

Genetics of Cholesterol Efflux

Iulia Iatan · Aurélien Palmyre · Sarah Alrasheed ·
Isabelle Ruel · Jacques Genest

© Springer Science+Business Media, LLC 2012

Abstract Plasma levels of high-density lipoprotein cholesterol (HDL-C) show an inverse association with coronary heart disease (CHD). As a biological trait, HDL-C is strongly genetically determined, with a heritability index ranging from 40% to 60%. HDL represents an appealing therapeutic target due to its beneficial pleiotropic effects in preventing CHD. This review focuses on the genetic basis of cellular cholesterol efflux, the rate-limiting step in HDL biogenesis. There are several monogenic disorders (e.g., Tangier

disease, caused by mutations within ABCA1) affecting HDL biogenesis. Importantly, many disorders of cellular cholesterol homeostasis cause a reduced HDL-C. We integrate information from family studies and linkage analyses with that derived from genome-wide association studies (GWAS) and review the recent identification of micro-RNAs (miRNA) involved in cellular cholesterol metabolism. The identification of genomic pathways related to HDL may help pave the way for novel therapeutic approaches to promote cellular cholesterol efflux as a therapeutic modality to prevent atherosclerosis.

I. Iatan
Cardiovascular Research Laboratories, Division of Cardiology,
Department of Biochemistry, Faculty of Medicine,
McGill University and McGill University Health Center /
Royal Victoria Hospital,
687 Pine Avenue West, Montreal, QC, H3A 1A1, Canada

A. Palmyre
Magistère Européen de Génétique,
Université Paris Diderot - Paris 7,
5 Thomas Mann street, 775013, Paris, France

S. Alrasheed
Cardiovascular Research Laboratories, Division of Cardiology,
Department of Human Genetics, Faculty of Medicine,
McGill University and McGill University Health Center /
Royal Victoria Hospital,
687 Pine Avenue West, Montreal, QC, H3A 1A1, Canada

I. Ruel
Cardiovascular Research Laboratories, Division of Cardiology,
McGill University Health Center / Royal Victoria Hospital,
687 Pine Avenue West, Montreal, QC, H3A 1A1, Canada

J. Genest (✉)
Faculty of Medicine, Novartis Chair in Medicine,
Cardiovascular Research Laboratories, Division of Cardiology,
McGill University and McGill University Health Center /
Royal Victoria Hospital,
687 Pine Avenue West, Montreal, QC, H3A 1A1, Canada
e-mail: jacques.genest@muhc.mcgill.ca

Keywords HDL · Coronary artery disease · Genetics ·
Cholesterol efflux · ABCA1 · Apo AI · ABCG1 · SR-BI ·
Cholesteryl ester hydrolases · NPC1 · NPC2 · LIPA ·
SMPD1 · StAR D3 · miRNA · Genome wide association
studies

Introduction

The study of genetic defects of high-density lipoprotein (HDL) has increased our knowledge of the mechanisms of HDL biogenesis and metabolism and has provided surprising insights on the contribution of genetic disorders of HDL in the pathogenesis of coronary heart disease (CHD). Indeed, since the identification of apolipoprotein (apo) AI Milano (Apo AI Arg173Cys, rs28931573), the once firmly held notion that genetically determined low HDL cholesterol (HDL-C) levels are associated with CHD has been questioned. The use of Mendelian randomization principles under which the existence of a causal relationship between HDL-C and CHD would imply that association between a gene variant and HDL-C levels will translate into the CHD risk expected from the effect on HDL-C, has also stirred

great controversy [1], despite the coherent association of HDL in heart disease. As such, the study of severe cases of HDL deficiency, such as Tangier disease and apo AI deficiency, has suggested that these forms are associated with premature CHD [2]. In this review, we examine significant genes identified from family studies and linkage analyses that pertain to cellular cholesterol efflux, along with candidate genes involved in cellular cholesterol homeostasis, and integrated information derived from genome-wide association studies (GWAS). Recent data on microRNAs (miRNAs) involved in cholesterol metabolism are also reviewed. The identification of novel genes involved in the complex cellular cholesterol efflux pathways may provide novel therapeutic targets to prevent atherosclerosis in man.

HDL Biogenesis

HDL is associated with protection against cardiovascular disease due to coronary artery disease and other forms of atherosclerosis. Evidence from animal and human studies suggests that HDL, as well as HDL-associated proteins and lipids, possess pleiotropic biological properties that contribute to their antiatherogenic effects. These include anti-inflammatory, antioxidant, and antithrombotic activities, as well as beneficial effects on vascular endothelial function, vascular endothelial cell proliferation, and survival and differentiation of endothelial progenitor cells [3, 4]. The best-recognized property of HDL is reverse cholesterol transport (RCT), the mechanism by which excess cellular cholesterol from peripheral cells, including vessel wall macrophages, is returned to the liver via specific transporters and receptors (ABCA1, ABCG1, ABCG5/8 and SR-BI), for excretion in the bile [3].

Two proteins play a vital role in the process of RCT and in the generation of HDL particles, a process we refer to as HDL biogenesis: apo AI, the major protein within HDL; and ABCA1, a large membrane-associated transporter responsible for cellular cholesterol efflux and apo AI lipidation [5]. This mechanism is remarkably conserved throughout evolution. Cellular cholesterol is toxic to cells and may activate at least two important events: cellular accumulation within the endoplasmic reticulum may trigger the unfolded protein response, leading to alarm, defense, or activation of apoptosis signaling pathways [6], and cholesterol crystals activate the NLRP3 inflammasome by first inducing lysosomal damage; in turn leading to the activation of interleukin-1 and -18 [7, 8]. Thus, the effects of HDL-mediated cholesterol efflux on specific cells such as macrophage point towards a role in innate immunity to protect cells against damage.

In this review, we examined novel and well-known genes and their products involved in cellular cholesterol efflux and provided some insights on the impact of genetic variations within these genes in man. First, key processes involved in cellular cholesterol efflux are summarized (Fig. 1) while presenting an update on candidate genes thus far identified in the cellular cholesterol efflux machinery (Table 1). Then, potential genes identified by GWAS and associated with HDL-C are discussed (Table 2).

Cholesteryl Ester Hydrolases

Arguably, the first step in removing cellular cholesterol involves the hydrolysis of cholesteryl esters (CE) by enzymes that cleave the fatty acyl chain. Neutral cholesterol ester hydrolase (nCEH1, also called KIAA1363) is the prototypical cellular enzyme involved in this process [9]. In macrophages, where most of the cholesterol is stored in lipid droplets as CE, hormone-sensitive lipase (LIPE) is considered to play a major role in the generation of free cholesterol destined for export in mouse, but not in humans. Interestingly, lysosomal cholesteryl ester hydrolases (CEHs) also hydrolyze CE obtained from circulating lipoproteins. At least one other nCEH has recently been identified [10], and the CE hydrolysis system is redundant [11]. This biological redundancy likely may explain why, so far, none of the genes involved in CE hydrolysis, the first step of RCT, have been identified in GWAS (Table 2), or why targeted gene deletion of LIPE does not reduce CE hydrolysis, supporting the concept of multiple redundant systems for cellular CE hydrolysis [12, 13].

ABCA1

The interaction between apo AI and ABCA1 is essential and rate-limiting for the initial step in RCT. Data suggest that ABCA1 acts as a phospholipid translocase and contributes to the formation of a non-raft membrane domain that facilitates the lipidation of apo AI and the formation of nascent HDL particles [14].

The ABCA1 gene was identified over a decade ago as the molecular basis for Tangier disease, a rare disorder of HDL deficiency in which cellular cholesterol efflux is severely reduced [15]. Many mutations in the ABCA1 gene have been identified in Tangier disease and the heterozygous form, familial HDL deficiency [16]. The prevalence of ABCA1 mutations in subjects with HDL deficiency is estimated at approximately 10% to 20% [17, 18]. Insights into the molecular physiology of ABCA1-mediated cellular cholesterol efflux have shown that ABCA1 is predominantly regulated by oxysterols via the

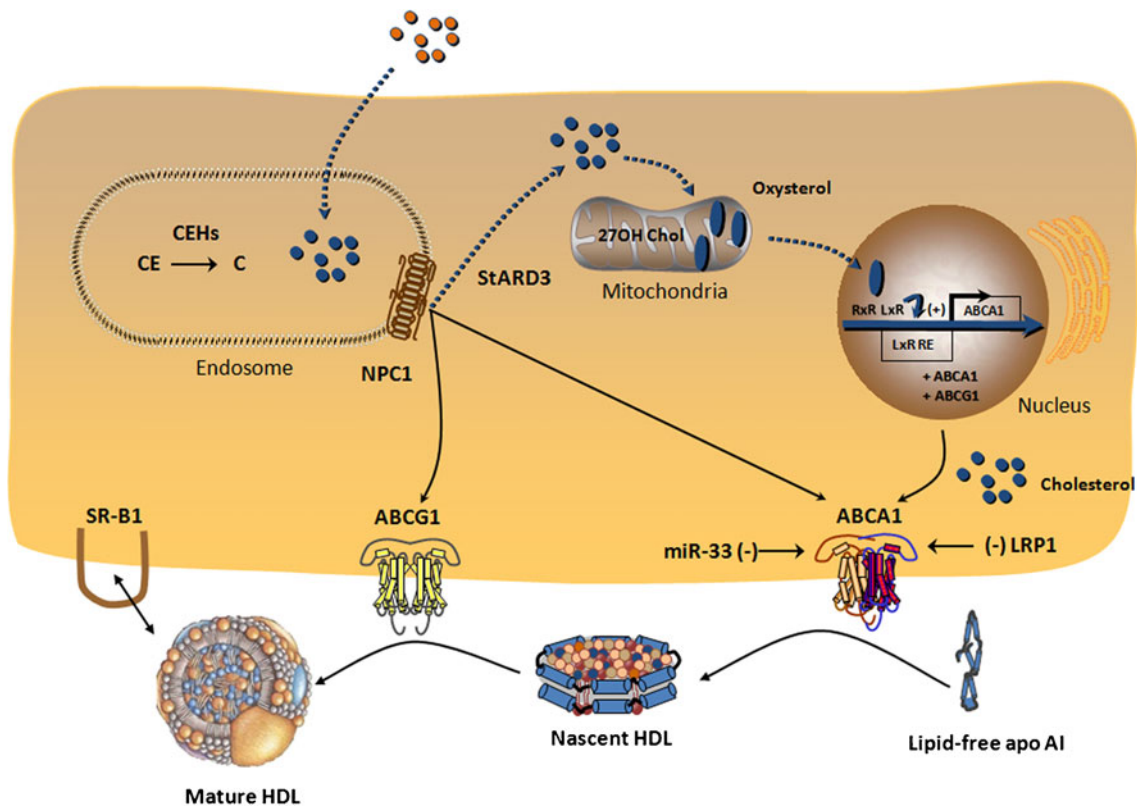


Fig. 1 Major genes involved in genetic disorders of cellular cholesterol efflux in man. To date, genetic disorders can be associated with defects of cholesterol transport in the cytosol in Niemann-Pick disease type C (NPC), defects in cellular cholesterol efflux to apolipoprotein (apo) AI caused by defective ATP-binding cassette (ABC) transporter A1 (Tangier disease), and mutations within apo AI and mutations within SR-BI. ABCG1—ATP-binding cassette transporter G1; C—free cholesterol;

CE—cholesteryl esters; CEHs—cholesteryl esterases (LIPE—hormone sensitive lipase; NCEH1—neutral cholesteryl ester hydrolase-1 also called KIAA1363 and other CE hydrolases); HDL—high-density lipoproteins; LRP1—low density lipoprotein receptor-related protein-1; LXR—liver X receptor; miR—micro RNA; RXR—retinoid X receptor; SR-BI—scavenger receptor B1; StARD3—steroidogenic acute regulator protein

liver specific receptor (LxR) pathway [19] and cyclic adenosine monophosphate (cAMP) in various cell lines [20]. Genetic control of ABCA1 appears to be predominantly

through the oxysterols/LxR pathway (Fig. 1). Attempts to modulate the LxR pathway to increase ABCA1 and HDL have been fraught with the multiple roles of LxR on fatty acid

Table 1 Genes and their proteins associated with cellular cholesterol efflux

Gene Symbol	Protein	GWAS ^a (Table 2)	Disease in humans
ABCA1	ATP binding cassette A1	Yes	Tangier disease
ABCG1	ATP binding cassette G1	No	–
Apo AI	Apolipoprotein AI	Yes	Apo AI deficiency
LIPE	Hormone sensitive lipase	No	–
LRP1	LDL receptor related protein-1	Yes	–
miR-33	Micro RNA -33	No	–
NCEH1	Neutral cholesteryl ester hydrolase	No	–
NPC1	Niemann Pick Type C -1	No	Niemann-Pick disease type C
SMPD1	Sphingomyelinase phosphodiesterase-1	No	Niemann-Pick disease types A & B
SR-BI	Scavenger Receptor BI	Yes	Elevated HDL-C
STARD3	Steroidogenic acute regulatory D3	Yes	–

^a Several of these genes are also found in GWAS associated with HDL-C (see Table 2) GWAS genome-wide association studies; HDL-C high-density lipoprotein cholesterol

Table 2 Results of GWAS showing 47 loci associated with HDL-C levels as a primary trait (with exception of the 9 last genes, where HDL-C was a secondary trait in the analysis)

Locus	Gene	Chromosome	Functions and/or biological processes associated with gene of interest
PABPC4	PolyAdenylate-Binding Protein Cytoplasmic 4	1	Platelet activation
ZNF648	Zinc Finger Protein 648	1	Transcription factor
GALNT2	UDP-N-acetyl-alpha-D-galactosamine: polypeptide Nacetylgalactosaminyltransferase 2 (GalNAc-T2)	1	Involved in O-linked oligosaccharide biosynthesis
IRS1	Insulin Receptor Substrate 1	2	Protein phosphorylated by insulin receptor tyrosine kinase, mutations in genes associated with type 2 diabetes and insulin resistance
COBLL1	COBL-Like protein 1	2	–
SLC39A8	Solute Carrier family 39 (Zinc transporter), MEMBER 8	4	Subfamily of proteins with structural characteristics of zinc transporters
ARL15	ADP-Ribosylation Factor-Like 15	5	–
C6orf106	Chromosome 6 Open Reading Frame 106	6	–
CITED2	CBP/p300-interacting transactivator, with Glu/Asp-rich carboxy-terminal domain 2	6	Inhibits transactivation of HIF1A-induced genes, mutations in genes are cause of cardiac septal defects
LPA	Lipoprotein (a)	6	Serine proteinase that inhibits tissue-type plasminogen activator I, elevated levels are linked to atherosclerosis
KLF14	Kruppel-like factor 14	7	Kruppel-like family of transcription factors
PPP1R3B	Protein Phosphatase 1, Regulatory subunit 3B	8	Regulation of glycogen synthase phosphatase activity; may be involved in type 2 diabetes
TRPS1	TrichoRhinoPhalangeal Syndrome I	8	Transcription factor that represses GATA-regulated genes, mutations in this genes are a cause of trichorhino-phalangeal syndrome types I-III
TTC39B	TetraTriCopeptide repeat domain 39B	9	Knockdown in mouse liver increases plasma HDL-C levels
ABCA1	ATP binding Cassette A1 ^a	9	Cholesterol efflux
AMPD3	Adenosine Monophosphate Deaminase 3	11	Deamination of AMP to IMP in red cells; purine nucleotide cycle
LRP4	Low density lipoprotein Receptor-related Protein 4	11	Regulator of Wnt signaling
PDE3A	Phosphodiesterase 3A	12	Mediates platelet aggregation and plays a role in cardiovascular function by regulating vascular smooth muscle contraction and relaxation
MVK	Mevalonate kinase ^a	12	Isoprenoid and sterol synthesis
SBNO1 1	Strawberry notch homolog 1	12	–
ZNF664	Zinc finger protein 664	12	Transcription factor
SCARB1	Scavenger receptor class B, member 1 ^a	12	Cholesterol uptake and efflux
LIPC	Hepatic lipase ^a	15	Triglyceride hydrolase; ligand/bridging factor for receptor-mediated lipoprotein uptake
LACTB	Lactamase, beta	15	Subunit of the mitochondrial ribosome (39S)
CETP	Cholesteryl Ester Transfer Protein	16	Transfer of cholesteryl ester from HDL to other lipoproteins
LCAT	Lecithin-cholesterol acyltransferase	16	Extracellular cholesterol esterifying enzyme; mutations in gene cause fish-eye disease and LCAT deficiency
CMIP	c-Maf-inducing protein	16	T-cell signaling pathway
STARD3	StAR-related lipid transfer (START) domain containing 3 ^a	17	Cellular cholesterol transport
ABCA8	ATP Binding Cassette A8	17	Member of the ABC1 subfamily (transport various molecules across extra- and intracellular membranes); function of protein has not yet been determined

Table 2 (continued)

Locus	Gene	Chromosome	Functions and/or biological processes associated with gene of interest
PGS1	Phosphatidylglycerophosphate synthase 1	17	–
LIPG	Endothelial lipase	18	Phospholipase
MC4R	Melanocortin 4 receptor	18	Interacts with adrenocorticotrop and MSH hormones
ANGPTL4	Angiopoietin-like 4	19	Regulator of glucose homeostasis, lipid metabolism, and insulin sensitivity
LOC55908	Hepatocellular carcinoma-associated gene TD26	19	–
LILRA3	Leukocyte Ig-like receptor, subfamily A, member 3	19	Soluble immunoreceptor expressed on monocytes and B cells
HNF4A	Hepatocyte nuclear factor 4, alpha	20	Nuclear transcription factor, regulates expression of several hepatic genes
PLTP	Phospholipid transfer protein	20	Transfers phospholipids from triglyceride-rich lipoproteins to HDL
UBE2L3	Ubiquitin-conjugating enzyme E2L 3	22	Ubiquitination of several protein
APOB	Apolipoprotein B	2	Major apolipoprotein of chylomicrons, VLDL and LDL
MLXIPL	MLX interacting protein-like	7	HLH leucine zipper transcription factor of the Myc/Max/Mad superfamily
LPL	Lipoprotein Lipase	8	Triglyceride hydrolase; ligand/bridging factor
TRIB1	Tribbles homolog 1	8	–
FADS1-2-3	Fatty acid desaturase	11	Regulate unsaturation of fatty acids
APOA1	Apolipoprotein AI ^a	11	Major protein of HDL; involved in cholesterol efflux, cofactor for LCAT
UBASH3B	Ubiquitin associated and SH3 domain containing B	11	Inhibit endocytosis of EGFR and PDGF
LRP1	Low density lipoprotein receptor-related protein 1	12	Multiligand cell surface receptor; involved in intracellular signaling, lipid homeostasis, clearance of apoptotic cells
APOE	Apolipoprotein E	19	Chylomicron remnants and VLDL remnants clearance by the liver

^a Gene is related to known efflux defects in humans

HDL-C high-density lipoprotein cholesterol

(Data from Teslovich et al. [27] the National Center for Biotechnology Information [86])

synthesis and regulation of sterol regulatory element-binding proteins (SREBP)-1c.

The regulation of ABCA1 differs between cell types [21]. In macrophages, a novel pathway has been identified. Chen et al. [22] examined the effect of low-density lipoprotein (LDL) loading in macrophages and found increased levels of phosphorylated specificity protein 1 (Sp1) and protein kinase C- ζ (PKC ζ), with increased amounts of Sp1 bound to the ABCA1 promoter. Inhibition of PKC ζ or mutations within the Sp1 binding site on ABCA1 attenuated the LDL-induced expression of ABCA1 and the increase in cellular cholesterol efflux [22].

In a recent study of the lipoprotein receptor related protein-1 (LRP1), a cell-surface receptor of the LDL-receptor superfamily, Basford et al. [23] examined the effects of liver-specific

LRP1 knockout (LRP1^{-/-}) in mice, and found lower levels of HDL-C. Primary hepatocytes from LRP1^{-/-} mice displayed reduced HDL secretion and decreased cell-surface localization of ABCA1 without a change in total cellular ABCA1 content. Their results are consistent with a decreased translocation of ABCA1 to the plasma membrane in LRP1^{-/-} mice hepatocytes (Fig. 1) [23].

Apo AI

Apo AI is the major apolipoprotein within HDL, accounting for approximately 70% of the protein mass. The interaction between Apo AI and ABCA1 is rate-limiting in the first step of the RCT and thus in promoting efflux of cholesterol from

cells (Fig. 1). It has recently been documented that ABCA1 acts by flipping phospholipids to the outer leaflet of the plasma bilayer, contributing to the formation of a 'high-capacity binding site', facilitating binding of Apo AI to this domain, and subsequent formation of nascent HDL particles [14•].

Mutations within Apo AI have been previously identified, with at least 47 variants affecting Apo AI structure [24, 25], some leading to a marked reduction in Apo AI and HDL-C levels [24], and concomitant CHD, whereas others have low HDL but no incidence of heart disease [26]. Apo AI was also previously identified in GWAS (Table 2) [27••], with a significant association for triglyceride (TG), total cholesterol (TC), HDL (as a secondary lipid trait), and LDL lipid levels. Nevertheless, despite the fact that apo AI genetic variations are well characterized, the role of Apo AI structure on ABCA1-mediated efflux has not been completely elucidated.

Considerable controversy in the past few years has emerged regarding the lipid-free and lipid-bound structure of Apo AI [28–30] offering different insights into the structure-function of Apo AI domains critical for nascent HDL particle assembly. As such, Huang et al. [31] have recently addressed Apo AI structure in spherical particles by applying chemical cross-linking and mass spectrometry to HDL particles, describing the first models of authentic human plasma HDL in which Apo AI assumes a cage-like structural framework closely resembling that in synthetic HDL. Specifically, Apo AI adopts intermolecular interactions in plasma HDL similar to interactions previously described in the double-belt and trefoil models derived in reconstituted systems, suggesting that HDL particle sizes are modulated through twisting motion of the resident Apo AI molecules [31].

Other studies have focused at better understanding the properties of the tertiary structure domains of Apo AI and their influence on Apo AI functionality in the RCT pathway [32•]. By generating two chimeric N-terminal domain-swap variants of mouse and human Apo AI, and expressing these hybrids in Apo AI-null mice, Alexander et al. [32•] evaluated their abilities to promote macrophage RCT in vivo, in comparison to wild-type (WT) human and mouse Apo AI. More cholesterol was observed to be significantly removed from the macrophages of the mouse-H Apo AI-expressing mice as compared to the other groups, as shown by a marked increase in fecal excretion via the RCT pathway. Furthermore, catalytic efficiencies (V_{\max}/K_m) of the Apo AI variants were determined and observed to be twofold higher in the mouse-H Apo AI than the WT human Apo AI and 3.5-fold higher than WT mouse Apo AI, suggesting a more efficient ABCA1-mediated cholesterol efflux. These findings, along with a detected increase in the rate of cholesterol uptake into hepatocytes, demonstrated that substitution of

the N-terminal domain of the human Apo AI with the mouse Apo AI counterpart created a “gain of function” Apo AI variant showing enhanced nascent HDL particle formation, a more efficient macrophage RCT, and potentially antiatherogenic apo AI. Additional insights into the effects of Apo AI structure on cholesterol efflux were described by the same group through examination of Apo AI C-terminal α -helix hydrophobicity influence on nascent HDL particle formation [33]. By engineering human Apo AI variants, Lyssenko et al. [33] observed that the decrease in Apo AI hydrophobicity of the non-polar face of the C-terminal amphipathic α -helix significantly reduced the catalytic efficacy of vesicle solubilisation and cholesterol efflux, forming large HDL particles, with reversing effects when hydrophobicity is restored [33].

In addition to structural modifications, several studies have documented that oxidative modification of Apo AI could also be a contributing factor in altering RCT levels [34, 35]. As such, Heinecke and Oram [36] have demonstrated that chlorination, but not nitration, of Apo AI through the myeloperoxidase pathway dramatically impaired the ability of Apo AI to interact directly with ABCA1 and to activate the Janus-kinase-2 signaling pathway, preventing Apo AI from promoting cellular cholesterol efflux in macrophages [36].

ABCG1

The ATP binding cassette transporter G1 (ABCG1) is expressed in a variety of tissues. Although it is predominantly located in the cytosol, ABCG1 promotes the lipidation of nascent HDL particles rather than lipid-free (or lipid-poor) apo AI, playing a significant role in the efflux transport of excess cholesterol to HDL in macrophages, thereby reducing atherosclerosis (Fig. 1). Gao et al. [37] recently identified the first amino acid residue that is critical for ABCG1-mediated cholesterol efflux. Indeed, the cysteine residue located at position 514 is highly conserved and a mutation at this position, although having no effect on the protein stability or trafficking, significantly decreased the efflux of cholesterol onto lipidated apo AI.

Apart from structural changes, ABCG1 expression is mainly regulated by the cellular cholesterol content: it has been shown to be increased following treatment with acetylated LDL (cholesterol-loading condition) and downregulated after treatment with HDL-3 [38]. Nuclear receptors such as the LxR and the peroxisome proliferator-activated receptors (PPARs) have been suggested to regulate the expression of ABCG1. More recently, a significant effort has been put on the study of the various regulators of ABCG1 expression through the LxR pathway, and their effect on the resultant cholesterol efflux. First, Rayner et al. [39••] showed that ABCG1 is targeted by miRNAs (see [miRNA](#) section below),

more specifically miR-33a. Hence, in mouse macrophages, miR-33 decreases ABCG1 expression, reducing cholesterol efflux to nascent HDL. Despite the presence of a miR-33a binding site in the human ABCG1 gene, the targeting of ABCG1 by miR-33a only appears to be marginal [39, 40]. The LxR pathway is also involved in the regulation of ABCG1 expression by group X secretory phospholipase A₂ (GX sPLA₂). In a recent study by Shridas et al. [41], ABCG1 expression and associated cholesterol efflux were found to be reduced after GX sPLA₂ overexpression or exogenous addition in mouse macrophages, suggesting a potential role of GX sPLA₂ in atherosclerotic lipid accumulation.

ABCG1-mediated cholesterol translocation also plays an important role in protection against endothelial dysfunction [42, 43], interaction of monocytes with endothelial cells [44], and regulation of insulin secretion [45, 46]. Increase in ABCG1 expression and concomitant cholesterol efflux capacity, in the presence of erythropoietin [47] or after weight loss [48] for instance, were respectively associated with a reduction in lipid accumulation in foam cells and an improvement in the atherogenic profile in human. As for insulin, it was shown to decrease HDL-mediated cholesterol efflux from macrophages from the suppression of nCEH and ABCG1 expressions [49] and to be involved in the insulin secretion process from pancreatic β cells [45]. These new insights on the regulation of ABCG1 present this transporter as having a novel potential target in the inhibition of the process of atherosclerosis in various human metabolic states, including obesity and diabetes.

Disorders of Cholesterol Trafficking: NPC1, NPC2, LIPA and SMPD1 Genes

Niemann-Pick disease type C (NPC) is a complex lysosomal storage disorder caused by mutations in either the *NPC1* or *NPC2* genes, characterized by the accumulation of unesterified cholesterol in the lysosomal compartment. Choi et al. [50] have reported that the cholesterol trafficking defect in cells from subjects with NPC disease results in a reduced activity of *ABCA1*. Low plasma HDL-C in *NPC1* disease patients occurs independent of their plasma triglyceride levels, suggesting that impaired *ABCA1* regulation as a consequence of reduced efflux of cholesterol out of the late endosome is the cause of the low HDL-C seen in NPC [50, 51]. Reduced plasma HDL-C was the most consistent lipoprotein abnormality found in male and female *NPC1* patients across age groups independent of changes in plasma triglycerides, representing a potential biomarker of *NPC1* disease severity [52]. *NPC1* cells also develop a secondary defect in acid sphingomyelinase (SMase) activity despite a normal acid SMase gene, (*SMPD1*) [53]. Recent advances have provided potential therapeutic approaches for the

treatment of NPC disease. In one study, SMase activity defect in fibroblasts from *NPC1* patients was corrected by *SMPD1* transfection or acid SMase enzyme replacement. Both approaches resulted in a dramatic reduction in lysosomal cholesterol [53]. Recently, the use of small histone deacetylase (HDAC) led to a correction of the NPC phenotype [54]. The HDAC inhibitor LBH589 (panobinostat) is currently undergoing clinical trials in man for several types of cancer. In cultured NPC1 fibroblasts, LBH589 restores cholesterol homeostasis, raising the possibility that this class of agents might be useful in a clinical setting [54].

Another form of *ABCA1* impairment is seen in mutations in the lysosomal acid lipase A (*LIPA*) gene that result in less than 5% of normal lysosomal acid lipase (LAL) activity causing cholesteryl ester storage disease (CESD) [55]. *LIPA* has been identified on chromosome 10q23 as a novel CHD susceptibility locus. Elevated *LIPA* expression itself was related to lower HDL-C levels that are attributed to reduced cholesterol efflux to apo AI. Furthermore, treatment of fibroblasts from normal patients with chloroquine to inhibit LAL activity was observed to reduce *ABCA1* expression and activity, similar to that of CESD cells [55]. LXR agonist treatment of CESD cells corrected *ABCA1* expression, but failed to correct LDL CE hydrolysis and cholesterol efflux to apo AI. Moreover, LDL-induced production of 27-hydroxycholesterol (27-HC) was decreased in CESD compared to normal fibroblasts. It was further determined that treatment of CESD cells with conditioned medium containing LAL from normal fibroblasts or with recombinant human LAL, rescued *ABCA1* expression, apo AI-mediated cholesterol efflux, HDL particle formation, and production of 27-HC [55].

Sphingomyelin (SM) plays an important role in the structural integrity of cellular membranes. Characterized by its high gel to liquid-crystalline phase transition temperature, SM often segregates with cholesterol within distinct sub-cellular compartment. As previously described, one of the genes involved in the modulation of SM levels is sphingomyelin phosphodiesterase 1, also known as acid sphingomyelinase (*SMPD1*, *MIM 607608*). It codes for the lysosomal and secretory SMase, a 70 kDa glycoprotein that hydrolyzes SM to ceramide and phosphorylcholine. Mutations in the *SMPD1* gene cause the autosomal recessive disorder of Niemann-Pick type I disease, which includes type A (*MIM 257200*) and B (*MIM 607616*) (NPA and NPB). Deficiency of lysosomal SMase results in a lysosomal accumulation of SM and a secondary accumulation of cholesterol. We have previously reported that mutations in *SMPD1* causing Niemann-Pick disease types A and B are associated with low HDL-C levels but normal cholesterol efflux under the experimental conditions used [56, 57].

Together, these findings provide further evidence that the rate of release of cholesterol from late endosomes/lysosomes is a critical regulator of cellular cholesterol

trafficking (Fig. 1) and this, in turn, may be critical in cellular cholesterol efflux pathways via the ABCA1 transporter.

SR-BI

The scavenger receptor BI (SR-BI) mediates the selective uptake of cholesteryl esters from HDL into hepatocytes and steroidogenic tissues [58, 59] (Fig. 1). The atheroprotective effects of SR-BI are therefore primarily attributable to its role in cholesterol efflux from lipid-laden macrophages to HDL [60–62] and in the delivery of HDL cholesteryl esters to the liver [63]. Interestingly though, SR-BI^{-/-} mice exhibit higher levels of HDL-C and increased atherosclerosis, a process attributed to a critical block in the RCT to the liver for biliary excretion [64]. In humans, several studies have described genetic *SCARB1* variants associated with HDL-C levels [65, 66], most notably in the recent GWAS by Teslovich et al. [27] (Table 2), where SR-BI was identified among the 95 loci significantly associated with lipid levels in more than 100,000 individuals of European ancestry (rs838880 $P=3 \times 10^{-14}$), confirming its critical role in lipid metabolism. Furthermore, Vergeer et al. [67] recently brought forth evidence that impaired SR-BI function can affect human physiology. Identifying a kindred with a functional missense mutation in SR-BI, P297S, they observed that carriers had significantly elevated HDL-C levels, although no considerable differences in lipid profile parameters such as atherosclerosis or carotid intima-media thickness. Importantly, primary murine hepatocytes expressing mutant SR-BI exhibited a marked reduction of 56% in cholesterol uptake from HDL of that of wild-type SR-BI, whereas P297S variants carriers showed reduced efflux capacity from monocyte-macrophages comparatively to non-carriers [67].

StARD3 (MLN64)

The steroidogenic acute regulatory proteins comprise at least 15 proteins involved in cellular cholesterol transport and homeostasis [68]. StARD3 (also called endosomal Metastatic Lymph Node protein 64) is involved in the key transfer of cholesterol from the late endosomal compartment, a step that occurs after the NPC1 protein. Recent data from Charman et al. [69] shows that StARD3 transports cholesterol from the endosome to the mitochondria and may provide an essential regulatory step between cholesterol, mitochondria-derived oxysterols, and regulation of the cellular cholesterol efflux machinery (Fig. 1). To date, no human disease linked to StARD3 has been identified. Moreover, targeted mutations of the StAR domain in mice do not lead to a defect in steroidogenesis or to a distinct phenotype [70]. However, in

a recent study, Borthwick et al [71] reported that the overexpression of StARD3 in macrophages induces an increase in ABCA1 mRNA and protein, and enhanced cellular cholesterol efflux to Apo AI. Interestingly, overexpression of StARD3 in macrophages also prevents the accumulation of cholesteryl esters in response to acetylated LDL [71]. In GWAS (Table 2), the StARD3 locus is associated with lower HDL-C levels, suggesting an important role for this protein in cellular cholesterol homeostasis and possibly, efflux.

ABCA8

The *ABCA8* gene, located on chromosome 17 (17q24) and coding for another membrane-associated protein member of the ABC superfamily, has also been identified in GWAS (Table 2). The function of this protein in cellular cholesterol efflux is however still unclear. The protein has been isolated in human brain microvessels [72] where it could play a role in intracellular lipid trafficking rather than trans-plasma membrane transport [73]. The gene could be regulated by extracellular signal-regulated kinases (ERK1 and 2) [74].

Novel Genes

The study of extensive kindred with HDL deficiency have identified several chromosomal regions harboring genes related to HDL metabolism [75–77, 78•]. We have recently reported a strong association between the *WWOX* gene locus and HDL cholesterol levels [79] ($P=6.9 \times 10^{-7}$). It remains to be determined if these genes are related to cholesterol homeostasis of cholesterol efflux pathways.

miRNAs

Recently, micro RNAs (miRNAs) as key regulators of lipoprotein metabolism have been uncovered [80, 81]. miRNAs are short (average 22 nucleotides) strands of RNA involved in the repression of gene transcription. Single miRNAs can regulate the transcription of multiple genes by virtue of target sequences usually located within the 3'UTR of target genes. One miRNA, miR-33, located within the *SREBP2* gene, has been shown to repress expression of ABCA1 and to lower HDL-C levels (Fig. 1). Using oligonucleotides directed against miR-33, Rayner et al. [39•, 82] showed that inhibition of miR-33 in the LDL-R^{-/-} mouse is associated with an increase in hepatic ABCA1 and ABCG1 expression, increase in HDL-C, increase in reverse cholesterol transport to the liver and feces, and reduced atherosclerosis. This provides compelling support for a therapeutic application in man. More recently, other miRNAs (miR-106b,

miR758) [81, 83] have been shown to control a variety of aspects of lipid metabolism, from HDL biogenesis to cholesterol efflux. These findings have thus highlighted the significant role that miRNAs play in HDL metabolism, opening new avenues for the treatment of CHD.

Genome-Wide Association Studies

The search for novel genes associated with a measurable trait has been greatly facilitated by GWAS. Using single-nucleotide polymorphisms (SNPs), common genetic variants in different individuals are examined to identify association with a specific, measurable biological trait. GWAS can identify SNPs (and other variations) in DNA, but do not necessarily imply causality between identified genes and the biological trait under study.

GWAS compare the frequency of common DNA variants between people with the trait or the disease (cases) and similar people without (controls). DNA is extracted from various cells in individual participants. High-throughput gene chips can now access over 2,000,000 SNPs, spanning the entire genome. The prevalence of genetic variants at each SNP is then examined between cases and controls and bioinformatics is then used to determine the statistical strength and significance of the observation. It should be emphasized that most of the SNP variations associated with a trait or disease do not always lay in the region of DNA that codes for proteins. They are usually in the large non-coding regions (introns or intergenic regions); such SNPs are probably not causal but may be in linkage disequilibrium with yet unknown genetic variants that have functional significance.

In contrast to careful analyses of individual SNPs, GWAS are hypothesis-free and, theoretically at least, bias-free. It should be noted that the sheer number of statistical tests performed presents an unprecedented potential for false-positive results. Despite these caveats, GWAS in the field of HDL genetics has reassuringly identified many of the same genes (or genomic regions) that harbor genes causing Mendelian disorders in man [78•] (Table 2). Thus, GWAS not only have the potential of finding common variants explaining a relatively small percentage of variation of a biological trait, but also to identify novel genes associated with disease in man.

The investigation of severe traits of HDL will be facilitated by two other techniques that allow even faster progress in the field of human genetics. “Deep” resequencing is a relatively novel technology that allows the sequencing of genomic DNA over a large chromosomal region (usually in millions of base pairs). Reference genome sequences for humans are available [84]. Identifying sequence variations and understanding their biological significance is therefore becoming a major focus of genetics. The second technique is exome-wide sequencing, where every exon (approximately 180,000) is sequenced by

several high-throughput technologies. Using reference genome sequences and powerful bioinformatics packages, one can identify genetic variants causing disease in humans [85•].

Conclusion

Evidence shows that cholesterol efflux and RCT play a major role in preventing atherosclerosis in humans. Multiple studies have also reported genetic data to explain variations in HDL-C levels associated with cellular cholesterol homeostasis. These studies are based on different approaches, including heritability of HDL-C levels in families, candidate genes involved in HDL metabolism, linkage analyses from genome-wide scans, and GWAS, among others. Furthermore, because of their importance in lipoprotein metabolism and vascular endothelial function, the identification of novel genes involved in cholesterol efflux pathways, is warranted. As such, novel genomic pathways related to HDL metabolism may provide novel therapeutic avenues to prevent CHD.

Disclosure No conflicts of interest relevant to this article were reported.

References

Papers of particular interest, published recently, have been highlighted as:

- Of importance
- Of major importance

1. Frikke-Schmidt R, Nordestgaard BG, Stene MC, Sethi AA, Remaley AT, Schnohr P, Grande P, Tybjaerg-Hansen A. Association of loss-of-function mutations in the ABCA1 gene with high-density lipoprotein cholesterol levels and risk of ischemic heart disease. *JAMA*. 2008;299:2524–32.
2. Asztalos BF, Tani M, Schaefer EJ. Metabolic and functional relevance of HDL subspecies. *Curr Opin Lipidol*. 2011;22:176–85.
3. Assmann G, Nofer JR. Atheroprotective effects of high-density lipoproteins. *Annu Rev Med*. 2003;54:321–41.
4. Iatan I, Alrasadi K, Ruel I, Alwaili K, Genest J. Effect of ABCA1 mutations on risk for myocardial infarction. *Curr Atheroscler Rep*. 2008;10:413–26.
5. Zannis VI, Chroni A, Krieger M. Role of apoA-I, ABCA1, LCAT, and SR-BI in the biogenesis of HDL. *J Mol Med (Berl)*. 2006;84:276–94.
6. Devries-Seimon T, Li Y, Yao PM, Stone E, Wang Y, Davis RJ, Flavell R, Tabas I. Cholesterol-induced macrophage apoptosis requires ER stress pathways and engagement of the type A scavenger receptor. *J Cell Biol*. 2005;171:61–73.
7. Duewell P, Kono H, Rayner KJ, Sirois CM, Vladimer G, Bauernfeind FG, Abela GS, Franchi L, Nunez G, Schnurr M, Espevik T, Lien E, Fitzgerald KA, Rock KL, Moore KJ, Wright SD, Homung V, Latz E. NLRP3 inflammasomes are required for atherogenesis and activated by cholesterol crystals. *Nature*. 2010;464:1357–61.

8. Rajamaki K, Lappalainen J, Oorni K, Valimaki E, Matikainen S, Kovanen PT, Eklund KK. Cholesterol crystals activate the NLRP3 inflammasome in human macrophages: a novel link between cholesterol metabolism and inflammation. *PLoS One*. 2010;5:e11765.
9. Okazaki H, Igarashi M, Nishi M, Sekiya M, Tajima M, Takase S, Takanashi M, Ohta K, Tamura Y, Okazaki S, Yahagi N, Ohashi K, Amemiya-Kudo M, Nakagawa Y, Nagai R, Kadowaki T, Osuga J, Ishibashi S. Identification of neutral cholesterol ester hydrolase, a key enzyme removing cholesterol from macrophages. *J Biol Chem*. 2008;283:33357–64.
10. Sekiya M, Osuga J, Igarashi M, Okazaki H, Ishibashi S. The role of neutral cholesterol ester hydrolysis in macrophage foam cells. *J Atheroscler Thromb*. 2011;18:359–64.
11. Quiroga AD, Lehner R. Role of endoplasmic reticulum neutral lipid hydrolases. *Trends Endocrinol Metab*. 2011;22:218–25.
12. Buchebner M, Pfeifer T, Rathke N, Chandak PG, Lass A, Schreiber R, Kratzer A, Zimmermann R, Sattler W, Koefeler H, Frohlich E, Kostner GM, Birner-Gruenberger R, Chiang KP, Haemmerle G, Zechner R, Levak-Frank S, Cravatt B, Kratky D. Cholesteryl ester hydrolase activity is abolished in HSL^{-/-} macrophages but unchanged in macrophages lacking KIAA1363. *J Lipid Res*. 2010;51:2896–908.
13. Kratky D. Neutral cholesterol ester hydrolases in macrophages: still a matter of debate. *Circ Res*. 2011;108:e13.
14. • Iatan I, Bailey D, Ruel I, Hafiane A, Campbell S, Krimbou L, Genest J: Membrane Microdomains Modulate Ligand Binding Activity of Oligomeric ABCA1 and ApoA-I-Mediated Lipid Removal: Molecular Evidence that ApoA-I Interaction with ABCA1 Activates the Phosphatidylcholine Biosynthesis Pathway. *J Lipid Res*. 2011. *This study examines the nature of ABCA1 as a phospholipid translocase, which in conjunction with the high capacity binding site contributes to the formation of a non-raft membrane domain that facilitates the lipidation of apo AI and the formation of nascent HDL particles.*
15. Brooks-Wilson A, Marcil M, Clee SM, Zhang LH, Roomp K, Van DM, Yu L, Brewer C, Collins JA, Molhuizen HO, Loubser O, Ouelette BF, Fichter K, Ashbourne-Excoffon KJ, Sensen CW, Scherer S, Mott S, Denis M, Martindale D, Frohlich J, Morgan K, Koop B, Pimstone S, Kastelein JJ, Genest Jr J, Hayden MR. Mutations in ABC1 in Tangier disease and familial high-density lipoprotein deficiency. *Nat Genet*. 1999;22:336–45.
16. D.N.Cooper EVBPDSADPKSMEM. The Human Gene Mutation Database at the Institute of Medical Genetics in Cardiff. 2011. Biobase Biological Database.
17. Candini C, Schimmel AW, Peter J, Bochem AE, Holleboom AG, Vergeer M, Dullaart RP, Dallinga-Thie GM, Hovingh GK, Khoo KL, Fasano T, Bocchi L, Calandra S, Kuivenhoven JA, Motazacker MM. Identification and characterization of novel loss of function mutations in ATP-binding cassette transporter A1 in patients with low plasma high-density lipoprotein cholesterol. *Atherosclerosis*. 2010;213:492–8.
18. Alrasadi K, Ruel IL, Marcil M, Genest J. Functional mutations of the ABCA1 gene in subjects of French-Canadian descent with HDL deficiency. *Atherosclerosis*. 2006;188:281–91.
19. Oram JF, Heinecke JW. ATP-binding cassette transporter A1: a cell cholesterol exporter that protects against cardiovascular disease. *Physiol Rev*. 2005;85:1343–72.
20. Haidar B, Denis M, Krimbou L, Marcil M, Genest Jr J. cAMP induces ABCA1 phosphorylation activity and promotes cholesterol efflux from fibroblasts. *J Lipid Res*. 2002;43:2087–94.
21. Denis M, Bissonnette R, Haidar B, Krimbou L, Bouvier M, Genest J. Expression, regulation, and activity of ABCA1 in human cell lines. *Mol Genet Metab*. 2003;78:265–74.
22. Chen X, Zhao Y, Guo Z, Zhou L, Okoro EU, Yang H. Transcriptional regulation of ATP-binding cassette transporter A1 expression by a novel signaling pathway. *J Biol Chem*. 2011;286:8917–23.
23. Basford JE, Wancata L, Hofmann SM, Silva RA, Davidson WS, Howles PN, Hui DY. Hepatic deficiency of low density lipoprotein receptor-related protein-1 reduces high density lipoprotein secretion and plasma levels in mice. *J Biol Chem*. 2011;286:13079–87.
24. Sorci-Thomas MG, Thomas MJ. The effects of altered apolipoprotein A-I structure on plasma HDL concentration. *Trends Cardiovasc Med*. 2002;12:121–8.
25. Dastani Z, Dangoisse C, Boucher B, Desbiens K, Krimbou L, Dufour R, Hegele RA, Pajukanta P, Engert JC, Genest J, Marcil M. A novel nonsense apolipoprotein A-I mutation (apoA-I(E136X)) causes low HDL cholesterol in French Canadians. *Atherosclerosis*. 2006;185:127–36.
26. Sirtori CR, Calabresi L, Franceschini G, Baldassarre D, Amato M, Johansson J, Salvetti M, Monteduro C, Zulli R, Muiesan ML, Agabiti-Rosei E. Cardiovascular status of carriers of the apolipoprotein A-I(Milano) mutant: the Limone sul Garda study. *Circulation*. 2001;103:1949–54.
27. •• Teslovich TM, Musunuru K, Smith AV et al. Biological, clinical and population relevance of 95 loci for blood lipids. *Nature* 2010, 466:707–713. *This is a significant genome-wide association study identifying 95 lipid-associated loci in 100,000 individuals of European ancestry, 59 of which showed, for the first time, a genome-wide significant association with lipid traits.*
28. Davidson WS, Silva RA. Apolipoprotein structural organization in high density lipoproteins: belts, bundles, hinges and hairpins. *Curr Opin Lipidol*. 2005;16:295–300.
29. Thomas MJ, Bhat S, Sorci-Thomas MG. Three-dimensional models of HDL apoA-I: implications for its assembly and function. *J Lipid Res*. 2008;49:1875–83.
30. Wu Z, Gogonea V, Lee X, Wagner MA, Li XM, Huang Y, Undurti A, May RP, Haertlein M, Moulin M, Gutsche I, Zaccai G, Didonato JA, Hazen SL. Double superhelix model of high density lipoprotein. *J Biol Chem*. 2009;284:36605–19.
31. Huang R, Silva RA, Jerome WG, Kontush A, Chapman MJ, Curtiss LK, Hodges TJ, Davidson WS. Apolipoprotein A-I structural organization in high-density lipoproteins isolated from human plasma. *Nat Struct Mol Biol*. 2011;18:416–22.
32. • Alexander ET, Vedhachalam C, Sankaranarayanan S, Llera-Moya M, Rothblat GH, Rader DJ, Phillips MC: Influence of apolipoprotein A-I domain structure on macrophage reverse cholesterol transport in mice. *Arterioscler.Thromb.Vasc.Biol*. 2011, 31:320–327. *This study examines the properties of the tertiary structure domains of apo AI and its influence on apo AI functionality in the reverse cholesterol pathway.*
33. Lyssenko NN, Hata M, Dhanasekaran P, Nickel M, Nguyen D, Chetty PS, Saito H, Lund-Katz S, Phillips MC: Influence of C-terminal alpha-helix hydrophobicity and aromatic amino acid content on apolipoprotein A-I functionality. *Biochim Biophys Acta* 2011.
34. Bergt C, Pennathur S, Fu X, Byun J, O'Brien K, McDonald TO, Singh P, Anantharamaiah GM, Chait A, Brunzell J, Geary RL, Oram JF, Heinecke JW. The myeloperoxidase product hypochlorous acid oxidizes HDL in the human artery wall and impairs ABCA1-dependent cholesterol transport. *Proc Natl Acad Sci U S A*. 2004;101:13032–7.
35. Shao B, Oda MN, Bergt C, Fu X, Green PS, Brot N, Oram JF, Heinecke JW. Myeloperoxidase impairs ABCA1-dependent cholesterol efflux through methionine oxidation and site-specific tyrosine chlorination of apolipoprotein A-I. *J Biol Chem*. 2006;281:9001–4.
36. Shao B, Tang C, Heinecke JW, Oram JF. Oxidation of apolipoprotein A-I by myeloperoxidase impairs the initial interactions with ABCA1 required for signaling and cholesterol export. *J Lipid Res*. 2010;51:1849–58.
37. Gao X, Gu H, Li G, Rye KA, Zhang DW: Identification of an amino acid residue in ATP-binding cassette transport G1

- critical for mediating cholesterol efflux. *Biochim Biophys Acta* 2011.
38. Klucken J, Buchler C, Orso E, Kaminski WE, Porsch-Ozcurumez M, Liebisch G, Kapinsky M, Diederich W, Drobnik W, Dean M, Allikmets R, Schmitz G. ABCG1 (ABC8), the human homolog of the *Drosophila* white gene, is a regulator of macrophage cholesterol and phospholipid transport. *Proc Natl Acad Sci U S A*. 2000;97:817–22.
 39. Rayner KJ, Suarez Y, Davalos A, Parathath S, Fitzgerald ML, Tamehiro N, Fisher EA, Moore KJ, Fernandez-Hernando C: MiR-33 contributes to the regulation of cholesterol homeostasis. *Science* 2010, 328:1570–1573. *This article describes the role of miR-33 as it appears in both HDL biogenesis and cellular cholesterol efflux.*
 40. Marquart TJ, Allen RM, Ory DS, Baldan A. miR-33 links SREBP-2 induction to repression of sterol transporters. *Proc Natl Acad Sci U S A*. 2010;107:12228–32.
 41. Shridas P, Bailey WM, Gizard F, Oslund RC, Gelb MH, Bruemmer D, Webb NR. Group X secretory phospholipase A2 negatively regulates ABCA1 and ABCG1 expression and cholesterol efflux in macrophages. *Arterioscler Thromb Vasc Biol*. 2010;30:2014–21.
 42. Terasaka N, Yu S, Yvan-Charvet L, Wang N, Mzhavia N, Langlois R, Pagler T, Li R, Welch CL, Goldberg IJ, Tall AR. ABCG1 and HDL protect against endothelial dysfunction in mice fed a high-cholesterol diet. *J Clin Invest*. 2008;118:3701–13.
 43. Terasaka N, Westerterp M, Koetsveld J, Fernandez-Hernando C, Yvan-Charvet L, Wang N, Sessa WC, Tall AR. ATP-binding cassette transporter G1 and high-density lipoprotein promote endothelial NO synthesis through a decrease in the interaction of caveolin-1 and endothelial NO synthase. *Arterioscler Thromb Vasc Biol*. 2010;30:2219–25.
 44. Whetzel AM, Sturek JM, Nagelin MH, Bolick DT, Gebre AK, Parks JS, Bruce AC, Skaflen MD, Hedrick CC. ABCG1 deficiency in mice promotes endothelial activation and monocyte-endothelial interactions. *Arterioscler Thromb Vasc Biol*. 2010;30:809–17.
 45. Sturek JM, Castle JD, Trace AP, Page LC, Castle AM, Evans-Molina C, Parks JS, Mirmira RG, Hedrick CC. An intracellular role for ABCG1-mediated cholesterol transport in the regulated secretory pathway of mouse pancreatic beta cells. *J Clin Invest*. 2010;120:2575–89.
 46. Fryirs MA, Barter PJ, Appavoo M, Tuch BE, Tabet F, Heather AK, Rye KA. Effects of high-density lipoproteins on pancreatic beta-cell insulin secretion. *Arterioscler Thromb Vasc Biol*. 2010;30:1642–8.
 47. Lu KY, Ching LC, Su KH, Yu YB, Kou YR, Hsiao SH, Huang YC, Chen CY, Cheng LC, Pan CC, Lee TS. Erythropoietin suppresses the formation of macrophage foam cells: role of liver X receptor alpha. *Circulation*. 2010;121:1828–37.
 48. Aron-Wisniewsky J, Julia Z, Poitou C, Bouillot JL, Basdevant A, Chapman MJ, Clement K, Guerin M. Effect of bariatric surgery-induced weight loss on SR-BI-, ABCG1-, and ABCA1-mediated cellular cholesterol efflux in obese women. *J Clin Endocrinol Metab*. 2011;96:1151–9.
 49. Yamashita M, Tamasawa N, Matsuki K, Tanabe J, Murakami H, Matsui J, Suda T. Insulin suppresses HDL-mediated cholesterol efflux from macrophages through inhibition of neutral cholesteryl ester hydrolase and ATP-binding cassette transporter G1 expressions. *J Atheroscler Thromb*. 2010;17:1183–9.
 50. Choi HY, Karten B, Chan T, Vance JE, Greer WL, Heidenreich RA, Garver WS, Francis GA. Impaired ABCA1-dependent lipid efflux and hypoalphalipoproteinemia in human Niemann-Pick type C disease. *J Biol Chem*. 2003;278:32569–77.
 51. Tangemo C, Weber D, Theiss S, Mengel E, Runz H. Niemann-Pick Type C disease: characterizing lipid levels in patients with variant lysosomal cholesterol storage. *J Lipid Res*. 2011;52:813–25.
 52. Garver WS, Jelinek D, Meaney FJ, Flynn J, Pettit KM, Shepherd G, Heidenreich RA, Vockley CM, Castro G, Francis GA. The National Niemann-Pick Type C1 Disease Database: correlation of lipid profiles, mutations, and biochemical phenotypes. *J Lipid Res*. 2010;51:406–15.
 53. Devlin C, Pipalia NH, Liao X, Schuchman EH, Maxfield FR, Tabas I. Improvement in lipid and protein trafficking in Niemann-Pick C1 cells by correction of a secondary enzyme defect. *Traffic*. 2010;11:601–15.
 54. Pipalia NH, Cosner CC, Huang A, Chatterjee A, Bourbon P, Farley N, Helquist P, Wiest O, Maxfield FR. Histone deacetylase inhibitor treatment dramatically reduces cholesterol accumulation in Niemann-Pick type C1 mutant human fibroblasts. *Proc Natl Acad Sci U S A*. 2011;108:5620–5.
 55. Bowden KL, Bilbey NJ, Bilawchuk LM, Boadu E, Sidhu R, Ory DS, Du H, Chan T, Francis GA. Lysosomal acid lipase deficiency impairs regulation of ABCA1 gene and formation of high density lipoproteins in cholesteryl ester storage disease. *J Biol Chem*. 2011;286:30624–35.
 56. Lee CY, Krimbou L, Vincent J, Bernard C, Larramee P, Genest Jr J, Marcil M. Compound heterozygosity at the sphingomyelin phosphodiesterase-1 (SMPD1) gene is associated with low HDL cholesterol. *Hum Genet*. 2003;112:552–62.
 57. Lee CY, Lesimple A, Denis M, Vincent J, Larsen A, Mamer O, Krimbou L, Genest J, Marcil M. Increased sphingomyelin content impairs HDL biogenesis and maturation in human Niemann-Pick disease type B. *J Lipid Res*. 2006;47:622–32.
 58. Trigatti B, Rigotti A, Krieger M. The role of the high-density lipoprotein receptor SR-BI in cholesterol metabolism. *Curr Opin Lipidol*. 2000;11:123–31.
 59. Krieger M. Charting the fate of the ‘good cholesterol’: identification and characterization of the high-density lipoprotein receptor SR-BI. *Annu Rev Biochem*. 1999;68:523–58.
 60. Covey SD, Krieger M, Wang W, Penman M, Trigatti BL. Scavenger receptor class B type I-mediated protection against atherosclerosis in LDL receptor-negative mice involves its expression in bone marrow-derived cells. *Arterioscler Thromb Vasc Biol*. 2003;23:1589–94.
 61. Ji Y, Jian B, Wang N, Sun Y, Moya ML, Phillips MC, Rothblat GH, Swaney JB, Tall AR. Scavenger receptor BI promotes high density lipoprotein-mediated cellular cholesterol efflux. *J Biol Chem*. 1997;272:20982–5.
 62. Van EM, Bos IS, Hildebrand RB, Van Rij BT, Van Berkel TJ. Dual role for scavenger receptor class B, type I on bone marrow-derived cells in atherosclerotic lesion development. *Am J Pathol*. 2004;165:785–94.
 63. Zhang Y, Da Silva JR, Reilly M, Billheimer JT, Rothblat GH, Rader DJ. Hepatic expression of scavenger receptor class B type I (SR-BI) is a positive regulator of macrophage reverse cholesterol transport in vivo. *J Clin Invest*. 2005;115:2870–4.
 64. Leiva A, Verdejo H, Benitez ML, Martinez A, Busso D, Rigotti A. Mechanisms regulating hepatic SR-BI expression and their impact on HDL metabolism. *Atherosclerosis*. 2011;217:299–307.
 65. Acton S, Osgood D, Donoghue M, Corella D, Pocovi M, Cenarro A, Mozas P, Keilty J, Squazzo S, Woolf EA, Ordovas JM. Association of polymorphisms at the SR-BI gene locus with plasma lipid levels and body mass index in a white population. *Arterioscler Thromb Vasc Biol*. 1999;19:1734–43.
 66. Hsu LA, Ko YL, Wu S, Teng MS, Peng TY, Chen CF, Chen CF, Lee YS. Association between a novel 11-base pair deletion mutation in the promoter region of the scavenger receptor class B type I gene and plasma HDL cholesterol levels in Taiwanese Chinese. *Arterioscler Thromb Vasc Biol*. 2003;23:1869–74.
 67. Vergeer M, Korpelaar SJ, Franssen R, Meurs I, Out R, Hovingh GK, Hoekstra M, Sierts JA, Dallinga-Thie GM, Motazacker MM, Holleboom AG, Van Berkel TJ, Kastelein JJ, Van EM, Kuivenhoven JA. Genetic variant of the scavenger receptor BI in humans. *N Engl J Med*. 2011;364:136–45.

68. Soccio RE, Breslow JL. StAR-related lipid transfer (START) proteins: mediators of intracellular lipid metabolism. *J Biol Chem*. 2003;278:22183–6.
69. Charman M, Kennedy BE, Osborne N, Karten B. MLN64 mediates egress of cholesterol from endosomes to mitochondria in the absence of functional Niemann-Pick Type C1 protein. *J Lipid Res*. 2010;51:1023–34.
70. Rigotti A, Cohen DE, Zanlungo S. STARTing to understand MLN64 function in cholesterol transport. *J Lipid Res*. 2010;51:2015–7.
71. Borthwick F, Allen AM, Taylor JM, Graham A. Overexpression of STARD3 in human monocyte/macrophages induces an anti-atherogenic lipid phenotype. *Clin Sci (Lond)*. 2010;119:265–72.
72. Shawahna R, Uchida Y, Declèves X, Ohtsuki S, Yousif S, Dauchy S, Jacob A, Chassoux F, Daumas-Duport C, Couraud PO, Terasaki T, Scherrmann JM. Transcriptomic and quantitative proteomic analysis of transporters and drug metabolizing enzymes in freshly isolated human brain microvessels. *Mol Pharm*. 2011;8:1332–41.
73. Kim WS, Weickert CS, Garner B. Role of ATP-binding cassette transporters in brain lipid transport and neurological disease. *J Neurochem*. 2008;104:1145–66.
74. Shukla A, Hillegass JM, MacPherson MB, Beuschel SL, Vacek PM, Pass HI, Carbone M, Testa JR, Mossman BT. Blocking of ERK1 and ERK2 sensitizes human mesothelioma cells to doxorubicin. *Mol Cancer*. 2010;9:314.
75. Dastani Z, Pajukanta P, Marcil M, Rudzicz N, Ruel I, Bailey SD, Lee JC, Lemire M, Faith J, Platko J, Rioux J, Hudson TJ, Gaudet D, Engert JC, Genest J. Fine mapping and association studies of a high-density lipoprotein cholesterol linkage region on chromosome 16 in French-Canadian subjects. *Eur J Hum Genet*. 2010;18:342–7.
76. Dastani Z, Quioque L, Plaisier C, Engert JC, Marcil M, Genest J, Pajukanta P. Evidence for a gene influencing high-density lipoprotein cholesterol on chromosome 4q31.21. *Arterioscler Thromb Vasc Biol*. 2006;26:392–7.
77. Iatan I, Dastani Z, Do R, Weissglas-Volkov D, Ruel I, Lee JC, Huertas-Vazquez A, Taskinen MR, Prat A, Seidah NG, Pajukanta P, Engert JC, Genest J. Genetic variation at the proprotein convertase subtilisin/kexin type 5 gene modulates high-density lipoprotein cholesterol levels. *Circ Cardiovasc Genet*. 2009;2:467–75.
78. •• Weissglas-Volkov D, Pajukanta P. Genetic causes of high and low serum HDL-cholesterol. *J Lipid Res*. 2010, 51:2032–2057. *This is an in-depth review of the genetic basis of HDL-C.*
79. Lee JC, Weissglas-Volkov D, Kyttala M, Dastani Z, Cantor RM, Sobel EM, Plaisier CL, Engert JC, van Greevenbroek MM, Kane JP, Malloy MJ, Pullinger CR, Huertas-Vazquez A, Aguilar-Salinas CA, Tusie-Luna T, de Bruin TW, Aouizerat BE, van der Kallen CC, Croce CM, Aqeilan RI, Marcil M, Viikari JS, Lehtimaki T, Raitakari OT, Kuusisto J, Laakso M, Taskinen MR, Genest J, Pajukanta P. WW-domain-containing oxidoreductase is associated with low plasma HDL-C levels. *Am J Hum Genet*. 2008;83:180–92.
80. Horie T, Ono K, Horiguchi M, Nishi H, Nakamura T, Nagao K, Kinoshita M, Kuwabara Y, Marusawa H, Iwanaga Y, Hasegawa K, Yokode M, Kimura T, Kita T. MicroRNA-33 encoded by an intron of sterol regulatory element-binding protein 2 (Srebp2) regulates HDL in vivo. *Proc Natl Acad Sci U S A*. 2010;107:17321–6.
81. Ramirez CM, Davalos A, Goedeke L, Salerno AG, Warrior N, Cirera-Salinas D, Suarez Y, Fernandez-Hernando C. MicroRNA-758 Regulates Cholesterol Efflux Through Posttranscriptional Repression of ATP-Binding Cassette Transporter A1. *Arterioscler Thromb Vasc Biol*. 2011.
82. Rayner KJ, Sheedy FJ, Esau CC, Hussain FN, Temel RE, Parathath S, van Gils JM, Rayner AJ, Chang AN, Suarez Y, Fernandez-Hernando C, Fisher EA, Moore KJ. Antagonism of miR-33 in mice promotes reverse cholesterol transport and regression of atherosclerosis. *J Clin Invest*. 2011;121:2921–31.
83. Kim J, Yoon H, Ramirez CM, Lee SM, Hoe HS, Fernandez-Hernando C, Kim J. miR-106b impairs cholesterol efflux and increases Abeta levels by repressing ABCA1 expression. *Exp Neurol*. 2011.
84. 1000 Genomes. <http://www.1000genomes.org/>. Accessed December 2011
85. •• Musunuru K, Pirruccello JP, Do R, Peloso GM, Guiducci C, Sougnez C, Garimella KV, Fisher S, Abreu J, Barry AJ, Fennell T, Banks E, Ambrogio L, Cibulskis K, Kernytzsky A, Gonzalez E, Rudzicz N, Engert JC, DePristo MA, Daly MJ, Cohen JC, Hobbs HH, Altshuler D, Schonfeld G, Gabriel SB, Yue P, Kathiresan S. Exome sequencing, ANGPTL3 mutations, and familial combined hypolipidemia. *N Engl J Med*. 2010, 363:2220–2227. *This article provides a remarkable example of the power of exome-wide sequencing for the identification of genetic defects.*
86. NCBI: National Center for Biotechnology Information. <http://www.ncbi.nlm.nih.gov/>. Accessed December 2011