | <i>CBP3</i> | required for assembly of cytochrome bc1 complex | YOR187w | <i>TUF1</i> | translation elongation factor TU, mitochondrial |
| YDL181w | <i>INH1</i> | inhibitor of mitochondrial ATPase | YEL059c-a | <i>SOM1</i> | required for mitochondrial lmp1 peptidase function | YJL102w | <i>MEF2</i> | translation elongation factor, mitochondrial |
| YMR267w | <i>PPA2</i> | inorganic pyrophosphatase, mitochondrial | YLR093w | <i>MBR1</i> | required for optimal growth on glycerol | YOL023w | <i>IFM1</i> | translation initiation factor 2, mitochondrial |
| YLL009c | <i>COX17</i> | intra-mitochondrial sorting protein involved in assembly of cytochrome oxidase | YLR067c | <i>PET309</i> | required for stability and translation of COX1 mRNA | YDL069c | <i>CBS1</i> | translational activator of cob mRNA |
| YGR028w | <i>MSP1</i> | involved in early maturation of pre-rRNA | YMR257c | <i>PET111</i> | required for translation of COX2 mRNA | YER153c | <i>PET122</i> | translational activator of cytochrome c oxidase subunit III |
| YJR034w | <i>PET191</i> | involved in mitochondrial inheritance | YBL080c | <i>PET112</i> | required to maintain RHO ⁺ mitochondrial DNA | YNR045w | <i>PET494</i> | translational activator, mitochondrial |
| YMR302c | <i>PRP12</i> | involved in mitochondrial morphology and inheritance | YBR185c | <i>MBA1</i> | respiratory chain assembly protein | YOR274w | <i>MOD5</i> | tRNA isopentenyltransferase |
| YOL009c | <i>MDM12</i> | involved in mitochondrial morphology and inheritance | YML091c | <i>PPM2</i> | ribonuclease P, mitochondrial | YER168c | <i>CCA1</i> | tRNA nucleotidyltransferase |
| YAL010c | <i>MDM10</i> | involved in mitochondrial morphology and inheritance | YKL170w | <i>MRPL38</i> | ribosomal protein L14, mitochondrial | YDR268w | <i>MSW1</i> | tryptophanyl-tRNA synthetase, mitochondrial |
| YIR021w | <i>MRS1</i> | involved in mitochondrial RNA splicing of COB mRNA | YJL063c | <i>MRPL8</i> | ribosomal protein L17, mitochondrial | YPL097w | <i>MSY1</i> | tyrosyl-tRNA synthetase |
| YBR037c | <i>SCO1</i> | involved in stabilization of Cox1p and Cox2p | YPR166c | <i>MRP2</i> | ribosomal protein S14 | YFR033c | <i>QCR6</i> | ubiquinol-cytochrome c reductase 17K protein |
| YNL066w | <i>SUN4</i> | involved in the aging process | YPL013c | | ribosomal protein S16, mitochondrial | YPR191w | <i>QCR2</i> | ubiquinol-cytochrome c reductase 40K subunit II |
| YCL017c | <i>NFS1</i> | involved in tRNA processing and mitochondrial metabolism | YBR251w | <i>MRPS5</i> | ribosomal protein S5, mitochondrial | YBL045c | <i>COR1</i> | ubiquinol-cytochrome c reductase 44K core protein |
| YNL037c | <i>IDH1</i> | isocitrate dehydrogenase (NAD ⁺) subunit 1, mitochondrial | YBR146w | <i>MRPS9</i> | ribosomal protein S9, mitochondrial | YHR001w-a | <i>QCR10</i> | ubiquinol-cytochrome c reductase 8.5K subunit |
| YOR136w | <i>IDH2</i> | isocitrate dehydrogenase (NAD ⁺) subunit 2, mitochondrial | YKR006c | <i>MRPL13</i> | ribosomal protein YmL13, mitochondrial | YGR174c | <i>CBP4</i> | ubiquinol-cytochrome c reductase assembly factor |
| YDL066w | <i>IDP1</i> | isocitrate dehydrogenase (NADP ⁺), mitochondrial | YNL005c | <i>MRPL2</i> | ribosomal protein YmL2, mitochondrial | YGL119w | <i>ABC1</i> | ubiquinol-cytochrome c reductase complex assembly protein |
| YPL040c | <i>ISM1</i> | isoleucine-tRNA ligase, mitochondrial | YKR085c | <i>MRPL20</i> | ribosomal protein YmL20, mitochondrial | YEL024w | <i>RIP1</i> | ubiquinol-cytochrome c reductase iron-sulphur protein |
| YLR355c | <i>ILV5</i> | ketol-acid reducto-isomerase | YBR282w | <i>MRPL27</i> | ribosomal protein YmL27, mitochondrial | YDR529c | <i>QCR7</i> | ubiquinol-cytochrome c reductase subunit 7 |
| YML054c | <i>CYB2</i> | lactate dehydrogenase cytochrome b2 | YDR462w | | ribosomal protein YmL27, mitochondrial | YGR183c | <i>QCR9</i> | ubiquinol-cytochrome c reductase subunit 9 |
| YLR382c | <i>NAM2</i> | leucine-tRNA ligase, mitochondrial | YNL252c | | ribosomal protein YmL30, mitochondrial | YJL166w | <i>QCR8</i> | ubiquinol-cytochrome c reductase subunit VIII |
| YOR196c | <i>LIP5</i> | lipic acid synthase | YKL138c | <i>MRPL31</i> | ribosomal protein YmL31, mitochondrial | YML021c | <i>UNG1</i> | uracil-DNA glycosylase |
| YNL073w | <i>MSK1</i> | lysyl-tRNA synthetase, mitochondrial | YCR003w | <i>MRPL32</i> | ribosomal protein YmL32, mitochondrial | YGR094w | <i>VAS1</i> | valyl-tRNA synthetase |
| YKL085w | <i>MDH1</i> | malate dehydrogenase, mitochondrial | YBR122c | <i>MRPL36</i> | ribosomal protein YmL36, mitochondrial | YIL114c | <i>POR2</i> | voltage-dependent anion channel (YVDAC2) |
| YOR130c | <i>ARG11</i> | member of the mitochondrial carrier family (MCF) | YBR268w | <i>MRPL37</i> | ribosomal protein YmL37, mitochondrial | YNL237w | <i>YTP1</i> | weak similarity to mitochondrial electron transport proteins |
| YDL198c | <i>YHM1</i> | member of the mitochondrial carrier family (MCF) | YPL173w | | ribosomal protein YmL40, mitochondrial | peroxisomal organization and biogenesis | | |
| YGR171c | <i>MSM1</i> | methionyl-tRNA synthetase | YGR076c | <i>MRPL25</i> | ribosomal protein YmR26 (YmL25), mitochondrial | YIL160c | <i>POT1</i> | acetyl-CoA C-acyltransferase, peroxisomal |
| YGR029w | <i>ERV1</i> | mitochondrial biogenesis and regulation of cell cycle | YMR225c | <i>MRPL44</i> | ribosomal protein YmR44, mitochondrial | YAL054c | <i>ACS1</i> | acetyl-CoA synthetase |
| YBR192w | <i>RIM2</i> | mitochondrial carrier protein (MCF) | YML025c | | ribosomal protein, mitochondrial | YGL205w | <i>POX1</i> | acyl-CoA oxidase |
| YPR058w | <i>YMC1</i> | mitochondrial carrier protein (MCF) | YDR347w | <i>MRP1</i> | ribosomal protein, mitochondrial | YBR222c | <i>PCS60</i> | AMP-binding protein, peroxisomal |
| YBR104w | <i>YMC2</i> | mitochondrial carrier protein (MCF) | YGR084c | <i>MRP13</i> | ribosomal protein, mitochondrial | YML042w | <i>CAT2</i> | carnitine O-acyltransferase |
| YOR020c | <i>HSP10</i> | mitochondrial chaperonin | YKL003c | <i>MRP17</i> | ribosomal protein, mitochondrial | YDR256c | <i>CTA1</i> | catalase A, peroxisomal |
| YJR144w | <i>MGM101</i> | mitochondrial genome maintenance protein | YDR405w | <i>MRP20</i> | ribosomal protein, mitochondrial | YCR005c | <i>CIT2</i> | citrate (si)-synthase, peroxisomal |
| YMR023c | <i>MSS1</i> | mitochondrial GTPase involved in expression of COX1 | YHL004w | <i>MRP4</i> | ribosomal protein, mitochondrial | YKR009c | <i>FOX2</i> | hydratase-dehydrogenase-epimerase, peroxisomal |
| YNR017w | <i>MAS6</i> | mitochondrial inner membrane import translocase subunit | YKL167c | <i>MRP49</i> | ribosomal protein, mitochondrial | YER065c | <i>ICL1</i> | isocitrate lyase |
| YJL143w | <i>TIM17</i> | mitochondrial inner membrane import translocase subunit | YKL142w | <i>MRP8</i> | ribosomal protein, mitochondrial | YKL188c | <i>PAT1</i> | long-chain-fatty-acid transporter |
| YIL022w | <i>TIM44</i> | mitochondrial inner membrane import translocase subunit | YLR312w-a | <i>MRPL15</i> | ribosomal protein, mitochondrial | | | |
| YOR035w | <i>IMP2</i> | mitochondrial inner membrane protease subunit | | | | | | |
| YKL134c | <i>(MIP1)</i> | mitochondrial intermediate peptidase | | | | | | |

YPL147w *PXA1* long-chain-fatty-acid transporter
 YER015w *FAA2* long-chain-fatty-acid-CoA ligase
 YDL078c *MDH3* malate dehydrogenase, peroxisomal
 YNL117w *MLS1* malate synthase 1
 YIR031c *DAL7* malate synthase 2
 YDR329c *PAS3* peroxisomal assembly protein
 YDR265w *PAS4* peroxisomal assembly protein
 YJL210w *PAS5* peroxisomal assembly protein
 YNL329c *PAS8* peroxisomal assembly protein
 YDR142c *PAS7* peroxisomal import protein
 YOL147c *PMP27* peroxisomal membrane protein
 YLR191w *PAS20* peroxisomal protein involved in protein import
 YDR244w *PAS10* putative peroxisomal targeting signal receptor
 YLR109w similarity to *C. bodinii* peroxisomal membrane protein 20K A
 YGR077c similarity to peroxisomal matrix protein Per1p
 YLR251w similarity to peroxisomal rat membrane protein PMP22
 YNL202w *SPS19* sporulation-specific protein
 YGR133w *PAS2* ubiquitin-conjugating enzyme
 YBR204c weak similarity to peroxisomal serine-active lipase

endosomal organization and biogenesis

YKL135c *APL2* β -adaptin
 YNL192w *CHS1* chitin synthase I
 YBR023c *CHS3* chitin synthase III
 YJR058c *APS2* clathrin-associated protein 17, small subunit
 YJR005w *APL1* clathrin-associated protein complex, β subunit
 YBR288c *APM3* clathrin-associated protein complex, medium subunit
 YLR170c *APS1* clathrin-associated protein complex, small subunit AP19
 YDL145c *RET1* coatomer complex α subunit
 YDR238c *SEC26* coatomer complex β subunit of secretory pathway vesicles
 YGL137w *SEC27* coatomer complex β subunit
 YFR051c *RET2* coatomer complex δ subunit
 YNL287w *SEC21* coatomer complex γ subunit
 YPL010w *RET3* coatomer complex ζ subunit
 YPR181c *SEC23* component of COPII coat of ER-golgi vesicles
 YDR170c *SEC7* component of non-clathrin vesicle coat
 YLR083c *EMP70* endosomal protein
 YPR017c *DSS4* GDP/GTP exchange factor for Sec4p
 YOR089c *VPS21* GTP-binding protein
 YDL137w *ARF2* GTP-binding protein of the ARF family
 YML001w *YPT7* GTP-binding protein of the RAB family
 YFL005w *SEC4* GTP-binding protein of the RAS superfamily
 YLR268w *SEC22* high copy suppressor of ypt1 null mutation
 YPL085w *SEC16* multidomain vesicle protein
 YLR208w *SEC13* protein transport protein
 YKL196c similarity to Sec22p
 YLR080w strong similarity to Emp47p
 YDR107c strong similarity to Emp70p
 YAL030w *SNC1* strong similarity to synaptic vesicle-associated membrane protein
 YOR327c *SNC2* strong similarity to synaptobrevin
 YBL050w *SEC17* transport vesicle fusion protein
 YNR006w *VPS27* vacuolar protein sorting-associated protein
 YBR080c *SEC18* vesicular-fusion protein, functional homologue of NSF
 YLR078c *BOS1* weak similarity to synaptobrevin (V-SNARE)

vacuolar and lysosomal organization and biogenesis

YGL156w *AMS1* α -mannosidase
 YBR286w *APE3* aminopeptidase Y, vacuolar
 YKL103c *LAP4* aminopeptidase yscI, vacuolar
 YPL154c *PEP4* aspartyl protease
 YDL128w *VCX1* Ca²⁺-transport (H⁺/Ca²⁺ exchange) protein, vacuolar
 YGL006w *PMC1* Ca²⁺-transporting P-type ATPase
 YMR297w *PRC1* carboxypeptidase Y, serine-type protease
 YHR028c *DAF2* dipeptidyl aminopeptidase B
 YDR135c *YCF1* glutathione S-conjugate transporter, vacuolar
 YJL172w *CPS1* Gly-X carboxypeptidase YCS
 YKL119c *VPH2* H⁺-ATPase assembly protein, vacuolar
 YHR039c-a *VMA10* H⁺-ATPase V0 domain 13K subunit, vacuolar
 YEL027w *CUP5* H⁺-ATPase V0 domain 17K subunit, vacuolar
 YPL234c *TFP3* H⁺-ATPase V0 domain 17K subunit, vacuolar
 YLR447c *VMA6* H⁺-ATPase V0 domain 36K subunit, vacuolar
 YOR270c *VPH1* H⁺-ATPase V0 domain 95K subunit, vacuolar
 YGR020c *VMA7* H⁺-ATPase V1 domain 14K subunit, vacuolar
 YOR332w *VMA4* H⁺-ATPase V1 domain 27K subunit, vacuolar
 YEL051w *VMA8* H⁺-ATPase V1 domain 32K subunit, vacuolar
 YKL080w *VMA5* H⁺-ATPase V1 domain 42K subunit, vacuolar

YPR036w *VMA13* H⁺-ATPase V1 domain 54K subunit, vacuolar
 YBR127c *VMA2* H⁺-ATPase V1 domain 60K subunit, vacuolar
 YDL185w *TFP1* H⁺-ATPase V1 domain 69K subunit, vacuolar
 YFR019w *FAB1* probable PI P 5-kinase
 YEL060c *PRB1* protease B, vacuolar
 YDR481c *PHO8* repressible vacuolar alkaline phosphatase
 YIL099w *SGA1* sporulation specific glucan 1,4-glucosidase
 YOR036w *PEP12* syntaxin (T-SNARE)
 YOR106w *VAM3* syntaxin related protein
 YMR231w *PEP5* vacuolar biogenesis protein
 YLR148w *PEP3* vacuolar membrane protein
 YGL212w *VAM7* vacuolar morphogenesis protein
 YJL053w *PEP8* vacuolar protein-sorting/targeting protein
 YJL154c *VPS35* vacuolar protein-sorting protein
 YDR323c *PEP7* vacuolar segregation protein
 YPL045w *VPS16* vacuolar sorting protein
 YDR495c *VPS3* vacuolar sorting protein
 YLR396c *VPS33* vacuolar sorting protein
 YPR173c *VPS4* vacuolar sorting protein

other cellular organization and biogenesis activities

YKL157w *APE2* aminopeptidase yscII
 YIL015w *BAR1* barrier pepsin
 YJL174w *KRE9* cell wall synthesis protein
 YBR092c *PHO3* constitutive acid phosphatase
 YLR286c *CTS1* endochitinase
 YLR300w *EXG1* exo-1,3-glucanase (I/II), major isoform
 YIR019c *STA1* extracellular α -1,4-glucan glucosidase
 YPL187w *Mfa1* mating pheromone α -1 factor
 YGL089c *Mfa2* mating pheromone α -2 factor
 YDR461w *Mfa1* mating pheromone a-factor 1
 YNL145w *Mfa2* mating pheromone a-factor 2
 YKL163w *PIR3* member of the Pir1p/Pir2p/Pir3p family
 YBR093c *PHO5* repressible acid phosphatase
 YKL164c *PIR1* required for tolerance to heat-shock
 YML008c *ERG6* S-adenosyl-methionine δ -24-sterol-c-methyltransferase
 YAR071w *PHO11* secreted acid phosphatase
 YHR215w *PHO12* secreted acid phosphatase
 YNL160w *YGP1* secreted glycoprotein
 YLR250w *SSP120* secretory protein
 YBR046c *ZTA1* similarity to ζ -crystallin
 YMR215w similarity to GAS1 protein
 YNL275w similarity to human band 3 anion transport protein
 YIL162w *SUC2* sucrose hydrolyzing enzyme

Signal transduction

pheromone response generation

YDR264c *AKR1* ankyrin repeat-containing protein
 YPL161c *BEM4* bud emergence protein
 YCLO27w *FUS1* cell fusion protein
 YNL053w *MSG5* dual-specificity protein phosphatase
 YJL157c *FAR1* factor arrest protein
 YAL041w *CDC24* GDP/GTP exchange factor for Cdc42p
 YHR005c *GPA1* GTP-binding protein α subunit of the pheromone pathway
 YOR212w *STE4* GTP-binding protein β subunit of the pheromone pathway
 YJR086w *STE18* GTP-binding protein γ subunit of the pheromone pathway
 YLR229c *CDC42* GTP-binding protein of RAS superfamily
 YNL173c *MDG1* GTP-binding protein of the pheromone pathway
 YER020w *GPA2* guanine nucleotide-binding regulatory protein
 YLR452c *SST2* involved in desensitization to α -factor pheromone
 YMR052w *FAR3* involved in pheromone-mediated cell cycle arrest
 YDR461w *Mfa1* mating pheromone a-factor 1
 YIL047c *SYG1* member of the major facilitator superfamily
 YBL016w *FUS3* mitogen-activated protein kinase (MAP kinase)
 YFL026w *STE2* pheromone α -factor receptor
 YKL178c *STE3* pheromone α -factor receptor
 YCLO32w *STE50* pheromone response pathway protein
 YDR103w *STE5* pheromone signal transduction pathway protein
 YGR040w *KSS1* ser/thr protein kinase of the MAP kinase family
 YLR362w *STE11* ser/thr protein kinase of the MEKK family
 YHL007c *STE20* ser/thr protein kinase of the pheromone pathway
 YDL159w *STE7* ser/thr/tyr protein kinase of MAP kinase kinase family
 YHR146w similarity to pheromone-response G-protein YNL173c
 YHR084w *STE12* transcriptional activator

morphogenesis

YBR200w *BEM1* bud emergence mediator
 YPL161c *BEM4* bud emergence protein
 YBR109c *CMD1* calmodulin
 YAL041w *CDC24* GDP/GTP exchange factor for Cdc42p
 YGR070w *ROM1* GDP/GTP exchange protein for Rho1p

YLR371w *ROM2* GDP/GTP exchange protein for Rho1p
 YPL116c *BEM3* GTPase-activating protein for Cdc42p and Rho1p
 YLR229c *CDC42* GTP-binding protein of RAS superfamily
 YPR165w *RHO1* GTP-binding protein of the RHO subfamily of RAS-like proteins
 YNL090w *RHO2* GTP-binding protein of the RHO subfamily of RAS-like proteins
 YPL106c *SSE1* heat-shock protein of HSP70 family
 YOR149c *SMP3* protein kinase C pathway protein
 YPL140c *MKK2* protein kinase of the MAP kinase kinase (MEK) family
 YDL135c *RD1* RHO GDP dissociation inhibitor with activity towards Rho1p
 YOR231w *MKK1* ser/thr protein kinase
 YBL105c *PKC1* ser/thr protein kinase
 YHR030c *SLT2* ser/thr protein kinase of MAP kinase family
 YJL095w *BCK1* ser/thr protein kinase of the MEKK family
 YER167w *BCK2* ser/thr protein kinase of the protein kinase C pathway
 YOL113w *SKM1* Ste20p/PAK-like protein kinase
 YGL106w strong similarity to calmodulins
 YKL161c strong similarity to ser/thr-specific protein kinase Slk2p

YPL089c *RLM1* transcription factor of the MADS box family
 YGL095c *VPS45* vacuolar protein sorting-associated protein

osmosensing

YDL022w *GPD1* glycerol-3-phosphate dehydrogenase (NAD⁺), cytoplasmic
 YFL014w *HSP12* heat-shock protein
 YER118c *SSU81* involved in the HOG1 high-osmolarity signal transduction pathway
 YCR073c *SSK22* MAP kinase kinase kinase
 YNR031c *SSK2* MAP kinase kinase kinase of the high-osmolarity signal transduction pathway
 YDL006w *PTC1* protein ser/thr phosphatase 2c
 YLR113w *HOG1* ser/thr protein kinase of MAP kinase (MAPK) family
 YHR030c *SLT2* ser/thr protein kinase of MAP kinase family
 YKL161c strong similarity to ser/thr-specific protein kinase Slk2p
 YIL147c *SLN1* two-component signal transducer
 YLR006c *SSK1* two-component signal transducer
 YJL128c *PBS2* tyrosine protein kinase of the MAP kinase kinase family
 YDR069c *DOA4* ubiquitin-specific protease

nutritional response

YJL005w *CYR1* adenylate cyclase
 YLR178c *TFS1* cdc25-dependent nutrient- and ammonia-response cell-cycle regulator
 YBR195c *MSI1* chromatin assembly complex, subunit p50
 YLL016w *SDC25* GDP/GTP exchange factor
 YLR310c *CDC25* GDP/GTP exchange factor for Ras1p and Ras2p
 YOR101w *RAS1* GTP-binding protein
 YNL098c *RAS2* GTP-binding protein
 YER020w *GPA2* guanine nucleotide-binding regulatory protein
 YPL106c *SSE1* heat-shock protein of HSP70 family
 YBR140c *IRA1* inhibitory regulator protein of the RAS-cyclic AMP pathway
 YIL119c *RPI1* negative regulator of RAS-cAMP pathway
 YIR026c *YVH1* protein tyrosine phosphatase
 YOR208w *PTP2* protein tyrosine-phosphatase
 YOL110w *SHR5* RAS suppressor
 YPL084w *BRO1* required for normal response to nutrient limitation
 YLR362w *STE11* ser/thr protein kinase of the MEKK family
 YDL159w *STE7* ser/thr/tyr protein kinase of MAP kinase kinase family
 YHR084w *STE12* transcriptional activator

other signal-transduction activities

YPL268w *PLC1* 1-phosphatidylinositol-4,5-bisphosphate phosphodiesterase
 YNL138w *SRV2* adenylate cyclase-associated protein, 70K
 YFR014c *CMK1* Ca²⁺/calmodulin-dependent ser/thr protein kinase, type I
 YOL016c *CMK2* Ca²⁺/calmodulin-dependent ser/thr protein kinase, type II
 YPL203w *TPK2* cAMP-dependent protein kinase 2, catalytic subunit
 YKL166c *TPK3* cAMP-dependent protein kinase 3, catalytic subunit
 YJL164c *SRA3* cAMP-dependent protein kinase, catalytic subunit 1
 YMR028w *TAP42* component of the Tor signalling pathway
 YML064c *TEM1* GTP-binding protein of the RAS superfamily
 YJL146w *IDS2* IME2-dependent signalling protein
 YJR066w *TOR1* phosphatidylinositol 3-kinase
 YKL203c *TOR2* phosphatidylinositol 3-kinase
 YLR240w *VPS34* phosphatidylinositol 3-kinase
 YNL267w *PIK1* phosphatidylinositol 4-kinase
 YNL106c *PIE3* phosphatidylinositol phosphate phosphatase
 YLR305c *STT4* phosphatidylinositol-4-kinase
 YDL101c protein kinase

YHR079c *IRE1* protein kinase
 YMR016c *SOK2* regulatory protein in the PKA signal transduction pathway
 YGR216c *GPI1* required for N-acetylglucosaminyl phosphatidylinositol synthesis
 YPL153c *SPK1* ser/thr/tyr protein kinase
 YHR046c similarity to bovine myo-inositol-1(or 4)-monophosphatase
 YBR260c similarity to *C. elegans* GTPase-activating protein
 YGR136w similarity to chicken growth factor receptor-binding protein GRB2 homologue
 YDR379w similarity to Dbm1p
 YDR208w *MSS4* similarity to human PI P 5-kinase
 YCR027c similarity to human Ras-related GTP-binding protein
 YDR287w similarity to inositol monophosphatase
 YLR150w *MPT4* specific affinity for guanine-rich quadruplex nucleic acids
 YPR054w *SMK1* sporulation-specific MAP kinase
 YNL132w strong similarity to *A. ambisexualis* antheridial steroid receptor
 YHR206w *SKN7* transcription factor with similarity to Hsf1p

Cell rescue

stress response generation

YFR052w *NIN1* 26S proteasome regulatory subunit
 YIL075c *SEN3* 26S proteasome regulatory subunit
 YDR074w *TPS2* α,α -trehalose-phosphate synthase, 105K subunit
 YBR126c *TPS1* α,α -trehalose-phosphate synthase, 56K subunit
 YPR026w *ATH1* acid trehalase, vacuolar
 YIL033c *SRA1* cAMP dependent protein kinase, regulatory subunit
 YDR477w *SNF1* carbon catabolite derepressing ser/thr protein kinase
 YHR135c *YCK1* casein kinase I isoform
 YNL154c *YCK2* casein kinase I isoform
 YGR088w *CTT1* catalase T, cytosolic
 YDR251w *PAM1* coiled-coil protein multicopy suppressor of loss of PP2A
 YOR010c *TIR2* cold shock induced protein
 YER011w *TIR1* cold-shock induced protein of the Tir1p, Tip1p family
 YDR155c *CPH1* cyclophilin (peptidylprolyl isomerase)
 YEL039c *CYC7* cytochrome c isoform 2
 YER062c *HOR2* DL-glycerol phosphatase
 YDR263c *DIN7* DNA damage inducible protein
 YGL021w *ALK1* DNA damage-responsive protein
 YAL015c *NTG1* DNA repair protein
 YDR519w *FKB2* FK506/rapamycin-binding protein of the ER
 YGR234w *YHB1* flavohemoglobin
 YIR037w *HYR1* glutathione peroxidase
 YLL043w *FPS1* glycerol channel protein
 YNL098c *RAS2* GTP-binding protein
 YHR064c heat-shock protein
 YMR173w *DDR48* heat-shock protein
 YOL052c-a *DDRA2* heat-shock protein
 YMR186w *HSC92* heat-shock protein
 YLL026w *HSP104* heat-shock protein
 YFL014w *HSP12* heat-shock protein
 YBR072w *HSP26* heat-shock protein
 YCR021c *HSP30* heat-shock protein
 YPL240c *HSP82* heat-shock protein
 YFL016c *MDJ1* heat-shock protein - chaperone
 YLR259c *HSP60* heat-shock protein - chaperone, mitochondrial
 YJR045c *SSC1* heat-shock protein 70-related protein, mitochondrial
 YDR258c *HSP78* heat-shock protein of clpB family of ATP-dependent proteases, mitochondrial
 YEL030w heat-shock protein of HSP70 family
 YAL005c *SSA1* heat-shock protein of HSP70 family
 YER103w *SSA4* heat-shock protein of HSP70 family
 YPL106c *SSE1* heat-shock protein of HSP70 family
 YLL024c *SSA2* heat-shock protein of HSP70 family, cytosolic
 YBL075c *SSA3* heat-shock protein of HSP70 family, cytosolic
 YBR169c *SSE2* heat-shock protein of the HSP70 family
 YGL073w *HSF1* heat-shock transcription factor
 YMR251w-a *HOR7* hyperosmolarity-responsive protein
 YMR273c *ZDS1* involved in negative regulation of cell polarity
 YER118c *SSU81* involved in the *HOG1* high-osmolarity signal transduction pathway
 YDR293c *SSD1* involved in the tolerance to high concentration of Ca²⁺
 YKL143w *LTV1* low-temperature viability protein
 YGR100w *MIC1* Mac1p interacting protein
 YCR073c *SSK22* MAP kinase kinase kinase
 YNR031c *SSK2* MAP kinase kinase kinase of the high-osmolarity signal transduction pathway
 YIL158c member of the Pir1p/Hsp150p/Pir3p family
 YIL159w member of the Pir1p/Hsp150p/Pir3p family
 YIL160c member of the Pir1p/Hsp150p/Pir3p family
 YKL163w *PIR3* member of the Pir1p/Pir2p/Pir3p family
 YMR021c *MAC1* metal binding activator
 YNL064c *YDJ1* mitochondrial and ER import protein

YGL178w *MPT5* multicopy suppressor of *pop2*
 YGR159c *NSR1* nuclear localization sequence binding protein
 YGL115w *SNF4* nuclear regulatory protein
 YJR051w *OSM1* osmotic growth protein
 YMR175w *SIP18* osmotic stress protein
 YBR070c *SAT2* osmotolerance protein
 YHR057c *CYP2* peptidyl-prolyl *cis-trans* isomerase
 YOR014w *RTS1* potential regulatory subunit of protein phosphatase 2A
 YMR174c *PAI3* protease A (*ysca*) inhibitor IA3
 YHR079c *IRE1* protein kinase
 YPL140c *MKK2* protein kinase of the MAP kinase kinase (MEK) family
 YOL064c *MET22* protein ser/thr phosphatase
 YDL006w *PTC1* protein ser/thr phosphatase 2c
 YDR436w *PPZ2* protein ser/thr phosphatase of the PP-1 family
 YOR208w *PTP2* protein-tyrosine-phosphatase
 YJR090c *GRR1* required for glucose repression and for glucose and cation transport
 YPL084w *BRO1* required for normal response to nutrient limitation
 YKL164c *PIR1* required for tolerance to heat-shock
 YPR005c *HAL1* salt-induced protein
 YNL160w *YGP1* secreted glycoprotein
 YGL190c *CDC55* ser/thr phosphatase 2A regulatory subunit B
 YML016c *PPZ1* ser/thr phosphatase required for normal osmoregulation
 YJL165c *HAL5* ser/thr protein kinase
 YOR231w *MKK1* ser/thr protein kinase
 YBL105c *PKC1* ser/thr protein kinase
 YLR113w *HOG1* ser/thr protein kinase of MAP kinase (MAPK) family
 YHR030c *SLT2* ser/thr protein kinase of MAP kinase family
 YDL025c ser/thr protein kinase of the DEAD/DEAH box family
 YJL095w *BCK1* ser/thr protein kinase of the MEKK family
 YER167w *BCK2* ser/thr protein kinase of the protein kinase C pathway
 YDR227w *SIR4* silencing regulatory protein
 YKL088w similarity to *C. tropicalis* hal3 protein, to C-term. of Sis2p and to hypothetical protein YOR054c
 YBR044c similarity to chaperonin HSP60 proteins
 YJR147w similarity to heat-shock transcription factors
 YGR249w *MGA1* similarity to heat-shock transcription factors
 YBR054w *YRO2* similarity to HSP30 heat-shock protein Yro1p
 YAR020c similarity to members of the Srp1p/Tip1p family
 YCR008w *SAT4* similarity to Npr1p and Hal5p protein kinases
 YOR009w similarity to SRP1 and TIR2 proteins
 YCR060w similarity to stress inducible protein Sti1p
 YMR037c stress responsive regulatory protein
 YOR027w *STI1* stress-induced protein
 YBL009w strong similarity to DNA damage responsive Alk1p
 YLR369w strong similarity to heat-shock protein 70-related proteins
 YHL046c strong similarity to members of the Srp1p/Tip1p family
 YIL011w strong similarity to members of the Srp1p/Tip1p family
 YOL161c strong similarity to members of the Srp1p/Tip1p family
 YJL223c *PAU1* strong similarity to members of the Srp1p/Tip1p family
 YEL049w *PAU2* strong similarity to members of the Srp1p/Tip1p family
 YCR104w *PAU3* strong similarity to members of the Srp1p/Tip1p family
 YFL020c *PAU5* strong similarity to members of the Srp1p/Tip1p family
 YLR461w *PAU4* strong similarity to members of the Srp1p/Tip1p family
 YNR076w *PAU6* strong similarity to members of the Srp1p/Tip1p family
 YDR033w strong similarity to putative heat-shock protein Yro2p
 YKL161c strong similarity to ser/thr-specific protein kinase Sit2p
 YOR054c strong similarity to Sis2 protein and *C. tropicalis* hal3 protein
 YBR067c *TIP1* temp.shock induced protein of the Srp1p/Tip1p family
 YGR144w *THI4* thiamine-repressed protein
 YBL093c *ROX3* transcription factor
 YGL181w *GTS1* transcription factor of the Gcs1p/Glo3p/Sps18p family
 YMR043w *MCM1* transcription factor of the MADS box family
 YHR206w *SKN7* transcription factor with similarity to Hsf1p
 YKL062w *MSN4* transcriptional activator
 YML007w *YAP1* transcriptional activator involved in oxidative stress response
 YIL147c *SLN1* two-component signal transducer
 YLR006c *SSK1* two-component signal transducer
 YJL128c *PBS2* tyrosine protein kinase of the MAP kinase kinase family
 YLL039c *UBI4* ubiquitin
 YMR022w *QR18* ubiquitin-conjugating enzyme
 YBR082c *UBC4* ubiquitin-conjugating enzyme

YDR059c *UBC5* ubiquitin-conjugating enzyme
 YER125w *RSP5* ubiquitin-protein ligase
 YKL210w *UBA1* ubiquitin-protein ligase
 DNA repair (direct repair, base excision repair and nucleotide excision repair)
 YER142c *MAG1* 3-methyladenine DNA glycosylase
 YML060w *OGG1* 8-oxoguanine DNA glycosylase
 YKL114c *APN1* AP endonuclease
 YJL092w *HPR5* ATP-dependent DNA helicase
 YPL204w *HRR25* casein kinase I, ser/thr/tyr protein kinase
 YPL022w *RAD1* component of the nucleotide excision repairosome
 YDL108w *KIN28* cyclin-dependent ser/thr protein kinase
 YOR386w *PHR1* deoxyribodipyrimidine photo-lyase
 YOR368w *RAD17* DNA damage checkpoint control protein
 YER176w DNA dependent ATPase/DNA helicase B
 YLR032w *RAD5* DNA helicase
 YIL143c *SSL2* DNA helicase
 YML061c *PIF1* DNA helicase involved in mitochondrial DNA repair and telomere length
 YER171w *RAD3* DNA helicase/ATPase
 YDL164c *CDC9* DNA ligase
 YEL019c *MMS21* DNA repair protein
 YAL015c *NTG1* DNA repair protein
 YMR137c *PSO2* DNA repair protein
 YML095c *RAD10* DNA repair protein
 YCR066w *RAD18* DNA repair protein
 YER095w *RAD51* DNA repair protein
 YDR076w *RAD55* DNA repair protein
 YDR004w *RAD57* DNA repair protein
 YOR346w *REV1* DNA repair protein
 YDR369c *XRS2* DNA repair protein
 YGL163c *RAD54* DNA-dependent ATPase of the Snf2p family
 YDL102w *CDC2* DNA-directed DNA polymerase δ , catalytic 125K subunit
 YFR023w *PES4* DNA-directed DNA polymerase ϵ suppressor
 YNL262w *POL2* DNA-directed DNA polymerase ϵ , catalytic subunit A
 YPL167c *REV3* DNA-directed DNA polymerase ζ
 YKR056w *RNC1* endo-exonuclease
 YOL043c *NTG2* endonuclease III-like glycosylase 2
 YER162c *RAD4* excision repair protein
 YLR288c *MEC3* G2-specific checkpoint protein
 YFL014w *HSP12* heat-shock protein
 YIL128w *MMS19* involved in repair and RNA polymerase transcription
 YMR035w *IMP2* mitochondrial inner membrane protease subunit
 YKR095w *MLP1* myosin-like protein related to Uso1p
 YMR201c *RAD14* nucleotide excision repair protein
 YBR114w *RAD16* nucleotide excision repair protein
 YEL037c *RAD23* nucleotide excision repair protein
 YJR052w *RAD7* nucleotide excision repair protein
 YDL200c *MGT1* O6-methylguanine DNA repair methyltransferase
 YDL101c *DUN1* protein kinase
 YML032c *RAD52* recombination and DNA repair protein
 YIL139c *REV7* required for DNA damage induced mutagenesis
 YBR073w *RDH54* required for meiosis
 YIL066c *RNR3* ribonucleotide reductase, repair inducible large subunit
 YPL153c *SPK1* ser/thr/tyr protein kinase
 YDR061w similarity to *E. coli* deoxyribodipyrimidine photolyase
 YAL019w *FUN30* similarity to helicases of the Snf2/Rad54 family
 YLR035c similarity to human mutL protein homologue
 YPR056w similarity to human transcription factor BTF2/TFIIH subunit p34
 YFR038w strong similarity to mouse lymphocyte specific helicase
 YOR206w strong similarity to Rad4p
 YGR258c *RAD2* structure-specific nuclease of the nucleotide excision repairosome
 YDL088c *ASM4* suppressor of temperature-sensitive mutations in Pol3p
 YDR311w *TFB1* TFIIH subunit (transcription initiation factor), 75K
 YPR025c *CCL1* TFIIH subunit (transcription initiation factor), cyclin C component
 YLR005w *SSL1* TFIIH subunit (transcription initiation factor), factor B
 YPL122c *TFB2* TFIIH subunit (transcription/repair factor)
 YDR460w *TFB3* TFIIH subunit (transcription/repair factor)
 YGL058w *RAD6* ubiquitin-conjugating enzyme
 detoxification
 YOL052c *SPE2* adenosylmethionine decarboxylase
 YFL050c *ALR2* aluminum resistance protein
 YML116w *ATR1* aminotriazole and 4-nitroquinoline resistance protein
 YNL259c *ATX1* antioxidant protein and metal homeostasis factor
 YLR398c *SK12* antiviral protein and putative helicase
 YKL004w *AUR1* aureobasidin-resistance protein
 YDR256c *CTA1* catalase A, peroxisomal
 YGR088w *CTT1* catalase T, cytosolic
 YJR104c *SOD1* copper-zinc superoxide dismutase
 YKR066c *CCP1* cytochrome c peroxidase
 YPR198w *SGE1* drug resistance protein
 YJL101c *GSH1* glutamate-cysteine ligase

YDR513w *TTR1* glutaredoxin
 YPL091w *GLR1* glutathione reductase (NADPH)
 YDR135c *YCF1* glutathione S-conjugate transporter, vacuolar

YGR197c *SNG1* involved in nitroguanidine resistance
 YDL168w *SFA1* long-chain alcohol dehydrogenase
 YCR023c member of major facilitator superfamily multidrug-resistance protein family 2

YNR055c *HOL1* member of major facilitator superfamily multidrug-resistance protein subfamily 1

YHR053c *CUP1A* metallothionein
 YHR055c *CUP1B* metallothionein
 YOR079c *ATX2* multicopy suppressor of SOD-linked defects

YDR011w *SNQ2* multidrug resistance protein
 YHR042w *NCP1* NADPH-cytochrome P450 reductase
 YOR018w *ROD1* O-dinitrobenzene, calcium and zinc resistance protein

YKL064w *MNR2* overexpression overcomes manganese toxicity

YOR266w *PNT1* pentamidin resistance factor
 YDR538w *PAD1* phenylacrylic acid decarboxylase
 YOR153w *PDR5* pleiotropic drug resistance protein
 YBL005w *PDR3* pleiotropic drug resistance regulatory protein

YGL016w *PDR6* pleiotropic drug resistance regulatory protein

YIL120w similarity to antibiotic resistance proteins
 YIL121w similarity to antibiotic resistance proteins
 YLR299w similarity to *B. subtilis* γ -glutamyltransferase

YHL040c similarity to *C. carbonum* toxin pump
 YHL047c similarity to *C. carbonum* toxin pump
 YBR180w similarity to drug resistance proteins
 YDL100c similarity to *E. coli* arsenical pump-driving ATPase

YLL015w similarity to metal resistance proteins
 YBR293w similarity to multidrug resistance proteins
 YGR138c similarity to multidrug resistance proteins
 YHR048w similarity to multidrug resistance proteins
 YKR103w similarity to multidrug resistance proteins
 YKR104w similarity to multidrug resistance proteins
 YLL028w similarity to multidrug resistance proteins
 YPR156c similarity to multidrug resistance proteins
 YOR273c similarity to resistance proteins
 YOR251c similarity to thiosulphate sulphurtransferases

YOR247w similarity to vanadate-sensitive suppressor Sys1p
 YJR025c strong similarity to 3-hydroxyanthranilate 3,4-dioxygenase

YOL130w strong similarity to Alr2p
 YMR279c strong similarity to aminotriazole resistance protein

YOR378w strong similarity to aminotriazole resistance protein

YBR008c strong similarity to benomyl/methotrexate resistance protein

YCL069w strong similarity to drug resistance protein SGE1

YKL026c strong similarity to glutathione peroxidase
 YBR244w strong similarity to glutathione peroxidases

YKL033w-a strong similarity to halacid-halidohydrolase

YFR022w strong similarity to Rod1p
 YLR046c strong similarity to Rta1p and Rtm1p protein

YER185w strong similarity to Rtm1p
 YKR105c strong similarity to Sge1p and hypothetical protein YCL069w

YJR015w strong similarity to Sng1p
 YNR070w strong similarity to Snq2p
 YBL064c strong similarity to thiol-specific antioxidant enzyme

YDR453c strong similarity to thiol-specific antioxidant protein

YGL254w *FZF1* sulphite resistance protein
 YPL092w *SSU1* sulphite sensitivity protein
 YHR008c *SOD2* superoxide dismutase (Mn), mitochondrial
 YOR031w *CRS5* suppressor of *cup1* deletion, metallothionein-like protein

YML028w *TSA1* thiol-specific antioxidant
 YLR043c *TRX1* thioredoxin I
 YGR209c *TRX2* thioredoxin II
 YGL013c *PDR1* transcription factor
 YMR043w *MCM1* transcription factor of the MADS box family

YDR423c *CAD1* transcriptional activator
 YPL163c *SVS1* vanadate sensitive suppressor
 YLL060c weak similarity to glutathione transferase
 YMR243c *ZRC1* zinc- and cadmium resistance protein

detoxification involving cytochrome P450

YMR015c *ERG5* C-22 sterol desaturase
 YDR402c *DIT2* cytochrome P450 56
 YHR007c *ERG11* cytochrome P450 lanosterol 14 α -demethylase

YLL057c similarity to *E. coli* dioxygenase
 YDR403w *DIT1* spore wall maturation protein

cell death and ageing

YOR101w *RAS1* GTP-binding protein
 YNL098c *RAS2* GTP-binding protein
 YOL025w *LAG2* involved in determining longevity
 YJL116c *NCA3* involved in regulation of synthesis of Atp6p and Atp8p

YNL066w *SUN4* involved in the ageing process
 YKR042w *UTH1* involved in the ageing process
 YHL003c *LAG1* longevity-assurance protein
 YER167w *BCK2* ser/thr protein kinase of the protein kinase C pathway

YDR227w *SIR4* silencing regulatory protein
 YKL008c strong similarity to Lag1p
 YIL123w strong similarity to Sun4p, Nca3p

degradation of exogenous polynucleotides

YGL213c *SKI8* antiviral protein of the β -transducin (WD-40) repeat family

other cell-rescue activities

YPR189w *SKI3* antiviral protein
 YJR069c *HAM1* controls 6-N-hydroxylaminopurine sensitivity and mutagenesis

YGR213c *RTA1* involved in 7-aminocholesterol resistance
 YKL110c *KTI12* involved in resistance to *K. lactis* killer toxin

YER076c similarity to killer toxin Khrlp
 YER187w similarity to killer toxin KHS

Unclassified proteins

YEL052w *AFG1* ATPase family gene
 YDR184c *BAT1* binds Aip3p

YCR072c β -transducin family (WD-40 repeat) protein
 YLR459w *CDC91* cell division control protein
 YJL079c *PRY1* contains homology to the plant PR-1 class of pathogen related proteins

YKR013w *PRY2* contains homology to the plant PR-1 class of pathogen related proteins
 YJL078c *PRY3* contains homology to the plant PR-1 class of pathogen related proteins

YOR302w *CPA1-1* CPA1 leader peptide
 YML113w *DAT1* datin, oligo(dA)/oligo(dT)-binding protein
 YKL020c *SPT23* dosage-dependent suppressor of Ty-induced promotor mutations

YJL168c *EZL1* enhancer of zeste-like
 YOR123c *LEO1* extremely hydrophilic protein
 YLL065w *GIN11* growth inhibitory protein
 YDL213c has an RNA recognition domain in the N-terminal region

YDR505c *PSP1* high copy suppressor of ts of mutations in DNA polymerase α

YNL087w homology to YOR086c
 YLR161w identical to hypothetical proteins YLR156w and YLR161w

YLR156w identical to hypothetical proteins YLR159w and YLR161w

YLR159w identical to hypothetical proteins YLR161w and YLR156w

YHR212c identical with hypothetical protein YAR060c

YHR214w identical with hypothetical protein YAR066w

YAR066w identical with hypothetical protein YIL169c
 YCR020c-a *MAK31* involved in stability of L-A dsRNA-containing particles

YPL049c *DIG1* MAP kinase-associated protein, down-regulator of invasive growth

YKL034w member of Kazal serine protease inhibitors family

YLR397c *AFG2* member of the Sec18p, Pas1p, Cdc48p, TBP-1 family of ATPases

YCR007c member of the YBR302c family

YGL036w *MTC2* Mtf1 two hybrid clone 2
 YMR200w *ROT1* mutant suppresses *tor2* mutation
 YAL059w *SIM1* mutants are hypersensitive to calcofluor white

YCR019w *MAK32* necessary for structural stability of L-A dsRNA-containing particles

YNL061w *NOP2* nucleolar protein
 YFR011c ochre suppressor tyr-tRNA
 YLR196w *PWP1* periodic tryptophan protein
 YDL235c *YPD1* phosphorelay intermediate between Sln1p and Ssk1p

YOR014w *RTS1* potential regulatory subunit of protein phosphatase 2A

YOR181w *LAS17* proline-rich protein
 YER129w *PAK1* protein kinase

YOR223w protein of unknown function
 YHR093w *AHT1* protein of unknown function
 YDL167c *ARP1* protein of unknown function
 YHR191c *CTF8* protein of unknown function
 YAL008w *FUN14* protein of unknown function
 YAR002w *FUN17* protein of unknown function
 YAR014c *FUN2* protein of unknown function
 YAL031c *FUN21* protein of unknown function
 YAL014c *FUN34* protein of unknown function
 YAR008w *FUN4* protein of unknown function
 YAL033w *FUN53* protein of unknown function
 YDL234c *GYP7* protein of unknown function
 YPL054w *LEE1* protein of unknown function
 YDR335w *MSN5* protein of unknown function
 YIR006c *PAN1* protein of unknown function
 YDL105w *QRI2* protein of unknown function
 YLR204w *QRI5* protein of unknown function
 YLL003w *SFI1* protein of unknown function
 YDR350c *TCM10* protein of unknown function
 YDL169c *UGX2* protein of unknown function
 YJR050w *UTR3* protein of unknown function
 YEL035c *UTR5* protein of unknown function

YJR067c *YAE1* protein of unknown function
 YGR172c *YIP1* protein of unknown function
 YNL044w *YIP3* protein of unknown function
 YBR111c *YSA1* protein of unknown function
 YGR123c *PPT1* protein ser/thr phosphatase
 YDL230w *PTP1* protein tyrosine phosphatase
 YER075c *PTP3* protein tyrosine phosphatase
 YPR073c *LTP1* putative phosphatase
 YNL128w regulatory protein of the β -transducin family
 YCL039w

YJR055w *HIT1* required for growth at high temperature
 YNL085w *MKT1* required for propagation of M2 dsRNA satellite of L-A virus

YLL046c *RNP1* ribonucleoprotein
 YOR046c *DBP5* RNA helicase

YDR466w ser/thr protein kinase
 YDR490c ser/thr protein kinase
 YGL179c ser/thr protein kinase
 YGL180w ser/thr protein kinase
 YKL168c ser/thr protein kinase
 YKL171w ser/thr protein kinase
 YLR063w ser/thr protein kinase
 YMR291w ser/thr protein kinase
 YNL161w ser/thr protein kinase
 YOL045w ser/thr protein kinase
 YOL100w ser/thr protein kinase

YAR018c *KIN3* ser/thr protein kinase
 YOR233w *KIN4* ser/thr protein kinase
 YCR091w *KIN82* ser/thr protein kinase
 YLL019c *KNS1* ser/thr protein kinase
 YHR082c *KSP1* ser/thr protein kinase
 YDL079c *MRK1* ser/thr protein kinase with similarity to Npr1p

YMR216c ser/thr protein kinase with similarity to *S. pombe* dsk1
 YKL116c ser/thr protein kinase with similarity to *S. pombe* nim1 protein

YBL056w *PTC3* ser/thr protein phosphatase PP2C
 YDR507c *GIN4* ser/thr protein kinase
 YKL126w *SPT1* ser/thr-specific protein kinase

YOR007c similarities to protein phosphatases
 YOR119c similarity to a *C. elegans* ZK632.3 protein
 YLR405w similarity to *A. brasilense* nifR3 protein
 YML079w similarity to *A. brasilense* nifR3 protein
 YML080w similarity to *A. brasilense* nifR3 protein
 YDR205w similarity to *A. eutrophus* cation efflux system membrane protein czd, rat zinc transport protein ZnT1 and Cot1p

YDL228c similarity to *A. klebsiana* glutamate dehydrogenase

YLL005c similarity to *A. thaliana* hyp1 protein
 YMR266w similarity to *A. thaliana* hyp1 protein
 YOL084w similarity to *A. thaliana* hyp1 protein
 YNL317w similarity to *A. thaliana* PRL1 protein
 YOL105c similarity to a-agglutinin core protein AGA1 and mucin proteins

YPL252c similarity to adrenodoxin and ferredoxin
 YMR110c similarity to aldehyde dehydrogenase
 YIL112w similarity to ankyrin and coiled-coil proteins

YLR064w similarity to *Anopheles* NADH-ubiquinone oxidoreductase, subunit 4

YMR153w similarity to Asm4p
 YDR349c similarity to aspartyl proteases
 YDR072c similarity to Aur1p
 YMR305c similarity to *B. japonicum* putative β -(6)-glucan transferase

YDR291w similarity to *B. subtilis* helicases
 YPR201w similarity to *B. subtilis* hypothetical protein
 YPR002w similarity to *B. subtilis* mmgE protein
 YPL258c similarity to *B. subtilis* transcriptional activator tenA, and strong similarity to hypothetical proteins YOL055c and YPR121w

YPR121w similarity to *B. subtilis* transcriptional activator tenA, strong similarity to hypothetical proteins YPL258c and YOL055c

YIL159w similarity to BNI1 protein
 YDL119c similarity to bovine Graves disease carrier protein

YNR068c similarity to Bul1p
 YNL218w similarity to *C. burnetii* trxB, spoIIIE and serS genes

YCR102c similarity to *C. carbonum* toxD gene
 YNL134c similarity to *C. carbonum* toxD gene
 YJR013w similarity to *C. elegans* B0491.1 protein
 YPR040w similarity to *C. elegans* C02C2.6 protein
 YGR257c similarity to *C. elegans* C16C10.1
 YGR054w similarity to *C. elegans* E04D5.1 protein
 YMR012w similarity to *C. elegans* hypothetical 139.9K protein F55H2.6

YLR022c similarity to *C. elegans* hypothetical protein

YNL127w similarity to *C. elegans* hypothetical protein

YKL095w *YJU2* similarity to *C. elegans* hypothetical protein

YML014w similarity to *C. elegans* hypothetical protein C14B1.5

YKL099c similarity to *C. elegans* hypothetical protein C16C10.2

YEL004w similarity to *C. elegans* hypothetical protein C53B4.6

YHL030w similarity to *C. elegans* hypothetical protein D2045.2

| | | | | | |
|-----------|--|-----------|---|-----------|---|
| YOL093w | similarity to <i>C. elegans</i> hypothetical protein F25H8.1 | YLR050c | similarity to human MAC30 C-terminus | YGL228w | similarity to hypothetical protein YFR039c |
| YDL008w | similarity to <i>C. elegans</i> hypothetical protein F35G12.9 | YCR071c | similarity to human NOF1 protein | YBR281c | similarity to hypothetical protein YFR044c |
| YKL151c | similarity to <i>C. elegans</i> hypothetical protein R1072 | YOR001w | similarity to human nucleolar 100K polyomysitis-scleroderma protein | YDL001w | similarity to hypothetical protein YFR048w, YDR282c and <i>S. pombe</i> hypothetical protein SPAC12G12.14 |
| YDR196c | similarity to <i>C. elegans</i> hypothetical protein T05G5.5 | YEL016c | similarity to human nucleotide pyrophosphatase | YPL216w | similarity to hypothetical protein YGL133w |
| YJL029c | similarity to <i>C. elegans</i> hypothetical protein T05G5.8 | YIL153w | similarity to human phosphotyrosyl phosphatase activator | YPL219w | similarity to hypothetical protein YGL134w |
| YNL223w | similarity to <i>C. elegans</i> hypothetical protein ZK792.1 | YDR280w | similarity to human PM-Scl-75 autoantigen | YDR444w | similarity to hypothetical protein YGL144c and YDL1009c |
| YOL003c | similarity to <i>C. elegans</i> hypothetical protein, YDR126w, YNL326c and YLR246w | YPR108w | similarity to human protein GPs1 | YLR047c | similarity to hypothetical protein YGL160w |
| YDR049w | similarity to <i>C. elegans</i> K06H7.3 protein | YLR206w | similarity to human protein KIAA0171 and hypothetical protein YDL161w | YER132c | similarity to hypothetical protein YGL197w |
| YGR278w | similarity to <i>C. elegans</i> LET-858 | YNL265c | similarity to human protein KIAA0174 | YFR039c | similarity to hypothetical protein YGL228w |
| YHR004c | similarity to <i>C. elegans</i> SPAC2F7.02c protein | YGR163w | similarity to human ragA protein | YHR036w | similarity to hypothetical protein YGL247w |
| YML059c | similarity to <i>C. elegans</i> ZK370.4 protein | YDR096w | similarity to human retinoblastoma binding protein 2 | YGR031w | similarity to hypothetical protein YGR015c |
| YNL168c | similarity to <i>C. elegans</i> ZK688.3 protein and <i>E. coli</i> hpcEp | YDR378c | similarity to human Sm protein F | YGR015c | similarity to hypothetical protein YGR031w |
| YPR188c | similarity to calmodulin and calmodulin-related proteins | YGR112w | similarity to human SURF-1 protein | YLR105c | similarity to hypothetical protein YGR071c |
| YPR128c | similarity to carrier protein FLX1 | YGR080w | similarity to human tyrosine kinase A6 | YLR373c | similarity to hypothetical protein YGR142w |
| YPL202c | similarity to cell size regulation protein Rcs1p | YDL060w | similarity to hypothetical <i>C. elegans</i> protein | YPR158w | similarity to hypothetical protein YGR142w |
| YPR154w | similarity to chicken growth factor receptor-binding protein GRB2 homologue | YDR183w | similarity to hypothetical <i>C. elegans</i> protein | YHR149c | similarity to hypothetical protein YGR221c |
| YNL091w | similarity to chicken h-caldesmon, Uso1p and hypothetical protein YKL201c | YDR427w | similarity to hypothetical <i>C. elegans</i> protein | YHR158c | similarity to hypothetical protein YGR238c |
| YPL176c | similarity to chinese hamster transferrin receptor protein | YDR473c | similarity to hypothetical <i>C. elegans</i> protein | YMR295c | similarity to hypothetical protein YGR273c |
| YDR259c | similarity to Cin5p | YER051w | similarity to hypothetical <i>C. elegans</i> protein | YMR310c | similarity to hypothetical protein YGR283c |
| YGL241w | similarity to Cse1p | YER051w | similarity to hypothetical <i>C. elegans</i> protein | YDR338c | similarity to hypothetical protein YHR032w |
| YNL305c | similarity to C-term. of <i>A. nidulans</i> regulatory protein (qutR) | YGR145w | similarity to hypothetical <i>C. elegans</i> protein | YGL247w | similarity to hypothetical protein YHR036w |
| YPR127w | similarity to C-term. of <i>N. tabacum</i> auxin-induced protein | YGR194c | similarity to hypothetical <i>C. elegans</i> protein | YDR309c | similarity to hypothetical protein YHR061c |
| YDL112w | similarity to C-terminus of human TRP-185 protein | YGR245c | similarity to hypothetical <i>C. elegans</i> protein | YNL075w | similarity to hypothetical protein YHR088w |
| YMR304w | similarity to <i>D. melanogaster</i> fat facets gene | YHL010c | similarity to hypothetical <i>C. elegans</i> protein | YDR348c | similarity to hypothetical protein YHR097c |
| YGR003w | similarity to <i>D. melanogaster</i> lin19 protein | YJR070c | similarity to hypothetical <i>C. elegans</i> protein | YNL144c | similarity to hypothetical protein YHR131c |
| YLL013c | similarity to <i>D. melanogaster</i> pumilio protein | YML094w | similarity to hypothetical <i>C. elegans</i> protein | YNL156c | similarity to hypothetical protein YHR133c |
| YNL023c | similarity to <i>D. melanogaster</i> shuttle craft protein | YMR002w | similarity to hypothetical <i>C. elegans</i> protein | YGR221c | similarity to hypothetical protein YHR149c |
| YKL201c | similarity to <i>D. melanogaster</i> sperm-tail-specific protein | YJR072c | similarity to hypothetical <i>C. elegans</i> protein | YOR147w | similarity to hypothetical protein YHR194w |
| YCL001w-a | similarity to Dom34p | YOL060c | similarity to hypothetical <i>C. elegans</i> protein and YLR243w | YER175c | similarity to hypothetical protein YHR209w |
| YIL103w | similarity to Dph2 protein | YDL097c | similarity to hypothetical <i>C. elegans</i> protein M02F4.4 | YJR108w | similarity to hypothetical protein YIL015c-a |
| YGL018c | similarity to <i>E. coli</i> dnaI homologue | YER004w | similarity to hypothetical <i>C. elegans</i> proteins | YER064c | similarity to hypothetical protein YIL056w |
| YHL014c | similarity to <i>E. coli</i> GTP-binding protein | YDL238c | similarity to hypothetical <i>E. coli</i> and <i>C. elegans</i> proteins | YLR036c | similarity to hypothetical protein YIL089w |
| YLR011w | similarity to <i>E. coli</i> hypothetical 20.4K protein | YDR013w | similarity to hypothetical human KIAA0186 protein | YFL034c-b | similarity to hypothetical protein YIL106w |
| YGL037c | similarity to <i>E. coli</i> hypothetical 23K protein | YDR496c | similarity to hypothetical human protein | YNL058c | similarity to hypothetical protein YIL117c |
| YOR131c | similarity to <i>E. coli</i> hypothetical 27K protein | YDR524c | similarity to hypothetical human protein | YNL074c | similarity to hypothetical protein YIL135c |
| YDR400w | similarity to <i>E. coli</i> hypothetical 33.7K protein in nfo-frau intergenic region | YDR524c | similarity to hypothetical human protein | YKR100c | similarity to hypothetical protein YIL158w |
| YDR539w | similarity to <i>E. coli</i> hypothetical 55.3K protein in rfh-rfe intergenic region | YHR098c | similarity to hypothetical human protein | YKR015c | similarity to hypothetical protein YIL043w |
| YDR332w | similarity to <i>E. coli</i> hypothetical protein | YOR294w | similarity to hypothetical human protein | YBR273c | similarity to hypothetical protein YIL048c |
| YGR021w | similarity to <i>E. coli</i> hypothetical protein | YOR215c | similarity to hypothetical <i>M. xanthus</i> protein | YLL031c | similarity to hypothetical protein YIL062w |
| YJL046w | similarity to <i>E. coli</i> lipote-protein ligase A | YDR105c | similarity to hypothetical mouse protein | YKR010c | similarity to hypothetical protein YIL076w |
| YIL003w | similarity to <i>E. coli</i> MRP protein | YDL193w | similarity to hypothetical <i>N. crassa</i> 32K protein | YKR019c | similarity to hypothetical protein YIL083w |
| YOR165w | similarity to <i>E. histolytica</i> surface lectin | YOR324c | similarity to hypothetical protein YAL028w | YKR029c | similarity to hypothetical protein YIL105p |
| YNR015w | similarity to <i>E. histolytica</i> nitrogen fixation regulatory protein-3 homologue | YOR371c | similarity to hypothetical protein YAL056w | YDR131c | similarity to hypothetical protein YIL149w |
| YER113c | similarity to Emp70p | YJR115w | similarity to hypothetical protein YBL059w | YDL123w | similarity to hypothetical protein YIL151c |
| YDR036c | similarity to enoyl-CoA hydratase | YER093c-a | similarity to hypothetical protein YBL101c | YBR162c | similarity to hypothetical protein YIL171c |
| YOR172w | similarity to finger protein YKL222c, YOR162c and YLR266c, weak similarity to transcription factors | YPR030w | similarity to hypothetical protein YBR002c | YJR030c | similarity to hypothetical protein YIL181w |
| YOR162c | similarity to finger proteins YKL222c, YOR162c and YLR266c, weak similarity to transcription factors | YMR101c | similarity to hypothetical protein YBR105c | YJL181w | similarity to hypothetical protein YJR300c |
| YOR338w | similarity to FUN19 protein | YJL171c | similarity to hypothetical protein YBR162c | YPR114w | similarity to hypothetical protein YKL041w |
| YLR193c | similarity to <i>G. gallus</i> px19 | YER145c | similarity to hypothetical protein YBR207w | YKL002w | similarity to hypothetical protein YKL050c |
| YKL054c | similarity to glutenin, high molecular weight subunit | YLR387c | similarity to hypothetical protein YBR267w | YMR031c | similarity to hypothetical protein YKL050c and human resin |
| YOL132w | similarity to glycopospholipid-anchored surface glycoprotein GAS1 | YJL048c | similarity to hypothetical protein YBR273c | YMR086w | similarity to hypothetical protein YKL105c |
| YHR168w | similarity to GTP-binding proteins | YFR044c | similarity to hypothetical protein YBR281c | YMR171c | similarity to hypothetical protein YKL124w |
| YDL033c | similarity to <i>H. influenzae</i> hypothetical protein HI0174 | YDL177c | similarity to hypothetical protein YCR059c | YMR115w | similarity to hypothetical protein YKL133c |
| YLL027w | similarity to <i>H. influenzae</i> hypothetical protein HI0376 | YDR282c | similarity to hypothetical protein YDL001w, YFR048w and <i>S. pombe</i> hypothetical protein SPAC12G12.14 | YNL101w | similarity to hypothetical protein YKL146w |
| YLR419w | similarity to helicases | YDR425w | similarity to hypothetical protein YDL113c | YJL076w | similarity to hypothetical protein YKR010c |
| YMR128w | similarity to helicases | YLR426w | similarity to hypothetical protein YDL114w | YJL043w | similarity to hypothetical protein YKR015c |
| YGR271w | similarity to Hfm1p | YJL151c | similarity to hypothetical protein YDL123w | YJL083w | similarity to hypothetical protein YKR019c |
| YDR080w | similarity to human (clone S53) mRNA, 3' end of cds | YDR233c | similarity to hypothetical protein YDL204w | YJL105w | similarity to hypothetical protein YKR029c |
| YLR051c | similarity to human acidic 82K protein | YER139c | similarity to hypothetical protein YDR066c | YIL158w | similarity to hypothetical protein YKR100c |
| YCR026c | similarity to human autotaxin | YNL249c | similarity to hypothetical protein YDR109c | YLR019w | similarity to hypothetical protein YKL010c |
| YIL059w | similarity to human Batten disease-related protein CLN3 | YLR099c | similarity to hypothetical protein YDR125c | YNL328c | similarity to hypothetical protein YLR008c |
| YHR099w | similarity to human DNA-PK | YJL149w | similarity to hypothetical protein YDR131c | YPL063w | similarity to hypothetical protein YLR019w and <i>S. pombe</i> hypothetical protein SPAC2F7.02c |
| YOL114c | similarity to human DS-1 protein | YLR238w | similarity to hypothetical protein YDR200c | YIL089w | similarity to hypothetical protein YLR036c |
| YJR036c | similarity to human E6-associated protein | YDL204w | similarity to hypothetical protein YDR233c | YLL023c | similarity to hypothetical protein YLR064w |
| YBR265w | similarity to human FV171 protein | YHR061c | similarity to hypothetical protein YDR309c | YFL043c | similarity to hypothetical protein YLR072w |
| YPL120w | similarity to human GT197 (partial ORF) | YHR080c | similarity to hypothetical protein YDR326c, YFL042c and YLR072w | YNR067c | similarity to hypothetical protein YLR144c |
| YPR023c | similarity to human hypothetical protein | YOR012w | similarity to hypothetical protein YDR391c | YDR501w | similarity to hypothetical protein YLR183c |
| YHR141w | similarity to human hypothetical protein 1 | YDL113c | similarity to hypothetical protein YDR425w | YNL278w | similarity to hypothetical protein YLR187w |
| YPL217c | similarity to human hypothetical protein KIAA0187 | YML018c | similarity to hypothetical protein YDR438w | YDR200c | similarity to hypothetical protein YLR238w |
| YJR125c | similarity to human KIAA0171 protein | YML033w | similarity to hypothetical protein YDR458c | YOR262w | similarity to hypothetical protein YLR243w |
| | | YML034w | similarity to hypothetical protein YDR458c | YDR126w | similarity to hypothetical protein YLR246w and YOL003c |
| | | YOR019w | similarity to hypothetical protein YDR474c | YOR137c | similarity to hypothetical protein YLR361c |
| | | YLR183c | similarity to hypothetical protein YDR501w | YGR071c | similarity to hypothetical protein YLR373c |
| | | YCL036w | similarity to hypothetical protein YDR514c | YPR117w | similarity to hypothetical protein YLR454w |
| | | YNR027w | similarity to hypothetical protein YEL029c | YDR458c | similarity to hypothetical protein YML034w |
| | | YGL197w | similarity to hypothetical protein YER132c | YJR054w | similarity to hypothetical protein YML047c |
| | | YDR066c | similarity to hypothetical protein YER139c | YPL184c | similarity to hypothetical protein YML117w |
| | | YBR207w | similarity to hypothetical protein YER145c | YKL050c | similarity to hypothetical protein YMR031c |
| | | YHR209w | similarity to hypothetical protein YER175c | YKL105c | similarity to hypothetical protein YMR086w |
| | | YGL262w | similarity to hypothetical protein YER187w and killer toxin KHS | YKL133c | similarity to hypothetical protein YMR115w |
| | | YPL100w | similarity to hypothetical protein YFR021w | YNL008c | similarity to hypothetical protein YMR119w |
| | | | | YLR031w | similarity to hypothetical protein YMR124w |
| | | | | YPL228w | similarity to hypothetical protein YMR180c |
| | | | | YML118w | similarity to hypothetical protein YMR285c |
| | | | | YGR273c | similarity to hypothetical protein YMR295c |
| | | | | YGR283c | similarity to hypothetical protein YMR310c |
| | | | | YMR119w | similarity to hypothetical protein YNL008c |
| | | | | YIL117c | similarity to hypothetical protein YNL058c |
| | | | | YHR088w | similarity to hypothetical protein YNL075w |
| | | | | YHR131c | similarity to hypothetical protein YNL144c |
| | | | | YHR133c | similarity to hypothetical protein YNL156c |
| | | | | YDL211c | similarity to hypothetical protein YNL176c |
| | | | | YLR187w | similarity to hypothetical protein YNL278w |
| | | | | YDR214w | similarity to hypothetical protein YNL328w |
| | | | | YLR008c | similarity to hypothetical protein YNL281c |
| | | | | YEL029c | similarity to hypothetical protein YNL027w |
| | | | | YFR012w | similarity to hypothetical protein YOL019w |
| | | | | YMR063w | similarity to hypothetical protein YOL082w |
| | | | | YOL083w | similarity to hypothetical protein YOR019w |
| | | | | YDR474c | |

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|---------|--|---------|--|-----------|--|
| YKR078w | similarity to hypothetical protein YOR069w | YNL006w | similarity to Met30p | YGL164c | similarity to <i>S. pombe</i> hypothetical protein SPAC31A2.10 |
| YHR194w | similarity to hypothetical protein YOR147w | YNL293w | similarity to Mic1p and human transforming protein tre-2, and strong similarity to hypothetical protein YOL112w | YPL236c | similarity to <i>S. pombe</i> hypothetical protein SPAC3H1.13 |
| YAL028w | similarity to hypothetical protein YOR324c | | similarity to microtubule-associated ser/thr protein kinases | YAL032c | <i>FUN20</i> similarity to <i>S. pombe</i> hypothetical protein SPAC8A4.06 |
| YLR361c | similarity to hypothetical protein YOR3329c | YNR047w | similarity to microtubule-interacting protein Mhp1p and to hypothetical protein YOR227w | YEL007w | similarity to <i>S. pombe</i> pac2 protein |
| YAL034c | <i>FUN19</i> similarity to hypothetical protein YOR338w | YPL137c | similarity to mitochondrial aldehyde dehydrogenase Ald1p | YLR247c | similarity to <i>S. pombe</i> rad8 protein |
| YAL056w | similarity to hypothetical protein YOR371c | YIL151c | similarity to mouse calcium-binding protein | YJL062w | similarity to <i>S. pombe</i> YAM3 protein |
| YFR021w | similarity to hypothetical protein YPL184c | YGR058w | similarity to mouse eps15R protein | YHR009c | similarity to <i>S. pombe</i> Z66568.C protein |
| YML117w | similarity to hypothetical protein YPL219w | YBL047c | similarity to mouse Tbc2 protein | YBL051c | similarity to <i>S. pombe</i> ZK1058.5 protein |
| YGL133w | similarity to hypothetical protein YPL228w | YLR200w | similarity to mouse MHC H-2K/t-w6-linked ORF | YOR240w | similarity to ser/thr kinases |
| YGL134w | similarity to hypothetical protein YPL229w | YLO23c | similarity to mouse nuclear receptor co-repressor N-Cor | YPL150w | similarity to ser/thr protein kinases |
| YMR180c | similarity to hypothetical protein YPR114w | YCR033w | similarity to mouse nucleolin | YGR052w | similarity to ser/thr protein kinases |
| YMR181c | similarity to hypothetical protein YPR117w | YER030w | similarity to mouse putative CCAAT binding factor 1 | YAL017w | <i>FUN31</i> similarity to ser/thr protein kinases |
| YJR116w | similarity to hypothetical protein YPR125w | YDR060w | similarity to mouse putative transmembrane protein FT27 | YOR090c | similarity to ser/thr protein phosphatases |
| YLR454w | similarity to hypothetical protein YPR158w | YBR187w | similarity to mouse Surf-4 protein | YBR059c | similarity to ser/thr-specific protein kinase Pak1p |
| YLO27c | similarity to hypothetical proteins YAL018c and YOL047c | YGR284c | similarity to mouse T10 protein | YBR274w | similarity to ser/thr-specific protein kinases |
| YOL047c | similarity to hypothetical proteins YAL018c and YOL048c | YGR127w | similarity to mouse Tbc1 protein | YLR118c | similarity to several esterases |
| YMR210w | similarity to hypothetical proteins YBR177c and YPL095c | YMR192w | similarity to mouse TEG-261 protein | YMR077c | similarity to SNF7 protein |
| YML052w | similarity to hypothetical proteins YDL222c and YNL194c | YPL249c | similarity to MPA43p | YLR313c | similarity to SPA2 protein |
| YLR246w | similarity to hypothetical proteins YDR126w, YNL326c and YOL003c | YHR181w | similarity to myosins | YIR033w | similarity to Spt23p |
| YER072w | similarity to hypothetical proteins YFL004w and YPL019c | YDR109c | similarity to <i>N. crassa</i> sulphur controller-2 | YBL109w | similarity to subtelomeric encoded proteins |
| YLR072w | similarity to hypothetical proteins YFL042c, YFL043c, YDR326c and YHR080c | YLO38c | similarity to NMD and CSE1 proteins | YEL074w | similarity to subtelomeric encoded proteins |
| YGL028c | similarity to hypothetical proteins YGR279c and YMR305c | YER085c | similarity to N-terminal part of Cdc39p | YGL263w | similarity to subtelomeric encoded proteins |
| YFL042c | similarity to hypothetical proteins YHR080c, YDR326c and YLR072w | YJL112w | similarity to nucleolar Nop2p | YHL042w | similarity to subtelomeric encoded proteins |
| YER072w | similarity to hypothetical proteins YHR090c and YHR090c | YDR395w | similarity to <i>O. aries</i> arylalkylamine N-acetyltransferase | YHL043w | similarity to subtelomeric encoded proteins |
| YLR072w | similarity to hypothetical proteins YKL200c and YKL201c | YPR072w | similarity to <i>P. aeruginosa</i> alkyl sulphatase | YHL044w | similarity to subtelomeric encoded proteins |
| YGL028c | similarity to hypothetical proteins YKR089c and YOR081c | YBL024w | similarity to <i>P. ciliare</i> possible apospory-associated protein | YIL177c | similarity to subtelomeric encoded proteins |
| YFL042c | similarity to hypothetical proteins YMR063w and YFR012w | YDR071c | similarity to <i>P. denitrificans</i> cobW protein | YKL219w | similarity to subtelomeric encoded proteins |
| YNL097c | similarity to hypothetical proteins YNL019c and YNL033w | YOL164w | similarity to <i>P. falciparum</i> 41-2 protein antigen | YMR326c | similarity to subtelomeric encoded proteins |
| YJR061w | similarity to hypothetical proteins YNL033w and YNL033w | YMR099c | similarity to <i>P. falciparum</i> mature-parasite-infected erythrocyte surface antigen MESA | YLR177w | similarity to suppressor protein Gin5p |
| YMR313c | similarity to hypothetical proteins YNL032w and YNL099c | YDR472w | similarity to <i>P. falciparum</i> merozoite cap protein-1 | YNL191w | similarity to <i>Synechocystis</i> hypothetical protein |
| YOL019w | similarity to hypothetical proteins YNL032w, YNL056w and YDR067c | YNL136w | similarity to <i>P. troglodytes</i> prot GOR | YGR036c | similarity to <i>T. denticola</i> phosphatase |
| YPR027c | similarity to hypothetical proteins YNL032w, YNL056w and YDR067c | YIL010w | similarity to Pie3p and hypothetical proteins YIL002c | YDR485c | similarity to trichohyalin |
| YNL056w | similarity to hypothetical proteins YOL002c and YDR492w | YLR107w | similarity to probable transcription factor Ask10p and hypothetical protein YPR115w, and strong similarity to hypothetical protein YIL105c | YOR195w | similarity to USO1 protein |
| YNL099c | similarity to hypothetical proteins YOL003c, YLR246w and <i>C. elegans</i> hypothetical protein ZK7571 | YOL065c | similarity to protein phosphatase 2C | YPL188w | <i>POS5</i> similarity to Utr1p and hypothetical protein YEL041w |
| YNL032w | similarity to hypothetical proteins YOR3141c and YNL087w and weak similarity to synaptogamines | YNL047c | similarity to putative cell surface glycoprotein Sed1p | YLR213c | similarity to UTR2 protein |
| YOL101c | similarity to hypothetical proteins YOR3165w and YNL095c | YBR125c | similarity to putative human GTP-binding protein HSR1 | YGR189c | similarity to Utr2p |
| YOL101c | similarity to hypothetical proteins YOR385w and YMR316w | YNR038w | similarity to <i>R. corallinus</i> N-ethylmelleine chlorohydrolase trzA | YGL220w | similarity to <i>V. alginolyticus</i> bolA protein |
| YMR316w | similarity to hypothetical proteins YOR385w and YNL165w | YER150w | similarity to <i>R. fascians</i> hypothetical protein 6 | YGR203w | similarity to <i>X. laevis</i> protein-tyrosin-phosphatase cdc homologue 2 and to hypothetical protein YPR200c |
| YDR083w | similarity to hypothetical <i>S. pombe</i> protein | YGL099w | similarity to RAD5 protein | YDR284c | similarity to YDR503c |
| YDR346c | similarity to hypothetical <i>S. pombe</i> protein | YPR062w | similarity to rat branched-chain α -ketoacid dehydrogenase kinase | YEL003w | similarity to Yke2p |
| YGR272c | similarity to hypothetical <i>S. pombe</i> protein | YJL055w | similarity to rat regucalcin | YIL135c | similarity to Ymk1p |
| YLR241w | similarity to hypothetical <i>S. pombe</i> protein | YOR191w | similarity to rat synaptic glycoprotein SC2 | YOL082w | similarity to YOL083w |
| YML005w | similarity to hypothetical <i>S. pombe</i> protein | YGL059w | similarity to ribosomal protein kinases | YDR534c | similarity to YOR383c, Sta1p and pig mucin |
| YOL098c | similarity to hypothetical <i>S. pombe</i> protein | YBR053c | similarity to RNA helicases | YBL101c | similarity to YPR030w |
| YOR091w | similarity to hypothetical <i>S. pombe</i> protein D83992.G | YDL015c | similarity to RNA-binding proteins | YCR062w | similarity to Ytp1p protein |
| YOL071w | similarity to hypothetical <i>S. pombe</i> protein SPAC12B10.06c | YBR028c | similarity to <i>S. acidocaldarius</i> ac2sac protein | YFR024c-a | strong similarity hypothetical protein YHR016c |
| YFR048w | similarity to hypothetical <i>S. pombe</i> protein SPAC12G12.14 and to YDL001w and YDR282c | YLR278c | similarity to <i>S. fumigata</i> Asp FII | YPL151c | strong similarity to <i>A. thaliana</i> PRL1 and PRL2 proteins |
| YOR322c | similarity to hypothetical <i>S. pombe</i> protein SPAC1F12.05 | YPR112c | similarity to <i>S. lincolnensis</i> lmbX protein | YOR034c | strong similarity to Akr1p |
| YOR250c | similarity to hypothetical <i>S. pombe</i> protein SPAC22H10.05c | YPR016c | similarity to <i>S. pombe</i> cek1 serine/threonine protein kinase | YOR374w | strong similarity to aldehyde dehydrogenase |
| YER143w | similarity to hypothetical <i>S. pombe</i> protein SPAC22H10.05c | YIL144w | similarity to <i>S. pombe</i> hypothetical protein | YIL113w | strong similarity to <i>C. albicans</i> dual-specificity phosphatase MSG5 |
| YDR504c | similarity to hypothetical <i>T. brucei</i> protein | YKR051w | similarity to <i>S. pombe</i> hypothetical protein | YLR460c | strong similarity to <i>C. carbonum</i> toxD protein |
| YDR223w | similarity to <i>Iffh1p</i> | YOL087c | similarity to <i>S. pombe</i> hypothetical protein SPAC18B11.05 | YKL013c | strong similarity to <i>C. elegans</i> hypothetical protein |
| YAL035w | similarity to <i>Iffm1p</i> | YBR004c | similarity to <i>S. pombe</i> hypothetical protein SPAC1D4.10 | YNL288w | strong similarity to <i>C. elegans</i> hypothetical protein |
| YOR109w | similarity to inositol polyphosphate 5-phosphatases | YKR079c | similarity to <i>S. pombe</i> hypothetical protein SPAC22G7.05 | YOL077c | strong similarity to <i>C. elegans</i> K12H4.3 protein |
| YEL013w | similarity to intracellular attachment proteins | YNL308c | similarity to <i>S. pombe</i> hypothetical protein SPAC24B11.08c | YGL080w | strong similarity to <i>C. elegans</i> R07E5.13 protein |
| YPR042c | similarity to Jsn1p | YNL310c | similarity to <i>S. pombe</i> hypothetical protein SPAC24H6.02c | YPR194c | strong similarity to C-term. of <i>S. pombe</i> isp4 protein |
| YMR226c | similarity to ketoreductases | YGR125w | similarity to <i>S. pombe</i> hypothetical protein SPAC24H6.11c | YAR023c | strong similarity to Fun55p, Fun59p, YGL051w, YCR007c, YGL053w, YAR031w and YAR028w |
| YGR232w | similarity to <i>L. mactans</i> α -latroinsectotoxin | YMR075w | similarity to <i>S. pombe</i> hypothetical protein SPAC2F7.07c | YAR028w | strong similarity to Fun55p, YGL053w, YCR007c, YAR031w, Fun59p and YGL051w |
| YER010c | similarity to <i>L. pneumophila</i> dlpA protein | YDR175c | similarity to <i>S. pombe</i> hypothetical protein SPAC2F7.15 | YGL051w | strong similarity to Fun59p |
| YGR210c | similarity to <i>M. capricolum</i> hypothetical protein SGC3 | YFL047w | similarity to <i>S. pombe</i> hypothetical protein SPAC2F7.18c | YAL036c | <i>FUN11</i> strong similarity to GTP-binding proteins |
| YMR095c | similarity to <i>M. leprae</i> hisH protein | YLR023c | similarity to <i>S. pombe</i> hypothetical protein SPAC30D11.11 | YMR290c | strong similarity to helices of the DEAD/DEAH box family |
| YBR079c | similarity to <i>M. musculus</i> p162 protein | YDR180w | similarity to <i>S. pombe</i> hypothetical protein SPAC31A2.05c | YDR276c | strong similarity to <i>Hordeum</i> btl101 protein |
| YNL335w | similarity to <i>M. verrucaria</i> cyanamide hydratase, identical to hypothetical protein YFL061w | | | YDL103c | strong similarity to human AgX-1 antigen |
| YLL034c | similarity to mammalian valosin | | | YDR373w | strong similarity to human BDR-1 protein and other calcium binding proteins |
| YMR166c | similarity to members of the mitochondrial carrier protein family | | | YNR053c | strong similarity to human breast tumor associated autoantigen |
| YIR041w | similarity to members of the Srp1p/Tip1p family | | | YDL120w | strong similarity to human frataxin (Friedreich's ataxia) |

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|-----------|--|-----------|---|---------|--|
| YPL152w | strong similarity to human phosphotyrosyl phosphatase activator | YDL109c | strong similarity to hypothetical protein YGL144c | YGR004w | strong similarity to hypothetical protein YLR324w |
| YPR028w | strong similarity to human protein TB2 | YER037w | strong similarity to hypothetical protein YGL224c | YGR010w | strong similarity to hypothetical protein YLR328w |
| YCL059c | strong similarity to human Rev interacting protein Rip-1 | YOR387c | strong similarity to hypothetical protein YGL258w | YGR038w | strong similarity to hypothetical protein YLR350w |
| YLR146c | strong similarity to human spermidine synthase | YLR324w | strong similarity to hypothetical protein YGR004w | YGR056w | strong similarity to hypothetical protein YLR357w |
| YNL200c | strong similarity to human TGR-CL10C | YLR328w | strong similarity to hypothetical protein YGR010w | YKL187c | strong similarity to hypothetical protein YLR413w |
| YBL036c | strong similarity to hypothetical <i>C. elegans</i> protein | YLR350w | strong similarity to hypothetical protein YGR038w | YPR172w | strong similarity to hypothetical protein YLR456w |
| YBL078c | strong similarity to hypothetical <i>C. elegans</i> protein | YLR357w | strong similarity to hypothetical protein YGR056w | YDR438w | strong similarity to hypothetical protein YML018c |
| YMR292w | strong similarity to hypothetical <i>C. elegans</i> protein | YPL004c | strong similarity to hypothetical protein YGR086c | YBR002c | strong similarity to hypothetical protein YMR101c |
| YDR430c | strong similarity to hypothetical <i>C. perfringens</i> protein | YPR157w | strong similarity to hypothetical protein YGR141w | YKL121w | strong similarity to hypothetical protein YMR102c |
| YOR365c | strong similarity to hypothetical protein YAL053w | YHR162w | strong similarity to hypothetical protein YGR243w | YPL224c | strong similarity to hypothetical protein YMR177w |
| YHR214w-a | strong similarity to hypothetical protein YAR068w | YBR300c | strong similarity to hypothetical protein YGR293c | YOR295w | strong similarity to hypothetical protein YMR233w |
| YPR032w | strong similarity to hypothetical protein YBL106c | YHR054c | strong similarity to hypothetical protein YHR056c | YKL046c | strong similarity to hypothetical protein YMR238w |
| YDR003w | strong similarity to hypothetical protein YBR005w | YDR358w | strong similarity to hypothetical protein YHR108w | YPL264c | strong similarity to hypothetical protein YMR253c |
| YDR210w | strong similarity to hypothetical protein YBR016w | YNL116w | strong similarity to hypothetical protein YHR115c | YGR279c | strong similarity to hypothetical protein YMR305c |
| YDL012c | strong similarity to hypothetical protein YBR016w and YDR210w | YGR238c | strong similarity to hypothetical protein YHR158c | YOR385w | strong similarity to hypothetical protein YMR316w |
| YDR018c | strong similarity to hypothetical protein YBR042c | YGR243w | strong similarity to hypothetical protein YHR162w | YNL034w | strong similarity to hypothetical protein YNL018c |
| YOL092w | strong similarity to hypothetical protein YBR147w | YHR199c | strong similarity to hypothetical protein YHR198c | YNL033w | strong similarity to hypothetical protein YNL019c |
| YPL095c | strong similarity to hypothetical protein YBR177c | YHR198c | strong similarity to hypothetical protein YHR199c | YNL019c | strong similarity to hypothetical protein YNL033w |
| YPL087w | strong similarity to hypothetical protein YBR183w | YAR060c | strong similarity to hypothetical protein YHR212c | YNL018c | strong similarity to hypothetical protein YNL034w |
| YGL056c | strong similarity to hypothetical protein YBR214w | YAR068w | strong similarity to hypothetical protein YHR214w-a | YIL109c | strong similarity to hypothetical protein YNL049c |
| YGL060w | strong similarity to hypothetical protein YBR216c | YPR071w | strong similarity to hypothetical protein YIL029c | YOR086c | strong similarity to hypothetical protein YNL087w and weak similarity to synaptogamines |
| YGL107c | strong similarity to hypothetical protein YBR238c | YER067w | strong similarity to hypothetical protein YIL057c | YOR092w | strong similarity to hypothetical protein YNL095c |
| YGL101w | strong similarity to hypothetical protein YBR242w | YDL175c | strong similarity to hypothetical protein YIL079c | YOR110w | strong similarity to hypothetical protein YNL108c |
| YIL058c | strong similarity to hypothetical protein YBR270c | YIL014c-a | strong similarity to hypothetical protein YIL102c | YHR115c | strong similarity to hypothetical protein YNL116w |
| YGR293c | strong similarity to hypothetical protein YBR300c | YNL049c | strong similarity to hypothetical protein YIL109c | YDL222c | strong similarity to hypothetical protein YNL194c and similarity to YML052w |
| YDR514c | strong similarity to hypothetical protein YCL036w | YOL162w | strong similarity to hypothetical protein YIL166c | YLR144c | strong similarity to hypothetical protein YNR067c |
| YGL144c | strong similarity to hypothetical protein YDL109c | YJL038c | strong similarity to hypothetical protein YJL037w | YDR492w | strong similarity to hypothetical protein YOL002c |
| YIL079c | strong similarity to hypothetical protein YDL175c | YJL037w | strong similarity to hypothetical protein YJL038c | YPR125w | strong similarity to hypothetical protein YOL027c |
| YNL194c | strong similarity to hypothetical protein YDL222c and similarity to hypothetical protein YML052w | YBR270c | strong similarity to hypothetical protein YJL068c | YBR147w | strong similarity to hypothetical protein YOL092w |
| YBR005w | strong similarity to hypothetical protein YDR003w | YKR018c | strong similarity to hypothetical protein YJL082w | YDR391c | strong similarity to hypothetical protein YOR013w |
| YBR042c | strong similarity to hypothetical protein YDR018c | YKR021w | strong similarity to hypothetical protein YJL084c | YAL007c | strong similarity to hypothetical protein YOR018c, similarity to hamster COP-coated vesicle membrane protein |
| YLR108c | strong similarity to hypothetical protein YDR132c | YKR053c | strong similarity to hypothetical protein YJL134w | YKR089c | strong similarity to hypothetical protein YOR081c |
| YPL235w | strong similarity to hypothetical protein YDR190c | YML047c | strong similarity to hypothetical protein YJR054w | YNL095c | strong similarity to hypothetical protein YOR092w |
| YNL281w | strong similarity to hypothetical protein YDR214w | YKL200c | strong similarity to hypothetical protein YJR061w | YNL108c | strong similarity to hypothetical protein YOR110w |
| YLR225c | strong similarity to hypothetical protein YDR222w | YDR399w | strong similarity to hypothetical protein YJR133w | YLR260w | strong similarity to hypothetical protein YOR171c |
| YHR032w | strong similarity to hypothetical protein YDR338c | YMR238w | strong similarity to hypothetical protein YKL046c | YLR270w | strong similarity to hypothetical protein YOR173w |
| YHR097c | strong similarity to hypothetical protein YDR348c | YMR040w | strong similarity to hypothetical protein YKL065c | YLR284c | strong similarity to hypothetical protein YOR180c |
| YHR108w | strong similarity to hypothetical protein YDR358w | YMR102c | strong similarity to hypothetical protein YKL121w | YLR243w | strong similarity to hypothetical protein YOR262w |
| YOR013w | strong similarity to hypothetical protein YDR391c | YLR413w | strong similarity to hypothetical protein YKL187c | YMR233w | strong similarity to hypothetical protein YOR295w |
| YJR133w | strong similarity to hypothetical protein YDR399w | YJL082w | strong similarity to hypothetical protein YKR018c | YGL258w | strong similarity to hypothetical protein YOR387c |
| YOL002c | strong similarity to hypothetical protein YDR492w | YJL084c | strong similarity to hypothetical protein YKR021w | YPL279c | strong similarity to hypothetical protein YOR390w |
| YOR383c | strong similarity to hypothetical protein YDR534c and similarity to <i>L. mexicana</i> secreted acid phosphatase 2 | YJL134w | strong similarity to hypothetical protein YKR053c | YGR086c | strong similarity to hypothetical protein YPL004c |
| YPR193c | strong similarity to hypothetical protein YEL066w | YOR081c | strong similarity to hypothetical protein YKR089c | YFL004w | strong similarity to hypothetical protein YPL019c |
| YGL224c | strong similarity to hypothetical protein YER037w | YIR013c | strong similarity to hypothetical protein YLR013w | YBR183w | strong similarity to hypothetical protein YPL087w |
| YIL057c | strong similarity to hypothetical protein YER067w | YLL010c | strong similarity to hypothetical protein YLR019w | YBR177c | strong similarity to hypothetical protein YPL095c |
| YPL019c | strong similarity to hypothetical protein YFL004w | YDR125c | strong similarity to hypothetical protein YLR099c | YOR227w | strong similarity to hypothetical protein YPL137c and to microtubule-interacting protein MHP1 |
| YMR096w | strong similarity to hypothetical protein YFL059w, YNL333w, and Para rubber tree ethylene-responsive protein 1 | YDR132c | strong similarity to hypothetical protein YLR108c | YGL084c | strong similarity to hypothetical protein YPL189w |
| YHR016c | strong similarity to hypothetical protein YFR024c-a | YDR185c | strong similarity to hypothetical protein YLR168c | YGL082w | strong similarity to hypothetical protein YPL191c |
| YBR214w | strong similarity to hypothetical protein YGL056c | YDL161w | strong similarity to hypothetical protein YLR208w and to human KIAA0171 protein | YGL139w | strong similarity to hypothetical protein YPL221w |
| YBR216c | strong similarity to hypothetical protein YGL060w | YDR222w | strong similarity to hypothetical protein YLR225c | YMR177w | strong similarity to hypothetical protein YPL224c |
| YPL191c | strong similarity to hypothetical protein YGL060w | YDR213w | strong similarity to hypothetical protein YLR228c | YDR190c | strong similarity to hypothetical protein YPL235w |
| YPL189w | strong similarity to hypothetical protein YGL082w | YOR171c | strong similarity to hypothetical protein YLR260w | YMR253c | strong similarity to hypothetical protein YPL264c |
| YBR242w | strong similarity to hypothetical protein YGL084c | YOR173w | strong similarity to hypothetical protein YLR270w | | |
| | strong similarity to hypothetical protein YGL101w | YOR180c | strong similarity to hypothetical protein YLR284c | | |

| | | | | | |
|---------|---|-----------|---|---------|--|
| YOR390w | strong similarity to hypothetical protein YPL279c | YHL017w | strong similarity to putative transmembrane protein PTM1 | YLR466w | strong similarity to subtelomeric encoded proteins |
| YIL029c | strong similarity to hypothetical protein YPR071w | YHR017w | strong similarity to <i>S. douglasii</i> YSD83 | YLR467w | strong similarity to subtelomeric encoded proteins |
| YDL242w | strong similarity to hypothetical protein YPR079w | YDL219w | strong similarity to <i>S. equisimilis</i> hypothetical protein | YML132w | strong similarity to subtelomeric encoded proteins |
| YGR141w | strong similarity to hypothetical protein YPR157w | YPL118w | strong similarity to <i>S. kluyveri</i> hypothetical protein | YNL336w | strong similarity to subtelomeric encoded proteins |
| YLR456w | strong similarity to hypothetical protein YPR172w | YNR046w | strong similarity to <i>S. pombe</i> hypothetical protein SPAC31A2.02 | YNL337w | strong similarity to subtelomeric encoded proteins |
| YEL066w | strong similarity to hypothetical protein YPR193c | YNL072w | strong similarity to <i>S. pombe</i> hypothetical protein SPAC4G9.02 | YNL338w | strong similarity to subtelomeric encoded proteins |
| YGL053w | strong similarity to hypothetical proteins YAR031, YGL051w, YAR028w, Fun55p and YCR007c | YDR032c | strong similarity to <i>S. pombe</i> obr1 | YNL339c | strong similarity to subtelomeric encoded proteins |
| YMR324c | strong similarity to hypothetical proteins YBL108w, YCR103c and YKL223w | YOL010w | strong similarity to <i>S. pombe</i> SPAC12G12.06c protein | YNR077c | strong similarity to subtelomeric encoded proteins |
| YNR048w | strong similarity to hypothetical proteins YCR094w and YNL323w | YOR163w | strong similarity to <i>S. pombe</i> SPAC13G6.14 protein | YOL158c | strong similarity to subtelomeric encoded proteins |
| YNL323w | strong similarity to hypothetical proteins YCR094w and YNR048w | YOR256c | strong similarity to secretory protein SSP134 | YPL282c | strong similarity to subtelomeric encoded proteins |
| YBR016w | strong similarity to hypothetical proteins YDL012c and YDR210w | YCL024w | strong similarity to ser/thr protein kinase GNP1 | YPL283c | strong similarity to subtelomeric encoded proteins |
| YNL334c | strong similarity to hypothetical proteins YFL060c and YMR095c | YOR310c | strong similarity to SIK1 protein | YPR202w | strong similarity to subtelomeric encoded proteins |
| YPL221w | strong similarity to hypothetical proteins YGL139w and YAL053w | YDR247w | strong similarity to Sks1p | YPR203w | strong similarity to subtelomeric encoded proteins |
| YDR326c | strong similarity to hypothetical proteins YHR080c, YFL042c and YLR072w | O7535 | strong similarity to subtelomeric encoded proteins | YPR204w | strong similarity to subtelomeric encoded proteins |
| YMR251w | strong similarity to hypothetical proteins YKR076w and YGR154c | YAL068c | strong similarity to subtelomeric encoded proteins | YNR075w | <i>EDL1</i> strong similarity to subtelomeric encoded proteins |
| YGR154c | strong similarity to hypothetical proteins YKR076w and YMR251w | YBL108w | strong similarity to subtelomeric encoded proteins | YER042w | strong similarity to transcription factors and peptide methionine sulphoxide reductases |
| YKR076w | strong similarity to hypothetical proteins YMR251w and YGR154c | YBL111c | strong similarity to subtelomeric encoded proteins | YJL186w | strong similarity to Ttp1p |
| YFL060c | strong similarity to hypothetical proteins YNL334c and YMR095c | YBL112c | strong similarity to subtelomeric encoded proteins | YEL041w | strong similarity to Utr1p |
| YCR094w | strong similarity to hypothetical proteins YNR048w and YNL323w | YBL113c | strong similarity to subtelomeric encoded proteins | YCR063w | strong similarity to <i>Xenopus</i> G10 and human edg-2 protein |
| YOR230w | strong similarity to hypothetical proteins YOR229w and YPL139c | YBL113c | strong similarity to subtelomeric encoded proteins | YAR027w | <i>FUN55</i> strong similarity to YAR028w, YCR007c, YGL053w, YAR031w, FUN59p and YGL051w |
| YPL139c | strong similarity to hypothetical proteins YOR230w and YOR229w | YBR302c | strong similarity to subtelomeric encoded proteins | YAR029w | <i>FUN57</i> strong similarity to YAR031w, YGL053w, Fun55p, Fun59p and YGL051w |
| YOR229w | strong similarity to hypothetical proteins YOR230w and YPL139c | YCL073c | strong similarity to subtelomeric encoded proteins | YAR033w | <i>FUN59</i> strong similarity to YGL051w, YGL053w, YAR031w, Fun55p, YAR028w and YCR007c |
| YAL053w | strong similarity to hypothetical proteins YOR365c, YGL139w, YPL221w | YCR103c | strong similarity to subtelomeric encoded proteins | YAR031w | strong similarity to YGL053w, Fun59p, YGL051w, Fun55p, YAR028w and YCR007c |
| YPL280w | strong similarity to hypothetical proteins YOR391c, YMR322c and YDR533c | YDL248w | strong similarity to subtelomeric encoded proteins | YIL102c | strong similarity to YIL014c-a |
| YMR321c | strong similarity to hypothetical proteins YPL273w and YLL062c | YDR542w | strong similarity to subtelomeric encoded proteins | YBR025c | strong similarity to Ylf1p |
| YLL062c | strong similarity to hypothetical proteins YPL273w, weak similarity to <i>M. leprae</i> meth2 protein | YDR543c | strong similarity to subtelomeric encoded proteins | YKL065c | strong similarity to YMR040w |
| YOR391c | strong similarity to hypothetical proteins YPL280w, YMR322c and YDR533c | YDR544c | strong similarity to subtelomeric encoded proteins | YOL112w | strong similarity to YNL293w, similarity to Mic1p and human transforming protein tre-2 |
| YDR533c | strong similarity to hypothetical proteins YPL280w, YOR391c and YMR322c | YDR545w | strong similarity to subtelomeric encoded proteins | YBL106c | strong similarity to YPR032w |
| YMR322c | strong similarity to hypothetical proteins YPL280w, YOR391c and YDR533c | YEL075c | strong similarity to subtelomeric encoded proteins | YLR045c | <i>STU2</i> suppressor of a cs tubulin mutation |
| YHR069c | strong similarity to hypothetical <i>S. pombe</i> and human proteins | YEL076c-b | strong similarity to subtelomeric encoded proteins | YER120w | <i>SCS2</i> suppressor of an inositol auxotrophic and a choline sensitive mutant |
| YMR288w | strong similarity to hypothetical <i>S. pombe</i> protein | YER188c-a | strong similarity to subtelomeric encoded proteins | YCR044c | suppressor of <i>cdc1-1</i> ts growth defect |
| YER049w | strong similarity to hypothetical <i>S. pombe</i> protein YER049W | YER189w | strong similarity to subtelomeric encoded proteins | YGL083w | suppressor of GTase mutant |
| YNL240c | strong similarity to <i>K. marxianus</i> LET1 protein | YER190w | strong similarity to subtelomeric encoded proteins | YDR510w | <i>SCY1</i> suppressor of <i>mi12</i> temperature-sensitive mutation |
| YBR301w | strong similarity to members of the Srp1p/Tip1p family | YFL062w | strong similarity to subtelomeric encoded proteins | YKL124w | suppressor of <i>shr3</i> |
| YGL261c | strong similarity to members of the Srp1p/Tip1p family | YFL063w | strong similarity to subtelomeric encoded proteins | YLR197w | suppressor of toxicity of Gal4-KB |
| YGR294w | strong similarity to members of the Srp1p/Tip1p family | YFL064c | strong similarity to subtelomeric encoded proteins | YOL102c | tRNA 2'-phosphotransferase |
| YIL176c | strong similarity to members of the Srp1p/Tip1p family | YFL065c | strong similarity to subtelomeric encoded proteins | YIL080w | Ty3-2 orf C fragment |
| YKL224c | strong similarity to members of the Srp1p/Tip1p family | YFL066c | strong similarity to subtelomeric encoded proteins | YBR015c | type II membrane protein |
| YLL025w | strong similarity to members of the Srp1p/Tip1p family | YGL260w | strong similarity to subtelomeric encoded proteins | YBR066c | weak similarity to <i>A. niger</i> carbon catabolite repressor protein |
| YLL064c | strong similarity to members of the Srp1p/Tip1p family | YGR295c | strong similarity to subtelomeric encoded proteins | YKL071w | weak similarity to <i>A. parasiticus</i> nor-1 protein |
| YLR037c | strong similarity to members of the Srp1p/Tip1p family | YGR296w | strong similarity to subtelomeric encoded proteins | YJL126w | weak similarity to <i>A. thaliana</i> nitrilase 3 |
| YMR325w | strong similarity to members of the Srp1p/Tip1p family | YHL045w | strong similarity to subtelomeric encoded proteins | YCR079w | weak similarity to <i>A. thaliana</i> protein phosphatase 2C |
| YOR394w | strong similarity to members of the Srp1p/Tip1p family | YHL048w | strong similarity to subtelomeric encoded proteins | YHR143w | weak similarity to a-agglutinin core protein AGA1 |
| YEL047c | strong similarity to Osm1p | YHR217c | strong similarity to subtelomeric encoded proteins | YOR353c | weak similarity to adenylate cyclases |
| YFL059w | strong similarity to Para rubber tree ethylene-responsive protein1 | YHR218w-a | strong similarity to subtelomeric encoded proteins | YER158c | weak similarity to Afr1p |
| YNL333w | strong similarity to Para rubber tree ethylene-responsive protein 1 and identical to hypothetical protein YFL059w | YIR040c | strong similarity to subtelomeric encoded proteins | YBR074w | weak similarity to aminopeptidase Y |
| YNR065c | strong similarity to Pep1p | YJL225c | strong similarity to subtelomeric encoded proteins | YCR051w | weak similarity to ankyrins |
| YNR066c | strong similarity to Pep1p | YJR161c | strong similarity to subtelomeric encoded proteins | YNR039c | weak similarity to <i>Anopheles</i> mitochondrial NADH dehydrogenase subunit 2 |
| YER089c | <i>PTC2</i> strong similarity to phosphoprotein phosphatases | YJL225c | strong similarity to subtelomeric encoded proteins | YMR152w | weak similarity to AST1 and AST2 protein |
| YPL141c | strong similarity to protein kinase Kin4p | YJR162c | strong similarity to subtelomeric encoded proteins | YJL109c | weak similarity to ATPase Drs2p |
| YOL128c | strong similarity to protein kinase Mck1p | YKL223w | strong similarity to subtelomeric encoded proteins | YIR002c | weak similarity to ATP-dependent RNA helicases |
| YNL020c | strong similarity to protein kinase PAK1 | YKL225w | strong similarity to subtelomeric encoded proteins | YMR211w | weak similarity to β tubulins |
| YJR150c | strong similarity to proteins of the Srp1p/Tip1p family | YLR462w | strong similarity to subtelomeric encoded proteins | YEL040w | weak similarity to <i>B. subtilis</i> 1,3-1,4--glucanase |
| YDL214c | strong similarity to putative protein kinase NPR1 | YLR463c | strong similarity to subtelomeric encoded proteins | YNL203c | weak similarity to <i>B. subtilis</i> CDPdialcylglycerol-serine O-phosphatidyltransferase |
| YEL077c | strong similarity to putative purine nucleotide-binding protein YIL177c | YLR464w | strong similarity to subtelomeric encoded proteins | YDR336w | weak similarity to <i>B. subtilis</i> hypothetical protein X |
| | | | | YOR111w | weak similarity to <i>B. subtilis</i> maf protein |
| | | | | YNR074c | weak similarity to <i>B. subtilis</i> nitrite reductase (nirB) |
| | | | | YNR004w | weak similarity to bovine interferon γ |
| | | | | YNL022c | weak similarity to <i>C. burnetii</i> FMU protein |
| | | | | YGL246c | weak similarity to <i>C. elegans</i> dom-3 protein |
| | | | | YKR071c | weak similarity to <i>C. elegans</i> hypothetical protein |
| | | | | YNL207w | weak similarity to <i>C. elegans</i> hypothetical protein ZK632.3 |
| | | | | YLR242c | weak similarity to <i>C. elegans</i> R05H5.5 protein and <i>T. borreli</i> apocytochrome b |

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|---------|---|---------|--|---------|--|
| YKL037w | weak similarity to <i>C. elegans</i> ubc-2 protein | YNR061c | weak similarity to hypothetical protein YDL218w | YNR022c | weak similarity to protein phosphatases |
| YHR012w | PEP11 weak similarity to <i>C. elegans</i> Z47357_A ZK1128.1 | YOR004w | weak similarity to hypothetical protein YDR339c | YBR276c | weak similarity to protein-tyrosine-phosphatase |
| YJR024c | weak similarity to <i>C. elegans</i> Z49131_E ZC373.5 protein | YMR010w | weak similarity to hypothetical protein YDR352w | YMR241w | weak similarity to putative carrier protein RIM2 |
| YBR086c | weak similarity to calcium and sodium channel proteins | YGR223c | weak similarity to hypothetical protein YFR021w | YJR044c | weak similarity to putative transport protein YKR103w |
| YGR225w | weak similarity to Cdc20p | YGR095c | weak similarity to hypothetical protein YGR195w | YBR103w | weak similarity to Pwp2p |
| YNR051c | weak similarity to chicken nucleolin | YHR160c | weak similarity to hypothetical protein YGR239c | YMR093w | weak similarity to Pwp2p |
| YMR272c | weak similarity to cytochrome b5 | YGR239c | weak similarity to hypothetical protein YHR160c | YNR064c | weak similarity to <i>R. capsulatus</i> bchO protein |
| YEL045c | weak similarity to cytochrome c oxidase III of <i>T. brucei</i> kinetoplast | YMR299c | weak similarity to hypothetical protein YJL062w | YEL018w | weak similarity to Rad50p |
| YHR142w | weak similarity to cytochrome c oxidases | YKL041w | weak similarity to hypothetical protein YLR002w | YGL244w | weak similarity to Rad50p |
| YDL035c | weak similarity to <i>D. discoideum</i> protein tyrosine phosphatase | YMR124w | weak similarity to hypothetical protein YLR031w | YHR022c | weak similarity to RAS-related protein |
| YGL242c | weak similarity to <i>D. melanogaster</i> ANK protein | YPL109c | weak similarity to hypothetical protein YLR253w | YLR351c | weak similarity to rat β -alanine synthase |
| YPR200c | weak similarity to <i>D. melanogaster</i> cdc25 protein, and similarity to hypothetical protein YGR203w | YBR168w | weak similarity to hypothetical protein YLR324w | YCR082w | weak similarity to Rbk1p |
| YHL041w | weak similarity to <i>D. melanogaster</i> hypothetical protein 6 | YNR014w | weak similarity to hypothetical protein YMR206w | YOR246c | weak similarity to reductases |
| YOR022c | weak similarity to <i>D. melanogaster</i> probable Ca ²⁺ transporter rdgB | YER186c | weak similarity to hypothetical protein YMR316w | YJL012c | weak similarity to regulatory protein PHO81 |
| YPL229w | weak similarity to <i>D. melanogaster</i> transcription factor shn | YJR129c | weak similarity to hypothetical protein YNL024c | YDL114w | weak similarity to <i>Rhizobium</i> nodulation protein nodG |
| YGL185c | weak similarity to dehydrogenases | YJL016w | weak similarity to hypothetical protein YNL278w and YLR187w | YGR068c | weak similarity to Rod1p |
| YLR222c | weak similarity to Dip2p | YMR206w | weak similarity to hypothetical protein YNR014w | YKL107w | weak similarity to <i>S. antibioticus</i> probable oxidoreductase |
| YER041w | weak similarity to DNA repair protein Rad2p | YDL218w | weak similarity to hypothetical protein YNR061c | YER093c | weak similarity to <i>S. epidermidis</i> PepB protein |
| YOR032c | weak similarity to DNA-binding proteins | YDR339c | weak similarity to hypothetical protein YOR004w | YMR065w | weak similarity to <i>S. pombe</i> hypothetical protein SPAC13C5.03 |
| YJL162c | weak similarity to dnaJ proteins | YLR253w | weak similarity to hypothetical protein YPL109c | YGR212w | weak similarity to <i>S. pombe</i> hypothetical protein SPAC18B11.03c |
| YBR220c | weak similarity to <i>E. coli</i> ampG protein | YPR151c | weak similarity to hypothetical protein YPL159c | YER127w | weak similarity to <i>S. pombe</i> hypothetical protein SPAC18B11.06 |
| YNL217w | weak similarity to <i>E. coli</i> bis(5'-nucleosyl)-tetraphosphatase | YPL159c | weak similarity to hypothetical protein YPR151c | YBR271w | weak similarity to <i>S. pombe</i> uvi22 protein and hypothetical protein YNL024c |
| YER126c | weak similarity to <i>E. coli</i> colicin N | YGR126w | weak similarity to hypothetical protein YPR151c | YLR311c | weak similarity to <i>S. tarentolae</i> cryptogene protein G4 |
| YGL136c | weak similarity to <i>E. coli</i> ftsJ protein | YLR391w | weak similarity to hypothetical proteins YAR068w and YHR214w-a | YLR380w | weak similarity to Sec14p |
| YKL094w | YJU3 weak similarity to <i>E. coli</i> hypothetical protein | YNL024c | weak similarity to hypothetical proteins YBR271w and YJR129c | YNL231c | weak similarity to Sec14p |
| YMR155w | weak similarity to <i>E. coli</i> hypothetical protein f402 | YAL018c | weak similarity to hypothetical proteins YOL047c and YOL048c | YJL123c | weak similarity to Sec7p |
| YER152c | weak similarity to <i>E. coli</i> hypothetical protein f470 | YDR352w | weak similarity to hypothetical proteins YOL092w, YBR147w and YMR010w | YKL055c | weak similarity to short-chain alcohol dehydrogenases |
| YJR019c | weak similarity to <i>E. coli</i> thioesterase II | YDR517w | weak similarity to hypothetical <i>S. pombe</i> protein | YNR030w | weak similarity to Smp3p |
| YPR067w | weak similarity to <i>F. alni</i> nitrogen fixation protein | YCR013c | weak similarity to <i>M. leprae</i> B1496_F1_41 protein | YDR486c | weak similarity to Snf7p |
| YDR063w | weak similarity to glia maturation factor β | YPL273w | weak similarity to <i>M. leprae</i> meth2 protein, and strong similarity to hypothetical protein YLL062c | YEL015w | weak similarity to Spa2p |
| YCL028w | weak similarity to glutenins, high molecular weight subunit | YEL043w | weak similarity to Mad1p | YHR177w | weak similarity to Spombe pac2 protein |
| YPL206c | weak similarity to glycerophosphoryl diester phosphodiesterases | YBR186w | weak similarity to members of CDC48/PAS1/SEC18 family of ATPases | YBR155w | weak similarity to stress-induced Sti1p |
| YOR272w | weak similarity to GTP-binding protein β subunits | YFL046w | weak similarity to middle part of <i>C. elegans</i> myosin heavy chain A | YJR101w | weak similarity to superoxide dismutases |
| YAL048c | weak similarity to GTP-binding proteins | YOR350c | MNE1 weak similarity to mitochondrial <i>L. illustris</i> cytochrome oxidase I | YBR099c | weak similarity to <i>T. brucei</i> mitochondrion hypothetical protein 6 |
| YPL093w | weak similarity to GTP-binding proteins | YMR111c | weak similarity to MSN1 protein | YJL145w | weak similarity to <i>T. pacificus</i> retinal-binding protein |
| YGL067w | weak similarity to <i>H. influenzae</i> hypothetical protein | YMR172w | weak similarity to MSN1 protein | YJL204c | weak similarity to Tor2p |
| YLR165c | weak similarity to <i>H. influenzae</i> hypothetical protein HI0176 | YDL223c | weak similarity to mucin | YER045c | weak similarity to transcription factor Sko1p |
| YLR239c | weak similarity to <i>H. influenzae</i> lipote biosynthesis protein B | YJL036w | weak similarity to Mvp1p | YDR520c | weak similarity to transcription factors |
| YNR062c | weak similarity to <i>H. influenzae</i> L-lactate permease (lctP) homologue | YNL063w | weak similarity to <i>Mycoplasma</i> protoporphyrinogen oxidase | YHR063c | weak similarity to translational activator CBS2 |
| YNL176c | weak similarity to Hkr1p | YLR309c | IMH1 weak similarity to myosin heavy chains | YGL004c | weak similarity to Tup1p |
| YBL032w | weak similarity to hnRNP complex protein homologue YBR233w | YBR156c | weak similarity to myosins | YJR046w | weak similarity to <i>Xenopus</i> vimentin 4 |
| YER033c | weak similarity to human BRCA2 early onset breast cancer gene | YAL022c | weak similarity to Na ⁺ /H ⁺ antiporter | YOL031c | weak similarity to <i>Y. lipolytica</i> Sls1 protein |
| YJL036w | weak similarity to human cAMP response element-binding protein | YDR413c | weak similarity to NADH dehydrogenase | YLL056c | weak similarity to <i>Y. pseudotuberculosis</i> CDP-3,6-dideoxy-D-glycero-L-glycero-4-hexulose-5-epimerase |
| YJL148w | weak similarity to human chromatin assembly factor I | YKR030w | weak similarity to NADH dehydrogenases | YCL063w | weak similarity to yeast translation regulator Gcd6p |
| YDR030c | weak similarity to human CSA protein | YPR174c | weak similarity to Nbp1p | YGR110w | weak similarity to YLR099c and YDR125c |
| YGL243w | weak similarity to human double-stranded RNA adenosine deaminase | YKR075c | weak similarity to negative regulator Srn1p/Hex2p | YDR090c | weak similarity to Yro2p |
| YJL091c | weak similarity to human G protein-coupled receptor | YCR047c | weak similarity to N-methyltransferases | YIL044c | weak similarity to zinc-finger protein GCS1 |
| YER063w | weak similarity to human heterogeneous ribonuclear particle protein U | YJL131c | weak similarity to non-epidermal <i>Xenopus</i> keratin, type I | YLR040c | weak similarity to hypothetical protein YL011w |
| YJR002w | weak similarity to human kinesin-related protein CENP-E | YHL024w | weak similarity to nuclear protein NOP4 | YNL212w | weak similarity to <i>C. cardunculus</i> cypr4 protein |
| YMR029c | weak similarity to human nuclear autoantigen | YLR126c | weak similarity to <i>P. aeruginosa</i> anthranilate synthase component II | | |
| YCL045c | weak similarity to human ORF | YHL021c | weak similarity to <i>P. aeruginosa</i> γ -butyrobetaine hydroxylase | | |
| YJL057c | weak similarity to human P1/eIF-2a protein kinase | YMR009w | weak similarity to <i>P. aeruginosa</i> regulatory protein mmsR | | |
| YNR008w | weak similarity to human phosphatidylcholine-sterol O-acyltransferase | YGR276c | weak similarity to <i>P. troglodytes</i> GOR protein | | |
| YJL132w | weak similarity to human phospholipase D | YER059w | weak similarity to Pho80p | | |
| YLL037w | weak similarity to human platelet-activating factor receptor | YOR281c | weak similarity to phosphoducins | | |
| YHR090c | weak similarity to human retinoblastoma binding protein 2 | YMR221c | weak similarity to photosystem II protein D2 | | |
| YOR064c | weak similarity to human retinoblastoma binding protein 2 | YBR094w | weak similarity to pig tubulin-tyrosine ligase | | |
| YMR131c | weak similarity to human retinoblastoma-binding protein | YOR287c | weak similarity to PITSLRE protein kinase isoforms | | |
| YLR272c | weak similarity to hypothetical human ORF | YBR151w | weak similarity to potato sucrose cleavage protein | | |
| YDL111c | weak similarity to hypothetical human protein | YML050w | weak similarity to potato sucrose cleavage protein | | |
| YCR059c | weak similarity to hypothetical protein YDL177c | YGR262c | weak similarity to protein kinases | | |
| | | YPR106w | weak similarity to protein kinases | | |

Genetic and physical maps of *Saccharomyces cerevisiae*

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Genetic and physical maps for the 16 chromosomes of *Saccharomyces cerevisiae* are presented. The genetic map is the result of 40 years of genetic analysis. The physical map was produced from the results of an international systematic sequencing effort. The data for the maps are accessible electronically from the *Saccharomyces* Genome Database (SGD: <http://genome-www.stanford.edu/Saccharomyces/>).

During the past 40 years, 11 compilations of mapping data for the yeast *Saccharomyces cerevisiae* have been made by R. K. Mortimer and colleagues^{1–11}. The last such compilation¹¹ included mapping data to 1991, and contained for the first time the results of physical as well as genetic mapping methods. Here we present the twelfth, and probably the last, such compilation. These final maps are based on the genetic information accumulated over the years^{1–11} and, for the physical mapping data, on an entirely new set of data: the complete genomic sequence of *S. cerevisiae*.

The genetic and physical maps were derived from two entirely different types of data. Genetic distances between genes were determined by tetrad analysis. Distances for gene–gene and gene–centromere linkages are expressed in centimorgans (cM) and were calculated using a maximum-likelihood equation¹² which yields values for map distance, an interference parameter, and error calculations for these two parameters. Mapping results on more than 2,600 named genes are presented. Physical distances are calculated directly from the complete DNA sequence. The precise values of all parameters (both tetrad analysis results and chromosomal base-pair coordinates) are available from the SGD.

Associations between open reading frames (ORFs) and corresponding mutations were made using a set of hybridization filters, originally produced by L. Riles and M. Olson¹³, which are now available from the American Type Culture Collection (<http://www.atcc.org/>). Other such associations were made by complementation experiments using cloned DNA fragments and/or sequence analysis of mutants. The data for some of these associations are published, but the documentation for all of them can be found on SGD.

Now that the entire yeast genome sequence is available, most revisions of the map will consist of associations between a biological function and an ORF. These associations will often involve the study of mutants of the gene. In the past, such an association invariably resulted in the naming of the gene; this process is likely to continue until all of the genes have been associated with a function and have thereby acquired a name. Because the genetic and physical maps are unlikely to change significantly, we see no need for any future publications; rather, we expect the electronic version of the maps to evolve into increasingly accurate guides to *S. cerevisiae* biology.

The maps shown here are also available in a continually updated electronic form from the SGD (<http://genome-www.stanford.edu/Saccharomyces/>), which will also provide directions to other useful information (gene names, aliases, phenotypes, mapping data, protein information, and curated compilations of published literature about genes). □

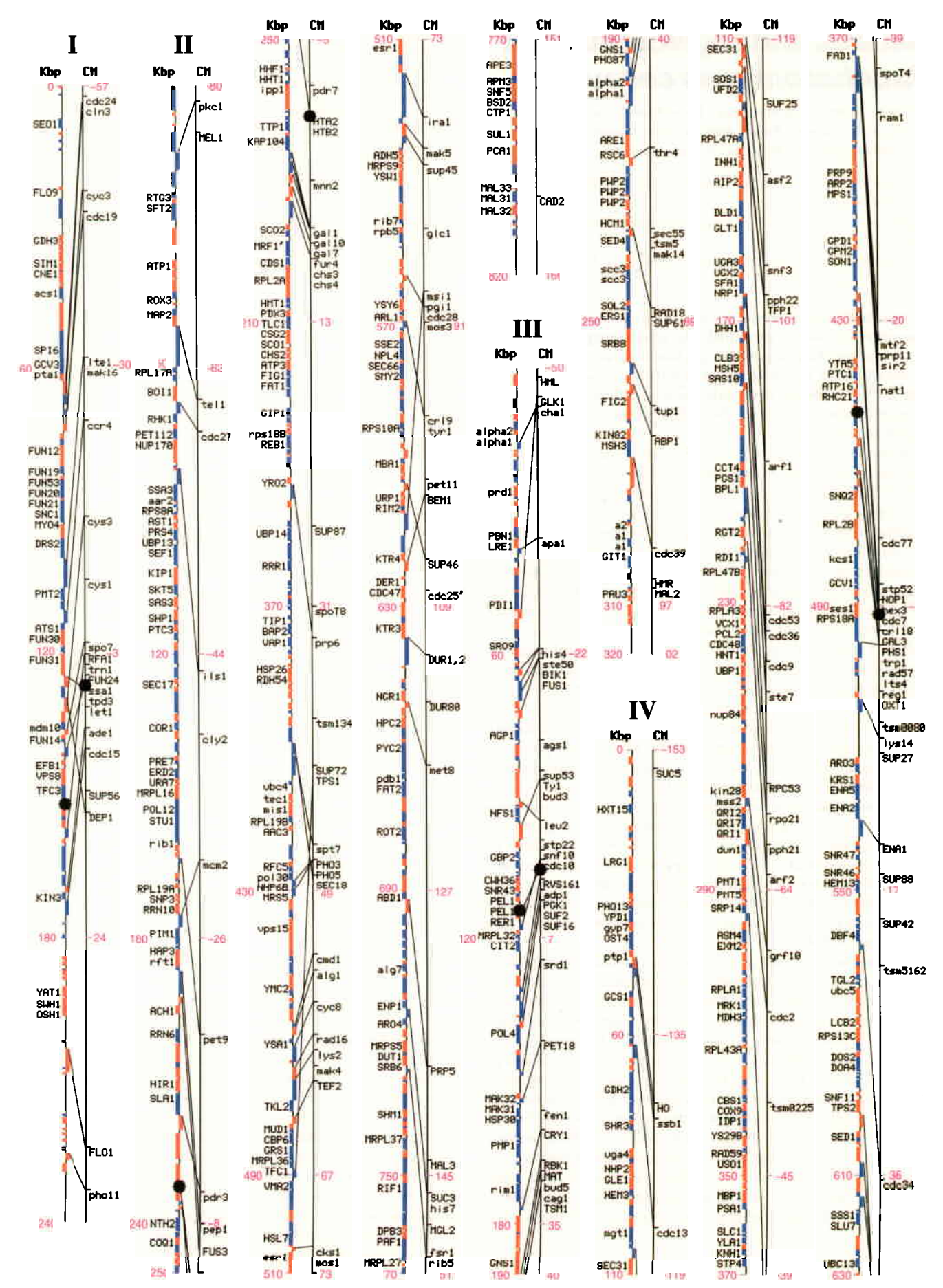
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Figures I–XVI (overleaf, pages 68–73): Genetic and physical maps, and their correlations, of the 16 *Saccharomyces cerevisiae* chromosomes. A parallel comparison of the physical map (left, in kilobase pairs) and the genetic map (right, in centimorgans) of each of the 16 chromosomes is illustrated. The information in this figure is available on the *Saccharomyces* Genome Database (<http://genome-www.stanford.edu/Saccharomyces/>). The physical map consists of coloured boxes that indicate ORFs. ORFs on the Watson strand (left telomere is the 5' end of this strand) are shown as red boxes, those on the Crick strand as blue boxes. Where it has been defined, the gene name of an ORF is indicated. The genetic map is based on data collected since 1991 by the SGD project, as well as on earlier data^{1–11}. Horizontal tick marks on the right of the genetic map line indicate positions of genes. Lines connect genetically mapped genes with their ORF on the physical map. A single name is listed for known synonyms.

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VII

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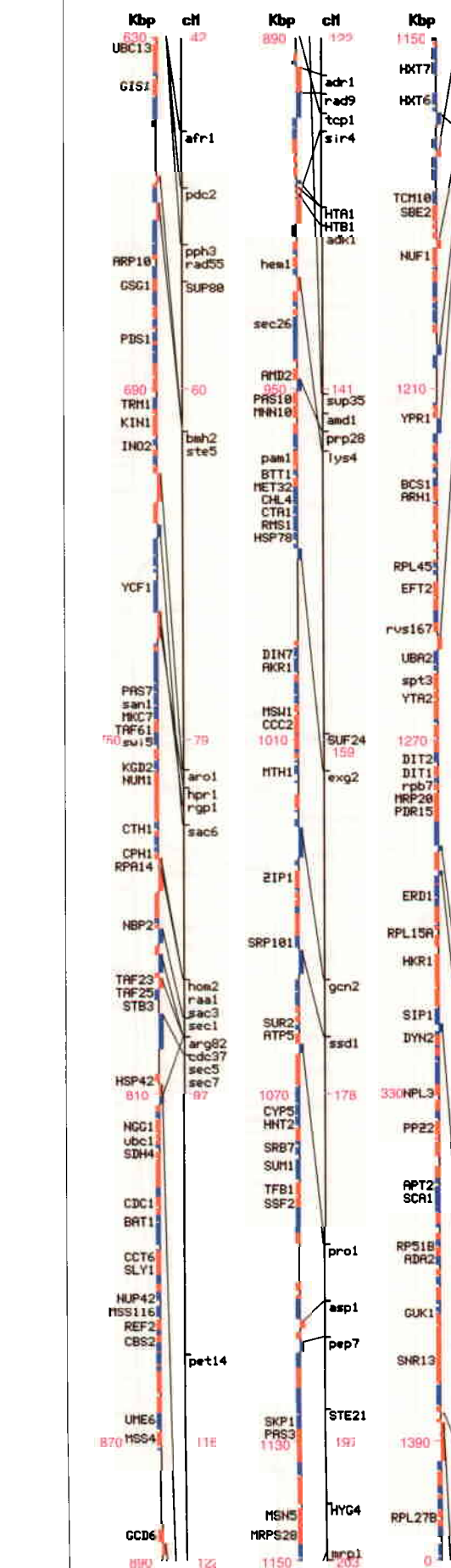
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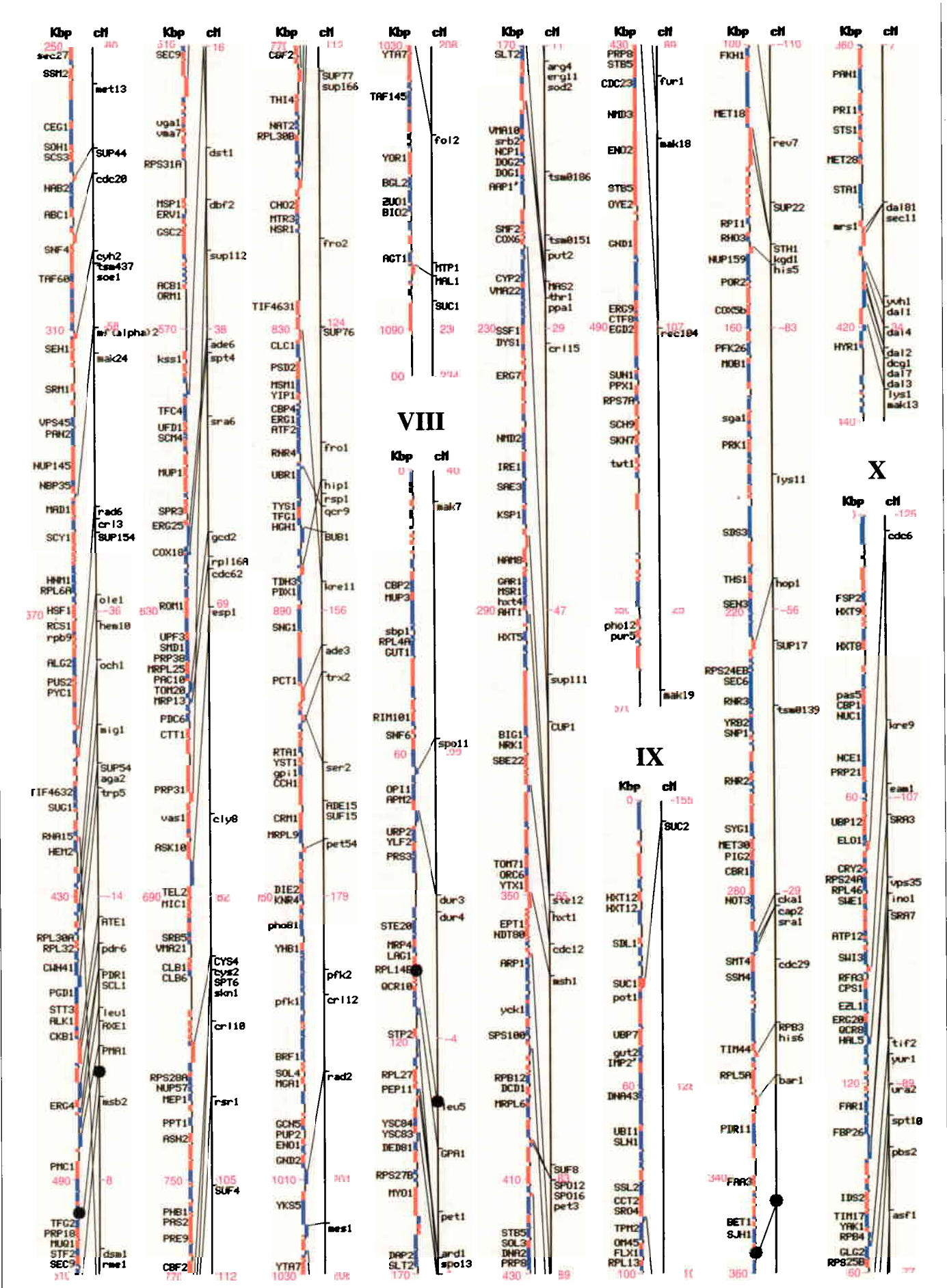
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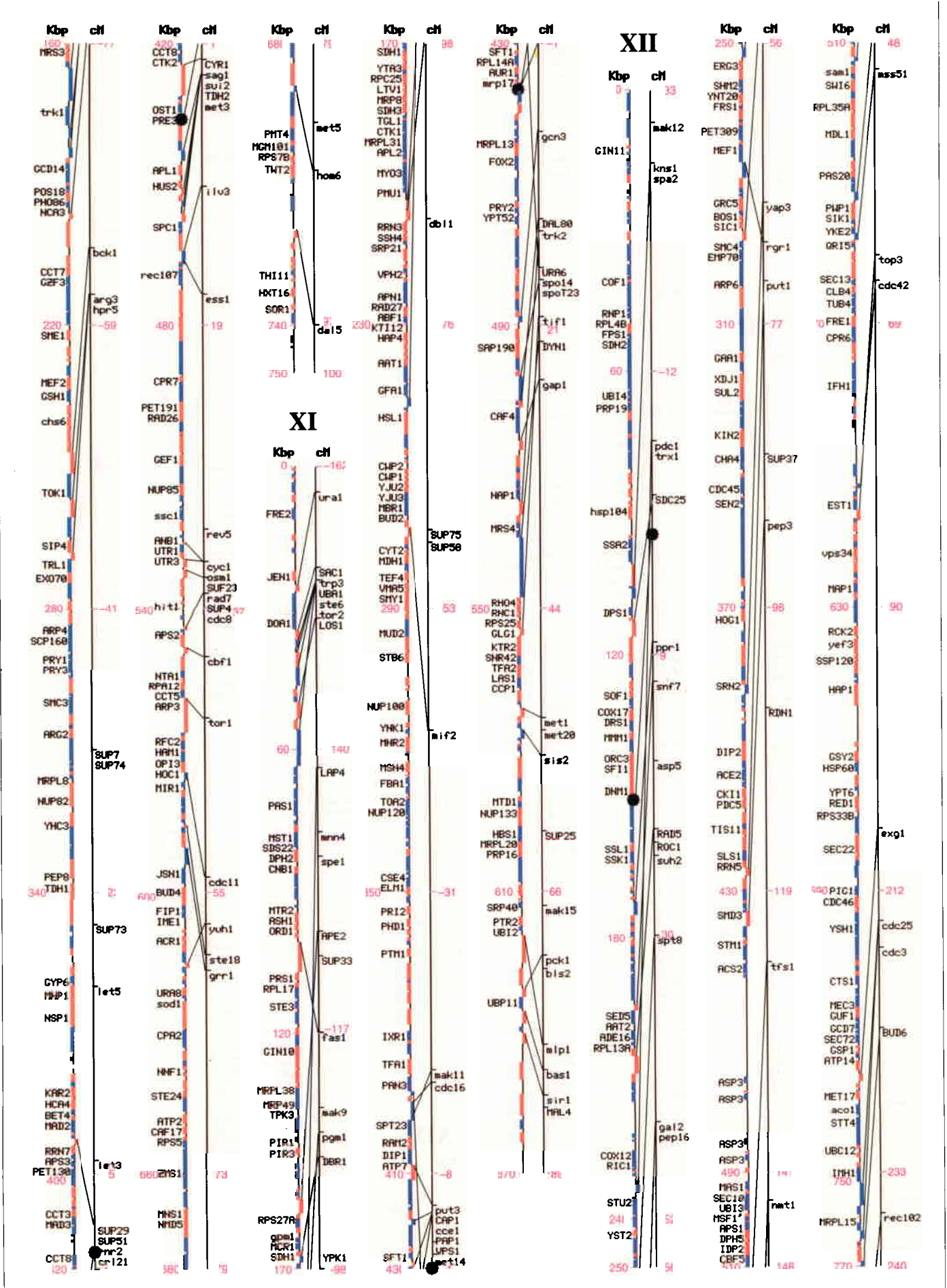


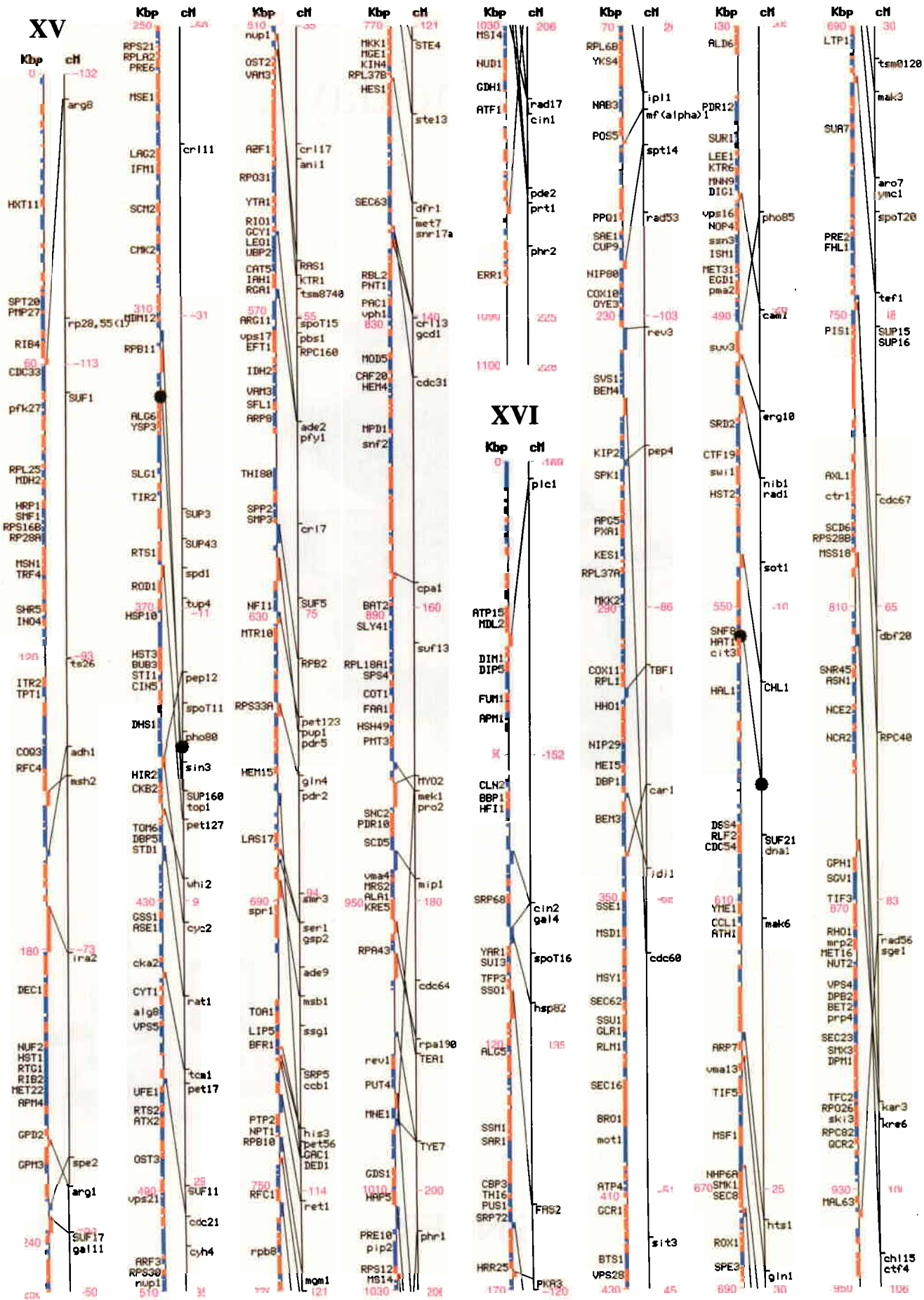
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The nucleotide sequence of *Saccharomyces cerevisiae* chromosome IV

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The complete DNA sequence of the yeast *Saccharomyces cerevisiae* chromosome IV has been determined. Apart from chromosome XII, which contains the 1–2 Mb rDNA cluster, chromosome IV is the longest *S. cerevisiae* chromosome. It was split into three parts, which were sequenced by a consortium from the European Community, the Sanger Centre, and groups from St Louis and Stanford in the United States. The sequence of 1,531,974 base pairs contains 796 predicted or known genes, 318 (39.9%) of which have been previously identified. Of the 478 new genes, 225 (28.3%) are homologous to previously identified genes and 253 (32%) have unknown functions or correspond to spurious open reading frames (ORFs). On average there is one gene approximately every two kilobases. Superimposed on alternating regional variations in G+C composition, there is a large central domain with a lower G+C content that contains all the yeast transposon (Ty) elements and most of the tRNA genes. Chromosome IV shares with chromosomes II, V, XII, XIII and XV some long clustered duplications which partly explain its origin.

The technique of determining the DNA sequence of large genomes has been unchanged for 21 years¹. Sequencing the yeast genome required considerable organization by the European Union, which initiated the grouping of 35 laboratories to sequence the first yeast chromosome² and coordinate an international effort to sequence the others. Chromosome IV had already been characterized both physically and genetically^{3,4}, and our sequence data presented are in good agreement with these preliminary data.

The average base composition of chromosome IV is 37.9% G+C, which is lower than for most of the yeast chromosomes (for example, 38.5% for chromosome III (ref. 2) and 38.3% for chromosome II (ref. 5)). Along the 1,513,914 base pairs of the chromosome there are alternating regions about 50 kilobases long of high and low G+C content (Fig. 1). This periodicity is not clearly associated with a variation in gene density, as has been observed for some other chromosomes^{6,7}. The central domain of chromosome IV (coordinates 500,000 to 1,215,000) has a much lower G+C value (37.4%) than the two flanking regions (38.2%); a similar observation has been made for the much smaller chromosome VI (ref. 8).

The low G+C content of the central domain seems to be correlated with the presence of Ty elements. All nine Ty1 or Ty2 elements, including a truncated form of Ty1, are localized between coordinates 450,000 and 1,190,000 (Fig. 1). Yeast transposons seem to insert into specific chromosomal regions^{9,10} where they are localized preferentially upstream of tRNA genes, as they might interact with the RNA polymerase III machinery¹¹. The density of tRNA genes in the central domain of chromosome IV is twice that in the flanking regions in which no Ty elements are found. A total of 27 tRNA genes are localized on each strand of the chromosome, 17 of which are located in the central domain. Of the 27 tRNA genes, 18 are in the vicinity of long terminal repeats (LTRs). Thus most of the tRNA genes, LTRs and Ty elements,

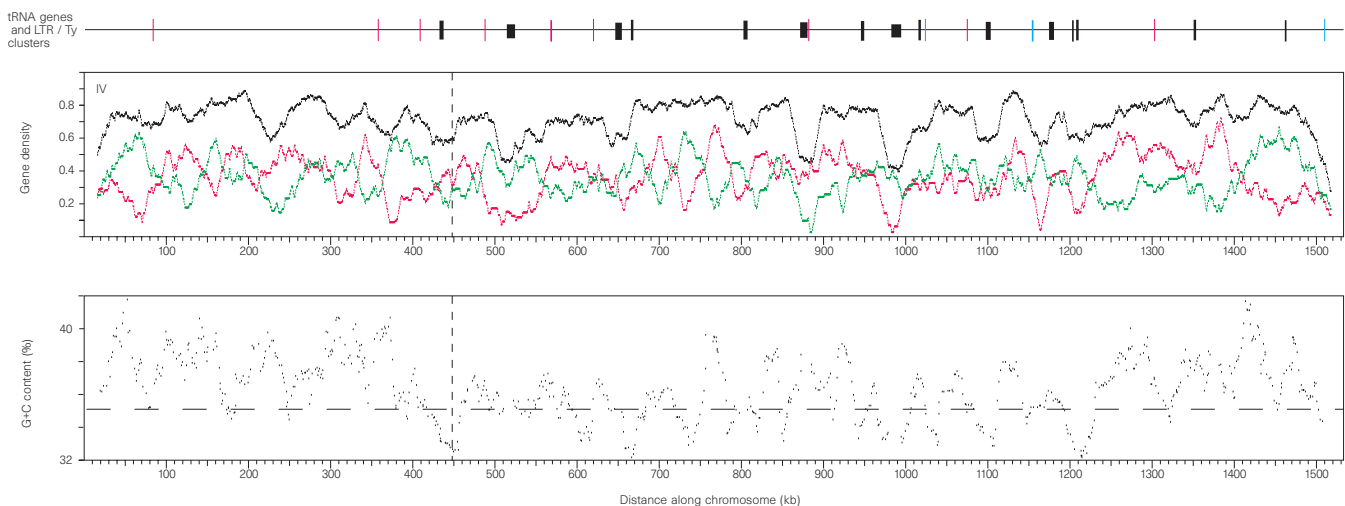


Figure 1 Overall molecular architecture of chromosome IV shows positions of tRNA genes, solo LTR or Ty elements (thin vertical lines), or clusters of them (thick vertical lines), along the chromosome map. Panels show variation of gene density (top) and base composition (bottom) along the sequence-based map of chromosome IV (scale in kilobases from the left telomere). Vertical broken lines indicate the centromere. Gene density is expressed as the probability for each nucleotide to be part of an ORF,

and was calculated using sliding windows of 30 kb (in steps of 0.5 kb) for the Watson strand alone (red line), the Crick strand alone (green line), and the sum of both (black line). G+C content was calculated from the silent positions of codons using a sliding window of 13 consecutive ORFs; the horizontal broken line indicates average G+C content (%) at silent positions of codons.

together with a lower G+C content, are found in this central domain. In chromosome II the 13 tRNA genes and three Ty elements are in AT-rich regions⁵.

The left telomere of chromosome IV is very similar to other yeast telomeres. Adjacent to the C₁₋₃A repeat are the usual STR-A, STR-B, STR-C, STR-D and the core X elements (435–904) shared by most of the telomeres¹². The left end of chromosome IV shares with the right end of chromosome X a large, nearly identical block of sequence similarity more than 19 kilobases long. This duplication includes five ORFs, which code for almost identical products. Indeed the sequences are so similar that we needed to exclude the possibility of contamination of the cosmid contig of chromosome IV by DNA sequences from chromosome X. To confirm our data, we established the genomic sequence of the junction between the duplicated sequence and the rest of the chromosome. Such subtelomeric duplications have often been observed in the yeast genome, suggesting either recent or continuous exchange of genetic information¹³. The right telomere has a less conventional structure with an internal TG₁₋₃ repeat.

Using the classical definition of ORFs (one ATG codon followed by at least 99 sense codons), 776 ORFs were recorded in the chromosome; there are also 20 ORFs shorter than 99 amino acids long, making a total of 796 ORFs. Small ORFs of between 25 and 99 codons were extracted and analysed for different properties (codon usage, homologies and ATG environment) to determine their function⁷; 15 had either a putative translation product that is homologous to proteins of other genes, or a codon adaptation index (CAI) greater than 0.2. Five other short ORFs, longer than 91 amino acids, are thought to be 'questionable'. These results support the choice of the threshold of more than 99 codons¹⁴, but show that some short ORFs must be considered¹⁵. Moreover, this evaluation has to take into account that at least 7% of the ORFs can be considered 'questionable'⁶. Disregarding the retrotransposons, this corresponds to a gene density of one ORF per 2,000 base pairs. The G+C regions correlate roughly, although not precisely, with the regions of increased density. There are approximately equal numbers of ORFs on the DNA strands (387 on the Watson and 409 on the Crick). However, the gene density is clearly not uniform on a given DNA strand (Fig. 1). This marked preference for an arrangement in which genes are on the same strand has already been observed for other chromosomes^{16,17} for which long runs of genes on one strand could be observed. Such gene-rich regions are mainly visible at the two ends of the chromosomes, with more uniform gene density in the central part of the chromosome (550,000 to 1,200,000).

Although not as great as predicted by an approximate calculation¹⁸, the high number of Ty and LTR elements in that region should give rise to large numbers of genome rearrangements (inversions, deletions and reciprocal translocations), which could explain this difference with the flanking regions. The construction of gene arrangement has led to general features that probably reflect important functional constraints. Thus the size of regions between ORFs is clearly dependent on the orientation of the flanking genes. In the case of divergent promoters, the mean size is 744 base pairs, whereas it is just 324 bp for convergent terminators. An intermediary situation (593 bp) is found for terminator–promoter combinations. This striking difference in inter-ORF size is probably due to the sequence requirements in the promoter regions for the regulation of gene expression. Nevertheless, the mean size of the inter-ORF region in the case of head-to-head gene orientations is small, and suggests that many divergent genes share common regulation signals. The *GAL1–GAL10* promoter was the first to be described¹⁹, but many other candidates for common regulation have been revealed by the systematic genome sequencing.

Based on the canonical sequences known to control the splicing process, 30 introns can be identified in genes coding for proteins²⁰. This represents 4% of genes having an intron in their ORF or in the 5' untranslated region, a figure close to the situation for the genome as a whole. Of these intron-containing genes, 12 code for ribosomal proteins, three for proteins of the actin family, three for proteins involved in the ubiquitin-dependent protein degradation system, and the rest are distributed between genes that do not necessarily have a high CAI.

Although dependent on the criteria used to estimate the significance of sequence similarities, roughly 30% of the 796 ORFs of chromosome IV are orphans²¹ with no sequence relatives in the available databases. This is one of the most exciting findings from the systematic sequencing approach of the yeast genome. Future work will tell us whether some of these genes are really 'yeast specific' and why they have escaped detection by the genetic approaches. The number of sequence orphans will no doubt decrease with the arrival of new sequence data²². As an example from chromosome IV, the ORF YDL120w, which had no relatives, is homologous to the human gene recently discovered to be involved in Friedreich's ataxia²³.

Chromosome IV is the longest chromosome in terms of coding sequences, and so might be expected to have features that are scattered on the other chromosomes. One of these features might be the non-uniform organization of the chromosome. The central domain (from 500,000 to

1,250,000) which makes up half of chromosome IV has several distinctive features. First, as with most of the chromosomes, it has more or less regularly spaced regions rich in G+C, but its central domain has a lower G+C content. Second, this central domain contains all of the Ty and most of the LTR elements found on the chromosome. Third, the central domain also contains 18 of the 27 tRNA genes, so its tRNA gene density is twice that of the rest of the chromosome. Finally, the DNA strand distribution of the ORFs is different in the central region when compared with that of the flanking regions. The ORF arrangement of this region might result from a greater genetic plasticity.

Analysis of structural relationships inside the yeast genome might provide an insight into eukaryotic genome organization and evolution. Redundancy is one of the most salient features of the yeast genome structure²⁴, and the DNA sequence of the whole yeast genome reveals several types of redundancy, probably originating from different biological processes. The most common form of redundancy involves individual genes that have a homologue in the genome; about 20% of the genes of chromosome IV are in this class. Second, there are clusters of very similar copies of a gene, often arranged in tandem; for example, there are five copies of the *ENAI* (or *PMR2*) gene on chromosome IV. Third, subtelomeric duplications are frequent and involve large regions of chromosomes that are very similar in both coding and non-coding regions¹³. Finally, clustered duplications are characterized by clusters of homologous genes in the same order, usually in the same orientation, and interspersed by long DNA fragments. Such paralogous regions have already been described between chromosomes III and XIV (ref. 25) and between chromosomes V and X (ref. 26). Only in the case of the duplication between chromosomes III and XIV is the gene order conserved. It is 15 kilobases long and contains four genes. The clustered duplications on chromosome IV are made up of at least 336 kilobase pairs, including 49 pairs of homologous genes. Chromosome IV shares large ordered cluster of homologous genes with chromosomes II, V, VIII, XII and XIII (ref. 27). A careful analysis of these duplications will no doubt tell us a great deal about the evolution of the yeast genome. In the largest interchromosomal clustered duplications, involving chromosome IV (coordinates 449,752–569,763) and chromosome II (238,164–407,122), the 18 gene pairs are all transcribed in the same direction. When known, most of the genes from a pair code for proteins with homologous but not identical functions (for example *GALI* and *GAL3*)¹⁹. Homologous genes from a clustered duplication can also be completely identical or totally different in their function. An extreme case of divergence involves YDR037w, which codes for a lysyl tRNA synthetase, and YBR060w_A, which is part of chromosome II, in which many stop codons interrupt an ORF, of which parts are homologous to YDR037w. Such a pseudogene could not be detected by searching the DNA sequence of chromosome II, as it has very short ORFs and no ATG codon. To our knowledge, this is the most degenerated yeast pseudogene yet discovered. This observation suggests that similar degenerated pseudogenes may have escaped previous analyses, and hence that the total number of pseudogenes may be underestimated.

A pair of genes from a clustered duplication can also differ in the presence of an intron. Both YDR055w and YBR078w are homologous to the gene *SPS2*, but only YBR078w has an intron, and the CAI of the two genes differs from 0.27 (YDR055w) to 0.61 (YBR078w), suggesting an unusual evolutionary process. The compared analysis of the interspersed DNA fragments is also very informative. For example, chromosome II has a Ty element where an LTR element is present at the equivalent position on chromosome IV, suggesting that the Ty element was lost from chromosome IV after the duplication process.

The greatly different degrees of similarities between the different gene pairs composing a duplicated region indicate that at least some of the duplications have evolved at very different rates, suggesting in some cases that gene conversion processes²⁸ have interfered with slower evolutionary processes. A careful quantitative analysis of the relative evolution rates of the different elements will be required to establish a chronological order of the different events. Nevertheless, evidence suggests that a first duplication event has been followed by the dispersal of the duplicated elements by the insertions of DNA fragments of various sizes and gene composi-

tions. Most of these clustered duplications in chromosome IV are localized in the pericentromeric region. The centromere itself is included in the longest duplicated region, which occurs between chromosomes IV (coordinates 450,000–570,000) and II (238,000–407,000). Similar localizations of clusters have already been noticed on other chromosomes^{7,17,25}. To explain the proximity of the centromere, it was suggested that the gene dispersion of the initial cluster of duplicated genes might be slower in the centromeric regions than nearer the telomere owing to the adverse effects of rearrangements on chromosome segregation²⁵. Alternatively, the centromeric duplications might have been essential steps in the construction of the yeast genome²⁵. In agreement with these ideas, the central domain of chromosome IV contains few traces of clustered redundancies, perhaps because of its genetic plasticity. These preliminary observations indicate that the availability of the complete sequence of the yeast genome will allow a greater understanding of the processes involved in creating the genome architecture. □

Methods

The sequence was assembled from a set of 44 partly overlapping cosmids and lambda phages from two independent contigs of chromosome IV. The 650-kb cosmid contig corresponding to the left part of chromosome IV was constructed mainly from a specific cosmid library obtained from a gel-purified chromosome portion (J. D. H. *et al.*, unpublished), and a few other cosmid clones from this contig came from a library²⁹. The rest of the chromosome sequence was established from a cosmid-lambda phage library (L. Riles & M. Olson, unpublished, and ref. 30). The two cosmid contigs were made from two closely related yeast strains: AB972, derived from S288C³⁰, and FY1679, a diploid strain issued from the cross between FY23 and FY73, both of which are isogenic with S288C except for the markers indicated. Sequence analysis of a large overlapping fragment (of about 170 kb) confirmed that the strains AB972 and FY1679 are very similar, as the number of base differences was below the estimated error rate. Only the extreme left telomeric regions of the two strains clearly differ, probably in the number of their TG₁₋₃ repeats (C. B. and C. J., unpublished). The telomeres were isolated independently and sequenced from a plasmid clone generated by integration at the TG₁₋₃ repeats of the telomere, followed by excision of the plasmid and capture of the flanking sequences¹⁵. Two gaps in the 650-kb left cosmid contig (constructed from the strain FY1679) were filled with lambda clones from the library constructed from AB972. They correspond to the regions 9,756–11360 and 363,100–368,150. The left 600-kb region was sequenced according to the rules followed by the European consortium and the 20 cosmids and phages were distributed to 18 contractors, whereas the central part was sequenced by the Sanger Centre (EMBL database SCCHRIV, accession no. Z71256); the rest of the chromosome was sequenced by groups from Washington University in St Louis and Stanford University in the United States.

There were very few base differences in the overlapping fragment sequenced in parallel by the Sanger Centre and by the European consortium, demonstrating that both approaches are reliable. However, a verification procedure was necessary because of the greater heterogeneity of the European approach. This was done on 25 regions of the left part of the chromosome, according to the protocol of G. V. (manuscript in preparation). This allows direct polymerase chain reaction (PCR) sequencing of a 300-bp region of the yeast genome limited by two previously designed oligonucleotides. We could thus correct a sequence in which a bacterial transposon had been inserted during the cloning process but no real sequence error could be detected at this final step of the sequencing project. Sequence errors could only be corrected after examination of the raw sequence data. From these data, the error rate of this part of the yeast chromosome IV sequence presented is less than four errors per 10 kb. In the central part of the chromosome the error rate is estimated as less than one error per 10 kb. Specific strategies were developed to sequence difficult parts of the chromosome. Thus, for example, to finish the regions between the two transposons located in cosmid 8142 (<http://www.sanger.ac.uk/~yeastpub/svw/sequencing.html>), a PCR product covering this region from strain MCYC2576 was sequenced. This strain, a gift from E. Louis, did not contain the transposons.

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The nucleotide sequence of *Saccharomyces cerevisiae* chromosome V

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Here we report the sequence of 569,202 base pairs of *Saccharomyces cerevisiae* chromosome V. Analysis of the sequence revealed a centromere, two telomeres and 271 open reading frames (ORFs) plus 13 tRNAs and four small nuclear RNAs. There are two Ty1 transposable elements, each of which contains an ORF (included in the count of 271). Of the ORFs, 78 (29%) are new, 81 (30%) have potential homologues in the public databases, and 112 (41%) are previously characterized yeast genes.

As part of an international collaborative effort to sequence the total genome of the yeast *Saccharomyces cerevisiae*, we have deduced the DNA sequence of 569,202 base pairs of yeast chromosome V. We used an overlapping set of recombinant yeast cosmid and lambda clones that together cover the entire chromosome (except for the extreme ends of the telomeres). A line drawing of chromosome V and the identification of the

recombinant DNAs sequenced are shown in Fig. 1. The sequence was broken arbitrarily into 11 slightly overlapping pieces for ease of handling and deposited in Genbank (see Fig. 1 for accession numbers).

Sequencing was accomplished in two phases: the 'shotgun' phase, using dye-primer chemistry, and the 'finishing' phase, using the polymerase chain reaction (PCR) and dye-terminator chemistry. There were no gaps in the sequence at the end of shotgun sequencing and assembly. The assembled, continuous sequence of chromosome V has 569,202 bp, starting from the guanine residue of the *Sau3A* site on the left vector boundary of the leftmost clone (1160 in Fig. 1). The 569-kilobase sequence is based on the results from 32,631 individual lanes of sequencing gels, or reads. The average depth of coverage was 12.5-fold. The minimum acceptable coverage was three, with at least one read from each strand.

After shotgun sequencing and assembly, problems remained in the sequence at a frequency of (roughly) two per kilobase and were of several types. They included the inability to count unambiguously the number of repeating units, such as poly (dA), and guanine compressions. There were also small regions in which only one of the two strands had been sequenced. These difficulties were resolved during the finishing phase.

After finishing, the 569-kb contig was checked against three external sets of data. First was the use of tetrad segregation data to derive a genetic map for yeast¹. The chromosome V gene order based on DNA sequence was in complete agreement with the tetrad segregation data. There were two locations on the genetic map (*CENV* at 151 kb and *PRO3* at 200 kb) where closely spaced loci had been mapped against distant markers and not against each other, resulting in ambiguities of relative locus order¹, which were resolved using the DNA sequence. The gene order across the centromere is *GLC3* tRNA-Arg *GCNA* *CENV* *MNM1*. In the region of *PRO3*, at 200 kb, the gene order is *PRO3* *GPA2* *GCD11* *CHO1* *GAL83*. Second, our sequence was compared to the *S. cerevisiae* sequences already deposited in Genbank, using both the FASTA and BLAST programs^{2,3}. In the rare cases of sequence difference, we re-examined our trace files. Remaining ambiguities were resolved using the same methodology as finishing. Third, we checked our data against the primary *EcoRI*/*HindIII* double-digestion fragment maps of the recombinant yeast DNAs⁴. Our sequence was examined for *EcoRI* and *HindIII* cleavage sites. Of 534 mapped fragments, there were only five discrepancies, which is a tribute to the care taken in preparing the cleavage sites map⁴. The five apparent discrepancies between the double-digest map⁴ and our sequence are: the map had doublets where the sequence predicts singlets after bases 272, 193; 280,936; and 441,102; the map has a fragment that was not found in the sequence after base 414,946; and the sequence is missing a cleavage site after base 506,807.

We examined all six possible reading frames of the 569-kb sequence for ORFs of at least 300 bp that began with a start codon and ended with a stop codon. As a special case, an ORF could be interrupted if there were yeast splice donor/acceptor/branchpoint sequences present at the appropriate intervals. The remaining sequence was examined using FASTA and BLAST for homology to sequences in the public databases. This enabled us to find small ORFs, as well as the centromere, 13 tRNAs, two Ty1 elements (which each contain an ORF), four small nuclear RNAs, many delta and delta-like elements, and the highly conserved X and Y' sequences characteristic of yeast telomeres (see refs 5, 6) at the far left and right ends.

Initially, 271 ORFs were identified in the 569-kb sequence, although this number has changed as evaluation continued. The 271 ORFs make up roughly 70% of the sequence, with an average of 2.1 kb per ORF. The 'average' ORF (1.4 kb) encodes 475 amino acids. Of the ORFs, 112 (41%) have been characterized previously, 81 (30%) have apparent homologues in the public databases, and 78 (29%) are new; six (2%) are spliced. Of the 81 apparent homologues, 55 of these are to other *S. cerevisiae* sequences.

The fractional G+C content of the 569,202 bp of chromosome V is 0.384. The combined ORF DNAs have a fractional G+C content of 0.401, and the combined 'non-ORF' DNA has a G+C content of 0.351.

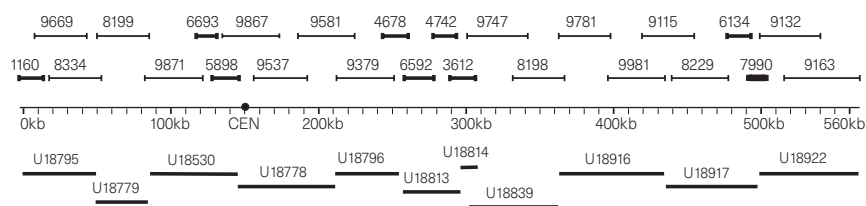


Figure 1 The central line representing *S. cerevisiae* chromosome V is marked in kilobase pairs, starts at the left at the guanine of the *Sau3A* site of the leftmost recombinant yeast DNA, and extends to the right to 570 kb. The centromere, *CEN*, is represented by a solid circle at ~151 kb. Above the line, placed across their map positions, are the individual recombinant yeast DNAs that were sequenced; 16 cosmids (thin lines), 8

lambdas (thick lines), and 1 plasmid (very thick line). Genbank accession numbers are placed below the line, above the bars indicating the corresponding map positions. From left to right, Genbank accession numbers are U18795, U18779, U18530, U18778, U18796, U18813, U18814, U18839, U18916, U18917 and U18922. There is some deliberate overlap between Genbank entries to maintain contiguity.

There are only two places on chromosome V where the quality of the sequence is not high. In the first case, at about 312 kb, there are ~50 bp of unique sequence bounded on both sides by poly (dA): poly (dT). *Taq* polymerase, and the other DNA polymerases tested, frequently terminated within the homopolymer, and seldom reached the short unique sequence. Therefore, we have only a few reads across the unique sequence. In the second case, at about 450 kb, there is a 5-kb stretch that contains many delta and delta-like sequences interspersed with a small amount of unique sequence. The clone containing this segment was shotgun sequenced to an average of 16-fold redundancy, yet there were relatively few reads in this region. Therefore, for PCR amplification, this 5-kb region was divided into many virtual parts, based on the positions of the unique sequences. Several custom primer pairs, and internal sequencing primers, were designed and synthesized for each part⁷. These were used in PCR amplification reactions with total yeast genomic DNA as the template. We have sequenced carefully across this region. For most bases, there is sequence from both strands.

There were three special cases that warrant further attention. First, a point mutation had occurred during either cloning or subsequent propagation in *Escherichia coli*. In an overlap region shared by two recombinant DNAs (lambda 5898 and cosmid 9867; Fig. 1), the sequence should be the same, but in this case there was one reproducible base difference. Lambda 5898 has a guanine residue where cosmid 9867 has an adenine residue. When total yeast genomic DNA was used as a template for PCR amplification, the product of which was used as a template for dye-terminator sequencing, the base at that position was an adenine. The traces showed no indication of a naturally occurring polymorphism. We therefore conclude that the guanine in lambda 5898 was the result of a mutation. In the second special case, we examined the ORFs for any apparently premature, in-frame stop codon, and found two that were puzzling. The first was the TGA stop codon at position (rounded-off) 352 kb. There are three rightward-reading frames, and this stop codon (TGA) is in the first of these. Following the TGA in this frame, there is a lysine codon (AAA) and a methionine codon (ATG) followed by a long ORF. In the second reading frame there are three stop codons: two (TGA, TAA) are next to each other, the third is five triplets further on; this frame is truly stopped. However, there is a long ORF starting with a methionine codon considerably upstream and ending at the double stop codons TGA, TAA. The third reading frame has many stop codons. The two ORFs share one base: A, the first base (ATG) of the first reading frame and the last base (AAA) of the second reading frame. In general, yeast ORFs are separated by several hundred bases. Except for a -1 frame shift in the first reading frame, there would be one ORF rather than two. However, despite sequencing through this position many times, the sequence, the TGA and the five A bases in a row remained invariant. The second apparently premature stop codon occurred in the ORF that corresponds to *FLO8* at ~375 kb. The TAG stop codon between YER108c and YER109c appears not just in the recombinant yeast DNA, but also in PCR amplifications from total yeast DNA. Thus it is possible that *FLO8* is not functional in yeast strain AB972. In the third special case, in the entire collection of recombinant yeast DNAs, only lambda 3612 (Fig. 1) covers this region. We found

lambda 3612 to be highly unstable, giving rise to non-random DNA deletions at extremely high frequency. Starting with 30 individual plaques from a primary stock, only one yielded lambda 3612 DNA without a detectable deletion, as judged by *EcoRI/HindIII* double-digestion patterns, but even that gave rise to deleted DNAs upon subsequent growth. Therefore, all of the lambda 3612 sequence came (uncharacteristically) from just one preparation of DNA.

A comparison of the DNA base sequence of chromosome V to that of the other *S. cerevisiae* chromosomes shows that there are two stretches that have similar genes in the same order on two other yeast chromosomes. A portion of the left arm of chromosome V, containing *CYC7* and *RAD*, shows the same relative gene order as chromosome X, but in the opposite orientation, as noted previously⁸. In addition, a 60-kb region of the left arm of chromosome IX contains nine genes or ORFs for which each has an apparent homologue within a 60-kb region of the right arm of chromosome V. The nine putative protein pairs and their calculated similarity (identity)⁹ are: (1) YIL045w/YER054c, 63 (44) %; (2) YIL050w/YER059w, 71 (52) %; (3) YIL051c/YER057c, 87 (70) %; (4) YIL053w/YER062c, 97 (92) %; (5) YIL056w/YER064c, 63 (47) %; (6) tRNA-ser, 100%; (7) YIL057c/YER067w, 85 (67) %; (8) *RNR3*/YER070w, 90 (82) %; and (9) YIL074c/YER081w, 95 (91) %. On chromosome V itself, the *FCY2* protein product¹⁰ and the putative YER060w translation product are 87 (75) % related.

When considering the accuracy of our 569,202-bp sequence of *S. cerevisiae* chromosome V, we must emphasize that (essentially) all of the sequence was determined from recombinant DNA propagated in *E. coli*. Even if our sequence of the recombinant DNAs were 100% accurate, there may be sequence differences between the recombinant DNAs and the yeast genome. We identified one apparent point mutation solely because it occurred within a region common to two recombinant DNAs. Other point mutations, occurring during cloning or propagation in *E. coli*, would probably not be detected.

There is a much more dramatic example of a discrepancy between the sequence of a recombinant DNA and the yeast genome. We deposited our chromosome V sequence in Genbank and SacchDB (<http://genome-www.stanford.edu/>) in December 1994. On 18 April 1996, we received an e-mail from J.-L. Souciet, J. de Montigny and S. Potier (CNRS, Strasbourg) politely telling us that ~2 kb were missing from our sequence. They had found that, in addition to *FCY2* (YER056c; encoding a purine-cytosine permease¹⁰) at ~267 kb and a closely related ORF (YER060w) at ~275 kb, there is also a third closely related ORF in this region (Genbank accession no. X97346) (Fig. 2.). Our sequence across the apparent deletion came from two libraries made from lambda 6592. The sequence is 12-fold deep, and there are no traces that diverge from our Genbank sequence. The Genbank sequence is therefore an accurate sequence of this particular recombinant DNA: lambda 6592. We had intended to sequence cosmid 9380. Repeated attempts to prepare 9380 yielded only minute amounts of DNA. We abandoned cosmid 9380, and instead sequenced three lambda DNAs: 4678, 6592 and 4742. From the tiny amount of cosmid 9380 DNA, we constructed (and sequenced) a *Sau3A* library. The data (traces) from the library of *Sau3A*-cleaved 9380 DNA were used within the assemblies

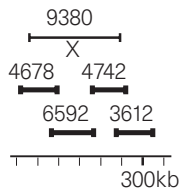


Figure 2 The bottom line is a schematic map of yeast chromosome V from 240 kb to 310 kb. Individual, sequenced recombinant DNAs are placed above the line and across the appropriate map positions. Cosmid 9380 covers map positions ~240 kb to 300 kb and overlaps lambda 3612. The latter is the only recombinant DNA in the entire Olson collection⁴³⁴ that covers map positions 300 kb to 310 kb. Because 9380 could only be produced in minute amounts, three lambda DNAs (thick lines) were substituted: 4678, 6592 and 4742. The X marks the position of the ~2 kb deletion in 6592 DNA.

of the three lambda DNAs. There were some data from the 9380 library that were left over, mostly vector without insert, low-quality traces, etc., which we put aside. We searched our 'left-over' 9380 traces for homology to unique sequences in Genbank X97346 and found one excellent (97.3% identity) match for 294 bp (bounded by *Sau3A* sites). We conclude that lambda 6592 has a deletion relative to the yeast genome and that one additional ORF should be added, bringing the ORF total to 272.

To complete the sequence of chromosome V, an insertion of 2,011 bp (Genbank accession number X97346) should be made at base 275,951 of our sequence as has been done in SacchDB. In addition, H. Wedler and R. Wambutt have sequenced the left (Genbank accession no. U73806) and right (Genbank accession no. U34775) telomeres of yeast chromosome V. Within SacchDB, 2,477 bp have been placed to the left of the leftmost base (the G of the leftmost *Sau3A* site) of our sequence. That G is no longer base 1 but base 2,478. Within the left telomere, there is an ORF, YEL077c, which brings the current ORF total to 273. Concomitantly in SacchDB, 3,181 bp have been placed to the right of our sequence.

Basically, there are two types of errors: random and systematic. If there is a random error in an individual sequence read, we will find and correct that error because we sequenced both strands to high redundancy (average of 12.5-fold). It is much more difficult to identify a systematic error that is inherent in, for example, the dye-primer chemistry, polyacrylamide gel electrophoresis, *Taq* polymerase, or base-calling software, that systematically misreads or deletes a base(s) within a particular sequence. *Taq* polymerase seemed to have systematic difficulties synthesizing across short repeating units; not only is the number of repeats often ambiguous, the sequence traces following a repeat are often diminished in signal quality. We believe that this observation reflects an inherent characteristic of *Taq* polymerase. Of the several DNA polymerases tested in an attempt to solve this problem, Amplitaq FS polymerase (Perkin-Elmer 402079) yielded the best number of good sequence calls, but did not solve the problem completely. However, the problem in counting the short repeating units unambiguously may not be a sequencing problem but in some cases may reflect true biological heterogeneity. A second possible systematic error arises from the well-known guanine compressions. Guanine compressions are usually identified when the base-calling software identifies fewer guanines on one strand than cytosines on the opposing strand. However, if two (or more) guanine compressions are positioned symmetrically on opposing strands, the compression on one strand is compensated by an analogous compression on the other strand. There are no 'extra' cytosines, and the existence of the compressions could be missed.

One reason for sequencing all of the *S. cerevisiae* DNA is that yeast is important as a model organism. A second reason is to test the approaches to, and develop technologies for, large-scale DNA sequencing in preparation for the sequencing of the human genome. In this regard, we would like to describe some important lessons learned during the sequencing of yeast chromosome V. First, 800 kb were shotgun sequenced to achieve 569,202 bp of contiguous sequence, an inefficiency of 40%. Considerable time and money would have been saved if the ends of the recombinant yeast DNAs had been mapped relative to each other (a 'sequence-ready'

contig of cosmid DNAs). Second, a large amount of freezer space was used in archiving recombinant M13 DNAs, a small percentage of which were later used as templates for finishing. An important reason in the delay of finishing was the cost of oligonucleotide primers for PCR. Finishing has been made economical by the availability of low-cost oligonucleotides⁷, so long-term storage of M13 DNAs is no longer necessary. Third, when the Yeast Genome Project was started, the conventional wisdom had that it was necessary to sequence a set of overlapping cosmids. However, we now know that the sequence of DNA as large as bacterial genomes can be assembled using a shotgun approach^{11,12}. If we started again, we would purify *S. cerevisiae* chromosome V directly by pulse-field gel electrophoresis, hydrodynamically shear the DNA to an average size of 1 kb (ref 13) and shotgun clone the sheared DNA directly into the M13 sequencing vector. The yeast genome could probably have been sequenced by the direct shotgun cloning of total genomic DNA to generate one M13 sequencing library. Individual cosmid and lambda clones could have been used to fill holes and resolve ambiguities. □

Methods

All of the *S. cerevisiae* recombinant DNAs sequenced in this study were constructed in the laboratory of M. Olson⁴¹⁴. With the exception of plasmid 7990, which was derived from two yeast strains¹⁵, all of the recombinant DNAs were derived from yeast strain AB972. Those recombinant DNAs with number designations less than 8000 are lambdas (except for plasmid 7990), those with numbers over 8000 are cosmids. We sequenced 16 cosmids (8198, 8199, 8229, 8334, 9115, 9132, 9163, 9379, 9537, 9581, 9669, 9747, 9781, 9867, 9871 and 9981), eight lambdas (1160, 3612, 4678, 4742, 5898, 6134, 6592 and 6693), and one plasmid (7990) (Fig. 1). We also obtained some sequence from another five cosmids (8063, 9268, 9380, 9495 which contained a large deletion and 9675) and two lambdas (3955 and 6052). These *S. cerevisiae* recombinant DNAs (except 9495) are available from the American Type Culture Collection.

The 'shotgun' sequencing strategy was to reduce randomly the size of the yeast recombinant DNAs ('inserts') to approximately 1 kb. The inserts were ligated to the M13 sequencing vector by using a 'linker-adaptor' system, which minimizes the formation of chimaeric DNAs. The recombinant M13 'sequencing library' was electroporated into *E. coli* and plated. Individual M13 plaques were picked and grown, and recombinant M13 DNAs were purified. (Our detailed laboratory protocols are freely available on the World-Wide Web at <http://sequence-www.stanford.edu>.)

The shotgun sequencing used dye-primer chemistry in cycle sequencing reactions, followed by fluorescence detection using an ABI 373A automated sequencer. Most of the sequence data from individual lanes ('traces' or 'reads') were edited automatically using custom software, with borderline cases being edited manually using the TED software¹⁶. Individual sequence reads were assembled using the XBAP program¹⁶. The final sequence was determined by editing manually the assembled reads.

Where there seemed to be overlapping ORFs, in either the same or in the opposite direction, the conventional assumption was made that yeast seldom uses both overlapping frames. Three criteria were used to determine which was the most likely ORF. First, using both FASTA and BLAST programs, each of the overlapping reading frames was examined for homology to known genes in the public databases; an ORF with homology was chosen over one without. Second, each organism has its own distinctive preference for certain codons over others. This preference can be expressed in arithmetic terms, as it is within the GeneFinder program (L. Hiller and P. Green, 1990-1993; documentation, software and yeast codon usage data files. Genome Sequencing Center, Washington University School of Medicine, St Louis, MO 63108, USA). GeneFinder was used to compare the codon usage for each of the overlapping ORFs. The ORF that more closely matched yeast's codon usage was chosen; in almost all cases, this distinction was unequivocal. Third, the longer ORF was selected. The nomenclature for yeast ORFs is composed of five letter/number combinations: Y (for yeast), E (the fifth letter of the alphabet for V), L or R (for the left or right arm, as defined genetically), a number (counted sequentially from the centromere in both directions), and w or c (for the transcribed strand); for example, the *URA3* gene, encoding orotidine-5'-phosphate decarboxylase, is YEL021w.

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The nucleotide sequence of *Saccharomyces cerevisiae* chromosome VII

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The complete nucleotide sequence of *Saccharomyces cerevisiae* chromosome VII has 572 predicted open reading frames (ORFs), of which 341 are new. No correlation was found between G+C content and gene density along the chromosome, and their variations are random. Of the ORFs, 17% show high similarity to human proteins. Almost half of the ORFs could be classified in functional categories, and there is a slight increase in the number of transcription (7.0%) and translation (5.2%) factors when com-

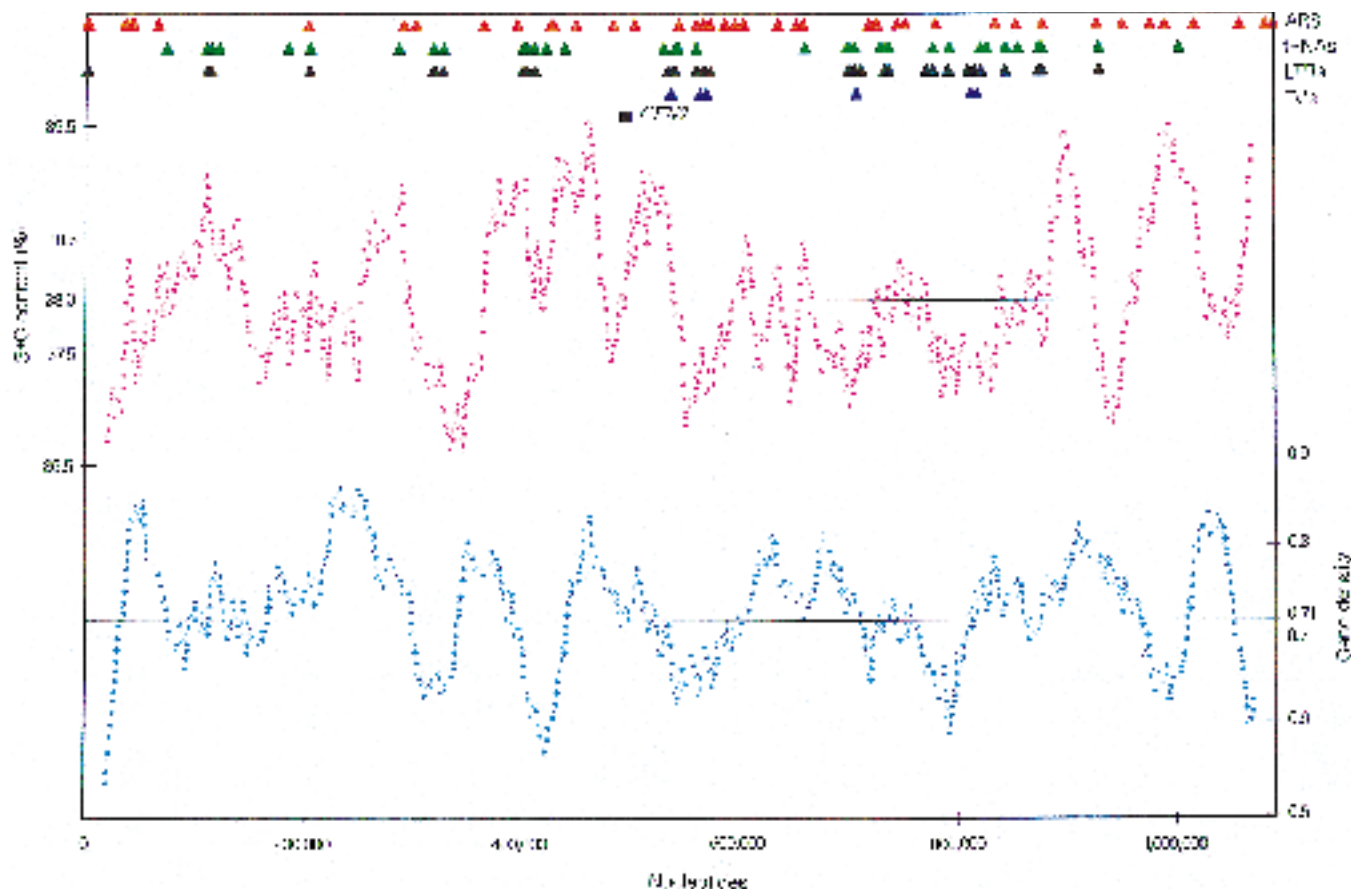


Figure 1 Top, position of genetic elements along chromosome VII. Middle, compositional variation curve; each point represents the average G+C content in a 40-kb sliding window (steps are 500 bp); the horizontal line represents the average G+C (38%). Bottom, gene density is expressed as

the fraction of nucleotides within ORFs versus the total number of nucleotides in sliding windows of 40 kb (steps are 500 bp); the horizontal line represents the average gene density (0.71).

pared with the complete *S. cerevisiae* genome. Accurate verification procedures demonstrate that there are less than two errors per 10,000 base pairs in the published sequence.

Before the publication in 1992 of the yeast chromosome III sequence¹, the only available *S. cerevisiae* genome sequence of appreciable size was a contig of 24 kilobases (kb) from chromosome VII (refs 2, 3). A 60-kb physical map covering the left arm of this chromosome between *CEN7* and *TRP5* markers had been built, allowing sequencing and transcriptional mapping of nine ORFs located in the 24-kb region spanning the *PMA1* and *ATE1* loci. Analysis of these data led to the estimation of a minimum number of 5,300 expressed genes in yeast. In this centromeric region a recombination frequency of 1 cM corresponded to an average distance of 3.3 kb, compared with 2.9 kb for the complete chromosome. These extrapolations have been confirmed by the complete 1,090,936 nucleotide sequence of chromosome VII, the fourth longest in *S. cerevisiae*.

Chromosome VII contains 564 ORFs of more than 99 codons, plus eight smaller, previously identified, ORFs. Of these 572 ORFs, 19 are predicted to carry an intron at their extreme 5' end. The *RPL6A* gene is interrupted by two introns, one of which codes for the small nuclear RNA39. Of the 572 ORFs, 152 (26.5%) had previously been characterized biochemically, and an additional 79 (14%) had been characterized phenotypically. Of these 231 known genes, disruption phenotypes have been reported in 140 cases, of which 37 are lethal. An additional 61 ORFs (11%) show a high similarity (FASTA score above 300 or one third of self score) to another ORF of known function. However, if the threshold FASTA score is lowered to 150, which is a significant value in many cases, another 20 ORFs (3.5%) could be envisaged to be of predictable function, raising the total number of ORFs of known or predictable function to 312 (54.6%). Of the remaining ORFs, 74 (12.9%) are similar to protein sequences of unknown function, and 186 ORFs (32.5%) show weak

or no significant similarity to any other protein sequence in the public data libraries. Finally, the expression of 63 ORFs (11%) is questionable owing to their partial overlap with other ORFs (44) or to a combination of small size (less than 150 codons) and low codon adaptation index (CAI below 0.110)⁴. However, these criteria are not absolute as at least four expressed ORFs from chromosome VII do not meet them: *AGA2* (87 codons; CAI, 0.089), *SOH1* (127 codons; CAI, 0.096), *SPT4* (102 codons; CAI, 0.109) and *VMA21* (77 codons; CAI, 0.109). Almost 30% of the ORFs from chromosome VII are redundant, as 166 show high similarity (FASTA score greater than 300 or one third of the self score) with other yeast genes.

The average ORF size is 468 codons, the longest (YGL195w, *GCN1*) being 2,672 codons. The average distance between ORFs located on the same strand is 514 base pairs when not containing tRNAs or long terminal repeats (LTRs) and six are longer than 2,000 bp. In the case of divergent promoters, the spacing is 553 bp, with two being longer than 2,000 bp. The mean size allocated to convergent terminators is only 304 bp, and one of these is longer than 2,000 bp. The mean G+C content of inter-ORFs regions is 33%. All of these values are very similar to those found for the complete genome⁵.

An attempt has been made to classify chromosome VII ORFs in functional categories (Table 1). Note that any given protein may belong to more than one category. The percentage of ORFs from chromosome VII in each functional category correlates with the functional distribution of ORFs within the complete genome, with a slightly higher content in the transcription (13 transcription factors) and protein synthesis (18 ribosomal proteins) categories.

The putative transmembrane spans have been computed with the KKD algorithm⁶ using a rather low threshold⁷ that takes into account not only the fully hydrophobic spans, but also the predicted amphipathic α -

Table 1 Functional categories of yeast ORFs from chromosome VII

| Functional category | ORFs | |
|------------------------------------|--------|------------|
| | Number | Percentage |
| Metabolism | 55 | 9.0 |
| Energy | 19 | 3.1 |
| DNA synthesis | 10 | 1.6 |
| Transcription | 43 | 7.0 |
| Protein synthesis | 32 | 5.2 |
| Protein destination | 18 | 2.9 |
| Transport facilitation (permeases) | 18 | 2.9 |
| Intracellular traffic | 17 | 2.8 |
| Cell structure | 19 | 3.1 |
| Organelle assembly | 11 | 1.8 |
| Signal transduction | 5 | 0.8 |
| Cell division | 24 | 3.9 |
| Cell rescue | 12 | 1.9 |
| Retrotransposons | 13 | 2.1 |
| Unclassified | 318 | 51.8 |
| Total | 614 | |

helices which, when present in a bundle, can contribute to the formation of a polar channel within the lipid bilayer. Of the 572 ORFs, 359 (63 %) show no predicted transmembrane spans or are known to be soluble, 79 (14 %) carry at least three putative spans or are known to be membrane bound, and 134 (23 %) have one or two predicted hydrophobic α -helices, a feature which does not necessarily mean that they are membrane-bound. All ORFs have been submitted to PSORT analysis⁸ to predict their subcellular localization. If we consider a high certainty score to be at least 0.8 for a given localization, and a low one to be less than 0.5 for all other possible localizations, only 76 ORFs match these criteria: 37 ORF products predicted to be in the nucleus, 16 in the endoplasmic reticulum, nine in the mitochondria, nine in the plasma membrane, two in the vacuole, two in the peroxisome, and one secreted outside the cell. For the 23 chromosome VII proteins of known subcellular localization, the PSORT prediction was correct in 14 cases (61 %). Note that three of the six ORFs that exhibited an erroneous nuclear localization were ribosomal proteins which are known to carry a nuclear localization signal to allow the biosynthesis of the ribosome particles⁹.

By applying the program PYTHIA¹⁰ to search for simple repeats within a gene, we detected at least 19 genes with regularly repeated nucleotides corresponding to repeated amino acids in the encoded proteins. Some of these regions are composed of single amino-acid repeats, including 21 aspartates (in YGL227w), 13 aspartates (YGL058w, *RAD6*), 11 and 8 glutamines (YGL066w), 18 asparagines (YGR233c), 15 asparagines (YGL014w), 10 asparagines (YGL013c, *PDR1*), 23 serines (YGR023w) or 16 serines (YGR130c). We also identified more complex repeats, such as (T S/N ATTT A/E S X₄)₁₁ in YGR296w (=Y'), (QQQP)₉ in YGL122c (*NAB2*), (DEEE)₃ in YGL164w, (AQ)₁₄ in YGL181w (*GTS1*), (TSSS)₉ in YGL028c, or (HN)₅ in YGL178w (*MPT5*).

A systematic search for similarities with human proteins was performed for the 572 ORFs of yeast chromosome VII. A total of 95 ORFs (16.6 %) show a very significant similarity (FASTA score higher than 300 or higher than one third of self score) with human proteins, of which these 79 (13.8%) correspond to known yeast proteins whose function is often closely related to the function of the human homologue. Similarities of some of the 16 previously unknown ORFs (2.8 %) with human proteins are shown in Table 2.

Several chromosome VII ORFs show a high degree of similarity with interesting proteins from other organisms. These include: YGL236c, similar to the glucose-inhibited division (*gidA*) protein of the bacterium *Escherichia coli*; YGL201c, similar to the intestinal DNA replication protein of the rat *Rattus norvegicus*; and YGL054c, similar to the Cni protein necessary for anterior–posterior and dorsal–ventral patterning in the fruit fly *Drosophila melanogaster*.

Another interesting feature of *S. cerevisiae* chromosome VII is the existence of a pseudogene, which has been confirmed by direct polymerase chain reaction (PCR) sequencing on the yeast genome. This pseudogene,

Table 2 Similarity of yeast chromosome VII ORFs of unknown function with human proteins

| Yeast ORF | Human protein |
|-----------|--|
| YGL150c | SNF2a transcription activator that cooperates with the oestrogen and retinoic acid receptors |
| YGL125w | methylenetetrahydrofolate reductase |
| YGL106w | calmodulin |
| YGL003c | probable cell-division control protein CDC 55 |
| YGR034w | new ribosomal protein similar to the human ribosomal protein L26 |
| YGR043c | transaldolase |
| YGR217w | first putative calcium channel in yeast similar to the human voltage-dependent L-type calcium channel $\alpha 1$ subunit |
| YGR231c | prohibitin, which inhibits DNA synthesis and regulates proliferation |
| YGR256w | phosphogluconate dehydrogenase |

The similarity threshold is a FASTA score higher than 300 or higher than one third of self score.

YGL259w, contains two frameshifts, and only one of the three ORFs is longer than 99 codons. However, all three parts show a high similarity with YIR039c, a hypothetical aspartyl proteinase. Another curious feature is the presence of three ORFs in the same frame separated by two stop codons. The rightmost one, YGL238w, corresponds to the *CSE1* chromosome segregation gene. The leftmost ORF, YGL241w, shows 17% identity over 1,053 amino acids with *CSE1*, whereas the central ORF shows no similarity with this protein. Finally, a possibly unique feature in the yeast genome is the tail-to-tail arrangement of the *SMD1* (YGR074w) and *PRP38* (YGR075c) genes, with their respective ORFs terminating on opposite strands without any intervening nucleotide between the stop codons¹¹. This region, as well as the *CSE1* region, have been verified by direct PCR sequencing on the yeast genome.

Chromosome VII contains six yeast retrotransposons: three Ty1s, one Ty2, one Ty3 and a pseudo-Ty, which contains, in addition to the normal frameshift separating the Ty1A and Ty1B coding sequences, two frameshifts splitting both ORFs in two parts. Of 35 tRNA genes identified, eight are interrupted by introns. All LTRs on chromosome VII are associated with a tRNA (for review, see ref. 12), except for the two Ty5 LTRs located close to the left telomere; however, 12 tRNAs are not associated with LTRs. The positions of tRNAs, Tys, LTRs and putative ARS consensus have been compared to G+C content and gene density along the chromosome (see Fig. 1). No clear correlation could be identified regarding the location of these genetic elements. Furthermore, no statistical correlation exists between the G+C content and the gene density curves (correlation coefficient, 0.04) when highly overlapping, neighbouring sliding windows (98.7 % overlap) are used. These G+C content and gene density variations are not significant, as the same analysis performed on several random mixes of the original sequence yields similar G+C content and gene density variations. A similar graph was obtained using non-overlapping neighbouring windows along the sequence after removal of Ty and LTR elements. In this case, a good correlation was found between the G+C content of the genes and the gene density (correlation coefficient, 0.98), as well as between the G+C content of silent positions of codons in genes and the gene density (correlation coefficient, 0.66). Finally, there is no significant difference in the G+C content of the coding regions on each strand: 40.07 % G+C and 305 ORFs on the Watson strand, and 39.95 % G+C and 267 ORFs on the Crick strand.

The physical map of chromosome VII has been constructed independently from the genetic map using the meganuclease *I-SceI* to produce *in vitro* nested fragmentations of the chromosome^{13,14}. The cosmids chosen for sequencing were screened from two genomic libraries¹⁴ and completed using a few cosmids from the physical map constructed by L. Riles (unpublished). The left telomere has been cloned¹⁵ and the right telomeric sequence was obtained from a PCR fragment amplified from a strain carrying the pEL61 plasmid¹⁵ integrated in the subtelomeric region.

The quality of the sequence of chromosome VII was assessed using dif-

ferent approaches. As well as partial overlaps between the regions sequenced by two laboratories, putative frameshift checking, alignment of the sequence with previously published data and a few random resequencing verifications were performed on cosmid subclones. A new method for verifying specific regions of the sequence was developed for this chromosome (G. V. *et al.*, manuscript in preparation) and applied to several other chromosomes. The extrapolation from the number of discrepancies observed in the overlaps (102,049 nucleotides, 9.4 %) to the whole sequence suggests that the nucleotide sequence of chromosome VII is 99.974 % accurate. The quality of the coding regions, where frameshifts are quite easy to check, is probably much higher than that of the intergenic regions. Indeed, the quality assessment procedure led to the correction of a total of 90 errors mainly located in the coding regions. A total of 56,344 bp (5.2 %) have been resequenced, and the comparison with the original data makes it possible to estimate that about 120 errors remain in the chromosome VII sequence. Parts of the sequence were published independently^{16–30} before assembly of the contig and application of the final quality controls; several other manuscripts are in the press. □

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The nucleotide sequence of *Saccharomyces cerevisiae* chromosome IX

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Large-scale systematic sequencing has generally depended on the availability of an ordered library of large-insert bacterial or viral genomic clones for the organism under study. The generation of these large insert libraries, and the location of each clone on a genome map, is a laborious and time-consuming process. In an effort to overcome these problems, several groups have successfully demonstrated the viability of the whole-genome random 'shotgun' method in large-scale sequencing of both viruses and prokaryotes^{1–5}. Here we report the sequence of *Saccharomyces cerevisiae* chromosome IX, determined in part by a whole-chromosome 'shotgun', and describe the particular difficulties encountered in the random 'shotgun' sequencing of an entire eukaryotic chromosome. Analysis of this sequence shows that chromosome IX contains 221 open reading frames (ORFs), of which approximately 30% have been sequenced previously. This chromosome shows features typical of a small *Saccharomyces cerevisiae* chromosome.

The sequence derived for chromosome IX is 439,886 nucleotides in length, and 71.6% codes for proteins or predicted proteins. There are 219 non-overlapping ORFs equal to or greater than 100 amino acids long, and a further two ORFs (YIL060W and YIL059C) that overlap; these are short, and both have a low codon adaptation index (CAI). Although it is unlikely that both are coding, one could not be selected above the other as more likely to encode a protein. A single Ty3-2 retrotransposon containing three ORFs is present on the left arm of chromosome IX (between bases 205,217 and 210,644), leaving 218 *S. cerevisiae*-derived ORFs encoded on this chromosome, of which 116 are on the Crick strand, and 102 (+ 3 transposon ORFs) are on the Watson strand. Of these, 66 (30.3%) have been sequenced previously. A further 68 (31.2%) have some similarity to genes in *S. cerevisiae* and other organisms for which some functional information is available. However, 74 (33.9%) of the predicted genes on this chromosome cannot be assigned even a putative function based on sequence similarity. These can be divided into two groups: those that show no similarity to current database entries (53, 24.3%), and those that are similar to predicted genes of unknown function (21, 9.6%). The remaining 10 (4.6%) are putative pseudogene ORFs.

The average length of a chromosome IX ORF is 476 codons, with an average of one ORF every 1,993 base pairs. The largest ORF on chromosome IX is YIL129C, which encodes a hypothetical protein of 2,376 amino acids. The YIL129C protein is similar to another hypothetical protein encoded on *Caenorhabditis elegans* chromosome III (EMBL database, accession numbers CEF21H11, U11279 and ORF F21H11.2) over a region of 2,009 amino acids. In total, 20 chromosome IX ORFs are longer than 1,000 codons. Short *S. cerevisiae* genes with no homology are difficult to detect⁶. On chromosome IX, five ORFs with less than 100 codons have been identified, but future analysis will probably reveal additional short coding regions. Less than 4% of the ORFs on chromosome IX are predicted to be spliced; eight ORFs contain introns. None of the tRNA genes on this chromosome are spliced.

Ten ORFs have been identified as contributing to five putative pseudogenes. These ORFs have very good homology to genes or predicted genes, but are separated from an adjacent ORF with homology to the same protein by internal stop codons or frameshifts. These areas have been sequenced on *S. cerevisiae* genomic DNA, and the frameshifts and stop codons confirmed. However, at least two pseudogene ORFs, YIL168W and YIL167W, probably constitute the single gene *SDLI*, which codes for a serine dehydratase⁷. This gene is not present elsewhere in the *S. cerevisiae* genome and is not essential in *S. cerevisiae*. A second putative pseudogene is highly similar to hexose transporter genes, and may also simply prove to be mutated in AB972 rather than being a true pseudogene. All putative pseudogenes are located near the telomeres of the chromosome.

For this and other *S. cerevisiae* chromosomes, the intergenic distance between two adjacent ORFs varies depending on their orientation with respect to each other. Of the adjacent ORFs on chromosome IX, 95 are arranged in tandem, 54 are divergent and 55 convergent. For ORFs arranged in tandem the intergenic distance averages 472 basepairs; ORFs with divergent promoters average 619 bp; and ORFs with convergent terminators average 421 bp. This is consistent with the greater information content required for transcription initiation and regulation than for transcription termination.

Chromosome IX also contains ten tRNAs, five solo delta elements, one solo sigma element, two sigma elements flanking a transposon, and a single solo tau element.

Reports describing the features of the other small yeast chromosomes suggest that they have used a variety of strategies to achieve a certain minimum length⁸⁻¹⁰, and this seems to be the case for chromosome IX. Its

right telomeric region is gene poor, with only 25.3% of the sequence contributing to ORFs over approximately the last 15 kilobases of the chromosome. Chromosomes I and VI, the two smallest yeast chromosomes, also have a low coding density in their telomeric regions. Ten ORFs on chromosome IX are thought to contribute to five putative pseudogenes, all of which are located in the telomeric regions (four in the left telomeric region and one in the right). Four of these putative pseudogenes have a high degree of similarity to sequences repeated elsewhere in the yeast genome (the remaining putative pseudogene may be a mutated copy of *SDLI*, as discussed previously). Two of these occur in a 21-kb region of the chromosome IX left telomere, which is duplicated almost exactly on the chromosome X left telomere. Comparison of these two pseudogene ORFs with the equivalent regions on chromosome X shows that one region is interrupted by a frameshift on that chromosome, but the second is a single ORF. The putative pseudogene on the right arm of the chromosome is highly similar to a single ORF on chromosome XIII. The centromere¹¹ of chromosome IX is located towards the right end of the chromosome between bases 355,627 and 355,744.

Sequencing has revealed the definitive chromosomal position for all genes on chromosome IX. The resulting physical map correlates well with the genetic map¹² for this chromosome. Local variations in the ratio of cM to kb occur throughout this chromosome. For example, the region bounded by the genes *REV7* and *HOP1* has a significantly lower ratio of 0.3, indicating that recombination events in this area are less frequent than for the chromosome as a whole.

A cluster of genes on this chromosome occurs within its smaller, right arm between bases 399,775 and 415,615 (YIR023W to YIR032C). Six of the ten ORFs in this region are involved in the allantoin degradation pathway^{13,14}.

Evidence of several interchromosomal duplications occurs on chromosome IX. As well as the large region common to the left telomeres of chromosomes IX and X already described, a smaller region at the right telomere shows good homology with the telomeres of several other chromosomes. There are also several other internal chromosomal regions with long-range homology to other chromosomes. The largest of these is an area common to chromosomes IX and XIV, occurring at 89,233–186,363 and 478,568–616,076, respectively, and containing 15 homologous ORFs. Smaller interchromosomal duplications have also occurred, including a region at 230,272–258,279 repeated on chromosome V at 273,881–305,178 and a region at 46,201–69,525 repeated on chromosome XI at 617,636–639,600.

A further long-range feature previously observed for *S. cerevisiae* is a variation in the percentage G+C content along the length of its chromosomes¹⁵. In most cases the G+C composition for the third base of each codon has been analysed, as this is less influenced by biases in amino acid composition. For several chromosomes a periodic variation in G+C content has been observed. A plot of third-position G+C composition for chromosome IX (Fig. 1) shows that, as for other chromosomes, G+C content varies along its length. Consistent with previous analyses, the region around the centromere of the chromosome has low G+C content; the highest content occurs in peaks separated by about 100 kb.

The occurrence of first-, second- and third-codon position G+C composition was analysed for individual ORFs, as well as G+C composition in intragenic regions and the total G+C content of the chromosome (Fig. 2). This was superimposed on a chromosome map showing individual ORFs, as the usual method for plotting G+C composition uses a large window to calculate the average third-position G+C content of several adjacent ORFs, making it difficult to ascertain whether high G+C composition is a general trend in a particular region or derives from a single ORF. This method correctly predicted the peaks in third-position G+C content already described for chromosome III (data not shown).

In regions producing peaks of G+C content several adjacent reading frames show higher than average third-position G+C. Specific areas noted include 1–6, 52–67, 93–97, 175–180, 266–276, 308–313, 380–385 and 421–424 kilobases, with up to six ORFs contributing to each region. The areas of high G+C content are much more pronounced in chromosome III, with many more reading frames over a much longer distance contributing

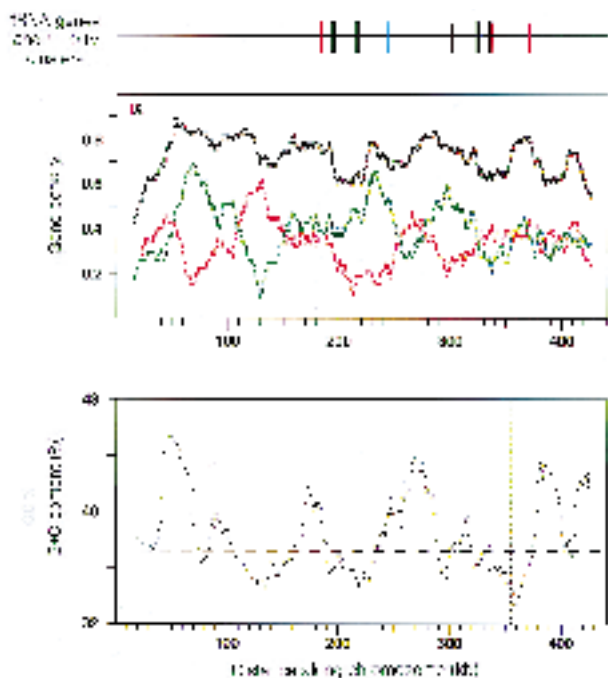


Figure 1 Overall molecular architecture of chromosome IX. The top line indicates positions of tRNA genes, solo long terminal repeat (LTR) or Ty elements (thin vertical lines) or clusters of them (thick vertical lines) along the chromosome map. The panels show variation in gene density (top) and base composition (bottom) along the sequence-based map of chromosome IX (scale in kilobases from left telomere). Vertical broken lines indicate the centromere. Gene density is expressed as the probability for each nucleotide to be part of an ORF, and was calculated using sliding windows of 30 kb (steps of 0.5 kb) for the Watson strand alone (red line), the Crick strand alone (green line), and the sum of both (black line). G+C richness (%) was calculated from the silent positions of codons using a sliding window of 13 consecutive ORFs (horizontal broken line indicates average percentage G+C at silent positions of codons, 35.8%).

to the two main peaks on each of the chromosome arms (results not shown).

As expected, the G+C content in intergenic regions is, on average, lower than that for coding DNA. However, it is not uniformly low, and local areas of high G+C content can be observed in some non-coding areas. The largest of these regions have been examined in detail for potential coding sequence. Although any ORFs present in these areas could conceivably be coding, they are all short, and standard methods for predicting coding sequences suggest that this is unlikely. A BLASTX search does not show any significant similarity to other *S. cerevisiae* ORFs, indicating that these areas are not remnants of ancient pseudogenes. Other possibilities have not yet been fully investigated.

Very few genes show high G+C content in position 2 of their codons. However, YIL169C, a putative glycoprotein, and YIR019C (*STAI*), a glucoamylase gene, show high second-position G+C content. Both genes code for proteins with a biased amino-acid composition. YIR019C is surrounded by regions low in G+C, but coincides with a high G+C peak on a plot of overall G+C content (results not shown), showing that a single gene can cause a peak on these plots. The role of these differences in base composition found in all yeast chromosomes has not been determined, although high G+C content might promote DNA replication or recombination.

The chromosome IX sequencing project served to highlight the difficulties involved in a whole-chromosome 'shotgun' project. This approach is described in detail in the Methods section. The purity of the starting chro-

mosomal DNA is critical to avoid redundant sequencing effort and to minimize problems in assembly. Contamination of the chromosome IX preparation with DNA from other chromosomes resulted in a large number of single reads in the database that were not from that chromosome, but which increased the complexity of sequence assembly, slowing down both assembly and subsequent manipulation of contigs. Re-running the PFG-purified chromosome on a second pulsed-field gel has been shown to improve significantly the purity of the DNA preparation¹⁶. Data from the whole-chromosome 'shotgun' helped to fill the gaps in the cosmid and lambda libraries, but on their own were difficult to manage. The cosmid clones provided a framework on which to build data from the whole-chromosome shotgun. □

Methods

The chromosome IX sequencing project was initiated using a lambda clone library generated by M. Olsen and L. Riles¹⁷. This was an inefficient approach because of short insert sizes (20 kb) and relatively large overlaps (about 5 kb) between lambda clones. Few cosmids were available at the time, so the approach of randomly 'shotgun' sequencing the entire chromosome was attempted.

Yeast genomic DNA was electrophoresed on pulsed-field gels¹⁸. To increase the purity of the chromosomal preparation, yeast plugs were excised early in the run after chromosome IX had entered the gel, reducing the background DNA level. Degradation of the material of higher molecular weight occurs with prolonged running of the gel and is the likely cause of contamination of material of lower

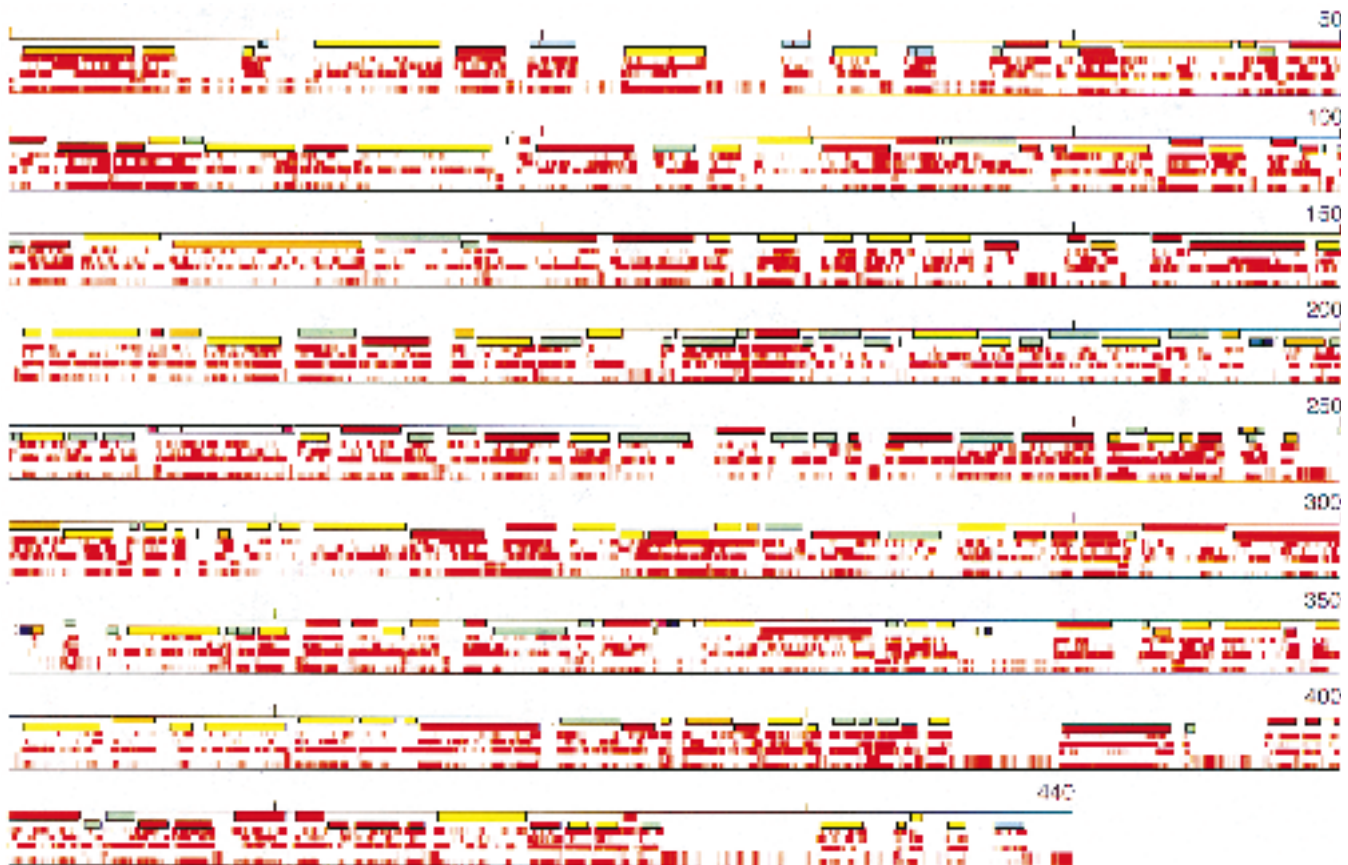


Figure 2 G+C composition of chromosome IX (drawn to scale). The top graduated line represents the chromosome split into 50-kb segments, with ORFs indicated below this as coloured boxes. ORFs located on the Watson strand are shown above those on the Crick strand. ORFs encoding previously identified genes are shown in red, those with similarities to known genes in yellow, those with similarities to hypothetical proteins in orange, and those with no significant similarities in green; pseudogene ORFs are shown in blue. tRNA genes are shown as white boxes, as are transposon-derived ORFs, with LTRs shown in dark blue (delta), turquoise (tau) and

pink (sigma). Below this, variations in G+C composition (calculated using a sliding window of 200 bases) are shown as bars, with gradations of red varying from 35% to 45% (P.R., unpublished data); areas lower than 35% G+C are white, and those over 45% are red. Five bars of G+C variation are shown: the lowest bar shows total G+C content; the second shows G+C content in intergenic regions alone; and the G+C composition in each of the three bases of each codon are shown above this, with the central of the five bars representing first-position G+C, the next representing second-position G+C, and the top bar showing third-position G+C content.

molecular weight. The chromosome IX band was excised under long-wave ultraviolet transillumination to minimize DNA damage. Chromosomal DNA was purified by melting and phenol extraction, sonicated and end repaired. Two libraries were prepared: fragments 1.4–2 kb in length were cloned into M13mp18, and fragments 6–9 kb long were cloned into the phagemid vector pBS. Over 10,000 independent M13 clones were sequenced and assembled into a database using the program XBAP¹⁹. Sequencing strategy and methods used for sequence assembly are as described^{19–25}. The lambda-clone consensus sequences were also entered into this database, which contained several thousand contigs at this stage, most of which contained a single gel read. We concluded that the chromosome IX DNA preparation was approximately 30% contaminated with DNA from other chromosomes, and that this was the source of most single-read contigs. This contamination, together with repetitive sequences in the database, caused many problems with the data assembly.

To overcome this problem, all single-read contigs were removed from the working copy of the chromosome IX database and collected in a secondary database. As further data was generated, the secondary database was periodically rescreened, and single reads were re-entered if they found matches in the primary database. This reduced the number of reads in the primary database to approximately 7,000, which represented coverage of the chromosome five times over. The database still contained several hundred contigs. At this stage a cosmid library covering most of chromosome IX became available¹⁷. The chromosomal 'shotgun' data were 'seeded' with reads from cosmid clones selected to give coverage over regions not previously sequenced by lambda clones. This approach also allowed the chromosome to be split up into manageable sections to solve double-stranding and compression problems. A minimal 'shotgun' of 300–500 reads was performed on each cosmid clone. Data from these cosmids were entered into the chromosome IX 'shotgun' database to contiguate the entire chromosome, and into separate cosmid databases for ease of handling, together with overlapping reads from the whole-chromosome shotgun. Each cosmid-sized project was contiguated, double stranded and all compressions were resolved.

Three regions of the chromosome remained unrepresented in either cosmid or lambda libraries: the left and right telomeres, and a region near the centre of the chromosome flanked by lambda clones 6569 and 3299. The right telomere was sequenced by primer walking using a plasmid clone²⁶. The left telomere was finished using data from the whole-chromosome 'shotgun' and some primer walking from polymerase chain reaction (PCR) products generated from PFGE-purified chromosome IX DNA. The gap near the centre of the chromosome was filled using data from the whole-chromosome 'shotgun' and by sequencing a 1 kb fragment generated by PCR from genomic DNA. The gap between the lambda clones 6569 and 3299 was approximately 7 kb.

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The nucleotide sequence of *Saccharomyces cerevisiae* chromosome XII

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The yeast *Saccharomyces cerevisiae* is the pre-eminent organism for the study of basic functions of eukaryotic cells¹. All of the genes of this simple eukaryotic cell have recently been revealed by an international collaborative effort to determine the complete DNA sequence of its nuclear genome. Here we describe some of the features of chromosome XII.

The nucleotide composition of the chromosome, which is 38.48% G+C overall, and gene density vary across the chromosome. This has been observed for other yeast chromosomes²⁻⁶ (Fig. 1). There are three main regions deficient in G+C, centred at approximately 150, 685 and 1,043 kilobases; one of these, as expected, coincides with the centromere. There is only one main peak of high G+C content, at approximately 473 kb, centred over the rDNA repeats. There does not seem to be any regularity in the variation in nucleotide composition, as may be the case for some other yeast chromosomes³.

Like other yeast chromosomes, 72% of chromosome XII is predicted to code for protein (considering only two copies of the rDNA cluster). The sequence contains 534 open reading frames (ORFs) of 100 or more sense codons (excluding the 13 ORFs contained within yeast transposable elements), distributed roughly equally on the two strands (255 on the Watson (top) strand and 279 on the Crick (bottom) strand). The average ORF size is 485 codons. The largest gene in the chromosome, *YLR106c*, containing 4,910 codons, is the largest in the yeast genome. The average distance between ORFs is 545 base pairs for the 121 divergently transcribed genes (promoters abutting), 282 bp for the 120 convergently transcribed genes (terminators abutting), and 493 bp for the 208 genes that are transcribed in the same direction (promoter abutting terminator). Of the ORFs, 17 (3.2% of the total) contain introns, all of which are at the extreme 5' end of the gene (except for *YLR464w*, a probable pseudogene). Two genes (*YLL057c* and *YLR388w*) may contain introns in the 5'-untranslated region of their mRNA. As expected⁷, about half of the intron-containing genes (9) encode ribosomal proteins.

Only 170 (31.8%) of the genes were previously identified. Of the 364 newly identified genes, 34 (6.4% of the total genes) are obviously similar to proteins of known function, and 54 (10.1%) are weakly similar to proteins of known function. Thus a function is known or can be predicted for 48.3% of the encoded proteins. A further 69 genes (12.9%) encode proteins similar to proteins of unknown function; 207 (38.8%) of the predicted proteins are not similar to other proteins.

Included in the predicted ORFs are 55 that are 'questionable', that is, they consist of fewer than 150 codons and have a codon adaptation index (CAI)⁸ of less than 0.110, or they overlap with another ORF. Of the 40 questionable ORFs that overlap with another ORF, the true gene can be predicted for 27 of these pairs, which include either a gene whose product is known (16 pairs) or whose predicted product is similar to another protein in the databases (11 pairs). There are therefore 13 overlapping ORFs that are suspect, although which of these ORFs is actually a gene awaits experimental determination.

Chromosome XII contains 22 tRNA genes, of which 7 are predicted to contain introns. Most of the tRNA genes are widely separated, although there are two clusters of three tRNA genes, each in a region of 9 kb to 13 kb (725,746-734,874 and 784,352-797,247). As expected⁹, many (12) of the tRNA genes are near yeast retrotransposons (Ty elements) or their isolated long terminal repeats (LTRs). Three known small nuclear RNAs, *SNR6*, *SNR30* and *SNR34* are encoded on chromosome XII. Four of the six retrotransposons on chromosome XII are of the Ty1 type and two are Ty2 elements. There are several complete or partial 'solo' retrotransposon LTRs, including nine delta elements, four sigma elements, and a tau element.

The subtelomeric regions of chromosome XII are typical¹⁰. The left subtelomeric region contains a 'core X' element, and subtelomeric repeats STR-D, C, B and A, along with two tandem Y' elements (short versions). The right subtelomeric region contains a core X element, the STR elements listed above, and 3-4 tandem Y' (long version) elements. The sequence of the first 1.5 and the last Y' elements were fused to give two copies of the Y' element in the presented sequence. Proximal to the core X are shared homologies with several other telomere regions. As with several other chromosomes, both chromosome ends contain members of the

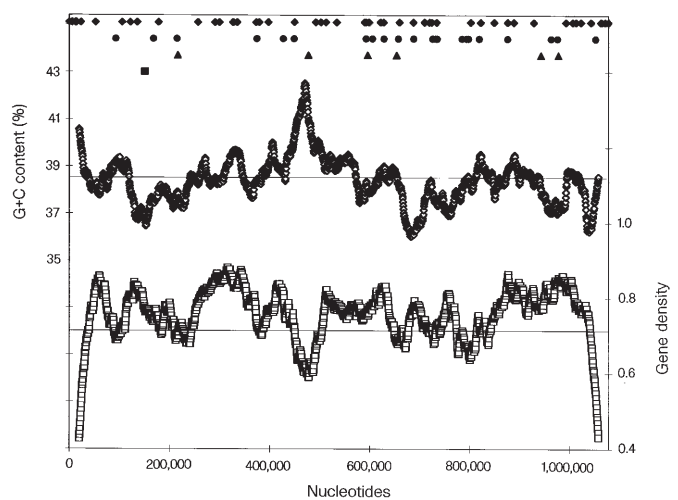


Figure 1 Top, non-coding elements of chromosome XII including autonomously replicating sequence (ARS) (filled diamonds); tRNA (filled circles); Ty element (filled triangles); centromere (filled square). Middle, G+C content as a percentage (open diamonds). Bottom, gene density (open squares).

PAU/TIP/SRP gene family¹¹

Chromosome XII is estimated to contain 100-200 copies of the 9,137 base pair rDNA repeat^{9,12-14}. Only one complete copy (the leftmost repeat in Fig. 2) and one nearly complete copy of the rDNA (the rightmost repeat in Fig. 2) are represented in the assembled sequence. In some strains these repeats seem to be interrupted by non-rDNA sequence¹⁴.

The boundaries of the rDNA repeats are in non-transcribed regions downstream of the 35S rDNA (the left, or centromere-proximal, boundary) and 5S rDNA (the right, or centromere-distal, boundary)¹⁵. The structure of the left boundary of the rDNA (nucleotide 21,811 in Fig. 2) is straightforward; the right boundary¹⁶ is more complicated. Immediately to the right of the rDNA repeats are several copies of a 3.6-kb repeat (one of which is interrupted by a Ty element) that includes the *ASP3* gene¹⁷ and ends with a nearly complete 5S rDNA gene (*5S^{var}* in Fig. 2). The precise number of copies of this 3.6-kb repeat in the genome is not known. The rightmost rDNA repeat ends in a 5S rDNA that adjoins a 3.6-kb repeat. Thus this rightmost rDNA repeat is lacking the 759 bp of sequence between the end of 5S rDNA and the end of the rDNA repeat (equivalent to nucleotides 30,186-30,947 in Fig. 2). The structure of the right rDNA junction differs in other yeast strains¹⁵.

The 5S rDNA gene in the 3.6-kb repeats lacks the non-transcribed regions of the gene. It begins two nucleotides upstream of the 5' end of 5S rRNA, and is missing the last four nucleotides of the 5S rRNA. These genes are labelled '5S^{var}' in Fig. 2 to indicate that they are incomplete. Immediately downstream of this gene is a run of 10 T residues that is reminiscent of the transcription termination sequence of RNA polymerase III (there are 29 T residues downstream of the 5S rDNA gene in the rDNA repeats). Because this gene seems to be missing the promoter and much of the terminator, it might represent a reverse-transcribed copy of the 5S rRNA that integrated into the genome. Nevertheless these genes produce 5S rRNA transcripts¹⁸.

One possible explanation of the structure of the right junction is that a reverse-transcribed copy of 5S rRNA is inserted into the genome near the right border of the rDNA cluster. This gene could then have been part of a 3.6-kb duplication. It is then easy to imagine a recombination event between an intact 5S rDNA gene in one of the rDNA repeats and a 5S^{var} rDNA in one of the 3.6-kb repeats that generated the right rDNA junction we sequenced. Other explanations for the origin of this junction have been proffered¹⁶.

To speed the completion of the sequence of this large chromosome, two groups collaborated on its sequencing. The rDNA repeats on chromosome XII served as a convenient point to divide the effort: the EU sequencing network¹⁹ determined the sequence of the chromosome to the

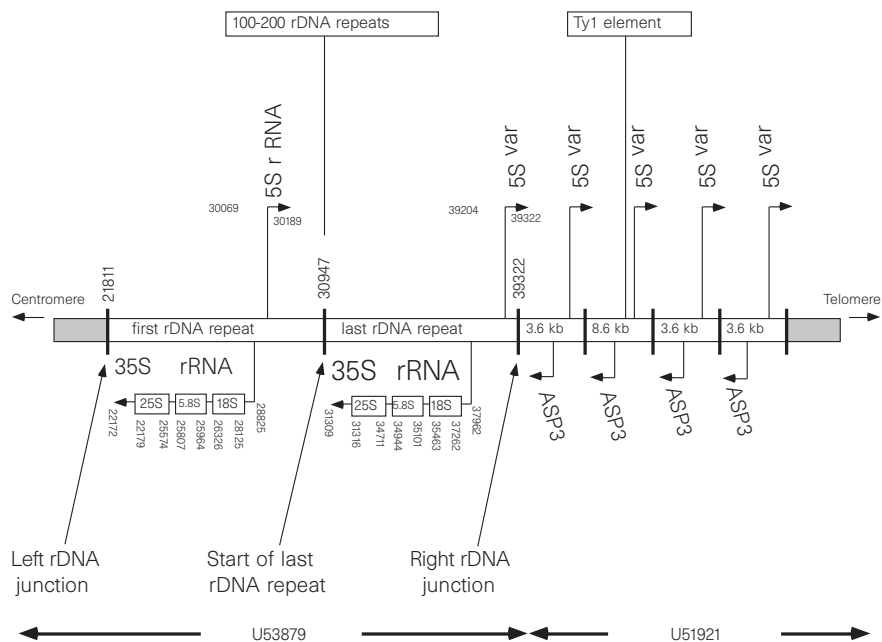


Figure 2 Diagram of the rDNA repeats and surrounding sequence, as assembled for cosmids YSCL9634 (left, GenBank accession no. U53879) and YSCL9362 (right, GenBank accession no. U51921). The numbers shown are the nucleotide coordinates for cosmid YSCL9634. The left rDNA junction (U53879 coordinate 21,811) begins at nucleotide 451,418 of chromosome XII; the right rDNA junction (U53879 coordinate 39,322) is at nucleotide 468,929. The sequence includes 1.92 rDNA repeats, representing the leftmost and rightmost copies in the genome. The remaining

100–200 rDNA repeats in the genome are represented as an insertion at coordinate 30,947. Only one complete 5S rDNA gene (in the left rDNA repeat) is included in this sequence (nucleotides 30,069–30,189); the 5S rDNA genes in the 3.6-kb repeats are variant genes. The 5S rDNA gene in the last rDNA repeat (nucleotides 39,204–39,322) includes all 5' non-translated sequences (like the normal 5S rDNA in the first repeat), but is missing sequences downstream of the 5S rRNA transcript (like the 5S rDNA genes in the 3.6-kb repeats).

left of the rDNA repeats; the sequence to the right was determined at the Washington University Genome Sequencing Center. The sequence of both strands of the entire 1,078,171 base-pair chromosome (but including only two copies of the rDNA repeats) was determined, nearly all the way to the telomeres.

The 781,865 nucleotides determined at Washington University came from 24 partly overlapping cosmid and two lambda clones²⁰. The sequence of each clone was determined by a 'shotgun' strategy followed by directed sequencing². The sequence of each clone was submitted to GenBank, and the entire non-overlapping sequence was assembled, analysed and annotated^{2–5}.

The 460,166 nucleotides to the left of the rDNA were determined by the EU network from a set of cosmid clones constructed from gel-purified chromosome XII DNA and mapped specifically for this purpose²¹. Sequencing was done by a directed approach that combines the advantages of primer walking (low redundancy) and 'shotgun' sequencing (use of a single primer)²². The sequence was determined from 'shotgun' sublibraries of 1-kb fragments of the cosmids that were then ordered by hybridization fingerprinting²². The sublibraries were arrayed on high-density filters, and sorted into smaller groups by hybridization with restriction fragments of the cosmids. Detailed mapping information was obtained by hybridizations with both oligodeoxynucleotides and pools of clone inserts amplified by the polymerase chain reaction (PCR).

The sequence of the left telomere region, including the TG_{1–3} sequence at the very end of the chromosome, was obtained from clones generated by integrating then excising a plasmid at the telomere, with capture of the flanking sequence¹⁰. The right telomere sequence was obtained by cycle sequencing of an anchored PCR product of the last Y' element from a strain whose chromosome end was specifically marked by unique vector sequence²³. The sequence of the very end of the Y' element (about 130 bp short of the end of the chromosome) was not determined.

Only the sequence of the leftmost rDNA repeat (see Fig. 2) and about 300 nucleotides across the junction of the first and second repeat was

determined. It was assembled appropriately to give the two rDNA repeats presented in Fig. 2 and in the database (GenBank accession no. U53879). The right junction sequence was not present in the cosmid closest to the rDNA on the right (YSCL9362; GenBank accession no. U51921), nor in two phage lambda clones that were mapped to this region. The structure of the junction was inferred from our ability to obtain a product of the expected size (the size of a 3.6-kb repeat) in PCR using an oligonucleotide primer in the 3.6-kb repeat (lying just to the right of 5S^{var}) and a primer unique to the rDNA repeat (lying just to the left of 5S rDNA). Our sequence was assembled from these results, and found to match the sequence of the previously determined junction¹⁶.

The complete, assembled, non-overlapping sequence of chromosome XII can be obtained at: <http://speedy.mips.biochem.mpg.de/mips/yeast/> and <http://genome-www.stanford.edu/Saccharomyces/>.

Verification of 71,072 bp of sequence determined by the EU network (64,001 bp of overlaps between cosmids sequenced independently, and 7,071 bp of selected region that were resequenced) revealed five mistakes per 10 kb, but most errors were clustered in just a few regions. Only 14 differences were found in 175,891 nucleotides that were sequenced independently by both groups; six of these were sequencing errors, leading to an error frequency of only one mistake per 29 kb. The origins of the remaining eight discrepancies were determined by sequencing PCR products of the genome of the two strains used to generate the clones. Seven of the differences are due to changes that arose in the clones, presumably during propagation in *Escherichia coli*; only one results from differences between the two yeast strains (which are isogenic, but were propagated separately for many years) used to generate the two sets of clones. Thus the number of errors in the sequence is equivalent to the number of errors resulting from propagation of the DNA in *E. coli* and yeast. □

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The nucleotide sequence of *Saccharomyces cerevisiae* chromosome XIII

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Systematic sequencing of the genome of *Saccharomyces cerevisiae* has revealed thousands of new predicted genes and allowed analysis of long-range features of chromosomal organization. Generally, genes and predicted genes seem to be distributed evenly throughout the genome, having no overall preference for DNA strand. Apart from the smaller chromosomes, which can have substantially lower gene density in their telomeric regions^{1–3}, there is a consistent average of one open reading frame (ORF) approximately every two kilobases. However, one of the most surprising findings for a eukaryote with approximately 6,000 genes was the amount of apparent redundancy in its genome. This redundancy occurs both between individual ORFs and over more extensive chromosome regions, which have been duplicated preserving gene order and orientation^{4–6}. Here we report the entire nucleotide sequence of chromosome XIII, the sixth-largest *S. cerevisiae* chromosome, and demonstrate that its features and organization are consistent with those observed for other *S. cerevisiae* chromosomes. Analysis revealed 459 ORFs, 284 have not been identified previously. Both intra- and interchromosomal duplications of regions of this chromosome have occurred.

Chromosome XIII of *S. cerevisiae* is 924,430 base pairs long, and contains 459 ORFs. Eight of these are TyA and TyB ORFs from four Ty1 retrotransposons present on chromosome XIII in strain AB972, two of

which are located on each arm of the chromosome, and these are excluded from further analyses. The average gene density on this chromosome is one ORF for every 1,997 base pairs of DNA, which correlates well with that observed for other *S. cerevisiae* chromosomes, with 74.2% of DNA on this chromosome contributing to ORFs. An average chromosome XIII ORF is 494 codons long.

Of the 451 *S. cerevisiae* ORFs on chromosome XIII, 167 (37.0%) encode previously identified proteins. A further 281 (62.3%) predicted genes have not been previously sequenced; 121 (26.8%) of these ORFs have similarities to genes for which some biochemical information is available. However, several of this category of ORF have their best protein similarity to a protein of unknown function. A total of 160 ORFs (35.5%) encode predicted proteins that are not significantly similar to proteins of known function. Because of the rapidly advancing progress of other systematic sequencing projects, many of these ORFs have homology to hypothetical proteins both in yeasts and higher organisms. A total of 51 predicted genes have similarity only to predicted proteins of unknown function. Although the majority are most similar to another *S. cerevisiae* hypothetical protein (50.9%), several have their best homology to an ORF identified in systematic sequencing of the yeast *Schizosaccharomyces pombe*⁷ (11.8%), or to predicted proteins in the nematode *Caenorhabditis elegans*⁸ (17%). Thus they are members of gene families whose function is currently unknown. There were no significant protein sequence similarities for 109 ORFs, of which 11 are thought to be questionable ORFs based on their length, codon adaptation index (CAI) value and positional base preferences.

During the systematic sequencing of other chromosomes, several putative pseudogenes were identified^{1,9}. These consisted of ORFs separated by a stop codon or frameshift from upstream or downstream sequences that shared a common homology to a single *S. cerevisiae* ORF. Most of these pseudogenes identified occur close to the telomeres of chromosomes. Three ORFs on chromosome XIII (YMR084W, YMR085W and YMR326C) have been classified as putative pseudogene ORFs. Of these, only YMR326C is located close to one of the chromosome telomeres; all three have strong similarity to sequences found elsewhere in the *S. cerevisiae* genome. These frameshifts have been confirmed by sequencing genomic DNA.

The average intergenic distance between adjacent ORFs depends on their relative orientation. This is certainly the case on chromosome XIII, in which 204 ORFs are arranged in tandem with an average intergenic distance of 450 base pairs. Of these, 110 are divergent and are an average 616 bp apart, and 111 are convergent and an average of 260 bp apart. This is consistent with a greater sequence requirement for the regulation of gene expression from promoter elements than for transcription termination.

Of the 451 ORFs on chromosome XIII, 24 (5.3%) are predicted to contain introns. There seems to be no preference for DNA strand, with 229 genes coded on the Watson strand and 222 on the Crick strand. There is no evidence of any significant clustering of related genes. However, there are several instances in which two very similar ORFs occur close to one another in tandem; for example, YMR169C and YMR170C/ALD2 (aldehyde dehydrogenases), and YMR006C and YMR008C/PLB1 (lysophospholipases).

The longest ORF on chromosome XIII is *HFA1*, (which is homologous to *FAS3*), a putative acetyl-CoA carboxylase that had been sequenced previously¹⁰ (2,123 codons). A total of 39 ORFs on this chromosome are more than 1,000 codons in length. *S. cerevisiae* genes of less than 100 codons with no homology are difficult to detect¹¹. On chromosome XIII, 10 ORFs shorter than 100 amino acids in length have been identified. The smallest of these is YMR248C, which is just 55 amino acids long, and may be spliced to a second small ORF immediately upstream. The smallest ORF on this chromosome that encodes a previously characterized protein is *COX7*, which is 59 amino acids long and encodes cytochrome oxidase polypeptide VII (ref.12).

Chromosome XIII encodes 21 predicted tRNA genes, of which six are spliced. In addition to the four Ty1 retrotransposons, several long terminal repeat (LTR) sequences are present, providing evidence of previous trans-

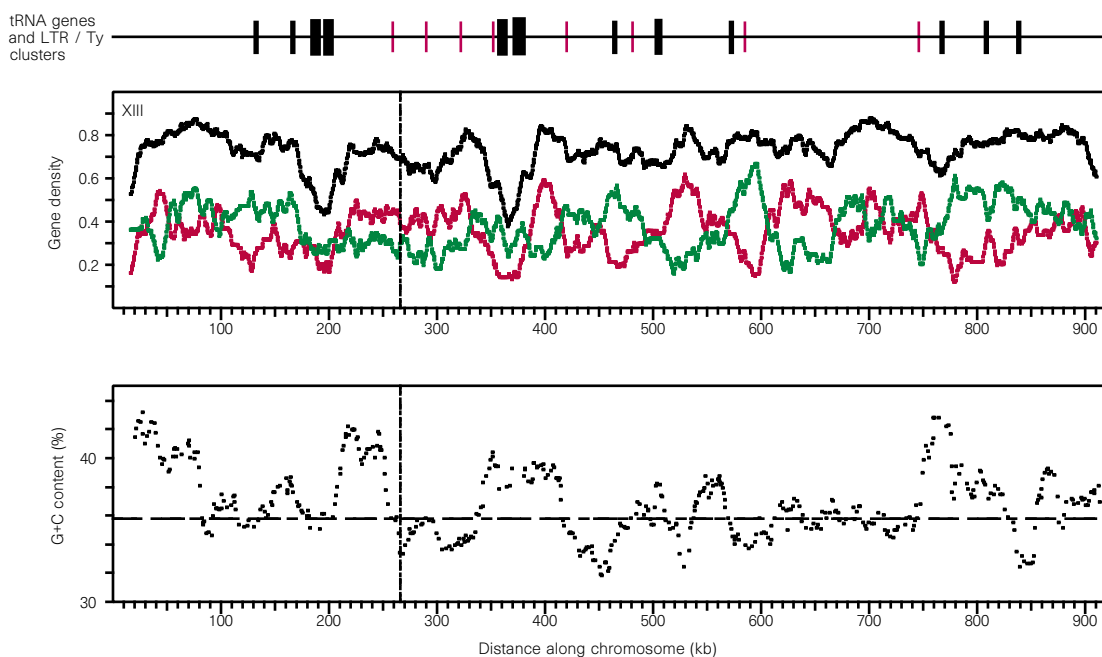


Figure 1 Overall molecular architecture of chromosome XIII. Variation in gene density (top) and base composition (bottom) along the sequence-based map of chromosome XIII (scale in kilobases from left telomere). Vertical broken lines indicate the position of the centromere. Gene density is expressed as the probability for each nucleotide to be part of an ORF. It was calculated using sliding windows of 30 kb (steps of 0.5 kb) for the Watson strand alone (red line), the Crick strand alone (green line), and the

sum of both strands (black line). Percentage of G+C was calculated from the silent positions of codons using a sliding window of 13 consecutive ORFs (horizontal broken line indicates average percentage G+C% at silent positions of codons as 35.8). Top line, positions of tRNA genes, solo LTR or Ty elements (thin small vertical lines), or clusters of them (thick small vertical lines) along the chromosome map.

position events on this chromosome. Of the 30 LTRs (whole or partial) identified on this chromosome, 29 are in close proximity to tRNA genes. There are 24 delta elements, with eight of these flanking the four retrotransposons and 16 solo elements. A further five LTRs resemble tau elements, and there is evidence of a single sigma element.

The telomeres of chromosome XIII are highly similar to the telomeres of several other *S. cerevisiae* chromosomes. Adjacent to the terminal $C_{1-3}A$ telomeric repeat at the left telomere is a Y' element, which is separated from a core X element by the subtelomeric repeats STR-D, STR-C, STR-B and STR-A¹³. The right telomere of chromosome XIII conforms to this structure but does not contain a Y' element. The right telomere shares a region of 4 kb in which the sequence is almost identical to that found at the right telomere of chromosome XV (EMBL database, accession no. SC23472). The centromere of chromosome XIII is located between bases 268,031 and 268,150; it conforms to the consensus sequence derived from the centromeres of other *S. cerevisiae* chromosomes¹⁴.

Comparison of the positions of genes on chromosome XIII with their corresponding genetically mapped loci¹⁵ shows that, as for most *S. cerevisiae* chromosomes, the two are generally in agreement. Several genes are incorrectly positioned on the genetic map in comparison with the sequence-derived map; for example, *van1* is much closer to the left telomere of chromosome XIII than expected. Small inversions are evident, for example between *UPF1* and *ADH3*, but no gross discrepancies have occurred as observed for chromosome XI (ref. 16). *MEL6* has been mapped to this chromosome, but in the *S. cerevisiae* strain S288C, from which AB972 was derived, this locus is missing. Also, the locus *SUP8* has been only tentatively assigned to the tRNA(Tyr) at base 837928 as a second tRNA(Tyr) is present on the left arm of the chromosome.

Chromosome XIII contains both intra- and interchromosomal duplications of both single ORFs and more extensive regions. The largest intra-chromosomal duplication consists of two regions approximately 40 kb to 50 kb in length containing six homologous genes in the same order and orientation with respect to each other; the duplication has occurred

between the left and right arms of the chromosome. This encompasses the region from base 32,334 to 73,917 and 790,207 to 840,147. Smaller instances of tandem gene duplication have also occurred within chromosome XIII; for example, YML125C and YML124C (*TUB3*) are similar to YML087C and YML085C (*TUB1*) on the short arm of the chromosome.

The largest interchromosomal duplication observed has occurred between a 200 kb region on the right arm of chromosome XIII and an equivalent region on the left arm of chromosome XI (in the opposite orientation). On chromosome XIII the region spans bases 303,238 to 502,733 and is bounded by ORFs YMR016C and YMR118C, on chromosome XI the coordinates of this repeat occur between bases 179,672 and 357,489. Of a total of approximately 90 genes, 17 show significant similarity and are in the same order and orientation on each chromosome. Further evidence of large scale duplications exists, including two separate regions on both chromosome IV and XVI.

Chromosome XIII has been analysed for variations in base composition, as described in ref. 9. The percentage G+C content in the third position of each codon varies throughout the length of chromosome XIII (Fig. 1), as observed through long-range analysis of other *S. cerevisiae* chromosomes. Detailed analysis at the level of individual ORFs (Fig. 2) shows that, as for chromosome IX, regions of high third-position G+C are shorter and contain fewer ORFs than regions of chromosome III that are rich in G+C. Comparison with chromosome IX shows that regions of high G+C are slightly less evident on chromosome XIII. This is also seen when plotting total G+C content for these two chromosomes: peaks of high G+C are generally lower in magnitude for chromosome XIII (results not shown).

Local areas of high G+C composition have also been observed in several of the intergenic regions of this chromosome, for example regions at approximately 119, 306, 541 and 653 kb are G+C rich. It is possible that these intergenic regions of high G+C may contain previously unidentified, small *S. cerevisiae* genes. These areas do not appear to correlate with sequences of high coding potential. However, this does not exclude the

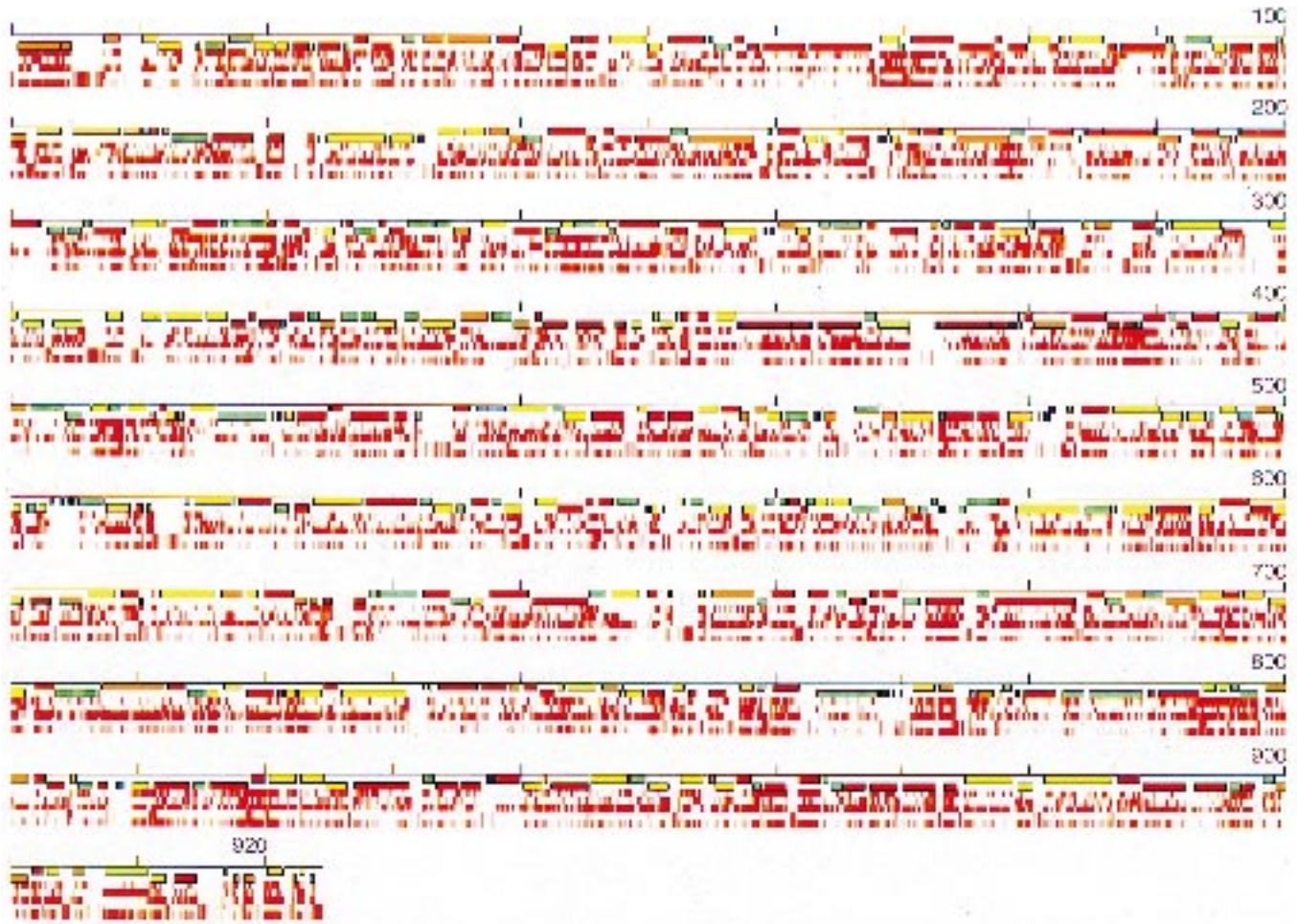


Figure 2 G+C composition of chromosome XIII. Chromosome XIII and its features are drawn to scale. The top graduated line represents the chromosome split into segments of 100 kb, with ORFs indicated below this as coloured boxes. ORFs located on the Watson strand are shown above those on the Crick strand. ORFs encoding previously identified genes are shown in red, those with similarities to hypothetical proteins in orange, and those with no significant similarities in green; pseudogene ORFs are shown in blue. tRNA genes and transposon-derived ORFs are shown in white boxes, with LTRs shown in dark blue (δ), turquoise (τ) and pink (σ).

Below this, variations in G+C composition (calculated using a sliding window of 200 bases) are shown as bars, with gradations of red varying from 35% to 45% G+C content (P. R., unpublished data). Areas of less than 35% are shown in white, with those over 45% red. Five bars of G+C variation are shown, the lowest bar shows total G+C content; above this a second shows G+C content in intergenic regions alone. The G+C composition in each of the three bases of each codon are shown above this, with the central of the five bars representing first position G+C, the next representing second position G+C, and the top bar showing third position G+C.

possibility that coding sequences exist in these areas. No similarities to other *S. cerevisiae* ORFs were found using the program BLASTX, indicating that these areas rich in G+C are not pseudogene remnants. Other possibilities for these G+C peaks in intergenic regions have not yet been fully investigated.

Few ORFs show a high incidence of high G+C content in the second position of their codons. The most pronounced example on chromosome XIII is YMR317W, which encodes a protein rich in serine and threonine residues, and is located close to the right telomere of the chromosome.

Analysis of other *S. cerevisiae* chromosomes has demonstrated that, in general, areas high in G+C content correlate with high gene density, and that regions around chromosome centromeres and telomeres appear to be G+C poor^{4-6,16}. This is also the case for chromosome XI11. However, the significance of these areas of high and low G+C content is not yet clear.

Two of the genes located on chromosome XIII have human homologues that have been implicated in certain forms of cancer. *MLH1* encodes a DNA mismatch repair protein that is very similar to a human protein defective in some forms of hereditary non-polyposis colon cancer¹⁷. In *S. cerevisiae*, null mutants of this gene are viable, but show an elevated rate of spontaneous mutations and an increased instability of simple repeat sequences^{18,19}. A second gene, *SGS1*, is homologous to the human *BLM* gene, which is involved in Bloom's syndrome²⁰. Both the

human and yeast genes encode DNA helicases. Mutations in the human gene confer a predisposition to many types of cancer and also cause other clinical defects²⁰. In *S. cerevisiae*, null mutants are viable, but again show genomic instability^{21,22}. Future analysis of these and other *S. cerevisiae* genes should assist the understanding of the molecular mechanisms underlying many human diseases.

Chromosome XIII is the largest *S. cerevisiae* chromosome to be sequenced by a single laboratory. Analysis shows that its features are typical of large *S. cerevisiae* chromosomes. □

Methods

Sequencing. Chromosome XIII was sequenced using a cosmid-based strategy that uses DNA isolated from the S288C-derived strain AB972. Methods used for sequence generation and assembly have been described in detail elsewhere^{8,23}. Cosmids were chosen from the map generated by L. Riles and M. Olson (personal communication), giving rise to two large contigs with a central gap. This was filled using lambda clones L-6543 and L-6223. The two chromosome telomeres, pEL161H (left) and pEL175H (right), were provided as plasmid clones²⁴ by E. Louis. Two further lambda clones, L-4987 and L-7056, were sequenced to complete the left end of the chromosome. Sequence from pEL161H overlapped L-4987, so no further gap filling was required at the left telomere of chromosome XIII. However, at the right telomere a gap between cosmid clone 9924 and plasmid pEL175H remained, and

was filled using long-range polymerase chain reaction (PCR) from *S. cerevisiae* genomic DNA. The sequence generated for this chromosome extends into the C₁₋₃A telomeric repeat sequences on both chromosome arms, although the exact number of these repeats has not been determined. Sequencing was considered to be finished when each base had been sequenced on both strands and all ambiguities had been resolved.

Analysis. For each completed clone, a consensus of the nucleotide sequence was generated in the Staden sequence assembly package XBAP²⁵, flanked by short regions of sequence overlapping neighbouring clones. This sequence was analysed primarily within the DIANA (Display and Analyse) package (T. Horsnell and B. B., unpublished), a sequence editor with a graphical interface. ORFs equal to or greater than 100 codons in length were marked and trimmed to their first methionine. Each ORF was screened against the SWIR database, a non-redundant compilation of the protein databases Swiss-Prot²⁶, TrEMBL²⁷ and WormPep, using the program FASTA²⁸ with limited optimization. The consensus sequence for each clone was screened against SWIR using BLASTX²⁹, and EMBL/EMNEW using BLASTN²⁹, to detect small ORFs less than 100 amino acids in length, other genome features, and local similarity. Some features were specifically identified; Prosite³⁰ amino-acid motifs (regular expression searching), transposon LTRs (CGC Wordsearch/Segments) and tRNAs (tRNA scan). Individual annotated clones were submitted to the EMBL database within days of being finished. The complete chromosomal sequence was built from overlapping clones and also submitted to the EMBL database as a single record (accession no. SCCHR XIII, Z271257).

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The nucleotide sequence of *Saccharomyces cerevisiae* chromosome XIV and its evolutionary implications

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In 1992 we started assembling an ordered library of cosmid clones from chromosome XIV of the yeast *Saccharomyces cerevisiae*. At that time, only 49 genes were known to be located on this chromosome¹ and we estimated that 80% to 90% of its genes were yet to be discovered. In 1993, a team of 20 European laboratories began the systematic sequence analysis of chromosome XIV. The completed and intensively checked final sequence of 784,328 base pairs was released in April, 1996 (ref. 2). Substantial parts had been published before³⁻²² or had previously been made available on request. The sequence contained 419 known or presumptive protein-coding genes, including two pseudogenes and three retrotransposons, 14 tRNA genes, and three small nuclear RNA genes. For 116 (30%) protein-coding sequences, one or more structural homologues were identified elsewhere in the yeast genome. Half of them belong to duplicated groups of 6-14 loosely linked genes, in most cases with conserved gene order and orientation (relaxed interchromosomal synteny). We have considered the possible evolutionary origins of this unexpected feature of yeast genome organization.

Figure 1 shows the map of cosmid, lambda and plasmid clones and of polymerase chain reaction (PCR) fragments from two unclonable regions which were used to determine the sequence of chromosome XIV. The final positions of genes listed in the 1992 map¹ are also presented changing the order of closely linked genes in only three regions. The assembled contig consists of 784,328 bp. The sequence of 180,983 bp (23%) was independently determined twice on both strands. These control sequences included 28 overlapping regions of cosmid and lambda clones (117,891 bp) as well as 108 selected regions, mainly at termini of open reading frames (ORFs), resequenced either on cosmids (54,540 bp) or by genomic PCR (8,552 bp). A total of 27 sequence mistakes were corrected. We estimate that the final sequence carries less than one error in every 10 kilobases, an estimate confirmed by a recent independent control analysis using 83 randomly picked genomic clones of chromosome XIV (G. Valle, unpublished data). Among the 40 kb sequenced, four deviations from our final sequence were noted: three single base-pair changes with neutral effects on coding regions (probably resulting from strain or clone differences), and only one confirmed sequence mistake. The left end of the chromosome carries telomeric repeat sequence (see below) and it is

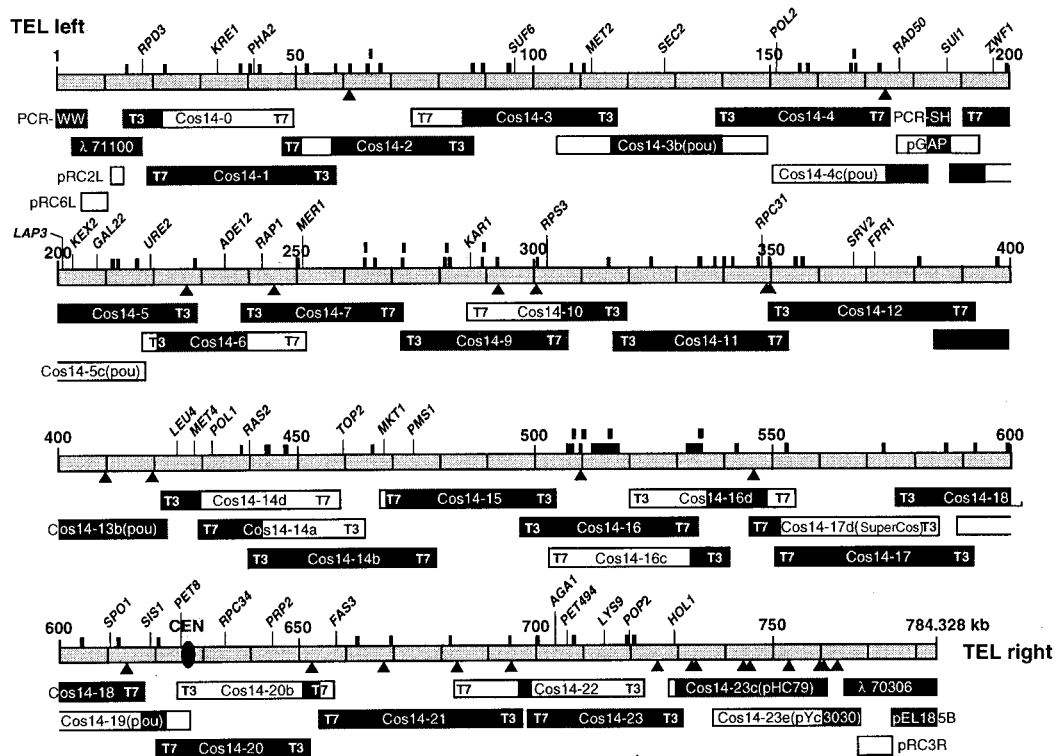


Figure 1 Physical map of subclones of chromosome XIV used for systematic DNA sequence analysis and final locations of originally genetically mapped genes¹. Position of cosmid clones (cos), lambda clones (λ), plasmid clones (p) and genomic PCR fragments (PCR) are drawn as overlapping bars. Sequenced regions are shown in black. The 108 short regions selected for verification analyses are shown as small bars (resequenced cosmid clones) or triangles (sequenced genomic PCR fragments) along the contig. All clones were derived from S288C strains, except plasmid pGAP (carrying a spontaneous nonsense mutation in the toxic YNL247w), which originates from strain A364a³⁶. Most cosmid clones with chromosome XIV DNA were isolated from cosmid libraries provided by B. Dujon³⁷ and R. Stucka³⁸, and mapped by a modified chromosome fragmentation approach^{39,40}. Several clones extending into or bridging remaining gaps were isolated by colony screening using non-radioactively labelled restriction fragments as hybridization probes. Two cosmid clones (14-17d and 14-23c) and both lambda clones carrying telomere DNA were obtained from L. Riles⁴¹ and the right telomere clone pEL185 was provided by E. Louis⁴². A more detailed description of the mapping strategy will be published elsewhere (K. Ham-

berg *et al.*, manuscript in preparation). The complete sequence can be retrieved from the EMBL database, accession nos Z71277-Z71692 or from the Martinsrieder website². Different parts were sequenced in different laboratories: 1-6035 (R. Wambutt); 3,203-17,700 (B. Obermeier); 13,990-22,212 (A. Goffeau); 18,699-58,748 (C. Gaillardin); 47,022-51,246 and 57,523-87,525 (R. J. Planta); 85,152-132,424 (N. N. Glansdorf); 130,724-187,891 (J. H. Hegemann); 183,004-187,900 (P. Philippsen); 187,809-192,153 (F. Del Rey); 192,154-195,234 (A. Jimenez); 190,506-229,360 (G. Valle); 220,854-239,907 (A. Dusterhöft); 238,582-273,742 (A. Goffeau); 271,932-319,898 (H. Domdey); 317,148-353,960 (C. Herbert); 349,559-393,039 (M. Jacquet); 384,059-421,858 (G. Valle); 421,188-443,100 (J.-L. Revuelta & F. Del Rey); 443,001-456,300 (A. Jimenez); 456,201-479,289 (J. P. G. Ballesta); 468,833-504,727 (P. Philippsen); 496,969-541,433 (M. Crouzet); 536,275-549,131 (C. Herbert); 545,180-592,214 (A. Dusterhöft); 575,858-617,912 (A. Urrestarazu); 617,105-622,324 (M. Crouzet); 620,016-652,539 (G. Volckaert.); 650,830-653,557 (R. J. Planta.); 651,995-654,446 (A. Dusterhöft); 654,389-731,357 (T. M. Pohl); 729,267-768,530 (A. Dusterhöft); 764,973-784,328 (A. Urrestarazu); 774,980-784,145 (C. Gaillardin).

Table 1 *S. cerevisiae* chromosome XIV ORFs and structurally homologous ORFs of other chromosomes

| ORF Chr.XIV [†] | ORF Chr.VI [†] | % identity/ stretch of amino acids | Biochemical or biological function (gene name) Chr. XIV ORF | Homologue in cluster duplication |
|--------------------------|-------------------------|---------------------------------------|--|---|
| 6-17.3 kb [†] | 6-15.5 kb | | | |
| YNL336w | YFL062w | 94.2% overall | unknown | unknown |
| YNL335w | YFL061w | 100 % overall | fungal cyanamide hydratase homologue | fungal cyanamide hydratase homologue |
| YNL334c | YFL060c | 99.1% overall | unknown, probable membrane protein | unknown, probable membrane protein |
| YNL333w | YFL059w | 99.7% overall | unknown | unknown |
| YNL332w | YFL058w | 99.7% overall | thiamine regulated protein homologue | thiamine regulated protein (THI5) |
| YNL331c | YFL057-56c | 87.0%/226 aa [‡] | probable aryl-alcohol reductase | probable aryl-alcohol reductase |
| ORF Chr.XIV | ORF Chr.XV (A) | % identity/ stretch of amino acids | | |
| 38.7-106.7 kb | 25.3-142.6 kb | | | |
| YNL318c | YOL156w | 38.4%/510 aa | hexose transporter (HXT14) | glucose transporter (LGT3) |
| YNL307c | YOL128c | 41.6%/316 aa | Ser/Thr/Tyr protein kinase (MCK1) | probable Ser/Thr protein kinase |
| YNL302c | YOL121c | 99.3% overall | ribosomal protein (RPS16A) | ribosomal protein (RPS16B) |
| YNL301c | YOL120c | 100% overall | ribosomal protein (RP28B) | ribosomal protein (RP28A) |
| YNL299w | YOL115w | 54.1%/556 aa | topoisomerase I related protein (TRF5) | topoisomerase I related protein (TRF4) |
| YNL298w | YOL113w | 61.2%/317 aa | Ser/Thr protein kinase (CLA4) | probable Ser/Thr protein kinase |
| YNL293w | YOL112w | 53.2%/417 aa | unknown | unknown |
| YNL290w | YOL094c | 34.4%/317 aa | replication factor C subunit (RFC3) | replication factor C subunit (RFC4) |
| YNL283c | YOL105c | 43.4%/302 aa | similarity to yeast chitinase | unknown |
| ORF Chr.XIV | ORF Chr.IV | % identity/ stretch of amino acids | | |
| 252.1-307 kb | 44.1-80.4 kb | | | |
| YNL209w | YDL229w | 99.3% overall | heat shock protein (SSB2) | heat shock protein (SSB1) |
| YNL204c | YDL226c | 30.5%/177 aa | sporul.spec.zinc finger protein (SPS18) | prolif.spec.zinc finger protein (GCS1) |
| YNL197c | YDL224c | 36.5%/581 aa | regulator of cell size (WHI3) | unknown |
| YNL194c | YDL222c | 52.0% overall | unknown, probable membrane protein | unknown, probable membrane protein |
| YNL183c | YDL214c | 42.6%/479 aa | Ser/Thr protein kinase (NPR1) | probable Ser/Thr protein kinase |
| YNL176c | YDL211c | 24.3%/292 aa | unknown, probable membrane protein | unknown, probable membrane protein |
| ORF Chr.XIV | ORF Chr.VIII | % identity/ stretch of amino acids | | |
| 309-410 kb | 390.3-341.4 kb | | | |
| YNL173c | YHR146w | 27.4%/351 aa | pheromone-response G protein | unknown, probable G protein |
| YNL162w | YHR141c | 100% overall | ribosomal protein (RPL41A) | ribosomal protein (RPL41A) |
| YNL160w | YHR139c | 45.0%/307 aa | secreted glycoprotein (YGP1) | sporulat.spec. wall maturation (SPS100) |
| YNL156c | YHR133c | 40.5%/205 aa | unknown | unknown |
| YNL154c | YHR135c | 70.9%/499 aa | casein kinase I isoform (YCK2) | casein kinase I (YCK1) |
| YNL144c | YHR131c | 36.2%/464 aa | unknown | unknown |
| YNL130c | YHR123w | 53.8% overall | diacylglyc.choline-P transferase (CPT1) | ethanolamin P-transferase (EPT1) |
| YNL121c | YHR117w | 49.3%/651 aa [§] | import recept.mito outer memb.(TOM70) | mitochondrial outer membrane protein |
| YNL116w | YHR115c | 55.4%/424 aa | unknown | unknown |
| ORF Chr.XIV | ORF Chr.XV (B) | % identity/ stretch of amino acids | | |
| 419-466 kb | 529-486.8 kb | | | |
| YNL108c | YOR110w | 65.0%/273 aa | unknown | unknown |
| YNL106c | YOR109w | 58.5%/979 aa | inositol phosphatase homologue | probable phosphatase |
| YNL104c | YOR108w | 88.5%/601 aa | 2-isopropyl malate synthase (LEU4) | 2-isopropyl malate synthase homologue |
| YNL098c | YOR101w | 55.8%/303 aa | GTP-binding protein (RAS2) | GTP-binding protein (RAS1) |
| YNL096c | YOR096w | 87.9% overall | ribosomal protein S7 homologue | ribosomal protein (RP30) |
| YNL095c | YOR092w | 55.3%/445 aa | unknown, probable membrane protein | unknown, probable membrane protein |
| YNL093w | YOR089c | 56.9%/209 aa | GTP-binding protein (YPT53) | GTP-binding protein (VPS21) |
| YNL090w | YOR089c | 56.9%/209 aa | GTP-binding protein (RHO2) | GTP-binding protein (VPS21) |
| YNL087w | YOR086c | 54.5% overall | unknown, probable membrane protein | unknown, probable membrane protein |
| ORF Chr.XIV | ORF Chr.IX | % identity/ stretch of amino acids | | |
| 478.6-597.6 kb | 89.3-202.1 kb | | | |
| YNL079c | YIL138c | 54.1%/159 aa | tropomyosin (TPM1) | tropomyosin (TPM2) |
| YNL074c | YIL135c | 23.2%/375 aa | unknown | unknown |
| YNL069c | YIL133c | 90.3% overall | ribosomal protein (RP23) | ribosomal protein (RP22) |
| YNL068c | YIL131c | 52.1%/190 aa | unknown, fork head domain (FKH2) | unknown, fork head domain (FKH1) |
| YNL066w | YIL123w | 62.9%/415 aa | β-glucosidase homologue (SUN4) | homologue of aging gene UTH1 |
| YNL065w | YIL121w | 47.7%/342 aa | cycloheximid resist.protein homologue | antibiotic resistance protein homologue |
| YNL065w | YIL120w | 43.0%/351 aa | cycloheximid resist.protein homologue | antibiotic resistance protein homologue |
| YNL058c | YIL117c | 37.3%/126 aa | unknown | unknown |
| YNL055c | YIL114c | 49.5% overall | outer mito membrane porin (OMP2) | OMP2 homologue |
| YNL053w | YIL113w | 48.8%/162 aa | protein phosphatase (MSG5) | protein-Tyr phosphatase homologue |
| YNL052w | YIL111w | 63.6% overall | cytochrome c oxidase (COX5A) | cytochrome c oxidase (COX5B) |
| YNL049c | YIL109c | 61.8%/566 aa | unknown | unknown |
| YNL047c | YIL105c | 54.3%/639 aa | unknown | unknown |
| YNL037c | YIL094c | 35.8%/296 aa | isocitrate dehydrogenase (IDH1) | isopropyl malate dehydrog. homologue |
| YNL029c | YIL085c | 55.7%/476 aa | mannosyl transferase homologue | mannosyl transferase homologue |
| YNL020c | YIL095w | 41.2%/636 aa | probable Ser/Thr protein kinase | probable Ser/Thr protein kinase |
| ORF Chr.XIV | ORF Chr.III | % identity/ stretch of amino acids | | |
| 623.4-753.7 kb | 101.7-301.8 kb | | | |
| YNL004w | YCL011c | 39.4%/409 aa | poly(A)bdg.protein homologue (TOM34) | probable TEL associated protein (GBP2) |
| YNR001c | YCR005c | 81.4%/441 aa | citrate synthase (CIT1) | peroxysomal citrate synthase (CIT2) |
| YNR002c | YCR010c | 77.7% overall | unknown, probable membrane protein | unknown, probable membrane protein |
| YNR013c | YCR037c | 48.1%/489 aa | unknown, probable membrane protein | probable phosphate transporter (PHO87) |
| YNR019w | YCR048w | 52.5%/459 aa | sterol acyltransferase (SAT1) | cholesterol acyltransferase (ARE1) |
| YNR023w | YCR052w | 29.5%/353 aa | unknown | unknown |
| YNR026c | YCR067c | 45.4%/388 aa | GTP-GDP exchange factor (SEC12) | ER protein (SEC4) |
| YNR028w | YCR069w | 33.2%/304 aa | peptidyl-prolyl isomerase homologue | peptidyl-prolyl cis-trans isom.(SCC3) |
| YNR031c | YCR073c | 53.6%/1172 aa | MAPKKK high osm.sign.transd. (SSK2) | MAP kinase kinase kinase (SSK2) |
| YNR034w | YCR073w-a | 76.9% overall | multicopy sup of los1-1 (SOL1) | Glcn-6-P deaminase homologue (SOL2) |
| YNR047w | YCR091w | 72.4%/424 aa | probable Ser/Thr protein kinase | probable Ser/Thr protein kinase (KIN82) |
| YNR048w | YCR094w | 65.4% overall | unknown | unknown |
| YNR065-66c | YCR099-101c | 64.8%/637 aa [†] | peptidase Y sorting protein (pseudogene) | peptidase Y sorting (PEP1 homologue) |

Degrees of homology were extracted from pairwise FASTA alignments of deduced protein sequences and are listed as percentage identity per stretch of amino acids.

[†]Y is included in Fig. 2 in the cluster duplication but is not listed in this table.

[‡]Coordinates of clusters.

[§]Overall homology between YNL331c and the sum of YFL056c and YFL057c (pseudogene in chromosome VI?).

[¶]Gaps introduced by the alignment algorithm may result in homology stretches slightly longer than the protein sequences.

^{††}Overall homology of the pseudogene (YNR065-YNR066c) to the sum of YCR099c, YCR100c and YCR101.

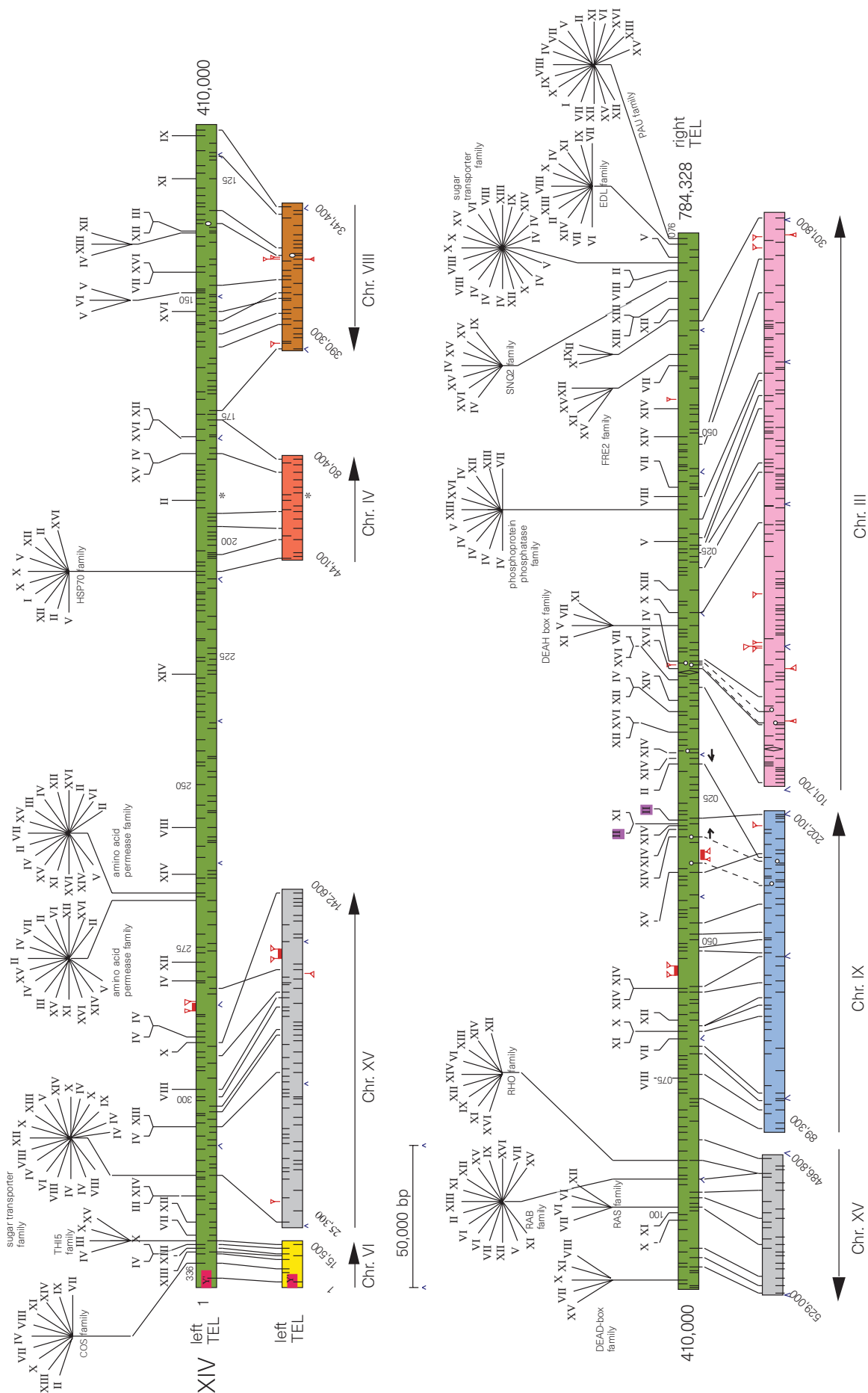


Figure 2 Map of chromosome XIV ORFs that are members of either multi-gene families or of pairs, triplets or quadruplets of structurally related *S. cerevisiae* ORFs. The green bar represents both strands of chromosome XIV, with centres of all ORFs (excluding Ty and four short telomere ORFs) drawn as vertical lines; 215 are coded by the upper strand and 195 by the lower strand. Vertical lines with open circles mark selected tRNA genes. Three-digit numbers beneath or above the green bar refer to the systematic ORF nomenclature starting with 1 at either side of the centromere (white dot at 628 kb). Lines above or below the green bar indicate ORFs with structural homologues elsewhere in the genome (at least 30% identity in 150 amino acids, or, in a few cases, 25% in 300 amino acids). Lines extending into branches mark multigene families, with roman numbers indicating members on different chromosomes (order of decreasing homology to the chromosomes XIV ORF from left to right or clockwise, respectively). Coloured bars below chromosome XIV display seven syntenic or partly syntenic segments of other chromosomes with accumulations of ORFs structurally related to and arranged similarly to chromosome XIV ORFs. Broken lines in three of these clusters connect positions of pairs of functionally identical tRNA genes. The star at 280 kb indicates a functional ARS element on chromosome XIV (Ref. 36) which seems to be positionally conserved on chromosome IV. Further details of these cluster duplications are given in Table 1. The red bar of the left telomere represents the ubiquitous Y' element found at many ends of *S. cerevisiae* chromosomes^{43,44}. Red triangles mark positions of solo delta sequences (remnants of Ty elements) and red bars flanked by triangles indicate Ty elements. The two black arrows at 570 kb and 600 kb indicate an intra-chromosomal highly conserved inverted repeat, involving in each repeat element one tRNA^{leu} gene and two new ORFs (YNL034w-YNL035w and YNL019c-YNL018c, respectively). The two marked chromosome II homologues and the corresponding chromosome XIV ORFs at 575 kb represent the two copies of the duplicated histone H3-H4 gene pair. As indicated there is an additional homologue to histone H3 on chromosome XI (*CSE4*), probably the yeast homologue of the human *CENP-A* gene⁴⁵.

possible that this end is a few hundred base pairs longer than indicated in Figure 1.

A systematic search of the chromosome XIV contig revealed 414 ORFs with 100 and more codons, including overlapping ORFs but excluding ORFs located within longer ORFs on either the same or the complementary strand. Chromosome XIV also has at least seven ORFs with less than 100 codons, of which four are known genes (*MFA2*, *TOM7*, *ATX1* and *PBI2*) and three show significant homology to known genes. A systematic nomenclature was given to all ORFs (excluding the six ORFs of the three Ty retrotransposons), indicating the organism (Y), the chromosome (N), the chromosome arm (L or R), the coding strand (Watson, w or Crick, c), and increasing numbers starting at the centromere; examples include YNL001w and YNR001c.

A simultaneous search for introns (using the EXPLORA program²³) revealed 16 intron-carrying genes, in two of which, (YNL066w and YNL065w) the introns are located in the non-translated 5' region^{24,25}. EXPLORA failed to locate an additional, experimentally verified intron in ORF YNL044w, because it has an unusual 5' splice sequence (EMBL database, accession nos X97400 and X97401).

Two pairs of adjacent ORFs (YNR065c and YNR066c; and YNR068c and YNR069c) were separated only by a stop codon; this was confirmed in both cases by genomic PCR. These pairs are rare examples of yeast pseudogenes, as highly conserved copies lacking internal stop codons are present on other chromosomes. Like their functional homologues these pseudogenes should be considered as single ORFs. Taking this into account, 419 ORFs are located on chromosome XIV, including six Ty elements and 23 questionable ORFs (short ORFs overlapping longer ones). The ORF density, not counting questionable ORFs, is one ORF per 1.98 kb (a total of 396 ORFs in 784 kb), and the average ORF size is 1.5 kb. These numbers are very similar to corresponding numbers obtained with other *S. cerevisiae* chromosomes. The ORF density (the ratio of ORF nucleotides to total nucleotides) fluctuates between 0.6 and 0.9. These fluctuations do not correlate with fluctuations in G+C content; five of

eight ORF density peaks coincide with regions of highest G+C content (39.4–40.0%) and the other three with regions of lowest G+C content (36.6–37.7%).

How many of the 396 non-questionable ORFs are new? Presently, functions are known, at least partly, for 149 ORFs (38%), based on detailed experiments or very high sequence homology to known genes^{2,26}. Most of these are involved in metabolism, cell growth, cell division, translation, transcription and intracellular transport, with a few involved in energy production, metabolite transport, protein modification, signal transduction, and stress response. Of the 247 new ORFs, some functional predictions can be made for 43 (11%), owing to homologies to characterized genes in *S. cerevisiae* or other organisms. Presumptive products coded by these 43 ORFs include: a human breast cancer-associated autoantigen homologue; a genetically linked cluster of three proteins (transporter, epimerase and reductase) for potential utilization of an unidentified mono- or oligosaccharide; four proteins with homology to prokaryotic ribosomal proteins; three protein kinases; three GTP-binding proteins; two protein phosphatases; two translation factors; two drug-resistance proteins; one actin homologue; one zinc finger; one peptidyl-prolyl isomerase; and ten with presumptive metabolic activities, such as cyanamide hydratase, mannosyl transferase, isocitrate dehydrogenase and inositol phosphatase. Further details can be found on the Martinsrieder website².

The functions of the other 204 ORFs (51%) cannot yet be predicted. One third of these code for presumptive membrane proteins, and more than four transmembrane domains are predicted for 18. Of the 204 new ORFs, 12 have homology to human expressed sequence tags (EST)^{27, 28} with FASTA scores of 200–760. Remarkably, two of the 23 questionable ORFs (YNL228w and YNL114c) also have significant homology to human EST sequences.

We used FASTA comparisons of all chromosome XIV ORFs (except the highly repetitive Y' and Ty ORFs) to all *S. cerevisiae* ORFs in order to establish the extent of gene duplications, and found that 116 ORFs shared structural homology with one or more ORFs elsewhere in the genome. For this search, structural homology was defined as over 30% identity in a stretch of 150 amino acids (in some cases, 25% identity in a stretch of at least 300 amino acids). Of these 116 ORFs, 67 belong to pairs of homologues, 32 to groups consisting of three or four homologues, and 17 are members of multigene families. ORFs from all chromosomes contribute to this picture of sequence homology (Fig. 2). The list of homologies based on FASTA analyses also revealed several regions of chromosome XIV with accumulations of homologous ORFs originating from distinct regions of six other chromosomes, and showing, with only a few exceptions, conserved gene orders and gene orientations. One of these apparently ancient duplications, involving ORFs of the left arms of chromosomes IX and XIV, respectively, had previously been reported^{19,29}. Duplications involving several genes had been described up to that time, mainly for relatively short subtelomeric and centromeric regions^{30–34}.

The extent of these types of duplications became apparent after the complete sequence information of the *S. cerevisiae* reference strain S288C was released². With respect to chromosome XIV, so-called gene cluster duplications were found in seven regions of 17 kb to 130 kb. The precise locations of the 67 pairs of ORF homologues in these seven cluster duplications are shown in Fig. 2, together with all other chromosome XIV ORFs for which structural homologues were found; five pairs of positionally conserved duplicated tRNA genes are also indicated. Probably half of these structural homologies among different chromosomes would have remained undetected in classical DNA hybridization experiments.

Complementary to the graphical display of the seven cluster duplications, we have determined the degree of homologies for each ORF pair and, if known or predictable, their functions (Table 1). ORFs displayed from left to right in Fig. 2 are listed from top to bottom in the table. An automated means of finding and displaying structurally homologous segments in genomes several million base pairs long involves the screening of sliding windows of 500 bases between pairs of chromosomes³⁵. This very efficient method was also applied to chromosome XIV, and most of the ORF pairs participating in cluster duplications were detected (K. Heumann, unpublished data). However, this automated approach still

requires manual editing to find all details of cluster duplications, such as multigene families, potentially inverted ORF members, more than averagely diverged ORFs, and tRNA genes.

The 17-kb subtelomeric cluster duplication between chromosomes XIV and VI (cluster duplications 14–6) consists entirely of highly conserved ORF pairs (average 96.6% amino-acid identity) and shows stringent synteny. The intergenic regions are also highly conserved, suggesting that the duplication of the six ORFs is a relatively recent event on an evolutionary timescale.

Most of the ORF pairs in the other six cluster duplications are much less conserved, and their promotor and terminator regions lack significant homologies, suggesting that they are ancient duplication events. Five of the highly conserved ORF pairs of these ancient duplications code for ribosomal proteins (average 95.3% amino-acid identity), one for two members of the 70K heat-shock protein family (99.3% amino-acid identity), one for two forms of iso-propyl malate synthase (88.5% amino-acid identity) and one for two forms of citrate synthase (81.4% amino-acid identity) (Table 1). Excluding these ORF pairs, which are apparently under high selection pressure to preserve their sequence information, the average homology of ORF pairs was determined for each of the cluster duplications. ORF pairs in cluster duplications CD14–15B and CD 14–3 (average 56% amino-acid identity) seem to be less diverged than ORF pairs in CD14–15A, CD14–8, CD14–9 (average 47.5% amino-acid identity) and CD14–4 (average 37% amino acid identity). However, there are too few ORF pairs to draw conclusions about different temporal orders for the cluster duplications involving chromosome XIV.

Could the six ancient cluster duplications, at the time of their creation, have looked similar to the recent cluster duplications between chromosomes XIV and VI, with perfect synteny of all ORFs? And could they have been shaped over evolutionary time by base-pair changes, insertions of new ORFs, deletions of some of the originally duplicated ORFs, inversions of single or groups of ORFs, and translocations to yield the present picture of 'relaxed synteny'? This is certainly possible if the now visible arrangements indeed originated from duplications of gene clusters, perhaps by long-range gene conversions or chromosome duplications. However, it remains possible that the evolutionary history of *S. cerevisiae* involved fusion of two ancient forms of yeast cells with smaller genomes already displaying sequence divergencies and some level of relaxed synteny and that, for most of the duplicated ORFs, one copy was lost over time because of a lack of selective advantage for *S. cerevisiae* to keep more than one copy. □

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The nucleotide sequence of *Saccharomyces cerevisiae* chromosome XV

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Chromosome XV was one of the last two chromosomes of *Saccharomyces cerevisiae* to be discovered¹. It is the third-largest yeast chromosome after chromosomes XII and IV, and is very similar in size to chromosome VII. It alone represents 9% of the yeast genome (8% if ribosomal DNA is included). When systematic sequencing of chromosome XV was started, 93 genes or markers were identified, and most of them were mapped². However, very little else was known about chromosome XV which, in contrast to shorter chromosomes, had not been the object of comprehensive genetic or molecular analysis. It was therefore decided to start sequencing chromosome XV only in the third phase of the European Yeast Genome Sequencing Programme, after experience was gained on chromosomes III, XI and II (refs 3–5). The sequence of chromosome XV has been determined from a set of partly overlapping cosmid clones derived from a unique yeast strain, and physically mapped at 3.3-kilobase resolution before sequencing. As well as numerous new open reading frames (ORFs) and genes encoding tRNA or small RNA molecules, the sequence of 1,091,283 base pairs confirms the high proportion of orphan genes and reveals a number of ancestral and successive duplications with other yeast chromosomes.

The DNA sequence of 1,091,283 nucleotides contains 560 ORFs, of at least 100 sense codons, that are not entirely included within a larger one (our standard basic definition; see ref. 4). If those corresponding to Ty or Y' elements are excluded, and intron predictions are considered (see below), 551 different ORFs remain. To these were added eight known genes shorter than 100 codons (*BAT2*, *CRS5*, *RPB10*, *RPS30B*, *RPS33A*,

SME1, *TOM6* and the *CPA1* leader), and a pseudogene 581 codons long (YOL153c) that contains two in-frame ochre codons. It is considered here because its putative translation product has significant homology with the Gly-X carboxypeptidase encoded by *CPS1*, and because, in another yeast strain, the two stop codons are replaced by two glutamine codons CAA⁶. Note that, in the present sequence, YOR031w (*CRS5*) also contains an in-frame ochre codon instead of the CAA codon found in other strains (the reality of the stop codon was verified by direct sequencing on yeast DNA). Other interesting but more complex cases of pseudogenes found in chromosome XV will be described elsewhere. Also note that YOL040c has been considered here (coordinates 253,147–253,575) instead of the larger antisense ORF within which it is entirely included because it corresponds to a known gene (*RPS21*).

The sequence also reveals 526 short ORFs (from 50 to 99 sense codons) not entirely included within larger ones, six of which are already considered above. In this size range, it is obviously difficult to distinguish actual genes from random occurrences. Using high codon bias (CAI > 0.2) and absence of a partial overlap with larger ORFs as predictive criteria, only six candidates remained. One of these shows very significant homology with the 60 codon-long gene *HOR7* of chromosome XIII. This ORF, named YOL052ca, has been added to the above list, bringing the grand total to 561. Of these ORFs 33 (5.9%) are 'questionable', based on their short size and low CAI (see ref. 4), and 18 of them partly overlap other ORFs, increasing their suspicious character.

The longest ORF of chromosome XV is YOL081w (the *IRA2* gene) with 3,079 codons. The two shortest ORFs, if the 25 amino-acid leader peptide of the *CPA1* gene is ignored, are YOR045w (*TOM6*) and YOL052ca, with 61 codons each. The average size of chromosome XV ORFs is 457 codons, very close to the figure observed for the entire yeast genome⁷. In total, 33 pairs and 5 trios of partly overlapping ORFs are found; 25 pairs are antiparallel, excluding the possibility of sequencing errors. Among the parallel pairs, YOR012w and YOR013w are suggestive of a frameshift error, or a pseudogene, as their products share homology with the amino terminus and the carboxy terminus of the product of YDR391c, respectively.

The overall density for protein-coding genes is 70.6%, slightly lower than the average for the whole yeast genome subtracted from rDNA⁷. Variations are observed along the chromosome, with two short regions showing gene density above 85% (centred around ~200 kb and ~950 kb, respectively; see Fig. 1). These regions also correspond to areas of high G+C content. As is generally observed in yeast, the two subtelomeric regions show low gene density.

The entire chromosome shows no significant strand coding bias, but important local variations are observed, with seven short regions (of ~30–50 kb each) showing a clearcut excess of ORFs on the Watson strand; eight others have an excess on the Crick strand (Fig. 1). Orientation of neighboring ORFs is random for the whole chromosome, with 150 diverging pairs, 149 converging pairs, and 261 tandemly arranged pairs (123 on the Watson strand, and 138 on the Crick strand). The longest tandem array contains 11 successive ORFs (YOR104w to YOR114w, coordinates 517,639–538,451).

A total of 13 introns have been identified, most of which occur in short ORFs with high codon bias. Chromosome XV introns are short, as is typical for yeast, ranging in size from 135 to 527 nucleotides (average 334 nucleotides). They have an average G+C content of 34.9%, significantly lower than that of the entire chromosome (38.2%) or its ORFs, (39.5%). Note that the possible occurrence of introns in the 5' untranslated region of the pre-mRNA molecules has not been examined systematically for lack of discriminative criteria among the numerous occurrences of the intron consensus 5'-GYMHGH-N1-TACTAAC-N2-YAG-3' in the sequence (294 occurrences if N1 and N2 are set shorter than or equal to 400 and 50 nucleotides, respectively, or more than 600 for limits of 1,000 and 180 nucleotides).

The 20 tRNA genes recognized correspond to 12 different amino acids and 22 different codons. Six tRNA genes contain introns. All tRNA genes are significantly richer in G+C content (47–63%) than the average yeast genome sequence. The frequent duplication of tRNA genes in yeast is

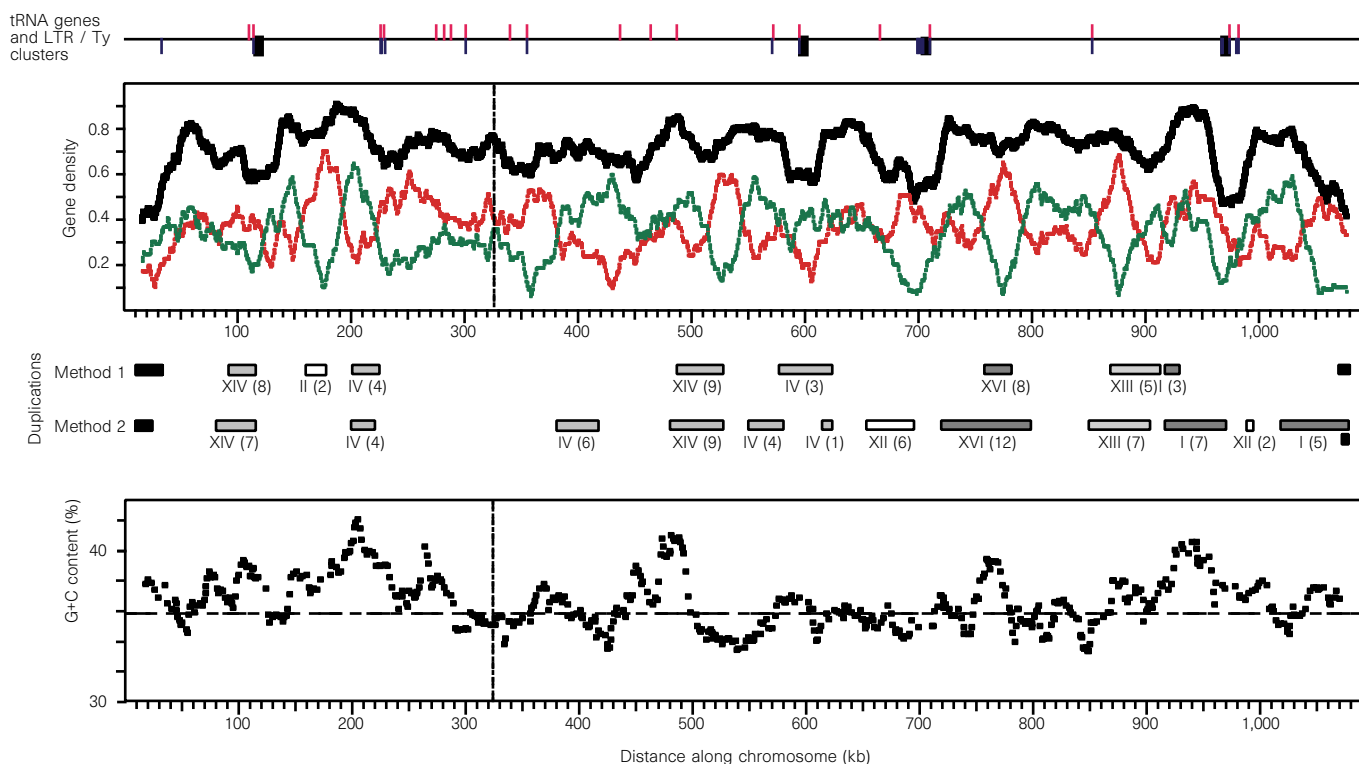


Figure 1 Variation in gene density (top) and base composition (bottom) along the sequence of chromosome XV (scale in kilobases from left telomere). Vertical broken lines indicate position of the centromere. Gene density is expressed as the probability that each nucleotide is part of an ORF, calculated using sliding windows of 30 kb (steps of 0.5 kb) for the Watson strand alone (red line), the Crick strand alone (green line), and their sum (black line). The Watson strand is oriented 5' to 3' from left to right on the chromosome map⁴. G+C content was calculated from silent codon positions using a sliding window of 13 consecutive ORFs (horizontal broken line indicates

average G+C content at these positions; 35.7%). Top line, positions of tRNA genes (thin bars above line), solo LTR (thin bars below line), and complete Ty elements (thick bars below line). Middle, positions of major 'ancient' chromosomal duplications. Blocks represent extent of clusters (method 1), or arrays of ORF pairs (method 2), as defined in text, with indication of the matching chromosome and the number of ORFs involved (in parentheses). Note that several blocks corresponding to a given chromosome are often intermingled with those corresponding to another. Blocks in subtelomeric position (filled) match with several distinct chromosomes.

noticeable on chromosome XV alone, with four gly-tRNA genes (*tG(GGY)OL1* and *tG(GGY)OL2* are identical in sequences, and *tG(GGG)ORI* is similar to the previous two), three pro-tRNA genes (identical in sequences except for their introns), two thr-tRNA genes, and two asn-tRNA genes, respectively identical in sequences; there are also two met-tRNA genes, but they differ in sequence. Duplicated tRNA genes are always found within different sequence environments (including different associated long terminal repeats (LTRs); see below). The tRNA genes are distributed throughout the chromosome (Fig. 1), as is generally the case for yeast which, unlike other organisms, does not show large clusters of tRNA genes in its genome.

Seven genes encoding small known RNA molecules were recognized from the sequence. One of these, *snR17A*, which encodes the U3 snRNA, contains an intron and is duplicated elsewhere in the yeast genome⁸. The downstream part of the gene encoding *snR35* partly overlaps the downstream part of an ORF of unknown function, *YOR222w*, the product of which shares similarity with ADP-ATP carrier proteins.

Four complete Ty elements (two Ty1 and two Ty2), 20 solo LTRs or remnants of them (12 delta, 6 sigma and 2 tau elements), and one Y' element were found. Solo LTRs, or complete Ty elements, are almost always located immediately upstream of tRNA genes (Fig. 1). Only one solo LTR element and one Ty1 stand alone. Consistent with general trends⁹, the two Ty2 elements are found at 'old' sites occupied by several solo LTRs associated with tRNA genes; the same is true for only one of the two Ty1 elements. Conversely, only 11 of the 20 tRNA genes have one or more LTR element upstream of their 5' end, the closest element always being within 200 base pairs of the tRNA gene (sigma elements are only 16–18 nucleotides upstream of tRNA genes).

In total, RNA-coding genes and transposons occupy only 1% of the chromosome XV sequence.

There remain 309,627 base pairs (28.4% of the chromosome) that we will call here 'intergenic regions'. Such regions contain, for a small part, structural chromosomal elements such as the centromere (coordinates 326,592–326,706), the telomeric (C₁₋₃A)_n repeats (coordinates 1–113 and 1,091,264–1,091,283) and two subtelomeric core X elements and their associated repeated elements (STR) (coordinates 114–847 and 1,083,914–1,084,611). But intergenic regions primarily contain promoters, terminators and transcriptional regulatory elements of the protein-coding genes, most of which have not yet been identified. Intergenic regions believed to contain promoter elements based on the orientation of flanking ORFs are noticeably longer (791 nucleotides on average between diverging ORFs) and are richer in G+C (36.2%) than intergenic regions containing putative terminators (421 nucleotides and 28.7% G+C). The presence of flanking RNA-coding genes does not alter this trend. The sequence also reveals 63 ARS consensus elements (5'-WTTTAYRTTTW-3') the activity of which remains to be examined; 39 of these occur in intergenic regions.

Chromosome XV contains few simple sequence iterations. The longest dinucleotide repeat is an alternating poly(AT) stretch of 20-mers (coordinates 45,691–45,730) within the intergenic region that separates the converging ORFs *YOL149w* (*DCP1*) and *YOL148c* (*SPT20*). Only 19 other cases exist of either dinucleotide repeats of at least 10-mers (all are alternating poly(AT) or mononucleotide repeats of at least 20-mers (all poly(A) or poly(T)). Similarly, few trinucleotide repeats are found, the longest being a 20-mer of the triplet CAA occurring within an ORF of unknown function, *YOR267c*, and encoding a poly-glutamine stretch. Six

Table 1 Assembly of the chromosome XV sequence from individual submissions

| Cosmids or DNA | Coordinates on final chromosome | Overlap | Strategy* | Reference |
|-----------------------------|---------------------------------|---------|-----------|-----------------------|
| telomeric plasmid / pEOA363 | 1-32687 | | S, A | |
| pEOA179 / pEOA461 | 24486-97824 | 8202 | N, A, M | 6, 14, 16, 17, 18, 20 |
| pEOA417 | 96924-140942 | 901 | S, M | 26 |
| pEOA228 | 139031-178337 | 1912 | S, A | 28, 29 |
| pEOA1044 | 177014-210234 | 1324 | N, W, A | in the press |
| pUOA1217 | 209185-235991 | 1050 | SS, M | 22 |
| pUOA1344 | 222408-256233 | 13584 | S, W, A | |
| pEOA321 | 253576-287613 | 2658 | N, A | in the press |
| pEOA215 | 286637-323078 | 977 | S, R | |
| pEOA156 | 321732-352202 | 1347 | S, P, A | 30 |
| pEOA303 / pEOA270 | 350740-408356 | 1463 | S, A | |
| pEOA272 | 391560-427841 | 16799 | S, A | |
| pEOA213 / pEOA217 | 415169-477887 | 12673 | SS, M | in the press |
| 11 cosmids† | 476475-606002 | 1413 | S, W, A | 27 and in the press |
| pEOA477 / pUOA1258 | 604545-660867 | 1458 | SS, W, M | 15, 21 |
| pUOA533 / pEOA378 / pEOA241 | 655864-741096 | 5004 | S, R | |
| pEOA423 / pEOA048 | 739215-799188 | 1882 | S, A | 8, 11 |
| pUOA1302 | 795488-832262 | 3701 | SS, W, M | in the press |
| pUOA1337 | 823760-861431 | 8503 | S, A | 19 |
| pEOA487 | 859329-895527 | 2103 | SS, W, M | in the press |
| pEOA284 | 892246-927955 | 3282 | SS, A | 24 |
| pUOA502 / pEOA232 | 927738-957182 | 218 | S, A | 23 |
| pEOA138 | 955761-996055 | 1422 | S, M | 25 |
| 5 cosmids‡ | 992145-1081258 | 3911 | SS, W, M | |
| Right PCR | 1080794-1091283 | 465 | P | |

*S, shotgun of cosmid or part thereof (SS); N, nested deletions; W, walking primer; P, PCR fragments; M, manual gels; A, automatic fluorescent; R, direct membrane blotting

†pEOA347, pUOA522, pEOA246, pEOA264, pEOA273, pEOA306, pEOA265, pEOA106, pEOA338, pEOA986 and pEOA1081.

‡pEOA387, pEOA360, pEOA343, pEOA434 and pEOA390.

other cases of trinucleotide repeats of at least 10-mers are found. Polymorphic variations in trinucleotide repeats have been described in yeast¹⁰. An example of such variations is given by two ORFs of unknown function, YOR229w and YOR230w, that represent an ancient and diverged tandem duplication (67% sequence identity), with an insertion of a long imperfect trinucleotide repeat in YOR229w that is absent from YOR230w¹¹.

Iterations of a few longer sequence motifs are also present. The clearest example is probably the near-perfect repeat of the 39 nucleotide-long unique sequence 5'-GAGCCTGATTCTGTGGCAGAAGATGAACCGGAGACTGAT-3', which occurs nine times between positions 30,935 and 31,309, at the beginning of YOL155c, an ORF of unknown function shows similarity to glucan-1,4- α -glucosidase. The repeats determine a serine-rich amino acid sequence. At the end of the same ORF (coordinates 28,905-29,279), another near perfect repeat of a unique long sequence, 5'-CAGTAGTGTATGWYTTNGGRGAARCASTRGTTKCKG-3', occurs four times. In both cases, degenerated copies of the unique sequence are also found beyond the main repeats.

Of the 561 identified ORFs, 212 (37.8%) correspond to known and functionally characterized genes; all of the others are new. By sequence comparison of their products with general databases, 34 of these (6.1% of the total) show significant homology to proteins of known biochemical and/or physiological function of either yeast or other organisms, and 69 others (12.3% of the total) show weak homology. There are 246 ORFs with products that have either no significant homologue (187 cases or 33.3% of total, among which 28 are questionable) or are homologous to proteins that are themselves of unknown function (59 cases or 10.5% of the total).

Using previously defined criteria¹², 239 ORF products (42.6% of total) are predicted to contain at least one transmembrane span, 170 of which are of unknown function. Two proteins have 12 predicted spans (SCM2, the product of YOL020w, and ALG8, the product of YOR067c), and 43 others have five predicted transmembrane spans or more (only 15 of them are functionally characterized).

Duplications are frequent in the yeast genome, and take several different forms that suggest distinct mechanisms of formation. Comparison of the chromosome XV sequence with the entire yeast genome (including chromosome XV), reveals 12 'clusters' of duplicated sequences that may represent ancestral chromosomal duplications subsequently modified. Such clusters range in size from 12 kb to 49.5 kb, and contain two to nine ORFs, making a total of 297.5 kb of the chromosome XV sequence (27% of total) that is duplicated on at least eight other chromosomes. Duplications with chromosomes IV and XIV are each represented by two clusters intermingled along the chromosome XV map, suggesting successive events of chromosome duplication or rearrangements. Among the four clusters that contain tRNA genes, there are two cases where homologous tRNA genes are conserved at equivalent positions on the other chromosome, further supporting, in such cases, the hypothesis of ancestral chromosomal duplications.

For chromosome XV, duplications were also examined using a second approach based on the systematic comparison of predicted translation products of all yeast ORFs against all others, followed by sequence alignments and estimation of their significance compared to randomizations, and to the overall distribution of similarity values for the entire yeast genome. A total of 193 different ORFs of chromosome XV (34.4%) were found to have at least one significant homologue in the yeast genome (including 28 pairs on the chromosome XV itself). Half of these have several homologues, forming gene families with various degrees of divergence. Among these, 35 ORFs, nearly all in a subtelomeric location, are members of large families with five partners or more (one of them has up to 26 partners). Large gene families include *HXT* genes, *PAU* genes and *RAS* genes, with more than 15 members each in the entire yeast genome. The chromosome XV ORF homologues include 426 different ORFs from the other chromosomes (7.8% of the yeast genome). Distribution of duplicated ORFs along chromosome XV using this method gives roughly the same results as the first method (Fig. 1).

Chromosome XV contains several local ORF duplications in tandem or inverted orientations. Those showing the highest degree of sequence

conservation are the tandem YOR229w and YOR230w already mentioned (63% amino-acid identity), and the inverted repeat YOR10c (*TIR2*) and YOR009w (62% identity). Other local duplications show significantly greater sequence divergence, suggesting more ancient events; they are represented by the tandems YOL083w and YOL082w (35% identity), YOR285w and YOR286w (37% identity), and YOL048c and YOL047c (38% identity, the latter ORF containing an intron). There also exists some 'local' duplications including pairs of ORFs, such as YOR162c and YOR172w (44% identity), or YOR381w and YOR384w (37% identity), that are separated by a few unrelated ORFs. They have also diverged, and may correspond to ancient local duplications that subsequently received intervening DNA.

When this work started, 81 genes or markers were genetically mapped to chromosome XV, and 12 others were assigned to it but unmapped². Seven of the unmapped genes and 55 of the mapped genes could be unambiguously assigned to ORFs or tRNA genes of the present sequence on the basis of previous partial sequence data, use of probes or gene function. Two genes, *ts26* and *PTP1*, originally mapped to chromosome XV, belong to chromosomes XII and IV (YLR268w and YDL230w, respectively). One gene, *TIR2* which corresponds to YOR10c, was originally named *SRP1*, creating confusion with YNL189w on chromosome XIV. Other than that, the original genetic map agrees fairly well with the present data, except for some local inversions of gene order, mostly around the centromere (*TOP1* and *SIN3* are on the left arm, *PEP12* is on the right) and in subtelomeric regions (*MEK1*, *PHR1* and *RAD17*), as is also observed for other chromosomes.

The chromosome XV sequence, like that of other yeast chromosomes, has been interpreted using criteria that are essentially predictive for ORFs, but are comparative with previously described sequences for other genetic features such as RNA-coding genes, Ty elements and the various chromosomal elements. It follows that the number of predicted ORFs is probably overestimated by a few percent compared with the number of actual protein-coding genes, whereas the identification of the other features should be considered as a minimum. Clear-cut identification of the questionable ORFs is not possible without independent experimental evidence, but it is suggested that they represent ~6% of all predicted ORFs. The *a posteriori* comparison of the predicted ORF products with general protein database entries forms the basis for the notion of 'orphan' genes⁷. Orphans are those protein-coding genes, predicted from the genomic sequence, that fail to show significant homology (at the chosen threshold value) when their translation products are compared to gene product sequences translated from all other genomic sequences present in public databases, whether they correspond to *S. cerevisiae* or any other organism. As is the case for other yeast chromosomes, a large fraction (33%) of chromosome XV ORFs are orphans without any significant structural homologue; while another 10.5% are orphans with structural homology to one or several other *S. cerevisiae* orphan(s). It is not surprising that, as with other genes, orphans can be duplicated in the yeast genome, or form diverged gene families. But what is more interesting is the significant deficit of these compared with the overall number of gene families in yeast. Of the 193 chromosome XV ORFs that are duplicated or parts of gene families, only 50 are orphans, compared with 83 predicted (43% of the 561 ORFs). The deficit of orphans among gene families (and the correlated excess of functionally characterized genes) is exactly opposite to the classical expectation from standard genetic screenings based on negative mutant phenotypes, which should tend to ignore isofunctional duplicated genes. One possible explanation for the bias observed may be that molecular methods, in contrast to classical genetic screenings, tend to facilitate the isolation of gene families that are structurally but not functionally related. Systematic functional analysis of the yeast genome, which is expected to follow the completion of the genomic sequence, should help to solve this important question. □

Methods

The sequence of chromosome XV was assembled from 46 cosmids covering the entire length of the chromosome except its left and right telomeres, which

were sequenced from a rescued plasmid and a polymerase chain reaction (PCR) genomic product, respectively (Table 1). The cosmid map will be described separately. Because chromosome XV comigrates in PFGE with chromosome VII (which is only 348 bp shorter), the technique of chromosome fragmentation based on the insertion of unique artificial *I-SceI* sites¹³ played a key role in the physical mapping of these two chromosomes.

Each segment of the chromosome was sequenced on both strands using the methods and strategies indicated in Table 1. Overlaps between sequences submitted by different laboratories range from 218 bp to 16,799 bp (average 3,765 bp). In nearly all cases, overlapping sequences were also entirely determined on both strands by each laboratory and were found to be 100% identical. Only three differences were found in a total of 90,360 bp. After re-examination of the sequences, only one real divergence remained (an A to G transition), probably resulting from a mutation in one of the sequenced cosmids. The present chromosome sequence includes an A at position 400,735 but a G is equally probable. This uncertainty affects YOR036w (*PEP12*), by changing a codon CAG (Gln) to CGG (Arg).

After assembly, the entire sequence was verified as follows. A total of 201 short segments (259 bp–400 bp long) were selected after examination of the sequence using, as criteria, the possible occurrence of frameshifts, compressions (particularly in G+C-rich regions), and the presence of oligomeric stretches of mono- or dinucleotide repeats. Selected segments were attributed anonymously to four different laboratories and resequenced following the protocol of G. V. (unpublished). In total, 64,370 bp were verified revealing 21 original errors (13 nucleotides omitted, 3 nucleotides in excess, and 5 substitutions). Taken together with overlaps between different laboratories, 14.2% of the chromosome (154,730 bp) has thus been sequenced twice independently. The average sequence accuracy is 99.98%. This figure is probably an underestimate, however, as verifications were directed to suspicious regions. Parts of the present sequence were published independently by the sequencers before assembly of the contig and application of final quality controls^{6,8,11,14–30}. Several other manuscripts are also in the press.

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The nucleotide sequence of *Saccharomyces cerevisiae* chromosome XVI

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The nucleotide sequence of the 948,061 base pairs of chromosome XVI has been determined, completing the sequence of the yeast genome. Chromosome XVI was the last yeast chromosome identified¹, and some of the genes mapped early to it, such as *GAL4*, *PEP4* and *RAD1* (ref. 2) have played important roles in the development of yeast biology. The architecture of this final chromo-

some seems to be typical of the large yeast chromosomes, and shows large duplications with other yeast chromosomes. Chromosome XVI contains 487 potential protein-encoding genes, 17 tRNA genes and two small nuclear RNA genes; 27% of the genes have significant similarities to human gene products, and 48% are new and of unknown biological function. Systematic efforts to explore gene function have begun.

There are 487 open reading frames (ORFs) on chromosome XVI, and 10 Ty-related ORFs. Of these ORFs, 17 have an intron, and ORF RPL6B (YPL198w), which encodes a ribosomal protein, has two introns. ORFs were identified using the working definition that they commence with an ATG and have at least a further 99 contiguous sense codons³, and were analysed using established procedures^{4,5}. Before systematic sequencing, there were 73 genes⁶, with 47 genes and their relative positions defining the genetic map, and an additional 26 genes located on the physical map. An additional 92 genes have formal genetic names, some of which had been previously cloned but not mapped to this chromosome or have been studied following their identification by systematic sequencing, for a total of 165 known genes. Thus only 33% of the total ORFs found on chromosome XVI had been identified before the completion of the sequence. Other genetic elements include: 17 tRNAs (9 of which are within 500base pairs of a long terminal repeat (LTR) element of a retrotransposon; 5 Ty retrotransposons; 15 delta elements including partial elements; 4 sigma and 2 tau elements; and 2 snRNAs.

The number of ORFs of known function is 194 (40%), of which 76 are functionally characterized proteins; 88 are known proteins that are not fully characterized; 26 have similarity to proteins of known biochemical and physiological function; 6 are homologous to proteins of known biochemical function; and 55 have a weak homology to known proteins. These weak similarities alone are insufficient to confidently assign function. This leaves 236 ORFs of unknown function (48%), of which 50 have homologues of unknown function, and 186 have no similarity to known proteins. There are 36 questionable ORFs, all of which partly overlap another ORF. All have a low codon adaptation index (CAI) of not greater than 0.18, are short (with an average length of 132 codons), and have no known homology with other proteins or are associated with no known phenotype. For four of these ORFs in two pairs (YPL034c and YPL035c, and YPR038w and YPR039w) it is unclear which, if any, are biologically meaningful. There are few apparent pseudogenes but these include YPL276w and YPL275w, which occur together in the genome as a frameshifted pair, both with homology to a formate dehydrogenase. This arrangement has been confirmed by direct sequencing of genomic DNA. Whether this region represents a mutation specific to strain S288C awaits experimental determination.

Functional categories have been compiled for chromosome XVI ORFs: 214 (43%) are classified in some form and 283 remain unclassified. The chromosome is sufficiently large to have a broad representation of all of the predominant functional groups. Detailed global genome classification and statistics for these functional assignments are tabulated and have been discussed elsewhere^{7,8}.

A robust amino-acid sequence motif is the presence of a predicted transmembrane domain, a region of a protein that spans a lipid bilayer. There are 181 ORFs with one or more predicted transmembrane domains on chromosome XVI (37%)⁹, a number close to that seen on chromosomes II and III (refs 3, 10). Of these, 68 are known functionally in some form, and this percentage of known membrane proteins (38%) is similar to that of all known ORFs. Most of these membrane-protein encoding ORFs contain one or two predicted transmembrane domains (99 and 39, respectively); 15 proteins have 7 or more predicted transmembrane domains, and Ypl006p has 13.

An immediate way to use the sequence information is to examine experimentally ORF function, and the Canadian group is compiling systematic transcript and gene-disruption data. For a section encoding 89 ORFs in the region spanned by YPL085w to YPR017c, 61 transcripts were detected in haploid cells grown on rich medium at 30°C. This level of ORF expression is similar to that seen on chromosomes I and III (refs 5, 11). Of the 117 genes on chromosome XVI that have been disrupted, 36

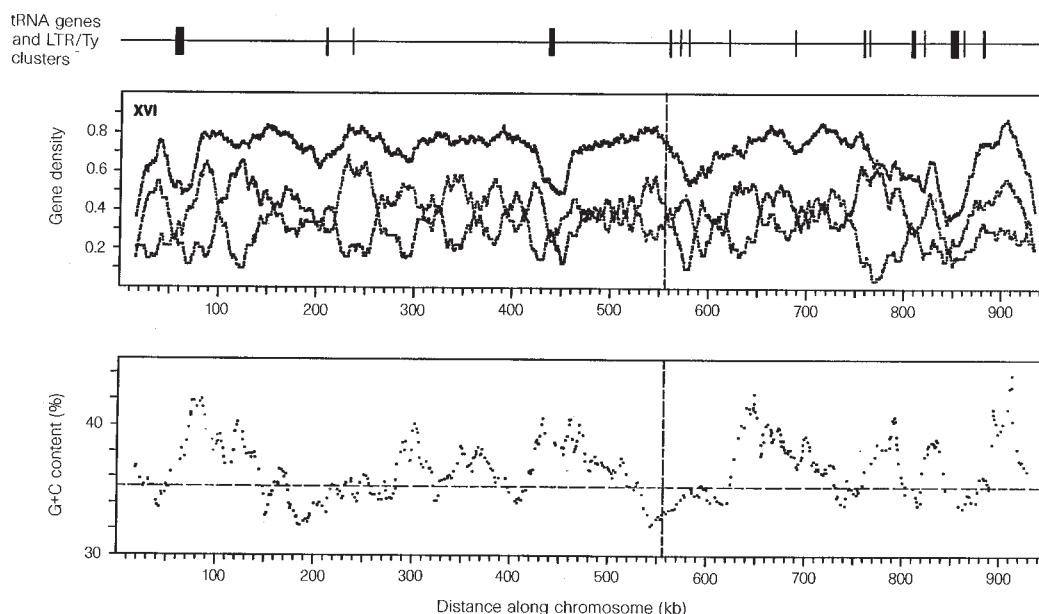


Figure 1 Molecular architecture of chromosome XVI. Top line shows positions of tRNA genes, solo LTR or Ty elements (thin vertical lines) or clusters of them (thick vertical lines) along the chromosome. Panels: variation of gene density (top) and base composition (bottom) along chromosome XVI (scale in kilobases from left telomere). Vertical broken lines

indicate position of the centromere. Gene density determined as for chromosome XV is shown for the Watson (medium line) or the Crick (thin line) strands, and the sum of both (thick line). G+C content was calculated as for chromosome XV (horizontal broken line).

(31%) are 'essential' for vegetative growth. Although this fraction is higher than the approximately 12% estimated for the entire genome¹², or found on chromosome I (ref. 5), it still represents only 7% of the ORFs on the chromosome. The final proportion of essential genes on chromosome XVI awaits a complete disruption set. This information is displayed and will be updated (web sites URL <http://www.mips.biochem.mpg.de> and URL <http://genome-www.stanford.edu>).

Several ORFs deserve mention, some because of size, some because they were anticipated but not previously found, and others because they occur in metazoans with phenotypes or functions that appear characteristic of the larger multicellular eukaryotes, and whose unexpected presence in yeast affords some insight into function. There are 130 ORFs (27%) with significant similarity to products or predicted products of human genes. Four ORF products (*PAL1*, *PHO85*, *ROX1* and *RAD1*) are similar to products of the human genes *ALD*, *RET*, *SRY* and *XPF* (*ERCC4*) that, when altered by mutation, lead to X-linked adrenoleukodystrophy, multiple endocrine neoplasia 2A, gonadal dysgenesis, and xeroderma pigmentosum, respectively^{13,14}. The largest ORF is YPR117w, specifying a 2,489-codon putative membrane protein of unknown function. Just one other ORF, *SEC18* (YPL085w), has more than 2,000 codons, and only 38 ORFs (8%) are larger than 1,000 codons. Identifying very small ORFs presents special problems^{3,15} and just six identified ORFs on chromosome XVI are less than 100 codons. The smallest known functional ORF, *ATP15* (YPL271w), has 62 codons and encodes an F1-ATP synthase epsilon subunit¹⁶. Other small ORFs undoubtedly exist on the chromosome, and remain to be uncovered by functional analysis.

YPL127c encodes an apparent H1 histone, a protein previously not thought to be present in yeast. Plant and animal H1 histones are known to be involved in the higher-order assembly of nucleosomes but, despite considerable work, the precise role of H1 histone remains unclear^{17,18}. Disruption of this single-copy yeast gene indicates that it is not essential for mitotic growth³². A detailed analysis of the possible role of this H1 histone in chromosome assembly, stability and the regulation of gene expression can now be explored.

YPR048w encodes a cytochrome P450 protein with similarity to human nitric oxide synthase, NOS. Human NOS is an amino-acid oxidoreductase that oxidizes the terminal N of arginine to give citrulline and nitric oxide. Nitric oxide (NO) in mammals behaves as a hormone, and is

involved in many processes including vasodilation and neurological activities¹⁹. The involvement of NO in yeast metabolism had not previously been imagined. One possibility is that it acts to regulate cell signalling through interaction with small GTP-binding proteins²⁰.

We found that 74.57% of the chromosomal DNA is involved in the coding of ORFs. These 497 ORFs have an average size of 474 codons, close to the average values seen for other large chromosomes^{3,4,15}. Of the 487 ORFs that are not Ty related, 251 are on the Watson strand and 236 are on the Crick strand, with no apparent strand bias³. There is, on average, an ORF every 1,908 bp over the chromosome. The positioning of ORFs relative to each other seems random: 111 ORFs are divergent, 118 are convergent, and 195 are tandemly arranged. Of the remaining ORFs, 38 are next to a non-ORF element, and in 12 cases two non-ORF elements are adjacent. With regard to intergenic spacing, the average length between tandemly arranged ORFs is 534 bp, for divergent ORFs it is 569 bp, and for convergent ORFs it is 340 bp. Only one region is apparently devoid of ORFs or other genetic entities for 3 kilobases or more: a subtelomeric 3,863 bp gap between YPL275w and YPL274w. There are no large non-coding regions comparable to those found in the subtelomeric regions of chromosomes I and VIII (refs 5,21).

There are no clustered gene families⁵ on chromosome XVI and, as found with other yeast chromosomes, there is little apparent functional clustering of genes. There are two exceptions: a 'syntenic' pair, *CIT3* (YPR001w) and YPR002w, conserving the arrangement of the *mmgD* and *mmgE* genes in *Bacillus subtilis* (GenBank accession no. U29084); and two adjacent cyclin genes, *CLB2* (YPR119w) and *CLB5* (YPR120c), an arrangement that is duplicated on chromosome VII as *CLB1* (YGR108w) and *CLB6* (YGR109c), and which forms part of a larger duplication between these two chromosomes (see below).

The G+C periodicity on chromosome XVI varies about an average content of 38.1% (Fig. 1). Gene density also fluctuates along the chromosome, although it shows little correlation with G+C periodicity. Overall the G+C content of coding regions is 39.5%, and for non-coding regions is 33.29%. The centromere lies in a region of low G+C content, and the Ty and tRNA element clusters lie in regions of low gene density; such long-range compositional variations have been discussed elsewhere²².

The centromere of chromosome XVI spans nucleotide residues 555,952–556,069, between *HAT1* and *CIT3*, making the chromosome

close to being metacentric. Both telomeres seem to be typical in structure. There are 47 potential origins of DNA replication that match the 11-bp ARS element consensus, although the actual autonomously replicating sequences (ARS) used in replicating the chromosome remain to be determined experimentally.

Like much of the yeast genome, regions of chromosome XVI are duplicated. There are some large-scale DNA duplications spanning 25 kb or more^{23,24}. The largest of these is on a 129-kb section of the right arm of chromosome XVI, nucleotide coordinates 731,001–860,000, where regions are duplicated onto a 129.5-kb section on the right arm of chromosome VII (nucleotide coordinates 648,001–777,500). Although removed from the comparison that identified this duplication, the region on chromosome XVI is rich in repetitive elements and contains three Ty elements, five additional LTRs and six tRNA genes. Such DNA duplications form large regions of partial gene synteny between these two chromosomes. An example is a section from nucleotide 834,000 to 860,000 on chromosome XVI and from 762,422 to 777,500 on chromosome VII. In the chromosome XVI interval from YPR154 to YPR159, four of the six genes, two delta elements and two tRNA genes, are syntenic with their chromosome VII counterparts, YGR136 to YGR143, with the exception of a tandem Ty1 element inserted between YPR158 and YPR159 on chromosome XVI. It has been suggested²⁵ that the origin of the *KRE6* (YPR159w)/*SKNI* (YGR143w) pair in this region resulted from some event involving duplication through transposition of the retrotransposon or tRNA elements. The origin of these major cluster homology regions remains unclear but probably consisted of more than one event; the subject is discussed elsewhere at a global genome level⁷. Evidence for duplications can also be found at the ORF level; 125 of the ORFs on chromosome XVI have one or more counterpart with significant similarity in the yeast genome. In addition to duplications, chromosome XVI also contains members of larger gene families, with at least 38 ORFs having similarity to two or more yeast ORFs. An example is *KTR6* (YPL053c), a member of a family of nine mannosyltransferase-encoding genes located on eight different chromosomes²⁶.

Establishment of the genetic and physical maps were critical precursors to obtaining the nucleotide sequence of the chromosomes. The genetic map of chromosome XVI is 251 cM in length, giving an average cM/kb value of 0.26, the smallest seen for a yeast chromosome, and close to that previously reported²⁷. It indicates that a chromosome XVI bivalent has, on average, ten crossovers per meiosis. Although the genetic map is generally correct, there are several discrepancies with the positions found on the sequenced chromosome. Of the 46 genes mapped genetically through the phenotypes of alleles, 14 (*spoT16*, *rad53*, *nib1*, *sot1*, *SUF21*, *dna1*, *mak6*, *tsm0120*, *ymc1*, *spoT20*, *SUP15*, *SUP16*, *cdc67* and *rad56*) remain to be identified on the sequenced chromosome.

Determination of the nucleotide sequence of chromosome XVI completes the sequence of the yeast genome, allowing a genome-wide analysis of a small and experimentally amenable eukaryotic organism. Such systematic studies should enhance our knowledge of cell function, and help us to understand the structure and function of eukaryotes with larger genomes. □

Methods

Chromosome XVI was sequenced using a set of overlapping cosmid and lambda clones based on a previous chromosome XVI physical map²⁸ (L. Riles and M.V. Olson, personal communication) from the S288C-derived strain AB972. Two gaps on the right arm and both telomeres were sequenced using polymerase chain reaction (PCR) products amplified from genomic DNA²⁹. Individual cosmids representing the portion mapped by the EU group were sequenced by contracting laboratories using a variety of subcloning, sequencing and assembly methods^{3, 4}. Explanations of the cloning, sequencing, assembly and quality control methods used by the other groups have been described^{12,13,30}. The telomeres were sequenced using specially devised procedures³¹. Assembly of the completed chromosome sequence was made in Martinsried or Montreal as described^{3-5,15}. Determination of overlapping sequence between groups indicated that seven differences were found and resolved in 81

kb of sequence. This allowed us to estimate the accuracy of the sequence to be conservatively within the three errors per 10 kb average for the yeast genome⁷.

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